

**DRAFT  
TOXICOLOGICAL PROFILE FOR  
ZINC**

U.S. DEPARTMENT OF HEALTH AND HUMAN SERVICES  
Public Health Service  
Agency for Toxic Substances and Disease Registry

September 2003

## **DISCLAIMER**

The use of company or product name(s) is for identification only and does not imply endorsement by the Agency for Toxic Substances and Disease Registry.

## UPDATE STATEMENT

A Toxicological Profile for zinc was released in 1994. This edition supersedes any previously released draft or final profile.

Toxicological profiles are revised and republished as necessary, but no less than once every three years. For information regarding the update status of previously released profiles, contact ATSDR at:

Agency for Toxic Substances and Disease Registry  
Division of Toxicology/Toxicology Information Branch  
1600 Clifton Road NE,  
Mailstop E-29  
Atlanta, Georgia 30333



## FOREWORD

This toxicological profile is prepared in accordance with guidelines developed by the Agency for Toxic Substances and Disease Registry (ATSDR) and the Environmental Protection Agency (EPA). The original guidelines were published in the *Federal Register* on April 17, 1987. Each profile will be revised and republished as necessary.

The ATSDR toxicological profile succinctly characterizes the toxicologic and adverse health effects information for the hazardous substance described therein. Each peer-reviewed profile identifies and reviews the key literature that describes a hazardous substance's toxicologic properties. Other pertinent literature is also presented, but is described in less detail than the key studies. The profile is not intended to be an exhaustive document; however, more comprehensive sources of specialty information are referenced.

The focus of the profiles is on health and toxicologic information; therefore, each toxicological profile begins with a public health statement that describes, in nontechnical language, a substance's relevant toxicological properties. Following the public health statement is information concerning levels of significant human exposure and, where known, significant health effects. The adequacy of information to determine a substance's health effects is described in a health effects summary. Data needs that are of significance to protection of public health are identified by ATSDR and EPA.

Each profile includes the following:

- (A) The examination, summary, and interpretation of available toxicologic information and epidemiologic evaluations on a hazardous substance to ascertain the levels of significant human exposure for the substance and the associated acute, subacute, and chronic health effects;
- (B) A determination of whether adequate information on the health effects of each substance is available or in the process of development to determine levels of exposure that present a significant risk to human health of acute, subacute, and chronic health effects; and
- (C) Where appropriate, identification of toxicologic testing needed to identify the types or levels of exposure that may present significant risk of adverse health effects in humans.


The principal audiences for the toxicological profiles are health professionals at the Federal, State, and local levels; interested private sector organizations and groups; and members of the public. We plan to revise these documents in response to public comments and as additional data become available. Therefore, we encourage comments that will make the toxicological profile series of the greatest use.

Comments should be sent to:

Agency for Toxic Substances and Disease Registry  
Division of Toxicology  
1600 Clifton Road, N.E.  
Mail Stop E-29  
Atlanta, Georgia 30333

The toxicological profiles are developed in response to the Superfund Amendments and Reauthorization Act (SARA) of 1986 (Public Law 99-499) which amended the Comprehensive Environmental Response, Compensation, and Liability Act of 1980 (CERCLA or Superfund). This public law directed ATSDR to prepare toxicological profiles for hazardous substances most commonly found at facilities on the CERCLA National Priorities List and that pose the most significant potential threat to human health, as determined by ATSDR and the EPA. The availability of the revised priority list of 275 hazardous substances was announced in the *Federal Register* on October 25, 2001 (66 FR 54014). For prior versions of the list of substances, see *Federal Register* notices dated April 17, 1987 (52 FR 12866); October 20, 1988 (53 FR 41280); October 26, 1989 (54 FR 43619); October 17, 1990 (55 FR 42067); October 17, 1991 (56 FR 52166); October 28, 1992 (57 FR 48801); February 28, 1994 (59 FR 9486); April 29, 1996 (61 FR 18744); November 17, 1997 (62 FR 61332); and October 21, 1999 (64 FR 56792). Section 104(i)(3) of CERCLA, as amended, directs the Administrator of ATSDR to prepare a toxicological profile for each substance on the list.

This profile reflects ATSDR's assessment of all relevant toxicologic testing and information that has been peer-reviewed. Staff of the Centers for Disease Control and Prevention and other Federal scientists have also reviewed the profile. In addition, this profile has been peer-reviewed by a nongovernmental panel and is being made available for public review. Final responsibility for the contents and views expressed in this toxicological profile resides with ATSDR.

  
Julie Louise Gerberding, M.D., M.P.H.  
Administrator  
Agency for Toxic Substances and  
Disease Registry

## QUICK REFERENCE FOR HEALTH CARE PROVIDERS

Toxicological Profiles are a unique compilation of toxicological information on a given hazardous substance. Each profile reflects a comprehensive and extensive evaluation, summary, and interpretation of available toxicologic and epidemiologic information on a substance. Health care providers treating patients potentially exposed to hazardous substances will find the following information helpful for fast answers to often-asked questions.

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### *Primary Chapters/Sections of Interest*

**Chapter 1: Public Health Statement:** The Public Health Statement can be a useful tool for educating patients about possible exposure to a hazardous substance. It explains a substance's relevant toxicologic properties in a nontechnical, question-and-answer format, and it includes a review of the general health effects observed following exposure.

**Chapter 2: Relevance to Public Health:** The Relevance to Public Health Section evaluates, interprets, and assesses the significance of toxicity data to human health.

**Chapter 3: Health Effects:** Specific health effects of a given hazardous compound are reported by type of health effect (death, systemic, immunologic, reproductive), by route of exposure, and by length of exposure (acute, intermediate, and chronic). In addition, both human and animal studies are reported in this section.

**NOTE:** Not all health effects reported in this section are necessarily observed in the clinical setting. Please refer to the Public Health Statement to identify general health effects observed following exposure.

**Pediatrics:** Four new sections have been added to each Toxicological Profile to address child health issues:

<b>Section 1.6</b>	<b>How Can (Chemical X) Affect Children?</b>
<b>Section 1.7</b>	<b>How Can Families Reduce the Risk of Exposure to (Chemical X)?</b>
<b>Section 3.7</b>	<b>Children's Susceptibility</b>
<b>Section 6.6</b>	<b>Exposures of Children</b>

### **Other Sections of Interest:**

<b>Section 3.8</b>	<b>Biomarkers of Exposure and Effect</b>
<b>Section 3.11</b>	<b>Methods for Reducing Toxic Effects</b>

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### **ATSDR Information Center**

**Phone:** 1-888-42-ATSDR or (404) 498-0110    **Fax:** (404) 498-0093  
**E-mail:** [atsdric@cdc.gov](mailto:atsdric@cdc.gov)    **Internet:** <http://www.atsdr.cdc.gov>

The following additional material can be ordered through the ATSDR Information Center:

*Case Studies in Environmental Medicine: Taking an Exposure History*—The importance of taking an exposure history and how to conduct one are described, and an example of a thorough exposure history is provided. Other case studies of interest include *Reproductive and Developmental Hazards*; *Skin Lesions and Environmental Exposures*; *Cholinesterase-Inhibiting Pesticide Toxicity*; and numerous chemical-specific case studies.

*Managing Hazardous Materials Incidents* is a three-volume set of recommendations for on-scene (prehospital) and hospital medical management of patients exposed during a hazardous materials incident. Volumes I and II are planning guides to assist first responders and hospital emergency department personnel in planning for incidents that involve hazardous materials. Volume III—*Medical Management Guidelines for Acute Chemical Exposures*—is a guide for health care professionals treating patients exposed to hazardous materials.

*Fact Sheets (ToxFAQs)* provide answers to frequently asked questions about toxic substances.

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### ***Other Agencies and Organizations***

*The National Center for Environmental Health (NCEH)* focuses on preventing or controlling disease, injury, and disability related to the interactions between people and their environment outside the workplace. Contact: NCEH, Mailstop F-29, 4770 Buford Highway, NE, Atlanta, GA 30341-3724 • Phone: 770-488-7000 • FAX: 770-488-7015.

*The National Institute for Occupational Safety and Health (NIOSH)* conducts research on occupational diseases and injuries, responds to requests for assistance by investigating problems of health and safety in the workplace, recommends standards to the Occupational Safety and Health Administration (OSHA) and the Mine Safety and Health Administration (MSHA), and trains professionals in occupational safety and health. Contact: NIOSH, 200 Independence Avenue, SW, Washington, DC 20201 • Phone: 800-356-4674 or NIOSH Technical Information Branch, Robert A. Taft Laboratory, Mailstop C-19, 4676 Columbia Parkway, Cincinnati, OH 45226-1998 • Phone: 800-35-NIOSH.

*The National Institute of Environmental Health Sciences (NIEHS)* is the principal federal agency for biomedical research on the effects of chemical, physical, and biologic environmental agents on human health and well-being. Contact: NIEHS, PO Box 12233, 104 T.W. Alexander Drive, Research Triangle Park, NC 27709 • Phone: 919-541-3212.

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### ***Referrals***

*The Association of Occupational and Environmental Clinics (AOEC)* has developed a network of clinics in the United States to provide expertise in occupational and environmental issues. Contact: AOEC, 1010 Vermont Avenue, NW, #513, Washington, DC 20005 • Phone: 202-347-4976 • FAX: 202-347-4950 • e-mail: AOEC@AOEC.ORG • Web Page: <http://www.aoec.org/>.

*The American College of Occupational and Environmental Medicine (ACOEM)* is an association of physicians and other health care providers specializing in the field of occupational and environmental medicine. Contact: ACOEM, 55 West Seegers Road, Arlington Heights, IL 60005 • Phone: 847-818-1800 • FAX: 847-818-9266.



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### THE PROFILE HAS UNDERGONE THE FOLLOWING ATSDR INTERNAL REVIEWS:

1. Health Effects Review. The Health Effects Review Committee examines the health effects chapter of each profile for consistency and accuracy in interpreting health effects and classifying end points.
2. Minimal Risk Level Review. The Minimal Risk Level Workgroup considers issues relevant to substance-specific minimal risk levels (MRLs), reviews the health effects database of each profile, and makes recommendations for derivation of MRLs.
3. Data Needs Review. The Research Implementation Branch reviews data needs sections to assure consistency across profiles and adherence to instructions in the Guidance.



## PEER REVIEW

A peer review panel was assembled for zinc. The panel consisted of the following members:

1. Olen Brown, Ph.D., University of Missouri-Columbia, Columbia, Missouri;
2. Robert Michael, Ph.D., RAM TRAC Corporation, Schenectady, New York; and
3. Gary Pascoe, Ph.D., DABT, Pascoe Environmental Consulting, Port Townsend, Washington.

These experts collectively have knowledge of zinc's physical and chemical properties, toxicokinetics, key health end points, mechanisms of action, human and animal exposure, and quantification of risk to humans. All reviewers were selected in conformity with the conditions for peer review specified in Section 104(I)(13) of the Comprehensive Environmental Response, Compensation, and Liability Act, as amended.

Scientists from the Agency for Toxic Substances and Disease Registry (ATSDR) have reviewed the peer reviewers' comments and determined which comments will be included in the profile. A listing of the peer reviewers' comments not incorporated in the profile, with a brief explanation of the rationale for their exclusion, exists as part of the administrative record for this compound. A list of databases reviewed and a list of unpublished documents cited are also included in the administrative record.

The citation of the peer review panel should not be understood to imply its approval of the profile's final content. The responsibility for the content of this profile lies with the ATSDR.



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## 1. PUBLIC HEALTH STATEMENT

This public health statement tells you about zinc and the effects of exposure. Zinc is an essential element needed by your body and is commonly found in nutritional supplements. However, taking too much zinc into the body can affect your health.

The Environmental Protection Agency (EPA) identifies the most serious hazardous waste sites in the nation. These sites make up the National Priorities List (NPL) and are the sites targeted for long-term federal cleanup activities. Zinc has been found in at least 953 of the 1,636 current or former NPL sites. However, the total number of NPL sites evaluated for zinc is not known. As more sites are evaluated, the sites at which zinc is found may increase. This information is important because exposure to zinc may harm you and because these sites may be sources of exposure.

When a substance is released from a large area, such as an industrial plant, or from a container, such as a drum or bottle, it enters the environment. This release does not always lead to exposure. You are exposed to a substance only when you come in contact with it. You may be exposed by breathing, eating, or drinking the substance, or by skin contact.

If you are exposed to zinc, many factors determine whether you'll be harmed. These factors include the dose (how much), the duration (how long), and how you come in contact with it. You must also consider the other chemicals you're exposed to and your age, sex, diet, family traits, lifestyle, and state of health.

### 1.1 WHAT IS ZINC?

Zinc is one of the most common elements in the earth's crust. Zinc is found in the air, soil, and water and is present in all foods. In its pure elemental (or metallic) form, zinc is a bluish-white, shiny metal. Powdered zinc is explosive and may burst into flames if stored in damp places. Metallic zinc has many uses in industry. A common use is to coat iron or other metals to prevent rust and corrosion. Metallic zinc is also mixed with other metals to form alloys such as brass and

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bronze. A zinc and copper alloy is used to make pennies in the United States. Metallic zinc is also used to make dry cell batteries.

Zinc can also combine with other elements, such as chlorine, oxygen, and sulfur, to form zinc compounds. Zinc compounds that may be found at hazardous waste sites are zinc chloride, zinc oxide, zinc sulfate, and zinc sulfide. Most zinc ore found naturally in the environment is in the form of zinc sulfide. Zinc compounds are widely used in industry. Zinc sulfide and zinc oxide are used to make white paints, ceramics, and other products. Zinc oxide is also used in producing rubber. Zinc compounds, such as zinc acetate, zinc chloride, and zinc sulfate, are used in preserving wood and in manufacturing and dyeing fabrics. Zinc chloride is also the major ingredient in smoke from smoke bombs. Zinc compounds are used by the drug industry as ingredients in some common products, such as vitamin supplements, sun blocks, diaper rash ointments, deodorants, athlete's foot preparations, acne and poison ivy preparations, and antidandruff shampoos. Information can be found on the chemical and physical properties of zinc in Chapter 4 and on its occurrence and fate in the environment in Chapter 6.

## 1.2 WHAT HAPPENS TO ZINC WHEN IT ENTERS THE ENVIRONMENT?

Zinc enters the air, water, and soil as a result of both natural processes and human activities. Most zinc enters the environment as the result of mining, purifying of zinc, lead, and cadmium ores, steel production, coal burning, and burning of wastes. These activities can increase zinc levels in the atmosphere. Waste streams from zinc and other metal manufacturing and zinc chemical industries, domestic waste water, and run-off from soil containing zinc can discharge zinc into waterways. The level of zinc in soil increases mainly from disposal of zinc wastes from metal manufacturing industries and coal ash from electric utilities. In air, zinc is present mostly as fine dust particles. This dust eventually settles over land and water. Rain and snow aid in removing zinc from air. Most of the zinc in lakes or rivers settles on the bottom. However, a small amount may remain either dissolved in water or as fine suspended particles. The level of dissolved zinc in water may increase as the acidity of water increases. Fish can collect zinc in their bodies from water containing zinc. Most of the zinc in soil is bound to the soil and does not dissolve in water. However, depending on the type of soil, some zinc may reach groundwater, and contamination of groundwater has occurred from hazardous waste sites. Zinc may be taken

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up by animals eating soil or drinking water containing zinc. For more information about what happens to zinc in the environment, see Chapter 6.

### 1.3 HOW MIGHT I BE EXPOSED TO ZINC?

Zinc is an essential element needed by your body in small amounts. We are exposed to zinc compounds in food. The average daily zinc intake through the diet in this country ranges from 5.2 to 16.2 milligrams (milligram = 0.001 gram). Food may contain levels of zinc ranging from approximately 2 parts of zinc per million (2 ppm) parts of foods (e.g., leafy vegetables) to 29 ppm (meats, fish, poultry). Zinc is also present in most drinking water. Drinking water or other beverages may contain high levels of zinc if they are stored in metal containers or flow through pipes that have been coated with zinc to resist rust. If you take more than the recommended daily amount of supplements containing zinc, you may have higher levels of zinc exposure.

In general, levels of zinc in air are relatively low and fairly constant. Average levels of zinc in the air throughout the United States are less than 1 microgram of zinc per cubic meter ( $\mu\text{g}/\text{m}^3$ ) of air, but range from 0.1 to 1.7  $\mu\text{g}/\text{m}^3$  in areas near cities. Air near industrial areas may have higher levels of zinc. The average zinc concentration for a 1-year period was 5  $\mu\text{g}/\text{m}^3$  in one area near an industrial source.

In addition to background exposure that all of us experience, about 150,000 people also have a source of occupational exposure to zinc that might elevate their total exposure significantly above the average background exposure. Jobs where people are exposed to zinc include zinc mining, smelting, and welding; manufacture of brass, bronze, or other zinc-containing alloys; manufacture of galvanized metals; and manufacture of machine parts, rubber, paint, linoleum, oilcloths, batteries, some kinds of glass and ceramics, and dyes. People at construction jobs, automobile mechanics, and painters are also exposed to zinc. For more information on exposure to zinc, see Chapter 6.

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**1.4 HOW CAN ZINC ENTER AND LEAVE MY BODY?**

Zinc can enter the body through the digestive tract when you eat food or drink water containing it. Zinc can also enter through your lungs if you inhale zinc dust or fumes from zinc-smelting or zinc-welding operations on your job. The amount of zinc that passes directly through the skin is relatively small. The most likely route of exposure near NPL waste sites is through drinking water containing a high amount of zinc. Zinc is stored throughout the body. Zinc increases in blood and bone most rapidly after exposure. Zinc may stay in the bone for many days after exposure. Normally, zinc leaves the body in urine and feces. More information on how zinc enters and leaves your body can be found in Chapter 3.

**1.5 HOW CAN ZINC AFFECT MY HEALTH?**

To protect the public from the harmful effects of toxic chemicals and to find ways to treat people who have been harmed, scientists use many tests.

One way to see if a chemical will hurt people is to learn how the chemical is absorbed, used, and released by the body; for some chemicals, animal testing may be necessary. Animal testing may also be used to identify health effects such as cancer or birth defects. Without laboratory animals, scientists would lose a basic method to get information needed to make wise decisions to protect public health. Scientists have the responsibility to treat research animals with care and compassion. Laws today protect the welfare of research animals, and scientists must comply with strict animal care guidelines.

Inhaling large amounts of zinc (as zinc dust or fumes from smelting or welding) can cause a specific short-term disease called metal fume fever. However, very little is known about the long-term effects of breathing zinc dust or fumes.

Taking too much zinc into the body through food, water, or dietary supplements can also affect health. The levels of zinc that produce adverse health effects are much higher than the Recommended Dietary Allowances (RDAs) for zinc of 11 mg/day for men and 8 mg/day for women. If large doses of zinc (10–15 times higher than the RDA) are taken by mouth even for a

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short time, stomach cramps, nausea, and vomiting may occur. Ingesting high levels of zinc for several months may cause anemia, damage the pancreas, and decrease levels of high-density lipoprotein (HDL) cholesterol.

Eating food containing very large amounts of zinc (1,000 times higher than the RDA) for several months caused many health effects in rats, mice, and ferrets, including anemia and injury to the pancreas and kidney. Rats that ate very large amounts of zinc became infertile. Rats that ate very large amounts of zinc after becoming pregnant had smaller babies. Putting low levels of certain zinc compounds, such as zinc acetate and zinc chloride, on the skin of rabbits, guinea pigs, and mice caused skin irritation. Skin irritation from exposure to these chemicals would probably occur in humans. EPA has determined that because of lack of information, zinc is not classifiable as to its human carcinogenicity.

Consuming too little zinc is at least as important a health problem as consuming too much zinc. Without enough zinc in the diet, people may experience loss of appetite, decreased sense of taste and smell, decreased immune function, slow wound healing, and skin sores. Too little zinc in the diet may also cause poorly developed sex organs and retarded growth in young men. If a pregnant woman does not get enough zinc, her babies may have birth defects.

More information on the health effects linked with exposure to higher-than-normal levels of zinc is presented in Chapter 3.

## 1.6 HOW CAN ZINC AFFECT CHILDREN?

This section discusses potential health effects from exposures during the period from conception to maturity at 18 years of age in humans.

Zinc is essential for proper growth and development of young children. Mothers who did not eat enough zinc during pregnancy had a higher frequency of birth defects and gave birth to smaller children (lower birth weight) than mothers whose zinc levels were sufficient. Very young children who did not receive enough zinc in the diet were smaller, both in length and in body weight, than children who ate enough zinc. Some foods, such as soy-based formulas, contain



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high levels of phytate, which can result in a decreased absorption of zinc in the diet. Too much of these foods may result in effects similar to those that occur when children receive too little zinc in the diet.

Little is known about whether children who eat too much zinc will react differently from adults who have ingested large amounts of zinc. A child who accidentally drank a large amount of a caustic zinc solution was found to have damage to his mouth and stomach, and later to his pancreas, but similar effects have been seen in adults who accidentally drank the same solution.

### **1.7 HOW CAN FAMILIES REDUCE THE RISK OF EXPOSURE TO ZINC?**

If your doctor finds that you have been exposed to significant amounts of zinc, ask whether your children might also be exposed. Your doctor might need to ask your state health department to investigate.

Children living near waste sites containing zinc are likely to be exposed to higher environmental levels of zinc through breathing, drinking contaminated drinking water, touching soil, and eating contaminated soil. Children sometimes eat dirt, which should be discouraged. Parents should supervise to see that children wash their hands frequently and before eating. Parents should consult their family physicians about whether (and how) hand-to-mouth behaviors in their children might be discouraged. A more complete discussion can be found in Section 3.11 of the profile.

Children and adults require a certain amount of zinc in the diet in order to remain healthy. However, overuse of some medicines or vitamin supplements containing zinc might be harmful; these medicines should always be used appropriately. If you are accidentally exposed to large amounts of zinc, consult a physician immediately.

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**1.8 IS THERE A MEDICAL TEST TO DETERMINE WHETHER I HAVE BEEN EXPOSED TO ZINC?**

A medical test exists that can determine whether your body fluids contain high levels of zinc. Samples of blood or feces can be collected in a doctor's office and sent to a laboratory that can measure zinc levels. It is easier for most laboratories to measure zinc in blood than in feces. The presence of high levels of zinc in the feces can mean recent high zinc exposure. High levels of zinc in the blood can mean high zinc consumption and/or high exposure. High zinc levels in blood or feces reflect the level of exposure to zinc. Measuring zinc levels in urine and saliva also may provide information about zinc exposure. Tests to measure zinc in hair may provide information on long-term zinc exposure; however, no useful correlation has been found between hair zinc levels and zinc exposure and these tests are not routinely used. More information on tests to measure zinc in the body can be found in Chapter 7.

**1.9 WHAT RECOMMENDATIONS HAS THE FEDERAL GOVERNMENT MADE TO PROTECT HUMAN HEALTH?**

The federal government develops regulations and recommendations to protect public health. Regulations can be enforced by law. Federal agencies that develop regulations for toxic substances include the Environmental Protection Agency (EPA), the Occupational Safety and Health Administration (OSHA), and the Food and Drug Administration (FDA). Recommendations provide valuable guidelines to protect public health but cannot be enforced by law. Federal organizations that develop recommendations for toxic substances include the Agency for Toxic Substances and Disease Registry (ATSDR) and the National Institute for Occupational Safety and Health (NIOSH).

Regulations and recommendations can be expressed in not-to-exceed levels in air, water, soil, or food that are usually based on levels that affect animals; then they are adjusted to help protect people. Sometimes these not-to-exceed levels differ among federal organizations because of different exposure times (an 8-hour workday or a 24-hour day), the use of different animal studies, or other factors.

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Recommendations and regulations are also periodically updated as more information becomes available. For the most current information, check with the federal agency or organization that provides it. Some regulations and recommendations for zinc include the following:

The federal government has set standards and guidelines to protect individuals from the potential health effects of excessive zinc. EPA has stated that drinking water should contain no more than 5 mg of zinc per liter of water (5 mg/L or 5 ppm) because of taste. Furthermore, any release of more than 1,000 pounds (or in some cases 5,000 pounds) of zinc or its compounds into the environment (i.e., water, soil, or air) must be reported to EPA.

The National Academy of Sciences (NAS) estimates an RDA for zinc of 11 mg/day (men). Eleven mg/day is the same as 0.16 mg per kilogram (kg) of body weight per day for an average adult male (70 kg). An RDA of 8 mg/day, or 0.13 mg per kg of body weight for an average adult female (60 kg), was established for women because they usually weigh less than men. Lower zinc intake was recommended for infants (2–3 mg/day) and children (5–9 mg/day) because of their lower average body weights. The RDA provides a level of adequate nutritional status for most of the population. Extra dietary levels of zinc are recommended for women during pregnancy and lactation. An RDA of 11–12 mg/day was set for pregnant women. Women who nurse their babies need 12–13 mg/day.

To protect workers, OSHA has set an average legal limit of 1 mg/m<sup>3</sup> for zinc chloride fumes and 5 mg/m<sup>3</sup> for zinc oxide (dusts and fumes) in workplace air during an 8-hour workday, 40-hour work week. This regulation means that the workroom air should contain no more than an average of 1 mg/m<sup>3</sup> of zinc chloride over an 8-hour working shift of a 40-hour work week. NIOSH similarly recommends that the level of zinc oxide in workplace air should not exceed an average of 1 mg/m<sup>3</sup> over a 10-hour period of a 40-hour work week. For more information on recommendations and standards for zinc exposure, see Chapter 8.

### 1.10 WHERE CAN I GET MORE INFORMATION?

If you have any more questions or concerns, please contact your community or state health or environmental quality department, or contact ATSDR at the address and phone number below.

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ATSDR can also tell you the location of occupational and environmental health clinics. These clinics specialize in recognizing, evaluating, and treating illnesses resulting from exposure to hazardous substances.

Toxicological profiles are also available on-line at [www.atsdr.cdc.gov](http://www.atsdr.cdc.gov) and on CD-ROM. You may request a copy of the ATSDR ToxProfiles CD-ROM by calling the information and technical assistance toll-free number at 1-888-42ATSDR (1-888-422-8737), by email at [atsdric@cdc.gov](mailto:atsdric@cdc.gov), or by writing at:

Agency for Toxic Substances and Disease Registry  
Division of Toxicology  
1600 Clifton Road NE  
Mailstop E-29  
Atlanta, GA 30333  
Fax: 1-404-498-0093

For-profit organizations may request a copy of final profiles from the following:

National Technical Information Service (NTIS)  
5285 Port Royal Road  
Springfield, VA 22161  
Phone: 1-800-553-6847 or 1-703-605-6000  
Web site: <http://www.ntis.gov/>



## 2. RELEVANCE TO PUBLIC HEALTH

### 2.1 BACKGROUND AND ENVIRONMENTAL EXPOSURES TO ZINC IN THE UNITED STATES

Zinc is ubiquitous in the environment, being released from both natural sources and sources of human origin. Release of zinc into the environment from sources of human origin are from mining, smelting of zinc, lead, and cadmium ores, steel production, coal burning, and burning of wastes. Natural sources of zinc include the weathering of zinc-containing rocks and soils. Ambient background air concentrations of zinc are generally  $<1 \mu\text{g}/\text{m}^3$ . Zinc is found in soils and surficial materials of the contiguous United States at concentrations between  $<5$  and  $2,900 \text{ mg}/\text{kg}$ , with a mean of  $60 \text{ mg}/\text{kg}$ . The zinc background concentrations in surface waters are usually  $<50 \mu\text{g}/\text{L}$ , but natural background concentrations in surface waters and groundwater can range from  $0.002$  to  $50 \text{ mg}/\text{L}$ .

Exposure of the general population to zinc is primarily by ingestion. The average daily intake of zinc from food in humans is  $5.2$ – $16.2 \text{ mg zinc}/\text{day}$ . Zinc is widespread in commonly consumed foods, but tends to be higher in those of animal origin, particularly some sea foods. Meat products contain relatively high concentrations of zinc, whereas fruits and vegetables have relatively low concentrations. Other possible pathways for zinc exposure are water and air. Sources of exposure include drinking water, food, air that is polluted, tobacco products, and occupational exposure. Individuals occupationally exposed to metallic zinc and zinc compounds are those involved in galvanizing, smelting, welding, or brass foundry operations.

### 2.2 SUMMARY OF HEALTH EFFECTS

The effects of inhalation exposure to zinc and zinc compounds vary somewhat with the chemical form of the zinc compound, but the majority of the effects seen will occur within the respiratory tract. Following inhalation of zinc oxide, and to a lesser extent zinc metal and many other zinc compounds, the most commonly reported effect is the development of “metal fume fever.” Metal fume fever is characterized by chest pain, cough, dyspnea, reduced lung volumes, nausea, chills, malaise, and leukocytosis. Symptoms generally appear a few hours after exposure, and are reversible 1–4 days following cessation of exposure. Exposure levels associated with the development of metal fume fever have not been identified, though are generally in the range of  $200$ – $500 \text{ mg zinc}/\text{m}^3$ . Acute experimental exposures to

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lower concentrations of zinc oxide (14 mg/m<sup>3</sup> for 8 hours or 45 mg zinc/m<sup>3</sup> for 20 minutes) and occupational exposures to similar concentrations (8–12 mg zinc/m<sup>3</sup> for 1–3 hours and 0.034 mg zinc/m<sup>3</sup> for 6–8 hours) did not produce symptoms of metal fume fever. In contrast, inhalation of zinc chloride generally results in more pronounced damage to the mucous membranes of the respiratory tract, without the effects normally seen in metal fume fever. Symptoms include dyspnea, cough, pleuritic chest pain, bilateral diffuse infiltrations, pneumothorax, and acute pneumonitis, resulting from respiratory tract irritation.

Acute, high-dose oral exposure to zinc compounds generally results in gastrointestinal distress, with symptoms including nausea, vomiting, abdominal cramps, and diarrhea; exposure levels resulting in these effects generally range from 2 to 8 mg zinc/kg/day. Ingestion of zinc chloride may also result in caustic burns of the gastrointestinal tract, particularly the mouth, esophagus, and stomach. Following longer-term exposure to lower doses (~0.5–2 mg zinc/kg/day) of zinc compounds, the observed symptoms generally result from a decreased absorption of copper from the diet, leading to systemic copper deficiency. The most noticeable manifestation of this is anemia, which has been reported in many studies of zinc supplementation in humans. Zinc administration has also resulted in reductions in leukocyte number and function. Available studies have also associated increased exposure to zinc with decreases in high-density lipoprotein (HDL) levels in humans; however, not all studies have confirmed this observation. Long-term consumption of excess zinc may also result in decreased iron status in women, though whether this is a primary or a secondary effect is not presently clear.

In most cases, dermal exposure to zinc or zinc compounds does not result in any noticeable toxic effects. Zinc oxide is used routinely in topical applications including sunscreens and creams designed to assist in wound healing. However, dermal exposure to zinc chloride, and to a lesser extent other zinc salts, can result in severe skin irritancy, characterized by parakeratosis, hyperkeratosis, inflammatory changes in the epidermis and superficial dermis, and acanthosis of the follicular epithelia.

Available studies have not presented evidence of reproductive or developmental effects in humans or animals following inhalation of zinc compounds. Effects on reproductive or developmental end points have been noted in animal studies by the oral route, but generally only at very high (>200 mg/kg/day) exposure levels.

Occupational studies of zinc inhalation have not shown an increase in the incidence of tumors in exposed workers, relative to controls. A single short-term study of zinc chloride inhalation showed an increase in

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tumor formation in mice, but not in guinea pigs; however, the study design was insufficient to allow for definitive determination of the potential carcinogenicity of zinc. Available studies of zinc-induced carcinogenic effects in humans and animals have not been sufficient to demonstrate an increase in cancer incidence following long-term oral exposure to zinc compounds.

The primary effects of zinc are on the respiratory and gastrointestinal systems, and on hematological endpoints. The reader is referred to Section 3.2, Discussion of Health Effects by Route of Exposure, for additional information on other health effects.

**Respiratory Effects.** In general, exposure to zinc and zinc compounds will result in significant effects on the respiratory system only when the exposure is by the inhalation route. A number of cases of acute, high-dose exposure to zinc chloride smoke from military smoke bombs have been reported. In an early study, cough, dyspnea, burning throat, diffuse infiltrates throughout the lung, chemical pneumonitis, and decreased vital capacity were reported. Other studies have also reported respiratory effects of zinc chloride inhalation, including dyspnea, cough, pleuritic chest pain, bilateral diffuse infiltrations, pneumothorax, and acute pneumonitis from respiratory tract irritation. In some studies, more severe effects have occurred, including ulcerative and edematous changes in mucous membranes, fibrosis, subpleural hemorrhage, advanced pulmonary fibrosis, and fatal respiratory distress syndrome.

Acute inhalation exposure to zinc oxide, and to a lesser extent other zinc compounds, may result in the development of “metal fume fever”, a condition that results in elevated body temperatures as well as a transient impairment of pulmonary function; the impairment generally ceases a few days after removal from exposure. The impairment of pulmonary function is characterized by reduced lung volumes, a decreased diffusing capacity of carbon monoxide, and an increase in bronchiolar leukocytes. Respiratory symptoms include dryness of the throat and coughing, progressing to substernal chest pain, cough, and dyspnea. In general, the symptoms of metal fume fever resolve within 1–4 days after cessation of exposure and do not lead to long-term respiratory effects. Inhalation of “ultrafine” zinc oxide particles may also result in metal fume fever, as well as histologic damage and inflammation of the lung periphery.

Several animal studies have been conducted to quantify specific effects after acute zinc oxide inhalation. As in human exposure, the respiratory system is the primary site of injury following inhalation exposure.



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**Gastrointestinal Effects.** Nausea was reported by humans exposed to high concentrations of zinc oxide fumes and zinc chloride smoke by inhalation, as well as following oral exposure to zinc chloride and zinc sulfate. Other gastrointestinal symptoms of excess zinc exposure include vomiting, abdominal cramps, and diarrhea, in several cases with blood. In general, exposure levels associated with gastrointestinal effects of zinc have not been reported. It is unclear in the majority of human studies whether the gastrointestinal effects seen following zinc inhalation were due to systemic zinc or were the result of direct contact with the gastrointestinal tract following mucociliary clearance of inhaled zinc particles and subsequent swallowing.

**Hematologic Effects.** Leukocytosis persisting for approximately 12 hours after fever dissipates is one of the hallmarks of metal fume fever. Such effects have been observed in a number of case reports of occupational and experimental exposure of humans to zinc oxide fumes. Decreased numbers of red blood cells and hemoglobin were found in several workers with 7–20 years of experience in the galvanizing industry. However, there was excess tobacco use and alcohol consumption by workers and possible concurrent exposure to other chemicals (chloride, sulfide), which confound the study results. No anemia was detected among 12 workers exposed for 4–21 years to zinc oxide fumes in the production of brass alloys. These workers may have also been exposed to magnesium, copper, and aluminum.

Treatment-related changes in hematological parameters have been observed in humans and animals after intermediate or chronic oral exposure to zinc or zinc-containing compounds. Zinc-induced hematological changes are the result of competition with copper during absorption from the small intestine, such that a prolonged increase in dietary zinc results in systemic copper deficiency, leading to anemia. Long-term administration (1–8 years) of zinc supplements has caused anemia in humans. A significant reduction in erythrocyte superoxide dismutase activity (47% decrease), hematocrit (4%), and serum ferritin (23%), compared to pretreatment levels, occurred in female subjects who received supplements of zinc gluconate for 10 weeks. Addition of iron supplements resulted in an increase in serum ferritin, but did not reverse the decrease in erythrocyte superoxide dismutase activity. In a similar study, a decrease in erythrocyte superoxide dismutase activity was reported in male volunteers receiving zinc gluconate supplements for 6 weeks.

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**2.3 MINIMAL RISK LEVELS (MRLs)*****Inhalation MRLs***

No inhalation MRLs have been derived for zinc. A number of acute-duration human studies have identified metal fume fever as an end point of concern; however, no study has clearly identified exposure levels that result in metal fume fever. Animal studies corroborate the effects observed in humans; however, the studies are generally limited in the methods utilized, and other possible targets of toxicity were not examined. Only one intermediate-duration inhalation study in humans was located. In this study, exposure levels were not reported; thus, the study could not be used as the basis for the derivation of an intermediate-duration MRL. No exposure-related effects on lung function were observed in a group of welders chronically exposed to zinc; however, the exposure level was not reported. Thus, no chronic-duration inhalation MRL could be derived.

***Oral MRLs***

No oral acute MRL was derived for zinc. A number of case reports involving acute exposure were located. These reports suggest that the gastrointestinal tract and the pancreas are end points of concern, and that the adrenal cortex and central nervous system may also be affected. However, a great deal of uncertainty exists regarding the exposure levels for these studies. An oral exposure study in sheep was also identified. Because sheep are ruminants, it is doubtful that they are a good model for human toxicity. As no studies sufficient for derivation of an acute oral MRL were available, no value was derived.

- An MRL of 0.3 mg zinc/kg/day has been derived for intermediate oral exposure to zinc.

Two studies in humans, one in men and one in women, have identified effects of oral zinc supplementation on body copper status in subjects given 50 mg supplemental zinc/day. In one, a decrease in erythrocyte superoxide dismutase activity was reported in male volunteers receiving zinc gluconate supplements for 6 weeks (Fischer et al. 1984). In the other, decreases in erythrocyte superoxide dismutase activity, serum ferritin, and hematocrit, were reported in women given daily supplements of zinc gluconate for 10 weeks (Yadrick et al. 1989). Zinc supplementation has been shown to decrease HDL levels with daily doses of at least 50 mg zinc for 12 weeks (Black et al. 1988; Chandra 1984; Hooper et al. 1980). These studies suggest that there is a dose-response trend and that 50 mg/day is the threshold lowest-observed-adverse-effect level (LOAEL) for zinc. Because of its duration, and because it evaluated a more thorough spectrum of end points, the 10-week study in women was chosen as the key study for derivation of an intermediate oral MRL. The MRL was therefore based on hematological

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effects, specifically decreases in erythrocyte superoxide dismutase activity, serum ferritin, and hematocrit, in women given daily supplements of 50 mg zinc as zinc gluconate for 10 weeks (Yadrick et al. 1989). The lowest-observed-adverse-effect level (LOAEL) of 0.83 mg/kg/day was calculated from the reported supplemental dose of 50 mg zinc/day, and based on a reference body weight of 60 kg for women. Initially, it was discussed that a correction for background zinc consumption would be included in the MRL calculation. However, when including the background (cited as 9.28 mg/day from the FDA total diet study [Pennington et al. 1996]), the value of the MRL was basically unchanged. Therefore, no background value was included since this adjustment did not occur in the primary study. The MRL was calculated by applying a composite uncertainty factor of 3 (for uncertainties regarding human variability and the use of a minimal LOAEL) to the lowest-observed-adverse-effect level (LOAEL) of 0.83 mg/kg/day.

- The intermediate oral MRL of 0.3 mg zinc/kg/day has been accepted as the chronic oral MRL.

The chronic oral MRL is expected to be without adverse effects when consumed on a daily basis over a long period of time; neither inducing nutritional deficiency in healthy, non-pregnant, adult humans ingesting the average American diet nor resulting in adverse effects from excess consumption. The MRL was not based on a chronic-duration oral study due to a lack of adequate long-term studies in humans and animals. The chronic human study by Hale et al. (1988) provides support for the Yadrick et al. (1989) study and suggests that hematological effects may occur at higher zinc doses. A significant decrease in red blood cells in females receiving daily supplements of up to 2 mg zinc/kg/day for an average of 8 years was reported by Hale et al. (1988). Furthermore, decreases in serum creatinine, total protein, and uric acid and an increase in mean corpuscular hemoglobin (MCH) were observed in the treated male and female subjects (mean age of 78 years) compared to controls. Several other studies have reported copper deficiency-induced anemia resulting from longer-term exposure to zinc, either via supplements or other sources (Broun et al. 1990; Gyorffy and Chan 1992; Hale et al. 1988; Hoffman et al. 1988; Patterson et al. 1985; Porter et al. 1977; Prasad et al. 1978; Ramadurai et al. 1993; Stroud 1991; Summerfield et al. 1992). However, these studies have generally been reports of a single individual, and the level of exposure has not been reliably identified, making them unsuitable for use in deriving a chronic MRL.

In general, the MRL is an estimate of the daily human exposure to a substance that is likely to be without an appreciable risk of adverse effects (noncarcinogenic) over a specified duration of exposure. The MRLs are based on soluble zinc salts; it is less likely that nonsoluble zinc compounds would have these effects at this level.

### 3. HEALTH EFFECTS

#### 3.1 INTRODUCTION

The primary purpose of this chapter is to provide public health officials, physicians, toxicologists, and other interested individuals and groups with an overall perspective on the toxicology of zinc. It contains descriptions and evaluations of toxicological studies and epidemiological investigations and provides conclusions, where possible, on the relevance of toxicity and toxicokinetic data to public health.

A glossary and list of acronyms, abbreviations, and symbols can be found at the end of this profile.

Zinc is an essential nutrient in humans and animals that is necessary for the function of a large number of metalloenzymes. These enzymes include alcohol dehydrogenase, alkaline phosphatase, carbonic anhydrase, leucine aminopeptidase, superoxide dismutase, and deoxyribonucleic acid (DNA) and ribonucleic acid (RNA) polymerase. As such, zinc is required for normal nucleic acid, protein, and membrane metabolism, as well as cell growth and division. Zinc also plays an essential role in the maintenance of nucleic acid structure of genes (zinc finger phenomenon). Zinc deficiency has been associated with dermatitis, anorexia, growth retardation, poor wound healing, hypogonadism with impaired reproductive capacity, impaired immune function, and depressed mental function; increased incidence of congenital malformations in infants has also been associated with zinc deficiency in the mothers (Cotran et al. 1989; Elinder 1986; Sandstead 1981). Zinc deficiency may also have an impact on carcinogenesis, though the direction of the influence seems to vary with the agent (Fong et al. 1978; Mathur 1979; Wallenius et al. 1979). Therefore, certain levels of zinc intake are recommended. The RDA for zinc is 11 mg/day in men and 8 mg/day in women (IOM 2002). Higher RDAs are recommended for women during pregnancy and lactation (12 mg/day for pregnant women and nursing women).

Just as zinc deficiency has been associated with adverse effects in humans and animals, overexposures to zinc also have been associated with toxic effects. This chapter contains a description of the toxic effects that have been associated with exposures to high levels of zinc and selected zinc compounds by the inhalation, oral, and dermal routes. Specifically, zinc chloride, zinc oxide, zinc sulfate, and zinc sulfide will be discussed. Other zinc compounds are discussed in this chapter whenever data regarding these

### 3. HEALTH EFFECTS

compounds add relevant information to the discussion on zinc. Any general comments regarding the lack of data on zinc refer to both zinc and its compounds.

Because there are differences in toxicity between the various zinc compounds following inhalation exposure, these compounds will be discussed under separate subheadings in Section 3.2.1 (Inhalation Exposure). After oral or dermal exposure, the toxicities are comparable for all zinc compounds. Therefore, in Section 3.2.2 (Oral Exposure) and Section 3.2.3 (Dermal Exposure), the discussion will not be subdivided, but the specific zinc compounds will be identified in each case.

## **3.2 DISCUSSION OF HEALTH EFFECTS BY ROUTE OF EXPOSURE**

To help public health professionals and others address the needs of persons living or working near hazardous waste sites, the information in this section is organized first by route of exposure (inhalation, oral, and dermal) and then by health effect (death, systemic, immunological, neurological, reproductive, developmental, genotoxic, and carcinogenic effects). These data are discussed in terms of three exposure periods: acute (14 days or less), intermediate (15–364 days), and chronic (365 days or more).

Levels of Significant Exposure for each route and duration are presented in tables and illustrated in figures. The points in the figures showing no-observed-adverse-effect levels (NOAELs) or lowest-observed-adverse-effect levels (LOAELs) reflect the actual doses (levels of exposure) used in the studies. LOAELs have been classified into "less serious" or "serious" effects. "Serious" effects are those that evoke failure in a biological system and can lead to morbidity or mortality (e.g., acute respiratory distress or death). "Less serious" effects are those that are not expected to cause significant dysfunction or death, or those whose significance to the organism is not entirely clear. ATSDR acknowledges that a considerable amount of judgment may be required in establishing whether an end point should be classified as a NOAEL, "less serious" LOAEL, or "serious" LOAEL, and that in some cases, there will be insufficient data to decide whether the effect is indicative of significant dysfunction. However, the Agency has established guidelines and policies that are used to classify these end points. ATSDR believes that there is sufficient merit in this approach to warrant an attempt at distinguishing between "less serious" and "serious" effects. The distinction between "less serious" effects and "serious" effects is considered to be important because it helps the users of the profiles to identify levels of exposure at which major health effects start to appear. LOAELs or NOAELs should also help in determining whether or not the effects vary with dose and/or duration, and place into perspective the possible significance of these effects to human health.

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The significance of the exposure levels shown in the Levels of Significant Exposure (LSE) tables and figures may differ depending on the user's perspective. Public health officials and others concerned with appropriate actions to take at hazardous waste sites may want information on levels of exposure associated with more subtle effects in humans or animals (LOAELs) or exposure levels below which no adverse effects (NOAELs) have been observed. Estimates of levels posing minimal risk to humans (Minimal Risk Levels or MRLs) may be of interest to health professionals and citizens alike.

Estimates of exposure levels posing minimal risk to humans (Minimal Risk Levels or MRLs) have been made for zinc. An MRL is defined as an estimate of daily human exposure to a substance that is likely to be without an appreciable risk of adverse effects (noncarcinogenic) over a specified duration of exposure. MRLs are derived when reliable and sufficient data exist to identify the target organ(s) of effect or the most sensitive health effect(s) for a specific duration within a given route of exposure. MRLs are based on noncancerous health effects only and do not consider carcinogenic effects. MRLs can be derived for acute, intermediate, and chronic duration exposures for inhalation and oral routes. Appropriate methodology does not exist to develop MRLs for dermal exposure.

Although methods have been established to derive these levels (Barnes and Dourson 1988; EPA 1990), uncertainties are associated with these techniques. Furthermore, ATSDR acknowledges additional uncertainties inherent in the application of the procedures to derive less than lifetime MRLs. As an example, acute inhalation MRLs may not be protective for health effects that are delayed in development or are acquired following repeated acute insults, such as hypersensitivity reactions, asthma, or chronic bronchitis. As these kinds of health effects data become available and methods to assess levels of significant human exposure improve, these MRLs will be revised.

A User's Guide has been provided at the end of this profile (see Appendix B). This guide should aid in the interpretation of the tables and figures for Levels of Significant Exposure and the MRLs.

#### **3.2.1 Inhalation Exposure**

##### **3.2.1.1 Death**

In humans, death has resulted from acute exposure to zinc compounds. When a high concentration (estimated at 33,000 mg zinc/m<sup>3</sup>) of zinc chloride smoke resulted from the explosion of many generators

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in a tunnel following a bombing raid in World War II, 10 of the 70 exposed people in the tunnel died within 4 days (Evans 1945). The smoke generated contained mainly zinc chloride, but exposure to other constituents, namely zinc oxide, hexachloroethane, calcium silicate, and an igniter, was also possible. Therefore, the deaths resulting from the explosion cannot be conclusively attributed to only exposure to zinc chloride. This is the only human study reporting an estimated exposure level that caused death. Another study reported the death of a fireman exposed to the contents of a smoke bomb in a closed environment (Milliken et al. 1963). The man died 18 days after exposure because of respiratory difficulty. Again, exposure to zinc chloride was simultaneous with exposure to other substances in the smoke. Two soldiers exposed without gas masks to zinc chloride smoke during military training developed severe adult respiratory distress syndrome (ARDS) and died 25–32 days after the incident (Hjortso et al. 1988). Diffuse microvascular obliteration, widespread occlusion of the pulmonary arteries, and extensive interstitial and intra-alveolar fibrosis were observed at autopsy. Zinc levels in major organs and tissues obtained during autopsy were within the normal range, and no zinc particles were observed by scanning electron microscopy. According to the authors, the fumes from the smoke bombs consisted mainly of zinc chloride. However, no exposure levels were estimated, and other substances were also present in the smoke.

A case study presented by Murray (1926) reported on an infant death due to bronchopneumonia resulting from inhalation, and possibly ingestion, of an unspecified amount of zinc stearate powder spilled from a container. However, it is unclear whether the death was due to the zinc content or whether aspiration bronchopneumonia would result from inhalation of similar powders that do not contain zinc.

In mice, the reported  $LCT_{50}$  (product of lethal concentration and time to kill 50% of animals) of zinc chloride is 11,800 mg-min/m<sup>3</sup> (Schenker et al. 1981). However, Schenker et al. (1981) did not provide information on how this value was determined. Following exposure to zinc chloride smoke for 3–20 weeks, mortality was 50% in mice exposed to 121.7 mg zinc/m<sup>3</sup> (compared to 20% in controls) and 22% in guinea pigs exposed to 119.3 mg zinc/m<sup>3</sup> (compared to 8% in controls) (Marrs et al. 1988). The smoke was similar to that described by Evans (1945) and also contained zinc oxide, hexachloroethane, and other compounds.

#### 3.2.1.2 Systemic Effects

No studies were located regarding musculoskeletal, endocrine, dermal, or body weight effects in humans or animals after inhalation exposure to zinc or zinc compounds. The systemic effects observed after

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inhalation exposure are discussed below. In most cases, the effects of zinc are discussed without separating effects caused by the individual zinc compounds. However, the respiratory effects of the individual zinc compounds are discussed separately because the nature of the respiratory toxicity differs depending on the particular compound to which one is exposed. The highest NOAEL values and all LOAEL values from each reliable study for systemic effects in each species and duration category are recorded in Table 3-1 and plotted in Figure 3-1.

**Respiratory Effects.**

**Zinc Oxide.** Metal fume fever, a well-documented acute disease induced by intense inhalation of metal oxides, especially zinc, impairs pulmonary function but does not progress to chronic lung disease (Brown 1988; Drinker and Drinker 1928; Malo et al. 1990). Symptoms generally appear within a few hours after acute exposure, usually with dryness of the throat and coughing (Drinker et al. 1927b). The most prominent respiratory effects of metal fume fever are substernal chest pain, cough, and dyspnea (Rohrs 1957). The impairment of pulmonary function is characterized by reduced lung volumes and a decreased diffusing capacity of carbon monoxide (Malo et al. 1990; Vogelmeier et al. 1987). The respiratory effects have been shown to be accompanied by an increase in bronchiolar leukocytes (Vogelmeier et al. 1987). The respiratory symptoms generally disappear in the exposed individual within 1–4 days (Brown 1988; Drinker et al. 1927b; Sturgis et al. 1927). Inhalation of zinc oxide is most likely to occur in occupational situations where zinc smelting or welding take place. Ultrafine zinc oxide particles (0.2–1.0  $\mu\text{m}$ ) originate from heating zinc beyond its boiling point in an oxidizing atmosphere. Upon inhalation, these small particles ( $<1 \mu\text{m}$ ) reach the alveoli and cause inflammation and tissue damage in the lung periphery (Brown 1988; Drinker et al. 1927b; Vogelmeier et al. 1987).

A number of studies have measured exposure levels associated with metal fume fever. Workers involved in pouring molten zinc reported shortness of breath and chest pains 2–12 hours following exposure to 320–580 mg zinc/m<sup>3</sup> as zinc oxide for 1–3 hours (Hammond 1944); the number of workers was not reported. Two volunteers had nasal passage irritation, cough, substernal chest pain, persistent rales of the lung base, and a decreased vital capacity for approximately 3–49 hours following acute inhalation (10–12 minutes) of 600 mg zinc/m<sup>3</sup> as zinc oxide (Sturgis et al. 1927). This study is limited due to an inadequate number of subjects, a lack of controls, and a lack of analysis of the final aerosol product. A subject experimentally exposed to zinc oxide fumes reported mild pain when breathing deeply the next day after a 5-hour exposure to 430 mg zinc/m<sup>3</sup> (Drinker et al. 1927a). Minimal changes in forced expiratory flow were observed 1 hour after a 15–30-minute exposure to 77 mg zinc/m<sup>3</sup> as zinc oxide (Blanc et al. 1991).



Table 3-1 Levels of Significant Exposure to Zinc - Inhalation

Key to figure <sup>a</sup>	Species (Strain)	Exposure/ Duration/ Frequency (Specific Route)	System	NOAEL (mg/m <sup>3</sup> )	LOAEL		Reference  Chemical Form
					Less Serious (mg/m <sup>3</sup> )	Serious (mg/m <sup>3</sup> )	
<b>ACUTE EXPOSURE</b>							
<b>Systemic</b>							
1	Human	15-30 min	Resp		77	(minimal change in pulmonary function)	Blanc et al. 1991 Zinc oxide
2	Human	1 d 2hr/d	Resp		3.9	(dry or sore throat, chest tightness)	Gordon et al. 1992 Zinc oxide
			Other		3.9	(fever/chills and headache)	
3	Human	2 hr	Resp	0.0036			Linn et al. 1981 Zinc amm sulfate
4	Human	6-8 hr (Occup)	Resp	0.034 M			Marquart et al. 1989 Zinc oxide
5	Human	10.5-12 min	Resp			600 M (decreased vital capacity)	Sturgis et al. 1927 Zinc oxide
			Gastro			600 M (nausea)	
			Hemato			600 M (increased leukocytes)	
6	Rat (Fischer- 344)	1 d 3hr/d	Resp		2.2	(increased LDH protein in bronchoalveolar lavage fluid)	Gordon et al. 1992 Zinc oxide
7	Gn Pig	1 hr	Resp			0.73 M (decrease in lung compliance)	Amdur et al. 1982 Zinc oxide
8	Gn Pig	1-3 d 3hr/d	Resp	1.8 M	4.7 M	(increased neutrophils, LDH, and protein in bronchoalveolar lavage fluid)	Conner et al. 1988 Zinc oxide

Table 3-1 Levels of Significant Exposure to Zinc - Inhalation

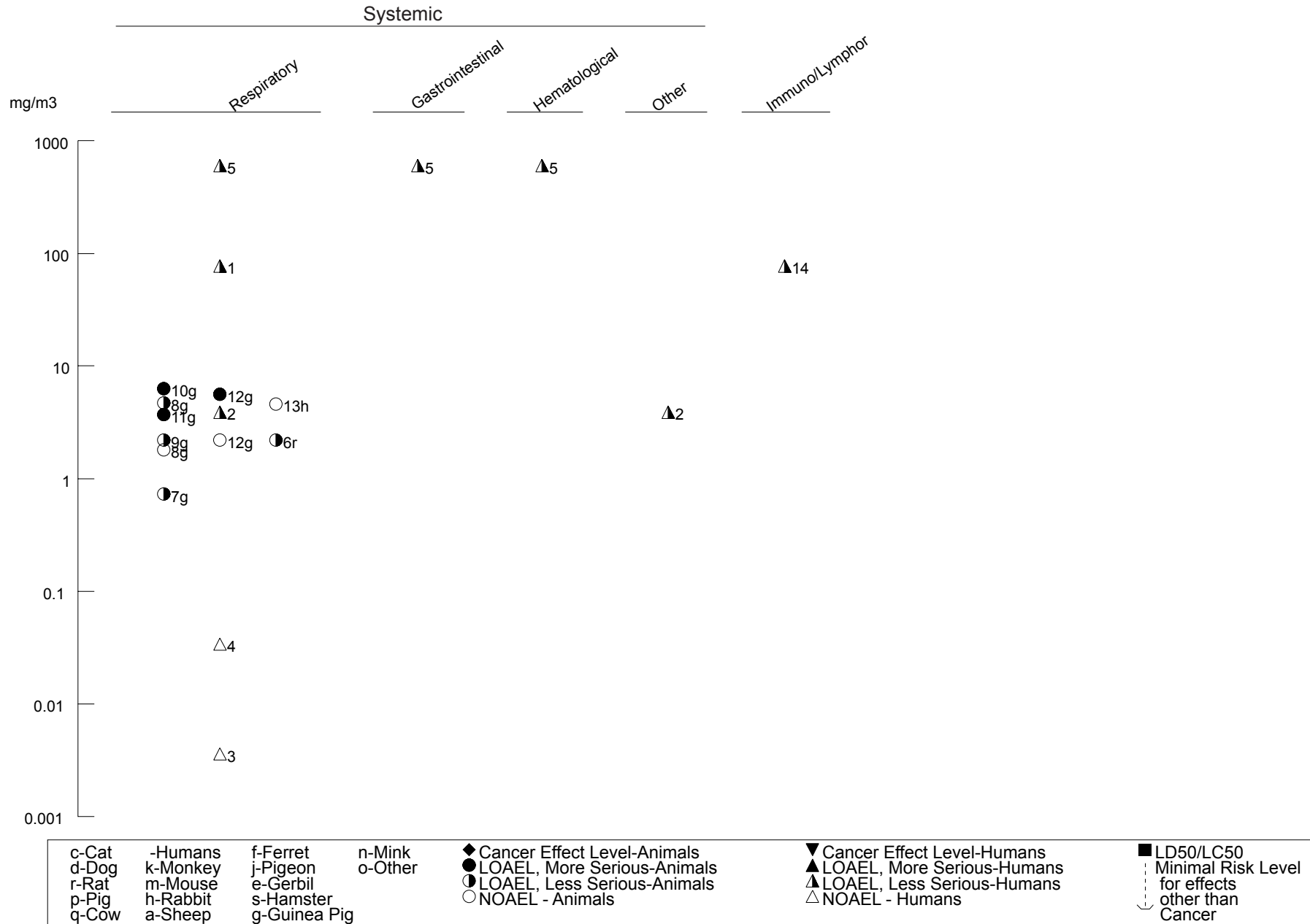
(continued)

Key to figure <sup>a</sup>	Species (Strain)	Exposure/ Duration/ Frequency (Specific Route)	System	LOAEL			Reference Chemical Form
				NOAEL (mg/m <sup>3</sup> )	Less Serious (mg/m <sup>3</sup> )	Serious (mg/m <sup>3</sup> )	
9	Gn Pig (Hartley)	1 d 3hr/d	Resp		2.2 (increased LDH and protein in bronchoalveolar lavage fluid)		Gordon et al. 1992 Zinc oxide
10	Gn Pig (Hartley)	3 hr	Resp			6.3 M (decreased functional residual capacity)	Lam et al. 1982 Zinc oxide
11	Gn Pig (Hartley)	6d 3hr/d	Resp			3.7 M (impaired lung function; inflammation; increased pulmonary resistance; increased lung weight)	Lam et al. 1985 Zinc oxide
12	Gn Pig (Hartley)	5 d 3hr/d	Resp	2.2 M		5.6 M (impaired lung function; increased lung weight)	Lam et al. 1988 Zinc oxide
13	Rabbit (New Zealand)	1 d 2hr/d	Resp	4.6			Gordon et al. 1992 Zinc oxide
14	Human	15-30 min			77 (increased number of leukocytes, T cells, T suppressor cells, and natural killer cells in bronchoalveolar lavage fluid)		Blanc et al. 1991 Zinc oxide

<sup>a</sup> The numbers corresponds to entries in Figure 3-1.

amm sulfate = ammonium sulfate; d = day(s); Gastro = gastrointestinal; Gn pig = guinea pig; Hemato = hematological; hr = hour(s); LDH = lactate dehydrogenase; LOAEL = lowest-observed-adverse-effect level; min = minute(s); NOAEL = no-observed-adverse-effect level; (occup) = occupational; Resp = respiratory

Figure 3-1. Levels of Significant Exposure to Zinc - Inhalation  
Acute ( $\leq 14$  days)



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Acute experimental exposures to lower concentrations of zinc oxide (14 mg/m<sup>3</sup> for 8 hours or 45 mg zinc/m<sup>3</sup> for 20 minutes) and occupational exposures to similar concentrations (8–12 mg zinc/m<sup>3</sup> for 1–3 hours and 0.034 mg zinc/m<sup>3</sup> for 6–8 hours) did not produce symptoms of metal fume fever (Drinker et al. 1927b; Hammond 1944; Marquart et al. 1989). In a single-blind experiment, exposure of subjects to 3.9 mg zinc/m<sup>3</sup> as zinc oxide resulted in sore throat and chest tightness but no impairment of pulmonary function (Gordon et al. 1992). It is speculated that subjects in other studies may have been less susceptible because of the development of tolerance to zinc (Gordon et al. 1992). Recurrent episodes of cough and dyspnea were reported in a former mild smoker 3 years after beginning work in a metal foundry where exposure to zinc oxide presumably occurred (Ameille et al. 1992). This case was distinguishable from metal fume fever because of the lack of tolerance to zinc (as shown by the late emergence of symptoms).

Several animal studies have been conducted to quantify specific effects after acute zinc oxide inhalation. As in human exposure, the respiratory system is the primary site of injury following inhalation exposure. Acute administration of 88–482 mg zinc/m<sup>3</sup> as zinc oxide to rats and rabbits resulted in the following pulmonary changes: grayish areas with pulmonary congestion, various degrees of peribronchial leukocytic infiltration, and bronchial exudate composed almost entirely of polymorphonuclear leukocytes (Drinker and Drinker 1928). Cats similarly exposed exhibited more severe effects including bronchopneumonia, leukocyte infiltration into alveoli, and grayish areas with congestion. During the exposure period, the cats demonstrated labored breathing and evidence of upper respiratory tract obstruction. A minimum effect level could not be determined for any species because the concentration varied widely (88–482 mg zinc/m<sup>3</sup>) during exposure.

Guinea pigs administered 0.73 mg zinc/m<sup>3</sup> as zinc oxide for 1 hour had a progressive decrease in lung compliance but no change in air flow resistance. These observations reflect a response in the lung periphery where submicrometer aerosols are likely to deposit (Amdur et al. 1982). The authors postulated that reduced compliance may be associated with human metal fume fever.

In contrast to the results of Amdur et al. (1982), no effects on ventilation, lung mechanics (respiratory frequency, tidal volume, pulmonary resistance, and pulmonary compliance), diffusing capacity of carbon monoxide, or most lung volume parameters were observed by Lam et al. (1982) following the exposure of guinea pigs to 6.3 mg zinc/m<sup>3</sup> as zinc oxide for 3 hours. However, functional residual capacity was significantly decreased. The discrepancy between the results of Amdur et al. (1982) and Lam et al.

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(1982) may be attributable to the use of anesthetized animals by Lam et al. (1982). In a later study, exposures of guinea pigs to 3.7 or 4.3 mg zinc/m<sup>3</sup> as zinc oxide for 3 hours/day, for 6 days, resulted in transient functional, morphological, and biochemical changes (Lam et al. 1985). Functional changes included increased flow resistance, decreased lung compliance, and decreased diffusing capacity, all of which returned to normal within 24–72 hours following exposure. The morphological changes (increased lung weight, inflammation involving the proximal portion of alveolar ducts and adjacent alveoli, interstitial thickening, and increased pulmonary macrophages and neutrophils in adjacent air spaces) were, however, still present at 72 hours. In guinea pigs with evidence of an inflammatory reaction involving the peripheral airways, DNA synthesis increased in bronchiolar cells. Similarly, exposure of guinea pigs to 5.6 mg zinc/m<sup>3</sup> as zinc oxide for 3 hours/day, for 5 days, resulted in gradual decreases in total lung capacity, vital capacity, and decreased carbon monoxide diffusing capacity (Lam et al. 1988); however, no effects were observed in guinea pigs exposed to 2.2 mg zinc/m<sup>3</sup>. The reason that effects have been seen in the guinea pig at exposure levels lower than humans may have to do with the structural features of the guinea pig lung. The bronchi and peripheral airways of guinea pigs have a thicker smooth muscle layer and only a small surface area covered by alveolar sacs compared to the bronchi and peripheral airways of other laboratory animals and humans. This makes the guinea pig more susceptible than other laboratory animals to functional impairment of the peripheral airways and should be noted in toxicity comparisons (Lam et al. 1982).

The bronchoalveolar lavage fluid of rats or guinea pigs exposed to 1.8 mg zinc/m<sup>3</sup> as zinc oxide for 3 hours contained increased levels of lactate dehydrogenase and total protein, suggesting effects on cell viability or membrane permeability (Gordon et al. 1992). Rabbits were not affected following a similar exposure to 4.6 mg zinc/m<sup>3</sup> for 2 hours. Guinea pigs had foci of inflammation after exposure to 4.7 mg zinc/m<sup>3</sup> for 3 days, and the bronchoalveolar lavage fluid contained increased levels of protein, angiotensin converting enzyme, and neutrophils (Conner et al. 1988). No significant changes in respiratory effects were observed in this study following exposure to 1.8 mg zinc/m<sup>3</sup> for 3 days.

***Zinc Chloride.*** Zinc chloride, a corrosive inorganic salt, is more damaging than zinc oxide to the mucous membranes of the nasopharynx and respiratory tract upon contact. Zinc chloride is a primary ingredient in smoke bombs used by the military for screening purposes, crowd dispersal, and occasionally in military and civilian fire-fighting exercises. Reports of serious respiratory injury have been reported to result from accidental inhalation of smoke from these bombs. These reports are of limited use in assessing the toxicity of zinc chloride because exposure to other compounds, usually hexachloroethane, zinc oxide, and calcium silicides, also occur. Furthermore, the specific concentrations inhaled are usually unknown.

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Despite these limitations, several case studies have described similar respiratory effects in humans following acute inhalation exposures. These effects include dyspnea, cough, pleuritic chest pain, bilateral diffuse infiltrations, pneumothorax, and acute pneumonitis from respiratory tract irritation (Johnson and Stonehill 1961; Matarese and Matthews 1966; Schenker et al. 1981; Zerahn et al. 1999). In the study by Johnson and Stonehill (1961), cough, dyspnea, burning throat, diffuse infiltrates throughout the lung, chemical pneumonitis, and decreased vital capacity were observed at an estimated zinc chloride exposure level of 4,075 mg/m<sup>3</sup> (1,955 mg zinc/m<sup>3</sup>). In other studies, more severe effects have occurred, including ulcerative and edematous changes in mucous membranes, fibrosis, subpleural hemorrhage, advanced pulmonary fibrosis, and fatal respiratory distress syndrome (Evans 1945; Hjortso et al. 1988; Homma et al. 1992; Milliken et al. 1963).

Focal alveolitis, consolidation, emphysema, infiltration with macrophages, and fibrosis were observed in guinea pigs that died following exposure to 119 mg zinc/m<sup>3</sup> as zinc chloride smoke for 1 hour/day, 5 days/week, for 3–20 weeks (Marrs et al. 1988); no changes were seen in guinea pigs that survived 13 months after the 20-week exposure. Thirteen months after a 20-week exposure, rats similarly exposed to 12.8 mg zinc/m<sup>3</sup> showed an increase in peribronchial inflammatory cell (lymphocytes and macrophage) infiltration. Mice exposed to 121.7 mg zinc/m<sup>3</sup> as zinc chloride smoke, but not to lower doses, for 1 hour/day, 5 days/week, showed increased macrophages and lymphocytes in the lungs (Marrs et al. 1988). The smoke also contained zinc oxide, hexachloroethane, and other compounds.

***Zinc Ammonium Sulfate.*** Zinc ammonium sulfate is a compound emitted during combustion of fossil fuels and is, therefore, found in the ambient air. Humans acutely exposed to a concentration of 0.0036 mg zinc/m<sup>3</sup> as zinc ammonium sulfate for 2 hours (Linn et al. 1981) exhibited minimal or no short-term respiratory effects (including minimal substernal irritation, throat irritation, and coughing in asthmatic subjects). However, most human exposures to an ambient air pollutant such as zinc ammonium sulfate are chronic, and this study provides little information about the health effects associated with typical exposures.

No studies were located regarding respiratory effects in animals after inhalation exposure to zinc ammonium sulfate.

***Zinc Stearate.*** Inhalation of zinc stearate powder resulted in aspiration bronchopneumonia in an infant (Murray 1926). However, it is unclear whether the bronchopneumonia resulted from the inhalation of zinc stearate powder specifically or from a nonspecific effect of the inhalation of powders.

## 3. HEALTH EFFECTS

No studies were located regarding respiratory effects in animals after inhalation exposure to zinc stearate.

**Cardiovascular Effects.** No atypical heart sounds or blood pressure abnormalities were observed in 24 employees occupationally exposed to concentrations as high as 130 mg zinc/m<sup>3</sup> of metallic zinc dust, zinc oxide dust, zinc sulfide dust, or lithophone dust (a combination of barium sulphate and ≈30% zinc sulphide) for 2–35.5 years (Batchelor et al. 1926). However, this study is limited because only selected employees were examined.

Only limited information was located regarding cardiovascular effects in animals following inhalation exposure to zinc. Routine gross and microscopic examination of the hearts of rats and mice revealed no adverse effects 13 months after exposure to 121.7 mg zinc/m<sup>3</sup> as zinc chloride smoke (also containing other compounds) for 1 hour/day, 5 days/week, for 20 weeks (Marrs et al. 1988). Similarly, no changes were observed in the hearts of guinea pigs exposed to 119.3 mg zinc/m<sup>3</sup> as zinc chloride smoke for 1 hour/day, 5 days/week, for 20 weeks, and then observed for an additional 17 months (Marrs et al. 1988).

**Gastrointestinal Effects.** Nausea was reported by humans exposed to high concentrations of zinc oxide fumes (Hammond 1944; Rohrs 1957; Sturgis et al. 1927) and zinc chloride smoke (Evans 1945; Johnson and Stonehill 1961; Schenker et al. 1981). The zinc chloride smoke also contained zinc oxide, hexachloroethane, and other compounds. In general, exposure levels associated with nausea have not been reported. However, exposures to 320 mg zinc/m<sup>3</sup> as zinc oxide for 1–3 hours (Hammond 1944) or 600 mg zinc/m<sup>3</sup> as zinc oxide for 10–12 minutes (Sturgis et al. 1927) were reported to have resulted in nausea; it should be noted, however, that the zinc used in these studies contained slight impurities (i.e., lead, magnesium). Autopsies of victims who died following exposure to very high concentrations of zinc chloride smoke revealed irritation of the stomach and intestines (Evans 1945). The smoke also contained zinc oxide, hexachloroethane, and other compounds. Workers in the galvanizing industry were found by McCord et al. (1926) to have a higher than expected incidence of gastrointestinal problems; however, these individuals may have been exposed to other chemicals (arsenic, hydrogen sulfide). Of 15 workers examined with 7–20 years of experience, 12 had frequent episodes of epigastric or abdominal pain, nausea, vomiting, ulcers, constipation, tarry stools, and/or gas. It is unclear whether these effects were due to systemic zinc or were the result of direct contact with the gastrointestinal tract following mucociliary clearance of inhaled zinc particles and subsequent swallowing. In contrast, 24 workers with 2–35.5 years of exposure to ≤130 mg zinc/m<sup>3</sup> as metallic zinc dust, zinc sulfide dust, zinc oxide, or lithophone dust reported no nausea or vomiting and only occasional mild abdominal discomfort that could

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not be attributed with certainty to zinc exposure (Batchelor et al. 1926). A study examining the acidity of the stomach contents after stimulation in controls and workers employed in the production of brass alloys showed that stomach acidity was similar in the two groups prior to stimulation but remained elevated for longer periods after stimulation in the exposed workers (Hamdi 1969). This was proposed to account for the gastric complaints of workers exposed to zinc fumes. Despite these findings, x-rays showed no lesions in the stomachs or duodenum of exposed workers.

The only information available regarding gastrointestinal effects in animals was found in a study by Marrs et al. (1988) in which rats and mice were exposed to 121.7 mg zinc/m<sup>3</sup> as zinc chloride smoke (which also contains zinc oxide, hexachlorophene, and other compounds) for 1 hour/day, 5 days/week, for 20 weeks, and then observed for an additional 13 months. In the same study, guinea pigs were exposed to 119.3 mg zinc/m<sup>3</sup> as zinc chloride smoke for 1 hour/day, 5 days/week, for 3 weeks. All animals were sacrificed at the end of 18 months. Routine gross and microscopic evaluation of the stomach and intestines at 18 months revealed no persistent adverse effects.

**Hematological Effects.** Leukocytosis persisting for approximately 12 hours after fever dissipates is one of the hallmarks of metal fume fever (Mueller and Seger 1985). Such effects have been observed in a number of case reports of occupational and experimental exposure of humans to zinc oxide fumes (Brown 1988; Drinker et al. 1927a; Malo et al. 1990; Rohrs 1957; Sturgis et al. 1927). Increased leukocyte counts were observed following experimental exposures to 430 mg zinc/m<sup>3</sup> as zinc oxide for 3 hours (Drinker et al. 1927a) or 600 mg zinc/m<sup>3</sup> as zinc oxide for 10–12 minutes (Sturgis et al. 1927). These studies are limited in that they used an inadequate number of subjects, lacked controls, and used impure zinc oxide.

Decreased numbers of red blood cells and hemoglobin were found in several workers with 7–20 years of experience in the galvanizing industry (McCord et al. 1926). However, there were excess tobacco use and alcohol consumption by workers and possible concurrent exposure to other chemicals (chloride, sulfide), which confound the study results. No anemia was detected among 12 workers exposed for 4–21 years to zinc oxide fumes in the production of brass alloys (Hamdi 1969). These workers may have also been exposed to magnesium, copper, and aluminum.

No studies were located regarding hematological effects in animals after inhalation exposure to zinc.



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**Hepatic Effects.** Routine blood chemistries and examinations revealed no liver disease among 12 workers with 4–21 years of exposure to zinc oxide fumes in the production of brass alloys (Hamdi 1969).

No adverse effects were observed during gross and microscopic examination of livers of rats and guinea pigs exposed to 121.7 mg zinc/m<sup>3</sup> or 119.3 mg zinc/m<sup>3</sup>, respectively, as zinc chloride smoke for 1 hour/day, 5 days/week, for 20 weeks, and sacrificed at the end of 18 months (Marrs et al. 1988). Significant increases in the incidence of fatty liver were observed in mice exposed to 12.8 or 121.7 mg zinc/m<sup>3</sup> as zinc chloride smoke using the same exposure paradigm; however, the incidence did not increase with dose (Marrs et al. 1988). The smoke contained other compounds in addition to zinc chloride.

**Renal Effects.** Urinalyses and histories of urinary function revealed no adverse effects in 24 workers exposed for 2–35.5 years to  $\leq 130$  mg zinc/m<sup>3</sup> as metallic zinc dust, zinc sulfide dust, zinc oxide, or lithophone dust (Batchelor et al. 1926).

No adverse effects were observed following gross and microscopic examination of kidneys from rats, mice, and guinea pigs exposed for 1 hour/day, 5 days/week, for 20 weeks, to concentrations as high as 121.7 or 119.3 mg zinc/m<sup>3</sup> as zinc chloride smoke (which also contained other compounds) and then sacrificed 13 months later (Marrs et al. 1988).

**Ocular Effects.** Reddened conjunctiva and corneal burns occurred in individuals exposed to high concentrations of zinc chloride smoke (estimated at 33,000 mg zinc/m<sup>3</sup>) when several smoke generators exploded in a tunnel during World War II (Evans 1945). The ocular effects may have been due to direct contact with the smoke.

**Homeostatic Effects.** A fever appearing 3–10 hours after exposure to zinc oxide fumes and lasting approximately 24–48 hours is characteristic of metal fume fever caused by zinc (Mueller and Seger 1985). Elevated body temperature has been observed in a number of experimental and occupational zinc oxide exposures (Brown 1988; Drinker et al. 1927a; Hammond 1944; Malo et al. 1990; Rohrs 1957; Sturgis et al. 1927; Vogelmeier et al. 1987). Using a number of exposure concentrations for various durations, Drinker et al. (1927b) found that the increase in body temperature was dependent on the exposure duration and concentration. Based on their data, they calculated that the threshold for pyrogenic effects was 45 mg zinc/m<sup>3</sup> for 20 minutes. This study is limited in that impurities were present in the zinc

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used and no statistical analysis was performed. Exposure to zinc chloride smoke (which also contains other compounds) has also been associated with fever (Hjortso et al. 1988; Matarese and Matthews 1966).

No studies were located regarding other systemic effects in animals following inhalation exposure to zinc.

#### **3.2.1.3 Immunological and Lymphoreticular Effects**

One report described hives and angioedema in a man exposed to zinc fumes at a zinc smelting plant (Farrell 1987). The author suggested that the patient had an immediate or delayed immunoglobulin E (IgE) response (or both) after a low dose of zinc fumes. Metal fume fever also resulted when the exposure increased. The signs and symptoms of toxicity were repeated in a challenge test conducted at the patient's home.

In a group of 14 welders acutely exposed to 77–153 mg zinc/m<sup>3</sup> as zinc oxide, significant correlations between the concentration of airborne zinc and the proportion of activated T cells, T helper cells, T inducer cells, T suppressor cells, and activated killer T cells were observed 20 hours after exposure (Blanc et al. 1991). In addition, significant increases in levels of polymorphonuclear leukocytes, macrophages, and all types of lymphocytes were observed in the bronchoalveolar lavage fluid 20 hours after exposure. Increased levels of lymphocytes, with a predominance of CD8 cells, in the bronchoalveolar lavage fluid were reported in a case study of a smelter exposed to unspecified levels of zinc fumes (Ameille et al. 1992).

The bronchoalveolar lavage fluid of rats or guinea pigs exposed to 2.2 mg zinc/m<sup>3</sup> for 3 hours contained increased levels of  $\beta$ -glucuronidase, suggesting a change in macrophage function (Gordon et al. 1992). Rabbits were not affected following a similar exposure to 4.6 mg zinc/m<sup>3</sup> for 2 hours. Rats, mice, and guinea pigs were exposed to concentrations as high as 119.3 or 121.7 mg zinc/m<sup>3</sup> as zinc chloride smoke for 1 hour/day, 5 days/week, for 20 weeks (Marrs et al. 1988). Routine gross and histopathologic examination of the lymph nodes, thymus, and spleen at the end of 18 months revealed no adverse effects. The smoke also contained zinc oxide, hexachlorophene, and other compounds.

#### **3.2.1.4 Neurological Effects**

Humans have reported nonspecific neurological effects such as headaches and malaise in association with other symptoms following inhalation of zinc oxide and in metal fume fever (Rohrs 1957; Sturgis et al.

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1927). Staggering gait, hallucinations, and hilarity were observed in an individual who intentionally inhaled aerosols of metallic paint containing copper and zinc (Wilde 1975). However, it is most likely that these effects were due to exposure to hydrocarbon propellant rather than zinc. Amr et al. (1997) reported an increase in neuropsychiatric symptoms, including fear of poisoning, headache, nervousness, insomnia, and changes in EEG, in workers who were occupationally-exposed to zinc phosphide for a period of many years; however, exposure levels were not reported, and no tests of statistical significance were performed.

No studies were located regarding neurological effects in animals after inhalation exposure to zinc.

#### **3.2.1.5 Reproductive Effects**

No studies were located regarding reproductive effects in humans after inhalation exposure to zinc.

Following an initial exposure of rats, mice, and guinea pigs to concentrations as high as 119.3 or 121.7 mg zinc/m<sup>3</sup> as zinc chloride smoke (which also contained other compounds) for 1 hour/day, 5 days/week, for 20 weeks; histological evaluation revealed no adverse effects on the mammary glands, ovaries, fallopian tubes, or uteri were observed at 18 months (Marrs et al. 1988).

#### **3.2.1.6 Developmental Effects**

No studies were located regarding developmental effects in humans or animals after inhalation exposure to zinc.

#### **3.2.1.7 Cancer**

In two epidemiological studies, workers did not have an increased incidence of cancers associated with occupational exposure (primarily inhalation exposure) to zinc (Logue et al. 1982; Neuberger and Hollowell 1982).

Workers in nine electrolytic zinc and copper refining plants were studied by Logue et al. (1982). The workers at two of these plants were exposed to zinc or zinc and copper; the other workers were exposed to copper. An association between cancer mortality and zinc exposure was not found.

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Excess lung cancer mortality associated with residence in an old lead/zinc mining and smelting area of the midwestern United States was studied by Neuberger and Hollowell (1982). The age- and sex-adjusted mortality rates were compared to state and national rates. The analysis determined that lung cancer mortality was elevated in the region but was not found to be associated with exposure to environmental levels of lead or zinc. Many confounding factors were not considered in the analysis, such as smoking, occupation, and duration of residence in the area in question.

Female Porton strain mice (98–100/group) exposed to 121.7 mg zinc/m<sup>3</sup> of a zinc oxide/hexachloroethane smoke mixture (which produces zinc chloride), 1 hour/day, 5 days/week, for 20 weeks had a statistically significant increase in the incidence of alveogenic carcinoma (30 versus 8% in control) thirteen months after the end of exposure (Marrs et al. 1988). No increased tumor incidences were seen in mice exposed to 1, 1.3, or 12.8 mg zinc/m<sup>3</sup>. Guinea pigs and rats were also tested with similar dose levels, and no significant carcinogenic response was observed. A number of factors limit the usefulness of this study, including the presence of several compounds in the smoke that may have carcinogenic potential, the use of only female animals, and the short duration of the exposure (20 weeks).

#### 3.2.2 Oral Exposure

Zinc has been orally administered in a variety of forms, such as zinc chloride, zinc sulfate, zinc oxide, powdered zinc, and others. Some of these compounds, such as zinc sulfate, have been administered in both hydrated and anhydrous forms. Study authors often do not state definitely which form was used in a particular study. Knowledge of the form used and its molecular weight is necessary to calculate the amount of elemental zinc administered under a given set of circumstances. If adequate information was not reported by the study authors, it was assumed that an anhydrous compound was used.

##### 3.2.2.1 Death

In a case report presented by Murray (1926), an infant died from bronchopneumonia resulting from inhalation and ingestion of an unspecified amount of zinc stearate powder spilled from a container. However, the cause of death (bronchopneumonia) suggests that it resulted from the inhalation exposure, rather than the oral exposure, and it is unclear whether the lung damage resulted from the inhalation of zinc stearate powder specifically or from a general effect of the inhalation of powders.

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The LD<sub>50</sub> values of several zinc compounds (ranging from 186 to 623 mg zinc/kg/day) have been determined in rats and mice (Domingo et al. 1988a). In general, mice appear to be more sensitive than rats to the lethal effects of zinc. In rats, zinc acetate was the most lethal compound tested; zinc nitrate, zinc chloride, and zinc sulfate (in order of decreasing toxicity) were less lethal. In mice, the most lethal compound was zinc acetate followed by zinc nitrate, zinc sulfate, and zinc chloride. Ingestion of 390 mg zinc/kg/day as zinc oxide in the diet for 3–13 days was lethal to 3 of 3 ferrets (Straube et al. 1980). An equivalent dose in humans would be approximately 27 g zinc/day (which would probably be intolerable to humans because of gastric discomfort). Death was reported in mice that consumed 1,110 mg zinc/kg/day as zinc sulfate in their diet for 13 weeks (Maita et al. 1981). Mortality was also observed in 20% of rats ingesting 191 mg zinc/kg/day as zinc acetate in drinking water for 3 months (Llobet et al. 1988a).

The LD<sub>50</sub> values and all LOAEL values from each reliable study for death in each species and duration category are recorded in Table 3-2 and plotted in Figure 3-2.

#### 3.2.2.2 Systemic Effects

Ingestion of zinc or zinc-containing compounds has resulted in a variety of systemic effects in the gastrointestinal and hematological systems and alterations in the blood lipid profile in humans and animals. In addition, lesions have been observed in the liver, pancreas, and kidneys of animals. No studies were located regarding respiratory, ocular, or metabolic effects in humans or animals after oral exposure to zinc.

Observed systemic effects after oral exposure are discussed below. The highest NOAEL values and all LOAEL values from each reliable study for systemic effects in each species and duration category are recorded in Table 3-2 and plotted in Figure 3-2. The effects discussed in case reports are not included in Table 3-2 or Figure 3-2 because of the small sample size and lack of control data.

**Cardiovascular Effects.** A number of studies in humans and animals have examined the effects of zinc on serum cholesterol and triglycerides. However, no studies regarding the direct relationship between excessive zinc intake and cardiac mortality were located. No effects on electrocardiographic results were found in a group of elderly subjects (>65 years of age) taking zinc supplements of up to 2 mg zinc/kg/day (Hale et al. 1988) or 0.71 mg zinc/kg/day (Czerwinski et al. 1974). There was also no

Table 3-2 Levels of Significant Exposure to Zinc - Oral

Key to figure <sup>a</sup>	Species (Strain)	Exposure/ Duration/ Frequency (Specific Route)	System	LOAEL			Reference Chemical Form
				NOAEL (mg/kg/day)	Less Serious (mg/kg/day)	Serious (mg/kg/day)	
<b>ACUTE EXPOSURE</b>							
<b>Death</b>							
1	Rat (Sprague- Dawley)	once (G)				237 M (LD50)	Domingo et al. 1988a Zinc acetate
2	Rat (Sprague- Dawley)	once (G)				623 M (LD50)	Domingo et al. 1988a Zinc sulfate
3	Rat (Sprague- Dawley)	once (G)				528 M (LD50)	Domingo et al. 1988a Zinc chloride
4	Rat (Sprague- Dawley)	once (G)				293 M (LD50)	Domingo et al. 1988a Zinc nitrate
5	Mouse (Swiss- Webster)	once (G)				337 M (LD50)	Domingo et al. 1988a Zinc sulfate
6	Mouse (Swiss- Webster)	once (G)				86 M (LD50)	Domingo et al. 1988a Zinc acetate
7	Mouse (Swiss- Webster)	once (G)				605 M (LD50)	Domingo et al. 1988a Zinc chloride
8	Mouse (Swiss- Webster)	once (G)				204 M (LD50)	Domingo et al. 1988a Zinc nitrate

Table 3-2 Levels of Significant Exposure to Zinc - Oral

(continued)

Key to figure <sup>a</sup>	Species (Strain)	Exposure/ Duration/ Frequency (Specific Route)	System	LOAEL			Reference Chemical Form
				NOAEL (mg/kg/day)	Less Serious (mg/kg/day)	Serious (mg/kg/day)	
9	Ferret	<2 wk (F)				390 (3/3 died)	Straube et al. 1980 Zinc oxide
<b>Systemic</b>							
10	Human	once (W)	Endocr		0.5 (decreased serum cortisol levels)		Brandao-Neto et al. 1990a Zinc sulfate
11	Human	once (W)	Gastro		6.7 (gastrointestinal distress; diarrhea)		Callender and Gentzkow 1937 Zinc oxide
12	Human	Single oral exposure (IN)	Gastro		6.8 M (Transient nausea, lasting approximately 6 hours)		Lewis and Kokan 1998 Zinc gluconate
13	Human	2 d (F)	Gastro	86 M			Murphy 1970 Zinc elemental
			Endocr		86 M (increased serum amylase, lipase)		
<b>Neurological</b>							
14	Rat	10 d 1x/d (G)			487 (minor neuronal degeneration; decreased acid phosphatase and acetylcholinesterase; increased thiamine pyrophosphatase)		Kozik et al. 1980 Zinc oxide
<b>INTERMEDIATE EXPOSURE</b>							
<b>Death</b>							
15	Rat	3 mo ad lib (W)				191 F (2/10 died)	Llobet et al. 1988a Zinc acetate

Table 3-2 Levels of Significant Exposure to Zinc - Oral

(continued)

Key to figure <sup>a</sup>	Species (Strain)	Exposure/ Duration/ Frequency (Specific Route)	System	LOAEL			Reference Chemical Form
				NOAEL (mg/kg/day)	Less Serious (mg/kg/day)	Serious (mg/kg/day)	
16	Mouse	13 wk ad lib (F)				1110 (5/24 died)	Maita et al. 1981 Zinc sulfate
	<b>Systemic</b>						
17	Human	12 wk 1x/d (C)	Other		0.71 M (decrease serum HDL-cholesterol)		Black et al. 1988 Zinc gluconate
18	Human	3 mo 7d/wk 1x/d (C)	Other	1.5			Bogden et al. 1988 Zinc acetate
19	Human	6 wk 2x/d (C)	Other		4.3 M (increased serum LDL-cholesterol; decreased serum HDL-cholesterol)		Chandra 1984 Zinc sulfate
20	Human	24 wk 7d/wk 3x/d (C)	Cardio	0.71			Czerwinski et al. 1974 Zinc sulfate
21	Human	6 wk 7d/wk 2x/d (C)	Hemato		0.71 (decreased superoxide dismutase activity)		Fischer et al. 1984 Zinc gluconate
22	Human	5 wk 2x/d (C)	Other		2.3 M (decreased serum HDL-cholesterol)		Hooper et al. 1980 Zinc sulfate



Table 3-2 Levels of Significant Exposure to Zinc - Oral

(continued)

Key to figure <sup>a</sup>	Species (Strain)	Exposure/ Duration/ Frequency (Specific Route)	System	LOAEL			Reference Chemical Form
				NOAEL (mg/kg/day)	Less Serious (mg/kg/day)	Serious (mg/kg/day)	
23	Human	6 wk 3x/d (F)	Gastro		2 (abdominal cramps; vomiting; nausea)		Samman and Roberts 1987 Zinc sulfate
24	Human	6 wk 7d/wk 3x/d (C)	Other	2.4			Samman and Roberts 1988 Zinc sulfate
25	Human	10 wk 7d/wk 2x/d (C)	Hemato		0.83 F <sup>b</sup> (decreased superoxide dismutase activity, hematocrit, and serum ferritin)		Yadrick et al. 1989 Zinc gluconate
26	Rat	14 wk (males) or 20 wk (females) 7 d/wk 1x/d (GW)	Bd Wt		7 M (decreased postpartum body weights in F0 animals)		Khan et al. 2001b Zinc chloride
27	Rat	6 wk 7d/wk ad lib (F)	Hemato		6 M (ceroplasmin reduced by 28%)		L'Abbe and Fischer 1984a Zinc sulfate

Table 3-2 Levels of Significant Exposure to Zinc - Oral

(continued)

Key to figure <sup>a</sup>	Species (Strain)	Exposure/ Duration/ Frequency (Specific Route)	System	LOAEL			Reference Chemical Form
				NOAEL (mg/kg/day)	Less Serious (mg/kg/day)	Serious (mg/kg/day)	
28	Rat (Sprague- Dawley)	3 mo ad lib (W)	Hemato	191 F			Llobet et al. 1988a Zinc acetate
			Hepatic	191 F			
			Renal	95 F		191 F (increased plasma creatine and urea levels; desquamation of epithelial cells of proximal tubules)	
			Bd Wt	191 F			
29	Rat	13 wk ad lib (F)	Gastro	565 F			Maita et al. 1981 Zinc sulfate
			Hemato	53 F	565 F (decreased hematocrit and WBC)		
			Musc/skel	565 F			
			Renal	565 F			
			Other	53 M		565 F (acinar cell necrosis and metaplasia in pancreas)	
30	Rat (Sprague- Dawley)	5 wk ad lib (F)	Hemato		500 (decreased Hb, hematocrit, MCH, MCHC; slightly increased WBC)		Smith and Larson 1946 Zinc carbonate
31	Rat (Sprague- Dawley)	6 wk ad lib (F)	Hemato		350 (decreased Hb)		Smith and Larson 1946 Zinc carbonate

Table 3-2 Levels of Significant Exposure to Zinc - Oral

(continued)

Key to figure <sup>a</sup>	Species (Strain)	Exposure/ Duration/ Frequency (Specific Route)	System	LOAEL		Reference Chemical Form	
				NOAEL (mg/kg/day)	Less Serious (mg/kg/day)		Serious (mg/kg/day)
32	Rat (Wistar)	4 wk 7d/wk ad lib (W)	Hemato		12 (decreased Hb and erythrocytes)	Zaporowska and Wasilewski 1992 Zinc chloride	
33	Mouse	5-14 mo ad lib (W)	Other		70 (hypertrophy and vacuolation of pancreas islet cells; hypertrophy and vacuolation of fasciculata cells in adrenal cortex)	Aughey et al. 1977 Zinc sulfate	
34	Mouse (ICR)	13 wk ad lib (F)	Gastro	104 M		1110 F (forestomach ulcers)	Maita et al. 1981 Zinc sulfate
			Hemato	104 M	1110 F (decreased WBC; anemia)		
			Renal	104 M	1110 F (unspecified regressive lesions)		
			Endocr	104 M		1110 F (acinar cell necrosis and metaplasia in pancreas)	
35	Mouse	9 mo ad lib (F)	Hemato			68 (severe anemia)	Walters and Roe 1965 Zinc oleate
36	Dog	9 mo ad lib (W)	Musc/skel	4 M			Anderson and Danylichuk 1979 Zinc oxide
37	Rabbit (New Zealand)	22 wk daily (F)	Hemato		174 M (slight decrease in Hb levels)		Bentley and Grubb 1991 Zinc carbonate
			Bd Wt	174 M			

Table 3-2 Levels of Significant Exposure to Zinc - Oral

(continued)

Key to figure <sup>a</sup>	Species (Strain)	Exposure/ Duration/ Frequency (Specific Route)	System	LOAEL			Reference Chemical Form
				NOAEL (mg/kg/day)	Less Serious (mg/kg/day)	Serious (mg/kg/day)	
38	Mink	144 d ppd70-214 (F)	Hemato	323.6			Aulerich et al. 1991 Zinc sulfate
			Hepatic	323.6			
			Renal	323.6			
			Bd Wt	323.6			
39	Cow	5 wk 2x/d ppd3-40 (F)	Hemato		64 M (decreased hematocrit levels)		Jenkins and Hidiroglou 1991 Zinc oxide
			Bd Wt	64 M		91 M (body weight gain decreased by 46%)	
40	Ferret	7-97 d ad lib (F)	Gastro	195		390 (intestinal hemorrhages)	Straube et al. 1980 Zinc oxide
			Hemato	65	195 (anemia)		
			Renal	65	195 (nephrosis)		
			Endocr	195	390 (pancreatitis)		
41	Human	3 mo 7d/wk 1x/d (C)					Bogden et al. 1988 Zinc acetate

Table 3-2 Levels of Significant Exposure to Zinc - Oral

(continued)

Key to figure <sup>a</sup>	Species (Strain)	Exposure/ Duration/ Frequency (Specific Route)	System	LOAEL			Reference Chemical Form
				NOAEL (mg/kg/day)	Less Serious (mg/kg/day)	Serious (mg/kg/day)	
42	Human	6 wk 2x/d (C)			4.3 M (impaired lymphocyte and polymorphonuclear leukocyte function)		Chandra 1984 Zinc sulfate
43	Human	1 mo 2x/d (C)		2.5			Duchateau et al. 1981 Zinc sulfate
44	Mouse	8 wk 7d/wk ad lib (F)		6.5			Fernandes et al. 1979 ns
45	Mouse (BALB/c)	continuously for 42 days (W)			136 Increases in direct plaque-forming activity of spleen cells and in lymphocyte proliferation in response to mitogen stimulation.		Lastra et al. 1997 ns
46	Mouse	4 wk 7d/wk ad lib (F)		76.9 F			Schiffer et al. 1991 Zinc sulfate
<b>Neurological</b>							
47	Mouse (Swiss-Webster)	drinking water for 60 days (W)			0.5 Increase in latency in inhibitory avoidance test.		Oliveira et al. 2001 Zinc acetate
<b>Reproductive</b>							
48	Human	Gwk 20 through parturition (C)		0.3 F			Mahomed et al. 1989 Zinc sulfate

Table 3-2 Levels of Significant Exposure to Zinc - Oral

(continued)

Key to figure <sup>a</sup>	Species (Strain)	Exposure/ Duration/ Frequency (Specific Route)	System	LOAEL			Reference Chemical Form
				NOAEL (mg/kg/day)	Less Serious (mg/kg/day)	Serious (mg/kg/day)	
49	Rat	8 wk 7d/wk ad lib (F)			25 M (altered sperm chromatin structure)		Evenson et al. 1993 Zinc chloride
50	Rat	14 wk (males) or 20 wk (females) 7 d/wk 1x/d (GW)		3.5 F	7 F (Decreased live pups per litter in all groups of treated rats)		Khan et al. 2001b Zinc chloride
51	Rat	18 d Gd0-18 ad lib (F)				200 F (increased pre-implantation loss)	Pal and Pal 1987 Zinc sulfate
52	Rat	150 d ad lib (F)		50		250 (no reproduction in females)	Sutton and Nelson 1937 Zinc carbonate
53	Mouse (BALB/c)	continuously for 42 days (W)		273 F			Lastra et al. 1997 ns
54	Mouse (ICR)	13 wk ad lib (F)		1110			Maita et al. 1981 Zinc sulfate
55	Mink	approx 25 wk ad lib (F)		20.8			Bleavins et al. 1983 Zinc sulfate

Table 3-2 Levels of Significant Exposure to Zinc - Oral

(continued)

Key to figure <sup>a</sup>	Species (Strain)	Exposure/ Duration/ Frequency (Specific Route)	System	LOAEL			Reference Chemical Form
				NOAEL (mg/kg/day)	Less Serious (mg/kg/day)	Serious (mg/kg/day)	
<b>Developmental</b>							
56	Human	11 wk 1x/d (C)		0.06 F			Kynast and Saling 1986 Zinc aspartate
57	Human	Gwk 20 through parturition (C)		0.3 F			Mahomed et al. 1989 Zinc sulfate
58	Human	last 15- 25 wk of preg- nancy 1x/d (C)		0.3 F			Simmer et al. 1991 Zinc citrate
59	Rat	7 wk Gd0-17 ad lib (F)		250 F			Kinnamon 1963 Zinc carbonate
60	Rat (Sprague- Dawley)	15 d Gd1-15 ad lib (F)				200 F (29% fetal resorption; decreased fetal weight)	Schlicker and Cox 1968 Zinc oxide
61	Rat	36 d Gd1-15 ad lib (F)		100 F			Schlicker and Cox 1968 Zinc oxide

Table 3-2 Levels of Significant Exposure to Zinc - Oral

(continued)

Key to figure <sup>a</sup>	Species (Strain)	Exposure/ Duration/ Frequency (Specific Route)	System	LOAEL			Reference Chemical Form
				NOAEL (mg/kg/day)	Less Serious (mg/kg/day)	Serious (mg/kg/day)	
62	Rat (Sprague- Dawley)	36 d Gd1-21 ad lib (F)				200 F (100% fetal resorption)	Schlicker and Cox 1968 Zinc oxide
63	Rat	150 d ad lib (F)		50		250 (increased stillbirths)	Sutton and Nelson 1937 Zinc carbonate
64	Rat (Sprague- Dawley)	20 d Gd0-20 ad lib (F)		25 F			Uriu-Hare et al. 1989 Zinc carbonate
65	Mouse	2 gen (F)			260 (alopecia; decreased hematocrit)		Mulhern et al. 1986 Zinc carbonate
66	Mink	approx 25 wk ad lib (F)		20.8			Bleavins et al. 1983 Zinc sulfate
<b>CHRONIC EXPOSURE</b>							
<b>Systemic</b>							
67	Human	(C)	Cardio	2			Hale et al. 1988
			Hemato		2 F (decreased RBC)		



Table 3-2 Levels of Significant Exposure to Zinc - Oral

(continued)

Key to figure <sup>a</sup>	Species (Strain)	Exposure/ Duration/ Frequency (Specific Route)	System	LOAEL			Reference Chemical Form
				NOAEL (mg/kg/day)	Less Serious (mg/kg/day)	Serious (mg/kg/day)	
			Endocr	2			

a The number corresponds to entries in Figure 3-2.

b Used to derive an intermediate-duration oral minimal risk level (MRL) of 0.3 mg/kg/day; The MRL was calculated by applying a composite uncertainty factor of 3 (for uncertainties regarding human variability and the use of a minimal LOAEL) to the lowest-observed-adverse-effect level (LOAEL) of 0.83 mg/kg/day. The intermediate oral MRL was adopted as the chronic oral MRL.

ad lib = ad libitum; approx = approximately; (C) = capsule; Cardio = cardiovascular; d - day(s); (F) = feed; (G) = gavage; Gastro = gastrointestinal; Gd = gestation day; gen = generation; Gwk = gestation week; Hb = hemoglobin; HDL = high density lipoprotein; Hemato = hematological; LD50 = lethal dose, 50% kill; LDL = low density lipoprotein; LOAEL = lowest-observed-adverse-effect level; MCH = mean corpuscular hemoglobin; MCHC = mean corpuscular hemoglobin concentration; mo = month(s); Musc/skel = musculoskeletal; NOAEL = no-observed-adverse-effect level; ppd - post partum day; RBC = red blood cell; (W) = drinking water; WBC = white blood cell; wk = week(s); x = time(s); yr = year(s)

Figure 3-2. Levels of Significant Exposure to Zinc - Oral  
Acute ( $\leq 14$  days)

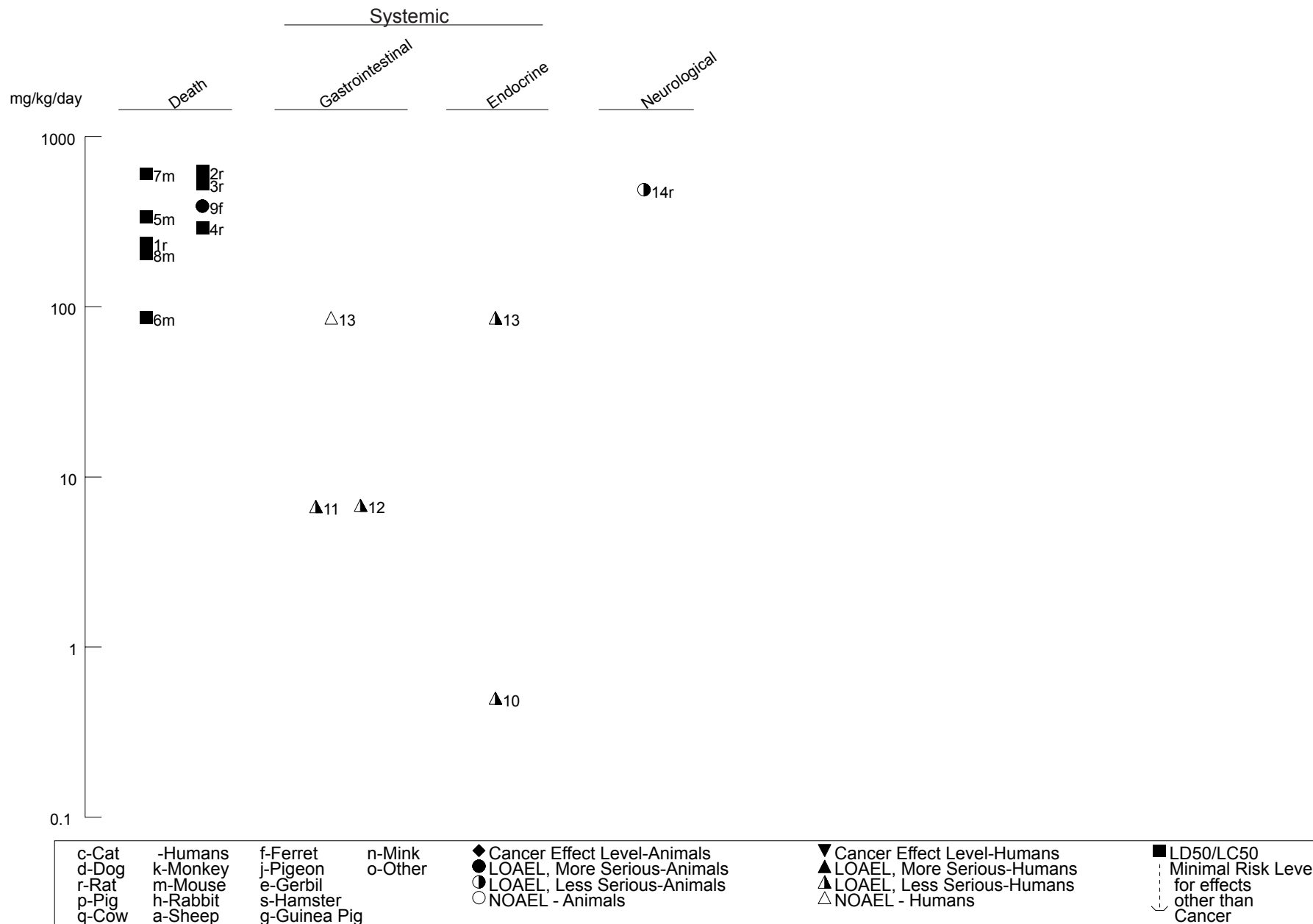


Figure 3-2. Levels of Significant Exposure to Zinc - Oral (Continued)

Intermediate (15-364 days)

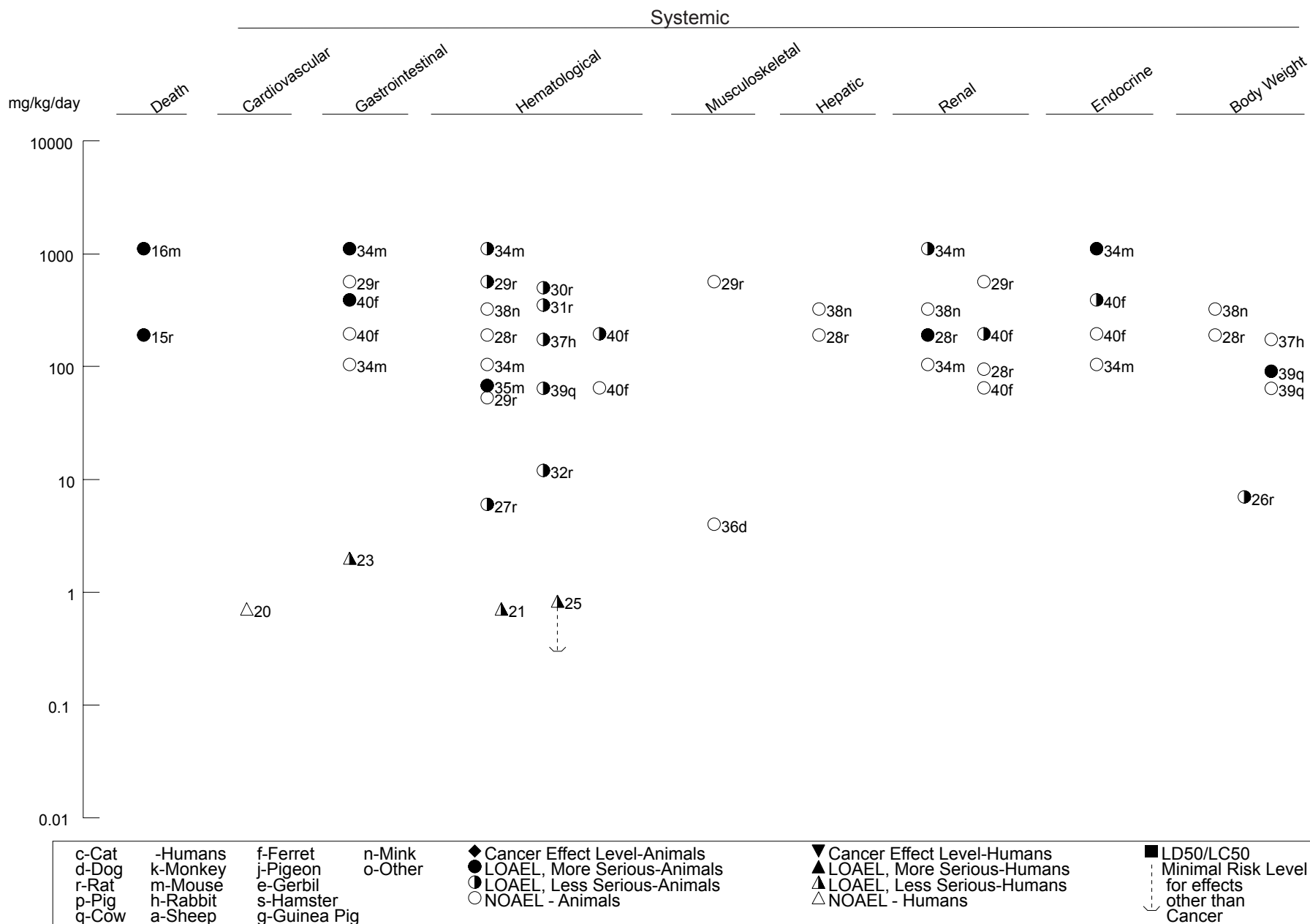


Figure 3-2. Levels of Significant Exposure to Zinc - Oral (Continued)  
Intermediate (15-364 days)

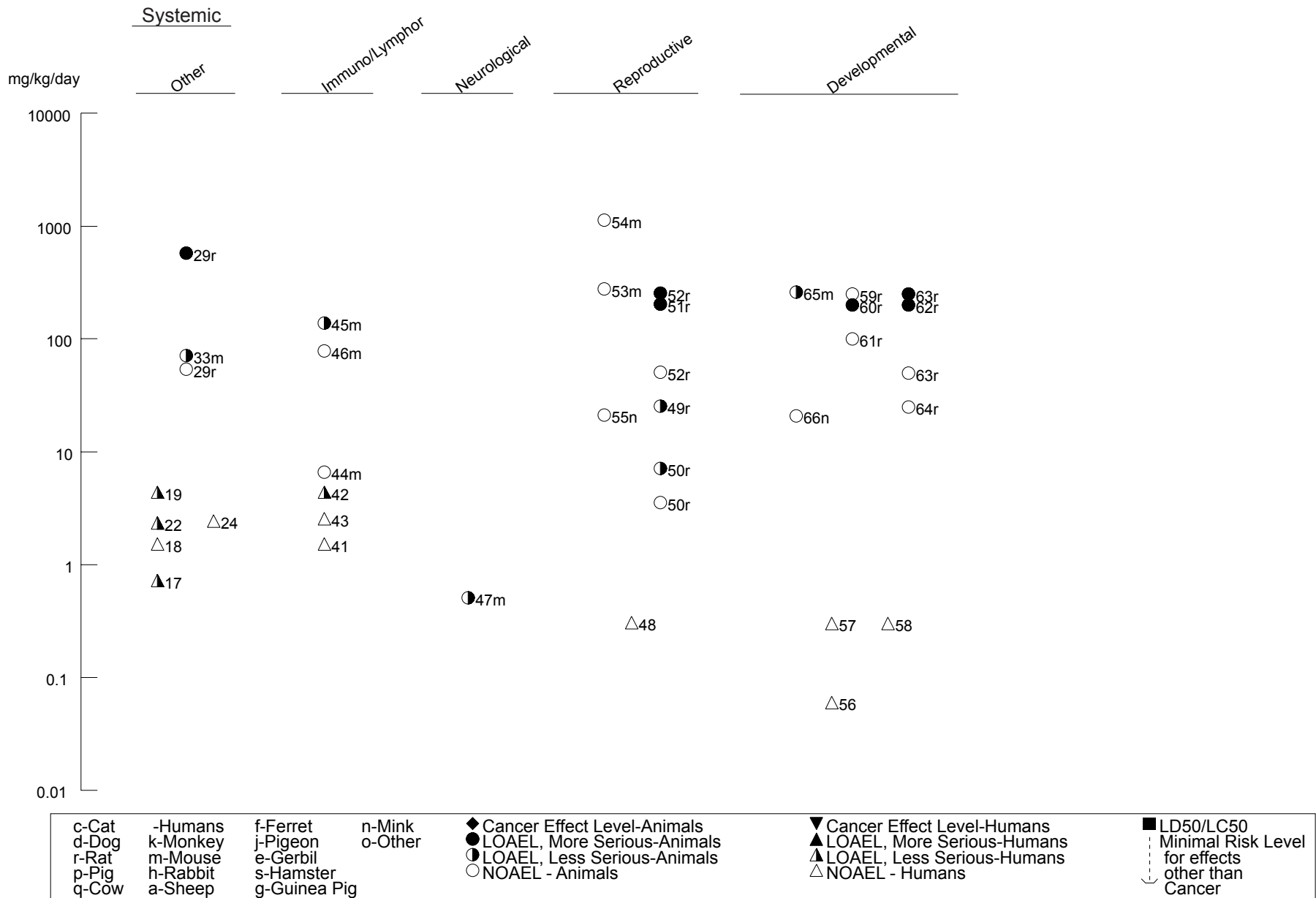
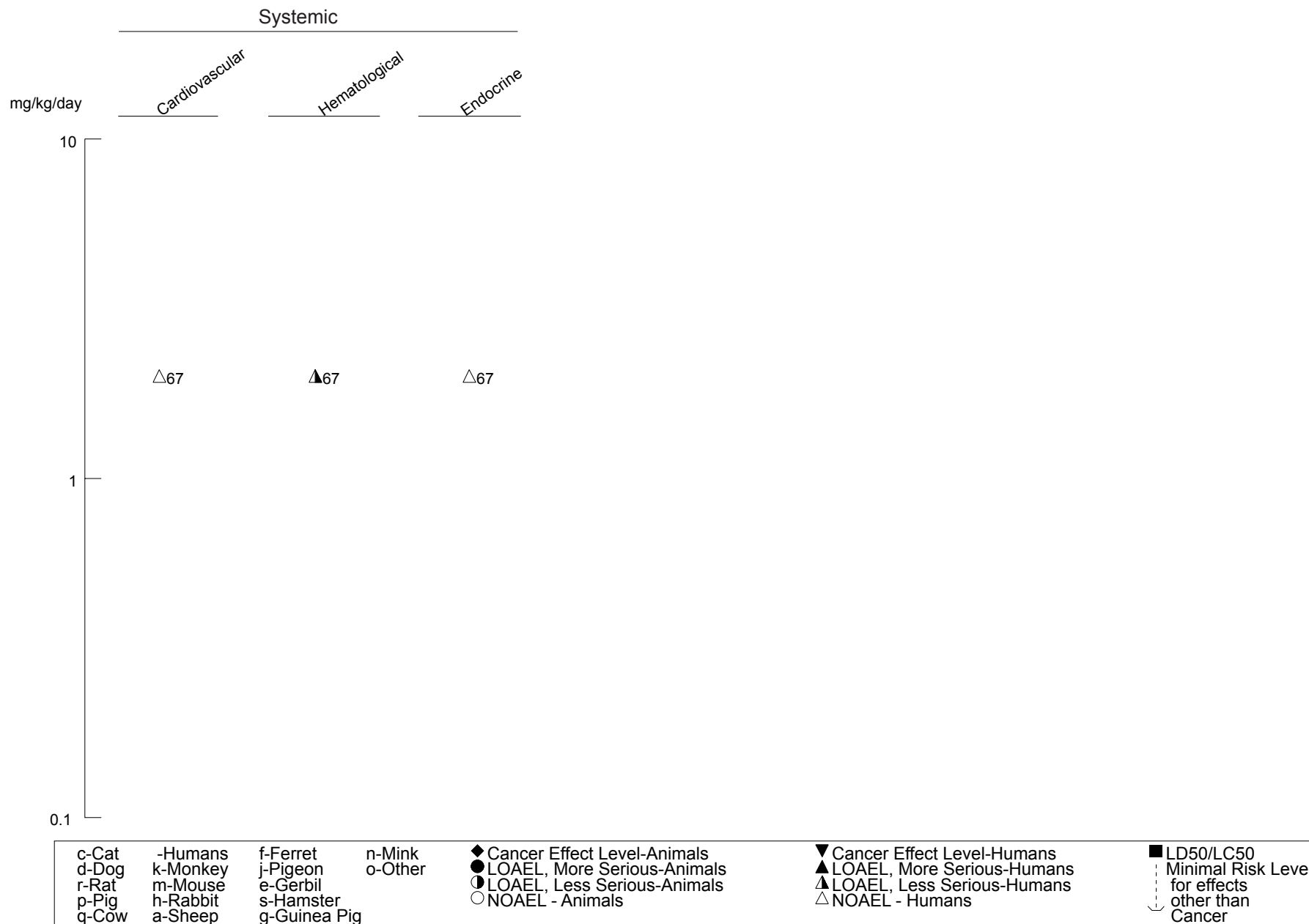


Figure 3-2. Levels of Significant Exposure to Zinc - Oral (*Continued*)

Chronic ( $\geq 365$  days)



### 3. HEALTH EFFECTS

effect on the frequency of cardiovascular disease (heart attack, heart failure, hypertension, or angina) in elderly subjects (>67 years of age) taking up to 2 mg zinc/kg/day for a mean of 8 years (Hale et al. 1988).

In one study, patients having inoperable severe occlusive vascular disease were administered 3.8 mg zinc/kg/day as zinc sulfate for at least 1 year (Henzel et al. 1971). Eighteen of the 24 patients experienced improvement in lower extremity blood flow and unchanged or decreased arterial pressure. Zinc's role in these improvements was not completely understood by the study authors. They hypothesized that when optimal zinc levels are provided to the ischemic limb, the activity of certain zinc enzymes promotes the reversal of tissue-dependent hypoxia and/or lactic acidemia in the muscles. It is also not known if this high dose of zinc was associated with any toxic effects.

No studies were located regarding cardiovascular effects in animals after oral exposure to zinc.

**Gastrointestinal Effects.** Several studies have suggested that zinc ingestion may cause symptoms of gastrointestinal distress or alterations in gastrointestinal tissues. For example, one individual who ingested about 3 ounces of a zinc chloride solution described acute symptoms that occurred almost immediately following contact with the compound, including burning and pain in the mouth and throat and vomiting (Chobanian 1981). Later, the patient exhibited pharyngitis, esophagitis, hypocalcemia, and elevated levels of amylase; the latter two alterations are suggestive of acute pancreatitis. The patient received intravenous hydration and calcium supplementation and recovered within 5 days. The material ingested was described as a "zinc chloride solution," and its concentration was not reported. Therefore, a dose level could not be established in this case.

Several cases of gastrointestinal disturbances have been reported after ingestion of zinc sulfate (Anonymous 1983; Brown et al. 1964; Moore 1978; Samman and Roberts 1987). Vomiting, abdominal cramps, and diarrhea, in several cases with blood, have been observed after ingestion of zinc sulfate. In one report, an English school girl ingested 440 mg zinc sulfate/day (2.6 mg zinc/kg/day) in capsules as a medically prescribed treatment for acne (Moore 1978). After taking each capsule, she experienced epigastric discomfort. A week later, she was admitted to the hospital after a fainting spell. She was diagnosed as anemic and subsequently passed melanic stools, indicative of gastrointestinal bleeding. Gastrointestinal upset (abdominal cramps, vomiting, nausea) occurred in 26 of 47 healthy volunteers following ingestion of zinc sulfate tablets (150 mg as zinc ion in three divided doses per day, 2 mg zinc/kg/day) for 6 weeks (Samman and Roberts 1987). A 17-year-old boy who ingested approximately 6.8 mg zinc/kg as zinc gluconate showed severe nausea and vomiting, but displayed no other symptoms,

## 3. HEALTH EFFECTS

and recovered within 7 hours of ingestion (Lewis and Kokan 1998). Ingestion of zinc oxide has also been associated with gastrointestinal distress (Anonymous 1983; Callender and Gentzkow 1937). In one case, 80% of the personnel of two army companies became ill with gastrointestinal distress and diarrhea after consuming limeade prepared in galvanized trash cans (Callender and Gentzkow 1937). The average dose was estimated to be 6.7–7.1 mg/kg. A second example was presented in a case involving school children in New Mexico who experienced nausea and vomiting after accidental excessive zinc intake (Anonymous 1983). These children had consumed punch containing high levels of zinc dissolved from galvanized hinges attached to tanks in which the punch was stored. A 16-year-old boy who ingested 12 g elemental zinc over a 2-day period (86 mg zinc/kg/day) experienced light-headedness, lethargy, staggering gait, and difficulty writing legibly, but no apparent gastrointestinal disturbances (Murphy 1970).

Gastrointestinal effects have also been observed in animals. Intestinal hemorrhages were observed in ferrets that ingested 390 mg zinc/kg/day as zinc oxide for 2 weeks (Straube et al. 1980). These ferrets exhibited a 75% reduction in food intake. No intestinal hemorrhaging was observed in ferrets fed 195 mg/kg/day for up to 21 days. Oral zinc sulfate exposures of intermediate duration in other experimental animals have also resulted in gastrointestinal effects. Mice fed a diet providing 1,110 mg zinc/kg/day for 13 weeks developed ulcers in the forestomach, but gastrointestinal effects were not observed in rats fed 565 mg zinc/kg/day for 13 weeks (Maita et al. 1981).

**Hematological Effects.** In a case report, acute exposure to 2.6 mg zinc/kg/day as zinc sulfate for 1 week resulted in anemia (Moore 1978). The authors of the report noted that the anemia may have been secondary to the gastrointestinal hemorrhages.

Treatment-related changes in hematological parameters have been observed in humans and animals after intermediate or chronic exposure to zinc or zinc-containing compounds. Long-term administration (1–8 years) of zinc supplements has caused anemia in humans (Broun et al. 1990; Gyorffy and Chan 1992; Hale et al. 1988; Hoffman et al. 1988; Patterson et al. 1985; Porter et al. 1977; Prasad et al. 1978; Ramadurai et al. 1993; Salzman et al. 2002; Stroud 1991; Summerfield et al. 1992). Exposure of one patient to 2 mg zinc/kg/day as zinc sulfate for 10 months resulted in anemia (Hoffman et al. 1988). A significant reduction in erythrocyte superoxide dismutase activity (47% decrease), hematocrit, and serum ferritin, compared to pretreatment levels, occurred in female subjects who received supplements (as capsules) of 50 mg zinc/day as zinc gluconate for 10 weeks (Yadrick et al. 1989); this study was selected as the principal study for derivation of the intermediate-duration MRL. A 15% decrease in erythrocyte superoxide dismutase activity was reported in male volunteers receiving 50 mg zinc/day as zinc gluconate

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for 6 weeks (Fischer et al. 1984). Healthy men given 200 mg zinc/day as elemental zinc for 6 weeks showed a reduction in lymphocyte stimulation response to phytohemagglutinin as well as chemotaxis and phagocytosis of bacteria by polymorphonuclear leukocytes (Chandra et al. 1984); however, no changes in lymphocyte cell number or in the proportion of lymphocyte populations were noted.

In animals, following oral administration of zinc compounds, decreased hemoglobin, hematocrit, erythrocyte, and/or leukocyte levels were observed in rats (Maita et al. 1981; Smith and Larson 1946), mice (Maita et al. 1981; Walters and Roe 1965), rabbits (Bentley and Grubb 1991), dogs (Drinker et al. 1927d; Meurs et al. 1991; Robinson et al. 1991), ferrets (Straube et al. 1980), and preruminant calves (Jenkins and Hidioglou 1991). In rats, the lowest LOAEL for decreased hemoglobin (85% of control value) and erythrocytes (90% of control values) is 12 mg zinc/kg/day as zinc chloride in a 4-week drinking water study with 2-month-old rats (Zaporowska and Wasilewski 1992). The highest NOAEL in rats is 191 mg zinc/kg/day as zinc acetate in a 3-month drinking water study (age of rats not specified) (Llobet et al. 1988a). The reason that the lowest LOAEL is less than the highest NOAEL in rats is unclear, but it may be because of the use of different zinc compounds or different rat strains or age. The second lowest rat LOAEL is 350 mg zinc/kg/day as zinc carbonate (Smith and Larson 1946). For mice, NOAEL and LOAEL values of 104 and 1,110 mg zinc/kg/day as zinc sulfate, respectively, were identified by Maita et al. (1981) in a 13-week feeding study. A serious LOAEL of 68 mg zinc/kg/day as zinc oleate was identified for severe anemia in a 9-month feeding study in mice (Walters and Roe 1965). It is not known if the difference in the LOAELs identified in the Maita et al. (1981) and Walters and Roe (1965) studies is due to the use of different zinc compounds, different basic diet formulations, different mouse strains, or different exposure durations. Slight decreases in hemoglobin levels were observed in rabbits fed 174 mg zinc/kg/day as zinc carbonate (Bentley and Grubb 1991). Zinc oxide consumption caused anemia in dogs (76.5 mg zinc/kg/day) (Drinker et al. 1927d), ferrets (195 mg zinc/kg/day) (Straube et al. 1980), and preruminant calves (64 mg zinc/kg/day) (Jenkins and Hidioglou 1991). Hematological alterations were not observed in cats exposed to up to 83.2 mg zinc/kg/day as zinc oxide (Drinker et al. 1927d) or in adult mink exposed to zinc at up to 297.4 mg zinc/kg/day as zinc oxide (Aulerich et al. 1991; Bleavins et al. 1983) or to rats exposed to 53 mg zinc/kg/day as zinc sulfate (Maita et al. 1981). However, decreases in hematocrit and lymphocytes were observed in the offspring of mink females that ingested a time-weighted-average dose of 20.8 mg zinc/kg/day as zinc sulfate for 10 weeks prior to conception and throughout gestation and lactation (Bleavins et al. 1983) indicating that very young mink may be more sensitive to the hematologic effects of zinc than adults. An increased number of weanling rats had low levels of ceruloplasmin, a copper serum protein, after administration of zinc sulfate for 6 weeks (L'Abbe and Fischer 1984a).



## 3. HEALTH EFFECTS

**Musculoskeletal Effects.** No studies were located regarding musculoskeletal effects in humans after oral exposure to zinc.

Rib biopsies revealed no treatment-related effects in dogs given 4 mg zinc/kg/day as zinc oxide in the diet for 9 months (Anderson and Danylchuk 1979). No lesions of the bones were observed in rats exposed to 565 mg zinc/kg/day as zinc sulfate during 13 weeks of exposure in the food (Maita et al. 1981).

**Hepatic Effects.** Ingestion of 3.5 mg/kg/day zinc sulfate for 18 weeks by 13 patients being treated for chronic venous leg ulcers was reported to have no effect on the results of liver function tests (Hallbook and Lanner 1972). However, the type of liver function tests was not specified and results were not presented to support this conclusion.

Several reports described changes in the serum lipid profile of humans exposed to zinc sulfate or gluconate for 3–12 months; however, the results are mixed. Ingestion of 2.3–4.3 mg zinc/kg/day for 5–6 weeks (Chandra 1984; Hooper et al. 1980) or 0.71 mg zinc/kg/day for 12 weeks (Black et al. 1988) reduced levels of high-density lipoprotein (HDL) cholesterol. In the study by Chandra (1984), a slight increase in low-density lipoprotein (LDL) cholesterol was observed in subjects who served as their own controls; measurements were taken prior to zinc supplementation and after a 10-week postexposure period. Serum cholesterol, triglyceride, and LDL cholesterol levels were not affected by zinc supplementation in the study by Black et al. (1988). However, in another study, zinc supplements depressed HDL cholesterol levels and raised LDL cholesterol levels in elderly subjects (>60 years of age), especially in those who exercised. This study was not well controlled, and the wide variation in doses of the supplemented group prevented the determination of a LOAEL (Goodwin et al. 1985). Young women with a total daily intake of 1.6 mg zinc/kg/day in a 2-month study had a transient decrease in HDL cholesterol (Freeland-Graves et al. 1980). In a double-blind crossover study of young men and women receiving 2.0 (men) or 2.4 (women) mg zinc/kg/day for 6 weeks, total HDL cholesterol was not affected, and LDL cholesterol was significantly decreased in the women (Samman and Roberts 1988). No effect on HDL cholesterol was seen in elderly men and women (60–89 years old) with a total daily intake (dietary zinc plus a zinc acetate supplement) of 1.5 mg/kg/day for 3 months (Bogden et al. 1988), but the subjects also received copper supplements (about 0.03 mg/kg). Another study (Hale et al. 1988) reported no differences in triglycerides and cholesterol levels in subjects ( $\geq 68$  years old) given zinc supplements of up to 2 mg/kg/day for an average of 8 years.

## 3. HEALTH EFFECTS

No histopathology or changes in serum enzyme levels (serum glutamic oxaloacetic transaminase, serum glutamic pyruvic transaminase, or alkaline phosphatase) were observed in rats receiving 191 mg zinc/kg/day as zinc acetate (Llobet et al. 1988a). Similarly, no histopathology was observed in rats administered 98.3 mg zinc/kg/day as zinc oxide, but an insufficient number of animals were tested (Drinker et al. 1927c). Sheep fed time-weighted-average doses of 19 mg zinc/kg/day as zinc oxide for 49–72 days developed hepatic effects, including necrotic hepatocytes and large quantities of hemosiderin in Kupffer cells (Allen et al. 1983). Because sheep are ruminants, it is not known if they are a good model for predicting human toxicity. No histological damage was observed in adult or young mink fed 327 or 324 mg zinc/kg/day, respectively, as zinc sulfate for 144 days (Aulerich et al. 1991).

Decreased hexobarbital sleeping times were reported by Kadiiska et al. (1985) in rats receiving 40 mg zinc/kg/day as zinc sulfate. This physiological response suggested an induction of microsomal enzymes.

Increases in serum cholesterol levels were observed in two studies where rats were fed either 2.8 or 10 mg zinc/kg/day as zinc acetate for 2–7 months (Katya-Katya et al. 1984; Klevay and Hyg 1973). Other studies have shown no effect on total cholesterol, HDL cholesterol, or serum triglyceride levels in rats ingesting 3 or 25 mg zinc/kg/day of unspecified zinc compounds (Fischer et al. 1980; Woo et al. 1983).

**Renal Effects.** Thirteen patients treated with zinc sulfate at 3.5 mg zinc/kg/day for 18 weeks for chronic venous leg ulcers had normal urinalyses (Hallbook and Lanner 1972). However, neither the specific parameters measured for the urinalysis nor the results were presented to support this conclusion. Furthermore, urinalysis may not be a sensitive indicator of renal function.

A number of intermediate-duration studies have demonstrated renal effects in animals exposed to zinc oxide, zinc sulfate, and zinc acetate. Zinc sulfate caused an increase in the absolute and relative kidney weights and regressive kidney lesions (not specified) in female mice that consumed 1,110 mg zinc/kg/day in the diet for 13 weeks, but no effects occurred in rats that consumed 565 mg zinc/kg/day or in mice that consumed 104 mg zinc/kg/day under similar conditions (Maita et al. 1981). Severe diffuse nephrosis was observed in ferrets exposed to 195 mg zinc/kg/day as zinc oxide in the diet (Straube et al. 1980). In rats exposed to 191 mg zinc/kg/day as zinc acetate for 3 months, epithelial cell damage in the glomerulus and proximal convoluted tubules and increased plasma creatinine and urea levels were observed (Llobet et al. 1988a). The NOAEL for the effects on creatinine and urea was 95 mg zinc/kg/day. It is unclear whether

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the microscopic changes were observed at lower doses. No histopathological changes in the kidneys were observed in three rats that drank water containing 98.3 mg zinc/kg/day as zinc oxide for 35–36 weeks (Drinker et al. 1927c); however, interpretation of the results of this study is severely limited by the small number of rats used. Renal tubular dilation, with proteinaceous casts and hemosiderin deposits, was observed in the kidneys of sheep that ingested 18 mg zinc/kg/day as zinc oxide for 49–72 days (Allen et al. 1983). It is not known if sheep are a good model for human toxicity because they are ruminants. No renal effects were observed in either adult mink consuming 326.7 mg zinc/kg/day as zinc sulfate or in young mink consuming 323.6 mg zinc/kg/day as zinc sulfate for 144 days (Aulerich et al. 1991). Minks exposed to 195 mg zinc/kg/day as zinc oxide for 7–97 days in the food developed a diffuse nephrosis, though it did not increase with increasing dose (Straube et al. 1980).

**Dermal Effects.** No studies were located regarding dermal/ocular effects in humans after oral exposure to zinc.

No dermal effects were seen in adult female minks given a time-weighted dose of 20.8 mg zinc/kg/day as zinc sulfate for 10 weeks prior to mating and then throughout gestation and lactation (Bleavins et al. 1983). However, the offspring of these animals showed graying of the fur around the eyes, ears, jaws, and genitals with a concomitant loss of hair and dermatosis in these areas during the weaning period. These conditions were reversible upon removal of treatment.

**Body Weight Effects.** No effects on body weight have been reported in humans following oral exposure to zinc. However, a 46% decrease in body weight gain was seen in preruminant calves that consumed 91 mg zinc/kg/day as zinc oxide for 5 weeks; there was no effect at 64 mg zinc/kg/day (Jenkins and Hidioglou 1991). The relevance of this effect to humans is unclear. Body weights of rabbits (Bentley and Grubb 1991), rats (Llobet et al. 1988a), and minks (Aulerich et al. 1991) were unaffected by dosing with 174, 191, and 326.7 mg zinc/kg/day, respectively, for 3–12 months. Decreased postpartum body weights in F0 animals were observed in rats exposed to 7 mg zinc/kg/day as zinc chloride for 20 weeks (Khan et al. 2001b).

#### 3.2.2.3 Immunological and Lymphoreticular Effects

Zinc plays a role in the normal development and maintenance of the immune system, such as in the lymphocyte response to mitogens and as a cofactor for the thymic hormone thymulin (Delafuente 1991; Fraker et al. 1986). Oral exposure to zinc at levels much higher than the RDA has impaired immune and

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inflammatory responses. This was observed in *in vivo* investigations of the immune competence of blood components taken from 11 healthy adult men after ingestion of 4.3 mg zinc/kg/day as zinc sulfate for 6 weeks. The mitogenic response elicited from peripheral blood lymphocytes and the chemotactic and phagocytic responses of polymorphonuclear leukocytes were impaired after zinc ingestion. No effects were seen on total numbers of lymphocytes or relative numbers of T cells, T cell subsets, or B cells (Chandra 1984). The relationship between these observations and decreased levels of immune competence that might lead to increased susceptibility to disease is unknown. Zinc supplements administered to elderly populations at doses up to 1.5 mg zinc/kg/day (Bogden et al. 1988) or 2.5 mg zinc/kg/day (Duchateau et al. 1981) resulted in either no effect or a beneficial effect on immune cell titers or delayed cutaneous hypersensitivity responses to specific antigens.

Decreased lymphocyte activity (incorporation of  $^3\text{H}$ -thymidine in response to concanavalin A) was reported in mink kits from dams who had ingested a time-weighted-average dose of 20.8 mg zinc/kg/day as zinc sulfate for 10 weeks prior to conception and throughout gestation and lactation (Bleavins et al. 1983). The dose to the kits is unknown. In contrast, no effect was observed on antibody titre (immunoglobulin G [IgG] and immunoglobulin M [IgM]) or the mitogenic response of splenic B cells isolated from mice fed 76.9 mg zinc/kg/day as zinc sulfate for 4 weeks and challenged with B cell antigens either *in vivo* or *in vitro* (Schiffer et al. 1991). The *in vitro* mitogenic response of T cells isolated from these mice was increased. There was no effect of the zinc supplement in the plaque forming cell assay or on cytotoxic T killer cell activity in mice exposed to 6.5 mg zinc/kg/day in the diet for 8 weeks (Fernandes et al. 1979). In mice exposed *in utero* to 136 mg zinc/kg/day, with exposure continuing postnatally, there were increases in direct plaque-forming activity of spleen cells and in lymphocyte proliferation in response to mitogen stimulation (Lastra et al. 1997). The highest NOAEL value in animals and the LOAEL value in humans for immunological effects after intermediate-duration oral exposure are recorded in Table 3-2 and plotted in Figure 3-2.

#### 3.2.2.4 Neurological Effects

Zinc appears to be necessary for normal brain function (Sandstead et al. 1983), but excess zinc is toxic. A 16-year-old boy who ingested  $\approx 86$  mg zinc/kg/day of metallic zinc over a 2-day period in an attempt to promote wound healing, developed signs and symptoms of lethargy, light-headedness, staggering, and difficulty in writing clearly (Murphy 1970). Lethargy was also observed in a 2-year-old child who ingested a zinc chloride solution ( $\approx 1,000$  mg zinc/kg) (Potter 1981). It is not known whether these observations represent direct effects on the nervous system.

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Very limited data were located regarding neurological effects in animals. Minor neuron degeneration and proliferation of oligodendroglia occurred in rats dosed with 487 mg zinc/kg/day as zinc oxide for 10 days (Kozik et al. 1980). Rats receiving 472 mg zinc/kg/day for 10 days had increased levels of secretory material in the neurosecretory nuclei of the hypothalamus (Kozik et al. 1981). Mice exposed postnatally to 0.5 mg zinc/kg/day as zinc acetate for 28 days showed no changes in memory formation, but showed a gradual decrease in learning extinction throughout the study (de Oliveira et al. 2001).

**3.2.2.5 Reproductive Effects**

Pregnant women receiving capsules containing 0.3 mg zinc/kg/day as zinc sulfate during the last two trimesters did not exhibit any reproductive effects (no changes in maternal body weight gain, blood pressure, postpartum hemorrhage, or infection) (Mahomed et al. 1989). No other studies were located regarding reproductive effects in humans after oral exposure to zinc.

No measurable effect on gestational length or litter size was observed when female mink ingested a time-weighted average dose of 20.8 mg zinc/kg/day as zinc sulfate (Bleavins et al. 1983). No histological alterations in the testes or ovaries were noted in mice fed zinc sulfate (1,110 mg zinc/kg/day) for 13 weeks (Maita et al. 1981). Male and female rats exposed by gavage to up to 14 mg zinc/kg/day as zinc chloride resulted in a nonsignificant decrease in fertility index in all groups that was not related to administered dose; in the two highest groups (7 and 14 mg zinc/kg/day), decreases in live pups per litter and pup weight at day 21 were also reported (Khan et al. 2001b). Male and female rats receiving 50 mg zinc/kg/day as zinc carbonate in the diet were reported to reproduce normally for several generations in a poorly documented study by Sutton and Nelson (1937). Rats fed 250 mg zinc/kg/day for 14–17 weeks mated successfully but had a higher than normal percentage of stillborn pups. A subsequent mating of the parental generation fed 250 mg zinc/kg/day for 5 months was unsuccessful. No reproduction occurred in rats fed 500 mg zinc/kg/day for 5 months (Sutton and Nelson 1937). The frequency of sperm with an altered chromatin structure was increased in rats fed 25 mg zinc/kg/day as zinc chloride for 8 weeks (Evenson et al. 1993). Pre-implantation loss increased in rats fed diets containing 200 mg zinc/kg/day as zinc sulfate on gestational days 0–18 (Pal and Pal 1987). When the rats received 200 mg zinc/kg/day 21 days prior to mating, no effects on implantation or other adverse reproductive effects were observed (Pal and Pal 1987). Similarly, exposure of up to 372 mg zinc/kg/day in mice prior to and throughout pregnancy did not result in changes in reproductive index (Lastra et al. 1997). The highest NOAEL

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values and all LOAEL values from each reliable study for reproductive effects in each species and duration category are recorded in Table 3-2 and plotted in Figure 3-2.

#### 3.2.2.6 Developmental Effects

Zinc is necessary for normal fetal growth and development. Fetal damage may result from zinc deficiency. Only one report in the literature suggested adverse developmental effects in humans due to exposure to excessive levels of zinc (Kumar 1976). Four women were given zinc supplements of 0.6 mg zinc/kg/day as zinc sulfate during the third trimester of pregnancy. Three of the women had premature deliveries, and one delivered a stillborn infant. However, the significance of these results cannot be determined because very few details were given regarding the study protocol, reproductive histories, and the nutritional status of the women. Other human studies have found no developmental effects in the newborns of mothers consuming 0.3 mg zinc/kg/day as zinc sulfate (Mahomed et al. 1989) or zinc citrate (Simmer et al. 1991) or 0.06 mg zinc/kg/day as zinc aspartate (Kynast and Saling 1986) during the last two trimesters. There has been a suggestion that increased serum zinc levels in pregnant women may be associated with an increase in neural tube defects (McMichael et al. 1994), but others have failed to confirm this association (Hambidge et al. 1993).

The developmental toxicity of zinc in experimental animals has been evaluated in a number of investigations. Exposure to high levels of zinc in the diet prior to and/or during gestation has been associated with increased fetal resorptions, reduced fetal weights, altered tissue concentrations of fetal iron and copper, and reduced growth in the offspring.

Administration of zinc in rats at 200 mg zinc/kg/day as zinc oxide in the diet for 21 days prior to mating and then throughout gestation resulted in resorption of all fetuses (Schlicker and Cox 1968). Fetal resorptions ranged from 4 to 29% when 200 mg zinc/kg/day was administered only during gestation (controls had no resorptions). When the dose was reduced to 100 mg zinc/kg/day starting 21 days prior to mating, there were no fetal resorptions, malformations, or growth reduction. In contrast, Kinnamon (1963) reported no resorptions, no difference in the number of offspring per litter, and no change in average wet weight of the fetuses from female rats fed 250 mg zinc/kg/day as zinc carbonate in the diet for 53 days before mating and during gestation. The reason for the differences in the results of these studies is unknown. No effect on fetal viability, size, or malformations was seen in fetuses from female rats fed 25 mg zinc/kg/day as zinc carbonate during gestational days 1–18 (Uriu-Hare et al. 1989).

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Administration of 200 mg zinc/kg/day to dams throughout gestation resulted in decreased growth and tissue levels of copper and iron in fetal rats (Cox et al. 1969; Schlicker and Cox 1968). In rats, at both 100 and 200 mg/kg/day during gestational days 1–18, maternal zinc levels increased. However, zinc tissue levels in the 22-day-old fetuses were not elevated at 100 mg/kg/day to dams, suggesting that the placenta was able to act as a barrier to zinc at the lower dietary level. In contrast, Ketcheson et al. (1969) showed that newborn and 14-day-old rats from mothers that had consumed 100 mg/kg/day throughout gestation had elevated levels of total zinc and decreased levels of iron. It is unclear whether the longer exposure to zinc during gestation or the suckling of newborn rats prior to sacrifice may have accounted for these differences.

Animal studies suggest that exposure to very high levels of dietary zinc is associated with reduced fetal weight, alopecia, decreased hematocrit, and copper deficiency in offspring. For example, second generation mice exposed to zinc carbonate during gestation and lactation (260 mg/kg/day in the maternal diet), and then continued on that diet for 8 weeks, had reduced body weight, alopecia, and signs of copper deficiency (e.g., lowered hematocrit and occasional achromotrichia [loss of hair color]) (Mulhern et al. 1986). Similarly, mink kits from dams that ingested a time-weighted-average dose of 20.8 mg zinc/kg/day as zinc sulfate also had alopecia and achromotrichia (Bleavins et al. 1983). It is likely that the alopecia resulted from zinc-induced copper deficiency, which is known to cause alopecia in monkeys (Obeck 1978). However, no adverse effects were observed in parental mice or mink. No effects on reproduction were reported in rats exposed to 50 mg zinc/kg/day as zinc carbonate; however, increased stillbirths were observed in rats exposed to 250 mg zinc/kg/day (Sutton and Nelson 1937).

The highest NOAEL values and all LOAEL values from each reliable study for developmental effects in each species and duration category are recorded in Table 3-2 and plotted in Figure 3-2.

#### 3.2.2.7 Cancer

Only limited data were found regarding relationships between the ingestion of zinc and its compounds by humans and the subsequent development of cancer. One study reported an association between an excess rate of gastric cancer in the people of North Wales (Great Britain) and the high zinc-to-copper ratio ( $\approx 30:1$ ) in the soil of household gardens (Stocks and Davies 1964). However, the inference that this excess in gastric cancer is causally associated with soil levels of zinc and copper is not consistent with another study. In a survey of cancer registry data (1954–1978) in Shipham, Somerset (Great Britain), an area that also has a high soil zinc-to-copper ratio ( $\approx 17:1$ ), the gastric cancer incidence rate was

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significantly lower than the regional rate (Philipp et al. 1982). It is probable that other factors, not considered by Stocks and Davies (1964), are associated with or coincidental to the high soil zinc-to-copper ratio confounded the results.

The carcinogenicity of zinc in experimental animals following oral exposure was evaluated by Walters and Roe (1965). The incidence of tumors was not increased in mice exposed to 951 mg zinc/kg/day as zinc sulfate in drinking water for 1 year compared to controls. However, important details regarding the study protocol were lacking including the age and sex of the mice, the number of mice at the beginning of the study, the purity of the test material, and a complete list of the organs and tissues examined at necropsy. The control mice developed intercurrent disease (ectromelia), which resulted in a number of deaths; supplementary control mice were added to the study, but they were not concurrent controls. The number of animals in treated and control groups surviving at 1 year (study termination) was small (22–28 mice/group). The exposure period (1 year) was less than the standard bioassay period (18–24 months). There were no data in the study (e.g., survival or body weight data) to indicate that a maximum tolerated dose was achieved. These limitations reduce the sensitivity of the study by Walters and Roe (1965) to detect a carcinogenic response.

Halme (1961) exposed tumor-resistant and tumor-susceptible strains of mice to zinc in drinking water. In a 5-generation study, groups of tumor-resistant mice (strain not specified) received 0, 10, 20, 50, 100, or 200 mg Zn/L as zinc chloride in the drinking water. The spontaneous tumor frequency for this strain of mice was 0.0004%. The tumor frequencies were reported as: F0=0.8%; F1=3.5%; F1 and F2=7.6%; and F3 and F4=25.7%. The majority of the tumors were seen in the 10- and 20-mg Zn dose groups. No individual or group tumor incidence data were reported, and a discussion of statistical analysis was not included. In the tumor-susceptible mice, strains C3H and A/Sn received 10–29 mg Zn/L in their drinking water for 2 years; 33/76 tumors were observed in the C3H strain (31 in females) and 24/74 tumors were observed in the A/Sn strain (20 in females). Most of the tumors were reported to be adenocarcinomas, but the tissues in which they occurred were not reported. The numbers of specific tumor types were not reported. The overall tumor frequencies (43.4% for C3H and 32.4% for A/Sn) were higher than the spontaneous frequency (15% for each strain), but statistical analyses were not reported.



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**3.2.3 Dermal Exposure****3.2.3.1 Death**

No studies were located regarding death in humans or animals after dermal exposure to zinc.

**3.2.3.2 Systemic Effects**

Zinc has been reported to promote the healing of burns and wounds when topically applied as zinc oxide or calamine lotion (Gordon et al. 1981). The mechanism by which this occurs was not discussed by the authors. Zinc oxide contained in an occlusive zinc tape dressing reduced the inflammatory reactions in the granulation tissue of wounded rats (Wetter et al. 1986). The authors speculated that zinc acted either by a continuous release of zinc ions or by modifying components involved in the tape's adhesive properties.

No studies were located regarding respiratory, cardiovascular, gastrointestinal, musculoskeletal, hepatic, renal, or other systemic effects in humans or animals after dermal exposure to zinc. The systemic effects observed after dermal exposure are discussed below. The NOAEL values and all LOAEL values from each reliable study for dermal effects in each species and duration category are recorded in Table 3-3.

**Hematological Effects.** A worker who had been employed making up zinc chloride solutions (concentrations not specified) with his hands was found to have microcytic anemia and decreased numbers of platelets (DuBray 1937).

No studies were located regarding hematological effects in animals after dermal exposure to zinc.

**Dermal Effects.** No signs of dermal irritation were observed in humans after a 25% zinc oxide patch (2.9 mg/cm<sup>2</sup>) was placed on the skin for 48 hours (Agren 1990). However, 14 out of 17 men who were employed in the bagging or packing of zinc oxide and whose skin was frequently covered with zinc oxide dust reported having experienced zinc oxide pox at least once (Turner 1921). The pox appeared as itchy papular-pustular eruptions in the pubic region, scrotum, inner surface of the thigh, and occasionally on the axilla and inner surface of the arms. The study author suggested that these lesions were due to clogging of glands by dust, perspiration, and bacteria when skin surfaces coated with these substances were rubbed together. In contrast, a case study of 24 workers exposed to dusts of either zinc oxide, zinc sulfide, or

Table 3-3 Levels of Significant Exposure to Zinc - Dermal

Species (Strain)	Exposure/ Duration/ Frequency (Specific Route)	System	NOAEL	LOAEL		Reference Chemical Form
				Less Serious	Serious	
<b>ACUTE EXPOSURE</b>						
<b>Systemic</b>						
Human	48 hr	Dermal	2.9 mg/cm <sup>2</sup>			Agren 1990 Zinc Oxide
Mouse	5 d	Dermal		0.48 M mg/cm <sup>2</sup>	(severe skin irritation)	Lansdown 1991 Zinc chloride
Mouse	5 d	Dermal		0.4 M mg/cm <sup>2</sup>	(Slight skin irritancy)	Lansdown 1991 Zinc sulfate
Mouse	5 d	Dermal	16 M mg/cm <sup>2</sup>			Lansdown 1991 Zinc Oxide
Mouse	5 d	Dermal		7.2 M mg/cm <sup>2</sup>	(moderate skin irritation)	Lansdown 1991 Zinc Acetate
Gn Pig	5 d	Dermal	0.4 M mg/cm <sup>2</sup>			Lansdown 1991 Zinc sulfate
Gn Pig	5 d	Dermal		0.48 M mg/cm <sup>2</sup>	(moderate skin irritation)	Lansdown 1991 Zinc chloride
Gn Pig	5 d	Dermal	16 M mg/cm <sup>2</sup>			Lansdown 1991 Zinc Oxide

Table 3-3 Levels of Significant Exposure to Zinc - Dermal

(continued)

Species (Strain)	Exposure/ Duration/ Frequency (Specific Route)	System	NOAEL	LOAEL		Reference Chemical Form
				Less Serious	Serious	
Gn Pig	5 d	Dermal	7.2 M mg/cm <sup>2</sup>			Lansdown 1991 Zinc Acetate
Rabbit	5 d	Dermal	0.4 M mg/cm <sup>2</sup>			Lansdown 1991 Zinc sulfate
Rabbit	5 d	Dermal	16 M mg/cm <sup>2</sup>			Lansdown 1991 Zinc Oxide
Rabbit	5 d	Dermal		7.2 M mg/cm <sup>2</sup>	(slight skin irritation - open patch test; severe skin irritation - occluded patch test)	Lansdown 1991 Zinc Acetate
Rabbit	5 d	Dermal		0.48 M mg/cm <sup>2</sup>	(severe skin irritation)	Lansdown 1991 Zinc chloride

d = day(s); Derm = dermal; Gn pig = Guinea pig; hr = hour(s); LOAEL = lowest-observed-adverse-effect level; NOAEL = no-observed-adverse-effect level

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metallic zinc revealed only 1 worker with papular pustular lesions on the axilla and inner thighs (Batchelor et al. 1926). The difference in the results was attributed to differences in the personal hygiene of the workers in the two studies.

The dermal irritancy of several zinc compounds was compared in mice, rabbits, and guinea pigs (Lansdown 1991). Of the six zinc compounds tested, zinc chloride had the greatest irritancy potential (severe irritation at 0.48 mg/cm<sup>2</sup>), followed by zinc acetate (moderate irritation at 7.2 mg/cm<sup>2</sup>) and zinc sulfate (slight irritation at 0.48 mg/cm<sup>2</sup>); no signs of irritation were observed following exposure to zinc oxide. Although zinc chloride is clearly the most irritating, the relative irritancy of zinc sulfate and zinc acetate was not determined because only one dose was tested and a different dose was used for each compound. The severe skin irritancy observed following application of zinc chloride was characterized by parakeratosis, hyperkeratosis, inflammatory changes in the epidermis and superficial dermis, and acanthosis of the follicular epithelia (Lansdown 1991).

**Ocular Effects.** In a case report, accidental splashing of a soldering paste containing 30% zinc chloride into the eye of a plumber produced an immediate reduction in visual acuity, hyperemia, hemorrhaging, conjunctival swelling, corneal opacity, bullous keratopathy, and spotting of the lens (Houle and Grant 1973). Most symptoms disappeared after 6 weeks, but residual lens opacities persisted for over a year after the exposure. Reddened conjunctivae and lacrimation were observed in 34 persons who were exposed to extremely high concentrations of zinc chloride smoke when several smoke generators exploded in a tunnel during World War II (Evans 1945). Two of the exposed persons had corneal burns and four had small vesicular burns on the forehead or wrist. Zinc chloride was the major component of the smoke. However, other components such as zinc oxide, hexachloroethane, calcium silicide, the igniter, or the heat of the explosion may have contributed to the injuries that were observed.

No studies were located regarding the following health effects in humans or animals after dermal exposure to zinc:

#### **3.2.3.3 Immunological and Lymphoreticular Effects**

#### **3.2.3.4 Neurological Effects**

#### **3.2.3.5 Reproductive Effects**

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**3.2.3.6 Developmental Effects****3.2.3.7 Cancer****3.3 GENOTOXICITY**

Genotoxicity studies conducted in a variety of test systems have failed to provide evidence for mutagenicity of zinc. However, there are indications of weak clastogenic effects following zinc exposure.

Chromosome aberrations were observed in the lymphocytes of 24 workers in a zinc smelting plant (Bauchinger et al. 1976). However, the workers had increased blood levels of lead and cadmium, and the clastogenic effect was attributed to cadmium exposure.

Results of *in vivo* studies are shown in Table 3-4. A dominant lethal study in mice failed to show a mutagenic potential for zinc. However, chromosomal aberrations have been observed in bone marrow cells following *in vivo* exposure to zinc (Vilkina et al. 1978). This effect was observed in rats exposed to 14.8 mg zinc/kg/day as zinc chlorate in drinking water (Kowalska-Wochna et al. 1988), mice given intraperitoneal injections of 3.6 mg zinc/kg/day as zinc chloride (Gupta et al. 1991), and mice exposed to zinc oxide by inhalation (Voroshilin et al. 1978). Chromosomal aberrations caused by zinc were observed in the bone marrow cells of mice maintained on a low-calcium diet (Deknudt and Gerber 1979). Calcium may be displaced by zinc in calcium-depleted conditions, leading to chromosome breaks and/or interfering in the repair process (Deknudt and Gerber 1979). *In vivo* exposure to zinc may also result in single-strand breaks, as measured by the Comet assay in mice (Banu et al. 2001). An increased incidence of sister chromatid exchange was observed in bone marrow cells of rats exposed to 17.5 mg zinc/kg/day as zinc chlorate in drinking water (Kowalska-Wochna et al. 1988).

Results of *in vitro* studies are shown in Table 3-5. Exposure to zinc as zinc sulfate or zinc chloride does not increase mutation frequencies in bacterial or mammalian cell culture test systems (Amacher and Paillet 1980; Gocke et al. 1981; Marzin and Vo Phi 1985; Nishioka 1975; Thompson et al. 1989; Venitt and Levy 1974; Wong 1988). Similarly, there was no convincing evidence of a clastogenic effect in human lymphocytes exposed to 0.0003–0.00003 M zinc chloride (Deknudt and Deminatti 1978).

**Table 3-4. Genotoxicity of Zinc *In Vivo***

Species (test system)	End point	Results	Reference
Mammalian systems:			
Mouse	Dominant lethal	–	Vilkina et al. 1978
Mouse	Single-strand DNA breaks	+	Banu et al. 2001
Mouse bone marrow	Chromosomal	+	Deknudt and Gerber 1979
Mouse	Chromosomal	+	Voroshilin et al. 1978
Rat bone marrow	Chromosomal aberrations	+	Kowalska-Wachna 1988
Mouse bone marrow	Chromosomal aberrations	+	Gupta et al. 1991
Rat bone marrow	Sister chromatid exchange	+	Kowalska-Wachna 1988
Mouse	Micronucleus	–	Gocke et al. 1981
Drosophila	Sex-linked recessive lethal	–	Gocke et al. 1981

– = negative result; + = positive result; DNA = deoxyribonucleic acid

**Table 3-5. Genotoxicity of Zinc *In Vitro***

Species (test system)	End point	Results		Reference
		With activation	Without activation	
Prokaryotic organisms:				
<i>Salmonella typhimurium</i> (TA102)	Gene mutation	Not tested	–	Marzin and Vo Phi 1985
<i>S. typhimurium</i> (TA98, TA102, TA1535, TA1537)	Gene mutation	– (S9)	–	Wong et al. 1988
<i>S. typhimurium</i> (TA1538, TA98, TA100, TA1537)	Gene mutation	– (S9)	–	Thompson et al. 1989
<i>S. typhimurium</i> (TA1535, TA1537, TA1538, TA98, TA100)	Gene mutation	– (S9)	–	Gocke et al. 1981
<i>Escherichia coli</i>	Gene mutation	Not tested	–	Nishioka 1975
<i>E. coli</i>	Gene mutation	Not tested	–	Venitt and Levy 1974
Mammalian cells:				
Mouse lymphoma	Gene mutation	Not tested	–	Amacher and Paillet 1980
Mouse lymphoma	Gene mutation	+ (S9)	+	Thompson et al. 1989
Human lymphocytes	Chromosomal aberrations	Not tested	+	Deknudt and Deminatti 1978
Chinese hamster ovary cells	Chromosomal aberrations	+ (S9)	+	Thompson et al. 1989

– = negative result; + = positive result

### 3.4 TOXICOKINETICS

There is limited information on the toxicokinetic properties of zinc following inhalation or dermal exposure. Increased zinc levels in the blood and urine of humans and in the tissue of animals after inhalation and dermal exposure to zinc, respectively, indicate that zinc is absorbed by these routes. The toxicokinetic properties of ingested zinc have been extensively studied. The absorption of zinc from the gastrointestinal tract is homeostatically regulated; under normal physiological conditions, 20–30% of ingested zinc is absorbed. Zinc uptake from the intestinal lumen involves passive diffusion and a carrier-mediated process. A number of factors influence the absorption of zinc; these include inhibitors, such as calcium, phosphorus, and dietary fiber and phytates (components of dietary fiber that may coprecipitate with zinc in the intestines), and enhancers, such as amino acids, picolinic acid, and prostaglandin E<sub>2</sub>. Once absorbed, zinc is widely distributed throughout the body. Zinc content is highest in muscle, bone, gastrointestinal tract, kidney, brain, skin, lung, heart, and pancreas. In plasma, two-thirds of the zinc is bound to albumin which represents the metabolically active pool of zinc. This pool of plasma zinc is frequently referred to as loosely bound zinc because albumin has the ability to give up bound zinc to tissues. Zinc is excreted in both urine and feces.

#### 3.4.1 Absorption

##### 3.4.1.1 Inhalation Exposure

Quantitative studies regarding absorption of zinc and zinc compounds after inhalation exposure in humans are limited. The absorption of inhaled zinc depends on the particle size and solubility. Elevated levels of zinc have been found in the blood and urine of workers exposed to zinc oxide fumes (Hamdi 1969).

The rates or percentages of absorption of inhaled zinc in animals are not available; however, studies provide data on zinc retention in the lungs. Zinc retention values were 19.8, 11.5, and 4.7% in the lungs of guinea pigs, rats, and rabbits, respectively, after inhalation exposure (nose-only) to 3.5–9.1 mg zinc/m<sup>3</sup> as zinc oxide aerosol for 2–3 hours (Gordon et al. 1992). The aerosol had a mass median diameter of 0.17 µm. The retention of zinc in lungs was dose related in male Wistar rats



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administered a single intratracheal instillation of 0.07–3.7 mg zinc/m<sup>3</sup> as zinc oxide (Hirano et al. 1989). A half-life of 14 hours was calculated.

The absorption of zinc oxide fumes lead to increased levels of zinc measured in the liver, kidney, and pancreas of cats exposed to zinc oxide fumes for durations ranging from 15 minutes to 3.25 hours (Drinker and Drinker 1928). The usefulness of the study is limited because reporting was inadequate and particle size of the zinc oxide aerosol was not determined. Some inhaled particles of zinc oxide are subject to ciliary clearance and swallowing. Thus, a portion of the inhaled zinc may ultimately be absorbed from the gastrointestinal tract.

#### **3.4.1.2 Oral Exposure**

Several studies have measured oral absorption rates of zinc in humans. Absorption ranged from 8% to 81% following short-term exposures to zinc supplements in the diet; differences in absorption are probably due to the type of diet (amount of zinc ingested, amount and kind of food eaten) (Aamodt et al. 1983; Hunt et al. 1991; Istfan et al. 1983; Reinhold et al. 1991; Sandstrom and Abrahamson 1989; Sandstrom and Cederblad 1980; Sandstrom and Sandberg 1992). For example, dietary protein facilitates zinc absorption; fractional zinc absorption ranged from 8% for low-protein rolls to 26% for high-protein rolls 3 days after individuals ingested 0.05 mg zinc/kg (Hunt et al. 1991).

Absorption of labeled zinc was 40.0–48.4% in male Wistar rats fed a diet containing 0.81 mg zinc/kg as zinc chloride or zinc carbonate (Galvez-Morros et al. 1992). Fractional absorption in immature organisms generally exceeds that in adults. In growing rats, on the basis of indirect calculation from isotope experiments, Weigand and Kirchgessner (1992) suggested surprisingly high absorption values of as much as 94.7%. It is likely that all these results were influenced by isotope exchange and do not provide estimates of net absorption.

The body's natural homeostatic mechanisms control zinc absorption from the gastrointestinal tract (Davies 1980). Persons with adequate nutritional levels of zinc absorb approximately 20–30% of all ingested zinc. Those who are zinc-deficient absorb greater proportions of administered zinc (Johnson et al. 1988; Spencer et al. 1985).

Absorption of zinc occurs from all segments of the intestine, although the largest proportion of zinc absorption occurs from the duodenum (Methfessel and Spencer 1973). The zinc absorption process

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includes both passive diffusion and a carrier-mediated process (Tacnet et al. 1990). The intestinal absorption of zinc appears to be a saturable carrier-mediated process at low zinc dose levels involving a cysteine-rich intestinal protein (CRIP) (Davies 1980; Gunshin et al. 1991; Hempe and Cousins 1992; Sturniolo et al. 1991). This protein binds zinc entering the intestinal cells from the lumen (Hempe and Cousins 1991). CRIP has a limited binding capacity for zinc and becomes saturated when zinc concentration in the intestine is high. Metallothionein, a metal-binding protein, may contribute to zinc homeostasis at higher zinc absorption. Like several other metals, zinc can induce metallothionein production in intestinal mucosal cells (Richards and Cousins 1975). Zinc binds to metallothionein, which remains in the mucosal cells lining the gastrointestinal tract, and the bound metal is excreted from the body upon sloughing off of these cells. Although the affinity of zinc for metallothionein is relatively low, the protein may serve to prevent absorption of excess zinc in the body (Foulkes and McMullen 1987). Absorption of zinc in rats is increased when metallothionein levels are lower (Flanagan et al. 1983). It is hypothesized that zinc entering luminal cells is associated with CRIP, and a small amount is bound to metallothionein; however, as the luminal zinc concentration increases, the proportion of cytosolic zinc associated with CRIP is decreased with a concomitant increase in zinc binding to metallothionein (Hempe and Cousins 1992). Further details on the influence of CRIP and metallothionein on zinc absorption are provided in Section 3.5, Mechanisms of Action.

Phytate and high phosphorus intakes in animals decrease zinc absorption. In humans, dairy products that contain both calcium and phosphorus decrease zinc absorption and plasma zinc concentration (Pecoud et al. 1975). Zinc binds to phosphate which results in coprecipitation of zinc with calcium phosphate in the intestines (Nelson et al. 1985). Dietary phytate also reduces zinc absorption. The addition of 400  $\mu\text{mol}$  phytate to the diet decreased zinc absorption from  $43.3 \pm 17.9\%$  in females fed bread containing 0.02 mg zinc/kg (zinc-65 isotope) to  $14.3 \pm 3.2\%$  (Sandstrom and Sandberg 1992). Rats given diets supplemented with radiolabeled zinc and phytate excreted significantly more zinc in the feces than rats given diets supplemented with radiolabeled zinc but without phytate (Davies and Nightingale 1975). The study authors suggested that the decrease in absorption was due to the formation of zinc-phytate complexes in the intestines. Phytate also reduced reabsorption of zinc secreted into the gastrointestinal tract of humans (Sandstrom and Sandberg 1992).

Endogenous substances, such as amino acids, can influence the absorption of zinc. Complexing of zinc with amino acids generally enhances its absorption in all segments of the intestine (Wapnir and Stiel 1986). Although neither zinc nor the amino acid proline are readily absorbed in the colon, complexing of zinc with proline during an *in vivo* intestinal perfusion in rats resulted in increased zinc absorption.

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Acrodermatitis enteropathica is a metabolic disorder that results in the malabsorption of zinc. However, when patients afflicted with this disorder were treated with human milk, zinc absorption was enhanced (Lombeck et al. 1975). It was reported by Evans (1980) that patients with acrodermatitis enteropathica have an impaired tryptophan metabolic pathway. Picolinic acid, a chief metabolite of tryptophan, is also a constituent of human milk. Picolinic acid is secreted by the pancreas into the intestinal lumen. A study by Boosalis et al. (1983) demonstrated that patients with pancreatic insufficiency had difficulty absorbing zinc administered as zinc sulfate. However, when these pancreatic-insufficient patients were given zinc as zinc picolinate, the extent of zinc absorption was similar to that of healthy controls. Zinc absorption may depend on the bioavailability of picolinic acid. Such a mandatory role of picolinic acid in absorption has not been confirmed (Bonewitz et al. 1982).

The addition of prostaglandin E<sub>2</sub> (PGE<sub>2</sub>) to the mucosal media of everted jejunal sacs from rats significantly increased zinc transport (Song and Adham 1979). In contrast, similar addition of prostaglandin F<sub>2</sub> (PGF<sub>2</sub>) significantly decreased zinc transport. Addition of PGF<sub>2</sub> to the serosal side of the jejunal sacs increased the transport of zinc to the mucosal side; PGE<sub>2</sub> decreased the serosal to mucosal transport of zinc. The mechanism by which prostaglandins regulate zinc transport has not been established (Song et al. 1992). The limitation of the *in vitro* study is the absence of vascular perfusion and consequent trapping of metals in the submucosal tissue. Hence, studies of absorption of heavy metals, including zinc, in everted sacs have limited physiological relevance (Foulkes 1984) but may provide information useful for the design of future *in vivo* experiments.

The presence of other trace metals (e.g., mercury, cadmium, copper) may also diminish zinc transport. Section 3.9 provides detailed information on the interaction of zinc with other metals.

#### 3.4.1.3 Dermal Exposure

Dermal absorption of zinc occurs, but its mechanism is not clearly defined. Studies are very limited regarding the absorption of zinc through the skin. Historically, zinc oxide has been used clinically to promote the healing of burns and wounds (Gordon et al. 1981). Absorption has been observed in burn patients treated with gauze dressings containing zinc oxide (Hallmans 1977). The pH of the skin, the amount of zinc applied, and the vehicle administered with zinc all affect the absorption of zinc (Agren 1990, 1991).

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Zinc chloride was also absorbed through the intact skin of the rat (Keen and Hurley 1977). Absorption of zinc sulfate was greater than zinc oxide following 4–48-hour dermal application to open wounds in Sprague-Dawley rats (Agren et al. 1991). About 12% of zinc oxide (0.25 mg zinc/cm<sup>2</sup>) from the dressing reached the wound while 65% of zinc sulfate (0.066 mg zinc/cm<sup>2</sup>) reached the wound. The data suggest that zinc oxide applied to wounds resulted in sustained delivery of zinc ions causing constant wound-tissue zinc levels. In contrast, zinc sulfate, being more water soluble than zinc oxide, is rapidly transferred into the blood and, therefore, caused decreased wound-tissue zinc levels (Agren et al. 1991).

#### 3.4.2 Distribution

Zinc is one of the most abundant trace metals in humans. It is found normally in all tissues and tissue fluids and is a cofactor in over 200 enzyme systems. Together, muscle and bone contain approximately 90% of the total amount of zinc in the body (~60% and 30%, respectively) (Wastney et al. 1986). Organs containing sizable concentrations of zinc are the liver, gastrointestinal tract, kidney, skin, lung, brain, heart, and pancreas (Bentley and Grubb 1991; Drinker and Drinker 1928; He et al. 1991; Llobet et al. 1988a). High concentrations of zinc were also detected in the prostate (Forssen 1972), retina, and sperm (Bentley and Grubb 1991). Zinc levels may vary considerably from one individual to another (Forssen 1972).

To some degree, the distribution of zinc in some tissues appears to be regulated by age (Schroeder et al. 1967). Zinc concentrations increase in the liver, pancreas, and prostate and decrease in the uterus and aorta with age. Levels in the kidneys and heart peak at approximately 40–50 years of age and then decline.

Zinc is present in blood plasma, erythrocytes, leukocytes, and platelets, but is chiefly localized within erythrocytes (of which 87% is in carbonic anhydrase, the major binding site) (Ohno et al. 1985). Zinc deficiency has been demonstrated to decrease the ability of erythrocytes to resist hemolysis *in vitro*. This finding suggests that zinc stabilizes the erythrocyte membrane. In plasma, two-thirds of the zinc is bound to albumin; the remainder is bound primarily to  $\alpha_2$ -macroglobulin (Bentley and Grubb 1991; Giroux et al. 1976; Wastney et al. 1986). It appears that the limited number of binding sites for zinc in plasma albumin and macroglobulin regulates the amount of zinc retained by the body (Andermann and Dietz 1982). Albumin-bound zinc has been correlated with plasma zinc levels, whereas  $\alpha_2$ -macroglobulin shows no correlation with plasma zinc levels.

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Hormones, such as the adrenocorticotrophic hormone (ACTH), appear to regulate the concentration of zinc in the liver. ACTH, secreted by the anterior pituitary gland, stimulates the secretion of glucocorticoids. Glucocorticoids, or hormones with glucocorticoid activity, have been shown *in vitro* to stimulate the net zinc uptake in cultured liver cells and at the same time activate the gene that regulates metallothionein synthesis (Failla and Cousins 1978). However, there are no *in vivo* data to support these *in vitro* findings. Metallothionein in the cells of the intestinal mucosa binds zinc, thus regulating its release into the blood.

The transfer of zinc across perfused placentas is slow; only  $\approx 3\%$  of maternal zinc reached the fetal compartment in 2 hours (Beer et al. 1992). The *in vitro* transfer of zinc between mother and fetus is bidirectional, with binding in the placenta (Beer et al. 1992). It is proposed that zinc uptake in the placenta involves a potassium/zinc transport system (Aslam and McArdle 1992). Newborns may also be exposed to zinc from their mothers by milk transfer of zinc during lactation (Rossowska and Nakamoto 1992).

#### 3.4.2.1 Inhalation Exposure

No studies were located regarding distribution in humans after inhalation exposure to zinc. However, occupational studies provided indirect evidence that zinc may distribute to tissues to produce systemic effects (Brown 1988; Drinker et al. 1927a; Malo et al. 1990; McCord et al. 1926; Rohrs 1957; Sturgis et al. 1927).

Zinc levels in the lungs of cats peaked immediately after acute exposure to 12–61 mg zinc/kg/day as zinc oxide for approximately 3 hours and remained high for 2 days postexposure, then dropped significantly thereafter (Drinker and Drinker 1928). Levels in pancreas, liver, and kidneys increased slowly.

#### 3.4.2.2 Oral Exposure

A single oral dose of 0.7 mg zinc/kg as zinc sulfate given to 11 individuals resulted in peak zinc levels in the plasma at 2–3 hours (Statter et al. 1988; Sturniolo et al. 1991). Similarly, Neve et al. (1991) reported peak serum zinc concentration at 2.3 hours with 0.7 mg zinc/kg as zinc sulfate.

Following feeding of 191 mg zinc/kg/day as zinc acetate to rats for 3 months, increased zinc levels were significant in the heart, spleen, kidneys, liver, bone, and blood (Llobet et al. 1988a). The greatest

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increases were in bone (258% of the control value) and blood (520% of the control value). Elevated zinc levels were found in the kidneys and liver of mice fed 76.9 mg zinc/kg/day as zinc sulfate (Schiffer et al. 1991) or 38 mg zinc/kg/day as zinc nitrate (Cooke et al. 1990) for approximately 1 month. The kidneys and pancreas had higher zinc levels than the liver and carcass of rats fed diets containing 1.1 mg/kg/day for an unspecified duration (Weigand and Kirchgessner 1992). Newborn, young, and adult mice receiving a single oral dose of 4.6 mg zinc/kg as zinc chloride generally had the highest levels of zinc in the liver, kidneys, lungs, bone, muscle, and carcass 1 day after dosing (He et al. 1991). However, the amount of zinc in the lungs, muscle, and femur decreased with age.

#### 3.4.2.3 Dermal Exposure

No studies were located regarding distribution in humans after dermal exposure to zinc.

Animal data on the distribution of zinc following dermal exposure are limited. Elevated serum zinc levels occurred with the application of zinc oxide or zinc sulfate to skin wounds of Sprague-Dawley rats for 4–48 hours (Agren et al. 1991). Serum zinc level peaked at 4 hours in rats treated with zinc sulfate, while levels were slightly elevated for 48 hours in rats administered zinc oxide. The differences may be attributed to the absorbability of the zinc compounds.

#### 3.4.3 Metabolism

Plasma provides a metabolically active transport compartment for zinc (Cousins 1985). Zinc is most often complexed to organic ligands (existing in loosely or firmly bound fractions) rather than free in solution as metallic ion (Gordon et al. 1981). Zinc is found in diffusible or nondiffusible forms in the blood (NAS/NRC 1979). In the diffusible form, approximately two-thirds of plasma zinc is freely exchangeable and loosely bound to albumin (Cousins 1985); the zinc-albumin complex has an association constant of about  $10^6$  (NAS/NRC 1979). The diffusible form of zinc also includes zinc bound to amino acids (primarily histidine and cysteine). The zinc-albumin complex is in equilibrium with the zinc-amino acid complex (Henkin 1974). The zinc-amino acid complex can be transported passively across tissue membranes to bind to proteins. An important binding protein in the kidney and liver is metallothionein, although other tissue-binding proteins may be present.

In the nondiffusible form, a small amount of circulating zinc is tightly bound to  $\alpha_2$ -macroglobulin in the plasma (Cousins 1985). Zinc is incorporated into and dissociated from  $\alpha_2$ -macroglobulin only in the liver

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(Henkin 1974). This zinc-protein complex has an association constant of  $>10^{10}$  (Henkin 1974; NAS/NRC 1979). The zinc bound to  $\alpha_2$ -macroglobulin is not freely exchangeable with other zinc ligands (i.e., zinc-albumin and zinc-amino acid complexes) in serum.

#### 3.4.4 Elimination and Excretion

##### 3.4.4.1 Inhalation Exposure

Information was limited regarding zinc excretion following inhalation exposure in humans. Workers exposed to zinc oxide fumes had elevated levels of zinc in the urine (Hamdi 1969) indicating that this is a route of excretion.

No studies were located regarding excretion in animals after inhalation exposure to zinc.

##### 3.4.4.2 Oral Exposure

The principal route of excretion of ingested zinc in humans is through the intestine (Davies and Nightingale 1975; Reinhold et al. 1991; Wastney et al. 1986). Zinc loss in the body is by secretion via the gut, and the remainder occurs in the urine (Wastney et al. 1986). Fecal excretion of zinc increases as intake increases (Spencer et al. 1985). Excretion of zinc in the urine also reflects zinc intake (Wastney et al. 1986). Minor routes of elimination are saliva secretion, hair loss, and sweat (Greger and Sickles 1979; Hambidge et al. 1972; Henkin et al. 1975a; Prasad et al. 1963a; Rivlin 1983).

There was a linear increase in fecal excretion of zinc in proportion to dietary intake in rats fed supplementations of 32 mg zinc/kg/day as zinc oxide for 7–42 days (Ansari et al. 1975) or 50–339 mg/kg/day for 21 days (Ansari et al. 1976). No differences in fecal excretion, total excretion, or retention of zinc were found among rats given diets containing different forms of zinc (Seal and Heaton 1983). Rats receiving 2.65 mg zinc/kg/day as zinc chloride, zinc sulfate, zinc phosphate, or zinc citrate, over a 4-day period excreted 87–98% of intake.

A study by Alexander et al. (1981) demonstrated that zinc is excreted in the bile of rats. Analysis of the bile indicated that the zinc is primarily complexed with reduced glutathione. Treatment of these rats with diethylmaleate, which conjugates with reduced glutathione and restricts its availability, depressed the

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biliary excretion of zinc. This depression confirms a relationship between zinc and glutathione and suggests that zinc is transferred from liver to bile by a glutathione-dependent process.

Other factors may affect zinc excretion. For example, low dietary intake of zinc or malnutrition can increase the urinary excretion of zinc. This release of zinc is a result of tissue breakdown and catabolism during starvation; and elevated urinary excretion of zinc may persist after intake levels return to normal (Spencer et al. 1976). Administration of histidine or high-protein diet may increase urinary zinc excretion; however, a corresponding increase in zinc absorption may maintain zinc balance in the body (Henkin et al. 1975b; Hunt et al. 1991).

#### **3.4.4.3 Dermal Exposure**

No studies were located regarding excretion in humans or animals after dermal exposure to zinc.

#### **3.4.5 Physiologically Based Pharmacokinetic (PBPK)/Pharmacodynamic (PD) Models**

Physiologically based pharmacokinetic (PBPK) models use mathematical descriptions of the uptake and disposition of chemical substances to quantitatively describe the relationships among critical biological processes (Krishnan et al. 1994). PBPK models are also called biologically based tissue dosimetry models. PBPK models are increasingly used in risk assessments, primarily to predict the concentration of potentially toxic moieties of a chemical that will be delivered to any given target tissue following various combinations of route, dose level, and test species (Clewell and Andersen 1985). Physiologically based pharmacodynamic (PBPD) models use mathematical descriptions of the dose-response function to quantitatively describe the relationship between target tissue dose and toxic end points.

PBPK/PD models refine our understanding of complex quantitative dose behaviors by helping to delineate and characterize the relationships between: (1) the external/exposure concentration and target tissue dose of the toxic moiety, and (2) the target tissue dose and observed responses (Andersen et al. 1987; Andersen and Krishnan 1994). These models are biologically and mechanistically based and can be used to extrapolate the pharmacokinetic behavior of chemical substances from high to low dose, from route to route, between species, and between subpopulations within a species. The biological basis of PBPK models results in more meaningful extrapolations than those generated with the more conventional use of uncertainty factors.



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The PBPK model for a chemical substance is developed in four interconnected steps: (1) model representation, (2) model parametrization, (3) model simulation, and (4) model validation (Krishnan and Andersen 1994). In the early 1990s, validated PBPK models were developed for a number of toxicologically important chemical substances, both volatile and nonvolatile (Krishnan and Andersen 1994; Leung 1993). PBPK models for a particular substance require estimates of the chemical substance-specific physicochemical parameters, and species-specific physiological and biological parameters. The numerical estimates of these model parameters are incorporated within a set of differential and algebraic equations that describe the pharmacokinetic processes. Solving these differential and algebraic equations provides the predictions of tissue dose. Computers then provide process simulations based on these solutions.

The structure and mathematical expressions used in PBPK models significantly simplify the true complexities of biological systems. If the uptake and disposition of the chemical substance(s) is adequately described, however, this simplification is desirable because data are often unavailable for many biological processes. A simplified scheme reduces the magnitude of cumulative uncertainty. The adequacy of the model is, therefore, of great importance, and model validation is essential to the use of PBPK models in risk assessment.

PBPK models improve the pharmacokinetic extrapolations used in risk assessments that identify the maximal (i.e., the safe) levels for human exposure to chemical substances (Andersen and Krishnan 1994). PBPK models provide a scientifically sound means to predict the target tissue dose of chemicals in humans who are exposed to environmental levels (for example, levels that might occur at hazardous waste sites) based on the results of studies where doses were higher or were administered in different species. Figure 3-3 shows a conceptualized representation of a PBPK model.

Validated PBPK models for zinc in animals or humans are not presently available.

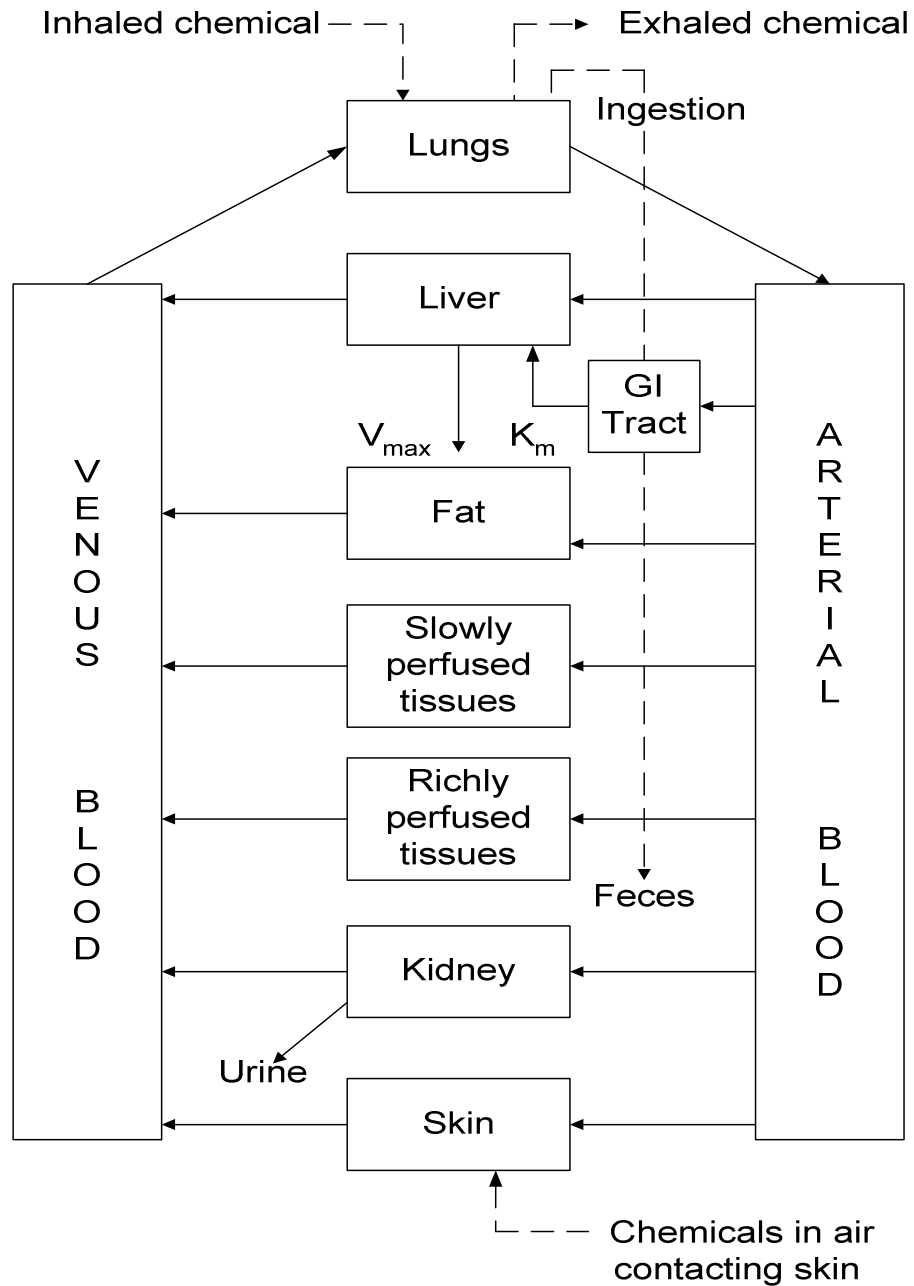
## **3.5 MECHANISMS OF ACTION**

### **3.5.1 Pharmacokinetic Mechanisms**

The absorption of zinc from the intestine is homeostatically controlled. A study by Hempe and Cousins (1992) found that CRIP, a diffusible intracellular zinc carrier, binds zinc in the mucosa during absorption; this process appears to be saturable (Gunshin et al. 1991; Hempe and Cousins 1992; Sturniolo et al.

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**Figure 3-3. Conceptual Representation of a Physiologically Based Pharmacokinetic (PBPK) Model for a Hypothetical Chemical Substance**



Source: adapted from Krishnan et al. 1994

Note: This is a conceptual representation of a physiologically based pharmacokinetic (PBPK) model for a hypothetical chemical substance. The chemical substance is shown to be absorbed via the skin, by inhalation, or by ingestion, metabolized in the liver, and excreted in the urine or by exhalation.

1991). Zinc transport in the intestinal lumen is also influenced by metallothionein which can inhibit zinc absorption by competing with CRIP for zinc (Hempe and Cousins 1992). CRIP binds about 40% of radiolabeled zinc entering the intestinal cells from the lumen in ligated loops of the small intestine of anesthetized rats when the zinc concentration is low (5  $\mu\text{M}$ ), but only 14% of the radiolabel when the concentration is high (300  $\mu\text{M}$ ) (Hempe and Cousins 1991). These findings suggest that CRIP has a limited binding capacity for zinc and becomes saturated when zinc concentration in the intestine is high (Hempe and Cousins 1992).

High luminal zinc concentrations may damage the brush border membrane, allowing zinc to enter the cell and bind nonspecifically to cell proteins and other ligands (Cousins 1985; Hempe and Cousins 1992). Within the intestinal lumen, a number of factors appear to influence the availability of zinc for absorption. Methionine, histidine, cysteine, reduced glutathione, citrate, and prostaglandin  $\text{E}_2$  increase the intestinal uptake of zinc (Song et al. 1992), whereas inorganic inhibitors of zinc absorption include cadmium, copper, calcium, and ferrous iron (Hamilton et al. 1978; Harford and Sarkar 1991; Ogiso et al. 1979; Spencer et al. 1992; Yoshida et al. 1993). The mechanism of inhibition has not been clearly elucidated, but it is believed to involve competition for zinc binding sites in the intestinal mucosal cells; an effect on charge distribution on the mucosal membrane has also been suggested (Foulkes 1985). The organic inhibitors, including phytate and some components of dietary fiber, are believed to complex with zinc and decrease its availability. In the mucosal cell, zinc is associated with metalloproteins, including metallothionein. The release of zinc from the intracellular protein ligands and its transfer to the blood may involve diffusion of complexes with glutathione and similar compounds (Foulkes 1993).

In the plasma, albumin is the primary carrier for zinc, with smaller amounts of zinc bound to  $\alpha_2$ -macroglobulin and amino acids (Giroux et al. 1976). The albumin-bound zinc represents the metabolically active pool of zinc. Zinc is loosely bound in plasma, and albumin-bound zinc can readily be given up to tissues; however, the mechanisms are not fully elucidated. Zinc is initially concentrated in the liver after ingestion, and is subsequently distributed throughout the body. The liver, pancreas, bone, kidney, and muscle are the major tissue storage sites. When plasma zinc levels are high, liver metallothionein synthesis is stimulated, which facilitates the retention of zinc by hepatocytes (Richards and Cousins 1975). A storage form of zinc has not been identified in soft tissues, with the possible exception of zinc metallothionein. Zinc in bone is relatively unavailable for use by other tissues.

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**3.5.2 Mechanisms of Toxicity**

Metal fume fever is the primary effect observed in workers exposed to zinc oxide fumes or dust (Blanc et al. 1991; Brown 1988; Drinker et al. 1927b; Vogelmeier et al. 1987). Metal fume fever usually occurs 3–10 hours after exposure, and the symptoms persist for 24–48 hours. The exact pathogenesis of metal fume fever is not known. It is believed to be an immune response to the inhaled zinc oxide (Mueller and Seger 1985). It has been suggested that the zinc oxide causes inflammation of the respiratory tract and the release of histamine or histamine-like substances. In response, an allergen-antibody complex is formed that may elicit an allergic reaction upon subsequent exposure to the allergen. In response to the allergen-antibody complex, an anti-antibody is formed. The anti-antibody dominates with continued exposure to the zinc oxide, thereby producing a tolerance. When the exposure is interrupted and re-exposure occurs, the allergen-antibody complex dominates, producing an allergic reaction and symptoms of metal fume fever (McCord 1960).

Oral exposure to high levels of zinc has caused anemia, decreased levels of HDL cholesterol, and pancreatic damage in humans (Black et al. 1988; Chandra 1984; Chobanian 1981; Hooper et al. 1980; Murphy 1970) and animals (Allen et al. 1983; Aughey et al. 1977; Drinker et al. 1927d; Katya-Katya et al. 1984; Klevay and Hyg 1973; Maita et al. 1981; Straube et al. 1980). The mechanisms involved in the pancreatic damage have not been elucidated. The anemia and possibly the decreased HDL cholesterol levels are thought to be caused by a zinc-induced copper deficiency. Although it is generally accepted that the anemia is the result of copper deficiency, the relationship between zinc and copper levels and HDL cholesterol levels has been extensively debated (Fischer et al. 1980; Katya-Katya et al. 1984; Klevay and Hyg 1973; Murthy and Petering 1976). However, that topic is beyond the scope of this profile.

**3.6 TOXICITIES MEDIATED THROUGH THE NEUROENDOCRINE AXIS**

Recently, attention has focused on the potential hazardous effects of certain chemicals on the endocrine system because of the ability of these chemicals to mimic or block endogenous hormones. Chemicals with this type of activity are most commonly referred to as *endocrine disruptors*. However, appropriate terminology to describe such effects remains controversial. The terminology *endocrine disruptors*, initially used by Colborn and Clement (1992), was also used in 1996 when Congress mandated the Environmental Protection Agency (EPA) to develop a screening program for “...certain substances [which] may have an effect produced by a naturally occurring estrogen, or other such endocrine

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effect[s]...”. To meet this mandate, EPA convened a panel called the Endocrine Disruptors Screening and Testing Advisory Committee (EDSTAC), which in 1998 completed its deliberations and made recommendations to EPA concerning *endocrine disruptors*. In 1999, the National Academy of Sciences released a report that referred to these same types of chemicals as *hormonally active agents*. The terminology *endocrine modulators* has also been used to convey the fact that effects caused by such chemicals may not necessarily be adverse. Many scientists agree that chemicals with the ability to disrupt or modulate the endocrine system are a potential threat to the health of humans, aquatic animals, and wildlife. However, others think that endocrine-active chemicals do not pose a significant health risk, particularly in view of the fact that hormone mimics exist in the natural environment. Examples of natural hormone mimics are the isoflavonoid phytoestrogens (Adlercreutz 1995; Livingston 1978; Mayr et al. 1992). These chemicals are derived from plants and are similar in structure and action to endogenous estrogen. Although the public health significance and descriptive terminology of substances capable of affecting the endocrine system remains controversial, scientists agree that these chemicals may affect the synthesis, secretion, transport, binding, action, or elimination of natural hormones in the body responsible for maintaining homeostasis, reproduction, development, and/or behavior (EPA 1997). Stated differently, such compounds may cause toxicities that are mediated through the neuroendocrine axis. As a result, these chemicals may play a role in altering, for example, metabolic, sexual, immune, and neurobehavioral function. Such chemicals are also thought to be involved in inducing breast, testicular, and prostate cancers, as well as endometriosis (Berger 1994; Giwercman et al. 1993; Hoel et al. 1992).

***Pancreas.*** Increased levels of serum amylase were observed in a man after accidental ingestion of about 3 ounces of a zinc chloride solution (Chobanian 1981). A 16-year-old boy who ingested an average of approximately 86 mg zinc/kg/day as metallic zinc for 2 days (114 mg/kg on the 1st day and 57 mg/kg on the 2nd day) had increased serum amylase and lipase (Murphy 1970).

In humans receiving a single low dose of zinc sulfate (0.5 mg zinc/kg/day), no changes in blood glucose or insulin levels were observed, and there were no differences in response to a glucose load (Brandao-Neto et al. 1990b).

Pancreatic abnormalities (islet cellular alterations, acinar cell necrosis, metaplasia, fibrosis, pancreatitis) resulting from zinc ingestion have been observed in rats (Maita et al. 1981), mice (Aughey et al. 1977; Maita et al. 1981), cats (Drinker et al. 1927d), ferrets (Straube et al. 1980), sheep (Allen et al. 1983), and birds (Kazacos and Van Vleet 1989; Lu et al. 1990). In dogs (Drinker et al. 1927d) and minks (Aulerich et al. 1991), histological changes in the pancreas have not been observed at doses comparable to or higher

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than the dose levels that caused abnormalities in rats, mice, cats, ferrets, and sheep. Degeneration of the acinar cells of the pancreas was observed in sheep by Allen et al. (1983) and in rats and mice by Maita et al. (1981). Since the pancreatic acinar cells secrete digestive juices into the small intestine, the increase in serum amylase and lipase observed in the human case reports (Chobanian 1981; Murphy 1970) would correspond to damage in this region of the pancreas.

In 2-month-old C3H mice exposed to 70 mg zinc/kg/day as zinc sulfate, hypertrophy and vacuolation of the  $\beta$ -cells of the pancreatic islets were observed beginning after 3 months of exposure and become more severe by 12 months (Aughey et al. 1977). The pancreatic islets secrete the hormones glucagon and insulin. No change in plasma levels of insulin and glucose was observed in this study after 6 months of exposure. No effect on islet cells was reported in rats exposed up to 565 mg/kg/day or mice exposed to 1110 mg/kg/day as zinc sulfate in a 13-week study by Maita et al. (1981), and Allen et al. (1983) reported that islet cells in sheep were generally unaffected, although occasional vacuolization occurred. Degeneration of acinar cells, but no effects on the islet cells, were found in ducklings (Kazacos and Van Vleet 1989); however, the relevance of this to humans is unclear. The data are too limited and contradictory to determine whether pancreatic islet cells are a primary target cell of zinc toxicity.

***Adrenal Gland.*** Decreased levels of serum cortisol (a hormone secreted by the adrenal cortex) were observed in humans after a single dose of 0.5 mg zinc/kg/day as zinc sulfate (Brandao-Neto et al. 1990b). No effects on the adrenal gland itself have been reported in humans.

In mice receiving 70 mg zinc/kg/day as zinc sulfate in the drinking water, hypertrophy and increased lipid content of the zona fasciculata cells of the adrenal cortex were observed as early as 3 months after the start of zinc supplementation (Aughey et al. 1977).

***Pituitary.*** No effects on pituitary function have been reported in humans following oral exposure to zinc. However, mice receiving 70 mg zinc/kg/day as zinc sulfate in the drinking water for 5–14 months had hypertrophy and increased granularity suggesting increased activity of the pituitary (Aughey et al. 1977). It is unclear whether the increased activity was a direct effect of the zinc or a reaction to decreased secretion from the adrenal cortex.

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**3.7 CHILDREN'S SUSCEPTIBILITY**

This section discusses potential health effects from exposures during the period from conception to maturity at 18 years of age in humans, when all biological systems will have fully developed. Potential effects on offspring resulting from exposures of parental germ cells are considered, as well as any indirect effects on the fetus and neonate resulting from maternal exposure during gestation and lactation. Relevant animal and *in vitro* models are also discussed.

Children are not small adults. They differ from adults in their exposures and may differ in their susceptibility to hazardous chemicals. Children's unique physiology and behavior can influence the extent of their exposure. Exposures of children are discussed in Section 6.6 Exposures of Children.

Children sometimes differ from adults in their susceptibility to hazardous chemicals, but whether there is a difference depends on the chemical (Guzelian et al. 1992; NRC 1993). Children may be more or less susceptible than adults to health effects, and the relationship may change with developmental age (Guzelian et al. 1992; NRC 1993). Vulnerability often depends on developmental stage. There are critical periods of structural and functional development during both prenatal and postnatal life and a particular structure or function will be most sensitive to disruption during its critical period(s). Damage may not be evident until a later stage of development. There are often differences in pharmacokinetics and metabolism between children and adults. For example, absorption may be different in neonates because of the immaturity of their gastrointestinal tract and their larger skin surface area in proportion to body weight (Morselli et al. 1980; NRC 1993); the gastrointestinal absorption of lead is greatest in infants and young children (Ziegler et al. 1978). Distribution of xenobiotics may be different; for example, infants have a larger proportion of their bodies as extracellular water and their brains and livers are proportionately larger (Altman and Dittmer 1974; Fomon 1966; Fomon et al. 1982; Owen and Brozek 1966; Widdowson and Dickerson 1964). The infant also has an immature blood-brain barrier (Adinolfi 1985; Johanson 1980) and probably an immature blood-testis barrier (Setchell and Waites 1975). Many xenobiotic metabolizing enzymes have distinctive developmental patterns. At various stages of growth and development, levels of particular enzymes may be higher or lower than those of adults, and sometimes unique enzymes may exist at particular developmental stages (Komori et al. 1990; Leeder and Kearns 1997; NRC 1993; Vieira et al. 1996). Whether differences in xenobiotic metabolism make the child more or less susceptible also depends on whether the relevant enzymes are involved in activation of the parent compound to its toxic form or in detoxification. There may also be differences in excretion, particularly in newborns who all have a low glomerular filtration rate and have not developed efficient tubular secretion and resorption capacities (Altman and Dittmer 1974; NRC 1993; West et al. 1948).

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Children and adults may differ in their capacity to repair damage from chemical insults. Children also have a longer remaining lifetime in which to express damage from chemicals; this potential is particularly relevant to cancer.

Certain characteristics of the developing human may increase exposure or susceptibility, whereas others may decrease susceptibility to the same chemical. For example, although infants breathe more air per kilogram of body weight than adults breathe, this difference might be somewhat counterbalanced by their alveoli being less developed, which results in a disproportionately smaller surface area for alveolar absorption (NRC 1993).

While a detailed discussion of zinc deficiency is beyond the scope of this document, there is considerably more information on the effects of zinc deficiency on the developing fetus in pregnant women than exists for the effects of excess zinc during pregnancy. Maternal zinc deficiency can result in intrauterine growth retardation, teratogenesis, or embryonic or fetal death (for review, see King 2000). Zinc supplementation during pregnancy is usually sufficient to prevent these outcomes. Similarly, zinc deficiency during early life can result in adverse effects, including skin rash, diarrhea, anorexia, and growth failure, with more severe instances resulting in detrimental effects on the immune and nervous systems (Krebs 1999); infants appear to be particularly sensitive to zinc deficiency.

A case study presented by Murray (1926) reported on an infant death due to bronchopneumonia resulting from inhalation, and possibly ingestion, of an unspecified amount of zinc stearate powder spilled from a container. However, it is unclear whether the death was due to the zinc content or whether aspiration bronchopneumonia would result from inhalation of similar powders that do not contain zinc. Other data on the effects of zinc inhalation in young children are not available.

The human data on the effects of excess zinc in children consist mainly of reports of acute ingestion. The primary symptoms in these subjects mimic those of adult exposure, consisting mainly of gastrointestinal disturbances (nausea, vomiting, epigastric discomfort), with occasional neurologic symptoms (Anonymous 1983; Lewis and Kokan 1998; Moore 1978; Murphy 1970). Data are not presently sufficient to determine whether children are more sensitive to these effects than adults.

The most sensitive animal model to zinc toxicity in young animals appears to be the mink. Young minks appear to be more sensitive to both the hematologic (decreased hematocrit and lymphocyte number) and dermal effects (graying of the fur and dermatosis) of oral zinc than adults (Bleavins et al. 1983). Other



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studies have examined the effects of zinc exposure in young animals (Drinker et al. 1927d; L'Abbe and Fischer 1984a; Maita et al. 1981), but have not provided data on adult animals similarly exposed for comparison. Additional data will be required to adequately assess the susceptibility of children to zinc exposure, relative to adults.

#### **3.8 BIOMARKERS OF EXPOSURE AND EFFECT**

Biomarkers are broadly defined as indicators signaling events in biologic systems or samples. They have been classified as markers of exposure, markers of effect, and markers of susceptibility (NAS/NRC 1989a).

Due to a nascent understanding of the use and interpretation of biomarkers, implementation of biomarkers as tools of exposure in the general population is very limited. A biomarker of exposure is a xenobiotic substance or its metabolite(s) or the product of an interaction between a xenobiotic agent and some target molecule(s) or cell(s) that is measured within a compartment of an organism (NAS/NRC 1989a). The preferred biomarkers of exposure are generally the substance itself or substance-specific metabolites in readily obtainable body fluid(s), or excreta. However, several factors can confound the use and interpretation of biomarkers of exposure. The body burden of a substance may be the result of exposures from more than one source. The substance being measured may be a metabolite of another xenobiotic substance (e.g., high urinary levels of phenol can result from exposure to several different aromatic compounds). Depending on the properties of the substance (e.g., biologic half-life) and environmental conditions (e.g., duration and route of exposure), the substance and all of its metabolites may have left the body by the time samples can be taken. It may be difficult to identify individuals exposed to hazardous substances that are commonly found in body tissues and fluids (e.g., essential mineral nutrients such as copper, zinc, and selenium). Biomarkers of exposure to zinc are discussed in Section 3.8.1.

Biomarkers of effect are defined as any measurable biochemical, physiologic, or other alteration within an organism that, depending on magnitude, can be recognized as an established or potential health impairment or disease (NAS/NRC 1989a). This definition encompasses biochemical or cellular signals of tissue dysfunction (e.g., increased liver enzyme activity or pathologic changes in female genital epithelial cells), as well as physiologic signs of dysfunction such as increased blood pressure or decreased lung capacity. Note that these markers are not often substance specific. They also may not be directly adverse, but can indicate potential health impairment (e.g., DNA adducts). Biomarkers of effects caused by zinc are discussed in Section 3.8.2.

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A biomarker of susceptibility is an indicator of an inherent or acquired limitation of an organism's ability to respond to the challenge of exposure to a specific xenobiotic substance. It can be an intrinsic genetic or other characteristic or a preexisting disease that results in an increase in absorbed dose, a decrease in the biologically effective dose, or a target tissue response. If biomarkers of susceptibility exist, they are discussed in Section 3.10 "Populations That Are Unusually Susceptible".

#### **3.8.1 Biomarkers Used to Identify or Quantify Exposure to Zinc**

There is no simple measure of zinc body burden. Under normal physiological conditions, the plasma/serum zinc level is  $\approx 1$   $\mu\text{g/mL}$  (NAS/NRC 1979) and the urinary level is 0.5 mg/g creatinine (Elinder 1986). Several studies have reported increased levels of zinc in the serum and urine of humans and animals after inhalation, oral, or dermal exposure to zinc (Agren et al. 1991; Bentley and Grubb 1991; Brandao-Neto et al. 1990a; Hallmans 1977; Hamdi 1969; Keen and Hurley 1977; Neve et al. 1991; Statter et al. 1988; Sturniolo et al. 1991). However, relationships between serum and/or urine levels and zinc exposure levels have not been established.

Hair and nail samples provide a lasting record of long-term metal intake possibly over weeks or months (Hayashi et al. 1993; Wilhelm et al. 1991). Mean zinc concentrations of 129–179  $\mu\text{g/g}$  have been estimated for nails (Hayashi et al. 1993; Wilhelm et al. 1991) and 102–258  $\mu\text{g/g}$  for hair (Folin et al. 1991; McBean et al. 1971; Provost et al. 1993; Wilhelm et al. 1991). Most investigators have found a poor correlation between hair and plasma zinc levels since the zinc in hair does not exchange with the body zinc pool (McBean et al. 1971; Rivlin 1983). Furthermore, measurements of zinc in hair can be affected by extraneous contamination of hair, contamination by sweat, location of hair sample (distance from scalp), hair coloring, and rate of hair growth (McBean et al. 1971; Rivlin 1983). Although the nail is considered more resistant to washing procedures than hair, external contamination and uncertainties regarding the length and period of exposure reflected by the observed zinc concentration limit this measurement as a biomarker of exposure for zinc (Wilhelm et al. 1991).

#### **3.8.2 Biomarkers Used to Characterize Effects Caused by Zinc**

The respiratory tract is the most sensitive target organ for zinc following inhalation exposure. Inhalation of zinc oxide results in a syndrome referred to as metal fume fever. Symptoms include fevers, chills, cough, listlessness, and metallic taste. Although oxides of several heavy metals (antimony, aluminum,

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arsenic, cadmium, cobalt, copper, iron, lead, magnesium, manganese, mercury, nickel, selenium, silver, and tin) and pyrolysis products of fluorocarbon polymers (polytetrafluoroethylene [Teflon] and fluorinated polyethylene propylene) also produce metal fume fever (Ellenhorn and Barceloux 1988), this group of symptoms may be used as a nonspecific biomarker to identify inhalation exposure to zinc oxide.

The target organs associated with oral zinc exposure include the gastrointestinal tract, blood, immune system, and pancreas. The toxic effects observed after oral exposure to zinc include nausea, vomiting, diarrhea, decreased hemoglobin and hematocrit levels, immune suppression, increased serum amylase and lipase, and decreased HDL cholesterol levels (a more detailed discussion of effects associated with exposure to zinc is presented in Section 3.2). However, nausea, vomiting, and diarrhea may be observed following exposure to any gastrointestinal irritant. Increases in serum amylase and lipase are also markers for pancreatic damage; therefore, any condition resulting in pancreatitis (i.e., biliary tract disease [gallstones], alcoholism, trauma, inflammation, blood-borne bacterial infections, viral infections, ischemia, and drugs such as azathioprine, thiazides, sulfonamides, and oral contraceptives) would result in similar increases in these enzymes (Cotran et al. 1989). A hypochromic microcytic anemia that is not responsive to iron supplements may indicate exposure to zinc; however, such anemia may also reflect copper, pyridoxine, or cobalt deficiency, lead intoxication, poor diet, or chronic blood loss (Suber 1989).

Thus, none of the above-mentioned effects observed after exposure to zinc is specific to zinc exposure. However, the combination of these toxic effects may be indicative of zinc overexposure. Additional information on the health effects of zinc may be found in Section 3.2.2. Additional information on biomarkers for renal, hepatobiliary, immune, and nervous system effects may be found in the CDC/ATSDR (1990) and OTA (1990) reports listed in Chapter 9.

Increased erythrocyte metallothionein may be an index of zinc exposure in humans (Grider et al. 1990). Daily supplementation of 50 mg zinc/day to subjects for at least 7 days caused a seven-fold increase in metallothionein concentration in erythrocytes. At least 3–4 days are required before an increase in metallothionein is observed. This biomarker of exposure is only useful for recent zinc exposure because the metallothionein levels decreased approximately a week after discontinuation of a 63-week supplementation of zinc (Grider et al. 1990). Fourteen days after discontinuation of zinc supplements, metallothionein levels were reduced by 61%.

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**3.9 INTERACTIONS WITH OTHER CHEMICALS**

Zinc is an essential element obtained from the diet. Many different metals and nutrients interact with the absorption, distribution, and excretion of zinc. However, information was not found concerning interactions that increase the toxicity of zinc or other substances in the presence of zinc (i.e., that cause the same amount of zinc to result in a greater toxic response). Zinc administration may increase the toxicity of lead; however, the data are conflicting (Cerklewski and Forbes 1976; Hsu et al. 1975). The toxicity of zinc is believed to be due to its interaction with copper, as explained below.

Metallothionein, a sulfhydryl-rich protein inducible by certain divalent cations and a variety of other agonists, is involved in the interaction between zinc and other metals such as copper (Wapnir and Balkman 1991). Inhibition of intestinal copper absorption by zinc may demonstrate competition between the two metals at the brush border of the lumen (Wapnir and Balkman 1991). Dietary intake of copper (1, 6, and 36 mg/kg) or zinc (5, 30, and 180 mg/kg) do not significantly alter the absorption of the other (Oestreicher and Cousins 1985), but when zinc levels are much higher than copper levels, copper absorption is depressed (Fischer et al. 1981). High levels of dietary zinc are known to induce *de novo* synthesis of metallothionein in the intestinal mucosal cell. Both copper and zinc appear to bind to the same metallothionein protein; however, copper has a higher affinity for metallothionein than zinc and displaces the zinc that is attached to the metallothionein (Ogiso et al. 1979). Copper complexed with metallothionein is retained in the mucosal cell, relatively unavailable for transfer to plasma, and is excreted in the feces when the mucosal cells are sloughed off (Fischer et al. 1981; L'Abbe and Fischer 1984b). A number of factors influence the effect of dietary zinc on copper metabolism, including the amount of copper and zinc in the diet, the zinc-to-copper ratio, age of the individual, and the duration of exposure to high zinc levels (Johnson and Flagg 1986).

Physiological interactions of zinc and cadmium have been discussed in a number of reviews (EPA 1980c; NAS 1980; Underwood 1977). Exposure to cadmium may cause changes in the distribution of zinc, with accumulation of zinc in the liver and kidney. This accumulation in the liver and kidney may result in a deficiency in other organs, particularly if the dietary intake of zinc is marginal. *In vitro* data demonstrate that zinc and cadmium enter renal proximal cells by a saturable, carrier-mediated process and a non-saturable pathway (Gachot and Poujeol 1992). At low cadmium doses, cadmium and zinc compete for a common transport carrier system in renal proximal cells. It is hypothesized that, at high doses, the subcellular microtubule system is disrupted by cadmium, which may interfere with changes in carrier configuration that are necessary for transport of the metals (modification of the cytoskeleton), and thereby lead to noncompetitive inhibition between cadmium and zinc (Gachot and Poujeol 1992). Combined

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treatment with cadmium and zinc in primary cultures of kidney cells resulted in enhanced toxicity of cadmium (Yoshida et al. 1993); however, pretreatment with a nontoxic concentration of zinc caused increased induction of metallothionein synthesis and partial protection against cadmium (Yoshida et al. 1993).

Cadmium is 10 times more efficient than zinc in metallothionein induction *in vitro* (Harford and Sarkar 1991). Induction by either cadmium or zinc alone is saturable; however, simultaneous administration of cadmium and zinc results in induction of metallothionein in an additive manner. The additive effect on metallothionein induction may involve binding of the metals either to two or more metallothionein promoter binding proteins or separate sites on the same promoter binding protein (Harford and Sarkar 1991).

Zinc acetate pretreatment in the mouse TRL-1215 cell line reduced single-strand DNA damage associated with cadmium exposure (Coogan et al. 1992). Diminished cadmium-induced DNA damage was not due to decreased cadmium burden in the zinc-pretreated cells. Instead, cadmium levels were actually greater than those in non-pretreated cells (Coogan et al. 1992). Metallothionein levels were elevated in these cells, suggesting that zinc pretreatment affects cadmium genotoxicity by inducing metallothionein which may sequester cadmium from genetic material. In contrast, simultaneous exposure to cadmium and zinc decreased cadmium accumulation in the cells, perhaps because of direct competition for a common transport mechanism (Coogan et al. 1992).

Zinc acetate reduced or prevented cadmium carcinogenesis in the prostate, in the testes, or at the injection site in rats (Gunn et al. 1963a, 1964; Waalkes et al. 1989). The effect of zinc on the cadmium-induced carcinogenesis appeared to be dependent on dose, route, and target site. Sustained levels of zinc inhibited cadmium-induced injection sarcomas but had no effect on the incidence of testicular Leydig cell tumors (Waalkes et al. 1989).

Excessive dietary zinc has been shown to induce a reversible copper deficiency and anemia in experimental animals (Magee and Matrone 1960; Murthy and Petering 1976; O'Dell 1969; Underwood 1977; Wapnir and Balkman 1991). Similar effects have been seen in humans receiving long-term treatment with zinc (Porter et al. 1977; Prasad et al. 1978). However, no significant decreases in plasma copper levels were observed in humans receiving zinc for 6 weeks or 6 months (Henkin et al. 1976; Samman and Roberts 1987) or in mice administered zinc for 1–12 weeks (Sutomo et al. 1992). A reduction in erythrocyte superoxide dismutase (an index of metabolically available copper), without a

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decrease in plasma copper levels, was exhibited following exposure to high amounts of ingested zinc (Fischer et al. 1984). These findings suggest that superoxide dismutase may be a sensitive indicator of zinc-copper interaction.

Cobalt has been demonstrated to induce seminiferous tubule damage and degeneration (vacuole formation, sloughing of cells, giant cell formation) in the testes of mice following exposure for 13 weeks (Anderson et al. 1993). Coadministration of cobalt and zinc chloride in the drinking water resulted in complete or partial protection in 90% of the animals. The sites of competitive interaction between zinc and cobalt were not established in the study; however, the authors postulated that the mechanism(s) may be similar to those involved in prevention of cadmium toxicity by zinc.

The effect of tin on heme biosynthesis appears to be dependent on the concentration of zinc (Chmielnicka et al. 1992). Oral administration of tin can affect the heme synthesis by inhibiting  $\delta$ -aminolevulinic acid dehydratase (ALAD) activity in blood. Zinc is required for ALAD activity and provides a protective role in heme synthesis by increasing the activity of ALAD. It is postulated that when the tin and zinc are coadministered, these metals are probably attaching to similar binding sites in the ALAD enzyme (Chmielnicka et al. 1992).

Calcium decreases the bioavailability of zinc; the converse is also true (Heth and Hoekstra 1965; Spencer et al. 1992). Oral zinc administration is associated with decreased calcium levels in the serum and in the bone of rats (Yamaguchi et al. 1983). Zinc inhibited calcium uptake in rat brush border membrane vesicles, possibly by competing directly at high-affinity calcium binding sites (Roth-Bassell and Clydesdale 1991). The interaction of calcium and zinc is apparently dose related; intestinal absorption of calcium at a low calcium intake (230 mg/day) was inhibited at a high zinc intake of 140 mg/day but not at a lower zinc intake of 100 mg/day (Spencer et al. 1992).

Pretreatment with zinc has been shown to reduce hepatotoxicity induced by xenobiotics such as acetaminophen, bromobenzene, carbon tetrachloride, D-galactosamine, gentamicin, and salicylate (Cagen and Klaassen 1979; Gunther et al. 1991; Hu et al. 1992; Szymanska et al. 1991; Yang et al. 1991). The protective effect of zinc against carbon tetrachloride toxicity is dose dependent at high dose levels of zinc, probably because of sequestering of toxic metabolites of carbon tetrachloride by metallothionein (Cagen and Klaassen 1979). Similarly, the protective action of zinc against bromobenzene and acetaminophen appears to be associated with elevated metallothionein levels (Szymanska et al. 1991). Inhibition of lipid peroxidation may be the basis for the protective effect of zinc against hepatic damage induced by

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D-galactosamine in rats (Hu et al. 1992). Zinc may be elevating NADPH (nicotinamide adenine dinucleotide phosphate) content in the cell, resulting in regeneration of glutathione, which increases the antioxidative ability of hepatic cells. Salicylate-induced hepatic alterations (increased lipid droplets and iron, reduced glycogen) (Gunther et al. 1991) and gentamicin-induced proximal tubular necrosis (Yang et al. 1991) were diminished in rats pretreated with injections of zinc chloride and zinc sulfate, respectively. This finding corresponded to a dramatic increase in metallothionein content with combined treatment of salicylate and zinc compared to a less significant increase with salicylate alone.

Animal studies suggest that the administration of zinc may also inhibit tumor growth. Forty weeks after exposure, the incidence of injection site sarcomas was 40–60% in rats receiving simultaneous intramuscular administration of nickel subsulfide and zinc oxide compared to an incidence of 100% following administration of nickel subsulfide alone (Kasprzak et al. 1988). Supplementing drinking water with zinc sulfate reduced the incidence of 9,10-dimethyl-1,2-benzanthracene-induced tumors in the cheek pouches of mice (Poswillo and Cohen 1971). Zinc decreased DNA synthesis in hepatomas induced by 3'-methyl-4-dimethylaminoazobenzene (Duncan and Dreosti 1975). The investigators speculated that the changes were due to inhibited cell division cycle at the level of DNA replication.

#### **3.10 POPULATIONS THAT ARE UNUSUALLY SUSCEPTIBLE**

A susceptible population will exhibit a different or enhanced response to zinc than will most persons exposed to the same level of zinc in the environment. Reasons may include genetic makeup, age, health and nutritional status, and exposure to other toxic substances (e.g., cigarette smoke). These parameters result in reduced detoxification or excretion of zinc, or compromised function of organs affected by zinc. Populations who are at greater risk due to their unusually high exposure to zinc are discussed in Section 6.7, Populations With Potentially High Exposures.

No specific data regarding human subpopulations that are unusually susceptible to the toxic effects of zinc were located. Healthy elderly people have been shown to have greater daily zinc intake than housebound elderly people (Bunker et al. 1987; Prasad 1988). Data from animal studies indicate that certain human subpopulations may be more susceptible to excess zinc because of zinc's depleting effect on copper (Underwood 1977). People who are malnourished or have a marginal copper status may be more susceptible to the effects of excessive zinc than people who are adequately nourished (Underwood 1977).

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Hepatic zinc levels are elevated in patients with hemochromatosis, a genetic disease associated with increased iron absorption from the intestine (Adams et al. 1991). The chronic iron loading that occurs could result in hepatic metallothionein induction leading to the accumulation of zinc because metallothionein has a greater affinity for zinc than iron. These individuals may, therefore, have a greater likelihood of developing toxicity with zinc exposure levels that do not normally result in any symptoms in the general population. However, available studies, including this one, have not correlated increased hepatic zinc with any adverse effects.

#### **3.11 METHODS FOR REDUCING TOXIC EFFECTS**

This section will describe clinical practice and research concerning methods for reducing toxic effects of exposure to zinc. However, because some of the treatments discussed may be experimental and unproven, this section should not be used as a guide for treatment of exposures to zinc. When specific exposures have occurred, poison control centers and medical toxicologists should be consulted for medical advice. The following texts provide specific information about treatment following exposures to zinc:

Ellenhorn MJ, Barceloux DG. 1988. Medical toxicology: Diagnosis and treatment of human poisoning. New York, NY: Elsevier, 879-880, 1064-1065.

##### **3.11.1 Reducing Peak Absorption Following Exposure**

General recommendations for the management and treatment of patients following acute exposure to zinc include removal of the victim from the contaminated area and removal and isolation of contaminated clothing, jewelry, and shoes (Bronstein and Currence 1988; Stutz and Janusz 1988). Excess contaminant is gently brushed away and excess liquids blotted with absorbent material. Measures that are appropriate to the route of exposure are taken to remove zinc from the body. Exposed eyes are flushed immediately with water, followed as soon as possible with irrigation of each eye with normal saline. Exposed skin is washed immediately with soapy water. Administration of ipecac to induce emesis, gastric lavage, ingestion of activated charcoal, and cathartics have been recommended to decrease the gastrointestinal absorption of zinc (Burkhart et al. 1990; Ellenhorn and Barceloux 1988). Because zinc causes nausea and vomiting following exposure by the oral route, use of emetic agents may be unnecessary. Ipecac administration may be contraindicated following ingestion of caustic zinc compounds such as zinc chloride. The large amounts of phosphorus and calcium in milk and cheese, and phytate in brown bread,



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may reduce absorption of zinc from the gastrointestinal tract (Pecoud et al. 1975). Therefore, if vomiting and diarrhea are not prohibitive, ingestion of dairy products or brown bread may also reduce gastrointestinal absorption of zinc. In a study of intestinal absorption of zinc in iron-deficient mice, the uptake of zinc from the gut was inhibited by adding iron to the duodenal loop system. The proposed mechanism was that iron and zinc shared a common gut mucosal binding site (Hamilton et al. 1978). However, it is unknown whether ingestion of iron supplements would be effective in reducing absorption of zinc overdoses.

#### 3.11.2 Reducing Body Burden

Zinc is an essential trace element that is normally found in tissues and fluids throughout the body and is under homeostatic control (NAS/NRC 1989b). Increased levels have been observed in the heart, spleen, kidneys, liver, bone, and blood of animals following subchronic oral exposure to zinc (Llobet et al. 1988a) indicating that some zinc accumulation occurs during excess intakes. The greatest increases were observed in bone and blood.

Administration of the chelating agent, calcium disodium ethylene diaminetetraacetate ( $\text{CaNa}_2\text{EDTA}$ ), is the treatment of choice for reducing the body burden of zinc in humans following exposure to high levels (Ellenhorn and Barceloux 1988). Ethylenediaminetetraacetic acid (EDTA), diethylenetriaminepentaacetic acid (DTPA), and dimercaprol (BAL) are the most common antidotes used in the treatment of human zinc intoxications (Llobet et al. 1989; Murphy 1970; Spencer and Rosoff 1966). Markedly elevated serum zinc levels in a young child who ingested a zinc chloride solution were normalized by intravenously administering a single small dose of  $\text{CaNa}_2\text{EDTA}$  (11.5 mg/kg) (Potter 1981). Use of chelation therapy (administration of BAL) was reported in a case study of a 16-year-old boy who ingested 12 g of metallic zinc (Murphy 1970). The boy exhibited lethargy and elevated blood zinc levels that were both reversed following intramuscular administration of BAL. Chelation therapy has been demonstrated to increase the urinary excretion of zinc 22-fold (Spencer and Rosoff 1966). Intravenous and nebulized *N*-acetylcysteine (another metal chelating agent) have also been observed to increase urinary zinc excretion and decrease plasma levels following inhalation of zinc chloride smoke (Hjortso et al. 1988).

The efficacy of 16 different chelating agents as possible antidotes for acute oral zinc exposure has been determined in mice (Llobet et al. 1988b). The most efficient chelators were DTPA, cyclohexanediamine-tetraacetic acid (CDTA), and EDTA. Increased urinary levels of zinc and decreased bone and liver zinc levels were observed following administration of the chelators. The maximum efficiency of the chelators

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was observed when they were administered from 10 minutes to 12 hours after zinc exposure (Domingo et al. 1988a, 1988b).

#### **3.11.3 Interfering with the Mechanism of Action for Toxic Effects**

Anemia has been observed in humans and animals after oral exposure to zinc. It has been postulated that excess zinc intake may result in copper deficiency (mechanisms of action are discussed in Section 3.5).

The anemia observed following zinc intake is believed to be caused by the copper deficiency.

Administration of copper in patients with zinc-related anemia has been shown to be effective in increasing the hemoglobin levels (Porter et al. 1977; Smith and Larson 1946).

The exact mechanism of metal fume fever (a syndrome consisting of a leukocytosis with chills, fever, cough, myalgias, headache, weakness, and dyspnea) is unknown (Ellenhorn and Barceloux 1988), but respiratory tract inflammation and the development of an immune complex reaction have been proposed (McCord 1960). Treatment is supportive (e.g., bed rest, analgesics, and antipyretics) (Mueller and Seger 1985).

In severe cases, inhalation of zinc chloride has resulted in advanced pulmonary fibrosis and fatal respiratory distress syndrome (Evans 1945; Hjortso et al. 1988; Milliken et al. 1963).

L-3,4-Dehydroproline was given to two soldiers after inhaling a high concentration of zinc chloride smoke (also contained other chemicals) in an attempt to inhibit collagen deposition in the lungs (Hjortso et al. 1988). This therapy did not prevent respiratory failure.

#### **3.12 ADEQUACY OF THE DATABASE**

Section 104(i)(5) of CERCLA, as amended, directs the Administrator of ATSDR (in consultation with the Administrator of EPA and agencies and programs of the Public Health Service) to assess whether adequate information on the health effects of zinc is available. Where adequate information is not available, ATSDR, in conjunction with the National Toxicology Program (NTP), is required to assure the initiation of a program of research designed to determine the health effects (and techniques for developing methods to determine such health effects) of zinc.

The following categories of possible data needs have been identified by a joint team of scientists from ATSDR, NTP, and EPA. They are defined as substance-specific informational needs that if met would

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reduce the uncertainties of human health assessment. This definition should not be interpreted to mean that all data needs discussed in this section must be filled. In the future, the identified data needs will be evaluated and prioritized, and a substance-specific research agenda will be proposed.

#### **3.12.1 Existing Information on Health Effects of Zinc**

The existing data on health effects of inhalation, oral, and dermal exposure of humans and animals to zinc are summarized in Figure 3-4. The purpose of this figure is to illustrate the existing information concerning the health effects of zinc. Each dot in the figure indicates that one or more studies provide information associated with that particular effect. The dot does not necessarily imply anything about the quality of the study or studies, nor should missing information in this figure be interpreted as a “data need”. A data need, as defined in ATSDR’s Decision Guide for Identifying Substance-Specific Data Needs Related to Toxicological Profiles (Agency for Toxic Substances and Disease Registry 1989), is substance-specific information necessary to conduct comprehensive public health assessments. Generally, ATSDR defines a data gap more broadly as any substance-specific information missing from the scientific literature.

Figure 3-4 indicates whether a particular health effect end point has been studied for a specific route and duration of exposure. There is little information concerning death in humans after inhalation, oral, or dermal exposure to zinc. However, several case studies report death after exposure to extremely high levels of zinc chloride and other components of zinc chloride smoke (Evans 1945; Hjortso et al. 1988; Milliken et al. 1963).

Systemic effects of acute inhalation exposure to generally unspecified levels of various zinc compounds in humans have been reported in several clinical case studies (Blanc et al. 1991; Brown 1988; Hjortso et al. 1988; Matarese and Matthews 1966; Vogelmeier et al. 1987). Case studies and experimental studies of systemic effects in humans following acute, intermediate, and chronic oral exposures are available (Anonymous 1983; Black et al. 1988; Brandao-Neto et al. 1990a; Chandra 1984; Chobanian 1981; Hale et al. 1988; Hallbook and Lanner 1972; Hoffman et al. 1988; Hooper et al. 1980; Malo et al. 1990; Moore 1978; Patterson et al. 1985; Porter et al. 1977; Potter 1981; Prasad et al. 1978). Experimental studies in humans following acute, intermediate, and chronic dermal exposures were located for hematological, dermal, and ocular effects (Agren 1990; Evans 1945; Fischer et al. 1984; Turner 1921; Yadrick et al. 1989).

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**Figure 3-4. Existing Information on Health Effects of Zinc**

	Systemic									
	Death	Acute	Intermediate	Chronic	Immunologic/Lymphoretic	Neurologic	Reproductive	Developmental	Genotoxic	Cancer
Inhalation	●	●		●	●	●			●	●
Oral	●	●	●	●	●	●	●			●
Dermal		●	●	●						

**Human**

	Systemic									
	Death	Acute	Intermediate	Chronic	Immunologic/Lymphoretic	Neurologic	Reproductive	Developmental	Genotoxic	Cancer
Inhalation	●	●	●		●		●		●	●
Oral	●	●	●	●	●	●	●	●	●	●
Dermal		●								

**Animal**

● Existing Studies

Information concerning respiratory effects of acute inhalation exposure to zinc in animals is available (Amdur et al. 1982; Drinker and Drinker 1928; Lam et al. 1982, 1988). One study (Marrs et al. 1988) was located regarding other systemic effects in animals following inhalation exposure to zinc for an intermediate exposure duration. Information regarding systemic effects of zinc following oral exposure in animals is available for acute, intermediate, and chronic exposure durations (Allen et al. 1983; Anderson and Danylchuk 1979; Aughey et al. 1977; Bentley and Grubb 1991; Domingo et al. 1988a; Drinker et al. 1927c; Jenkins and Hidioglou 1991; Katya-Katya et al. 1984; Klevay and Hyg 1973; Llobet et al. 1988a; Maita et al. 1981; Straube et al. 1980; Walters and Roe 1965). One acute dermal study evaluated dermal irritancy in animals (Lansdown 1991).

Immunological effects were reported in humans following inhalation exposure to zinc oxide (Blanc et al. 1991; Farrell 1987). Another study reported potential adverse immunological effects following oral exposure of humans (Chandra 1984). Clinical symptoms suggestive of neurological effects have been reported by humans following inhalation exposure (Rohrs 1957; Sturgis et al. 1927; Wilde 1975) or oral exposure (Anonymous 1983; Murphy 1970; Potter 1981) to zinc. There were studies that examined reproductive and developmental effects in women orally exposed to zinc during their pregnancies (Kynast and Saling 1986; Mahomed et al. 1989; Simmer et al. 1991).

One study examined immunological and reproductive effects in animals following inhalation exposure to zinc chloride (Marrs et al. 1988). Immunological and neurological end points were evaluated in animals following oral exposure to zinc (Bleavins et al. 1983; Kozik et al. 1980, 1981; Schiffer et al. 1991). Information regarding developmental and reproductive effects in animals after oral exposure to zinc is available (Cox et al. 1969; Ketcheson et al. 1969; Kinnamon 1963; Mulhern et al. 1986; Pal and Pal 1987; Schlicker and Cox 1968; Sutton and Nelson 1937). Studies regarding genotoxicity in animals after inhalation and oral exposures to zinc are limited (Gupta et al. 1991; Kowalska-Wochna et al. 1988; Voroshilin et al. 1978).

Epidemiological studies regarding carcinogenicity after inhalation and oral exposure to zinc are available (Logue et al. 1982; Neuberger and Hollowell 1982; Philipp et al. 1982; Stocks and Davies 1964); however, they were not well controlled and the data are of little significance. Studies are available regarding carcinogenicity in animals after inhalation and oral exposure to zinc (Marrs et al. 1988; Walters and Roe 1965). However, the studies have several deficiencies that limit their usefulness.

## 3. HEALTH EFFECTS

**3.12.2 Identification of Data Needs**

**Acute-Duration Exposure.** Symptoms of metal fume fever (headache, fever, leukocytosis, myalgias) have been observed in humans acutely exposed to airborne zinc oxide (Blanc et al. 1991; Brown 1988; Drinker et al. 1927b; Sturgis et al. 1927). Acute oral exposure to zinc has resulted in gastrointestinal disturbances (abdominal pain, nausea, vomiting, esophageal erosion), evidence of pancreatic damage (increased serum amylase and lipase levels), and decreased levels of serum cortisol in humans (Anonymous 1983; Brandao-Neto et al. 1990a; Chobanian 1981; Murphy 1970; Potter 1981). Acute dermal exposure to zinc oxide has not been shown to be irritating to human skin (Agren 1990). Toxic effects similar to those observed for metal fume fever have been observed in guinea pigs (Amdur et al. 1982; Lam et al. 1985). In addition to LD<sub>50</sub> data, only one reliable study assessed the acute oral toxicity of zinc compounds in animals. Pancreatic, gastrointestinal, and liver damage were observed in sheep (Allen et al. 1983). It is doubtful that sheep (ruminant animals) are an appropriate model for toxicity of orally administered zinc in humans. The dermal toxicity of several zinc compounds has been tested in rabbits, guinea pigs, and mice (Lansdown 1991). Zinc acetate, zinc chloride, and zinc sulfate have irritating properties. Skin irritation was not observed in rabbits, guinea pigs, or mice after zinc oxide paste application (Lansdown 1991).

The animal data (Amdur et al. 1982; Drinker and Drinker 1928; Lam et al. 1982, 1988) corroborate occupational exposure studies that indicate metal fume fever is an end point of concern. However, other possible targets of toxicity have not been examined. Thus, an acute inhalation MRL cannot be derived. A large amount of the human oral exposure data is in the form of case reports, and a great deal of uncertainty exists regarding the dose levels. The uncertainty about whether sheep are a good model for humans precludes using these data to derive an oral MRL for acute-duration exposure. Additional studies involving acute exposure to zinc compounds by all routes of exposure would be helpful to identify target organ and dose-response relationships. There are groups who may be exposed to zinc at hazardous waste sites for brief periods; therefore, this information is important.

**Intermediate-Duration Exposure.** Metal fume fever was observed in an individual exposed to zinc fumes and zinc powder for approximately 1 month (Malo et al. 1990). Anemia and decreased levels of HDL cholesterol have been observed in humans taking high doses of zinc supplements (Chandra 1984; Hoffman et al. 1988; Hooper et al. 1980). Intermediate-duration dermal exposure to zinc oxide dust has resulted in pustular lesions, but these lesions were attributed to clogging of the sebaceous glands resulting from poor hygiene (Turner 1921). Rats, mice, and guinea pigs exposed to smoke containing zinc chloride and other compounds had evidence of lung irritation (Marrs et al. 1988). No intermediate-duration

### 3. HEALTH EFFECTS

animal dermal studies were located. In animals that ingested zinc for an intermediate duration, anemia and kidney and pancreas damage were observed (Bentley and Grubb 1991; Drinker et al. 1927d; Jenkins and Hidioglou 1991; Llobet et al. 1988a; Maita et al. 1981; Straube et al. 1980).

Only one case report regarding human intermediate-duration inhalation exposure was located, and this study did not report the exposure level (Malo et al. 1990). Thus, an intermediate-duration inhalation MRL could not be derived. The less serious LOAELs (decreased serum HDL cholesterol) identified in the Hooper et al. (1980) and Chandra (1984) human oral exposure studies could be used as the basis of an intermediate-duration oral MRL; however, evidence regarding this effect is inconsistent (Bogden et al. 1988; Hale et al. 1988; Samman and Roberts 1988). In addition, the resulting MRL would be  $\approx$  10-fold lower than the RDA of 11 mg/day (0.21 mg zinc/kg/day). An intermediate-duration oral MRL was derived for zinc based on hematological effects (decreased hematocrit, serum ferritin, and erythrocyte superoxide dismutase) in women given zinc gluconate supplements for 10 weeks (Yadrick et al. 1989). The toxic effects of intermediate-duration exposure to zinc compounds are relatively well characterized for the oral route. There are insufficient toxicokinetic data to determine if the toxic effects observed following oral exposure would occur following inhalation or dermal exposure. Inhalation and dermal studies would be useful to determine possible target organs and dose-response relationships. There are populations surrounding hazardous waste sites that might be exposed to zinc compounds for similar durations.

**Chronic-Duration Exposure and Cancer.** No exposure-related effects on lung function were observed in a group of welders chronically exposed to zinc (Marquart et al. 1989). Anemia has been observed in humans following ingestion of high doses of zinc supplements (Broun et al. 1990; Hale et al. 1988; Porter et al. 1977; Prasad et al. 1978). Chronic-duration dermal exposure to zinc oxide dust has resulted in pustular lesions, but these were attributed to clogging of the sebaceous glands resulting from poor hygiene (Batchelor et al. 1926). No chronic-duration inhalation or dermal studies in animals were located. Pancreatic damage was observed in mice after chronic exposure to zinc sulfate in drinking water (Aughey et al. 1977).

A chronic-duration inhalation MRL could not be derived for zinc because neither of the inhalation studies reported the levels of airborne zinc. Due to a lack of adequate chronic-duration oral studies, the intermediate-duration oral MRL was adopted as the chronic-duration oral MRL, based on hematological effects (decreased hematocrit, serum ferritin, and erythrocyte dismutase) in women given zinc gluconate

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supplements for 10 weeks (Yadrick et al. 1989). Additional studies involving chronic exposure to zinc compounds by all routes of exposure would be helpful to identify dose-response relationships.

Although there are several human and animal carcinogenicity studies, the limitations of these studies preclude their use in assessing the carcinogenicity of zinc (Logue et al. 1982; Neuberger and Hollowell 1982; Walters and Roe 1965). Carcinogenicity studies by all routes of exposure would be useful.

**Genotoxicity.** Several *in vitro* microbial gene mutation assays were negative (Marzin and Vo Phi 1985; Nishioka 1975; Thompson et al. 1989; Venitt and Levy 1974; Wong 1988), but evidence from gene mutation assays in mammalian cells is mixed (Amacher and Paillet 1980; Thompson et al. 1989). An increase in the occurrence of chromosomal aberrations was observed *in vitro* in human lymphocytes (Deknudt and Deminatti 1978) and *in vivo* in rats and mice (Deknudt and Gerber 1979; Gupta et al. 1991; Kowalska-Wochna et al. 1988; Voroshilin et al. 1978). Increased sister chromatid exchange was observed *in vivo* in rat bone marrow (Kowalska-Wochna et al. 1988). However, while there are sufficient *in vivo* data establishing the clastogenicity of zinc, data regarding the mutagenicity of zinc are conflicting. Studies designed to assay different types of genotoxicity (i.e., mutagenicity in mammalian cells, effect of excess zinc on DNA replication) would be useful for determining the genotoxic potential of zinc.

**Reproductive Toxicity.** No complications occurred in the pregnancies of women exposed to daily doses of zinc sulfide during the last two trimesters (Mahomed et al. 1989). No studies were located regarding the reproductive toxicity of zinc in humans after inhalation or dermal exposure. Increased pre-implantation loss and reproductive dysfunction in rats were observed in oral exposure studies (Pal and Pal 1987; Sutton and Nelson 1937). No histological changes in reproductive organs were observed in rats, mice, or guinea pigs following inhalation exposure to zinc chloride smoke, but reproductive function was not assessed (Marrs et al. 1988). No dermal reproductive toxicity studies in animals were located. Inhalation and dermal studies assessing reproductive function would be useful to determine whether zinc has the potential to cause reproductive effects by these routes. An oral reproductive toxicity study in a different animal strain as well as a multigeneration study, including reproductive organ pathology, would be useful for determining whether oral zinc exposure is likely to cause reproductive toxicity in humans.

**Developmental Toxicity.** No studies were located regarding the potential of zinc to cause developmental effects in humans after inhalation or dermal exposure. In a very brief report of a human study in which pregnant women received high-doses of zinc supplements during the last trimester of pregnancy, an increased incidence of stillbirths and one premature delivery were observed (Kumar 1976).



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This study, however, has many limitations. Increased fetal resorptions were observed in rats after oral exposure to zinc (Schlicker and Cox 1968). No studies were located regarding developmental toxicity in animals after inhalation or dermal exposure to zinc. Additional inhalation, oral, and dermal exposure studies in animals would be useful to predict whether developmental effects should be a concern for humans exposed to zinc.

**Immunotoxicity.** Metal fume fever is believed to be an immune response to zinc oxide. A correlation between the concentration of airborne zinc and the number of all types of T cells (helper, inducer, suppressor, and killer) in the bronchoalveolar lavage fluid of humans, possibly related to the onset of metal fume fever, was observed in an acute-duration inhalation study (Blanc et al. 1991). Impaired immune response in humans has been reported in an intermediate-duration oral study (Chandra 1984). No immune effects were observed in mice after oral exposure to zinc (Schiffer et al. 1991). There is some limited information to suggest that the immune system is a target of zinc toxicity. A battery of immune function tests after inhalation, oral, and dermal exposure to zinc compounds would be useful in determining if zinc is immunotoxic.

**Neurotoxicity.** Staggering gait and hallucinations were reported in an individual who intentionally inhaled metallic paint aerosols (Wilde 1975). Because there was simultaneous exposure to copper and hydrocarbons, this study cannot be used to assess the neurotoxic potential of zinc. Nonspecific signs and symptoms of neurotoxicity (light-headedness, dizziness, headache, and lethargy) have been reported by humans following acute oral exposure to zinc (Murphy 1970; Potter 1981). Very limited data suggest that high oral doses of zinc can cause minor neuron degeneration and alteration of secretion of the hypothalamus in rats (Kozik et al. 1980, 1981). No studies were located regarding neurotoxic effects in animals after inhalation or dermal exposure to zinc. Additional studies by all routes of exposure would be useful to determine if exposure to zinc compounds would result in neurotoxicity.

**Epidemiological and Human Dosimetry Studies.** Acute high-level exposure to zinc by inhalation resulted in respiratory irritation and metal fume fever (Blanc et al. 1991; Hjortso et al. 1988; Johnson and Stonehill 1961; Linn et al. 1981; Schenker et al. 1981; Sturgis et al. 1927). Welders are a subpopulation of workers who have a high potential for exposure to zinc oxide. Most of the available studies did not report exposure levels or used a small number of subjects. Studies that correlate occupational exposure to zinc with health effects would be useful. A number of human oral exposure studies have shown that excess levels of zinc can result in anemia, pancreatic damage, decreased serum HDL cholesterol levels, and immunotoxicity (Black et al. 1988; Chandra 1984; Hooper et al. 1980).

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There are insufficient data for establishing dose-response relationships. Studies designed to establish dose-response relationships would be useful for establishing cause/effect relationships and future monitoring of individuals living near hazardous waste sites.

#### **Biomarkers of Exposure and Effect.**

*Exposure.* Increased serum and urine levels of zinc were observed in humans and animals after inhalation, oral, or dermal exposure to zinc (Bentley and Grubb 1991; Brandao-Neto et al. 1990b; Hallmans 1977; Keen and Hurley 1977). However, the relationships between zinc exposure levels and the levels of zinc in biological fluids have not been established. Hair and nail samples may be a potential biomarker for long-term zinc exposure (McBean et al. 1971; Rivlin 1983; Wilhelm et al. 1991); however, no correlation has been demonstrated between these parameters and zinc exposure levels. Development of a biomarker with more exposure and dose data would aid in future medical surveillance that could lead to better detection of zinc exposure.

*Effect.* Several potential biomarkers for the effects of zinc have been identified. These include increased levels of serum amylases and lipase, indicative of pancreatic damage; non-iron responsive anemia; and decreased HDL cholesterol levels (Suber 1989). However, these biomarkers of effect are not specific for zinc. These biomarkers cannot be used for dosimetry. A potential biomarker of exposure for recent exposures to zinc is increased erythrocyte metallothionein concentrations (Grider et al. 1990). Further investigation of serum biomarkers of effect, particularly for chronic exposure, in zinc-exposed populations would be useful to determine whether exposed populations may be experiencing adverse health effects as the result of zinc exposures.

**Absorption, Distribution, Metabolism, and Excretion.** Absorption of zinc in humans after oral exposure to high levels has been well described (Aamodt et al. 1983; Hunt et al. 1991; Reinhold et al. 1991; Sandstrom and Abrahamson 1989; Sandstrom and Cederblad 1980; Sandstrom and Sandberg 1992; Spencer et al. 1985). However, quantitative evidence of zinc absorption in humans after inhalation or dermal exposure is very limited. It is known that workers exposed to zinc oxide fumes who experience toxic effects have elevated levels of zinc in plasma and urine (Hamdi 1969). However, it remains to be established whether the elevated levels are the result of the pulmonary absorption or of the swallowing of particles leading to gastrointestinal absorption. Toxic effects have also been observed in humans after dermal exposure (DuBray 1937), indicating dermal absorption.

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Information regarding the absorption of zinc in animals following inhalation exposure was limited to lung retention data (Gordon et al. 1992; Hirano et al. 1989). However, there was information to assess the extent of absorption following oral exposure (Davies 1980; Galvez-Morros et al. 1992; Johnson et al. 1988; Weigand and Kirchgessner 1992). Evidence is limited regarding dermal absorption in animals, but it indicates that zinc sulfate and zinc oxide can penetrate the skin (Agren 1990; Agren et al. 1991; Gordon et al. 1981; Hallmans 1977). Mechanistic data on the oral absorption is reported by Hempe and Cousins (1992); however, there is a lack of information regarding the mechanism of action of inhalation and dermal exposures.

Information on physiological levels and zinc distribution following subtoxic short-term exposures to zinc in humans and animals is abundant (NAS/NRC 1979; Wastney et al. 1986). Blood levels of zinc have been determined in humans following oral exposure to zinc sulfate (Neve et al. 1991; Statter et al. 1988; Sturniolo et al. 1991). Increased zinc tissue content has been seen after short-term oral exposure in humans (Cooke et al. 1990; He et al. 1991; Llobet et al. 1988a; Schiffer et al. 1991; Weigand and Kirchgessner 1992). Studies on tissue distribution in humans following high exposure to zinc for inhalation, oral, and dermal would be useful. There were no studies regarding blood or tissue distribution after acute, high-level exposures to zinc in animals following inhalation or dermal exposure. Additional mechanistic data on the transfer of zinc from respiratory and dermal absorption sites to the blood would be useful.

The principal excretion route of ingested zinc is through the intestines (Davies and Nightingale 1975; Reinhold et al. 1991; Wastney et al. 1986). There is a lack of information regarding the excretion of zinc in both animals and humans following inhalation and dermal exposure.

Therefore, additional studies designed to assess the toxicokinetic properties of zinc following inhalation and dermal exposures would be useful.

**Comparative Toxicokinetics.** Data suggest that humans and animals have similar target organs of zinc toxicity (Allen et al. 1983; Aughey et al. 1977; Black et al. 1988; Blanc et al. 1991; Brown 1988; Chandra 1984; Chobanian 1981; Drinker et al. 1927b, 1927d; Hoffman et al. 1988; Hooper et al. 1980; Katya-Katya et al. 1984; Klevay and Hyg 1973; Lam et al. 1982, 1985, 1988; Maita et al. 1981; Moore 1978; Murphy 1970; Smith and Larson 1946; Straube et al. 1980; Sturgis et al. 1927). Toxicokinetic studies have been performed in both humans and animals following oral exposure; however, data are limited for inhalation and dermal exposures. The animal model used most often to evaluate the

### 3. HEALTH EFFECTS

toxicokinetics of zinc are rats (Agren et al. 1991; Alexander et al. 1981; Galvez-Morros et al. 1992; Hirano et al. 1989; Llobet et al. 1988a; Weigand and Kirchgessner 1992) and may be a good model for assessing the kinetics of zinc in humans.

**Methods for Reducing Toxic Effects.** No established methods or treatments for reducing the absorption of zinc were located. Studies that examined the effectiveness of emetics and cathartics in the prevention of zinc absorption would be useful. Once absorbed from the gastrointestinal tract, zinc bound to plasma albumin is distributed to the rest of the body. Zinc has a high affinity for proteins, and a number of chelating agents are effective in increasing urinary excretion of zinc following acute- and intermediate-duration administrations (Domingo et al. 1988a, 1988b; Llobet et al. 1989). Studies designed to examine the effectiveness of chelating agents following chronic zinc exposure would be useful in determining treatments to reduce the zinc body burden. Very little information is known about the absorption and distribution of zinc following inhalation or dermal exposure. Studies to determine the mechanisms of absorption and distribution would be useful for developing treatments or methods for reducing the toxic effects of zinc after inhalation or dermal exposure.

Although the exact mechanisms of many of the toxic actions of zinc are not known, the pathogenesis of metal fume fever following inhalation exposure (McCord 1960; Mueller and Seger 1985) and anemia following oral exposure (Prasad et al. 1978) are known. Studies to more clearly elucidate the mechanisms involved in metal fume fever and anemia and to determine the mechanisms involved in pancreatic damage and decreased HDL cholesterol levels would be useful. Therapy for metal fume fever is mainly supportive (Mueller and Seger 1985). Administration of copper has been shown to be effective in alleviating zinc-induced anemia (Porter et al. 1977). Research into methods useful for mitigating metal fume fever and other adverse effects of zinc would be helpful.

**Children's Susceptibility.** While a considerable amount of data are available on the effects of zinc deficiency on the growth and development of children, less is known about the effects of excess zinc on children. Accidental acute oral exposures result in mainly gastrointestinal symptoms, including nausea, vomiting, and epigastric discomfort (Anonymous 1983; Lewis and Kokan 1998; Moore 1978; Murphy 1970). Data are not presently available to determine whether children are more susceptible to these effects than adults. Similarly, additional animal studies examining the effects of similar exposure on young and mature animals would be useful to further clarify possible mechanisms of childhood susceptibility, if it exists.

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Child health data needs relating to exposure are discussed in 6.8.1 Identification of Data Needs: Exposures of Children.

#### **3.12.3 Ongoing Studies**

A selection of ongoing studies, located in the Federal Research in Progress database (FEDRIP 2003), is presented in Table 3-6.

**Table 3-6. Ongoing Studies on Zinc Health Effects<sup>a</sup>**

Investigator	Institute	Research area
Abrams, SA	Baylor College of Medicine, Houston, Texas	Zinc metabolism in health and chronic inflammatory bowel disease children
Black, MM	University of Maryland, Baltimore, Maryland	Effect of micronutrient supplementation on children's growth, immune functioning, and morbidity
Blanchard, K	Department of Veterans Affairs, Medical Center, Shreveport, Louisiana	Efficacy and safety of oral zinc therapy in patients with Polycythemia Vera
Blumenthal, SS	Department of Veterans Affairs, Medical Center, Milwaukee, Wisconsin	Cadmium, zinc, metallothionein, and kidney cytotoxicity
Bobilya, DJ	University of New Hampshire, Durham, New Hampshire	Evaluation of zinc transport by using an <i>in vitro</i> model of the blood brain barrier under different conditions of zinc status
Bobilya, DJ	University of New Hampshire, Durham, New Hampshire	Testing to determine whether co-transport with albumin is a significant route for zinc transport by endothelial cells
Brewer, GJ	University of Michigan at Ann Arbor, Ann Arbor, Michigan	Studies on the treatment of Wilson's disease with zinc
Brown, KH	University of California, Nutrition, Davis, California	Bioavailability of vitamin A and zinc from selected foods of potential use for intervention programs in populations at high risk of deficiency
Brown, KH	University of California, Nutrition, Davis, California	Determination of the safety and efficacy of three levels of zinc supplementation, provided with or without supplemental copper
Brown, NM	Northwestern University, Evanston, Illinois	Combination of fluorescent microscopy studies with biophysical and proteomic approaches to identify zinc rich cellular compartments and isolate the proteins associated with these vesicles
Choi, DW	Washington University, St. Louis, Missouri	Zinc and ischemic brain injury
Choi, DW	Washington University, St. Louis, Missouri	Study of Zn <sup>2+</sup> -mediated neurotoxicity
Cline, TR	Purdue University, Animal Science, West Lafayette, Indiana	Measurement of the effects of fasting, diet particle size and elevated levels of zinc on growth and stomach morphology in young pigs
Disilvestro, RA	Ohio State University, College of Human Ecology, Columbus, Ohio	Determine whether stress-induced accumulation of certain radicals is affected by copper and zinc consumption in rats
Disilvestro, RA	Ohio State University, College of Human Ecology, Columbus, Ohio	Zinc supplementation in Crohn's disease patients
Fraker, PJ	Michigan State University, East Lansing, Michigan	Identification of the underlying mechanisms that cause the lymphopenia and reduced host defense that accompanies zinc deficiency in humans and animals

## 3. HEALTH EFFECTS

**Table 3-6. Ongoing Studies on Zinc Health Effects<sup>a</sup>**

Investigator	Institute	Research area
Freake, HC	University of Connecticut, Nutritional Sciences, Storrs, Connecticut	Effects of zinc on nuclear actions of thyroid hormone
Griffiths, JK	Tufts University Boston, Boston, Massachusetts	Examination of how vitamin A and zinc supplementation interact in improving immunity, fostering growth, and preventing infection, in populations at risk for malnutrition and vitamin A and zinc deficiency
Guo, MG	University of Vermont, Nutritional Sciences, Burlington, Vermont	Determination of whether the solubility of minerals added as organic salts of Zn, Fe, and Cu is greater than that of formulae prepared using inorganic ones
Hennig, B	University of Kentucky, Animal Science, Lexington, Kentucky	Examination of the antiatherogenic properties of zinc
Hennig, B	University of Kentucky, Animal Science, Lexington, Kentucky	Interference of zinc with the generation of an oxidative environment mediated by fatty acids
Johnson, MA	University of Georgia, College of Family and Consumer Science, Athens, Georgia	Examination of the influence of supplements of copper, zinc, and/or manganese on indices of bone formation and bone resorption in postmenopausal women
Keen, CL	University of California Davis, Davis, California	Examination of potential mechanisms by which maternal and embryonic zinc deficiency arise, and how this deficiency results in abnormal development and growth
King, LM	ARS, Germplasm and Physiology Lab, Beltsville, Maryland	Mechanisms of zinc and calcium regulation of sperm storage in the turkey
Lee, J-M	Washington University, St. Louis, Missouri	Role of zinc in focal ischemic brain injury
Lei, DK	University of Maryland, Human Nutrition and Food Science, College Park, Maryland	Modulation of p53 human tumor suppressor gene expression by zinc status
MacDonald, RS	University of Missouri, Food Science and Engineering, Columbia, Missouri	Examination of the cellular and molecular mechanisms that become limiting in humans and animals when they are deprived of the essential nutrient zinc
Mody, I	University of California Los Angeles, Los Angeles, California	Pathological consequence of the plastic conversion of zinc (Zn <sup>2+</sup> )-insensitive synaptic GABA/A receptors into Zn <sup>2+</sup> -sensitive ones
Moser-Veillon, PB	University of Maryland, Nutrition and Food Science, College Park, Maryland	Zinc needs and homeostasis during lactation
Onstad, CA	Agricultural Research Service, Houston, Texas	Assessment of the effects of low zinc intake compared with a zinc intake consistent with the RDA on zinc absorption and kinetics in 9–13-year-old girls

## 3. HEALTH EFFECTS

**Table 3-6. Ongoing Studies on Zinc Health Effects<sup>a</sup>**

Investigator	Institute	Research area
Onstad, CA	Agricultural Research Service, Houston, Texas	Measurement of the content of Ca, Mg, Fe, and Zn in existing germ-plasm of selected food crops to characterize genetic diversity
Panemangalore, M	Kentucky State University, Human Nutrition Research Program, Frankfort, Kentucky	Evaluation of the use of prophyrin profiles, ceruloplasmin, superoxide dismutase in serum or blood cells as biomarkers of zinc and copper status in humans and animals
Reeves, PG	Agricultural Research Service, Grand Forks, North Dakota	Studies to determine the correlation between sperm motility and heavy metals in semen, blood, urine, plasma, and saliva
Sazawal, S	Johns Hopkins University, Baltimore, Maryland	Role of zinc in childhood growth and development and the effects of zinc deficiency on childhood morbidity
Spears, JW	North Carolina State University, Animal Science, Raleigh, North Carolina	Determination of the effect of dietary level and source of zinc and copper on growth, reproduction, mineral status, and mineral excretion during the productive life span of female swine
Tankanow, RM	University of Michigan at Ann Arbor, Ann Arbor, Michigan	Zinc gluconate glycine lozenges and vitamin c effects on common cold
Thompson, RB	University of Maryland, Baltimore, Maryland	Development of a group of optical probes for studying zinc in neural tissue by fluorescence microscopy
Tielsch, JM	Johns Hopkins University, Baltimore, Maryland	Examination of the role of micronutrient deficiency on the health and well-being of women and children in underdeveloped areas of the world
Wagner, GJ	University of Kentucky, Agronomy, Lexington, Kentucky	Study of the mechanisms for vacuolar storage/sequestration of Cd, Zn, Mn, and Ni
Weiss, JH	University of California Irvine, Irvine, California	Ca <sup>2+</sup> , Zn <sup>2+</sup> , and selective excitotoxic neurodegeneration

<sup>a</sup>Source: FEDRIP 2003

FEDRIP = Federal Research in Progress





## 4. CHEMICAL AND PHYSICAL INFORMATION

### 4.1 CHEMICAL IDENTITY

Information concerning the chemical identity of elemental zinc and zinc compounds is listed in Table 4-1.

Zinc is a naturally occurring element found in the earth's surface rocks. Because of its reactivity, zinc metal is not found as the free element in nature. There are approximately 55 mineralized forms of zinc. The most important zinc minerals in the world are sphalerite ( $ZnS$ ), smithsonite ( $ZnCO_3$ ), and hemimorphite ( $Zn_4Si_2O_7(OH_2)H_2O$ ). Zinc appears in Group IIB of the periodic table and has two common oxidation states,  $Zn(0)$  and  $Zn(+2)$ . Zinc forms a variety of different compounds, such as zinc chloride, zinc oxide, and zinc sulfate (Goodwin 1998; Ohnesorge and Wilhelm 1991; WHO 2001).

### 4.2 PHYSICAL AND CHEMICAL PROPERTIES

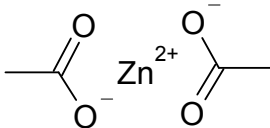
Information regarding the physical and chemical properties of elemental zinc and zinc compounds is located in Table 4-2.

Zinc is a lustrous, blue-white metal that burns in air with a bluish-green flame. It is stable in dry air, but upon exposure to moist air, it becomes covered with a film of zinc oxide or basic carbonate (e.g.,  $2ZnCO_3 \cdot 3Zn(OH)_2$ ) isolating the underlying metal and retarding further corrosion. Consistent with being high on the electromotive series, zinc forms quite stable compounds and, as such, resembles magnesium. Bonding in zinc compounds tends to be covalent, as in the sulfide and oxide (Goodwin 1998). In solution, four to six ligands can be coordinated with the zinc ion. Zinc has a strong tendency to react with acidic, alkaline, and inorganic compounds. Since zinc is amphoteric (i.e., capable of reacting chemically either as an acid or a base), it also forms zincates (e.g.,  $[Zn(OH)_3H_2O]^-$  and  $[Zn(OH)_4]^{2-}$ ) (Goodwin 1998; Ohnesorge and Wilhelm 1991; WHO 2001).

In humans and animals, zinc is an essential nutrient that plays a role in membrane stability, in over 300 enzymes, and in the metabolism of proteins and nucleic acids (WHO 2001).

## 4. CHEMICAL AND PHYSICAL INFORMATION

**Table 4-1. Chemical Identity of Zinc and Selected Compounds<sup>a</sup>**

Characteristic	Zinc	Zinc acetate	Zinc chloride
Synonyms	Zinc dust; zinc powder	Acetic acid, zinc salt; acetic acid, zinc(II) salt; dicarbomethoxyzinc; octan zinecnaty [Czech]; zinc diacetate; zinc(II) acetate	Butter of zinc; chlorure de zinc (French); zinc (Chlorure de) (French); zinc butter; zinc chloride (ZnCl <sub>2</sub> ); zinc dichloride; zinco (cloruro di) (Italian); zinkchlorid (German); zinkchloride (Dutch)
Registered trade name(s)	Asarco; L 15; Blue powder; CI 77945; CI pigment Metal 6; Emanay zinc dust; Granular zinc; JASAD; Merrillite; PASCO	No data	Tinning flux (DOT) <sup>b</sup> ; AI3-0440; Zintrace
Chemical formula	Zn	Zn(C <sub>2</sub> H <sub>3</sub> O <sub>2</sub> ) <sub>2</sub>	ZnCl <sub>2</sub>
Chemical structure	Zn		Cl-Zn-Cl
Identification numbers:			
CAS registry	7440-66-6	557-34-6 (anhydrous) 5970-45-6 (dihydrate)	7646-85-7
NIOSH RTECS	ZG8600000	AK1500000 (anhydrous) ZG8750000 (dihydrate)	ZH1400000
EPA hazardous waste	No data	No data	No data
OHM/TADS	7216955	No data	7216957
DOT/UN/NA/IMCO shipping	Zinc, powder or dust, UN 1436; zinc, powder or dust, zinc ashes, IMO4.3; zinc ashes, UN 1435	Zinc acetate, environmental hazardous substance, solid, NOS, UN 3077	Zinc chloride, anhydrous, UN 2331; zinc chloride, solution, UN 1840; zinc chloride, anhydrous, solution, IMO 8.3
HSDB	1344	1043	1050
NCI	No data	No data	No data

## 4. CHEMICAL AND PHYSICAL INFORMATION

**Table 4-1. Chemical Identity of Zinc and Selected Compounds<sup>a</sup>**

Characteristic	Zinc chromate	Zinc cyanide
Synonyms	Basic zinc chromate; chromic acid, zinc salt(1:1); chromic acid, zinc salt; chromium zinc oxide; zinc chrome yellow; zinc chromate; zinc chromate AM; zinc chromate C; zinc chromate O; zinc chromate Z; zinc chromate(VI) hydroxide; zinc chrome; zinc chrome (anti-corrosion); zinc chromium oxide; zinc hydroxychromate; zinc tetraoxychromate	Cyanure de zinc (French); zinc dicyanide
Registered trade name(s)	Pigment yellow 36; buttercup yellow; zinc tetraoxychromate 76A; zinc tetraoxychromate 780B; zinc yellow; ZTO; zincro ZTO	No data
Chemical formula	ZnCrO <sub>4</sub>	Zn(CN) <sub>2</sub>
Chemical structure		
Identification numbers:		
CAS registry	13530-65-9	557-21-1
NIOSH RTECS	GB3290000	ZH1575000
EPA hazardous waste	No data	P121; an acute hazardous waste when a discarded commercial chemical product or manufacturing chemical intermediate or an off-specification commercial chemical product or a manufacturing chemical intermediate. D003; a waste containing zinc cyanide may (or may not) be characterized a hazardous waste following testing for the reactivity characteristics as prescribed by RCRA regulations
OHM/TADS	No data	No data
DOT/UN/NA/IMCO shipping	No data	Zinc cyanide, UN 1713; Zinc cyanide, IMO 6.1
HSDB	6188	1051
NCI	77955	No data

## 4. CHEMICAL AND PHYSICAL INFORMATION

**Table 4-1. Chemical Identity of Zinc and Selected Compounds<sup>a</sup>**

Characteristic	Zinc hydroxide	Zinc oxide
Synonyms	Zinc dihydroxide	Zinc monoxide; zincum oxydatum; zinci oxydum; zinci oxicum; cynku tlenek (Polish)
Registered trade name(s)	No data	Actox 14; Actox 16; Actox 216; Al3-00277; Akro-Zinc Bar85 <sup>b</sup> ; Amalox; Amaloz; Azo 22; Azodox; Blanc de Zinc; Cadox XX 78; Caswell No 920; Chinese White; CI 77947; CI Pigment White 4; Electroxx 2500; Emanay Zinc Oxide; Emar; Felling Zinc Oxide; Flores de Zinci; Flowers of Zinc; GIAP 10; Green Seal-8; Hubbuck's White; Kadox 15; Kadox-25; Kadox 72; Outmine; Ozide; Ozlo; Permanent White; Philosopher's Wool; Powder Base 900; Protox 166; Protox 168; Protox 169; Protox Type 166; Protox Type 167; Protox Type 168; Protox Type 169; Protox Type 267; Protox Type 268; Red Seal; Red seal-9; Snow White; Unichem ZO; Vandem VAC; Vandem VOC; Vandem VPC; C-Weiss 8 (German); White Seal-7; XX 78; XX 203; XX 601; Zinca 20; Zinc White; Zincoid; Zn 0701T; Calamine <sup>b</sup> ; Zincite <sup>b</sup>
Chemical formula	Zn(OH) <sub>2</sub>	ZnO
Chemical structure	HO—Zn—OH	Zn=O
Identification numbers:		
CAS registry	20427-58-1	1314-13-2
NIOSH RTECS	ZH3853000	ZH4810000
EPA hazardous waste	No data	No data
OHM/TADS	No data	No data
DOT/UN/NA/IMCO shipping	No data	No data
HSDB	No data	5024
NCI	No data	No data

## 4. CHEMICAL AND PHYSICAL INFORMATION

**Table 4-1. Chemical Identity of Zinc and Selected Compounds<sup>a</sup>**

Characteristic	Zinc phosphate	Zinc sulfate
Synonyms	Zinc ortho-phosphate; neutral zinc phosphate; tribasic zinc phosphate; trizinc diphosphate; zinc acid phosphate; zinc phosphate (3:2)	Sulfate de zinc (French); sulfuric acid zinc salt; sulfuric acid, zinc salt (1:1); white copperas; white vitriol; zinc sulfate; zinc vitriol; zinci sulfas; zincum sulfuricum
Registered trade name(s)	Bonderite 181; Bonderite 40; Bonderite 880; C.I. Pigment White 32; Delaphos; Delaphos 2M; Fleck's Extraordinary; Fleck's Extraordinary cement; Granodine 16NC; Granodine 80; Heucophos ZP 10; LF Bowsei PW 2; Man-Gill 51339; Man-Gill 51355; Microphos 90; Phoshinox PZ 06; Pigment White 32; Sicor ZNP/M; ZPF; Sicor ZNP/S; Virchem 931; Weather coat 1000; ZP-DL; ZP-SB	Bonazen <sup>b</sup> ; Medizinc; Bufopto Zinc sulfate; Op-thal-zin; Opraex; Solvenzink; Verazinc; zincate; Zincomed; Zinkosite; AI3-03967; Orazinc; Zinc-200; Zinklet; Neozin; Optised; Prefrin-Z; Visine-AC; Zincfrin; Zink-Gro
Chemical formula	Zn <sub>3</sub> (PO <sub>4</sub> ) <sub>2</sub>	ZnSO <sub>4</sub> <sup>c</sup>
Chemical structure		
Identification numbers:		
CAS registry	7779-90-0	7733-02-0
NIOSH RTECS	TD0590000	ZH5260000
EPA hazardous waste	No data	No data
OHM/TADS	No data	7216958
DOT/UN/NA/IMCO shipping	No data	NA 9161
HSDB	No data	1063
NCI	No data	No data

## 4. CHEMICAL AND PHYSICAL INFORMATION

**Table 4-1. Chemical Identity of Zinc and Selected Compounds<sup>a</sup>**

Characterstic	Zinc sulfide
Synonyms	Wurtzite (alpha) <sup>b</sup> ; sphalerite (beta) <sup>b</sup> ; zinc monosulfide; zinc blende; zinc sulphide
Registered trade name(s)	Albalith; Irtran Z; Irtran 2; CI Pigment White 7; Sachtolith; Sachtolith HD-S; Cleartran
Chemical formula	ZnS
Chemical structure	Zn=S
Identification numbers:	
CAS registry	1314-98-3
NIOSH RTECS	ZH5400000
EPA hazardous waste	D003
OHM/TADS	No data
DOT/UN/NA/IMCO shipping	UN 3077; Zinc sulfide, environmentally hazardous substance, solid, NO; UN 3082; zinc sulfide, environmentally hazardous substance, liquid, NOS
HSDB	5802
NCI	No data

<sup>a</sup>Unless otherwise specified, all data from Chemfinder 2003; ChemID 2003; HSDB 2003; NIOSH 1990; and RTECS 2003

<sup>b</sup>HSDB 1990

<sup>c</sup>O'Neil et al. 2001

CAS = Chemical Abstracts Service; DOT/UN/NA/IMCO = Department of Transportation/United Nations/North America/International Maritime Dangerous Goods Code; EPA = Environmental Protection Agency; HSDB = Hazardous Substances Data Bank; NCI = National Cancer Institute; NIOSH = National Institute for Occupational Safety and Health; OHM/TADS = Oil and Hazardous Materials/Technical Assistance Data System; RCRA=Resource Conservation and Recovery Act; RTECS = Registry of Toxic Effects of Chemical Substances

## 4. CHEMICAL AND PHYSICAL INFORMATION

**Table 4-2. Physical and Chemical Properties of Zinc and Selected Compounds<sup>a</sup>**

Property	Zinc	Zinc acetate	Zinc chloride
Molecular weight	65.38	183.48	136.29
Color	Bluish-white, lustrous	White granules	White granules <sup>b</sup>
Physical state	Solid metal	Solid	Solid
Melting point	419.5 °C	237 °C (decomposes)	290 °C
Boiling point	908 °C	No applicable	732 °C
Density (g/cm <sup>3</sup> )	7.14 at 25 °C	1.735	2.907 at 25 °C
Odor	No data	Faint acetous odor <sup>c</sup>	Odorless; fume has acrid odor <sup>c</sup>
Odor threshold:			
Water	No data	No data	No data
Air	No data	No data	No data
Solubility:			
Water	Insoluble <sup>d</sup>	4.0x10 <sup>4</sup> mg/L at 25 °C; 6.7x10 <sup>4</sup> mg/L at 100 °C <sup>c</sup>	4.32x10 <sup>6</sup> mg/L at 25 °C; 6.14x10 <sup>6</sup> mg/L at 100 °C <sup>c</sup>
Other solvent(s)	Soluble in acetic acid and alkali	33 mg/L in alcohol	1 g/1.3 mL alcohol; 1 g/2 mL glyderol; 1 g/0.25 mL 2% hydro-chloroacetic acid
Partition coefficients:			
K <sub>d</sub> (mL/g)	0.1–8,000 <sup>e</sup> ; 40 (average) <sup>f</sup> ; 39 in sandy loam soil; 12.2 in sandy soil <sup>g</sup>	No data	No data
K <sub>ow</sub>	No data	No data	No data
K <sub>oc</sub>	No data	No data	No data
Vapor pressure	1 mm Hg at 487 °C	Not data	Not data
Henry's law constant	Not applicable	Not applicable	Not applicable
Autoignition temperature	No data	No data	Not flammable <sup>h</sup>
Flashpoint	No data	No data	Not flammable <sup>h</sup>
Flammability limits	No data	No data	Not flammable <sup>h</sup>
Conversion factor	Not applicable	mg Zn(C <sub>2</sub> H <sub>3</sub> O <sub>2</sub> ) <sub>2</sub> x 0.36 = mg Zn	mg ZnSO <sub>4</sub> x 0.40 = mg Zn
Explosive limits	No data	No data	No data



## 4. CHEMICAL AND PHYSICAL INFORMATION

**Table 4-2. Physical and Chemical Properties of Zinc and Selected Compounds<sup>a</sup>**

Property	Zinc chromate	Zinc cyanide	Zinc hydroxide
Molecular weight	181.37	117.42	99.40
Color	Lemon-yellow	White <sup>c</sup>	Colorless <sup>c</sup>
Physical state	Solid	Powder <sup>c</sup>	Solid <sup>c</sup>
Melting point	No data	800 °C (decomposes) <sup>c</sup>	Decomposes at 125 °C <sup>c</sup>
Boiling point	No data	Not applicable	Not applicable
Density (g/cm <sup>3</sup> )	3.40	1.852 <sup>c</sup>	3.053 <sup>c</sup>
Odor	Odorless	No data	No data
Odor threshold:			
Water	No data	No data	No data
Air	No data	No data	No data
Solubility:			
Water	Insoluble in cold water; sparingly soluble	Insoluble <sup>c</sup> ; 50 mg/L at 20 °C <sup>c</sup>	Almost insoluble <sup>c</sup>
Other solvent(s)	Soluble in acids, liquid ammonia; insoluble in acetone	Soluble in dilute mineral acids	No data
Partition coefficients:			
K <sub>d</sub> (mL/g)	No data	No data	No data
K <sub>ow</sub>	No data	No data	No data
K <sub>oc</sub>	No data	No data	No data
Vapor pressure	No data	No data	No data
Henry's law constant	Not applicable	Not applicable	Not applicable
Autoignition temperature	No data	No data	No data
Flashpoint	No data	No data	No data
Flammability limits	No data	No data	No data
Conversion factor	mg ZnCrO <sub>4</sub> x 0.36 = mg Zn	mg Zn(CN) <sub>2</sub> x 0.56 = mg Zn	mg Zn(OH) <sub>2</sub> x 0.66 = mg Zn
Explosive limits	No data	No data	No data

## 4. CHEMICAL AND PHYSICAL INFORMATION

**Table 4-2. Physical and Chemical Properties of Zinc and Selected Compounds<sup>a</sup>**

Property	Zinc oxide	Zinc phosphate	Zinc sulfate
Molecular weight	81.38	386.11	161.44
Color	White/yellowish-white	White <sup>c</sup>	Colorless <sup>i</sup>
Physical state	Solid	Powder <sup>c</sup>	Solid
Melting point	1975 °C	900 °C <sup>c</sup>	680 °C (decomposes)
Boiling point	Sublimes	No data	No applicable
Density (g/cm <sup>3</sup> )	5.607 at 20 °C	3.998 at 15 °C <sup>c</sup>	3.54 at 25 °C
Odor	Odorless	Odorless	Not determined
Odor threshold:			
Water	No data	No data	No data
Air	No data	No data	No data
Solubility:			
Water	1.6 mg/L at 29 °C <sup>i</sup>	Insoluble <sup>j</sup>	Soluble in cold and hot water <sup>i</sup> ; 4.19x10 <sup>5</sup> mg/L at 0 °C; 9.1x10 <sup>5</sup> mg/L at 70 °C
Other solvent(s)	Soluble in dilute acetic or mineral acids, ammonia, ammonium carbonate, fixed alkali hydroxide solution, and ammonium chloride <sup>i</sup> ; insoluble in alcohol <sup>i</sup>	Soluble in dilute mineral acids, ammonium hydroxide and alkali hydroxide solutions; insoluble in alcohol	Slightly soluble in alcohol; soluble in methanol and glycerol <sup>i</sup> ; 1 g/2.5 mL glycerol
Partition coefficients:			
K <sub>d</sub>	No data	No data	No data
K <sub>ow</sub>	No data	No data	No data
K <sub>oc</sub>	No data	No data	No data
Vapor pressure	Not data	No data	No data
Henry's law constant	Not applicable	No data	Not applicable
Autoignition temperature	Not flammable <sup>h</sup>	No data	Not flammable <sup>h</sup>
Flashpoint	Not flammable <sup>h</sup>	No data	Not flammable <sup>h</sup>
Flammability limits	Not flammable <sup>h</sup>	No data	Not flammable <sup>h</sup>
Conversion factor	mg ZnO x 0.80 = mg Zn	mg Zn <sub>3</sub> (PO <sub>4</sub> ) <sub>2</sub> x 0.51 = mg Zn	mg ZnSO <sub>4</sub> x 0.40 = mg Zn
Explosive limits	No data	No data	No data

## 4. CHEMICAL AND PHYSICAL INFORMATION

**Table 4-2. Physical and Chemical Properties of Zinc and Selected Compounds<sup>a</sup>**

Property	Zinc sulfide (α)	Zinc sulfide (γ)
Molecular weight	97.45	97.45
Color	Colorless <sup>i</sup>	Colorless
Physical state	Solid	Solid
Melting point	1,700±20 °C	No data
Boiling point	1,185 °C at 1 atm	1,185 °C at 1 atm
Density (g/cm <sup>3</sup> )	3.98 at 20 °C <sup>i</sup> ; 4.087 at 25 °C	4.102 at 25 °C
Odor	No data	No data
Odor threshold:		
Water	No data	No data
Air	No data	No data
Solubility:		
Water	6.9 mg/L at 18 °C <sup>i</sup>	6.5 mg/L at 18 °C <sup>i</sup>
Organic solvents	Very soluble in alcohol; soluble in dilute mineral acids; insoluble in acetic acid; insoluble in alkalis	Very soluble in alcohol; soluble in dilute mineral acids; insoluble in alkalis
Partition coefficients:		
K <sub>d</sub>	No data	No data
K <sub>ow</sub>	No data	No data
K <sub>oc</sub>	No data	No data
Vapor pressure	No data	No data
Henry's law constant	Not applicable	Not applicable
Autoignition temperature	No data	No data
Flashpoint	No data	No data
Flammability limits	No data	No data
Conversion factor	mg ZnS x 0.67 = mg Zn	mg ZnS x 0.67 = mg Zn
Explosive limits	No data	No data

<sup>a</sup>Information obtained from O'Neil et al. (2001) except where noted.

<sup>b</sup>ACGIH 1991

<sup>c</sup>Lewis 1997

<sup>d</sup>HSDB 2003

<sup>e</sup>Baes and Sharp 1983

<sup>f</sup>Baes et al. 1984

<sup>g</sup>Gerritse et al. 1982

<sup>h</sup>Weiss 1986

<sup>i</sup>Weast 1988

<sup>j</sup>Goodwin 1998

Zn = zinc; Zn(C<sub>2</sub>H<sub>3</sub>O<sub>2</sub>)<sub>2</sub> = zinc acetate; ZnCl<sub>2</sub> = zinc chloride; ZnCrO<sub>4</sub> = zinc chromate; Zn(CN)<sub>2</sub> = zinc cyanide; Zn(OH)<sub>2</sub> = zinc hydroxide; ZnO = zinc oxide; Zn<sub>3</sub>(PO<sub>4</sub>)<sub>2</sub> = zinc phosphate; ZnS = zinc sulfide; ZnSO<sub>4</sub> = zinc sulfate

## 5. PRODUCTION, IMPORT/EXPORT, USE, AND DISPOSAL

### 5.1 PRODUCTION

Zinc is widely distributed in nature, constituting 20–200 ppm (by weight) of the earth's crust (Goodwin 1998), but it is not found as elemental zinc in nature (Lloyd and Showak 1984). The procedure used to mine zinc varies with the composition of the ore. The mineral sphalerite (ZnS) provides ca. 90% of the zinc produced today (Goodwin 1998). Zinc ore is mined using both underground mining and open pit mining (Stokinger 1981). The mined zinc ores are too low in zinc content for direct reduction to refined metal; thus, they are first concentrated. Production of concentrates requires crushing and grinding followed by gravity or magnetic methods of separation or flotation. These processes may be combined, depending on the complexity of the ore. A caustic-leach process is used to decrease the extent of metal loss during the concentration process. In this process, the metal is leached by caustic soda, the resulting electrolyte is purified with zinc dust and lime, and the zinc is electrodeposited. The crude zinc may be dissolved in sulfuric acid and purified by electrodeposition. Two processes are used to produce metallic zinc from the ore concentrates that are not subjected to caustic soda leaching. In one process, the ore concentrate containing zinc sulfide is roasted in the presence of air to produce zinc oxide, which is combined with coke or coal and retorted to approximately 1,100 °C to produce metallic zinc. In the other process, the roasted zinc oxide is leached with sulfuric acid, and the solution is electrolyzed to produce zinc of >99.9% purity. The electrolytic processing of zinc is replacing smelting as the most commonly used process (Lloyd and Showak 1984; Stokinger 1981).

Continued low zinc prices in 2001 have resulted in operation reductions and facility closures across the United States. By the end of 2001, 12 mines in 5 states were in operation in the United States. Alaska was the leading zinc-mining state, followed by (in descending order) Tennessee, Missouri, New York, and Montana. Alaska also had the largest production of recoverable zinc in the United States in 2001, followed by Missouri, Montana, and New York. In 2001, three companies operated three primary zinc refineries (Zinc Corporation of America, Monaco, Pennsylvania; Big River Zinc Corporation, Sauget, Illinois; and Pasminco Ltd., Clarksville, Tennessee) (USGS 2001). Tables 5-1 and 5-2 summarize the facilities that manufacture or process zinc and zinc compounds, respectively, in the United States. The information in this table was obtained from the Toxics Release Inventory (TRI), and it summarizes the reported release data for 2001 (TRI01 2003). However, this list does not include all facilities that manufacture or process zinc and zinc compounds. Tables 5-1 and 5-2 also list the maximum amounts of

## 5. PRODUCTION, IMPORT/EXPORT, USE, AND DISPOSAL

**Table 5-1. Facilities that Produce, Process, or Use Zinc**

State <sup>a</sup>	Number of facilities	Minimum amount on site in pounds <sup>b</sup>	Maximum amount on site in pounds <sup>b</sup>	Activities and uses <sup>c</sup>
AL	10	1,000	999,999	1, 4, 5, 7, 8, 11
AR	7	1,000	999,999	1, 4, 5, 7, 8, 12, 14
AZ	2	1,000	999,999	8
CA	16	1,000	9,999,999	1, 3, 4, 5, 7, 8, 12
CO	2	10,000	999,999	2, 3, 8
CT	3	1,000	99,999	1, 5, 6
FL	3	1,000	99,999	8, 12
GA	8	1,000	99,999	1, 2, 3, 5, 7, 8
IA	7	0	999,999	1, 5, 8, 13
ID	2	10,000	49,999,999	1, 3, 5, 12
IL	19	100	499,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11
IN	16	100	999,999	1, 5, 6, 7, 8
KS	6	0	99,999	1, 8, 12, 13
KY	12	100	999,999	1, 2, 3, 4, 5, 7, 8, 11, 12, 13
LA	9	100	999,999	1, 5, 7, 8, 10, 11, 12
MA	5	1,000	99,999	1, 5, 7, 8, 9, 12
MD	2	100,000	999,999	7
ME	1	10,000	99,999	8
MI	14	100	999,999	1, 2, 3, 5, 7, 8, 10, 12
MN	3	10,000	99,999	8
MO	3	100	999,999	1, 5, 7, 8
MS	4	1,000	999,999	7, 8, 9
NC	10	0	9,999,999	1, 4, 8, 9, 13, 14
NE	2	10,000	99,999	1, 3, 5, 7, 9, 13
NH	1	10,000	99,999	1, 5
NJ	7	1,000	49,999,999	1, 2, 3, 5, 6, 8
NY	7	0	999,999	1, 2, 3, 5, 7, 8, 9
OH	31	0	9,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 12, 13
OK	10	0	999,999	1, 2, 3, 5, 7, 8, 9, 11, 12
OR	1	10,000	99,999	12
PA	24	0	999,999	1, 2, 3, 4, 5, 7, 8, 9, 10, 11, 12, 13, 14
RI	2	100	99,999	1, 5, 8
SC	7	100	999,999	1, 5, 8, 9, 11, 12, 13
SD	2	10,000	99,999	1, 5, 8, 12, 14
TN	10	0	49,999,999	1, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14
TX	17	1,000	999,999	1, 2, 3, 5, 6, 7, 8, 9, 10, 11, 12, 14

## 5. PRODUCTION, IMPORT/EXPORT, USE, AND DISPOSAL

**Table 5-1. Facilities that Produce, Process, or Use Zinc**

State <sup>a</sup>	Number of facilities	Minimum amount on site in pounds <sup>b</sup>	Maximum amount on site in pounds <sup>b</sup>	Activities and uses <sup>c</sup>
UT	3	10,000	99,999	2, 5, 7, 10, 12
VA	5	1,000	99,999	2, 3, 7, 8, 11
WA	1	10,000	99,999	7
WI	14	0	999,999	1, 2, 3, 4, 5, 7, 8, 9, 11
WV	8	100	9,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 12
WY	2	1,000	99,999	1, 4, 9, 12

Source: TRI01 2003

<sup>a</sup>Post office state abbreviations used

<sup>b</sup>Amounts on site reported by facilities in each state

<sup>c</sup>Activities/Uses:

- |                          |                          |                             |
|--------------------------|--------------------------|-----------------------------|
| 1. Produce               | 6. Impurity              | 11. Chemical Processing Aid |
| 2. Import                | 7. Reactant              | 12. Manufacturing Aid       |
| 3. Onsite use/processing | 8. Formulation Component | 13. Ancillary/Other Uses    |
| 4. Sale/Distribution     | 9. Article Component     | 14. Process Impurity        |
| 5. Byproduct             | 10. Repackaging          |                             |

## 5. PRODUCTION, IMPORT/EXPORT, USE, AND DISPOSAL

**Table 5-2. Facilities that Produce, Process, or Use Zinc Compounds**

State <sup>a</sup>	Number of facilities	Minimum amount on site in pounds <sup>b</sup>	Maximum amount on site in pounds <sup>b</sup>	Activities and uses <sup>c</sup>
AK	4	1,000	10,000,000,000	1, 3, 5, 7, 9, 10, 12, 13
AL	68	100	99,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14
AR	59	0	499,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14
AZ	17	1,000	10,000,000,000	1, 2, 3, 4, 5, 6, 7, 8, 9, 11, 12, 13, 14
CA	90	0	9,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14
CO	14	100	49,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 12, 13, 14
CT	33	100	999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12
DE	11	1,000	999,999	1, 2, 4, 5, 7, 8, 10, 12, 13, 14
FL	44	0	9,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14
GA	75	0	9,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13
HI	2	1,000	99,999	1, 5, 7, 9
IA	58	100	49,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14
ID	8	10,000	9,999,999	1, 2, 3, 5, 7, 8, 10
IL	193	0	499,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14
IN	134	0	499,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14
KS	28	0	999,999	1, 2, 3, 5, 6, 7, 8, 9, 10, 11, 12, 13
KY	75	0	999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14
LA	60	0	49,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14
MA	41	100	9,999,999	1, 2, 3, 4, 5, 6, 7, 8, 12, 14
MD	23	1,000	9,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13
ME	7	100	999,999	1, 3, 5, 6, 8, 11, 12, 13
MI	142	0	49,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14
MN	36	100	9,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14
MO	72	0	499,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14
MS	39	100	999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13
MT	5	100	10,000,000,000	1, 2, 3, 4, 5, 6, 9, 12, 13, 14
NC	66	100	9,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 11, 12, 13, 14
ND	4	1,000	99,999	1, 5, 9, 12, 13
NE	27	0	9,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13
NH	9	1,000	999,999	1, 5, 7, 8, 9, 10
NJ	61	0	49,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13
NM	9	1,000	9,999,999	1, 3, 4, 5, 8, 9, 10, 12, 13
NV	20	0	999,999,999	1, 3, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14
NY	62	100	10,000,000,000	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13
OH	261	0	9,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14
OK	37	1,000	9,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14
OR	21	0	9,999,999	1, 2, 3, 4, 5, 7, 8, 9, 10, 11, 12, 13, 14
PA	152	0	49,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14
RI	8	1,000	999,999	1, 3, 5, 6, 7, 8, 12
SC	65	0	9,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14

## 5. PRODUCTION, IMPORT/EXPORT, USE, AND DISPOSAL

**Table 5-2. Facilities that Produce, Process, or Use Zinc Compounds**

State <sup>a</sup>	Number of facilities	Minimum amount on site in pounds <sup>b</sup>	Maximum amount on site in pounds <sup>b</sup>	Activities and uses <sup>c</sup>
SD	3	1,000	9,999,999	1, 5, 7, 9, 10, 13
TN	99	0	499,999,999	1, 2, 3, 4, 5, 6, 7, 8, 10, 11, 12
TX	170	0	99,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14
UT	18	100	499,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14
VA	56	0	9,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14
VT	2	1,000	99,999	6, 7
WA	20	0	9,999,999	1, 3, 4, 5, 6, 7, 8, 9, 10, 12, 13
WI	87	0	9,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14
WV	25	0	9,999,999	1, 3, 4, 5, 7, 8, 9, 10, 11, 12, 13, 14
WY	6	0	9,999,999	1, 4, 5, 6, 9, 12, 13

Source: TRI01 2003

<sup>a</sup>Post office state abbreviations used<sup>b</sup>Amounts on site reported by facilities in each state<sup>c</sup>Activities/Uses:

- |                          |                          |                             |
|--------------------------|--------------------------|-----------------------------|
| 1. Produce               | 6. Impurity              | 11. Chemical Processing Aid |
| 2. Import                | 7. Reactant              | 12. Manufacturing Aid       |
| 3. Onsite use/processing | 8. Formulation Component | 13. Ancillary/Other Uses    |
| 4. Sale/Distribution     | 9. Article Component     | 14. Process Impurity        |
| 5. Byproduct             | 10. Repackaging          |                             |



## 5. PRODUCTION, IMPORT/EXPORT, USE, AND DISPOSAL

zinc and zinc compounds, respectively, that are present at these sites and the end uses of zinc. In 2001, approximately 799,000 metric tons of zinc was produced in the United States from domestic ores. The estimated world production from mines in 2001 was 8,850,000 metric tons. The world production of zinc has increased from 1997 to 2001 (USGS 2001).

Zinc is available in many commercial forms, including ingots, lumps, sheets, wire, shot, strips, sticks, granules, granulated zinc (obtained when molten metal is poured into cold water), and powder (O'Neil et al. 2001).

### 5.2 IMPORT/EXPORT

In 2001, approximately 813,000 metric tons of zinc were imported to the United States as refined slab zinc, 84,000 metric tons were imported as ores and concentrates, and 7,240 metric tons were imported as rolled zinc. In 2001, the United States imported less refined slab and rolled zinc than in the previous 4 years, and more ores and concentrate than in the previous 4 years (USGS 2001).

In 2001, an estimated 696,000 metric tons of ores and concentrates, 1,180 metric tons of slab zinc, and 5,700 metric tons of rolled zinc were exported from the United States. In contrast, exports of ores and concentrates reached approximately 23,000 metric tons in 1985 and 461,000 in 1997 (DOI 1988, 1991; USGS 2001). In 2001, the United States exported the largest amounts of zinc ores and concentrates to Belgium (151,000 metric tons), Japan (141,000 metric tons), and South Korea (141,000 metric tons) (USGS 2001).

### 5.3 USE

Zinc metal is used most commonly as a protective coating of other metals, such as iron and steel. Methods, in general, include hot-dip galvanizing, continuous-line galvanizing, electro-galvanizing, zinc plating, zinc spraying, and painting with zinc-bearing paints. Some examples of galvanized materials include nails, water towers, and electrical transmission towers. Because zinc metal lacks strength, it is frequently alloyed with other metals (e.g., aluminum, copper, titanium, and magnesium) to impart a range of properties. When zinc metal is the primary component of the alloy, it is called a 'zinc-base' alloy, which is primarily used for casting and wrought applications. Other important applications of zinc alloys are in dye-casting, construction, and in other alloys (e.g., brass and bronze) which may be found in electrical components of many household goods. Also, alloys containing zinc and copper are used to

## 5. PRODUCTION, IMPORT/EXPORT, USE, AND DISPOSAL

make U.S. one-cent coins. Zinc metal dust is widely used in paint coatings, as a catalyst, and as a reducing and precipitating agent in organic and analytical chemistry (Goodwin 1998). As shown in Table 5-3, in 2001, the reported consumption of zinc by industry was 281,000 metric tons (51.4% of total consumption) for galvanizing; 91,200 metric tons (16.8% of total consumption) for zinc-based alloys; and 74,400 metric tons (13.7% of total consumption) for brass and bronze (USGS 2001).

Zinc compounds have dental, medical, and household applications. In pharmaceuticals, zinc salts are used as solubilizing agents in many drugs, including insulin (Lloyd 1984; Lloyd and Showak 1984; Windholz 1983). Zinc compounds are utilized therapeutically in human medicine in the treatment of zinc deficiency (Keen and Hurley 1977). Zinc oxide accounts for the largest use of zinc compounds, and is used primarily by the rubber industry as a vulcanization activator and accelerator and to slow rubber aging by neutralizing sulfur and organic acids formed by oxidation. It also acts in rubber as a reinforcing agent, a heat conductor, a white pigment, and an absorber of UV light. In paints, zinc oxide serves as a mildewstat, acid buffer, and a pigment. It is used in animal feed as a zinc supplement and as a fertilizer-additive for zinc-deficient soils. Zinc oxide is used in cosmetics and drugs primarily for its fungicide properties, and in dentistry in dental cements. It is also used in ceramics, in glass manufacture, as a catalyst in organic synthesis, and in coated photocopy paper (Goodwin 1998). The largest uses of zinc chloride in the United States are in wood preservation, solder fluxes, and batteries. Solutions of zinc chloride are widely used in mercerizing cotton and as a mordant in dyeing. In medicine, zinc chloride is used as an antiseptic, disinfectant, deodorant, and in dental cements. Other uses are in organic synthesis, as a dehydrant, in rubber vulcanization, and in oil refining (Goodwin 1998). Zinc chloride is a primary ingredient in smoke bombs used for crowd dispersal, in fire-fighting exercises (by both military and civilian communities), and by the military for screening purposes (WHO 2001). Zinc sulfate is used in fertilizers, sprays, and animal feed as a trace element and disease-control agent. It is used in the manufacture of rayon (as crendulating agent), as a starting material for many zinc chemicals, in textile dyeing and printing, in flotation reagents, for electrogalvanizing, in paper bleaching, and in glue (Goodwin 1998). Zinc sulfide is used as a phosphor (watches, TV screens), a white pigment, and in dental materials (especially in form of lithopone) (O'Neil et al. 2001). Uses for zinc acetate are as a wood preservative, a mordant for antiseptics, a catalyst, and a waterproofing agent. Zinc cyanide has two uses: electroplating and gold extraction. The primary uses of zinc phosphate are in preparation of metal coatings and as a dental cement (Goodwin 1998). Zinc chromate is used in pigments. Zinc hydroxide uses are as an intermediate, as an absorbent in surgical dressings, and in rubber compounding (Lewis 1997).

## 5. PRODUCTION, IMPORT/EXPORT, USE, AND DISPOSAL

**Table 5-3. Distribution of U.S. Zinc Consumption in 2001**

Use	Total (metric tons)	Percent of total
Galvanizing	281,000	51.7
Zinc-based alloys	91,200	16.8
Brass and bronze	74,400	13.7
Total <sup>a</sup>	543,000	100.0

Source: USGS 2001

<sup>a</sup>Data are rounded to more than three significant figures; may not add totals shown.

## 5. PRODUCTION, IMPORT/EXPORT, USE, AND DISPOSAL

**5.4 DISPOSAL**

Zinc processing plants have attempted to limit releases to the environment by using techniques such as water reuse, control of particulate emissions, and filtration thickener overflow. In addition, liquid effluents are limed and allowed to settle so that zinc can precipitate out as the hydroxide (Lloyd and Showak 1984). Waste products containing zinc are also being used as a source of zinc for electrogalvanizing (Jolly 1988). Disposal procedures for spills include ferric hydroxide precipitation and cement-based fixation processes; the latter method is very effective in rendering zinc contaminants insoluble (Dawson and Mercer 1986). Unsalvageable zinc waste may be buried in an approved landfill while salvageable zinc is typically recycled. In 2002, an estimated 370,000 tons of zinc were recovered from waste and scrap in the United States; about 30% was recovered in the form of slab zinc and the remainder was recovered in alloys, oxide, and chemicals. Of the total amount of scrap recycled, 300,000 tons was derived from new scrap and 70,000 tons were derived from old scrap. About 25,000 tons of scrap in the United States were exported mainly to China, Canada, and Taiwan (USGS 2003).

In 1989, EPA applied its revised interpretation of the Bevill Amendment (exclusion) to solid waste from the extraction, beneficiation, and processing of ores and minerals. The slag from the primary zinc processing is the only zinc-related waste remaining in the Bevill exclusion (DOI 1991).



## 6. POTENTIAL FOR HUMAN EXPOSURE

### 6.1 OVERVIEW

Zinc has been identified in at least 953 of the 1,636 hazardous waste sites that have been proposed for inclusion on the EPA National Priorities List (NPL) (HazDat 2003). However, the number of sites evaluated for zinc is not known. The frequency of these sites can be seen in Figure 6-1. Of these sites, 941 are located within the United States, and 2, 9, and 1 sites are located in the Commonwealths of Guam, Puerto Rico, and the Virgin Islands, respectively (not shown).

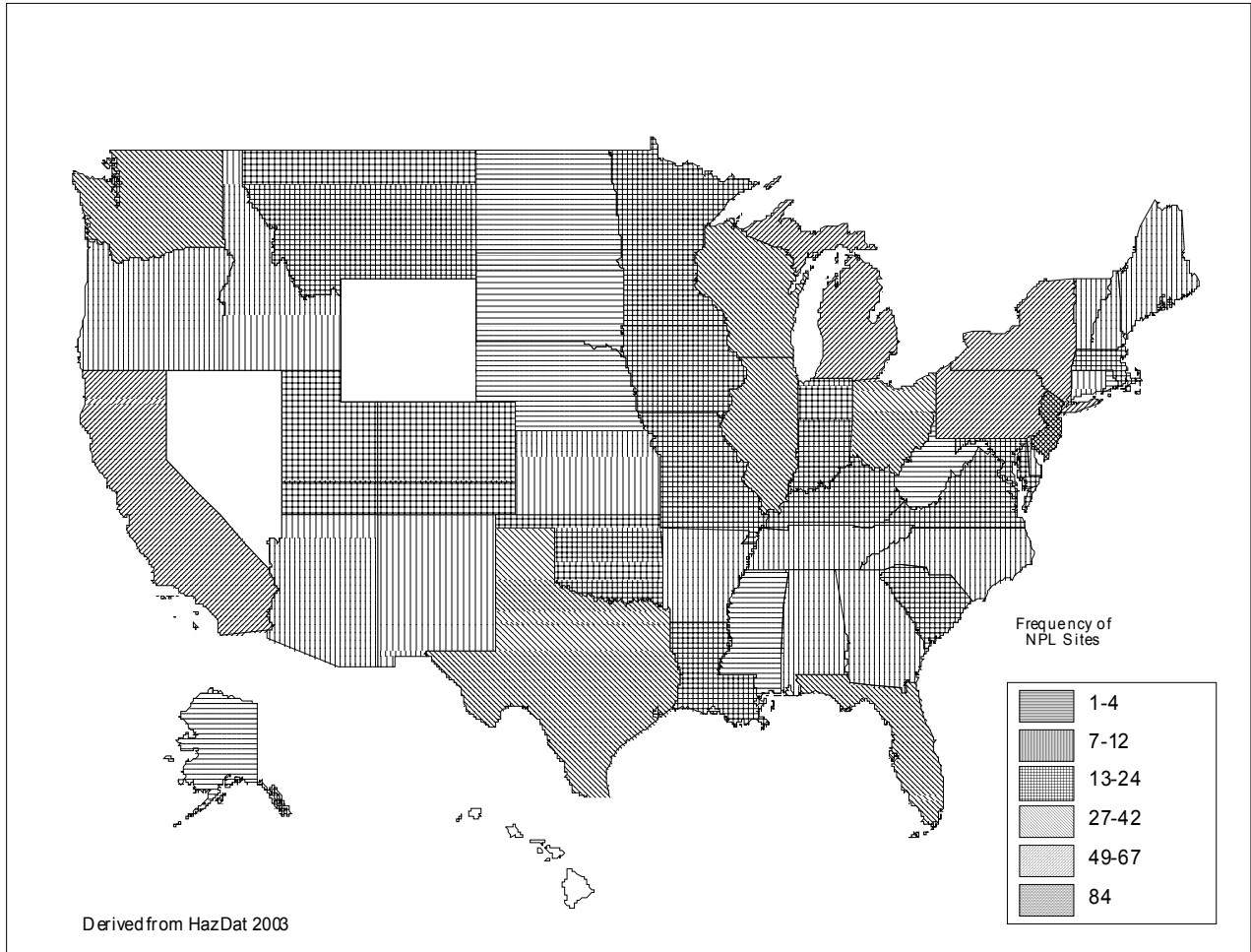
Zinc is an element commonly found in the earth's crust. It is released to the environment from both natural and anthropogenic sources; however, releases from anthropogenic sources are greater than those from natural sources. The primary anthropogenic sources of zinc in the environment (air, water, soil) are related to mining and metallurgic operations involving zinc and use of commercial products containing zinc. Worldwide, releases to soil are probably the greatest source of zinc in the environment. The most important sources of zinc in soil are discharges of smelter slags and wastes, mine tailings, coal and bottom fly ash, and the use of commercial products such as fertilizers and wood preservatives that contain zinc. Zinc does not volatilize from soil. Although zinc usually remains adsorbed to soil, leaching has been reported at waste disposal sites. Zinc does not volatilize from water but is deposited primarily in sediments through adsorption and precipitation. Severe zinc contamination tends to be confined to areas near emission sources.

Zinc is capable of forming complexes with a variety of organic and inorganic groups (ligands). Biological activity can affect the mobility of zinc in the aquatic environment, although the biota contains relatively little zinc compared to the sediments. Zinc bioconcentrates moderately in aquatic organisms; bioconcentration is higher in crustaceans and bivalve species than in fish. Zinc does not concentrate in plants, and it does not biomagnify through terrestrial food chains.

There are few data regarding the speciation of zinc released to the atmosphere. Zinc is removed from the air by dry and wet deposition, but zinc particles with small diameters and low densities suspended in the atmosphere travel long distances from emission sources.

6. POTENTIAL FOR HUMAN EXPOSURE

**Figure 6-1. Frequency of NPL Sites with Zinc Contamination**



## 6. POTENTIAL FOR HUMAN EXPOSURE

Zinc has been detected in air, surface water, groundwater, and soil; the frequency of detection and the concentrations are greatest near source areas (e.g., hazardous waste sites and industrial areas such as lead smelters). In a survey by the National Air Surveillance Network, the mean concentration of zinc in the air in the United States in 1977–1979 was 0.02–0.16  $\mu\text{g}/\text{m}^3$  for urban air compared to 0.01–0.05  $\mu\text{g}/\text{m}^3$  for rural air. The concentrations of zinc in the air of remote areas range from <0.003 to 0.027  $\mu\text{g}/\text{m}^3$ . The mean concentrations of zinc in ambient water and drinking water range from 0.02 to 0.05 mg/L and from 0.01 to 0.1 mg/L, respectively. The concentration of zinc in drinking water can often be higher than the concentration in the raw water from which the drinking water was obtained because zinc may leach from transmission and distribution pipes. The concentration of zinc in standing water from galvanized household water pipes was  $\leq 1.3$  mg/L (Sharrett et al. 1982a). The concentration of zinc in cultivated soils in the United States ranged from <5 to 400 mg/kg, with a mean of 36 mg/kg, compared to a range of <10–2,000 mg/kg, with a mean of 51 mg/kg, in uncultivated soils; this probably results from the differences in soils used for farming rather than the use of zinc in agriculture. Concentrations of zinc can be high in soils from contaminated sites, such as waste dumps.

The concentrations of zinc in various foods and human tissues have also been determined. In a 1980–1982 survey of total diet samples, FDA estimated that the average intake of zinc from food (including water) for an adult was 0.23 mg/kg-day. FDA concluded that the daily intake of zinc from the inhalation of ambient air is negligible compared to the daily intake from food. Certain population groups may be exposed to higher concentrations of zinc than the general population. People who work in coal mines, people who work with the refining and smelting of nonferrous metals, and people who live near waste sites and metal smelting operations may be exposed to high levels of zinc. People who consume large amounts of foods high in zinc content, such as oysters and mussels, may also be exposed to elevated levels of zinc. The higher exposure may or may not be manifested as increased body burden in the exposed individuals.

## 6.2 RELEASES TO THE ENVIRONMENT

Zinc is commonly found in the earth's crust, and natural releases to the environment can be significant. In addition, zinc is one of the most widely used metals in the world. The major industrial sources of zinc include electroplating, smelting and ore processing, and drainage from both active and inactive mining operations (Mirenda 1986). Furthermore, zinc is an important component of brass, bronze, die casting



## 6. POTENTIAL FOR HUMAN EXPOSURE

metal, other alloys, rubber, and paints. The environmental releases of zinc from sources of human origin far exceed the releases from natural sources (Fishbein 1981).

**6.2.1 Air**

Natural emissions of zinc and its compounds to air are mainly due to windborne soil particles, volcanic emissions, and forest fires, and are estimated to be 19,000, 9,600, and 7,600 metric tons/year, respectively. Other natural sources of zinc in air are biogenic emissions and sea salt sprays with annual amounts estimated to be 8,100 and 440 metric tons, respectively (Nriagu 1989).

Anthropogenic releases of zinc and its compounds to the atmosphere are from dust and fumes from mining, zinc production facilities, processing of zinc-bearing raw materials (e.g., lead smelters), brass works, coal and fuel combustion, refuse incineration, and iron and steel production (EPA 1980d; Ragaini et al. 1977). In urban East St. Louis, Illinois, industrial complexes, such as smelters, accounted for about 86% of fine and coarse particulate matter emitted into air. In southeastern Chicago, Illinois, incinerator emissions accounted for 86% of fine particulate in the atmosphere while urban dust account for 93% of the coarse particulate emissions (Sweet et al. 1993). Estimated atmospheric zinc loss is 100 g/ton of zinc mined, and most of the loss comes from handling raw and concentrated ore and wind erosion of tailing piles (Lloyd and Showak 1984). Average zinc emissions to the atmosphere from stationary sources in the United States were 151,000 tons/year (137,000 metric tons/year) for 1969–1971 (Fishbein 1981). Based on emission studies in Western Europe, the United States, Canada, and the former Soviet Union, total worldwide zinc emissions to air were calculated to range from 70,250 to 193,500 metric tons in 1983. Emissions from the non-ferrous metal industry account for the largest fraction of zinc emitted (50–70%) (Nriagu and Pacyna 1988). However, emissions have decreased considerably since the 1970s and 1980s as a result of improvements in contemporary zinc production facilities (WHO 2001).

According to the TRI, estimated totals of 870,792 pounds (395 metric tons) of zinc (dust and fume) and 6,159,692 pounds (2,790 metric tons) of zinc compounds, amounting to about 2.51 and 0.81%, respectively, of the total environmental on-site releases, were discharged into the atmosphere in the United States in 2001 from mining, manufacturing, processing, and electrical power generation industries listed in Tables 6-1 and 6-2 (TRI01 2003). Data for stack/point source emissions indicate releases of 495,206 pounds (224 metric tons) of zinc (dust and fume) and 4,617,457 pounds (2094 metric tons) of zinc compounds, while data for fugitive source emission indicate a release of 375,586 pounds (170 metric tons) of zinc (dust and fume) and 1,542,235 pounds (700 metric tons) of zinc compounds. The TRI data

## 6. POTENTIAL FOR HUMAN EXPOSURE

**Table 6-1. Releases to the Environment from Facilities that Produce, Process, or Use Zinc**

Reported amounts released in pounds per year <sup>a</sup>								
State <sup>b</sup>	Number of facilities	Air <sup>c</sup>	Water	Under-ground injection	Land	Total on-site release <sup>d</sup>	Total off-site release <sup>e</sup>	Total on and off-site release
AL	12	22,700	568	0	9,936	33,204	16,937	50,141
AR	11	11,995	36	0	96,221	108,252	21,764	130,016
AZ	2	500	No data	0	0	500	0	500
CA	19	9,804	8	0	1,468,045	1,477,857	4,327	1,482,184
CO	2	793	263	0	0	1,056	2,326	3,382
CT	5	864	No data	0	0	864	0	864
DE	1	No data	No data	No data	No data	No data	No data	0
FL	5	90,870	250	0	0	91,120	7,625	98,745
GA	13	271	0	0	3	274	2,915	3,189
IA	11	16,857	0	0	0	16,857	112,641	129,498
ID	2	6,933	No data	0	24,925,639	24,932,572	550	24,933,122
IL	25	188,120	5	0	37,815	225,940	1,415,368	1,641,308
IN	24	13,484	15	0	26,228	39,727	227,484	267,211
KS	6	33,065	0	0	0	33,065	29,070	62,135
KY	13	57,834	No data	44,518	46,018	148,370	73,355	221,725
LA	14	17,455	No data	0	47	17,502	1,263,055	1,280,557
MA	7	662	No data	0	0	662	947	1,609
MD	4	83	No data	0	0	83	4,200	4,283
ME	1	0	No data	0	0	0	9,193	9,193
MI	16	12,631	12	0	250,089	262,732	87,151	349,883
MN	3	278	No data	0	0	278	0	278
MO	5	326	No data	0	0	326	5,350	5,676

## 6. POTENTIAL FOR HUMAN EXPOSURE

**Table 6-1. Releases to the Environment from Facilities that Produce, Process, or Use Zinc**

Reported amounts released in pounds per year <sup>a</sup>								
State <sup>b</sup>	Number of facilities	Air <sup>c</sup>	Water	Under-ground injection	Land	Total on-site release <sup>d</sup>	Total off-site release <sup>e</sup>	Total on and off-site release
MS	5	1,755	1,398	0	11,279	14,432	255	14,687
MT	1	No data	No data	No data	No data	No data	No data	0
NC	14	2,700	No data	0	0	2,700	1,694,446	1,697,146
NE	2	3,596	No data	0	27,000	30,596	5	30,601
NH	1	1,366	5	0	0	1,371	5	1,376
NJ	7	9,585	0	0	0	9,585	18,561	28,146
NV	1	No data	No data	No data	No data	No data	No data	0
NY	8	8,809	No data	0	0	8,809	1,521	10,330
OH	36	89,244	7,017	0	6,567,340	6,663,601	5,308,872	11,972,473
OK	13	12,524	177	0	12,726	25,427	1,700	27,127
OR	1	0	No data	0	29,879	29,879	0	29,879
PA	25	84,915	207	0	0	85,122	634,798	719,920
RI	2	881	No data	0	0	881	0	881
SC	7	57,346	481	0	18,255	76,082	69,239	145,321
SD	2	368	No data	0	250	618	1,061	1,679
TN	15	4,479	136	0	55,323	59,938	27,842	87,780
TX	19	34,148	4,009	0	0	38,157	9,760	47,917
UT	3	446	No data	0	19,772	20,218	1,813	22,031
VA	6	3,723	750	0	23,454	27,927	9,407	37,334
WA	2	30	No data	0	0	30	537	567
WI	15	7,103	0	0	0	7,103	222,990	230,093

## 6. POTENTIAL FOR HUMAN EXPOSURE

**Table 6-1. Releases to the Environment from Facilities that Produce, Process, or Use Zinc**

Reported amounts released in pounds per year <sup>a</sup>								
State <sup>b</sup>	Number of facilities	Air <sup>c</sup>	Water	Under-ground injection	Land	Total on-site release <sup>d</sup>	Total off-site release <sup>e</sup>	Total on and off-site release
WV	9	62,061	89	0	31,517	93,667	5,698	99,365
WY	2	188	0	0	38,607	38,795	5,177	43,972
Total	399	870,792	15,426	44,518	33,695,443	34,626,179	11,297,950	45,924,130

Source: TRI01 2003

<sup>a</sup>Data in TRI are maximum amounts released by each facility.

<sup>b</sup>Post office state abbreviations are used.

<sup>c</sup>The sum of fugitive and stack releases are included in releases to air by a given facility.

<sup>d</sup>The sum of all releases of the chemical to air, land, water, and underground injection wells.

<sup>e</sup>Total amount of chemical transferred off-site, including to publicly owned treatment works (POTW).

## 6. POTENTIAL FOR HUMAN EXPOSURE

**Table 6-2. Releases to the Environment from Facilities that Produce, Process, or Use Zinc Compounds**

Reported amounts released in pounds per year <sup>a</sup>								
State <sup>b</sup>	Number of facilities	Air <sup>c</sup>	Water	Under-ground injection	Land	Total on-site release <sup>d</sup>	Total off-site release <sup>e</sup>	Total on and off-site release
AK	5	58,859	845	9,100,000	300,902,429	310,062,133	0	310,062,133
AL	90	133,635	75,179	0	7,144,741	7,353,555	4,211,129	11,564,684
AR	79	66,586	39,130	7,438	297,440	410,594	9,278,540	9,689,134
AZ	23	25,146	454	0	67,699,826	67,725,426	29,895	67,755,321
CA	122	33,969	13,382	1,350	1,000,432	1,049,133	343,282	1,392,415
CO	21	53,177	10,165	0	2,957,817	3,021,159	110,727	3,131,886
CT	34	5,144	5,392	0	116	10,652	338,393	349,045
DE	14	12,982	8,786	0	42,638	64,406	74,924	139,330
FL	68	44,320	17,043	0	632,355	693,718	746,895	1,440,613
GA	109	223,022	156,428	0	716,491	1,095,941	484,292	1,580,233
HI	2	22	No data	5	0	27	32,774	32,801
IA	94	84,368	5,741	0	416,906	507,015	1,099,111	1,606,126
ID	12	41,262	2,320	0	5,233,884	5,277,466	4,657	5,282,123
IL	239	441,009	14,881	327	23,396,088	23,852,305	6,836,025	30,688,330
IN	162	420,434	1,083,235	0	7,741,995	9,245,664	48,482,012	57,727,676
KS	42	39,981	401	18,367	153,956	212,705	312,778	525,483
KY	89	172,598	47,669	0	2,507,548	2,727,815	836,467	3,564,282
LA	72	100,444	61,605	683	1,378,694	1,541,426	940,794	2,482,220
MA	49	9,930	1,065	0	62,026	73,021	530,875	603,896
MD	31	18,216	6,569	10,819	118,267	153,871	205,102	358,973

## 6. POTENTIAL FOR HUMAN EXPOSURE

**Table 6-2. Releases to the Environment from Facilities that Produce, Process, or Use Zinc Compounds**

Reported amounts released in pounds per year <sup>a</sup>								
State <sup>b</sup>	Number of facilities	Air <sup>c</sup>	Water	Under-ground injection	Land	Total on-site release <sup>d</sup>	Total off-site release <sup>e</sup>	Total on and off-site release
ME	7	15,568	16,601	0	199,158	231,327	400,793	632,120
MI	157	232,287	21,331	13,000	653,887	920,505	29,953,155	30,873,660
MN	58	55,945	4,626	0	436,020	496,591	1,327,338	1,823,929
MO	97	470,171	15,915	0	31,123,423	31,609,509	526,720	32,136,229
MS	55	542,446	19,442	200,000	230,946	992,834	734,718	1,727,552
MT	7	6,624	14	0	24,009,658	24,016,296	756,919	24,773,215
NC	96	64,514	18,139	0	709,595	792,248	1,559,035	2,351,283
ND	6	746	22	0	63,000	63,768	104,345	168,113
NE	40	101,586	1,392	0	112,420	215,398	526,296	741,694
NH	9	586	35	0	1,300	1,921	11,141	13,062
NJ	78	23,020	19,076	0	100,036	142,132	17,857,518	17,999,650
NM	11	7,469	2,000	0	6,708,333	6,717,802	119,842	6,837,644
NV	23	8,747	230	42	148,683,690	148,692,709	35,208	148,727,917
NY	78	58,622	51,211	0	131,191	241,024	1,089,487	1,330,511
OH	311	188,750	49,562	300,000	2,371,120	2,909,432	12,700,672	15,610,104
OK	50	28,394	5,693	1,812	680,876	716,775	1,198,227	1,915,002
OR	34	20,045	17,770	0	70,153	107,968	96,957	204,925
PA	192	1,294,230	47,185	0	1,331,395	2,672,810	33,002,255	35,675,065
RI	8	187	No data	0	0	187	80,697	80,884
SC	75	91,961	42,668	0	798,294	932,923	13,912,886	14,845,809

## 6. POTENTIAL FOR HUMAN EXPOSURE

**Table 6-2. Releases to the Environment from Facilities that Produce, Process, or Use Zinc Compounds**

Reported amounts released in pounds per year <sup>a</sup>								
State <sup>b</sup>	Number of facilities	Air <sup>c</sup>	Water	Under-ground injection	Land	Total on-site release <sup>d</sup>	Total off-site release <sup>e</sup>	Total on and off-site release
SD	8	5,041	No data	0	2,100,000	2,105,041	32,085	2,137,126
TN	115	350,666	84,221	5	16,734,935	17,169,827	1,674,456	18,844,282
TX	219	386,877	48,651	3,083	1,210,518	1,649,129	4,589,566	6,238,695
UT	23	9,496	8,672	0	75,447,094	75,465,262	6,955	75,472,217
VA	66	112,681	25,675	0	1,021,023	1,159,379	3,914,939	5,074,318
VT	3	0	No data	0	0	0	22,917	22,917
WA	29	22,578	11,874	0	123,724	158,176	232,927	391,103
WI	109	56,022	4,975	0	192,559	253,556	3,454,411	3,707,967
WV	29	13,579	104,917	0	829,381	947,877	300,340	1,248,217
WY	7	4,584	224	0	133,230	138,038	28,066	166,104
<b>Total</b>	<b>3,372</b>	<b>6,159,692</b>	<b>2,172,966</b>	<b>9,656,931</b>	<b>738,610,608</b>	<b>756,600,197</b>	<b>205,468,291</b>	<b>962,068,488</b>

Source: TRI01 2003

<sup>a</sup>Data in TRI are maximum amounts released by each facility.<sup>b</sup>Post office state abbreviations are used.<sup>c</sup>The sum of fugitive and stack releases are included in releases to air by a given facility.<sup>d</sup>The sum of all releases of the chemical to air, land, water, and underground injection wells.<sup>e</sup>Total amount of chemical transferred off-site, including to publicly owned treatment works (POTW).

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should be used with caution since only certain facilities are required to report. This is not an exhaustive list.

Zinc has been identified in air at 36 sites collected from the 953 NPL hazardous waste sites where it was detected in some environmental media (HazDat 2003).

### 6.2.2 Water

Zinc and its compounds are found in the earth's crust and are present in most rocks, certain minerals, and some carbonate sediments. As a result of weathering of these materials, soluble compounds of zinc are formed and may be released to water (NAS 1977). The largest input of zinc to water results from erosion of soil particles containing natural traces of zinc (45,400 metric tons/year) (EPA 1980d). Erosion resulting from human activities accounts for 70% of this soil loss; geologic or natural erosion constitutes the other 30% (EPA 1980d). However, this source of low levels of zinc is widely dispersed and is, therefore, unlikely to elevate aquatic concentrations significantly. Zinc flux to the oceans from high temperature hydrothermal fluids in mid-ocean ridges has been estimated to be approximately 681,000 metric tons/year (WHO 2001).

Urban runoff, mine drainage, and municipal and industrial effluents are smaller but more concentrated sources of zinc in water. Davis et al. (2001) estimated the zinc loadings in urban stormwater runoff. In this study, buildings and automobiles were found to contribute 95% of loadings (0.646 kg/ha-year) to storm water runoff in urban environments. Data from this study are summarized in Table 6-3. The Nationwide Urban Runoff Program (NURP), initiated to evaluate the significance of priority pollutants in urban storm water runoff, reports a frequency of detection for zinc of 95%, with a concentration range of 0.01–2.4 mg/L (Cole et al. 1984). Industries that discharge large quantities of zinc directly to water include iron and steel, zinc smelting, plastics, and electroplating (EPA 1980d). The arithmetic mean concentration of zinc in influents of 239 waste water treatment plants in the United States was 0.7 mg/L, with minimum and maximum concentrations of 0.0001 and 28.7 mg/L, respectively (Minear et al. 1981). Accidental zinc discharges to water are most often associated with smelting and refining operations. Zinc is present with cadmium and lead in these processes (NAS 1977). Urban runoff and drainage from inactive mines account for approximately 5,250 and 4,060 metric tons/year, respectively, of the total releases of zinc to water (EPA 1980d). Drainage from active mining areas is considerably less than from inactive areas because of the disposal methods currently employed. Hazardous waste sites, in which zinc has been improperly disposed of, are additional sources of the element.



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**Table 6-3. Zinc Loadings in Urban Stormwater Runoff<sup>a</sup>**

	Rate	Unit value	Loading (kg/ha-year)	Percent of total
Buildings				
Siding	180,000 m <sup>2</sup> /ha-year	2,100 µg/m <sup>2</sup>		58
Roof	450,000 L/ha-year	100 µg/L		7
Total			0.423	65
Autos				
Brakes	240,000 km/ha-year	88 µg/km	0.021	3
Tires	48,000 g tire/ha-year	3,400 µg/g	0.163	25
Oil leakage	48 L-oil/ha-year	1.25x10 <sup>5</sup> mg/L	0.006	1
Total			0.190	29
Total buildings and autos			0.613	95
Wet deposition			0.013	2
Dry deposition			0.020	3
<i>Total</i>			0.646	100

<sup>a</sup>Source: Davis et al. (2001)

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Metals, such as zinc, also enter estuaries from many natural and man-made sources. Three important sources of zinc input into surface water are metal manufacturing (33,000–178,000 metric tons/year), domestic waste water (21,000–58,000 metric tons/year), and atmospheric fallout (2,600–31,000 metric tons/year). On an annual worldwide basis, an estimated 77,000–375,000 metric tons of zinc are discharged into water from anthropogenic sources (Nriagu and Pacyna 1988). Publicly owned treatment works are the largest total point source for zinc discharges. Publicly owned treatment works receive zinc contributions from the water supply and distribution system corrosion, combined sewer area runoff, industrial wastes, and human excrement (EPA 1980d). Crawford et al. (1995) reported the direct inputs of zinc contamination to the Newark Bay Estuary as follows (kg/day): municipal treatment systems, 272.0; industry, 14.34; combined sewer overflows, 141.5; storm water runoff, 164.6; and tributary flow, 307. Indirect inputs were 934.7 kg/day. The flux of zinc into the Hudson River Estuary from sewage has decreased from 924 kg/day in 1974 to 285 kg/day in 1997 as a result of improvements in controlling discharges from municipal and industrial waste water treatment plants since the Clean Water Act was enacted in 1972 (Sanudo-Wilhelmy and Gill 1999).

According to the TRI, estimated totals of 15,426 pounds (7 metric tons) of zinc (dust and fume) and 2,172,966 pounds (986 metric tons) of zinc compounds, amounting to about 0.044 and 0.287%, respectively, of the total environmental on-site releases, were discharged into surface water in the United States in 2001 from mining, manufacturing, processing, and electrical power generation industries listed in Tables 6-1 and 6-2. Estimated totals of 44,518 pounds (20 metric tons) of zinc (dust and fume) and 9,656,931 pounds (4,380 metric tons) of zinc compounds, amounting to about 0.13 and 1.28% of the total environmental on-site releases, respectively, were injected underground in the United States in 2001 from mining, manufacturing, processing, and Resource Conservation and Recovery Act (RCRA)/Solvent Recovery industries listed in Tables 6-1 and 6-2 (TRI01 2003). The TRI data should be used with caution since only certain facilities are required to report. This is not an exhaustive list.

The concentration of zinc in drinking water may increase as a result of the distribution system and household plumbing (EPA 1987c). Common piping materials used in distribution systems often contain zinc, as well as other metals and alloys. Trace metals may enter the water through corrosion products or simply by the dissolution of small amounts of metals with which the water comes in contact. Reactions with materials of the distribution system, particularly in soft low-pH waters, very often have produced concentrations of zinc in tap water much greater than those in the raw or treated waters at the plant of

## 6. POTENTIAL FOR HUMAN EXPOSURE

origin (NAS 1977). The total quantity of annual releases of zinc from these sources has not been estimated.

Zinc has been identified in surface water and groundwater at 385 and 668 sites, respectively, collected from the 953 NPL hazardous waste sites where it was detected in some environmental media (HazDat 2003).

### 6.2.3 Soil

Limited information is available on total releases of zinc to soil. Zinc is often present in soils and grasses as a result of atmospheric deposition. Furthermore, approximately 22,000 tons (20,000 metric tons) of zinc is used in fertilizers each year in the United States (NAS 1977). The extent to which zinc may run off into soil, rivers, and streams has not been evaluated. Hazardous waste sites are additional sources of zinc in soil. Municipal sludges applied to cropland soils can also be an important source of trace metals, including zinc (Chang et al. 1987).

On a worldwide basis, an estimated 1,193,000–3,294,000 metric tons of zinc per year are released to soil from anthropogenic sources (Nriagu and Pacyna 1988). The four most important sources of zinc in soil were estimated to be smelter slugs and wastes, mine tailings, coal and bottom fly ash, and the discharge of commercial products such as fertilizers.

Tire debris contains significant quantities of zinc, which may contaminate soils near roads. For example, snow collected on soil near an expressway in Montréal, Québec (Canada) contained higher levels zinc near the expressway. At 15 m from expressway, a snow pack concentration of 0.143 mg/L was measured, while at 150 m from the expressway, the concentration of zinc in snow was 0.029 mg/L (Loranger et al. 1996). Laboratory experiments indicated that a significant fraction of zinc may be released from tire rubber debris. Soil pH limits the mobilization of zinc in soil. Thus, zinc from tire debris will be less available and become immobile with soil interactions (Smolders and Degryse 2002).

Metallic zinc may yield soluble zinc compounds under acidic conditions where the zinc hydroxide-carbonate layer is attacked from pollutants such as sulfur dioxide. Metallic zinc is washed off slowly and forms a diffuse source of zinc release to soils. Other releases of zinc include the use of sacrificial anodes in soil to protect steel structures from corrosion (WHO 2001).

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According to the TRI, estimated totals of 33,695,443 pounds (15,300 metric tons) of zinc (dust and fume) and 738,610,608 pounds (335,000 metric tons) of zinc compounds, amounting to about 97 and 98%, respectively, of the total environmental on-site releases, were released to land in the United States in 2001 from mining, manufacturing, processing, and electrical power generation industries (Tables 6-1 and 6-2). Another 11,297,950 pounds (5,120 metric tons) of zinc (dust and fume) and 205,468,291 pounds (93,200 metric tons) of zinc compounds were transferred to off-site treatment, storage, and disposal facilities. The TRI data should be used with caution since only certain types of facilities are required to report. This is not an exhaustive list.

Zinc has been identified in soil and sediment at 508 and 358 sites, respectively, collected from the 953 NPL hazardous waste sites where it was detected in some environmental media (HazDat 2003).

### 6.3 ENVIRONMENTAL FATE

Zinc occurs in the environment mainly in the +2 oxidation state (Lindsay 1979). Sorption is the dominant reaction, resulting in the enrichment of zinc in suspended and bed sediments (EPA 1979d). Zinc in aerobic waters is partitioned into sediments through sorption onto hydrous iron and manganese oxides, clay minerals, and organic material. The efficiency of these materials in removing zinc from solution varies according to their concentrations, pH, redox potential (Eh), salinity, nature and concentrations of complexing ligands, cation exchange capacity, and the concentration of zinc. Precipitation of soluble zinc compounds appears to be significant only under reducing conditions in highly polluted water. Generally, at lower pH values, zinc remains as the free ion. The free ion ( $Zn^{+2}$ ) tends to be adsorbed and transported by suspended solids in unpolluted waters. In polluted waters in which the concentration of zinc is high, removal of zinc by precipitation of the hydroxide is possible, particularly when the pH is greater than 8 (EPA 1979d). In anaerobic environments and in the presence of sulfide ions, precipitation of zinc sulfide limits the mobility of zinc. The relative mobility of zinc in soil is determined by the same factors that affect its transport in aquatic systems (i.e., solubility of the compound, pH, and salinity) (Clement 1985).

Zinc is an essential nutrient that is present in all organisms. Although biota appears to be a minor reservoir of zinc relative to soils and sediments, microbial decomposition of biota in water can produce ligands, such as humic acids, that can affect the mobility of zinc in the aquatic environment through zinc precipitation and adsorption (EPA 1979d).

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Zinc concentrations in the air are relatively low, except near industrial sources such as smelters. No estimate for the atmospheric lifetime of zinc is available at this time, but the fact that zinc is transported long distances in air indicates that its lifetime in air is at least on the order of days.

### 6.3.1 Transport and Partitioning

*Air.* In the atmosphere, zinc exists primarily in an oxidized form bound to aerosols, with the size of zinc particulates determined by the source of zinc emission (Nriagu and Davidson 1980; Sweet et al. 1993). A major proportion of zinc released from industrial processes is adsorbed on particulates that are small enough to be in the respirable range (Dorn et al. 1976). Wind-blown dust transports zinc bound to soil particulates into the atmosphere (EPA 1980d). The particulates may also contain other materials (Pacyna et al. 1989; Saltzman et al. 1985).

Zinc-bearing particles in the atmosphere are transported to soil and water by wet deposition (rain and snow) and dry deposition (gravitational settling and deposition on water and soil surfaces). Zinc particles with low dry deposition velocities (i.e., particles with small diameters and low densities) can be transported from their emission source to distant regions (Pacyna et al. 1989). The atmospheric wet deposition of zinc (and other trace metals) was examined at two Maryland Chesapeake Bay sites from June 1990 to July 1991 as part of the Chesapeake Bay Atmospheric Deposition Study (Scudlark et al. 1994). The average annual wet deposition at these two sites was 1,335  $\mu\text{g}/\text{m}^2\text{-year}$  with 99% attributed to anthropogenic sources. As part of the Atmospheric Exchange over Lakes and Oceans Study (AEOLOS), dry deposition fluxes of zinc were measured over the southern basin of Lake Michigan near the urban area of Chicago and the non-urban area of South Haven, Michigan (Paode et al. 1998). In 1993, the average measured zinc fluxes were 200  $\mu\text{g}/\text{m}^2\text{-day}$  in Chicago; 10  $\mu\text{g}/\text{m}^2\text{-day}$  over southern Lake Michigan; and 4  $\mu\text{g}/\text{m}^2\text{-day}$  in South Haven, Michigan. Between 1993 and 1995, Shahin et al. (2000) estimated the dry deposition flux of zinc in Chicago to be  $4.4 \times 10^4$   $\mu\text{g}/\text{m}^2\text{-year}$ . Atmospheric deposition rates of zinc for Lakes Superior, Erie, and Ontario were reported to be 3,310, 2,180, and 5,650  $\mu\text{g}/\text{m}^2\text{-year}$ , respectively (Nriagu et al. 1996). Detection of zinc in rainwaters confirms the importance of wet precipitation in the removal of zinc particles from the atmosphere (Aten et al. 1983; Colin et al. 1990; Dasch and Wolff 1989; Golomb et al. 1997; Heaton et al. 1990). Golomb et al. (1997) measured the atmospheric deposition of zinc at Nahant, Massachusetts (near urban area of Boston) and at Truro, Massachusetts (on Cape Cod) for the years 1992–1993. Results indicated that wet deposition was a significant fraction of the total atmospheric deposition of zinc for Nahant (28%) and Truro (40%).

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**Water.** In water, zinc occurs in the environment primarily in the +2 oxidation state. It dissolves in acids to form hydrated  $Zn^{+2}$  cations and in strong bases to form zincate anions, which are hydroxo complexes, e.g.,  $[Zn(OH)_3]$ ,  $[Zn(OH)_4]^{2-}$ , and  $[Zn(OH)_4(H_2O)_2]^{2-}$  (O'Neil et al. 2001). In most waters, zinc exists primarily as the hydrated form of the divalent cation. However, the metal often forms complexes with a variety of organic and inorganic ligands (EPA 1979d, 1984b, 1987c).

Zinc can occur in both suspended and dissolved forms in surface water. Dissolved zinc may occur as the free (hydrated) zinc ion or as dissolved complexes and compounds with varying degrees of stability. Suspended (i.e., undissolved) zinc may be dissolved with changes in water conditions (e.g., pH, Eh, solution speciation) or may sorb on to suspended matter. Gundersen and Steinnes (2003) reported that <10% of zinc was sorbed on particles or colloids in river water from two rivers with average pHs of 3.1 and 5.1 (rivers with mining activity near Roes, Norway), whereas 21% of zinc occurred in sorbed form in six pH neutral rivers.

In the aquatic environment, zinc partitions to sediments or suspended solids in surface waters through sorption onto hydrous iron and manganese oxides, clay minerals, and organic material. Reservoirs located downstream from lead-zinc mining and milling areas were found to contain higher concentrations of zinc than reservoirs in other areas, and the zinc was more highly concentrated in reservoir bottom sediments than in the surrounding soils (Pita and Hyne 1975). In addition, the zinc content in sediment closely correlated with the depth, organic content, and clay content of the sediments. Phosphates and iron hydroxides affect the transfer of metals (including zinc) from river water to the sediments, according to a study by Houba et al. (1983). In this study, zinc was bound predominantly to carbonate and amorphous matter (iron, aluminum, and manganese hydroxides). In addition, mobile components of naturally occurring organic matter contributed to the increase in the metal hydroxide-bound fraction.

The transport of zinc in the aquatic environment is controlled by anion species. In natural waters, complexing agents, such as humic acid, can bind zinc. The stability of zinc complexes depends on the pH of the water and the nature of the complex. The dissociation of the complex may determine the amount of free zinc ions in solution. Zinc-humic acid complexes may be 50% dissociated at pH 5.5 and the dissociation rate may be higher as the pH decreases (Guy and Chakrabarti 1976). Therefore, as the pH of the water decreases, the concentration of zinc ions in the water phase increases at the same rate as that of the release of zinc from the sediment. The magnesium found in the silicate minerals of igneous rocks is often replaced with the divalent zinc ion; consequently, weathering of this zinc-containing bedrock gives rise to  $Zn^{+2}$  in solution. The hydrated cation is the dominant form when the pH is  $\leq 9$  (EPA 1979d).

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The tendency of zinc to be sorbed is affected not only by the nature and concentration of the sorbent but also by pH and salinity (EPA 1979d). Zinc tends to sorb more readily at a high pH ( $\text{pH} > 7$ ) than at a low pH (EPA 1979d). Desorption of zinc from sediments occurs as salinity increases (Helz et al. 1975), apparently because of displacement of the adsorbed zinc ions by alkali and alkaline earth cations, which are abundant in brackish and saline waters (EPA 1979d). In column leaching tests with sediment collected from the banks of the Rhone River, the presence of dissolved organic matter and pH was found to be the factors controlling the adsorption and mobility of zinc (Bourg and Darmendrail 1992).

A small fraction of zinc will exist in the aquatic phase as soluble inorganic zinc compounds (e.g., zinc chloride, zinc sulfate). Soluble inorganic zinc compounds hydrolyze in solution, forming zinc hydroxide precipitates. Hydrolysis may lower pH, but the buffering action present in most natural water prevents a significant alteration in pH. The precipitation of zinc hydroxide and zinc carbonate was studied by Patterson et al. (1977), who found that zinc hydroxide precipitates faster than zinc carbonate. Zinc carbonate is soluble in pure water at 25 °C at concentrations of  $\leq 107$  mg zinc/L. The hydroxide is soluble only at concentrations of  $\leq 0.2$  mg zinc/L. As a result, some of the inorganic forms of zinc that are expected to be present in water are basic carbonate ( $\text{Zn}_2[\text{OH}]_2\text{CO}_3$ ), hydroxide ( $\text{Zn}[\text{OH}]_2$ ), and silicate ( $\text{Zn}_2\text{SiO}_4$ ) (Florence 1980; NAS 1977). When the pH is  $\geq 8$ , most of these compounds will precipitate; however, as the pH decreases, more and more of these compounds will dissolve and remain in the water phase (EPA 1979d).

The effect of pH on the mobilization of zinc in a few highly acidic clean lakes has been studied (Sprenger et al. 1987; White and Driscoll 1987). In these lakes, in which the pH was  $\leq 3.6$ , concentrations of zinc were elevated in the water column, and the concentration of zinc in the upper layer of sediment was substantially lower than values reported for other lakes at higher pH values. The relatively higher concentration of zinc in the water column compared to the sediment may be the result of lower adsorption of zinc on oxide surfaces due to low pH, solubilization of inorganic zinc from the sediment layer, and the dissociation of bound organic complexes of zinc present in the sediment and their subsequent release into the water phase.

The precipitation of zinc sulfide affects the mobility of zinc in reducing environments, especially when hydrogen sulfide is formed. The precipitation of the hydroxide, carbonate, or basic sulfate may become more significant at high concentrations of zinc. Hesterberg et al. (1997) reports that zinc (hydr)oxides and not sulfides are the dominant species in aquifers solids under reducing conditions. The hydroxides

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and hydrous oxides of iron and manganese are often components of the clay fraction of sediments and often exist as coatings on the surfaces of other minerals (NAS 1977). Zinc may coprecipitate with hydrous oxides when reduced iron or manganese oxides are oxidized. As the new solids are formed, they can trap various ions in their crystal lattices (EPA 1979d).

**Soil.** The redox status of the soil may shift zinc partitioning. Reductive dissolution of iron and manganese (hydr)oxides under suboxic conditions release zinc into the aqueous phase; the persistence of suboxic conditions may then lead to a repartitioning of zinc into sulfide and carbonate solids. Bostick et al. (2001) describe zinc speciation in contaminated wetland soil that undergoes seasonal flooding. In dry oxidized soils, the authors found that zinc was associated with (hydr)oxide phases, while in flooded systems, zinc was associated with sulfides and carbonates. Reversible change occurred with flooding from dry soil. However, a small fraction of zinc became recalcitrant with (hydr)oxides fraction.

Zinc sorbs strongly onto soil particulates. Little water-soluble and exchangeable heavy metals were found in soil irrigated with raw waste water (Schalscha et al. 1982). Although considerable amounts of metals were added to the soil in soluble and exchangeable forms during waste-water irrigation, they were converted into the less chemically active forms (i.e., organically bonded and inorganic precipitates). Further examination showed that zinc accumulation in soil resulting from waste disposal occurred primarily as inorganic precipitates.

The mobility of zinc in soil depends on the solubility of the speciated forms of the element and on soil properties such as cation exchange capacity, pH, redox potential, and chemical species present in soil; under anaerobic conditions, zinc sulfide is the controlling species (EPA 1980d; Kalbasi et al. 1978). Since zinc sulfide is insoluble, the mobility of zinc in anaerobic soil is low. In a study of the effect of pH on zinc solubility, Saeed and Fox (1977) showed that, when the pH is <7, an inverse relationship exists between the pH and the amount of zinc in solution. As negative charges on soil surfaces increase with increasing pH, additional sites for zinc adsorption are activated and the amount of zinc in solution decreases. The active zinc species in the adsorbed state is the singly charged zinc hydroxide species (i.e.,  $Zn[OH]^+$ ) (Sanders and Kherbawy 1987). Other investigators have also shown that the mobility of zinc in soil increases at lower soil pH under oxidizing conditions and at a lower cation exchange capacity of soil (Bergkvist et al. 1989; Hermann and Neumann-Mahlkau 1985; Tyler and McBride 1982). On the other hand, the amount of zinc in solution generally increases when the pH is >7 in soils high in organic matter. This is a result of the release of organically complexed zinc, reduced zinc adsorption at higher pH, or an increase in the concentration of chelating agents in soil (Saeed and Fox 1977). For calcareous soils, the



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relationship between zinc solubility and pH is nonlinear. At a high pH, zinc in solution is precipitated as  $Zn(OH)_2$ , zinc carbonate ( $ZnCO_3$ ), or calcium zincate (Saeed and Fox 1977). Clay and metal oxides are capable of sorbing zinc and tend to retard its mobility in soil. Warwick et al. (1998) studied the mobility of zinc ions in sand. Zinc was more mobile at pH 4 than at pH 6.5 as a consequence of sorption. Goethite (i.e., iron oxyhydroxide) caused a greater decrease in mobility, and increased retardation was also observed with humic acid.

Distribution constants ( $K_d$  = concentration of sorbed zinc/concentration of zinc in solution) for zinc in soil range widely from 0.1 to 8,000 L/kg (or mL/g) (Baes and Sharp 1983; Bunzl and Schimmack 1989; Gao et al. 1997; Gerritse et al. 1982; Janssen et al. 1997).  $K_d$  values of  $100 \pm 770$  mL/g for sandy loam soil and  $0.2 \pm 4$  mL/g for sandy soils were reported by Gerritse et al. (1982).  $K_d$  values ranging from 0.1 to 8,000 mL/g were reported by Baes and Sharp (1983).  $K_d$  values for zinc of 140 and 41 L/kg were determined for the O-horizon (organic layer) and E-horizon (silty sand), respectively, of a podzol forest soil (Bunzl and Schimmack 1989). Field-based  $K_p$  ranged from 6 to 6,762 L/kg for 20 Dutch agricultural soils (Janssen et al. 1997).  $K_d$  values for nine soils treated with sewage sludge supernatant ranged from 0.034 to 1.359 L/g at pH 4.5 while at pH 6.5,  $K_d$  values ranged from 0.425 to 2.896 L/g (Gao et al. 1997).

Zinc in a soluble form (e.g., zinc sulfate) is moderately mobile in most soils. However, relatively little land-disposed zinc at waste sites is in the soluble form. Thus, mobility is limited by a slow rate of dissolution. Consequently, movement towards groundwater is expected to be slow unless zinc is applied to soil in soluble form (such as in agricultural applications) or accompanied by corrosive substances (such as in mine tailings) (EPA 1980d). Yet, soil conditions not suitable for zinc sorption may lead to leaching. Low pH (pH <7) and high ionic strength of the leaching solution favor desorption (EPA 1987c; Saeed and Fox 1977).

Consequently, zinc primarily remains in recalcitrant, immobile forms in contaminated soils (Chlopecka et al. 1996; Kabala and Singh 2001; Kaminiski and Landsberger 2000b; Ma and Rao 1997a). Ma and Rao (1997) studied the chemical fractionation of zinc in nine soils from various U.S. locations contaminated by agriculture and industrial activities. Zinc was found to be concentrated in the residual (or recalcitrant) fraction (range, 55.8–97.6%), which reflected greater tendency of zinc to become unavailable once in soils. However, some zinc was found in exchangeable and carbonate fractions at levels ranging from 0.73 to 25%, which suggests that some zinc may be available to plants. In soil from East St. Louis, Illinois, an area heavily contaminated with metals, sequential extraction analysis of soil revealed that the largest fraction of zinc was partitioned in the iron-manganese oxide fraction (47.2%) followed by

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carbonate (37.9%) and organic (14.1%) and exchangeable (0.8%) fractions (Kaminiski and Landsberger 2000b). Kabala and Singh (2001) reported that water-soluble zinc was present in only very small amounts (<1%) in most contaminated soils. However, concentrations of exchangeable zinc were significantly higher in surface horizons (4–19% of total zinc) than in subsurface layers. In subsurface horizons of the studied soils, zinc was concentrated in the residual (or recalcitrant) fraction. The percentage of residual zinc ranged from 45% in silty soils to 94% in clay-loam soil. The non-residual fractions prevailed only in the surface horizons in both contaminated and uncontaminated soil (65–91% of total zinc) (Kabala and Singh 2001). Soils from Southwestern Poland subjected to severe metal contamination contained zinc at concentrations ranging from 20 to 10,000 mg/kg (Chlopecka et al. 1996). Acidic soils (pH<5.6) contained a greater fraction of exchangeable zinc, while for other soils (pH>5.6), zinc was found primarily in the oxide and residual (or recalcitrant) fractions with moderate amounts in the organic, carbonate, and exchangeable forms.

Zinc is an essential nutrient and occurs in the tissues of organisms, even at normal ambient water and soil concentrations. Zinc can accumulate in freshwater animals at 51–1,130 times the concentration present in the water (EPA 1987c). Microcosm studies indicate, in general, that zinc does not biomagnify through food chains (Biddinger and Gloss 1984; EPA 1979d; Hegstrom and West 1989). Furthermore, although zinc actively bioaccumulates in aquatic systems, biota appears to represent a relatively minor sink compared to sediments. Steady-state zinc bioconcentration factors (BCFs) for 12 aquatic species range from 4 to 24,000 (EPA 1987c). Crustaceans and fish can accumulate zinc from both water and food. A BCF of 1,000 was reported for both aquatic plants and fish, and a value of 10,000 was reported for aquatic invertebrates (Fishbein 1981). The order of enrichment of zinc in different aquatic organisms was as follows (zinc concentrations in  $\mu\text{g/g}$  dry weight appear in parentheses): fish (25), shrimp (50), mussel (60), periphyton (260), zooplankton (330), and oyster (3,300) (Ramelow et al. 1989). The high enrichment in oysters may be due to their ingestion of particulate matter containing higher concentrations of zinc than ambient water. Other investigators have also indicated that organisms associated with sediments have higher zinc concentrations than organisms living in the aqueous layer (Biddinger and Gloss 1984). With respect to bioconcentration from soil by terrestrial plants, invertebrates, and mammals, BCFs of 0.4, 8, and 0.6, respectively, have been reported. The concentration of zinc in plants depends on the plant species, soil pH, and the composition of the soil (Dudka and Chlopecka 1990; Rudd et al. 1988). Plant species do not concentrate zinc above the levels present in soil (Levine et al. 1989).

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**6.3.2 Transformation and Degradation**

As an element, zinc does not degrade in the environment. Degradation of an element is a nuclear process by definition, and stable elements, such as zinc, typically undergo such processes only at insignificant rates in the environment. Zinc can change from one form to another, sometimes reversibly, in numerous chemical reactions that can proceed under a wide range of common environmental conditions.

**6.3.2.1 Air**

The chemical interaction of zinc compounds in the atmosphere may change the anionic speciation of the compound. Atmospheric interactions are greatest for particles with small aerodynamic diameters (Fishbein 1981). Zinc is found in the atmosphere at the highest concentrations in the smallest particles (Fishbein 1981). Atmospheric emissions of zinc, consisting primarily of zinc sorbed to submicron particulate matter in the form of zinc oxide (ZnO), are expected to dissipate quickly as a result of deposition to soil and surface waters (EPA 1980d).

In the atmosphere, zinc-bearing particles may undergo chemical transformation before deposition. The association of zinc particles in aerosols in Arizona was studied, and five zinc-bearing particles were identified with an automated scanning electron microscope (Anderson et al. 1988). These particles, in decreasing order of concentration in the aerosol, were zinc sulfide (ZnS), ferrous zinc ( $\text{Fe}_x\text{Zn}_y$ ), zinc phosphides ( $\text{Zn}_3\text{P}_2$ ), zinc chloride ( $\text{ZnCl}_2$ ), and metallic zinc (Zn). The presence of zinc sulfide in an area adjacent to mining and smelting activities was not unanticipated, but no conclusion regarding the speciation of zinc in the atmosphere could be drawn from this investigation. However, the relative concentration of zinc ions in rainwater from a rural area was approximately 10 times higher than in airborne particulates (Aten et al. 1983). This finding suggests that zinc sulfide in the atmosphere is oxidized to a more water-soluble form, zinc sulfate.

**6.3.2.2 Water**

Zinc is in the +2 form in aqueous solution and exhibits amphoteric properties; zinc metal and compounds dissolve in acids to form hydrated  $\text{Zn}^{+2}$  cations and in strong bases to form zincate anions, which are hydroxo complexes, such as  $[\text{Zn}(\text{OH})_3]^-$ ,  $[\text{Zn}(\text{OH})_4]^{2-}$ , and  $[\text{Zn}(\text{OH})_4(\text{H}_2\text{O})_2]^{2-}$  (EPA 1979d; O'Neil et al. 2001). However, at the pH of most natural waters, the formation of anionic zinc species is not likely.

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A small part of the available zinc may partition into the aquatic phase through the formation of soluble zinc chloride and sulfate compounds. These compounds hydrolyze in solution to form the hydroxide or hydrated zinc oxide precipitate with a resultant decrease in pH. The decrease in pH can increase the solubility of zinc hydroxide and increase the zinc concentration in water. However, the buffering action of most natural waters prevents any significant change of pH due to the hydrolysis reactions. As a result, in the water phase, the solubility of its carbonate and hydroxide is likely to control the availability of zinc. It was reported by Patterson et al. (1977) that  $Zn(OH)_2$  precipitates faster than  $ZnCO_3$ . Zinc is not directly affected by changes in Eh; however, the valences and reactivity of ligands reacting with zinc are affected by Eh. Zinc is an active reducing agent for many metal ions such as iron ( $Fe^{+3}$ ) and permanganate ( $MnO_4^{-2}$ ) ions (Stokinger 1981). As a result of the reducing reactions, the manganese oxides and ferric salts may precipitate and, in the process, may entrap soluble zinc in the precipitate, thereby reducing the zinc concentration in the water phase.

Because alkyl zinc compounds are unstable in water and oxygen, biomethylation of zinc compounds in aquatic ecosystems probably does not occur (EPA 1979d). Insoluble zinc compounds (e.g., zinc oxide) are solubilized indirectly under anaerobic conditions with reduction of iron sulfides, which reduces the solution pH (Couillard et al. 1994; Francis and Dodge 1988). No evidence was found that photolysis in the aquatic environment significantly affects the fate of zinc compounds.

### 6.3.2.3 Sediment and Soil

Zinc undergoes reactions in sediment and soil involving precipitation/dissolution, complexation/dissociation, and adsorption/desorption. These reactions are controlled by the pH, redox potential (Eh), the concentration of zinc ions and other ions in the soil pore water, the number and type of adsorption sites associated with the solid phase, and the organic ligands present that are capable of forming complexes with zinc. In acidic sediments and soils, more zinc is available in ionic forms, and cation exchange processes influence its fate. Depending on the nature and concentrations of other mobile metals in sediments and soils, competition for the binding sites probably occurs. In the absence of suitable binding sites, zinc may be mobilized (ICF 1986). In alkaline soils, the chemistry of zinc is dominated by interactions with organic ligands.

Biological degradation of zinc complexes in soil is necessary for the normal operation of ecosystems to facilitate the recycling of zinc from litter, feces, and dead organisms. In some environments, bacteria and

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fungi are able to oxidize zinc sulfide producing zinc sulfate, which will solubilize in the soil solution (WHO 2001).

#### 6.3.2.4 Other Media

During composting of organic wastes, zinc remains in mobile and bioavailable forms. Zinc carbonates are formed, although not at the expense of zinc sulfide levels, which remain unaltered during the composting process (Ciba et al. 1997). No further data were located in the literature for the transformation and degradation of zinc in other media.

### 6.4 LEVELS MONITORED OR ESTIMATED IN THE ENVIRONMENT

#### 6.4.1 Air

Zinc concentrations in air are relatively low and fairly constant except near sources such as smelters. Average atmospheric concentrations of zinc resulting from releases from automobiles, fuel combustion, incineration, soil erosion, and industrial, commercial, and construction activity throughout the United States generally are  $<1 \mu\text{g}/\text{m}^3$  (EPA 1980d; Lloyd and Showak 1984). In 1990, the median concentration of zinc in air samples collected across Minnesota was  $0.012 \mu\text{g}/\text{m}^3$  (maximum,  $0.187 \mu\text{g}/\text{m}^3$ ) (Pratt et al. 2000). At six measurement sites in Columbus, Ohio, the mean atmospheric particulate concentration of zinc for samples collected in 1989 was  $0.01 \pm 0.01 \mu\text{g}/\text{m}^3$  (Spicer et al. 1996). Data on zinc concentrations in New York City during 1972–1975 show that the average atmospheric zinc concentration ranged from  $0.293$  to  $0.380 \mu\text{g}/\text{m}^3$  annually (Lioy et al. 1978). An average ambient zinc concentration of  $0.127 \mu\text{g}/\text{m}^3$  (concentration range,  $0.027$ – $0.500 \mu\text{g}/\text{m}^3$ ) was determined from analyses of particulate samples collected at nine air monitoring sites in the San Francisco Bay area (John et al. 1973). The concentrations of zinc in atmospheric samples collected from seven cities in the United States during 1968–1971 ranged from  $0.17$  to  $0.67 \mu\text{g}/\text{m}^3$ , whereas the concentrations at two rural sites ranged from  $0.02$  to  $0.16 \mu\text{g}/\text{m}^3$  (Saltzman et al. 1985). The concentrations of zinc during 1977–1979 from the National Air Surveillance Networks were reported by Evans et al. (1984). The arithmetic mean zinc concentrations in urban areas in the United States ranged from  $0.02$  to  $0.16 \mu\text{g}/\text{m}^3$ , whereas the concentrations in non-urban areas ranged from  $0.01$  to  $0.05 \mu\text{g}/\text{m}^3$ . The geometric mean concentrations of zinc from three urban areas in New Jersey monitored in 1981–1982 ranged from  $0.07$  to  $0.59 \mu\text{g}/\text{m}^3$ , whereas the concentrations at a rural site ranged from  $0.02$  to  $0.06 \mu\text{g}/\text{m}^3$  (Daisey 1987). Davidson et al. (1988) measured the

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atmospheric zinc concentrations at Great Smoky Mountains and Olympic National Parks where crustal weathering, sea spray, and long-range transport of zinc were likely to influence concentrations. The average atmospheric concentrations of zinc were 0.0033 and 0.0089  $\mu\text{g}/\text{m}^3$  for Great Smoky Mountains in 1979 and Olympic National Parks in 1980, respectively. The reported concentration range of zinc in air at remote sites (arctic) was <0.003–0.027  $\mu\text{g}/\text{m}^3$  (Barrie and Hoff 1985; Duce et al. 1975; Zoller et al. 1974). In aerosol samples of the lower troposphere collected over the Southern Bight of the North Sea between September 1988 and October 1989, the average zinc concentration was 0.067  $\mu\text{g}/\text{m}^3$  (standard deviation, 0.054  $\mu\text{g}/\text{m}^3$ ; range, 0.003–0.22  $\mu\text{g}/\text{m}^3$ ; n=108 samples) (Injuk et al. 1992). The concentration of atmospheric zinc is usually lower in winter than in summer (Barrie and Hoff 1985; Daisey 1987).

Indoor air from other regions of the world have been reported to contain zinc in particulate matter at low levels. In 1991, household dust sampled from Bahrain in Persian Gulf region contained zinc at a concentration of 64.4  $\mu\text{g}/\text{g}$  (Akhter and Madany 1993). As part of the Southeastern Brazil Indoor Air Quality Study (SEBIAQS) in the summer of 1993, indoor and outdoor air samples were collected at 12 sites in the cities of São Paulo and Rio de Janeiro, Brazil. Indoor air particulate samples had higher levels of zinc than outdoor samples. The concentration of zinc in indoor air particulates from São Paulo and Rio de Janeiro, Brazil were 0.046–0.30 and 0.036–0.38  $\mu\text{g}/\text{m}^3$ , respectively, while outdoor samples ranged from not detected to 0.23 and from 0.05 to 0.29  $\mu\text{g}/\text{m}^3$ , respectively (Miguel et al. 1995).

Although data are sparse, higher-than-background concentrations have been reported near iron- and steel-producing factories and zinc, lead, and copper smelters. During zinc smelting operations, concentrated zinc ore goes through a roasting procedure to convert zinc sulfide to zinc oxide. This process accounts for a large portion of the total atmospheric zinc emission during primary production (EPA 1980d). About 1.5 miles from a smelter in Kellogg, Idaho, Ragaini et al. (1977) detected high annual mean concentrations of zinc in ambient air of 5  $\mu\text{g}/\text{m}^3$ . The 24-hour values for zinc ranged from 0.27 to 15.7  $\mu\text{g}/\text{m}^3$ ; the average lead and cadmium concentrations at this smelter site were 11 and 0.8  $\mu\text{g}/\text{m}^3$ , respectively, indicating severe environmental pollution. Higher concentrations of zinc in the vicinity of a copper smelter than in reference areas were also reported by Patterson et al. (1977).

#### 6.4.2 Water

In general, zinc is more concentrated in the sediments of streams and rivers than in the water column. It is reported by NAS (1977) that zinc is probably detectable in 75% of all water samples from various locations. The zinc background concentrations in surface waters are usually <50  $\mu\text{g}/\text{L}$  (EPA 1980d), but

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concentrations in surface waters and groundwater can range from 0.002 to 50 mg/L (NAS 1977). Table 6-4 summarizes the typical concentrations of dissolved zinc in rivers of the United States (Shiller and Boyle 1985). The concentration of dissolved zinc in water from three Adirondack lakes was highest for low pH waters (Heit et al. 1989). Lake water from Darts Lake (pH 5.1–5.4) contained 7.9 ng zinc/mL while Moss Lake, a lake with variable acidity (pH 5.8–6.7), contained zinc at 2.9 ng/mL and Rondoxe Lake, a neutral lake (pH 6.5–6.8), contained 2.5 ng zinc/mL.

In many locations (e.g., New England, the southeast, the Missouri River basin, the Rio Grande River basin, and the Upper Colorado River basin), higher-than-background concentrations of zinc are common and appear to be correlated with mining activities in these areas and/or geological areas rich in zinc (EPA 1980d). However, in all river basins, there are some locations with zinc concentrations of 0.1–1.0 mg/L (EPA 1980d). In the Upper Rio Grande River, the dissolved zinc concentration upriver of Willow Creek, which drains a metal mining district, was approximately 2–3 µg/L. Immediately downstream of the Willow Creek confluence, zinc concentrations were >20 µg/L and elevated concentrations occurred for the next 100 km (Taylor et al. 2001). The concentrations of zinc in water samples from Whitewood Creek, South Dakota, were measured by Hale (1977). The samples were collected upstream from the discharge of a local mining company. In 42 analyses, zinc concentrations ranged from <0.004 to 0.048 mg/L with a mean concentration of 0.018 mg/L. May et al. (2001) reported that water samples analyzed in 1996 from this contaminated watershed contained zinc at concentrations of 2.4–21 and 3.8–30 µg/L for filtered and unfiltered samples, respectively.

The average levels of dissolved zinc in water from Lakes Superior, Erie, and Ontario were 277, 87, and 160 ng/L, respectively, for samples collected between 1991 (Lake Superior) and 1993 (Lakes Ontario and Erie) (Nriagu et al. 1996). Coale and Flegal (1989) reported that concentration of dissolved zinc in water from Lakes Erie and Ontario ranged from  $3 \times 10^{-6}$  to  $1.1 \times 10^{-4}$  mg/L. In summer, there is a marked depletion of zinc in the epilimnion (i.e., area of warmest water) of Great Lakes off-shore waters, which may be attributed to biological processes (Nriagu et al. 1996). Groundwater from a shallow alluvial aquifer beneath a major urban center (Denver, Colorado) contained dissolved zinc at a median concentration of 3 µg/L (range, 2–28 µg/L) (Bruce and McMahon 1996).

Scudlark et al. (1994) reported that the average concentration of zinc in water from the Chesapeake Bay was  $1.21 \pm 0.95$  µg/L (n=5) for samples collected from 1990 to 1991. The median concentration of zinc in the Hudson River estuary decreased from 200 nM (1.6 ng/L) in 1974 to approximately 25 nM (13 ng/L) in 1997 (Sañudo-Wilhelmy and Gill 1999). The declining levels of zinc and other metals is a result of the

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**Table 6-4. Dissolved Zinc in Rivers of the United States<sup>a</sup>**

River <sup>b</sup>	Date <sup>c</sup>	pH	Zinc (µg/L at 20 °C) <sup>d</sup>
<u>Ohio Valley</u>			
Allegheny (Pittsburgh, PA)	May 1984	6.86	1.89, 2.02
Monongahela (Pittsburgh, PA)	May 1984	7.22	2.54, 2.87
Ohio (Wheeling, WV)	May 1984	7.34	3.19, 3.19
Muskingum (Zanesville, OH)	May 1984	7.55	1.04, 1.24
Muskingum (Marietta, OH)	May 1984	7.66	0.63, 0.63
Kanawha (Winfield, WV)	May 1984	7.4	0.33, 0.35
Big Sandy (Louisa, KY)	May 1984	7.15	0.33, 0.33
Ohio (Greenup Dam)	May 1984	7.42	0.78, 0.91
Sciotto (Portsmouth, OH)	May 1984	7.87	1.30, 1.43
Little Miami (Milford, OH)	May 1984	8.1	0.85, 1.04
Licking (Alexandria, KY)	May 1984	7.63	0.07
Great Miami (Cleves, OH)	May 1984	8.0	4.24, 4.30
Whitewater (Elizabethtown, OH)	May 1984	7.95	0.16
Ohio (Warsaw, KY)	May 1984	7.45	0.39, 0.42
Kentucky (Lockport, KY)	May 1984	7.28	0.12, 0.15
Ohio (Cannelton Dam)	May 1984	7.27	0.61
Green (Beach Grove, KY)	May 1984	7.32	0.16, 0.16
Wabash (New Harmony, IN)	May 1984	8.1	0.49
Cumberland (Barkley Dam)	May 1984	7.44	0.10, 0.10
Tennessee (Kentucky Dam)	May 1984	7.10	0.12, 0.12
Ohio (Mound City, IL)	May 1984	7.49	0.29, 0.29
<u>Mississippi River</u>			
Mississippi (Cape Girardeau, MO)	May 1984	7.70	0.19, 0.23
Mississippi (Baton Rouge, LA)	Sept 1983	8.1	0.10, 0.12
Mississippi (Baton Rouge, LA)	Apr 1984	7.72	0.18, 0.19
Atchafalaya (Krotz Springs, LA)	Apr 1984	7.6	0.18
<u>Other U.S. rivers</u>			
Connecticut (Old Saybrook, CT)	Apr 1983	7.1	0.91, 1.04
Mullica	Aug 1983	5.81	2.54, 2.61
Merrimack (West Newbury, MA)	Feb 1983	6.73	13.04, 13.04
Vermilion, (Lafayette, LA)	Apr 1984	7.1	0.25, 0.27
Delaware (West Trenton, NJ)	Apr 1984	7.38	3.91, 3.98
Delaware (Philadelphia, PA)	Apr 1984	7.05	13.04, 15.65
Schuykill (Philadelphia, PA)	Apr 1984	7.58	4.56, 4.89
Susquehanna (Holtwood, PA)	Apr 1984	7.54	0.78, 0.85
Potomac (Great Falls, MD)	Apr 1984	7.75	0.54, 0.55

<sup>a</sup>Source: Shiller and Boyle (1985)<sup>b</sup>Post office state abbreviations are used.<sup>c</sup>Apr = April; Aug = August; Feb = February; Sept = September<sup>d</sup>Calculated from nmol/kg using density of water at 20 °C (0.99707 g/mL) and zinc molecular weight of 65.39 g/mole.



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decreased metal flux to the estuary from sewage effluents. Zinc concentrations in remote regions of the Atlantic Ocean ranged from 0.023 to 0.097  $\mu\text{g/L}$  in the Northeast region and averaged 0.004  $\mu\text{g/L}$  in the Northwest region (Helmers and Schrens 1995). Yeats (1988) reported that the concentrations of dissolved zinc in ocean water from the Sargasso Sea and Northeast Pacific were 0.3–3.0 nM (20–200 ng/L) in 1984 and 3.6–9.2 nM (240–600 ng/L) in 1981, respectively. Seawater from lagoons of the Gulf of Mexico contained average dissolved zinc concentrations of 2.37, 5.12, and 9.76  $\mu\text{g/L}$  for locations at Alvarado, San Andres, and Sedue (in Mexico), respectively (Vazquez et al. 1995). The concentration of zinc in surficial seawater from the Indian River lagoon (Florida) ranged from 0.01 to 6.6  $\mu\text{g/L}$  (average,  $0.8 \pm 1.4 \mu\text{g/L}$ ) in 1992 (Trocine and Trefry 1996).

Zinc concentration in precipitation from remote regions of the Atlantic Ocean ranged from 0.359 to 3.93  $\mu\text{g/L}$  (Helmers and Schrens 1995). Heaton et al. (1990) reported that precipitation collected from three locations Rhode Island between 1985 and 1988 contained zinc at median concentration of 4.5 ppb ( $n=269$ ). Levels of zinc were higher in samples collected in warm periods (5.8 ppb) versus cold periods (3.7 ppb). Nearly all zinc was dissolved in these samples. Trace amounts of zinc were measured in cloud water ( $n=3$ ; range,  $<10\text{--}43 \mu\text{g/L}$ ) collected from Whiteface Mountain (Adirondacks Region, New York) in the summer of 1987 (Khwaja et al. 1995). Municipal waste incineration was the primary source of zinc in these wet deposition samples. Snow near an expressway in Montréal, Québec, Canada, contained zinc at average concentrations of 0.143, 0.33, 0.034, and 0.029 mg/L at 15, 20, 125, and 150 meters from the roadway, respectively (Loranger et al. 1996). Higher zinc concentrations near expressway were the result of road dust from tire abrasion.

Available data suggest that zinc concentrations in drinking water are far less than levels required to meet a daily intake level of 11 mg/day (assuming an adult water consumption of 2 L/day) (IOM 2002).

Concentrations of zinc in drinking water can be higher than levels in surface waters. Concentrations of 0.002–1.2 mg/L were detected in 77% of 1,577 surface water samples while levels of 0.003–2.0 mg/L were found in 380 drinking water samples (NAS 1977). Higher concentrations in drinking waters are a result of water treatment and of contamination from plumbing of the water distribution system.

Zinc was found in drinking water at levels as great as several mg/L as a result of galvanized pipes and tanks in alkaline-water distribution systems. Drinking water samples from galvanized pipe plumbing systems in Seattle, Washington, contained zinc concentrations of 0.128–1.279 mg/L; these levels were >10 times higher than those in homes with copper pipe plumbing systems (Sharrett et al. 1982a). Forty-three tap-water samples collected from homes in Dallas, Texas and analyzed for trace metals reported

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maximum, minimum, median, and average zinc concentrations of 0.049, 0.005, 0.011, and 0.0124 mg zinc/L, respectively (NAS 1977). High zinc concentrations in these water samples were believed to be due to the household plumbing. In a study investigating associations between inorganic constituents of drinking water and cardiovascular diseases, Greathouse and Osborne (1980) collected and analyzed tap water samples in 35 geographic areas in the United States; 100 to 110 tap-water samples were collected from each area. The maximum, minimum, and mean concentrations were 1.447, 0.025, and 0.144 mg zinc/L, respectively. Seventy-five percent of the zinc values were below 0.236 mg/L. Other investigators have attributed the higher concentrations of zinc in household tap waters, compared to finished drinking water, to distribution and transmission lines (Maessen et al. 1985; Ohanian 1986; Schock and Neff 1988).

The median concentration of zinc in leachate from municipal landfills in the United States ranged from 0.68 to 1.7 mg/L with a high concentration of 250 mg/L (Roy 1994).

#### 6.4.3 Sediment and Soil

Zinc is found in soils and surficial materials of the conterminous United States at concentrations between <5 and 2,900 mg/kg, with a mean of 60 mg/kg (Schacklette and Boerngen 1984). Zinc concentrations measured across the United States ranged from <5 to 400 mg/kg and from <10 to 2,000 mg/kg, with corresponding means of 36 and 51 mg/kg in cultivated and uncultivated subsurface soils, respectively (Connor and Shacklette 1975); however, these differences in zinc concentration may be attributed to differences in the soils prior to use (and not to cultivation). The sampling survey was designed to determine zinc concentrations of surficial materials unaltered from their natural condition. Chen et al. (1999) determined the baseline concentration of zinc in 448 representative Florida surface soils as part of the Florida Cooperative Soil Survey Program. Baseline soil samples represent natural elemental concentrations without human influence. The mean concentration of zinc was  $8.35 \pm 13.8$  mg/kg (range, 0.9–169 mg/kg) in archived soil samples from this study.

Soils near highways and smelters contained high zinc concentrations as a result of deposition of zinc released in tire abrasion and stack emissions (EPA 1980d; Norrström and Jacks 1999). Urban alluvial soils from New Orleans, Louisiana had higher levels of zinc as a result of highway traffic (130 µg/g) than freshly deposited lower Mississippi River delta spillway alluvium (11.1 µg/g) (Mielke et al. 1999, 2000).

A study was designed by Hutchinson and Wai (1979) to investigate the distribution of cadmium, lead, and zinc in the soil and vegetation at two reclaimed waste dumps from phosphate ore mines in southeastern

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Idaho. Zinc concentrations in the soil of the waste dumps averaged from  $443 \pm 210$  to  $1,112 \pm 124$  mg/kg. These values were high compared to those found in the control plot ( $54 \pm 16$  mg/kg). Zinc concentrations in vegetation from the reclaimed waste dumps were also high compared to the control plot. Moderate-to-high levels of zinc contamination were found in leafy vegetables (lettuce) and their supporting soil in a zone with a 0–5-km radius around a copper smelter (Beavington 1975). The mean concentrations of zinc in 17 soil samples and 12 lettuce samples collected in this zone were  $229 \pm 17$  and  $316 \pm 64$  mg/kg dry weight, respectively. Significant relationships were found between the distance from the smelter and the levels of easily extractable zinc in the soil, and between the distance from the smelter and the content of zinc in herbage. The concentration of zinc in soil at the Palmerton zinc smelter site in eastern Pennsylvania was determined 6 years after zinc smelting was terminated in 1980 (Storm et al. 1994). Levels in soils were highest (4,160 mg/kg) at sites close to the former smelter and decreased with distance. Zinc concentrations in urban top soils from the western half of East St. Louis, Illinois (a city with historical industrial activities such as smelting of non-ferrous metals) ranged from 79 to 10,360  $\mu\text{g/g}$  with an average concentration of 1,034  $\mu\text{g/g}$  (Kaminski and Landsberger 2000). Concentrations of zinc in soil irrigated with waste water or river water were measured by Schalscha et al. (1982). The total concentration of zinc in waste water-treated soils was 228 mg/kg. The total concentration of zinc in soils irrigated with river water ranged from 103 to 136 mg/kg.

Soils around galvanized water and electrical transmission towers have been reported to have elevated levels of zinc (Jones and Burgess 1984). Near Peterborough, Ontario, Canada, soil nearest to a galvanized water tower contained zinc at a concentration of  $11,480 \pm 2,966$   $\mu\text{g/g}$  dry weight, while the concentration of zinc in soil 50 meters from the tower was  $54 \pm 16$   $\mu\text{g/g}$  dry weight.

Municipal sludge and municipal incineration ash contain considerably higher levels of zinc than uncontaminated soils (Mumma et al. 1984, 1990, 1991). Therefore, application of sludge and municipal ash to soil will elevate the levels of zinc in these soils. The mean concentrations (mg/kg) of zinc according to four land use types were as follows: agricultural, 25; suburban residential, 75; mixed industrial/residential, 157; and industrial inner urban area, 360 (Haines 1984).

Zinc in water is transported to the sediment in the adsorbed or precipitated phase; the concentration of zinc in sediments of most waters is higher than the zinc concentration in aqueous phase. From 1992 to 1996, streambed sediments samples were collected from 541 locations at more than 50 river basins across the conterminous United States (illustrated in Table 6-5) as part of the National Water-Quality Assessment Program (Rice 1999; USGS 2002). The median zinc concentration in these sediments was

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110 µg/g dry weight (range, <4.0 to 9,000 µg/g dry weight). Samples collected from urban settings were enriched in zinc relative to agricultural or forest settings. The highest median concentration was observed in the Upper Colorado River Basin while the lowest was observed in Central Nebraska Basins (USGS 2002). Bed sediments from the South Platte River basin sampled from 1992 to 1993 contained zinc at concentrations ranging from 82 to 3,700 µg/g dry weight (average, 454 µg/g dry weight) (Heiny and Tat 1997). The highest concentrations were observed near the urban region around Denver, Colorado and in the Rocky Mountains. In 1979–1980, as part of the Apalachicola River Quality Assessment, fine grained sediment (<20 µm particle size) of the Apalachicola River was reported to contain zinc at a median concentration of 70 µg/g dry weight (n=15; range, 20–150 µg/g dry weight) (Elder and Mattraw 1984). Surficial lake sediments from four locations in Rock Mountain National Park contained zinc at mean concentrations ranging from 72±4 to 125±3 µg/g dry weight (Heit et al. 1984). The geometric mean and range of zinc levels in lake sediment from 189 sites in 52 Quebec and Ontario, Canada lakes were 125.2 and 3.0–559.9 µg/g dry weight, respectively (Rowan and Kalff 1993). The concentrations of zinc in sediments of the upper Columbia River, British Columbia, ranged from 45 to 51 mg/kg, while zinc concentrations in sediments from Lake Roosevelt, Washington were 60–26,840 mg/kg (Johnson et al. 1990). The higher zinc concentrations in lake sediments were due to discharges from a lead-zinc smelter and a refinery. Contaminated sediments in the West Branch of the Grand Calumet River (Indiana/Illinois), a river system heavily impacted by various industrial activities for many years, were found to contain zinc at concentrations ranging from 325 to 9,281 ppm (mean, 1,270±1,097 ppm) (Cahill and Unger 1993). Sediment samples collected from streams in the Black Hills, South Dakota, an area impacted by gold mining operations, contained zinc at levels ranging from 3.8 to 250 µg/g dry weight (May et al. 2001).

Marine sediments also contain elevated concentrations of zinc with respect to concentrations of zinc in seawater. The concentration of zinc in Hamilton Harbor sediments ranged from 1,050 to 2,900 mg/kg, compared to zinc concentrations of 6–48 µg/L in the aqueous phase (Mayer and Manning 1990). Surficial sediments from the Newark Bay Estuary, Hackensack River, Newark Bay, Arthur Kill, and Kill van Kull contained zinc at mean concentrations of 739.5±243.9, 426±600.5, 489.8±238.1, 769.1±715.2, and 331.3±213.7 ppm, respectively (Crawford et al. 1995). Sediment samples collected from the Hudson River Estuary in 1991 contained zinc at levels of 27–215 and 400–2,500 mg/kg in bottom and suspended sediment, respectively (Gibbs 1994). Marine sediment samples from the border region of Baja California (Mexico) and California (United States) contained zinc at concentration levels ranging from 39 to

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**Table 6-5. Median Zinc Levels in Bed Sediment from River Basins of the United States<sup>a</sup>**

NAWQA study unit <sup>b</sup>	Median concentration of zinc (µg/g dry weight)
Acadian-Ponchartrain (ACAD)	120
Albemarle-Pamlico Drainage (ALBE)	99
Allegheny and Monongahela River Basins (ALMN)	195
Apalachicola-Chattahoochee-Flint River Basin (ACFB)	130
Central Arizona Basins (CAZB)	160
Central Columbia Plateau (CCPT)	82
Central Nebraska Basins (CNBR)	69
Connecticut, Housatonic, and Thames River Basins (CONN)	200
Cook Inlet Basin (COOK)	110
Delaware River Basin (DELR)	290
Eastern lowas Basin (EIWA)	72.5
Georgia-Florida Coastal Plain (GAFL)	100
Great and Little Miami River Basins (MIAM)	130
Hudson River Basin (HDSN)	180
Kanawha-New River Basin (KANA)	200
Lake Erie-Lake St. Clair Drainage (LERI)	120
Long Island and New Jersey Coastal Drainages (LINJ)	245
Lower Illinois River Basin (LIRB)	88
Lower Susquehanna River Basin (LSUS)	300
Lower Tennessee River Basin (LTEN)	84
Mississippi Embayment (MISE)	91.5
Mobile River and Tributaries (MOBL)	110
Nevada Basin and Range (NVBR)	100
New England Coastal Basins (NECB)	295
Northwern Rockies Intermontane Basins (NROK)	108
Oahu (OAHU)	375
Ozark Plateaus (OZRK)	90
Potomac River Basin (POTO)	130
Puget Sound Basin (PUGT)	130
Red River of the North (REDN)	95
Rio Grande Valley (RIOG)	82.5
Sacramento River Basin (SACR)	120
San Joaquin-Tulare Basin (SANJ)	110
Santa Ana Basin (SANA)	160
Santee Basin and Costal Drainages (SANT)	94
South Central Texas (SCTX)	77
South Platte River Basin (SPLT)	180
Trinity River Basin (TRIN)	77.5
Upper Colorado River Basin (UCOL)	940
Upper Illinois River Basin (UIRB)	110
Upper Mississippi Basin (UMIS)	110
Upper Snake River Basin (USNK)	81
Upper Tennessee River Basin (UTEN)	140
Western Lake Michigan Drainage (WMIC)	98

## 6. POTENTIAL FOR HUMAN EXPOSURE

**Table 6-5. Median Zinc Levels in Bed Sediment from River Basins of the United States<sup>a</sup>**

NAWQA study unit <sup>b</sup>	Median concentration of zinc (µg/g dry weight)
White River Basin (WHIT)	100
Willamette River Basin (WILL)	120
<i>National median</i>	<i>110</i>

<sup>a</sup>Source: USGS 2000<sup>b</sup>( ) = acronym for study unit

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188 µg/g dry weight (mean, 68.3 µg/g dry weight) in the fine fraction (<63 µm) of sediment. A relative enrichment of >350% was observed with respect to non-polluted sediments of the region (Villascusa-Celaya et al. 2000). Marine sediment in coastal areas of Mexico (Pacific Ocean and Gulf of Mexico) was found to contain zinc at mean concentrations ranging from 4.0 to 227.0 µg/g dry weight (Villanueva and Botello 1998). Soto-Jimenez and Páez-Osuna (2001) reported mean concentrations of zinc ranging from 84.3±38.7 to 359±76.5 mg/kg dry weight for marine sediments collected in November 1994 from Mazatlán Harbor, Mexico (southeastern Gulf of California). This harbor receives land runoff and untreated or partially treated industrial, shipping, and domestic effluents from local point sources. During the period of 1984–1985, marine sediment samples were collected from the San Andres lagoon of the Gulf of Mexico, which is located near two industrial ports and industrial effluent is discharged into the lagoon year round. Sediments from this region were found to contain zinc at a concentration of 10.1 mg/kg dry weight (Vasquez et al. 1994).

#### 6.4.4 Other Environmental Media

As part of the National Water Quality Assessment (NAWQA) Program, the concentration of zinc in various species of fish was measured (USGS 2000a, 2000b, 2001). The concentration of zinc in fish fillet sampled from the Lower Tennessee River Basin ranged from 3.0 to 46.0 µg/g dry weight for 79 of 102 positive detections from 1980 to 1998 (USGS 2001). Fish fillets collected from the Clark Fork-Pend Oreille and Spokane River Basins (Washington, Idaho, and Montana) contained zinc at concentrations ranging from 11 to 36 µg/g dry weight (n=15; median 16 µg/g dry weight) in 1998 (USGS 2000b). In the National Contaminant Biomonitoring Program, the geometric mean concentration of zinc in various whole fish was 21.7 mg/kg wet weight (Schmitt and Brumbaugh 1990). Of all fish tested (e.g., bloater, sucker, white perch, bass, catfish, etc.), common carp showed the highest level of zinc. No significant trend in the level of zinc in whole fish was observed during 1978–1984. Blevens and Pancorbo (1986) determined the zinc concentrations in muscle tissue of fish from aquatic systems in east Tennessee, from 1980 to 1984. Mean levels of zinc in fish from Nolichucky and Little Chucky Creeks ranged from 12 to 19 ppm wet weight. Fish from Watauga and Boone Lakes had a range of mean zinc concentrations of 8.3–12 ppm wet weight; in the Holston River Basin, the range of mean concentrations of zinc was 4.6–28 ppm wet weight. The mean concentration of zinc in muscle tissue of tuna (*Thunnus thynnus*) collected from the northwest Atlantic Ocean was 17 µg/g dry weight (range, 12–25 µg/g dry weight) in 1990 (Hellou et al. 1992).

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Zinc will not concentrate in fish tissues with exposure to elevated concentrations. The concentration of zinc in yellow perch (*Perca flavescens*) from six acidic lakes in northwestern New Jersey ranged from 26.1 to 66.2 mg/kg dry weight (Sprenger et al. 1988). Although the concentrations of mercury and lead in fish from acidic lakes were higher compared to fish collected from non-acidic lakes, the concentrations of zinc showed no significant difference. Similarly, high concentrations of zinc were not found in white suckers (*Catostomus commersoni*) and brown bullheads (*Ictalurus nebulosus*) collected from two acidic Adirondack lakes in New York (Heit and Klusek 1985). Fish from the Milltown Reservoir Superfund Site in Montana (characterized by elevated concentrations of metals in wetland soils, surface water, and groundwater) contained zinc in whole body tissues at a concentration of 26.3 mg/kg wet weight (Pascoe et al. 1996). Redear sunfish (*Lepomis microlophus*), largemouth bass (*Micropterus salmoides*), and bluegill sunfish (*Lepomis macrochirus*) were collected from stormwater ponds and natural lakes and ponds in Orlando, Florida between 1991 and 1992 (Campbell 1994). The mean concentrations of zinc in whole fish collected from stormwater ponds were 42.2, 29.99, and 36.1 mg/kg wet weight for redear sunfish, largemouth bass, and bluegill sunfish, respectively. At natural lakes and ponds (controls sites), the mean concentrations of zinc were 24.83, 21.18, and 30.72 mg/kg wet weight for redear sunfish, largemouth bass, and bluegill sunfish, respectively.

Bivalves and other sessile estuarine organisms are often used as a measure of contamination of estuarine water because they usually contain higher levels of metals than fish. The arithmetic mean concentration of zinc in oysters (*Crassostrea virginica*) from the Mississippi Sound collected in 1988 was 640 mg/kg wet weight (Lytle and Lytle 1990). Oysters collected from the San Andres lagoon (Gulf of Mexico) from 1984 to 1985 contained zinc at a concentration of 3,180 mg/kg dry weight (Vazquez et al. 1994). The mean concentration of zinc in oysters (*C. virginica*) collected from the U.S. coastline of the Gulf of Mexico during 1986–1988 was 2,150 mg/kg dry weight (Presley et al. 1990). In a nationwide mussel watch program, the mean concentrations of zinc in molluscs (*Mytilus edulis*) around the coast of the United States during 1976–1988 ranged from 67 to 3,700 mg/kg dry weight (Lauenstein et al. 1990). Although the concentration on a nationwide basis varied depending on sampling sites, the level of zinc showed little evidence of statistically significant change during 1976–1988. Clams endevors (*Corbicula manilmsis*) collected as part of the Apalachicola River Quality Assessment between 1979 and 1980 contained zinc at a median concentration of 20 µg/g dry weight (range, 2.1–26 µg/g dry weight) (Elder and Matraw 1984). Blue crabs (*Callinectes sapidus*) from the Quinnipac and Connecticut Rivers (Connecticut), which are mostly harvested for personal consumption, contained zinc in muscle and hepatopancreas tissues at concentrations of 31–33 and 27–28 mg/kg wet weight, respectively (Jop et al. 1997).



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Vegetation may accumulate higher levels zinc if grown on contaminated soils. Jones et al. (1988) found that corn plants and young corn plants (*Zea mays*) grown beneath and close to a galvanized electrical transmission tower had elevated concentrations of zinc due to corrosion of the zinc protective layer on the steel. Corn seedlings grown in a highly contaminated soil (1,425 µg zinc/g soil) a meter from the tower had zinc concentration in shoots and roots of 484±103 and 1,330±250 µg/g dry weight, respectively. In contrast, seedlings grown in soil 50 meters from tower (67.3 µg zinc/g soil) had zinc concentrations in shoots and roots of 25.3±4.2 and 21.0±2.6 µg/g dry weight, respectively. Bache et al. (1991) found concentrations of zinc were highest in grass samples (*Phleum pratense L.*; *Agropyron repens L.*; *Bromus inermis L.*; *Phalaris arundinacea L.*) collected immediately adjacent to the a municipal waste incinerator (135.7 µg/g dry weight) compared to grass samples collected upwind or a distance from the incinerator (17.82–73.78 µg/g dry weight). Grasses collected from the Milltown Reservoir Superfund Site in Montana contained zinc at concentrations of 153.7 and 882.1 mg/kg for above- and below-ground samples, respectively (Pascoe et al. 1996). In contrast, control samples from a reference area contained zinc at concentrations of 71.8 and 36.2 mg/kg for above- and below-ground samples, respectively. Other studies have not shown significant correlations between zinc concentrations in soils and vegetation (Fytianos et al. 2001; Schuhmacher et al. 1998).

Sewage sludge and compost, which may be used in agriculture as a soil amendment, have high levels of zinc. The average concentrations of zinc in municipal solid waste compost and sewage from the United States were found to be 609 and 1,202 mg/kg dry weight, respectively (He et al. 1995). The median concentration of zinc found in residential compost from Toronto, Canada was 190 mg/kg dry weight (range, 100–410 mg/kg dry weight) (Evans and Tan 1998). Biowaste, composed of organic waste products from indoors and outdoors, contained zinc at concentrations of 120±25, 129±13, and 338±58 mg/kg dry matter in the >5 mm, 1–5 mm, and <0.05 mm fractions, respectively (Veeken and Hamelers 2002).

Other environmental concentrations of significance include coal and paint. Coal from the United States was found to contain zinc at a mean concentration of 53±440 ppm (n=7,908; maximum=19,000 ppm) (Finkelman 1999). Zinc was present at a median concentration of 31,101 µg/g (n=31; range, 52–98,056 µg/g) in paint from historic old homes of New Orleans, Louisiana (Mielke et al. 2001).

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**6.5 GENERAL POPULATION AND OCCUPATIONAL EXPOSURE**

**General Population.** Zinc is essential element needed by the body in small amounts and ranks as one of the most abundant trace metals in humans. Sources of exposure to zinc include ingestion of food, drinking water, food, polluted air, tobacco products, and occupational exposure, with ingestion of food being the primary route of exposure. NAS established the RDA for zinc at 11 mg/day for men and 8 mg/day for women (IOM 2002).

The average daily intake (AVDI) of zinc in humans is on the order of 5.2–16.2 mg zinc/day (Pennington et al. 1986). The dietary intake of an average teenage male has been estimated to be 0.27 mg zinc/(kg-day). Dietary supplements may provide up to an additional 1 mg zinc/(kg-day) (EPA 1980d). In an extensive survey of foods in the total diets of individuals in the United States, conducted by FDA during 1982–1984, the following values for daily zinc intakes (mg/day) were estimated in eight age and sex groups: 6–11-month-old infants, 5.24; 2-year-old children, 7.37; 14–16-year-old girls, 9.90; 14–16-year-old boys, 15.61; 25–30-year-old women, 9.56; 25–30-year-old men, 16.15; 60–65-year-old women, 8.51; and 60–85-year-old men, 12.64 (Pennington et al. 1986). FDA included drinking water in the total diet. In 1986, the average daily intakes of 40 African-Americans (age 21–65 years old living in Washington D.C. area) were  $7.7 \pm 0.4$  mg for men and  $9.1 \pm 0.7$  mg for women (Ellis et al. 1997). These results are comparable with data from the USDA's Continuing Survey of Food Intakes by Individuals (CSFII) and the National Health and Nutrition Examination Survey (NHANES) III, which reported average daily intakes as follows: NHANES III, 40–49 year old value, male (12.3 mg/day), female (8.5 mg/day); CSFII, >20-year-old value, male (12.9 mg/day), female, (8.3 mg/day). The estimated average daily intakes of zinc were reported to be 14, 11, 14, and 13.2 mg/day in France, Spain, Sweden, and Belgium, respectively (Biego et al. 1998). Using a market basket method, the average daily intakes of zinc for residents of Japan were estimated as 8,700 and 8,500  $\mu\text{g/day}$  for the years 1991 and 1992, respectively (Tsuda et al. 1995).

After a review of the literature, the National Research Council concluded that zinc concentrations in drinking water are generally well below 5 mg/L (NAS 1977). Assuming a daily intake of 2 L of water and an average body weight of 70 kg, a daily intake of less than 0.14 mg zinc/kg-day from drinking water can be estimated. Based on a body weight of 70 kg, the mean daily intakes of zinc in drinking water for residents of homes with galvanized and copper pipe plumbing systems in Seattle, Washington, were estimated to be 0.017–0.028 and 0.002–0.006 mg/kg-day, respectively (Sharrett et al. 1982b).

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Food is the major source of zinc for the general population (EPA 1987c). Zinc is widespread in commonly consumed foods but tends to be higher in those of animal origin, particularly some seafoods (e.g., one serving of oysters will more than meet the daily dietary requirements of zinc) (NAS/NRC 1979). Meat products contain relatively high concentrations of zinc, whereas fruits and vegetables have relatively low concentrations. Meats, fish, and poultry contained an average of 24.5 mg zinc/kg, whereas grains (or cereal products) and potatoes contained 8 and 6 mg/kg, respectively (Mahaffey et al. 1975). Zinc was present in all of the examined food classes. A diet of dairy products, meat, fish, poultry, grains, and cereals provides approximately 77% of the daily zinc intake. Data reported by the Food Safety and Inspection Service of the U.S. Department of Agriculture indicate that zinc was detected in 99.4–100% of the samples of healthy livestock and poultry randomly selected from among the specimens presented for slaughter in 1985–1986. Zinc concentrations in muscle tissue ranged from 0.20 ppm in young turkeys (n=61) to 1.92 ppm in heifers/steers (n=287) (Coleman et al. 1992). In a review of zinc levels in vegetables and other foods and beverages of plant origin, Weigert (1991) reported the following average concentrations (mg/kg): wheat, 41; rye, 13; rice, 8–20; potatoes, 3.51; vegetables, 4.31; fruit, 1.66; mushrooms, 9.7; cocoa, 35; tea, 35; and coffee, 6.7. Zinc is found in onions, peas, and potatoes from Denmark at mean concentrations of 3.4, 3.3–5.5, and 7.9 mg/kg fresh weight, respectively (Bibak et al. 1998a, 1998b; 1999). Zinc has also been detected in wines from Seville, Spain, at concentrations of 0.3–5.40 µg/mL, while concentrations of zinc in sherry wines ranged from 0.12 to 5.08 mg/L (López-Artiguez et al. 1990, 1996). As part of the U.S. FDA's Total Diet Study (TDS), market basket food items from locations in United States were sampled from 1991 through 1999 (FDA 2001). Results from this survey are shown in Table 6-6. In France, the estimated dietary intake of zinc from different foods were determined as follows (µg/day): vegetables, 807; fruits, 143; beverages, 143; cereals, 2,572; fish-crustaceans, 795; meat-poultry-eggs, 8,318; milk-dairy products, 1,127; condiments-sugar-oils, 140; canned foods, 383; and total, 14,429 (Biego et al. 1998). The largest percentage of zinc is from meat-poultry-eggs (58%) followed by cereals (18%) and milk-dairy products (8%).

Federal regulations permit the use of zinc acetate, zinc oxide, and zinc sulfide as components of adhesives, coatings, or rubber packaging materials intended for food contact (FDA 1987b, 1987c, 1987d). Federal regulations also permit the use of zinc chloride, zinc oxide, zinc stearate, and zinc sulfate as GRAS (Generally Recognized As Safe) food additives when they are used "in accordance with good manufacturing practices" (FDA 1987e, 1987f, 1987g, 1987h, 1987i, 1987j). In addition, the use of zinc oxide as a color additive in drugs and cosmetics is also permitted with certain restrictions (FDA 1987a).

## 6. POTENTIAL FOR HUMAN EXPOSURE

**Table 6-6. U.S. Food and Drug Administration—Total Diet Study (TDS)—Market Baskets 91-3 Through 99-1<sup>a</sup>**

Zinc—summary of results									
TDS food description	TDS food number	Number of results	Number of not detected	Number of traces	Mean (mg/kg)	Standard deviation (mg/kg)	Minimum (mg/kg)	Maximum (mg/kg)	Median (mg/kg)
	Overall:	6,756	480	926	9.4	18.5	0	226	3.9
Whole milk, fluid	1	26	0	0	3.5	0.012	0	0.044	0.02
Lowfat (2% fat) milk, fluid	2	26	0	0	3.9	0.4	3.0	4.9	3.9
Chocolate milk, fluid	3	26	0	0	4.1	0.7	2.7	6.1	4.1
Skim milk, fluid	4	26	0	0	3.9	0.9	1.6	6.1	4.0
Plain yogurt, lowfat	6	26	0	0	5.6	0.5	4.6	6.9	5.7
Chocolate milk shake, fast-food	7	26	0	0	4.7	0.6	3.3	5.8	4.7
Evaporated milk, canned	8	26	0	0	7.4	0.7	5.3	8.9	7.5
American, processed cheese	10	26	0	0	27.9	2.8	23.4	33.2	27.4
Cottage cheese, 4% milkfat	11	26	0	0	3.8	0.6	2.8	5.2	3.8
Cheddar cheese	12	26	0	0	36.8	4.8	18.5	45.0	36.8
Ground beef, pan-cooked	13	26	0	0	58.4	6.0	48.8	71.3	58.6
Beef chuck roast, baked	14	26	0	0	81	11	56.7	103	82.5
Beef steak, loin, pan-cooked	16	26	0	0	56.6	7.9	39.7	72.9	56.0
Ham, baked	17	26	0	0	21.7	3.6	13.8	29.3	21.1
Pork chop, pan-cooked	18	26	0	0	27.7	3.9	22.0	36.6	27.4
Pork sausage, pan-cooked	19	26	0	0	28.6	3.2	23.5	36.6	28.1
Pork bacon, pan-cooked	20	26	0	0	27.7	4.0	20.6	36.0	27.0
Pork roast, baked	21	26	0	0	29.8	5.8	22.1	43.7	30.0
Lamb chop, pan-cooked	22	26	0	0	52	13	33.0	86.0	51.0

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TDS food description	TDS food number	Number of results	Number of not detected	Number of traces	Mean (mg/kg)	Standard deviation (mg/kg)	Minimum (mg/kg)	Maximum (mg/kg)	Median (mg/kg)
Chicken, fried (breast, leg, and thigh) homemade	24	26	0	0	19.0	2.4	14.1	23.2	18.9
Turkey breast, roasted	26	26	0	0	13.9	1.7	11.4	18.1	13.6
Liver, beef, fried	27	26	0	0	56	18	33.5	114	52.0
Frankfurters, beef, boiled	28	26	0	0	22.8	2.3	18.5	27.7	22.9
Bologna, sliced	29	26	0	0	14.0	2.8	9.9	19.7	13.8
Salami, sliced	30	26	0	0	20.8	5.2	13.4	32.5	19.5
Tuna, canned in oil	32	26	0	0	6.0	1.6	3.4	10.7	6.0
Fish sticks, frozen, heated	34	26	0	0	4.4	0.8	2.0	5.8	4.4
Eggs, scrambled	35	26	0	0	11.0	1.2	8.2	13.0	11.1
Eggs, fried	36	26	0	0	13.1	0.9	11.3	14.9	13.1
Eggs, boiled	37	26	0	0	13.5	1.5	11.6	16.8	13.3
Pinto beans, dry boiled	38	26	0	0	9.4	0.8	7.6	11.1	9.4
Pork and beans, canned	39	26	0	0	7.9	2.3	4.6	13.6	7.8
Lima beans, immature, frozen, boiled	42	26	0	0	6.4	0.6	5.4	7.7	6.3
Green peas, fresh/frozen, boiled	46	26	0	0	6.2	1.2	2.8	7.8	6.3
Peanut butter, smooth	47	26	0	0	26.9	1.9	22.6	30.7	27.0
Peanuts, dry roasted	48	26	0	0	29.5	2.0	25.2	33.2	29.6
White rice, cooked	50	26	0	0	5.0	0.9	3.0	6.1	5.1

## 6. POTENTIAL FOR HUMAN EXPOSURE

**Table 6-6. U.S. Food and Drug Administration—Total Diet Study (TDS)—Market Baskets 91-3 Through 99-1<sup>a</sup>**

Zinc—summary of results									
TDS food description	TDS food number	Number of results	Number of not detected	Number of traces	Mean (mg/kg)	Standard deviation (mg/kg)	Minimum (mg/kg)	Maximum (mg/kg)	Median (mg/kg)
Oatmeal, quick (1–3 minutes), cooked	51	26	0	0	5.2	1.0	3.3	7.6	5.0
Wheat cereal, farina, quick (1–3 minutes), cooked	52	26	0	3	2.3	1.3	0.7	6.1	1.9
Corngrits, regular, cooked	53	26	0	5	1.4	0.7	0.6	3.7	1.3
Corn, fresh/frozen, boiled	54	26	0	0	4.0	0.7	2.6	5.5	3.9
Cream style corn, canned	56	26	0	0	3.0	0.9	2.0	6.0	2.6
Popcorn, popped in oil	57	26	0	0	21.4	4.3	14.2	28.4	21.7
White bread	58	26	0	0	6.4	0.9	3.5	8.1	6.3
White roll	59	26	0	0	8.3	1.0	5.8	9.9	8.2
Cornbread, homemade	60	26	0	0	6.1	0.8	4.7	7.9	6.0
Biscuit, from refrigerated dough, baked	61	26	0	0	4.2	0.8	2.2	7.0	4.1
Whole wheat bread	62	26	0	0	16.6	2.5	12.1	20.8	17.1
Tortilla, flour	63	26	0	0	5.4	1.4	3.7	10.9	5.1
Rye bread	64	26	0	0	9.6	1.0	6.8	11.8	9.6
Blueberry muffin, commercial	65	26	0	0	4.2	0.6	2.9	5.0	4.3
Saltine crackers	66	26	0	0	6.9	0.7	5.6	7.9	6.9
Corn chips	67	26	0	0	13.8	1.6	11.4	17.8	13.6
Pancake from mix	68	26	0	0	6.1	0.9	3.6	8.0	6.1
Egg noodles, boiled	69	26	0	0	6.2	1.1	4.3	8.7	6.0
Macaroni, boiled	70	26	0	0	4.5	0.6	3.6	6.3	4.4
Corn flakes	71	26	0	0	8.2	14.3	1.2	54.5	1.7

## 6. POTENTIAL FOR HUMAN EXPOSURE

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Zinc—summary of results									
TDS food description	TDS food number	Number of results	Number of not detected	Number of traces	Mean (mg/kg)	Standard deviation (mg/kg)	Minimum (mg/kg)	Maximum (mg/kg)	Median (mg/kg)
Fruit-flavored, sweetened cereal	72	26	0	0	147	35.7	41.2	211	156
Shredded wheat cereal	73	26	0	0	28.4	5.1	22.3	43.4	27.1
Raisin bran cereal	74	26	0	0	99	26	40.2	156	102
Crisped rice cereal	75	26	0	0	20	18	9.1	91.5	12.8
Granola cereal	76	26	0	0	85	43	17.9	191	86.7
Oat ring cereal	77	26	0	0	130	50	25.7	226	147
Apple, red, raw	78	26	16	10	0.2	0.2	0	0.8	0
Orange, raw	79	26	0	20	0.6	0.1	0.4	1.0	0.6
Banana, raw	80	26	0	0	1.6	0.2	1.4	2.0	1.6
Watermelon, raw	81	26	0	21	0.7	0.2	0.3	1.1	0.7
Peach, raw	83	26	0	6	1.6	1.2	0.7	5.2	1.2
Applesauce, bottled	84	26	15	11	0.2	0.2	0	0.6	0
Pear, raw	85	26	0	10	1.0	0.2	0.6	1.5	1.0
Strawberries, raw	86	26	0	8	1.0	0.2	0.7	1.4	1.0
Fruit cocktail, canned in heavy syrup	87	26	0	24	0.5	0.2	0.4	1.0	0.5
Grapes, red/green, seedless, raw	88	26	0	22	0.6	0.2	0.3	1.1	0.5
Cantaloupe, raw	89	26	0	3	1.4	0.3	0.8	2.1	1.4
Plums, raw	91	26	0	16	0.9	0.2	0.5	1.2	0.8
Grapefruit, raw	92	26	0	23	0.6	0.2	0.3	0.9	0.5
Pineapple, canned in juice	93	26	0	22	0.8	0.2	0.5	1.2	0.8
Sweet cherries, raw	94	20	0	14	0.8	0.3	0.4	1.5	0.7
Raisins, dried	95	26	0	6	2.1	0.4	1.5	3.1	2.1
Prunes, dried	96	26	0	0	4.4	0.5	3.3	5.4	4.3

## 6. POTENTIAL FOR HUMAN EXPOSURE

**Table 6-6. U.S. Food and Drug Administration—Total Diet Study (TDS)—Market Baskets 91-3 Through 99-1<sup>a</sup>**

Zinc—summary of results									
TDS food description	TDS food number	Number of results	Number of not detected	Number of traces	Mean (mg/kg)	Standard deviation (mg/kg)	Minimum (mg/kg)	Maximum (mg/kg)	Median (mg/kg)
Avocado, raw	97	26	0	0	6.5	1.6	4.0	9.2	6.2
Orange juice, from frozen concentrate	98	26	2	23	0.4	0.2	0	0.8	0.4
Apple juice, bottled	99	26	8	18	0.2	0.2	0	0.5	0.2
Grapefruit juice, from frozen concentrate	100	26	0	24	0.4	0.1	0.2	0.8	0.4
Prune juice, bottled	103	26	0	0	1.8	1.9	1.1	10.7	1.4
Lemonade, from frozen concentrate	105	26	18	8	0.08	0.13	0	0.4	0
Spinach, fresh/frozen, boiled	107	26	0	0	4.3	1.4	2.6	8.8	3.9
Collards, fresh/frozen	108	26	0	1	2.3	0.8	1.0	4.5	2.1
Iceberg lettuce, raw	109	26	0	3	1.4	0.4	0.6	2.2	1.4
Cabbage, fresh, boiled	110	26	0	14	1.0	0.4	0.4	2.4	0.9
Coleslaw with dressing, homemade	111	26	0	17	1.6	0.4	0.9	2.4	1.6
Sauerkraut, canned	112	26	0	10	1.1	0.3	0.5	1.9	1.1
Broccoli, fresh/frozen, boiled	113	26	0	0	1.8	0.4	1.2	2.5	1.8
Celery, raw	114	26	0	21	0.8	0.2	0.3	1.3	0.8
Asparagus, fresh/frozen, boiled	115	26	0	0	3.9	0.7	2.7	5.2	3.8
Cauliflower, fresh/frozen, boiled	116	26	0	0	1.5	0.3	1.1	2.1	1.5
Tomato, red, raw	117	26	0	5	1.3	0.3	0.6	1.9	1.2
Tomato sauce, plain, bottled	119	26	0	0	1.8	0.4	1.1	2.6	1.7



## 6. POTENTIAL FOR HUMAN EXPOSURE

**Table 6-6. U.S. Food and Drug Administration—Total Diet Study (TDS)—Market Baskets 91-3 Through 99-1<sup>a</sup>**

Zinc—summary of results									
TDS food description	TDS food number	Number of results	Number of not detected	Number of traces	Mean (mg/kg)	Standard deviation (mg/kg)	Minimum (mg/kg)	Maximum (mg/kg)	Median (mg/kg)
Green beans, fresh/frozen, boiled	121	26	0	0	2.0	0.4	1.5	2.8	2.0
Cucumber, raw	123	26	0	5	1.1	0.2	0.7	1.6	1.1
Summer squash, fresh/frozen, boiled	124	26	0	0	1.9	0.4	1.3	2.9	1.9
Green pepper, raw	125	26	0	7	1.2	0.3	0.6	1.9	1.2
Winter squash, fresh/frozen, baked, mashed	126	26	1	0	2.2	0.7	0	3.4	2.2
Onion, mature, raw	128	26	0	5	1.4	0.5	0.7	2.2	1.5
Radish, raw	132	26	0	9	1.2	0.3	0.7	2.0	1.1
French fries, frozen, heated	134	26	0	0	3.3	0.5	2.6	4.1	3.2
Mashed potatoes, from flakes	135	26	0	0	2.2	0.3	1.7	2.7	2.2
White potato, boiled without skin	136	26	0	0	2.1	0.5	1.4	3.2	1.9
White potato, baked with skin	137	26	0	0	3.3	0.8	1.9	5.5	3.2
Potato chips	138	26	0	0	9.7	1.6	6.5	12.7	9.3
Scalloped potatoes, homemade	139	26	0	0	3.6	0.4	2.8	4.3	3.4
Sweet potato, fresh, baked	140	26	0	0	2.7	0.4	1.8	3.5	2.8
Spaghetti with tomato sauce and meatballs, homemade	142	26	0	0	8.5	1.3	6.2	10.5	8.5

## 6. POTENTIAL FOR HUMAN EXPOSURE

**Table 6-6. U.S. Food and Drug Administration—Total Diet Study (TDS)—Market Baskets 91-3 Through 99-1<sup>a</sup>**

Zinc—summary of results									
TDS food description	TDS food number	Number of results	Number of not detected	Number of traces	Mean (mg/kg)	Standard deviation (mg/kg)	Minimum (mg/kg)	Maximum (mg/kg)	Median (mg/kg)
Beef stew with potatoes, carrots, and onion, homemade	143	26	0	0	22.2	6.7	10.1	38.2	22.1
Macaroni and cheese, from box mix	146	26	0	0	5.8	1.3	2.9	7.7	5.9
Quarter-pound hamburger on bun, fast-food	147	26	0	0	27.6	3.5	21.5	34.5	27.4
Meatloaf, homemade	148	26	0	0	45.0	5.8	30.4	52.0	46.3
Spaghetti with tomato sauce, canned	149	26	0	0	3.0	1.5	2.0	9.9	2.7
Lasagna with meat, homemade	151	26	0	0	14.9	2.2	11.2	18.7	15.3
Chicken potpie, frozen, heated	152	26	0	0	4.5	0.8	3.3	7.1	4.3
Chicken noodle soup, canned, condensed, prepared with water	155	26	0	7	1.3	0.5	0.4	2.4	1.2
Tomato soup, canned, condensed, prepared with water	156	26	0	10	1.1	0.2	0.7	1.7	1.0
Vegetable beef soup, canned, condensed, prepared with water	157	26	0	0	3.3	0.7	1.8	4.9	3.3
White sauce, homemade	160	26	0	0	3.8	0.5	2.8	4.7	3.8
Dill cucumber pickles	161	26	1	14	0.8	0.3	0	1.3	0.9

## 6. POTENTIAL FOR HUMAN EXPOSURE

**Table 6-6. U.S. Food and Drug Administration—Total Diet Study (TDS)—Market Baskets 91-3 Through 99-1<sup>a</sup>**

Zinc—summary of results									
TDS food description	TDS food number	Number of results	Number of not detected	Number of traces	Mean (mg/kg)	Standard deviation (mg/kg)	Minimum (mg/kg)	Maximum (mg/kg)	Median (mg/kg)
Margarine, stick, regular (salted)	162	26	11	15	0.4	0.3	0	1.0	0.3
Butter, regular (salted)	164	26	0	19	0.9	0.4	0.5	2.6	0.7
Mayonnaise, regular, bottled	166	26	0	13	2.0	0.4	1.4	3.0	1.9
Half & half cream	167	26	0	0	3.1	0.4	2.2	3.9	3.3
Cream substitute, frozen	168	26	19	7	0.1	0.2	0	0.6	0
White sugar, granulated	169	26	24	2	0	0.1	0	0.4	0
Pancake syrup	170	26	5	10	1.0	0.9	0	3.8	0.9
Honey	172	26	0	12	1.2	0.6	0.6	2.9	1.0
Tomato catsup	173	26	0	0	1.9	0.3	1.5	3.0	1.9
Chocolate pudding, from instant mix	175	26	0	0	4.4	0.5	2.5	5.2	4.5
Vanilla flavored light ice cream	177	26	0	0	3.9	1.0	1.8	5.7	3.9
Chocolate cake with chocolate icing, commercial	178	26	0	0	5.5	0.8	3.9	7.1	5.3
Yellow cake with white icing, prepared from cake and icing mixes	179	26	0	0	2.2	0.5	1.2	3.5	2.2
Sweet roll/Danish, commercial	182	26	0	0	6.0	0.8	4.5	8.2	5.8
Chocolate chip cookies, commercial	183	26	0	0	7.0	1.0	5.3	10.0	6.9

## 6. POTENTIAL FOR HUMAN EXPOSURE

**Table 6-6. U.S. Food and Drug Administration—Total Diet Study (TDS)—Market Baskets 91-3 Through 99-1<sup>a</sup>**

Zinc—summary of results									
TDS food description	TDS food number	Number of results	Number of not detected	Number of traces	Mean (mg/kg)	Standard deviation (mg/kg)	Minimum (mg/kg)	Maximum (mg/kg)	Median (mg/kg)
Sandwich cookies with creme filling, commercial	184	26	0	0	5.1	1.4	2.7	8.2	4.8
Apple pie, fresh/frozen, commercial	185	26	0	0	1.8	0.3	1.2	2.7	1.8
Pumpkin pie, fresh/frozen, commercial	186	26	0	0	4.4	0.5	3.6	5.4	4.4
Milk chocolate candy bar, plain	187	26	0	0	13.4	0.9	11.6	14.9	13.5
Caramel candy	188	26	0	0	4.1	1.2	2.0	6.5	3.9
Gelatin dessert, any flavor	190	26	24	2	0	0.1	0	0.3	0
Cola carbonated beverage	191	26	25	1	0	0.1	0	0.3	0
Fruit drink, from powder	193	26	26	0	0	0	0	0	0
Low-calorie cola carbonated beverage	194	26	23	3	0	0.1	0	0.3	0
Coffee, decaffeinated, from instant	196	26	19	7	0	0.1	0	0.2	0
Tea, from tea bag	197	26	5	20	0.1	0.1	0	0.4	0.1
Beer	198	26	20	6	0	0.1	0	0.3	0
Dry table wine	199	26	2	8	0.5	0.3	0	1.5	0.5
Whiskey	200	26	25	1	0	0	0	0.1	0
Tap water	201	26	14	9	0.07	0.13	0	0.6	0
Milk-based infant formula, high iron, ready-to-feed	202	26	0	0	6.9	0.7	5.7	9.1	6.8

## 6. POTENTIAL FOR HUMAN EXPOSURE

**Table 6-6. U.S. Food and Drug Administration—Total Diet Study (TDS)—Market Baskets 91-3 Through 99-1<sup>a</sup>**

Zinc—summary of results									
TDS food description	TDS food number	Number of results	Number of not detected	Number of traces	Mean (mg/kg)	Standard deviation (mg/kg)	Minimum (mg/kg)	Maximum (mg/kg)	Median (mg/kg)
Milk-based infant formula, low iron, ready-to-feed	203	26	0	0	6.9	1.2	3.8	9.4	6.9
Beef, strained/junior	205	26	0	0	29.9	3.9	19.7	36.4	29.9
Chicken, strained/junior, with/without broth or gravy	207	26	0	0	11.9	2.0	6.4	14.9	11.8
Chicken/turkey with vegetables, high/lean meat, strained/junior	208	2	0	0	6.5	0.1	6.4	6.6	6.5
Beef with vegetables, high/lean meat, strained/junior	209	2	0	0	13.3	1.0	12.6	14.0	13.3
Ham with vegetables, high/lean meat, strained/junior	210	2	0	0	10.3	0.6	9.9	10.7	10.3
Vegetables and beef, strained/junior	211	26	0	0	4.3	0.9	2.3	5.6	4.7
Vegetables and chicken, strained/junior	212	26	0	0	3.3	0.6	1.9	5.0	3.3
Vegetables and ham, strained/junior	213	26	0	0	2.7	0.4	2.0	3.6	2.7

## 6. POTENTIAL FOR HUMAN EXPOSURE

**Table 6-6. U.S. Food and Drug Administration—Total Diet Study (TDS)—Market Baskets 91-3 Through 99-1<sup>a</sup>**

Zinc—summary of results									
TDS food description	TDS food number	Number of results	Number of not detected	Number of traces	Mean (mg/kg)	Standard deviation (mg/kg)	Minimum (mg/kg)	Maximum (mg/kg)	Median (mg/kg)
Chicken noodle dinner, strained/ junior	214	26	0	0	3.6	0.5	2.7	4.6	3.6
Macaroni, tomatoes, and beef, strained/ junior	215	26	0	0	3.9	0.5	3.0	5.3	3.9
Turkey and rice, strained/ junior	216	26	0	0	4.4	0.9	3.0	6.3	4.3
Carrots, strained/ junior	218	26	0	1	1.8	0.4	0.7	2.5	1.8
Green beans, strained/ junior	219	26	0	0	2.2	0.4	1.4	3.0	2.3
Mixed vegetables, strained/ junior	220	26	0	0	2.1	0.4	1.3	3.2	2.0
Sweet potatoes, strained/ junior	221	26	0	0	1.8	0.3	1.4	2.5	1.7
Creamed corn, strained/ junior	222	26	0	0	2.5	0.4	1.6	3.1	2.6
Peas, strained/ junior	223	26	0	0	5.6	0.8	4.1	7.0	5.5
Creamed spinach, strained/ junior	224	26	0	0	5.0	1.1	3.0	8.0	5.0
Applesauce, strained/ junior	225	26	10	15	0.3	0.3	0	1.0	0.3
Peaches, strained/ junior	226	26	0	15	0.9	0.3	0.4	1.7	0.8
Pears, strained/ junior	227	26	0	12	1.1	0.2	0.7	1.5	1.0

## 6. POTENTIAL FOR HUMAN EXPOSURE

**Table 6-6. U.S. Food and Drug Administration—Total Diet Study (TDS)—Market Baskets 91-3 Through 99-1<sup>a</sup>**

Zinc—summary of results									
TDS food description	TDS food number	Number of results	Number of not detected	Number of traces	Mean (mg/kg)	Standard deviation (mg/kg)	Minimum (mg/kg)	Maximum (mg/kg)	Median (mg/kg)
Apple juice, strained	230	26	17	9	0.1	0.2	0	0.6	0
Orange juice, strained	231	26	1	25	0.5	0.1	0	0.7	0.5
Custard pudding, strained/ junior	232	26	0	0	3.2	0.6	2.5	4.8	3.0
Fruit dessert/ pudding, strained/ junior	233	26	1	20	0.6	0.4	0	1.9	0.5
Fruit-flavored yogurt, lowfat (fruit mixed in)	235	26	0	0	4.4	0.5	3.3	5.6	4.4
Swiss cheese	236	26	0	0	44.1	3.2	35.6	48.9	44.5
Cream cheese	237	26	0	0	5.3	1.0	2.7	6.7	5.4
Veal cutlet, pan-cooked	238	26	0	0	39.6	8.7	16.7	62.9	39.6
Ham luncheon meat, sliced	239	26	0	0	18.8	2.9	12.1	26.0	19.0
Chicken breast, roasted	240	26	0	0	8.9	2.2	7.1	18.4	8.5
Chicken nuggets, fast-food	241	26	0	0	8.3	1.6	5.4	13.1	8.2
Chicken, fried (breast, leg, and thigh), fast-food	242	26	0	0	12.2	1.5	8.3	15.7	12.5
Haddock, pan-cooked	243	20	0	0	4.7	0.9	3.7	8.1	4.5
Shrimp, boiled	244	26	0	0	16.7	7.6	7.2	43.7	14.6
Kidney beans, dry, boiled	245	26	0	0	10.4	1.3	8.1	14.4	10.3
Peas, mature, dry, boiled	246	26	0	0	10.7	1.1	8.8	12.9	10.5

## 6. POTENTIAL FOR HUMAN EXPOSURE

**Table 6-6. U.S. Food and Drug Administration—Total Diet Study (TDS)—Market Baskets 91-3 Through 99-1<sup>a</sup>**

Zinc—summary of results									
TDS food description	TDS food number	Number of results	Number of not detected	Number of traces	Mean (mg/kg)	Standard deviation (mg/kg)	Minimum (mg/kg)	Maximum (mg/kg)	Median (mg/kg)
Mixed nuts, no peanuts, dry roasted	247	26	0	0	40.6	6.1	23.2	62.6	40.2
Cracked wheat bread	248	26	0	0	11.5	2.3	7.9	19.1	11.3
Bagel, plain	249	26	0	0	7.8	1.4	5.0	12.9	7.6
English muffin, plain, toasted	250	26	0	0	7.9	1.3	5.7	10.7	7.8
Graham crackers	251	26	0	0	7.7	1.4	5.3	10.3	7.5
Butter-type crackers	252	26	0	0	5.4	0.6	4.4	6.4	5.5
Apricot, raw	253	21	0	0	2.0	0.5	1.3	3.2	1.9
Peach, canned in light/medium syrup	254	26	1	25	0.6	0.2	0	0.9	0.6
Pear, canned in light syrup	255	26	0	26	0.6	0.1	0.3	0.8	0.6
Pineapple juice, from frozen concentrate	256	26	0	18	0.8	0.1	0.5	1.1	0.7
Grape juice, from frozen concentrate	257	26	3	23	0.4	0.2	0	0.6	0.5
French fries, fast-food	258	26	0	0	4.8	1.0	3.2	8.1	4.7
Carrot, fresh, boiled	259	26	0	0	1.8	0.4	1.1	2.7	1.7
Tomato, stewed, canned	260	26	0	1	1.3	0.1	1.0	1.6	1.3
Tomato juice, bottled	261	26	0	5	1.2	0.2	0.8	1.7	1.1
Beets, fresh/frozen, boiled	262	26	0	0	3.5	0.9	2.2	5.5	3.3
Brussels sprouts, fresh/frozen, boiled	263	26	0	0	2.5	0.5	1.7	3.7	2.3
Mushrooms, raw	264	26	0	0	3.9	0.8	2.6	5.5	4.0



## 6. POTENTIAL FOR HUMAN EXPOSURE

**Table 6-6. U.S. Food and Drug Administration—Total Diet Study (TDS)—Market Baskets 91-3 Through 99-1<sup>a</sup>**

Zinc—summary of results									
TDS food description	TDS food number	Number of results	Number of not detected	Number of traces	Mean (mg/kg)	Standard deviation (mg/kg)	Minimum (mg/kg)	Maximum (mg/kg)	Median (mg/kg)
Eggplant, fresh, boiled	265	26	0	10	1.0	0.3	0.6	1.9	1.0
Turnip, fresh/frozen, boiled	266	26	0	3	1.3	0.4	0.6	2.5	1.2
Okra, fresh/frozen, boiled	267	26	0	0	4.3	0.9	2.2	6.5	4.3
Mixed vegetables, frozen, boiled	268	26	0	0	3.6	0.7	2.0	5.3	3.7
Beef stroganoff, homemade	269	26	0	0	16.4	2.7	11.5	21.2	16.4
Green peppers stuffed with beef and rice, homemade	270	26	0	0	13.2	4.8	6.8	30.1	11.9
Chili con carne with beans, homemade	271	26	0	0	18.5	4.5	11.8	35.2	17.7
Tuna noodle casserole, homemade	272	25	0	0	6.0	0.9	4.0	7.7	5.9
Salisbury steak with gravy, potatoes, and vegetable, frozen meal, heated	273	26	0	0	9.2	1.7	4.9	13.0	9.2
Turkey with gravy, dressing, potatoes, and vegetable, frozen meal, heated	274	26	0	0	6.3	1.1	4.5	9.1	6.0
Quarter-pound cheeseburger on bun, fast-food	275	26	0	0	26.8	3.2	22.2	33.5	26.3

## 6. POTENTIAL FOR HUMAN EXPOSURE

**Table 6-6. U.S. Food and Drug Administration—Total Diet Study (TDS)—Market Baskets 91-3 Through 99-1<sup>a</sup>**

Zinc—summary of results									
TDS food description	TDS food number	Number of results	Number of not detected	Number of traces	Mean (mg/kg)	Standard deviation (mg/kg)	Minimum (mg/kg)	Maximum (mg/kg)	Median (mg/kg)
Fish sandwich on bun, fast-food	276	26	0	0	5.7	0.7	4.4	7.7	5.6
Frankfurter on bun, fast-food	277	26	0	0	14.8	2.7	10.7	21.6	14.5
Egg, cheese, and ham on English muffin, fast-food	278	26	0	0	13.4	1.4	11.4	17.2	13.1
Taco/tostada, from Mexican carry-out	279	26	0	0	22.7	3.8	12.2	30.0	23.0
Cheese pizza, regular crust, from pizza carry-out	280	26	0	0	17.1	1.9	13.9	21.2	16.9
Cheese and pepperoni pizza, regular crust, from pizza carry-out	281	26	0	0	17.4	2.1	13.6	21.8	17.1
Beef chow mein, from Chinese carry-out	282	26	0	0	11.3	2.1	7.6	16.0	11.3
Bean with bacon/pork soup, canned, condensed, prepared with water	283	26	0	0	3.8	0.8	2.0	5.5	4.0
Mushroom soup, canned, condensed, prepared with whole milk	284	26	0	0	2.7	0.6	2.0	4.1	2.6

## 6. POTENTIAL FOR HUMAN EXPOSURE

**Table 6-6. U.S. Food and Drug Administration—Total Diet Study (TDS)—Market Baskets 91-3 Through 99-1<sup>a</sup>**

Zinc—summary of results									
TDS food description	TDS food number	Number of results	Number of not detected	Number of traces	Mean (mg/kg)	Standard deviation (mg/kg)	Minimum (mg/kg)	Maximum (mg/kg)	Median (mg/kg)
Clam chowder, New England, canned, condensed, prepared with whole milk	285	26	0	0	4.0	0.8	3.1	7.5	3.8
Vanilla ice cream	286	26	0	0	3.3	0.8	1.8	5.0	3.4
Fruit flavor sherbet	287	26	0	10	1.1	0.4	0.4	2.6	1.0
Popsicle, any flavor	288	26	22	4	0.07	0.18	0	0.6	0
Chocolate snack cake with chocolate icing	289	26	0	0	5.9	1.2	3.9	8.4	5.8
Cake doughnuts with icing, any flavor, from doughnut store	290	26	0	0	4.6	0.8	3.0	6.6	4.4
Brownies, commercial	291	26	0	0	6.7	1.1	4.8	9.3	6.7
Sugar cookies, commercial	292	26	0	0	4.1	0.7	3.1	6.1	4.0
Suckers, any flavor	293	26	20	6	0.1	0.2	0	0.5	0
Pretzels, hard, salted, any shape	294	26	0	0	9.1	1.3	5.7	11.4	8.7
Chocolate syrup dessert topping	295	26	0	0	6.7	0.8	5.5	8.4	6.7
Jelly, any flavor	296	26	8	18	0.3	0.2	0	0.7	0.4
Sweet cucumber pickles	297	26	2	21	0.7	0.3	0	1.7	0.7
Yellow mustard	298	26	0	0	6.5	0.6	5.1	7.5	6.4

## 6. POTENTIAL FOR HUMAN EXPOSURE

**Table 6-6. U.S. Food and Drug Administration—Total Diet Study (TDS)—Market Baskets 91-3 Through 99-1<sup>a</sup>**

Zinc—summary of results									
TDS food description	TDS food number	Number of results	Number of not detected	Number of traces	Mean (mg/kg)	Standard deviation (mg/kg)	Minimum (mg/kg)	Maximum (mg/kg)	Median (mg/kg)
Black olives	299	26	0	5	1.3	0.4	0.5	2.2	1.4
Sour cream	300	26	0	0	3.7	0.5	2.9	4.5	3.7
Brown gravy, homemade	301	26	0	0	5.0	2.2	2.0	9.5	4.5
French salad dressing, regular	302	26	0	17	0.93	0.37	0.3	2.2	0.9
Italian salad dressing, low-calorie	303	26	2	20	0.66	0.45	0	2.0	0.5
Olive/safflower oil	304	25	9	15	0.4	0.5	0	2.1	0.3
Coffee, from ground	305	26	24	1	0.2	0.8	0	3.9	0
Fruit-flavored carbonated beverage	306	26	23	2	0.1	0.5	0	2.5	0
Fruit drink, canned	307	26	18	8	0.1	0.1	0	0.4	0
Martini	308	26	16	10	0.06	0.08	0	0.3	0
Soy-based infant formula, ready-to-feed	309	26	0	0	6.9	1.4	4.8	11.6	6.9
Egg yolk, strained/junior	310	12	0	0	22.2	1.7	19.4	26.0	22.2
Rice infant cereal, instant, prepared with whole milk	311	26	0	0	6.1	1.3	3.1	8.8	6.1
Rice cereal, strained/junior	312	20	0	1	2.6	2.6	0.8	8.5	1.5
Bananas with tapioca, strained/junior	313	26	0	20	0.8	0.2	0.3	1.3	0.8
Beets, strained/junior	314	15	0	0	2.3	0.4	1.5	3.0	2.4

## 6. POTENTIAL FOR HUMAN EXPOSURE

**Table 6-6. U.S. Food and Drug Administration—Total Diet Study (TDS)—Market Baskets 91-3 Through 99-1<sup>a</sup>**

Zinc—summary of results									
TDS food description	TDS food number	Number of results	Number of not detected	Number of traces	Mean (mg/kg)	Standard deviation (mg/kg)	Minimum (mg/kg)	Maximum (mg/kg)	Median (mg/kg)
Split peas with vegetables and ham/bacon, strained/ junior	316	26	0	0	2.9	0.6	2.2	4.1	2.9
Teething biscuits	317	26	0	0	6.9	2.1	4.5	14.0	6.4
Salmon, steaks or filets, fresh or frozen, baked	318	6	0	0	4.6	0.6	4.0	5.3	4.8
Rice cereal with apple, strained/ junior	319	4	0	0	19.2	16.5	3.8	38.4	17.4
Squash, strained/ junior	320	4	0	0	1.5	0.2	1.4	1.7	1.5

<sup>a</sup> Source: FDA 2001

## 6. POTENTIAL FOR HUMAN EXPOSURE

Negligible quantities of zinc are inhaled in ambient air. Exposure to airborne zinc is largely occupational through the inhalation of industrial dusts or fumes. Individuals occupationally exposed to metallic zinc and zinc compounds are those involved in galvanizing, smelting, welding, or brass foundry operations. In such operations, zinc as ore or metal and its alloys are often exposed in an oxidizing atmosphere to temperatures near the metal's boiling point of 907 °C. This heating results in the formation of fresh zinc oxide particles (0.2–1.0 µm), which may subsequently be inhaled. Inhalation of zinc oxide particles and fumes by workers can result in metal fume fever (Martin et al. 1999). Inhalation was reported to be the most probable route of exposure to zinc for 26 lead smelter workers found to have significantly ( $p < 0.01$ ) elevated blood plasma levels of zinc. Mean plasma zinc concentrations were 12.9 mmol/L (range, 9.8–16.7) for the workers versus 10.9 mmol/L (range, 8.1–14.6) for a non-lead-exposed control group (Vasikaran et al. 1992). Twenty workers in a zinc foundry in Baiyin, China were investigated for exposure to zinc oxide fumes (Martin et al. 1999). Eighteen of the workers had worked at the foundry since its opening 6 years earlier. Thirteen of the subjects reported at least one of the symptoms associated with metal fume fever during their tenure at the foundry. Workers were examined before the start of the shift, in the middle of the shift, and after the shift. Despite zinc exposures as high as 36.3 mg/m<sup>3</sup> over <4 hours and a mean air sample concentration of 3.16 mg/m<sup>3</sup>, no cases of metal fume fever were observed for these workers. Concentrations of zinc in serum and urine for these workers averaged 11.515 µmol/L (752.85 µg/L) and 3.6705 µmol/L (239.98 µg/L), respectively, during the period of an 8-hour shift.

Zinc is found in human tissues and body fluids. As part of the 1982 National Human Adipose Tissue Survey (NHATS) conducted in the United States, the concentration of zinc in adipose tissue ranged from 1.1 to 6.0 µg/g (EPA 1986). The mean concentration of zinc was 6.95±1.08 µg/mL in whole blood samples from residents of Baajoz, Spain (a region with low environmental pollution) with zinc levels increasing with age (Moreno et al. 1999). Individuals <30, 30–45, and >45 years old had whole blood zinc concentrations of 4.85, 6.85, and 7.32 µg/mL, respectively. Blood and serum collected from 372 adolescents (15 years old) from the Swedish cities of Uppsala and Trollhättan contained zinc at median concentrations of 6.1 and 0.99 mg/L, respectively (Bárány et al. 2002). The mean concentration of zinc in the fingernails and toenails of populations from the United States, Canada, and Japan were 105, 109, and 94 mg/kg, respectively (Takagi et al. 1988). Hayashi et al. (1993) reported that human fingernail samples from Japanese individuals had higher mean levels of zinc in the spring (145–149 µg/g) compared to winter (122–136 µg/g). The geometric mean concentrations of zinc in toenails (129 mg/kg) and scalp hair (108 mg/kg) of pre-school children in Germany were about the same (Wilhelm et al. 1991). The total concentrations of zinc in 29 body tissues of 55 human cadavers were measured (Saltzman et al. 1990).

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The lowest concentration (mean of  $1.5 \pm 2.2$  mg/kg wet weight) of zinc in both males and females was found in adipose tissues, while the highest concentrations were detected in the skull of males (mean of 54.3 mg/kg wet weight) and in the skeletal muscle of females (mean of 59.0 mg/kg wet weight). The mean concentrations of zinc in the feces of low-income urban Hispanics and rural Blacks in the United States were 75 and 94 mg/kg wet weight, respectively (Prevost et al. 1985). Body tissue and fluid samples were collected from two non-occupationally exposed individuals living in the Los Angeles, California area (Krishnan and Que Hee 1992). Ear wax, blood plasma, sweat, and skin from these individuals contained zinc at levels of 88–103, 0.79–1.7, 0.50–1.58, and 15.6–1,000  $\mu\text{g/g}$  dry weight, respectively.

Concentrations of zinc in human milk are affected by the stages lactation. Arnaud and Favier (1995) found that the level of zinc in human milk will rise to a peak 2-days postpartum ( $183 \pm 70$   $\mu\text{mol/L}$  or  $12.0 \pm 4$  mg/L) and then decline during the duration of lactation (e.g., at 6-days postpartum,  $77 \pm 22$   $\mu\text{mol/L}$  or  $5.0 \pm 1.4$  mg/L). At 6-months, the concentration of zinc in human milk is only 12% of its initial levels (Dórea 2002). Wasowicz et al. (2001) observed an inverse relationship between zinc levels in blood plasma and human milk for lactating women from Poland. Mean levels of zinc in blood plasma increased from  $0.51 \pm 0.13$  mg/L (0–4 days postpartum) to  $0.76 \pm 0.20$  mg/L (10–30 days postpartum), while mean levels of zinc in human milk decreased from  $8.2 \pm 2.8$  mg/L (0–4 days postpartum) to  $1.4 \pm 0.7$  mg/L (10–30 days postpartum).

Zinc levels in maternal blood are normally higher than levels in cord blood. Maternal and cord blood of 56 mothers living in Singapore were analyzed for zinc (Ong et al. 1993). For these mothers, the mean concentrations of zinc were  $4.97 \pm 1.15$  and  $1.58 \pm 0.45$  mg/L in maternal and cord blood, respectively. During the period of 1993–1997, Raghunath et al. (2000) determined the concentration of zinc in maternal and cord blood for 148 mothers (20–25 years old) living Mumbai City, India. The mean concentrations of zinc were 6.335 and 2.527 mg/L in maternal and cord blood, respectively.

**Occupational.** As part of the National Occupational Exposure Survey (NOES) conducted from 1980 to 1983, NIOSH statistically estimated that 269 workers (including 22 women) in 22 plants were potentially exposed to elemental zinc in the workplace; also, 133,608 workers (including 17,586 women) in 6,157 plants were potentially exposed to other forms of zinc (of undefined composition) in the workplace (NIOSH 1984b). All of the workers exposed to elemental zinc were employed in the fabricated metal products industry as millwrights or assemblers. The largest numbers of workers exposed to other forms of zinc worked in the primary metal industries, with fabricated metal products, with transportation

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equipment, with stone, clay, and glass products, and in special trade contractors industries. Occupational groups with the largest numbers of exposed workers were miscellaneous machine operators (not elsewhere classified or not specified), molding and casting machine operators, janitors and cleaners, and machinists. Exposure estimates were derived from observations of the actual use of the compound and the use of trade name products known to contain the compound.

## 6.6 EXPOSURES OF CHILDREN

This section focuses on exposures from conception to maturity at 18 years in humans. Differences from adults in susceptibility to hazardous substances are discussed in 3.7 Children's Susceptibility.

Children are not small adults. A child's exposure may differ from an adult's exposure in many ways. Children drink more fluids, eat more food, breathe more air per kilogram of body weight, and have a larger skin surface in proportion to their body volume. A child's diet often differs from that of adults. The developing human's source of nutrition changes with age: from placental nourishment to breast milk or formula to the diet of older children who eat more of certain types of foods than adults. A child's behavior and lifestyle also influence exposure. Children crawl on the floor, put things in their mouths, sometimes eat inappropriate things (such as dirt or paint chips), and spend more time outdoors. Children also are closer to the ground, and they do not use the judgment of adults to avoid hazards (NRC 1993).

As for adults, sources of exposure for children to zinc include ingestion of food, drinking water, and polluted air, with ingestion of food being the primary route of exposure. In an extensive survey of foods in the total diets of children in the United States, conducted by FDA during 1982–1984, the following values for daily zinc intakes (mg/day) were estimated: 6–11-month-old infants, 5.24; 2-year-old children, 7.37; 14–16-year-old girls, 9.90; and 14–16-year-old boys, 15.61 (Pennington et al. 1986). The FDA also included drinking water in the total diet. Hair samples collected from children (10–12 years old) living rural and industrial areas of southern Poland and analyzed for zinc (Zachwieja et al. 1995). Hair samples of children from Kraków-Shakina (urban areas), Tarnów-Czechowice-Dziedzice (industrial areas), and rural areas contained zinc at concentrations of 171.5, 185.0, and 244.6 ppm, respectively (Zachwieja et al. 1995). Whole blood samples from children (3–6 years old) living in Mumbai and Hyderabad, industrialized urban areas of India, contained zinc at mean concentrations of 398.9 and 483.4 µg/dL, respectively (Tripathi et al. 2001).



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At waste sites, zinc that is found in excess of natural background levels is most likely to be in soil, and presents a special hazard for young children. Hand-to-mouth activity and eating contaminated dirt will result in oral exposure to zinc. The hazard in this case depends on the form of zinc that is present at the waste site. Zinc in soil at waste sites is in both soluble and insoluble forms; zinc in insoluble forms would be expected to be less available than more soluble forms.

Zinc exposure to children from parents' work clothes, skin, hair, tools, or other objects from the workplace is possible if the parent uses zinc or its compounds at work. Household products or products used in crafts, hobbies, or cottage industries which contain galvanized materials (e.g., nails) or zinc-containing paint will have significant amounts of zinc. Hand-to-mouth activity, chewing, and eating these materials may result in higher exposure to zinc. However, no cases of home exposure to zinc were located in the literature.

### 6.7 POPULATIONS WITH POTENTIALLY HIGH EXPOSURES

Certain populations receive greater-than-average exposures to zinc from environmental sources. For example, higher levels of zinc have been reported in soil and water near waste sites, metal smelters, and areas exposed to untreated waste water (Hutchinson and Wai 1979; Ragaini et al. 1977; Schalscha et al. 1982). Other populations at risk of high exposure are those that have galvanized plumbing in their residences, and those that intentionally consume large doses of zinc as a dietary supplement. Patients who receive chronic treatment with drugs containing zinc salts (such as injectable insulin) are exposed to higher zinc levels than the general population. Allergic reactions to the zinc in insulin have been reported (Bruni et al. 1986). People in certain occupations (e.g., non-ferrous metal smelting) are likely to be exposed to higher concentrations of zinc than the general population (see Section 6.5). However, the higher exposure may not be indicative of a long-term increase in body burden. For example, the median zinc concentration in the lung tissues of 21 Swedish workers previously employed in the refining and smelting of nonferrous metals was about the same as in a control group (11.0 versus 10.7 mg/kg wet weight) (Hewitt 1988). On the other hand, the median concentration of zinc in lung tissues of eight deceased coal miners from England was 72 mg/kg wet weight compared to a median value of 54 mg/kg wet weight for a control group (Hewitt 1988); however, the study author did not provide any evidence that the difference in zinc concentrations in the lungs of unexposed controls is statistically significant.

Individuals who smoke or who use zinc supplementation will have greater exposure to zinc. Zinc was measured in samples of cigarette tobacco from the United States at concentrations ranging from 30 to

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69 µg/g (Jenkins 1986). Levels of zinc in smoke from these cigarettes ranged from 0.34 to 1.21 µg/cigarette. Individuals, who use high-dose zinc supplementation as a potential treatment for age-related macular degeneration, will have higher exposures to zinc. Hiller et al. (1995) reported that zinc supplements will have an effect on the concentration of serum zinc in the body.

## 6.8 ADEQUACY OF THE DATABASE

Section 104(i)(5) of CERCLA, as amended, directs the Administrator of ATSDR (in consultation with the Administrator of EPA and agencies and programs of the Public Health Service) to assess whether adequate information on the health effects of zinc is available. Where adequate information is not available, ATSDR, in conjunction with the National Toxicology Program (NTP), is required to assure the initiation of a program of research designed to determine the health effects (and techniques for developing methods to determine such health effects) of zinc.

The following categories of possible data needs have been identified by a joint team of scientists from ATSDR, NTP, and EPA. They are defined as substance-specific informational needs that if met would reduce the uncertainties of human health assessment. This definition should not be interpreted to mean that all data needs discussed in this section must be filled. In the future, the identified data needs will be evaluated and prioritized, and a substance-specific research agenda will be proposed.

### 6.8.1 Identification of Data Needs

**Physical and Chemical Properties.** Data are available that adequately characterize the physical and chemical properties of the various forms of zinc to permit estimation of their environmental fate (ACGIH 1991; Baes and Sharp 1983; Baes et al. 1984; Gerritse et al. 1982; HSDB 1986, 1990; NIOSH 1990; Weast 1988; Weiss 1986; Windholz 1983).

**Production, Import/Export, Use, Release, and Disposal.** Information about current and future production of zinc and zinc compounds is available. Zinc is also one of the most widely used metals in the world (Mirenda 1986). In 2001, approximately 799,000 metric tons of zinc were produced in the United States from domestic ores. The estimated world production from mines in 2001 was 8,850,000 metric tons (USGS 2001). Information on the use of zinc and its compounds in the home, environment, and workplace is available. Zinc is most commonly used as a protective coating for other metals. It is also used in alloys such as bronze and brass, for electrical apparatus, and in organic chemical

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extractions. Zinc salts have numerous applications, including wood preservation. Zinc chloride is a primary ingredient in smoke bombs. In pharmaceuticals, zinc salts are used as solubilizing agents in drugs, including insulin (Lloyd 1984; Lloyd and Showak 1984; Windholz 1983). Zinc oxide is found in ointments used to treat burns and infectious and skin diseases (EPA 1987d). Zinc is also utilized therapeutically in human medicine in the treatment of zinc deficiency (Elinder 1986). Information on typical releases of zinc and its compounds in the home, environment, and workplace, and which environmental media are likely to be contaminated with significant quantities of zinc are available. Zinc is ubiquitous in the environment. Both natural releases and releases of human origin to the environment can be significant (EPA 1980d; Fishbein 1981; Mirenda 1986; NAS 1977; Nriagu 1989; Ragaini et al. 1977; TRI01 2003). Soils and sediments are likely to contain significant quantities of zinc and its compounds (Connor and Shacklette 1975; Rice 1999; Shacklette and Boerngen 1984; USGS 2002). Current disposal methods are efficient (Dawson and Mercer 1986; Lloyd and Showak 1984). No data were located regarding the amount of zinc being disposed. Rules and regulations regarding the disposal of zinc are available (Dawson and Mercer 1986; DOI 1991). According to the Emergency Planning and Community Right-to-Know Act of 1986, 42 U.S.C. Section 11023, industries are required to submit chemical release and off-site transfer information to EPA. The Toxics Release Inventory (TRI) contains this information for 2001. Environmental releases of zinc and zinc compounds from manufacturing and processing facilities required to report their releases are listed in Tables 6-1 and 6-2. This database is updated yearly and should provide a list of industrial production facilities and emissions.

**Environmental Fate.** Zinc partitions to the air, water, and soil (EPA 1979d; Guy and Chakrabarti 1976; Houba et al. 1983; Pita and Hyne 1975). Zinc occurs in the environment mainly in the +2 oxidation state (Lindsey 1979). Adsorption is the dominant fate of zinc, resulting in enrichment of zinc in suspended and bed sediments (EPA 1979d). The mobility of zinc in soil has been characterized (Baes and Sharp 1983; Bergkvist et al. 1989; EPA 1980d; Hermann and Neumann-Mahlkau 1985; Kalbasi et al. 1978; Saeed and Fox 1977; Tyler and McBride 1982). No estimate for the atmospheric lifetime of zinc is available. Development of pertinent data on the atmospheric processes important for zinc speciation in the atmosphere would be helpful. Development of this information would permit construction of a comprehensive model for the transport and interaction of zinc not only in air but in other media as well. Transformation in air and water can occur as a result of changes in chemical speciation (Anderson et al. 1988; EPA 1979d; EPA 1980d; Stokinger 1981). Data that describe the transformation processes for zinc in soil or the fate of zinc in soil are needed. A model of zinc flux from all environmental compartments would be useful for providing information on the overall environmental fate of zinc.

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The primary anthropogenic sources of zinc in the environment (i.e., air, water, soil) are related to mining and metallurgic operations involving zinc and use of commercial products containing zinc (EPA 1980d; NAS 1977; Nriagu and Pacyna 1988; Ragaini et al. 1977; TRI01 2003). Zinc has been detected in air, surface water, groundwater, and soil, with the frequency of detection and the concentrations greatest near source areas (e.g., hazardous waste sites and industrial areas such as lead smelters) (EPA 1980d; HazDat 2003; Lioy et al. 1978; Lloyd and Showak 1984; Mumma et al. 1984, 1990, 1991; NAS 1977).

**Bioavailability from Environmental Media.** Zinc can be absorbed following inhalation (Drinker and Drinker 1928; Hamdi 1969), ingestion (Aamodt et al. 1983; Davies 1980; Johnson et al. 1988; Methfessel and Spencer 1973; NAS/NRC 1979; Spencer et al. 1985), or dermal contact (Agren 1990; Gordon et al. 1981; Hallmans 1977; Keen and Hurley 1977). No estimates of the bioavailability of zinc after inhalation of zinc particles in air, ingestion from water and soil, or skin contact with bath water or soil were located. The bioavailability of zinc is higher in media with a low pH, as a result of increased zinc solubility and ionization. If zinc is partly present in an irreversibly adsorbed state in soil, this part is not available for skin absorption. It would be useful to develop quantitative data on the bioavailability of zinc from various environmental media.

**Food Chain Bioaccumulation.** Zinc bioconcentrates moderately in aquatic organisms, and this bioconcentration is higher in crustaceans and bivalve species than in fish (EPA 1987c; Ramelow et al. 1989). Zinc may concentrate in plants grown on contaminated soils. However, it does not biomagnify through the terrestrial food chain (Biddinger and Gloss 1984; EPA 1979d; Hegstrom and West 1989; Levine et al. 1989).

**Exposure Levels in Environmental Media.** Zinc has been detected in air (Barrie and Hoff 1985; Duce et al. 1975; EPA 1980d; Evans et al. 1984; John et al. 1973; Lioy et al. 1978; Lloyd and Showak 1984; Patterson et al. 1977; Pratt et al. 2000; Ragaini et al. 1977; Saltzman et al. 1985; Spicer et al. 1996; Zoller et al. 1974), water (Bruce and McMahon 1996; Coale and Flegal 1989; Cole et al. 1984; EPA 1980d; Hale 1977; HazDat 2003; Heit et al. 1989; Maessen et al. 1985; Minear et al. 1981; NAS 1977; Nriagu et al. 1996; Ohanian 1986; Sañudo-Wilhelmy and Gill 1999; Schock and Neff 1988; Scudlark et al. 1994; Shiller and Boyle 1985; Taylor et al. 2001; Windom et al. 1991), soil (Beavington 1975; Chen et al. 1999; Connor and Shacklette 1975; EPA 1980d; Haines 1984; HazDat 2003; Johnson et al. 1990; Mayer and Manning 1990; Mielke et al. 1999, 2000; Mumma et al. 1984, 1990, 1991; Norrström and Jacks 1999; Schalscha et al. 1982; Storm et al. 1994), and food (Coleman et al. 1992; FDA 2001; Gartrell et al. 1986a; Mahaffey et al. 1975; Weigert 1991). However, since most of the data are not current, i.e.,

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within the last 3 years, additional data would be useful to provide a more complete characterization of human exposure and the trend in zinc concentrations in various environmental media. Estimates have been made for human intake of zinc from food and drinking water (EPA 1980d; Gartrell et al. 1986a; IOM 2002; Pennington et al. 1986; Sharrett et al. 1982a, 1982b). Further data are needed on estimated daily intakes from inhalation resulting from occupational exposures.

Reliable monitoring data for the levels of zinc in contaminated media at hazardous waste sites are needed so that the information obtained on levels of zinc in the environment can be used in combination with the known body burden of zinc to assess the potential risk of adverse health effects in populations living in the vicinity of hazardous waste sites.

**Exposure Levels in Humans.** Zinc has been detected in fingernails, toenails, hair, all tissues, organs, skull and skeletal muscle, blood, feces, urine, sweat, and saliva (Greger and Sickles 1979; Hambidge et al. 1972; Henkin et al. 1975a; Llobet et al. 1988a; NAS/NRC 1979; Prasad et al. 1963a; Prevost et al. 1985; Saltzman et al. 1990; Schroeder et al. 1967; Takagi et al. 1988; Wastney et al. 1986; Wilhelm et al. 1991). Most of the data on occupational exposure levels of zinc are outdated (NIOSH 1976, 1984b). Additional information on potentially exposed workers and exposure levels would provide a more accurate characterization of occupational exposures in the United States. Current biological monitoring data on zinc are needed for populations surrounding hazardous waste sites. This information is necessary for assessing the need to conduct health studies on these populations.

**Exposures of Children.** Limited data are available regarding the exposure and body burdens of children to zinc. Children, like adults, are primarily exposed to zinc through the diet. Zinc was identified in the postpartum human milk of women at concentrations of 5 to 12 mg/L (Arnaud and Favier 1995). In an extensive survey of foods in the total diets of individuals in the United States, conducted by FDA during 1982–1984, the following values for daily zinc intakes (mg/day) were estimated for children: 6–11-month-old infants, 5.24; 2-year-old children, 7.37; 14–16-year-old girls, 9.90; and 14–16-year-old boys, 15.61 (Pennington et al. 1986). Since zinc is found in soil and children ingest soil either intentionally through pica or unintentionally through hand-to-mouth activity, pica is a unique exposure pathway for children. While zinc is found in home products such as paint, ointments, galvanized metals, coins, and dietary supplements, this exposure route should be low and will not disproportionately affect children. Continued monitoring data are necessary to understand potentially dangerous routes of childhood exposure. Child health data needs relating to susceptibility are discussed in 3.12.2 Identification of Data Needs: Children's Susceptibility.

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**Exposure Registries.** No exposure registries for zinc were located. This substance is not currently one of the compounds for which a subregistry has been established in the National Exposure Registry. The substance will be considered in the future when chemical selection is made for subregistries to be established. The information that is amassed in the National Exposure Registry facilitates the epidemiological research needed to assess adverse health outcomes that may be related to exposure to this substance.

### 6.8.2 Ongoing Studies

The Federal Research in Progress (FEDRIP 2003) database provides additional information obtainable from a few ongoing studies that may fill in some of the data needs identified in Section 6.8.1. These studies are summarized in Table 6-7.

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**Table 6-7. Ongoing Studies on the Environmental Effects of Zinc<sup>a</sup>**

Investigator	Affiliation	Study	Sponsor
Ahner BA	Cornell University, Biological and Environmental Engineering	Monitoring the bioavailability of toxic metals in soil	USDA
Basta N, Raun WR	Oklahoma State University, Agronomy	Chemistry and bioavailability of waste constituents in soils	USDA
Basta NT	Oklahoma State University, Agronomy	Heavy metal and trace element chemistry in soils: Chemical speciation and bioavailability	USDA
Bleam WF, Helmke PA	University of Wisconsin Soil Science	Verifying and quantifying the specific complexation of metals to humic substances	USDA
Chaney RL	Beltsville Agricultural Research Center	Characterization and remediation of potential trace element and phosphate risks from contaminated soils	USDA
Chaney RL	Beltsville Agricultural Research Center	Development of methods to control heavy metal contents in soils at benign or beneficial levels	USDA
Chaney RL	Beltsville Agricultural Research Center	Long-term phytoavailability and bioavailability of soil metals	USDA
Chaney RL, Angle JS	University of Maryland, Agronomy	Phytoavailability and bioavailability of heavy metals from heavy metal contaminated soil	USDA
Cox FR	North Carolina State University, Soil Science	Effects of P, Cu, and Zn from animal waste and fertilizer on crop responses and soil test interpretations	USDA
Fish RH	Lawrence Berkeley Laboratory, University of California	Removal and recovery of toxic metal ions from aqueous streams by utilization of polymer pendant ligands	DOE
Guo MG, Tyzbir R	University Of Vermont, Nutritional Sciences	Solubility and distribution of trace elements in milk based infant formula.	USDA
Harsh JB, Zamora BA, Kuo S, Pan W, Stevens RG, Flury M	Washington State University, Crop and Soil Sciences	Physical chemical state and plant availability of uranium, lead, cadmium, zinc, and arsenic in selected Washington soils	USDA
Heil D	Colorado State University, Soil and Crop Science	Biogeochemistry and management of salts and potentially toxic trace elements in arid-zone soils, sediments, and water	USDA
Helmke PA, Bleam WF	University of Wisconsin Soil Science	Reactions controlling free ion activities and solubility of soil trace elements	USDA
Hesterberg DL	North Carolina State University, Soil Science	Molecular-scale characterization and fate of soil contaminants	USDA
Kinraide TB	Agricultural Research Service	The role of binding and electrostatic attraction to roots in the uptake of heavy metals by plants	USDA

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**Table 6-7. Ongoing Studies on the Environmental Effects of Zinc<sup>a</sup>**

Investigator	Affiliation	Study	Sponsor
Kochian LV, Paolillo DJ	Cornell University, Plant Biology	Mechanisms of aluminum tolerance and heavy metal accumulation in plants	USDA
Kochian LV	Agricultural Research Service	Investigation of heavy metal bioaccumulation in plants grown on metal-polluted soils	USDA
Kochian LV	Agricultural Research Service	Mechanisms of heavy metal and radionuclide hyper-accumulation and bioavailability in higher plants	USDA
Kpombrekou- Ademawou K, Ankumah RO	Tuskegee University, Agriculture and Home Economics	Trace elements in broiler littered soils: fate and effects on nitrogen transformation	USDA
Kuo S	Washington State University, Puyallup Research and Extension Center	Chemistry and bioavailability of waste constituents in soils	USDA
Little RE	NIEHS, National Institutes of Health	Environmental pollution in eastern and central Europe	NIH
McBride MB	Cornell University, Soil, Crop and Atmospheric Science	Heavy metal solubility in contaminated soils	USDA
McBride MB	Cornell University, Soil, Crop and Atmospheric Science	Reaction and availability of toxic metals in soils	USDA
Norvell WA, Duxbury JM	Cornell University	Plant availability and geographical distribution of essential and toxic elements	USDA
Norvell WA, Welch RM, Degloria SD	Cornell University	Bioavailability and geographic distribution of nutritionally important elements in crops and soils	USDA
Odom JW	Auburn University, Agronomy and Soils	Occurrence, measurement and mapping of plant micronutrient and trace elements in Alabama soils	USDA
Parker DR	University of California, Environmental Sciences	Predicting trace-metal bioavailability from soil solution speciation: can it be done	USDA
Pierzynski GM	Kansas State University, Agronomy	Chemistry and bioavailability of waste constituents in soils	USDA
Ross DS	University of Vermont, Plant and Soil Science	Soil manganese oxides: Oxidation and retention of contaminant metals and organics	USDA
Salt DE	Purdue University, Horticulture	A dissection of the molecular mechanisms underlying metal hyperaccumulation in plants	USDA
Slaton NA	University of Arkansas, Crop, Soil and Environmental Sciences	Evaluation of fertilization practices, soil fertility, and plant nutrition for crops produced in Arkansas	USDA



## 6. POTENTIAL FOR HUMAN EXPOSURE

**Table 6-7. Ongoing Studies on the Environmental Effects of Zinc<sup>a</sup>**

Investigator	Affiliation	Study	Sponsor
Sparks DL, Ford RG	University of Delaware, Plant and Soil Sciences	Influence of aging and competitive sorption on stabilization of metals via surface precipitation in soils	USDA
Thompson ML	Iowa State University, Agronomy	Co-migration of metals and dissolved humic substances in aquifer material	USDA
Thompson ML	Iowa State University, Agronomy	Sustainable and environmentally safe management of soil resources	USDA
Welch RM, Norvell WA, Kochian LV	Agricultural Research Service	Agricultural approaches to human health through understanding soil-plant- human/animal food systems	USDA
Zelazny LW	Virginia Polytechnic Institute, Crop and Soil Environmental Sciences	Soil mineralogical controls on nutrient availability and mobility	USDA

<sup>a</sup>Source: FEDRIP 2003

DOE = Department of Energy; FEDRIP = Federal Research in Progress Database; NIEHS = National Institute of Environmental Health Services; NIH = National Institute of Health; USDA = United States Department of Agriculture

## 7. ANALYTICAL METHODS

The purpose of this chapter is to describe the analytical methods that are available for detecting, measuring, and/or monitoring zinc, its metabolites, and other biomarkers of exposure and effects of zinc. The intent is not to provide an exhaustive list of analytical methods. Rather, the intention is to identify well-established methods that are used as standard methods of analysis. Many of the analytical methods used for environmental samples are the methods approved by federal agencies and organizations such as EPA and the National Institute for Occupational Safety and Health (NIOSH). Other methods presented in this chapter are those that are approved by groups such as the Association of Official Analytical Chemists (AOAC) and the American Public Health Association (APHA). Additionally, analytical methods are included that modify previously used methods to obtain lower detection limits and/or to improve accuracy and precision.

Zinc is ubiquitous in both the environment and the laboratory. Since many biological and environmental samples contain low levels of zinc, it is easy to contaminate samples. Thus, it is imperative that special precautions be taken to avoid sample contamination in order to obtain accurate results and ensure the integrity of samples. Precautions must be taken to avoid contamination during sample collection and analysis from sources such as sampling and filtration equipment, inadequate reagent purity, and atmospheric deposition. For ultratrace analysis, the use of a clean-room laboratory with a laminar flow work station is highly recommended to avoid contamination of samples and standards with airborne particulates. In blood analysis, collection tubes are potential sources of zinc contamination (Delves 1981). An example of failure to institute proper measures to control sample contamination, which led to inaccuracies in reported data, was described by Windom et al. (1991). Methods that can be used to avoid reporting erroneous results include interlaboratory data comparison (Galloway et al. 1983) or use of standard reference materials, such as certified SRM 1549 (nonfat powdered milk) available from the National Institute of Standards and Technology (Perry 1990).

Zinc concentrations are typically quantified using instrumental methods such as atomic absorption, emission, or mass spectroscopies; x-ray fluorescence; electro-analytical techniques (e.g., stripping voltammetry); and neutron activation analysis.

## 7. ANALYTICAL METHODS

**7.1 BIOLOGICAL MATERIALS**

Table 7-1 lists the applicable analytical methods used for determining zinc in biological fluids and tissues.

Inductively coupled plasma-atomic emission spectroscopy (ICP-AES) is used for zinc determinations in blood and tissue samples (NIOSH Method 8005) and in urine (NIOSH Method 8310). Detection limits in blood and tissue are 1 µg/100 g and 0.2 µg/g, respectively, with recoveries of 100% (NIOSH 1994). Sample preparation involves acid digestion with concentrated acids. Detection of zinc in urine samples requires extraction of the metals with a polydithiocarbamate resin prior to digestion and analysis (NIOSH 1984). Detection limits in urine are 0.1 µg/sample. Inductively coupled plasma-mass spectroscopy (ICP-MS) has been used to determine the concentration of zinc in milk samples and brain tissue (Panayi et al. 2002; Patterson et al. 1992). Detection limits are 0.06 µg/sample for milk and 10.7 ng/g (for a 150 mg sample) for brain tissue samples. Recoveries ranged from 99–111% for brain tissue samples (Panayi et al. 2002).

Atomic absorption spectrometry (AAS) is a common and simple laboratory technique capable of routine zinc analysis of biological samples including bone, liver, hair, blood, and urine. Graphite furnace AAS (GF-AAS) is more sensitive than flame AAS and has been used to determine very low levels of zinc (detection limit, 0.052 µmol/L) in human milk (Arnaud et al. 1991). GF-AAS has been used to determine zinc in human semen. Recovery (96–104%) was good, and preparation by microwave wet acid dissolution was more accurate than the standard water dilution method (Alvarado et al. 1991). Zinc concentrations in liver have been accurately quantified by flame AAS. Homogenization of tissue samples coupled with flame AAS resulted in 100% recoveries, accuracies of 0–3%, and a detection limit of 0.04 mg/L (Luterotti et al. 1992). AAS has also been used to determine zinc in bloodstains on filter paper. This method is accurate, reproducible, and acceptable for routine clinical testing using both dry ashing and direct extraction sample preparation (Fan et al. 1991).

The use of stable isotopes or tracers to study zinc absorption in humans with subsequent analysis by mass spectrometry has been reported in the literature. Analysis of fecal samples obtained 3 and 6 days after the administration of zinc-65 isotope in food showed that between 45 and 75% of zinc isotope was absorbed (Johnson 1982). The results indicated satisfactory detection of the zinc-67 isotope in human feces, while the zinc-70 isotope was not as detectable. Better precision and recovery were obtained for the zinc-67 isotope (2.4% CV [coefficient of variation]; >95% recovery) than for the zinc-70 isotope (38%

## 7. ANALYTICAL METHODS

**Table 7-1. Analytical Methods for Determining Zinc in Biological Materials**

Sample matrix	Preparation method	Analytical method	Sample detection limit	Percent recovery	Reference
Blood or tissue	Acid digestion with HNO <sub>3</sub> /HClO <sub>4</sub> , H <sub>2</sub> SO <sub>4</sub> , measure at 213.9 nm	ICP-AES	1 µg/100 g (blood); 0.2 µg/g (tissue)	103	NIOSH 1994 (method 8005)
Urine	Acid digestion of oxygen plasma ashing; extract with polydithiocarbamate resin; measure at 213.9 nm	ICP-AES	0.1 µg/sample	100	NIOSH 1994 (method 8310)
Semen	Microwave wet acid digestion	GF-AAS	400 µg/L	96–104	Alvarado et al. 1991
Fingernails	Digest nail samples with concentrated nitric acid; heat at 65 °C for 1 hour; cool and dilute with deionized water	GF-AAS	No data	No data	Sohler et al. 1976
Liver	Acid digestion with mixtures of different acids; distill volatile elements	Radio-chemical NAA	No data	98	Lievens et al. 1977
Liver	Homogenize sample with water; add HCl; shake; centrifuge; dilute	Flame AAS	40 µg/L	100	Luterotti et al. 1992
Muscle tissue	Mineralize sample in muffle furnace; dissolve in HNO <sub>3</sub>	FIA	3 µg/L	No data	Fernandez et al. 1992b
Blood	Separate serum from blood by centrifugation; transfer a portion of serum into an ampule of highly pure silica and dry; irradiate capsules at a thermal neutron density of 5x10 <sup>3</sup> n/cm <sup>-2</sup> /second <sup>-1</sup>	Instrumental NAA	No data	>100	Jurgensen and Behne 1977
Blood	Feed radiotracer <sup>65</sup> zinc; measure zinc activity in blood at 14 days	Tracer technique	No data	88	Watson et al. 1987
Blood serum and red blood cells	Feed <sup>68</sup> zinc and <sup>70</sup> zinc and measure blood levels in a 24-hour sample and a sample taken immediately after zinc administration; wet ash sample; add APDC precipitant; dissolve precipitate in HNO <sub>3</sub> irradiate	Isotope tracer technique	No data	No data	Janghorbani et al. 1981
Blood	Feed <sup>65</sup> ZnCl <sub>2</sub> orally; measure zinc blood levels and whole blood count	Radiotracer technique—whole blood count and blood level measurement	No data	88	Watson et al. 1987

## 7. ANALYTICAL METHODS

**Table 7-1. Analytical Methods for Determining Zinc in Biological Materials**

Sample matrix	Preparation method	Analytical method	Sample detection limit	Percent recovery	Reference
Bloodstain	Place drop of blood on filter paper; cut away excess paper; optional dry ash; add HCl; shake	Flame AAS	No data	No data	Fan et al. 1991
Thoracic aorta, lung, myocardium, spleen	Homogenize sample; complete wet ashing with HNO <sub>3</sub>	Flame AAS	No data	No data	Marks et al. 1972
Brain tissue	Digest with HNO <sub>3</sub> using microwave digestion; dilute	ICP-MS	32 mg/L (10.7 ng/g for 150 mg sample)	99–111	Panayi et al. 2002
Feces	Give <sup>67</sup> Zn through diet; treat fecal samples with H <sub>2</sub> O <sub>2</sub> ; prepare chelates	Isotope tracer technique	No data	>95 ( <sup>67</sup> Zn); 71 ( <sup>70</sup> Zn)	Johnson 1982
Feces	Feed <sup>70</sup> Zn, <sup>68</sup> Zn, and <sup>64</sup> Zn orally; homogenize sample; evaporate; ash; HNO <sub>3</sub> digestion; boil; evaporate; add HCl; transfer to anion exchange column; prepare eluate; irradiate	Isotope tracer technique; NAA	No data	No data	Ni et al. 1991
Bone	Acid digestion of dried bone ash with concentrated HNO <sub>3</sub> ; evaporate to dryness and add more concentrated HNO <sub>3</sub> ; remove silica residue by filtration; transfer samples to polyethylene bottles	Flame AAS	No data	No data	Szpunar et al. 1978
Hair	Digest clean sample in acid mixture	Flame AAS	20 µg/g	No data	Wilhelm et al. 1991
Hair	Rinse sample with hexane; wet or dry ash with HNO <sub>3</sub>	EDXRF	0.001 µg/L	No data	Folin et al. 1991
Hair	Rinse sample with hexane; wet or dry ash with HNO <sub>3</sub>	Flame AAS	0.001 µg/L	No data	Folin et al. 1991
Hair	Digest clean sample in acid mixture	ICP-AES	No data	81–102	Takagi et al. 1988
Serum (animal)	Add Brij 35 to sample; mix	Flame AAS	~0.6 µg/mL	No data	AOAC 1990 (method 991.11)
Serum and plasma	Separate serum and plasma by centrifugation; keep stored in glass tubes at -20 °C until analysis; thaw to room temperature prior to analysis	Flame AAS	No data	No data	Shaw et al. 1982

## 7. ANALYTICAL METHODS

**Table 7-1. Analytical Methods for Determining Zinc in Biological Materials**

Sample matrix	Preparation method	Analytical method	Sample detection limit	Percent recovery	Reference
Milk	Ash; lyophilize; wet-ash with HNO <sub>3</sub> ; add H <sub>2</sub> O <sub>2</sub> ; dry; dissolve in HCl and NH <sub>4</sub> Cl; extract with DDDC	ICP-MS	0.06 µg/sample	No data	Patterson et al. 1992
Milk	Dilute sample with Triton X-100	GF-AAS	0.052 µmol/L	86–106	Arnaud et al. 1991
Saliva	Lashley cup place over one of the Stenson's ducts; secretion stimulated with lemon candies; discard first 5–10 mL; collect ≈120 mL	GF-AAS	No data	No data	Langmyhr et al. 1979

AAS = atomic absorption spectroscopy; AES = atomic emission spectroscopy; APDC = ammonium pyroldine dithiocarbamate; Brij 35 = polyoxyethylene (35) lauryl ether; DDDC = diethylammonium diethyldithiocarbamate; EDXRF = energy dispersive x-ray fluorescence; FIA = flow injection analysis; GF = graphite furnace; HCl = hydrochloric acid; HClO<sub>4</sub> = perchloric acid; HNO<sub>3</sub> = nitric acid; H<sub>2</sub>O<sub>2</sub> = hydrogen peroxide; H<sub>2</sub>SO<sub>4</sub> = sulfuric acid; ICP = inductively coupled plasma spectroscopy; MS = mass spectrometry; NAA = neutron activation analysis; NH<sub>4</sub>Cl = ammonium chloride; Zn = zinc; ZnCl<sub>2</sub> = zinc chloride

## 7. ANALYTICAL METHODS

CV; 71% recovery). Sample detection limits were not reported. Total reported sample preparation time was <2 hours, and it took only 5–10 minutes to analyze each sample on the mass spectrometer.

Multi-elemental analysis has been used to detect zinc and other trace metals in biological fluids and tissues. For determination of metallic constituents in biological samples, such as liver, samples were digested with mixtures of different acids, volatile elements were distilled by selective distillation, and a cleanup step was performed using ion exchange chromatography prior to assay by neutron activation analysis (NAA) (Lievens et al. 1977). Recovery (98%) and precision (<10% CV) were excellent. Although the limit of detection for zinc was not reported, based on the reported results, this method can detect levels ranging from the low- to the sub-ppm range (Lievens et al. 1977). The NAA technique has also been used to detect zinc in urine and blood samples. Jurgensen and Behne (1977) used the technique to measure human serum levels of trace elements including zinc. Recovery and precision for this method are very good. Sensitivity was not reported.

A practical method, based on NAA, was developed for accurate measurement of the stable isotopes zinc-68 and zinc-70 in human plasma and red blood cells (Janghorbani et al. 1981). This method can provide an alternative to the use of radiolabeled zinc. It is more complex and time consuming than those used to measure radiolabeled zinc levels. As with any isotopic method, isotope exchange may invalidate calculation of net absorption, but this potential problem was not investigated. Precision was very good (<10%). Sensitivity and accuracy were not reported.

Radionuclide studies offer an additional method to investigate the factors that affect trace element absorption. Radioactivity emitted by the radionuclide was measured in blood 14 days after the oral ingestion of zinc-65 and compared with the amount of radioactivity emission determined by whole-body counting (Watson et al. 1987). The results indicated that, where whole-body counting facilities were not available, measurement of radioactivity emitted in blood was a reasonable alternative for the prediction of zinc absorption. Recovery for this method was adequate (88%); precision was acceptable (<17% CV). The limit of detection for zinc was not reported.

Other analytical methods include flow injection analysis (FIA). FIA has been used to determine very low levels of zinc in muscle tissue. This method provides very high sensitivity, low detection limits (3 ng/mL), good precision, and high selectivity at trace levels (Fernandez et al. 1992b).

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Animal and human tissues samples are usually analyzed without drying and concentrations are reported as wet weight. For some samples, freeze-drying has been used. Care should be taken during the acid dissolution of blood and urine samples as frothing of natural surfactants during digestion can lead to losses. This problem can be prevented by allowing the sample to stand overnight after the addition of acid (WHO 2001).

## 7.2 ENVIRONMENTAL SAMPLES

Table 7-2 lists the methods used for analyzing zinc in environmental samples.

ICP-AES is used to determine concentrations of zinc in air (NIOSH method 7300), water (EPA methods 3120 B, 6010 C, 200.7; APHA methods 3120B, 3125B, 3130B), solid wastes (AOAC method 990.08), and soil (EPA methods 6010, 3050) (AOAC 1998; APHA 1998; EPA 1986a, 1994; NIOSH 1994). Detection limits in air, water, and solid wastes are 0.6, 2, and, 2 µg/L, respectively (AOAC 1998; EPA 1994; NIOSH 1994). Preparation for water samples typically involves acid digestion with concentrated acids. The concentration of zinc in soil was determined by ICP-AES coupled with an ammonium bicarbonate-diethylenetriaminepentaacetic acid ( $\text{NH}_4\text{HCO}_3$ -DTPA) extraction procedure. This method can be used to screen soils for zinc (Boon and Soltanpour 1991). ICP-MS has been used to determine the concentration of zinc in water (EPA methods 200.8, 1638; APHA method 3125 B), (APHA 1998; EPA 1994, 1996). Detection limits have been reported to be as low as 0.017 µg/L using  $^{66}\text{Zn}$  isotope. Recoveries range from 99 to 117% (APHA 1998).

Flame AAS has been used to determine zinc concentrations in natural waters (Fishman 1966). AAS is a rapid method of measuring zinc, with a detection limit of 0.005 ppm. Brooks et al. (1967) demonstrated a simple extraction system consisting of two reagents, ammonium pyrrolidine dithiocarbamate (APDC) and methyl isobutylketone (MIBK), with subsequent analysis by flame AAS to measure particulate and "soluble" zinc in seawater. Sensitivity was in the sub-ppm range, and precision was good (3% CV). Flame AAS, coupled with microwave digestion and GF-AAS, has been used to determine the concentration of zinc in food and shellfish samples. Limits of detection ranged from 0.12 to 0.24 ppm, with recoveries ranging from 80 to 113%. Precision and recovery using microwave digestion were comparable to traditional wet ashing and superior to dry ashing in shellfish samples (AOAC 1984; McCarthy and Ellis 1991; Morales-Rubio et al. 1992). GF-AAS was also used to determine low levels of zinc in beer. Recovery (94–106%) and precision (4.2% CV) were excellent. Sensitivity was not reported (Wagner et al. 1991). Flame AAS has been used to measure heavy metals, including zinc, in various oil



## 7. ANALYTICAL METHODS

**Table 7-2. Analytical Methods for Determining Zinc in Environmental Samples**

Sample matrix	Preparation method	Analytical method	Sample detection limit	Percent recovery	Reference
Air	Collect air particulates on Teflon filters; digest with HNO <sub>3</sub>	NAA (non-destructive)	No data	No data	Zoller et al. 1974
Air	Collect sample on cellulose filter; wet ash filter with HNO <sub>3</sub> /HClO <sub>4</sub> ; dilute	ICP-AES	0.6 ng/mL	94–101	NIOSH 1994 (method 7300)
Air	Collect sample on cellulose filter; wet ash with HNO <sub>3</sub> ; dilute	Flame AAS	3 µg/sample	No data	NIOSH 1994 (method 7030)
Air (as zinc oxide)	Collect sample on PVC-acrylonitrile filter	XRD	5 µg/sample	No data	NIOSH 1994 (method 7502)
Atmospheric aerosols	Collect sample on cellulose filter; digest with HNO <sub>3</sub> ; filter; dry; add HNO <sub>3</sub> ; adjust pH; add KNO <sub>3</sub>	Anodic stripping voltammetry	13.7 µg/L	No data	Casassas et al. 1991
Water	Acidify; dilute	ICP-MS	0.14 µg/L	No data	EPA 1996 (method 1638)
Water	Reflux with HNO <sub>3</sub> /HCl; dilute	GF-AAS	0.14 µg/L	No data	EPA 1996 (method 1639)
Water	Acidify (digest if necessary); dilute	ICP-AES	2 µg/L	No data	EPA 1994 (method 200.7)
Water	Acidify (digest if necessary); dilute	ICP-MS	1.8 µg/L	No data	EPA 1994 (method 200.8)
Water	Acid digestion; dilute	ICP-AES	1.2 µg/L	No data	EPA 2000 (method 6010 C)
Water	Dissolve in HCl; dilute	Flame AAS	5 µg/L	No data	APHA 1998 (method 3111 B)
Water	Chelation with ammonium pyrrolidine dithiocarbamate and extraction into MIBK	Flame AAS	No data	No data	APHA 1998 (method 3111C)
Water	Acidify; dilute	ICP-AES	2 µg/L	No data	APHA 1998 (method 3120 B)
Water	Acidify; dilute	ICP-MS	0.017 µg/L ( <sup>66</sup> Zn) 0.020 µg/L ( <sup>68</sup> Zn)	99–117 ( <sup>66</sup> Zn) 98–116 ( <sup>66</sup> Zn)	APHA 1998 (method 3125 B)
Water	Dilute with HNO <sub>3</sub>	Anodic stripping voltammetry	<1 µg/L	No data	APHA 1998 (method 3130 B)

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**Table 7-2. Analytical Methods for Determining Zinc in Environmental Samples**

Sample matrix	Preparation method	Analytical method	Sample detection limit	Percent recovery	Reference
Water	Add sodium ascorbate; KCN; zircon (2-carboxy-2'-hydroxy-5'-sulfoformazyl benzene)	Colormetry	No data	No data	APHA 1998 (method 3500-Zn B)
Water and waste water	Acid digestion	Flame AAS	5 µg/L	No data	EPA 1979 (method 289.1)
Water and waste water	Acidify; dilute	GF-AAS	0.05 µg/L	No data	EPA 1979 (method 289.2)
Water	Mineralize sample in muffle furnace; dissolve in HNO <sub>3</sub>	FIA	3 µg/L	No data	Fernandez et al. 1992b
Seawater	APDC-MIBK extraction	Flame AAS	0.05 ppb	No data	Brooks et al. 1967
Seawater	Take a sample digest in the electrochemical cell; adjust pH; add chelating agent and aerate	Cathodic stripping voltammetry	7x0 <sup>-11</sup> M	No data	van den Berg 1986
Crude oil	Digest sample with HNO <sub>3</sub> ; extract with MIBK or dilute with MIBK	Flame AAS	0.8 µg/g	No data	Elson et al. 1981
Soil, solid waste, sludges	Acid digestion	ICP-AES or flame AAS	2 µg/L (in solution)	102.5 at 80 µg/L	EPA 1986a (methods 6010 and 3050)
Soil, solid waste and sludges	None	Flame AAS	0.005 µg/L	No data	EPA 1986a (method 7950)
Solid wastes	No data	ICP-AES	2 µg/L	No data	AOAC 1998 (method 990.08)
Soil	Extract with DTPA and NH <sub>4</sub> HCO <sub>3</sub> -DTPA	ICP-AES	No data	No data	Boon and Soltanpour 1991
Plants	Digest samples with acids	Flame AAS	No data	No data	AOAC 1984 (method 3.013)
Plants	Digest samples with acid; extract with dithiozone reagent and CCl <sub>4</sub> ; add HCl and CCl <sub>4</sub> ; read at 525 nm for mixed-color method and at 535 nm for single-color method	Mixed and single color methods – spectrophotometric analysis	No data	No data	AOAC 1984 (methods 3.054 and 3.061)
Food	Digest sample with acid mixtures; remove sulfide, nickel, and cobalt; add dithioxone and CCl <sub>4</sub> ; measure transmission at 540 nm	Colorimetry	No data	No data	AOAC 1984 (method 25.168)

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**Table 7-2. Analytical Methods for Determining Zinc in Environmental Samples**

Sample matrix	Preparation method	Analytical method	Sample detection limit	Percent recovery	Reference
Food	Wet ash using Kjeldahl digestion HNO <sub>3</sub> /H <sub>2</sub> SO <sub>4</sub> with heat; dilute; alternatively, dry ash; dissolve in HCl with heat	Flame AAS	No data	No data	AOAC 1990 (method 969.32)
Food	Digest samples with acid mixtures; dilute	Flame AAS	No data	No data	AOAC 1990 (method 986.15)
Food	Dry ash sample in muffle oven; dilute with HNO <sub>3</sub>	Flame AAS; Flame AES	0.24 µg/g	97–100	Morales-Rubio et al. 1992
Food	Clarify; de-gas; dilute with deionized water; add HNO <sub>3</sub> to solid samples	GF-AAS	No data	90–113	Wagner et al. 1991
Food	Blend; lyophilize; grind; oven-dry; press into pellets	EDXRF	0.8 ppm	No data	Nielson et al. 1991
Shellfish	HNO <sub>3</sub> digestion in microwave; dilute	Flame AAS	0.12 ppm	80	McCarthy and Ellis 1991

AAS = atomic absorption spectroscopy; AES = atomic emission spectrometry; APDC = ammonium pyroline dithiocarbamate; CCl<sub>4</sub> = carbon tetrachloride; DTPA = diethylenetriaminepentaacetic acid; EDXRF = energy dispersive x-ray fluorescence; FIA = flow-injection analysis; GF = graphite furnace; HCl = hydrochloric acid; HClO<sub>4</sub> = perchloric acid; HNO<sub>3</sub> = nitric acid; H<sub>2</sub>SO<sub>4</sub> = sulfuric acid; ICP = inductively coupled plasma spectroscopy; KCN = potassium cyanide; KNO<sub>3</sub> = potassium nitrite; MIBK = methyl isobutyl ketone; NAA = neutron activation analysis; NH<sub>4</sub>HCO<sub>3</sub>-DTPA = ammonium bicarbonate-diethylenetriaminepentaacetic acid; PVC = polyvinyl chloride; XRD = x-ray diffraction; Zn = zinc.

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samples collected at different stages of oil refining (Elson et al. 1981). These samples were prepared using three techniques (digestion, extraction, and dilution) prior to AAS analysis; recovery from crude oil was higher with wet digestion. Sensitivity for zinc was in the low-ppm range.

Cathodic stripping voltammetry, also known as adsorption voltammetry, has been used to detect various metal ions in a  $10^{-10}$ – $10^{-11}$  M range in seawater (van den Berg 1986). APDC was used as a chelating agent for zinc. Because of the great sensitivity and specificity of APDC for zinc, it can be detected directly in the unaltered sample. Similarly, differential pulse cathodic stripping voltammetry (DPCSV) and differential pulse anodic stripping voltammetry (DPASV) after complexation with APDC have been used for determining zinc speciation at nanomolar concentrations in ocean waters (Donat and Bruland 1990). Anodic stripping voltammetry (ASV) has been used to detect zinc and other metal ions simultaneously at trace levels in atmospheric aerosols. This method is primarily used for small samples with very low concentrations of zinc. The limit of detection was 13.7 ng/L (Casassas et al. 1991).

An ion chromatographic method has been proposed for simultaneous determination of several elements including zinc in soil (Basta and Tabatabai 1990). In this method, after preliminary sample treatment, the metals are separated by ion chromatography, and the separated elements are quantified by ultraviolet-visible detection of zinc-PAR (4-[2-pyridylazo] resorcinol) colored complexes. The limit of detection for zinc by this method was 5 ppb in soil extract. Precision was  $\leq 2.5\%$  CV.

Other analytical methods include energy dispersive x-ray fluorescence (EDXRF). This technique has been used to detect zinc in dried food samples with better precision (e.g., detection limit, 0.8 ppm) than AAS methods (Nielson et al. 1991).

### 7.3 ADEQUACY OF THE DATABASE

Section 104(i)(5) of CERCLA, as amended, directs the Administrator of ATSDR (in consultation with the Administrator of EPA and agencies and programs of the Public Health Service) to assess whether adequate information on the health effects of zinc is available. Where adequate information is not available, ATSDR, in conjunction with the National Toxicology Program (NTP), is required to assure the initiation of a program of research designed to determine the health effects (and techniques for developing methods to determine such health effects) of zinc.

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The following categories of possible data needs have been identified by a joint team of scientists from ATSDR, NTP, and EPA. They are defined as substance-specific informational needs that if met would reduce the uncertainties of human health assessment. This definition should not be interpreted to mean that all data needs discussed in this section must be filled. In the future, the identified data needs will be evaluated and prioritized, and a substance-specific research agenda will be proposed.

### 7.3.1 Identification of Data Needs

#### Methods for Determining Biomarkers of Exposure and Effect.

**Exposure.** ICP-AES, ICP-MS, and AAS are the most commonly used analytical methods to determine zinc levels in plasma, bone, fingernails, hair, and other biological tissues and body fluids (Alvarado et al. 1991; AOAC 1990; Arnaud et al. 1991; Fan et al. 1991; Folin et al. 1991; Langmyhr et al. 1979; Luterotti et al. 1992; Marks et al. 1972; NIOSH 1984a, 1994; Panayi et al. 2002; Patterson et al. 1992; Shaw et al. 1982; Sohler et al. 1976; Szpunar et al. 1978; Takagi et al. 1988; Wilhelm et al. 1991). These methods generally are sensitive enough to measure background levels in the population and levels at which biological effects occur. However, improved sensitivity and recovery data are needed in order to better evaluate the relationship between body and environmental exposure levels of zinc. Other methods that are specific for measuring zinc in biological fluids and tissues include NAA, FIA, and isotope tracers techniques (Fernandez et al. 1992b; Janghorbani et al. 1981; Johnson 1982; Lievens et al. 1977; NIOSH 1984a; Watson et al. 1987). Sensitivity and/or recovery data for these methods are needed to more fully evaluate the reliability of these methods as predictors of environmental exposure.

**Effect.** Although several biomarkers for the effects of zinc have been identified (increased levels of serum amylases and lipase, non-iron responsive anemia, and decreased HDL cholesterol levels), these biomarkers of effect are not specific for zinc (Cotran et al. 1989; Suber 1989). Standard laboratory tests are available that can measure these biomarkers (Henry 1984). These methods are sensitive, accurate, and reliable enough to measure background levels in the population and levels at which biological effects occur. The development of methods for determining biomarkers of effect specific for zinc would be beneficial in assessing whether an individual has been exposed to elevated levels of zinc.

#### Methods for Determining Parent Compounds and Degradation Products in Environmental

**Media.** Methods of adequate sensitivity and specificity are available for determining levels of zinc in environmental media (AOAC 1984; APHA 1998; Basta and Tabatabai 1990; Brooks et al. 1967; Casassas

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et al. 1991; Donat and Bruland 1990; Elson et al. 1981; EPA 1979c, 1986a, 1994, 1996, 2000; Fishman 1966; McCarthy and Ellis 1991; Morales-Rubio et al. 1992; Nielson et al. 1991; NIOSH 1994; van den Berg 1986; Wagner et al. 1991; Zoller et al. 1974). Most of these methods are precise and sensitive enough to measure background levels in the environment and levels at which health effects occur. Some methods can distinguish between soluble zinc, insoluble zinc, and chelated zinc in water (Donat and Bruland 1990).

### 7.3.2 Ongoing Studies

The information in Table 7-3 was found as a result of a search of the Federal Research in Progress database (FEDRIP 2003).

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**Table 7-3. Ongoing Studies on Analytical Methods for Zinc<sup>a</sup>**

Investigator	Affiliation	Study	Sponsor
Michel RG, Freake HC, Zinn SA, Kumar CV, Bruchner C	University of Connecticut	Capillary electrophoresis to enable zinc speciation for studies of zinc homeostasis	USDA
Panemangalore M	Kentucky State University, Human Nutrition Research Program	Evaluation of biomarkers of zinc and copper status in animals and humans	USDA

<sup>a</sup>Source: FEDRIP 2003

FEDRIP = Federal Research in Progress Database; USDA = United States Department of Agriculture

## 8. REGULATIONS AND ADVISORIES

Zinc (fume and dust) and its compounds are on the list of chemicals appearing in "Toxic Chemicals Subject to Section 313 of the Emergency Planning and Community Right-to-Know Act of 1986" (EPA 1991d).

The national and state regulations and guidelines pertaining to zinc and compounds in air, water, and food are summarized in Table 8-1. No international regulations or guidelines applicable to zinc or its compounds were found.

ATSDR has derived intermediate oral MRL for zinc based on decreased erythrocyte superoxide dismutase, a sensitive indicator of body copper status, in women given supplements containing zinc gluconate for 10 weeks (Yadrick et al. 1989). The intermediate oral MRL has been adopted as the chronic oral MRL.

EPA has derived oral reference doses (RfDs) of 0.3 mg/kg/day for zinc (IRIS 2003). EPA has not derived an inhalation reference concentration (RfC) for zinc.



## 8. REGULATIONS AND ADVISORIES

**Table 8-1. Regulations and Guidelines Applicable to Zinc and Zinc Compounds**

Agency	Description	Information	Reference
<u>INTERNATIONAL</u>			
Guidelines:			
IARC	Carcinogenicity classification	No data	
WHO	Drinking water and air quality guidelines	No data	
<u>NATIONAL</u>			
Regulations and Guidelines:			
a. Air			
ACGIH	TLV (8-hour TWA)	No data	
EPA	Hazardous air pollutant pursuant to Section 112 of the Clean Air Act	Zinc Zinc oxide	EPA 2003e 40 CFR 61.01
NIOSH	REL (10-hour TWA)		NIOSH 2003a,2003b
	Zinc chloride (fume)	1 mg/m <sup>3</sup>	
	Zinc oxide (dust and fume)	5 mg/m <sup>3</sup>	
	STEL (15-minute TWA)		
	Zinc chloride (fume)	2 mg/m <sup>3</sup>	
	Zinc oxide (fume)	10 mg/m <sup>3</sup>	
OSHA	Ceiling		OSHA 2003a 29 CFR 1910.1000, Table Z-1
	Zinc oxide (dust)	15 mg/m <sup>3</sup>	
	IDLH		
	Zinc chloride (fume)	50 mg/m <sup>3</sup>	
	Zinc oxide	500 mg/m <sup>3</sup>	
	PEL (8-hour TWA) for general industry		
	Zinc chloride (fume)	1 mg/m <sup>3</sup>	
	Zinc oxide (fume and respirable fraction of dust)	5 mg/m <sup>3</sup>	
	Zinc oxide (total dust)	15 mg/m <sup>3</sup>	
	PEL (8-hour TWA) for construction industry		
Zinc chloride (fume)	1 mg/m <sup>3</sup>		
Zinc oxide (fume and respirable fraction of dust)	5 mg/m <sup>3</sup>		
Zinc oxide (total dust)	15 mg/m <sup>3</sup>		
PEL (8-hour TWA) for shipyard industry			
Zinc chloride (fume)	1 mg/m <sup>3</sup>		
Zinc oxide (fume and respirable fraction of dust)	5 mg/m <sup>3</sup>		
Zinc oxide (total dust)	15 mg/m <sup>3</sup>		
b. Water			
EPA	Drinking water health advisories		EPA 2002
	1-day (10-kg child)		
	10-day (10-kg child)	6 mg/L	
	DWEL <sup>a</sup>	6 mg/L	
	Lifetime <sup>b</sup>	10 mg/L 2 mg/L	

## 8. REGULATIONS AND ADVISORIES

**Table 8-1. Regulations and Guidelines Applicable to Zinc and Zinc Compounds**

Agency	Description	Information	Reference
<u>NATIONAL</u> (cont.)			
EPA	Hazardous substance in accordance with Section 311 (b)(2)(A) of the Clean Water Act	Zinc chloride Zinc sulfate	EPA 2003k 40 CFR 116.4
	Hazardous substance in accordance with Section 311 of the Clean Water Act; reportable quantities		EPA 2003g 40 CFR 117.3
	Zinc chloride	1,000 pounds	
	Zinc sulfate	1,000 pounds	
	National secondary drinking water regulations; secondary MCL for zinc	5 mg/L	EPA 2003f 40 CFR 143.3
	Pollutants of initial focus in the Great Lakes Water Quality Initiative	Zinc	EPA 2003l 40 CFR 132, Table 6
	Reportable quantities of hazardous substances designated pursuant to Section 311 of the Clean Water Act		
	Zinc chloride	1,000 pounds	
	Zinc sulfate	1,000 pounds	
	Toxic pollutant designation pursuant to Section 307(a)(1) of the Clean Water Act	Zinc and compounds	EPA 2003c 40 CFR 401.15
c. Food			
EPA	Tolerances for residues (ppm) of a fungicide (mancozeb), which contains 20% manganese, 2.5% zinc, and 77.5% ethylene-bisdi-thiocarbamate		EPA 2003j 40 CFR 180.176
	Apple	7	
	Asparagus	0.1	
	Banana	4	
	Banana, pulp (no peel)	0.5	
	Barley, grain	5	
	Barley, milled feed fractions	20	
	Barley, straw	25	
	Beet, sugar	2	
	Beet, sugar tops	65	
	Carrots	2	
	Celery	5	
	Corn, fodder and forage	5	
	Corn, grain	0.1	
	Cottonseed	0.5	
	Crabapple	10	
	Cranberry	7	
	Cucumber	4	
	Fennel	10	
	Fresh Corn	0.5	

## 8. REGULATIONS AND ADVISORIES

**Table 8-1. Regulations and Guidelines Applicable to Zinc and Zinc Compounds**

Agency	Description	Information	Reference
<u>NATIONAL</u> (cont.)			
EPA	Tolerances for residues (cont.)		EPA 2003j
	Grape	7	40 CFR 180.176
	Kidney	0.5	
	Liver	0.5	
	Melon	4	
	Oat, bran and milled feed fractions	20	
	Oat, grain	5	
	Oat, straw	25	
	Onion, dry bulb	0.5	
	Papaya	10	
	Peanut	0.5	
	Peanut vine hay	65	
	Pear	10	
	Popcorn grain	0.5	
	Quince	10	
	Rye, grain	5	
	Rye, milled feed fractions	20	
	Rye, straw	25	
	Squash, summer	4	
	Tomato	4	
	Wheat, grain	5	
	Wheat, milled feed fractions	20	
	Wheat, straw	25	
FDA	Bottled drinking water (allowable concentration of zinc); mineral water is exempt based on aesthetically allowable levels and do not relate to health concern	5.0 mg/L	FDA 2003a 21 CFR 165.110
	Drug products containing certain active ingredients offered over-the-counter (OTC) for certain uses; ingredients used in skin protectant drug products	Zinc chloride Zinc oxide Zinc sulfate	FDA 2003c 21 CFR 310.545
	Nutrition labeling of food; statement of the amount per serving of zinc calculated as a percent of the RDI and expressed as percent of Daily Value	15 mg	FDA 2003b 21 CFR 101.9(c)(8)
	Substances generally recognized as safe; trace minerals added to animal feeds as nutritional dietary supplements when added at levels consistent with good feeding practices	Zinc chloride Zinc oxide Zinc sulfate	FDA 2003d 21 CFR 582.80

## 8. REGULATIONS AND ADVISORIES

**Table 8-1. Regulations and Guidelines Applicable to Zinc and Zinc Compounds**

Agency	Description	Information	Reference
<u>NATIONAL</u> (cont.)			
FDA	Zinc oxide and zinc sulfate are substances generally recognized as safe when used in accordance with good manufacturing practices		FDA 2003e 21 CFR 182.8991  FDA 2003e 21 CFR 182.8997
d. Other			
EPA	Carcinogenicity classification	D <sup>c</sup>	IRIS 2003
	RfD	3x10 <sup>-1</sup> mg/kg/day	IRIS 2003
	Community right-to-know; release reporting; effective date of reporting for zinc (fume and dust)	01/01/87	EPA 2003j 40 CFR 372.65
	Designated as a hazardous substance pursuant to Section 311(b)(2) of the Clean Water Act; reportable quantity Zinc and compounds	Not assigned	EPA 2003b 40 CFR 302.4
	Designated as a hazardous substance pursuant to Section 307(a) of the Clean Water Act; reportable quantities Zinc chloride Zinc sulfate	1,000 pounds 1,000 pounds	EPA 2003b 40 CFR 302.4
	Hazardous constituent for municipal solid waste landfills	Zinc	EPA 2003a 40 CFR 258, Appendix II
	Land disposal restrictions; universal treatment standards for zinc		EPA 2003d 40 CFR 268.48
	Waste water standard	2.61 mg/L	
	Non-waste water standard	4.3 mg/L TCLP	
	Standards for owners and operators of hazardous waste TSD facilities; groundwater monitoring for zinc	Suggested <u>method</u> 6010 7950 7951	<u>PQL</u> 20 µg/L 50 µg/L 0.5 µg/L
			EPA 2003h 40 CFR 264, Appendix IX
<u>STATE</u>			
a. Air	No data		
b. Water			
Arizona	Drinking water guideline (zinc and zinc compounds)	5 mg/L	HSDB 2003
Illinois	Drinking water standard (zinc and zinc compounds)	5 mg/L	HSDB 2003
Minnesota	Drinking water guideline (zinc and zinc compounds)	2 mg/L	HSDB 2003

## 8. REGULATIONS AND ADVISORIES

**Table 8-1. Regulations and Guidelines Applicable to Zinc and Zinc Compounds**

Agency	Description	Information	Reference
<u>STATE</u> (cont.)			
c. Food	No data		
d. Other	No data		

<sup>a</sup>DWEL: a lifetime exposure concentration protective of adverse, non-cancer health effects, that assumes all of the exposure to a contaminant is from drinking water.

<sup>b</sup>Lifetime: the concentration of a chemical in drinking water that is not expected to cause any adverse noncarcinogenic effects for a lifetime of exposure. The Lifetime HA is based on exposure of a 70-kg adult consuming 2 L water/day.

<sup>c</sup>D: not classifiable as to human carcinogenicity

ACGIH = American Conference of Governmental Industrial Hygienists; CFR = Code of Federal Regulations; DWEL = drinking water equivalent level; EPA = Environmental Protection Agency; FDA = Food and Drug Administration; HA = health advisory; HSDB = Hazardous Substances Data Bank; IARC = International Agency for Research on Cancer; IDLH = immediately dangerous to life or health; IRIS = Integrated Risk Information System; MCL = maximum contaminant level; NIOSH = National Institute for Occupational Safety and Health; OSHA = Occupational Safety and Health Administration; PEL = permissible exposure limit; PQL = practical quantitation level; RDI = recommended daily intake; REL = recommended exposure limit; RfD = reference dose; STEL = short-term exposure limit; TCLP = toxicity characteristic leachate procedure; TLV = threshold limit values; TSD = treatment, storage, and disposal; TWA = time-weighted average; WHO = World Health Organization

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## 10. GLOSSARY

**Absorption**—The taking up of liquids by solids, or of gases by solids or liquids.

**Acute Exposure**—Exposure to a chemical for a duration of 14 days or less, as specified in the Toxicological Profiles.

**Adsorption**—The adhesion in an extremely thin layer of molecules (as of gases, solutes, or liquids) to the surfaces of solid bodies or liquids with which they are in contact.

**Adsorption Coefficient ( $K_{oc}$ )**—The ratio of the amount of a chemical adsorbed per unit weight of organic carbon in the soil or sediment to the concentration of the chemical in solution at equilibrium.

**Adsorption Ratio ( $K_d$ )**—The amount of a chemical adsorbed by a sediment or soil (i.e., the solid phase) divided by the amount of chemical in the solution phase, which is in equilibrium with the solid phase, at a fixed solid/solution ratio. It is generally expressed in micrograms of chemical sorbed per gram of soil or sediment.

**Anthropogenic**—Caused by human activities.

**Benchmark Dose (BMD)**—Usually defined as the lower confidence limit on the dose that produces a specified magnitude of changes in a specified adverse response. For example, a  $BMD_{10}$  would be the dose at the 95% lower confidence limit on a 10% response, and the benchmark response (BMR) would be 10%. The BMD is determined by modeling the dose response curve in the region of the dose response relationship where biologically observable data are feasible.

**Benchmark Dose Model**—A statistical dose-response model applied to either experimental toxicological or epidemiological data to calculate a BMD.

**Bioconcentration Factor (BCF)**—The quotient of the concentration of a chemical in aquatic organisms at a specific time or during a discrete time period of exposure divided by the concentration in the surrounding water at the same time or during the same period.

**Biomarkers**—Broadly defined as indicators signaling events in biologic systems or samples. They have been classified as markers of exposure, markers of effect, and markers of susceptibility.

**Cancer Effect Level (CEL)**—The lowest dose of chemical in a study, or group of studies, that produces significant increases in the incidence of cancer (or tumors) between the exposed population and its appropriate control.

**Carcinogen**—A chemical capable of inducing cancer.

**Case-Control Study**—A type of epidemiological study which examines the relationship between a particular outcome (disease or condition) and a variety of potential causative agents (such as toxic chemicals). In a case-controlled study, a group of people with a specified and well-defined outcome is identified and compared to a similar group of people without outcome.

**Case Report**—Describes a single individual with a particular disease or exposure. These may suggest some potential topics for scientific research but are not actual research studies.

## 10. GLOSSARY

**Case Series**—Describes the experience of a small number of individuals with the same disease or exposure. These may suggest potential topics for scientific research but are not actual research studies.

**Ceiling Value**—A concentration of a substance that should not be exceeded, even instantaneously.

**Chronic Exposure**—Exposure to a chemical for 365 days or more, as specified in the Toxicological Profiles.

**Cohort Study**—A type of epidemiological study of a specific group or groups of people who have had a common insult (e.g., exposure to an agent suspected of causing disease or a common disease) and are followed forward from exposure to outcome. At least one exposed group is compared to one unexposed group.

**Cross-sectional Study**—A type of epidemiological study of a group or groups which examines the relationship between exposure and outcome to a chemical or to chemicals at one point in time.

**Data Needs**—Substance-specific informational needs that if met would reduce the uncertainties of human health assessment.

**Developmental Toxicity**—The occurrence of adverse effects on the developing organism that may result from exposure to a chemical prior to conception (either parent), during prenatal development, or postnatally to the time of sexual maturation. Adverse developmental effects may be detected at any point in the life span of the organism.

**Dose-Response Relationship**—The quantitative relationship between the amount of exposure to a toxicant and the incidence of the adverse effects.

**Embryotoxicity and Fetotoxicity**—Any toxic effect on the conceptus as a result of prenatal exposure to a chemical; the distinguishing feature between the two terms is the stage of development during which the insult occurs. The terms, as used here, include malformations and variations, altered growth, and *in utero* death.

**Environmental Protection Agency (EPA) Health Advisory**—An estimate of acceptable drinking water levels for a chemical substance based on health effects information. A health advisory is not a legally enforceable federal standard, but serves as technical guidance to assist federal, state, and local officials.

**Epidemiology**—Refers to the investigation of factors that determine the frequency and distribution of disease or other health-related conditions within a defined human population during a specified period.

**Genotoxicity**—A specific adverse effect on the genome of living cells that, upon the duplication of affected cells, can be expressed as a mutagenic, clastogenic, or carcinogenic event because of specific alteration of the molecular structure of the genome.

**Half-life**—A measure of rate for the time required to eliminate one half of a quantity of a chemical from the body or environmental media.

**Immediately Dangerous to Life or Health (IDLH)**—The maximum environmental concentration of a contaminant from which one could escape within 30 minutes without any escape-impairing symptoms or irreversible health effects.

## 10. GLOSSARY

**Incidence**—The ratio of individuals in a population who develop a specified condition to the total number of individuals in that population who could have developed that condition in a specified time period.

**Intermediate Exposure**—Exposure to a chemical for a duration of 15–364 days, as specified in the Toxicological Profiles.

**Immunologic Toxicity**—The occurrence of adverse effects on the immune system that may result from exposure to environmental agents such as chemicals.

**Immunological Effects**—Functional changes in the immune response.

**In Vitro**—Isolated from the living organism and artificially maintained, as in a test tube.

**In Vivo**—Occurring within the living organism.

**Lethal Concentration<sub>(LO)</sub> (LC<sub>LO</sub>)**—The lowest concentration of a chemical in air which has been reported to have caused death in humans or animals.

**Lethal Concentration<sub>(50)</sub> (LC<sub>50</sub>)**—A calculated concentration of a chemical in air to which exposure for a specific length of time is expected to cause death in 50% of a defined experimental animal population.

**Lethal Dose<sub>(LO)</sub> (LD<sub>LO</sub>)**—The lowest dose of a chemical introduced by a route other than inhalation that has been reported to have caused death in humans or animals.

**Lethal Dose<sub>(50)</sub> (LD<sub>50</sub>)**—The dose of a chemical which has been calculated to cause death in 50% of a defined experimental animal population.

**Lethal Time<sub>(50)</sub> (LT<sub>50</sub>)**—A calculated period of time within which a specific concentration of a chemical is expected to cause death in 50% of a defined experimental animal population.

**Lowest-Observed-Adverse-Effect Level (LOAEL)**—The lowest exposure level of chemical in a study, or group of studies, that produces statistically or biologically significant increases in frequency or severity of adverse effects between the exposed population and its appropriate control.

**Lymphoreticular Effects**—Represent morphological effects involving lymphatic tissues such as the lymph nodes, spleen, and thymus.

**Malformations**—Permanent structural changes that may adversely affect survival, development, or function.

**Minimal Risk Level (MRL)**—An estimate of daily human exposure to a hazardous substance that is likely to be without an appreciable risk of adverse noncancer health effects over a specified route and duration of exposure.

**Modifying Factor (MF)**—A value (greater than zero) that is applied to the derivation of a minimal risk level (MRL) to reflect additional concerns about the database that are not covered by the uncertainty factors. The default value for a MF is 1.

**Morbidity**—State of being diseased; morbidity rate is the incidence or prevalence of disease in a specific population.

## 10. GLOSSARY

**Mortality**—Death; mortality rate is a measure of the number of deaths in a population during a specified interval of time.

**Mutagen**—A substance that causes mutations. A mutation is a change in the DNA sequence of a cell's DNA. Mutations can lead to birth defects, miscarriages, or cancer.

**Necropsy**—The gross examination of the organs and tissues of a dead body to determine the cause of death or pathological conditions.

**Neurotoxicity**—The occurrence of adverse effects on the nervous system following exposure to a chemical.

**No-Observed-Adverse-Effect Level (NOAEL)**—The dose of a chemical at which there were no statistically or biologically significant increases in frequency or severity of adverse effects seen between the exposed population and its appropriate control. Effects may be produced at this dose, but they are not considered to be adverse.

**Octanol-Water Partition Coefficient ( $K_{ow}$ )**—The equilibrium ratio of the concentrations of a chemical in *n*-octanol and water, in dilute solution.

**Odds Ratio (OR)**—A means of measuring the association between an exposure (such as toxic substances and a disease or condition) which represents the best estimate of relative risk (risk as a ratio of the incidence among subjects exposed to a particular risk factor divided by the incidence among subjects who were not exposed to the risk factor). An odds ratio of greater than 1 is considered to indicate greater risk of disease in the exposed group compared to the unexposed.

**Organophosphate or Organophosphorus Compound**—A phosphorus containing organic compound and especially a pesticide that acts by inhibiting cholinesterase.

**Permissible Exposure Limit (PEL)**—An Occupational Safety and Health Administration (OSHA) allowable exposure level in workplace air averaged over an 8-hour shift of a 40-hour workweek.

**Pesticide**—General classification of chemicals specifically developed and produced for use in the control of agricultural and public health pests.

**Pharmacokinetics**—The science of quantitatively predicting the fate (disposition) of an exogenous substance in an organism. Utilizing computational techniques, it provides the means of studying the absorption, distribution, metabolism and excretion of chemicals by the body.

**Pharmacokinetic Model**—A set of equations that can be used to describe the time course of a parent chemical or metabolite in an animal system. There are two types of pharmacokinetic models: data-based and physiologically-based. A data-based model divides the animal system into a series of compartments which, in general, do not represent real, identifiable anatomic regions of the body whereby the physiologically-based model compartments represent real anatomic regions of the body.

**Physiologically Based Pharmacodynamic (PBPD) Model**—A type of physiologically-based dose-response model which quantitatively describes the relationship between target tissue dose and toxic end points. These models advance the importance of physiologically based models in that they clearly describe the biological effect (response) produced by the system following exposure to an exogenous substance.

## 10. GLOSSARY

**Physiologically Based Pharmacokinetic (PBPK) Model**—Comprised of a series of compartments representing organs or tissue groups with realistic weights and blood flows. These models require a variety of physiological information: tissue volumes, blood flow rates to tissues, cardiac output, alveolar ventilation rates and, possibly membrane permeabilities. The models also utilize biochemical information such as air/blood partition coefficients, and metabolic parameters. PBPK models are also called biologically based tissue dosimetry models.

**Prevalence**—The number of cases of a disease or condition in a population at one point in time.

**Prospective Study**—A type of cohort study in which the pertinent observations are made on events occurring after the start of the study. A group is followed over time.

**$q_1^*$** —The upper-bound estimate of the low-dose slope of the dose-response curve as determined by the multistage procedure. The  $q_1^*$  can be used to calculate an estimate of carcinogenic potency, the incremental excess cancer risk per unit of exposure (usually  $\mu\text{g/L}$  for water,  $\text{mg/kg/day}$  for food, and  $\mu\text{g/m}^3$  for air).

**Recommended Exposure Limit (REL)**—A National Institute for Occupational Safety and Health (NIOSH) time-weighted average (TWA) concentrations for up to a 10-hour workday during a 40-hour workweek.

**Reference Concentration (RfC)**—An estimate (with uncertainty spanning perhaps an order of magnitude) of a continuous inhalation exposure to the human population (including sensitive subgroups) that is likely to be without an appreciable risk of deleterious noncancer health effects during a lifetime. The inhalation reference concentration is for continuous inhalation exposures and is appropriately expressed in units of  $\text{mg/m}^3$  or ppm.

**Reference Dose (RfD)**—An estimate (with uncertainty spanning perhaps an order of magnitude) of the daily exposure of the human population to a potential hazard that is likely to be without risk of deleterious effects during a lifetime. The RfD is operationally derived from the no-observed-adverse-effect level (NOAEL—from animal and human studies) by a consistent application of uncertainty factors that reflect various types of data used to estimate RfDs and an additional modifying factor, which is based on a professional judgment of the entire database on the chemical. The RfDs are not applicable to nonthreshold effects such as cancer.

**Reportable Quantity (RQ)**—The quantity of a hazardous substance that is considered reportable under the Comprehensive Environmental Response, Compensation, and Liability Act (CERCLA). Reportable quantities are (1) 1 pound or greater or (2) for selected substances, an amount established by regulation either under CERCLA or under Section 311 of the Clean Water Act. Quantities are measured over a 24-hour period.

**Reproductive Toxicity**—The occurrence of adverse effects on the reproductive system that may result from exposure to a chemical. The toxicity may be directed to the reproductive organs and/or the related endocrine system. The manifestation of such toxicity may be noted as alterations in sexual behavior, fertility, pregnancy outcomes, or modifications in other functions that are dependent on the integrity of this system.

## 10. GLOSSARY

**Retrospective Study**—A type of cohort study based on a group of persons known to have been exposed at some time in the past. Data are collected from routinely recorded events, up to the time the study is undertaken. Retrospective studies are limited to causal factors that can be ascertained from existing records and/or examining survivors of the cohort.

**Risk**—The possibility or chance that some adverse effect will result from a given exposure to a chemical.

**Risk Factor**—An aspect of personal behavior or lifestyle, an environmental exposure, or an inborn or inherited characteristic, that is associated with an increased occurrence of disease or other health-related event or condition.

**Risk Ratio**—The ratio of the risk among persons with specific risk factors compared to the risk among persons without risk factors. A risk ratio greater than 1 indicates greater risk of disease in the exposed group compared to the unexposed.

**Short-Term Exposure Limit (STEL)**—The American Conference of Governmental Industrial Hygienists (ACGIH) maximum concentration to which workers can be exposed for up to 15 minutes continually. No more than four excursions are allowed per day, and there must be at least 60 minutes between exposure periods. The daily Threshold Limit Value - Time Weighted Average (TLV-TWA) may not be exceeded.

**Standardized Mortality Ratio (SMR)**—A ratio of the observed number of deaths and the expected number of deaths in a specific standard population.

**Target Organ Toxicity**—This term covers a broad range of adverse effects on target organs or physiological systems (e.g., renal, cardiovascular) extending from those arising through a single limited exposure to those assumed over a lifetime of exposure to a chemical.

**Teratogen**—A chemical that causes structural defects that affect the development of an organism.

**Threshold Limit Value (TLV)**—An American Conference of Governmental Industrial Hygienists (ACGIH) concentration of a substance to which most workers can be exposed without adverse effect. The TLV may be expressed as a Time Weighted Average (TWA), as a Short-Term Exposure Limit (STEL), or as a ceiling limit (CL).

**Time-Weighted Average (TWA)**—An allowable exposure concentration averaged over a normal 8-hour workday or 40-hour workweek.

**Toxic Dose<sub>(50)</sub> (TD<sub>50</sub>)**—A calculated dose of a chemical, introduced by a route other than inhalation, which is expected to cause a specific toxic effect in 50% of a defined experimental animal population.

**Toxicokinetic**—The study of the absorption, distribution and elimination of toxic compounds in the living organism.

## 10. GLOSSARY

**Uncertainty Factor (UF)**—A factor used in operationally deriving the Minimal Risk Level (MRL) or Reference Dose (RfD) or Reference Concentration (RfC) from experimental data. UFs are intended to account for (1) the variation in sensitivity among the members of the human population, (2) the uncertainty in extrapolating animal data to the case of human, (3) the uncertainty in extrapolating from data obtained in a study that is of less than lifetime exposure, and (4) the uncertainty in using lowest-observed-adverse-effect level (LOAEL) data rather than no-observed-adverse-effect level (NOAEL) data. A default for each individual UF is 10; if complete certainty in data exists, a value of one can be used; however a reduced UF of three may be used on a case-by-case basis, three being the approximate logarithmic average of 10 and 1.

**Xenobiotic**—Any chemical that is foreign to the biological system.



## **APPENDIX A. ATSDR MINIMAL RISK LEVELS AND WORKSHEETS**

The Comprehensive Environmental Response, Compensation, and Liability Act (CERCLA) [42 U.S.C. 9601 et seq.], as amended by the Superfund Amendments and Reauthorization Act (SARA) [Pub. L. 99–499], requires that the Agency for Toxic Substances and Disease Registry (ATSDR) develop jointly with the U.S. Environmental Protection Agency (EPA), in order of priority, a list of hazardous substances most commonly found at facilities on the CERCLA National Priorities List (NPL); prepare toxicological profiles for each substance included on the priority list of hazardous substances; and assure the initiation of a research program to fill identified data needs associated with the substances.

The toxicological profiles include an examination, summary, and interpretation of available toxicological information and epidemiologic evaluations of a hazardous substance. During the development of toxicological profiles, Minimal Risk Levels (MRLs) are derived when reliable and sufficient data exist to identify the target organ(s) of effect or the most sensitive health effect(s) for a specific duration for a given route of exposure. An MRL is an estimate of the daily human exposure to a hazardous substance that is likely to be without appreciable risk of adverse noncancer health effects over a specified duration of exposure. MRLs are based on noncancer health effects only and are not based on a consideration of cancer effects. These substance-specific estimates, which are intended to serve as screening levels, are used by ATSDR health assessors to identify contaminants and potential health effects that may be of concern at hazardous waste sites. It is important to note that MRLs are not intended to define clean-up or action levels.

MRLs are derived for hazardous substances using the no-observed-adverse-effect level/uncertainty factor approach. They are below levels that might cause adverse health effects in the people most sensitive to such chemical-induced effects. MRLs are derived for acute (1–14 days), intermediate (15–364 days), and chronic (365 days and longer) durations and for the oral and inhalation routes of exposure. Currently, MRLs for the dermal route of exposure are not derived because ATSDR has not yet identified a method suitable for this route of exposure. MRLs are generally based on the most sensitive chemical-induced end point considered to be of relevance to humans. Serious health effects (such as irreparable damage to the liver or kidneys, or birth defects) are not used as a basis for establishing MRLs. Exposure to a level above the MRL does not mean that adverse health effects will occur.

MRLs are intended only to serve as a screening tool to help public health professionals decide where to look more closely. They may also be viewed as a mechanism to identify those hazardous waste sites that

## APPENDIX A

are not expected to cause adverse health effects. Most MRLs contain a degree of uncertainty because of the lack of precise toxicological information on the people who might be most sensitive (e.g., infants, elderly, nutritionally or immunologically compromised) to the effects of hazardous substances. ATSDR uses a conservative (i.e., protective) approach to address this uncertainty consistent with the public health principle of prevention. Although human data are preferred, MRLs often must be based on animal studies because relevant human studies are lacking. In the absence of evidence to the contrary, ATSDR assumes that humans are more sensitive to the effects of hazardous substance than animals and that certain persons may be particularly sensitive. Thus, the resulting MRL may be as much as a hundredfold below levels that have been shown to be nontoxic in laboratory animals.

Proposed MRLs undergo a rigorous review process: Health Effects/MRL Workgroup reviews within the Division of Toxicology, expert panel peer reviews, and agency wide MRL Workgroup reviews, with participation from other federal agencies and comments from the public. They are subject to change as new information becomes available concomitant with updating the toxicological profiles. Thus, MRLs in the most recent toxicological profiles supersede previously published levels. For additional information regarding MRLs, please contact the Division of Toxicology, Agency for Toxic Substances and Disease Registry, 1600 Clifton Road NE, Mailstop E-29, Atlanta, Georgia 30333.

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**MINIMAL RISK LEVEL (MRL) WORKSHEET**

Chemical name: Zinc and compounds  
 CAS number:  
 Date: August 29, 2003  
 Profile status: Draft 3 Pre-Public  
 Route:  Inhalation  Oral  
 Duration:  Acute  Intermediate  Chronic  
 Key to figure: 25  
 Species: Human  
 MRL: 0.3  mg/kg/day  ppm  mg/m<sup>3</sup>

Reference: Yadrick MK, Kenney MA, Winterfelt EA. 1989. Iron, copper, and zinc status: Response to supplementation with zinc or zinc and iron in adult females. Am J Clin Nutr 49:145-150.

Experimental design: Eighteen healthy women, ages 25–40 years, were given zinc gluconate supplements twice daily (50 mg supplemental zinc/day, or 0.83 mg supplemental zinc/kg/day assuming a 60-kg mean body weight for healthy women) for a 10-week period (Yadrick et al. 1989). Blood was drawn from each subject prior to treatment for use as a referent. Erythrocyte superoxide dismutase (ESOD) activity declined over the 10-week supplementation period and, at 10 weeks, was significantly ( $p < 0.05$ ) lower (47% decrease) than pretreatment values; a decrease at 6 weeks of exposure (15%) was not statistically significant. ESOD levels are considered to be a sensitive indicator of systemic copper status. Ceruloplasmin levels were not altered. Serum zinc was significantly increased at both 6 and 10 weeks. There was also a significant decline in serum ferritin (23%) and hematocrit (4%) values at 10 weeks. In women similarly-exposed but also receiving 0.42 mg supplemental iron/day, serum ferritin was increased at both 6 and 10 weeks of exposure, while serum ESOD levels were significantly decreased at 6 weeks (24%) and 10 weeks (47%).

Effects noted in study and corresponding doses: A significant decrease in erythrocyte superoxide dismutase levels was seen at 0.83 mg Zn/kg/day at both time points evaluated.

Dose end point used for MRL derivation: Yadrick et al. (1989) reported a significant decrease in ESOD, which is considered to be a sensitive indicator of body copper status, in women exposed to 50 mg supplemental zinc/day. Because the observed effect is considered to be a precursor event to the more severe symptoms seen with zinc-induced copper deficiency, rather than a toxic effect of itself, the 50 mg supplemental zinc/day value is considered to be a minimal LOAEL. Assuming a reference female body weight of 60 kg, this represents 0.83 mg Zn/kg/day.

NOAEL  LOAEL

Uncertainty factors used in MRL derivation:

1  3  10 (for use of a LOAEL)  
 1  3  10 (for extrapolation from animals to humans)  
 1  3  10 (for human variability)

\* A composite uncertainty factor of 3 was used to represent the use of a minimal LOAEL and the possible existence of susceptible subpopulations.

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The intermediate oral MRL for zinc is derived as follows:

$$\text{MRL} = \text{NOAEL} \div \text{UF}$$

$$\text{MRL} = 0.83 \text{ mg Zn/kg/day} \div 3$$

$$\text{MRL} = 0.3 \text{ mg Zn/kg/day}$$

Was a conversion factor used from ppm in food or water to a mg/body weight dose? No.

Was a conversion used from intermittent to continuous exposure? If so, explain: No.

If an inhalation study in animals, list conversion factors used in determining human equivalent dose: Not Applicable.

Other additional studies or pertinent information that lend support to this MRL:

Fischer et al. (1984) gave groups of 13 healthy adult male volunteers (ages not specified) 0 (cornstarch) or 25 mg supplemental zinc (as zinc gluconate) twice daily for 6 weeks (0 or 0.71 mg supplemental zinc/kg/day). Nonfasting blood samples were taken at the beginning and at biweekly intervals and tested for measures of copper status. ESOD activity decreased after 4 weeks in the supplement group and was significantly lower than controls by 6 weeks. An inverse correlation between plasma zinc levels and erythrocyte superoxide dismutase activity was also observed at 6 weeks. Plasma copper levels and levels of ferroxidase activity did not change during the course of the study.

Groups of 9, 13, or 9 healthy white men were administered 0, 50, or 75 mg/day supplemental zinc as zinc gluconate (total zinc intakes were 0.16, 0.85, and 1.10 mg supplemental zinc/kg/day, respectively, based on mean group body weights) for 12 weeks (Black et al. 1988). The subjects were given instructions to avoid foods high in calcium, fiber, and phytic acid, dietary constituents that are known to decrease zinc absorption. Subjects were also told to restrict their intake of zinc-rich foods in order to minimize the variation in daily dietary zinc. There was a general decline in the mean serum HDL-cholesterol for the 75-mg supplement group between weeks 6 and 12. HDL values for this group were significantly lower than those for the placebo group at weeks 6 and 12 ( $p < 0.05$ ). There was also a decline in the HDL values for the 50-mg group between weeks 8 through 12; however, this decline was not significantly different from that for the controls until the 12th week of treatment. Serum zinc, copper, total cholesterol, LDL-cholesterol, and triglycerides did not appear to be affected by treatment.

Freeland-Graves et al. (1982) exposed groups of eight healthy women to 0, 15, 50, or 100 mg supplemental zinc as zinc acetate (approximately 0, 0.25, 0.83, or 1.7 mg supplemental zinc/kg/day, assuming a reference female body weight of 60 kg) daily for 60 days. In the highest exposure group only, plasma HDL-cholesterol was significantly reduced at 4 weeks of exposure, but not at any other timepoint examined. A correlation between dietary zinc and whole-blood copper was observed in treated subjects. In the 50 and 100 mg groups, some bloating, nausea, and abdominal cramps were reported unless the supplement was taken with a large glass of water at mealtime.

Prasad et al. (1978) fed a patient with sickle cell anemia supplements of 150–200 mg supplemental zinc/day for 2 years. The supplement resulted in copper deficiency; serum copper and plasma ceruloplasmin levels were decreased. When copper was administered, the plasma ceruloplasmin levels became normal. In a follow-up study, of 13 patients on zinc therapy (similar treatment levels assumed), 7 patients had ceruloplasmin levels at the lower limit of normal after 24 weeks of dosing.

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## APPENDIX B. USER'S GUIDE

### Chapter 1

#### Public Health Statement

This chapter of the profile is a health effects summary written in non-technical language. Its intended audience is the general public especially people living in the vicinity of a hazardous waste site or chemical release. If the Public Health Statement were removed from the rest of the document, it would still communicate to the lay public essential information about the chemical.

The major headings in the Public Health Statement are useful to find specific topics of concern. The topics are written in a question and answer format. The answer to each question includes a sentence that will direct the reader to chapters in the profile that will provide more information on the given topic.

### Chapter 2

#### Relevance to Public Health

This chapter provides a health effects summary based on evaluations of existing toxicologic, epidemiologic, and toxicokinetic information. This summary is designed to present interpretive, weight-of-evidence discussions for human health end points by addressing the following questions.

1. What effects are known to occur in humans?
2. What effects observed in animals are likely to be of concern to humans?
3. What exposure conditions are likely to be of concern to humans, especially around hazardous waste sites?

The chapter covers end points in the same order they appear within the Discussion of Health Effects by Route of Exposure section, by route (inhalation, oral, dermal) and within route by effect. Human data are presented first, then animal data. Both are organized by duration (acute, intermediate, chronic). *In vitro* data and data from parenteral routes (intramuscular, intravenous, subcutaneous, etc.) are also considered in this chapter. If data are located in the scientific literature, a table of genotoxicity information is included.

The carcinogenic potential of the profiled substance is qualitatively evaluated, when appropriate, using existing toxicokinetic, genotoxic, and carcinogenic data. ATSDR does not currently assess cancer potency or perform cancer risk assessments. Minimal risk levels (MRLs) for noncancer end points (if derived) and the end points from which they were derived are indicated and discussed.

Limitations to existing scientific literature that prevent a satisfactory evaluation of the relevance to public health are identified in the Chapter 3 Data Needs section.

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**Interpretation of Minimal Risk Levels**

Where sufficient toxicologic information is available, we have derived minimal risk levels (MRLs) for inhalation and oral routes of entry at each duration of exposure (acute, intermediate, and chronic). These MRLs are not meant to support regulatory action; but to acquaint health professionals with exposure levels at which adverse health effects are not expected to occur in humans.

They should help physicians and public health officials determine the safety of a community living near a chemical emission, given the concentration of a contaminant in air or the estimated daily dose in water. MRLs are based largely on toxicological studies in animals and on reports of human occupational exposure.

MRL users should be familiar with the toxicologic information on which the number is based. Chapter 2, "Relevance to Public Health," contains basic information known about the substance. Other sections such as Chapter 3 Section 3.9, "Interactions with Other Substances," and Section 3.10, "Populations that are Unusually Susceptible" provide important supplemental information.

MRL users should also understand the MRL derivation methodology. MRLs are derived using a modified version of the risk assessment methodology the Environmental Protection Agency (EPA) provides (Barnes and Dourson 1988) to determine reference doses for lifetime exposure (RfDs).

To derive an MRL, ATSDR generally selects the most sensitive end point which, in its best judgement, represents the most sensitive human health effect for a given exposure route and duration. ATSDR cannot make this judgement or derive an MRL unless information (quantitative or qualitative) is available for all potential systemic, neurological, and developmental effects. If this information and reliable quantitative data on the chosen end point are available, ATSDR derives an MRL using the most sensitive species (when information from multiple species is available) with the highest NOAEL that does not exceed any adverse effect levels. When a NOAEL is not available, a lowest-observed-adverse-effect level (LOAEL) can be used to derive an MRL, and an uncertainty factor (UF) of 10 must be employed. Additional uncertainty factors of 10 must be used both for human variability to protect sensitive subpopulations (people who are most susceptible to the health effects caused by the substance) and for interspecies variability (extrapolation from animals to humans). In deriving an MRL, these individual uncertainty factors are multiplied together. The product is then divided into the inhalation concentration or oral dosage selected from the study. Uncertainty factors used in developing a substance-specific MRL are provided in the footnotes of the LSE Tables.

**Chapter 3****Health Effects****Tables and Figures for Levels of Significant Exposure (LSE)**

Tables (3-1, 3-2, and 3-3) and figures (3-1 and 3-2) are used to summarize health effects and illustrate graphically levels of exposure associated with those effects. These levels cover health effects observed at increasing dose concentrations and durations, differences in response by species, minimal risk levels (MRLs) to humans for noncancer end points, and EPA's estimated range associated with an upper-bound individual lifetime cancer risk of 1 in 10,000 to 1 in 10,000,000. Use the LSE tables and figures for a quick review of the health effects and to locate data for a specific exposure scenario. The LSE tables and figures should always be used in conjunction with the text. All entries in these tables and figures represent studies that provide reliable, quantitative estimates of No-Observed-Adverse-Effect Levels (NOAELs), Lowest-Observed-Adverse-Effect Levels (LOAELs), or Cancer Effect Levels (CELs).

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The legends presented below demonstrate the application of these tables and figures. Representative examples of LSE Table 3-1 and Figure 3-1 are shown. The numbers in the left column of the legends correspond to the numbers in the example table and figure.

**LEGEND****See LSE Table 3-1**

- (1) **Route of Exposure** One of the first considerations when reviewing the toxicity of a substance using these tables and figures should be the relevant and appropriate route of exposure. When sufficient data exists, three LSE tables and two LSE figures are presented in the document. The three LSE tables present data on the three principal routes of exposure, i.e., inhalation, oral, and dermal (LSE Table 3-1, 3-2, and 3-3, respectively). LSE figures are limited to the inhalation (LSE Figure 3-1) and oral (LSE Figure 3-2) routes. Not all substances will have data on each route of exposure and will not therefore have all five of the tables and figures.
- (2) **Exposure Period** Three exposure periods - acute (less than 15 days), intermediate (15–364 days), and chronic (365 days or more) are presented within each relevant route of exposure. In this example, an inhalation study of intermediate exposure duration is reported. For quick reference to health effects occurring from a known length of exposure, locate the applicable exposure period within the LSE table and figure.
- (3) **Health Effect** The major categories of health effects included in LSE tables and figures are death, systemic, immunological, neurological, developmental, reproductive, and cancer. NOAELs and LOAELs can be reported in the tables and figures for all effects but cancer. Systemic effects are further defined in the "System" column of the LSE table (see key number 18).
- (4) **Key to Figure** Each key number in the LSE table links study information to one or more data points using the same key number in the corresponding LSE figure. In this example, the study represented by key number 18 has been used to derive a NOAEL and a Less Serious LOAEL (also see the 2 "18r" data points in Figure 3-1).
- (5) **Species** The test species, whether animal or human, are identified in this column. Chapter 2, "Relevance to Public Health," covers the relevance of animal data to human toxicity and Section 3.4, "Toxicokinetics," contains any available information on comparative toxicokinetics. Although NOAELs and LOAELs are species specific, the levels are extrapolated to equivalent human doses to derive an MRL.
- (6) **Exposure Frequency/Duration** The duration of the study and the weekly and daily exposure regimen are provided in this column. This permits comparison of NOAELs and LOAELs from different studies. In this case (key number 18), rats were exposed to 1,1,2,2-tetrachloroethane via inhalation for 6 hours per day, 5 days per week, for 3 weeks. For a more complete review of the dosing regimen refer to the appropriate sections of the text or the original reference paper, i.e., Nitschke et al. 1981.
- (7) **System** This column further defines the systemic effects. These systems include: respiratory, cardiovascular, gastrointestinal, hematological, musculoskeletal, hepatic, renal, and dermal/ocular. "Other" refers to any systemic effect (e.g., a decrease in body weight) not covered in these systems. In the example of key number 18, 1 systemic effect (respiratory) was investigated.

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- (8) NOAEL A No-Observed-Adverse-Effect Level (NOAEL) is the highest exposure level at which no harmful effects were seen in the organ system studied. Key number 18 reports a NOAEL of 3 ppm for the respiratory system which was used to derive an intermediate exposure, inhalation MRL of 0.005 ppm (see footnote "b").
- (9) LOAEL A Lowest-Observed-Adverse-Effect Level (LOAEL) is the lowest dose used in the study that caused a harmful health effect. LOAELs have been classified into "Less Serious" and "Serious" effects. These distinctions help readers identify the levels of exposure at which adverse health effects first appear and the gradation of effects with increasing dose. A brief description of the specific end point used to quantify the adverse effect accompanies the LOAEL. The respiratory effect reported in key number 18 (hyperplasia) is a Less serious LOAEL of 10 ppm. MRLs are not derived from Serious LOAELs.
- (10) Reference The complete reference citation is given in Chapter 9 of the profile.
- (11) CEL A Cancer Effect Level (CEL) is the lowest exposure level associated with the onset of carcinogenesis in experimental or epidemiologic studies. CELs are always considered serious effects. The LSE tables and figures do not contain NOAELs for cancer, but the text may report doses not causing measurable cancer increases.
- (12) Footnotes Explanations of abbreviations or reference notes for data in the LSE tables are found in the footnotes. Footnote "b" indicates the NOAEL of 3 ppm in key number 18 was used to derive an MRL of 0.005 ppm.

**LEGEND****See Figure 3-1**

LSE figures graphically illustrate the data presented in the corresponding LSE tables. Figures help the reader quickly compare health effects according to exposure concentrations for particular exposure periods.

- (13) Exposure Period The same exposure periods appear as in the LSE table. In this example, health effects observed within the intermediate and chronic exposure periods are illustrated.
- (14) Health Effect These are the categories of health effects for which reliable quantitative data exists. The same health effects appear in the LSE table.
- (15) Levels of Exposure concentrations or doses for each health effect in the LSE tables are graphically displayed in the LSE figures. Exposure concentration or dose is measured on the log scale "y" axis. Inhalation exposure is reported in mg/m<sup>3</sup> or ppm and oral exposure is reported in mg/kg/day.
- (16) NOAEL In this example, the open circle designated 18r identifies a NOAEL critical end point in the rat upon which an intermediate inhalation exposure MRL is based. The key number 18 corresponds to the entry in the LSE table. The dashed descending arrow indicates the extrapolation from the exposure level of 3 ppm (see entry 18 in the Table) to the MRL of 0.005 ppm (see footnote "b" in the LSE table).



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- (17) CEL Key number 38r is 1 of 3 studies for which Cancer Effect Levels were derived. The diamond symbol refers to a Cancer Effect Level for the test species-mouse. The number 38 corresponds to the entry in the LSE table.
- (18) Estimated Upper-Bound Human Cancer Risk Levels This is the range associated with the upper-bound for lifetime cancer risk of 1 in 10,000 to 1 in 10,000,000. These risk levels are derived from the EPA's Human Health Assessment Group's upper-bound estimates of the slope of the cancer dose response curve at low dose levels ( $q_1^*$ ).
- (19) Key to LSE Figure The Key explains the abbreviations and symbols used in the figure.

**SAMPLE**

1 →

**TABLE 3-1. Levels of Significant Exposure to [Chemical x] - Inhalation**

Key to figure <sup>a</sup>	Species	Exposure frequency/ duration	System	NOAEL (ppm)	LOAEL (effect)		Reference
					Less serious (ppm)	Serious (ppm)	
<b>INTERMEDIATE EXPOSURE</b>							
	5	6	7	8	9		10
3 →	Systemic	↓	↓	↓	↓	↓	↓
4 →	18	Rat	13 wk 5 d/wk 6 hr/d	Resp	3 <sup>b</sup>	10 (hyperplasia)	Nitschke et al. 1981
<hr style="border-top: 1px dashed black;"/>							
<b>CHRONIC EXPOSURE</b>							
<b>Cancer</b>							
					11		
					↓		
	38	Rat	18 mo 5 d/wk 7 hr/d			20 (CEL, multiple organs)	Wong et al. 1982
	39	Rat	89-104 wk 5 d/wk 6 hr/d			10 (CEL, lung tumors, nasal tumors)	NTP 1982
	40	Mouse	79-103 wk 5 d/wk 6 hr/d			10 (CEL, lung tumors, hemangiosarcomas)	NTP 1982

12 →

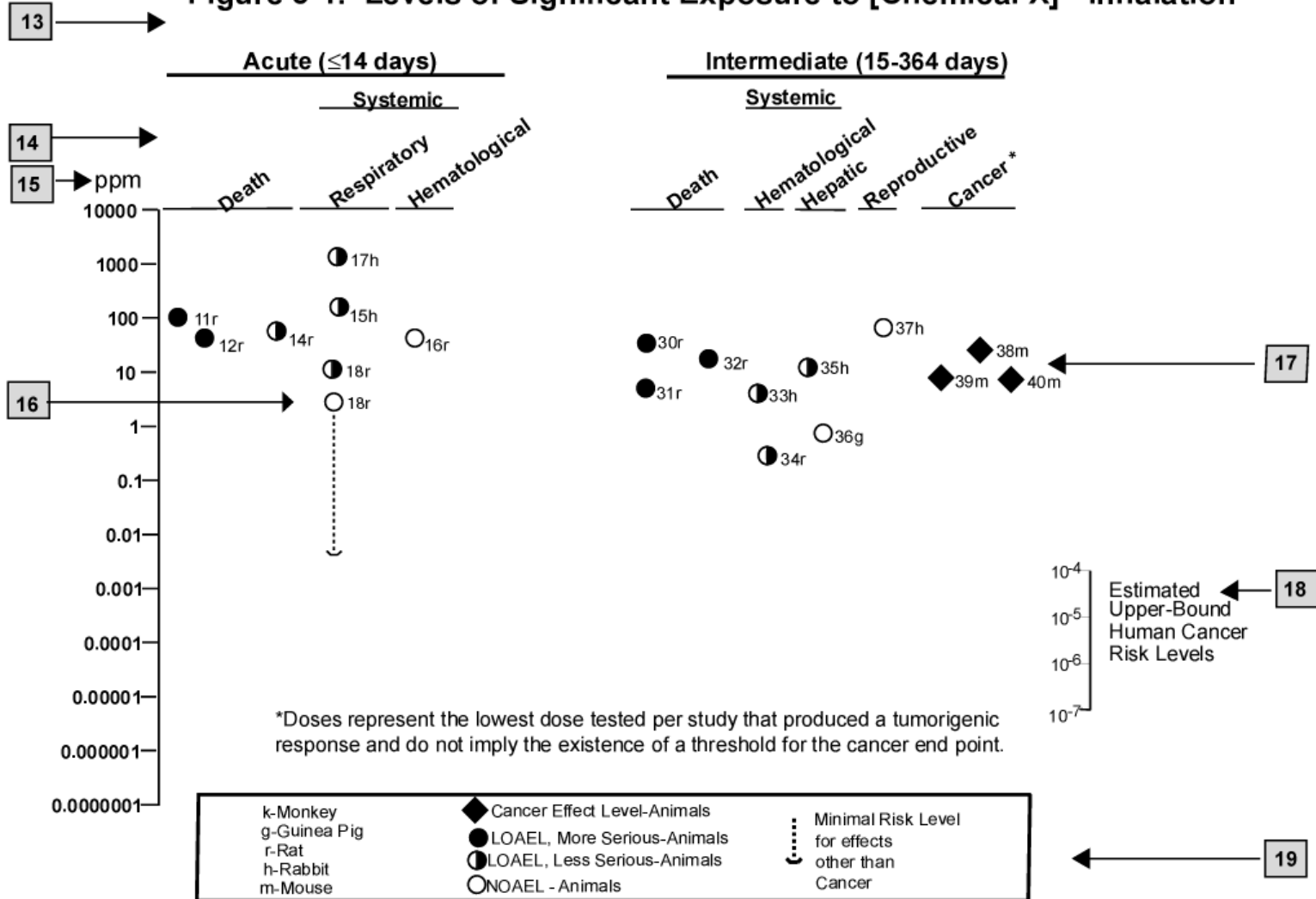
<sup>a</sup> The number corresponds to entries in Figure 3-1.  
<sup>b</sup> Used to derive an intermediate inhalation Minimal Risk Level (MRL) of  $5 \times 10^{-3}$  ppm; dose adjusted for intermittent exposure and divided by an uncertainty factor of 100 (10 for extrapolation from animal to humans, 10 for human variability).

\*\*\*DRAFT FOR PUBLIC COMMENT\*\*\*

APPENDIX B

# SAMPLE

### Figure 3-1. Levels of Significant Exposure to [Chemical X] - Inhalation



\*\*\*DRAFT FOR PUBLIC COMMENT\*\*\*

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## APPENDIX C. ACRONYMS, ABBREVIATIONS, AND SYMBOLS

ACOEM	American College of Occupational and Environmental Medicine
ACGIH	American Conference of Governmental Industrial Hygienists
ADI	acceptable daily intake
ADME	absorption, distribution, metabolism, and excretion
AED	atomic emission detection
AOEC	Association of Occupational and Environmental Clinics
AFID	alkali flame ionization detector
AFOSH	Air Force Office of Safety and Health
ALT	alanine aminotransferase
AML	acute myeloid leukemia
AOAC	Association of Official Analytical Chemists
AP	alkaline phosphatase
APHA	American Public Health Association
AST	aspartate aminotransferase
atm	atmosphere
ATSDR	Agency for Toxic Substances and Disease Registry
AWQC	Ambient Water Quality Criteria
BAT	best available technology
BCF	bioconcentration factor
BEI	Biological Exposure Index
BSC	Board of Scientific Counselors
C	centigrade
CAA	Clean Air Act
CAG	Cancer Assessment Group of the U.S. Environmental Protection Agency
CAS	Chemical Abstract Services
CDC	Centers for Disease Control and Prevention
CEL	cancer effect level
CELDS	Computer-Environmental Legislative Data System
CERCLA	Comprehensive Environmental Response, Compensation, and Liability Act
CFR	Code of Federal Regulations
Ci	curie
CI	confidence interval
CL	ceiling limit value
CLP	Contract Laboratory Program
cm	centimeter
CML	chronic myeloid leukemia
CPSC	Consumer Products Safety Commission
CWA	Clean Water Act
DHEW	Department of Health, Education, and Welfare
DHHS	Department of Health and Human Services
DNA	deoxyribonucleic acid
DOD	Department of Defense
DOE	Department of Energy
DOL	Department of Labor
DOT	Department of Transportation
DOT/UN/ NA/IMCO	Department of Transportation/United Nations/ North America/International Maritime Dangerous Goods Code
DWEL	drinking water exposure level

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ECD	electron capture detection
ECG/EKG	electrocardiogram
EEG	electroencephalogram
EEGL	Emergency Exposure Guidance Level
EPA	Environmental Protection Agency
F	Fahrenheit
F <sub>1</sub>	first-filial generation
FAO	Food and Agricultural Organization of the United Nations
FDA	Food and Drug Administration
FEMA	Federal Emergency Management Agency
FIFRA	Federal Insecticide, Fungicide, and Rodenticide Act
FPD	flame photometric detection
fpm	feet per minute
FR	<i>Federal Register</i>
FSH	follicle stimulating hormone
g	gram
GC	gas chromatography
gd	gestational day
GLC	gas liquid chromatography
GPC	gel permeation chromatography
HPLC	high-performance liquid chromatography
HRGC	high resolution gas chromatography
HSDB	Hazardous Substance Data Bank
IARC	International Agency for Research on Cancer
IDLH	immediately dangerous to life and health
ILO	International Labor Organization
IRIS	Integrated Risk Information System
K <sub>d</sub>	adsorption ratio
kg	kilogram
K <sub>oc</sub>	organic carbon partition coefficient
K <sub>ow</sub>	octanol-water partition coefficient
L	liter
LC	liquid chromatography
LC <sub>Lo</sub>	lethal concentration, low
LC <sub>50</sub>	lethal concentration, 50% kill
LD <sub>Lo</sub>	lethal dose, low
LD <sub>50</sub>	lethal dose, 50% kill
LDH	lactic dehydrogenase
LH	luteinizing hormone
LT <sub>50</sub>	lethal time, 50% kill
LOAEL	lowest-observed-adverse-effect level
LSE	Levels of Significant Exposure
m	meter
MA	<i>trans,trans</i> -muconic acid
MAL	maximum allowable level
mCi	millicurie
MCL	maximum contaminant level
MCLG	maximum contaminant level goal
MFO	mixed function oxidase
mg	milligram
mL	milliliter

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mm	millimeter
mmHg	millimeters of mercury
mmol	millimole
mppcf	millions of particles per cubic foot
MRL	Minimal Risk Level
MS	mass spectrometry
NAAQS	National Ambient Air Quality Standard
NAS	National Academy of Science
NATICH	National Air Toxics Information Clearinghouse
NATO	North Atlantic Treaty Organization
NCE	normochromatic erythrocytes
NCEH	National Center for Environmental Health
NCI	National Cancer Institute
ND	not detected
NFPA	National Fire Protection Association
ng	nanogram
NIEHS	National Institute of Environmental Health Sciences
NIOSH	National Institute for Occupational Safety and Health
NIOSHTIC	NIOSH's Computerized Information Retrieval System
NLM	National Library of Medicine
nm	nanometer
NHANES	National Health and Nutrition Examination Survey
nmol	nanomole
NOAEL	no-observed-adverse-effect level
NOES	National Occupational Exposure Survey
NOHS	National Occupational Hazard Survey
NPD	nitrogen phosphorus detection
NPDES	National Pollutant Discharge Elimination System
NPL	National Priorities List
NR	not reported
NRC	National Research Council
NS	not specified
NSPS	New Source Performance Standards
NTIS	National Technical Information Service
NTP	National Toxicology Program
ODW	Office of Drinking Water, EPA
OERR	Office of Emergency and Remedial Response, EPA
OHM/TADS	Oil and Hazardous Materials/Technical Assistance Data System
OPP	Office of Pesticide Programs, EPA
OPPTS	Office of Prevention, Pesticides and Toxic Substances, EPA
OPPT	Office of Pollution Prevention and Toxics, EPA
OR	odds ratio
OSHA	Occupational Safety and Health Administration
OSW	Office of Solid Waste, EPA
OW	Office of Water
OWRS	Office of Water Regulations and Standards, EPA
PAH	polycyclic aromatic hydrocarbon
PBPD	physiologically based pharmacodynamic
PBPK	physiologically based pharmacokinetic
PCE	polychromatic erythrocytes
PEL	permissible exposure limit

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pg	picogram
PHS	Public Health Service
PID	photo ionization detector
pmol	picomole
PMR	proportionate mortality ratio
ppb	parts per billion
ppm	parts per million
ppt	parts per trillion
PSNS	pretreatment standards for new sources
RBC	red blood cell
REL	recommended exposure level/limit
RfC	reference concentration
RfD	reference dose
RNA	ribonucleic acid
RTECS	Registry of Toxic Effects of Chemical Substances
RQ	reportable quantity
SARA	Superfund Amendments and Reauthorization Act
SCE	sister chromatid exchange
SGOT	serum glutamic oxaloacetic transaminase
SGPT	serum glutamic pyruvic transaminase
SIC	standard industrial classification
SIM	selected ion monitoring
SMCL	secondary maximum contaminant level
SMR	standardized mortality ratio
SNARL	suggested no adverse response level
SPEGL	Short-Term Public Emergency Guidance Level
STEL	short term exposure limit
STORET	Storage and Retrieval
TD <sub>50</sub>	toxic dose, 50% specific toxic effect
TLV	threshold limit value
TOC	total organic carbon
TPQ	threshold planning quantity
TRI	Toxics Release Inventory
TSCA	Toxic Substances Control Act
TWA	time-weighted average
UF	uncertainty factor
U.S.	United States
USDA	United States Department of Agriculture
USGS	United States Geological Survey
VOC	volatile organic compound
WBC	white blood cell
WHO	World Health Organization

## APPENDIX C

>	greater than
≥	greater than or equal to
=	equal to
<	less than
≤	less than or equal to
%	percent
α	alpha
β	beta
γ	gamma
δ	delta
μm	micrometer
μg	microgram
q <sub>1</sub> *	cancer slope factor
-	negative
+	positive
(+)	weakly positive result
(-)	weakly negative result