



DEPARTMENT OF HEALTH AND HUMAN SERVICES

Understanding Antimicrobial Resistance in Your Facility: the Patient Report and the Cumulative Antibiogram!

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**currently working with CDC's Division of Laboratory Systems through an Interagency Personnel Agreement*

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At the conclusion of this talk, you will be able to.....

- ◆ Discuss use of NCCLS standards for antimicrobial susceptibility testing (AST) and reporting in clinical and public health laboratories.
- ◆ Describe effective reporting of antimicrobial resistance on routine laboratory reports.



At the conclusion of this talk, you will be able to.....(con't)

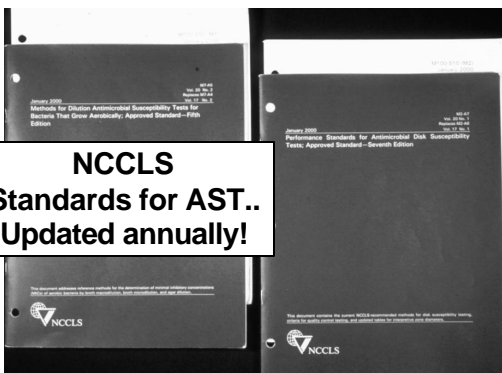
- ◆ Discuss analysis and presentation of cumulative antimicrobial susceptibility test data (antibiogram) at local healthcare facilities.



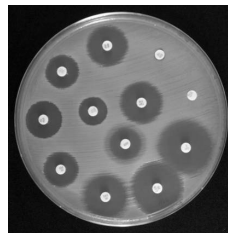
Current AST Methods NCCLS

NCCLS

NCCLS Standards for AST.. Updated annually!

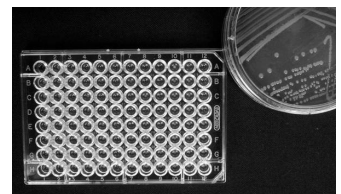



NCCLS Methods



Disk Diffusion (Kirby Bauer)

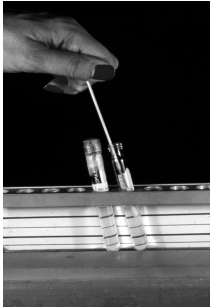
MIC






Select colonies

Prepare inoculum suspension





Disk Diffusion Testing





Swab plate

Remove sample

Add disks

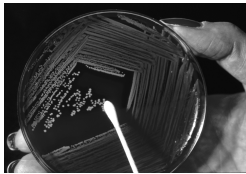
Incubate overnight

Measure zones

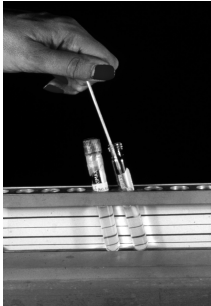
**NCCLS
Zone Interpretive Criteria (mm)**

Drug	Disk content (ug)	Res	Int	Susc
cefazolin	30	£ 14	15-17	≥ 18
gentamicin	10	£ 12	13-14	≥ 15

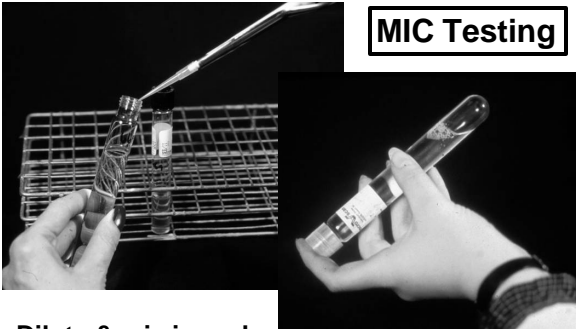


Select colonies

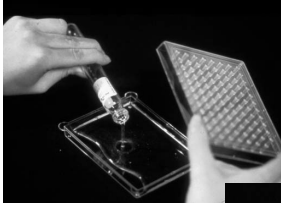
Prepare inoculum suspension



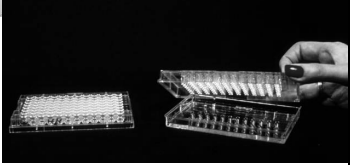
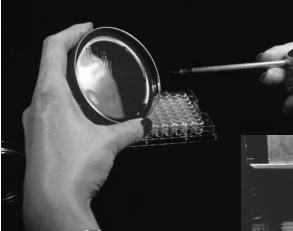
MIC Testing



Dilute & mix inoculum suspension




Pour inoculum into reservoir and inoculate MIC tray

Inoculate purity plate

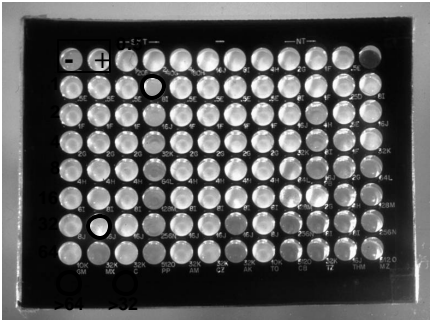
Incubate overnight



Read MICs



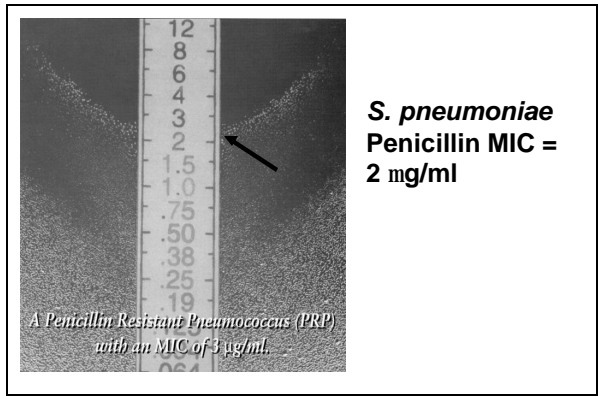
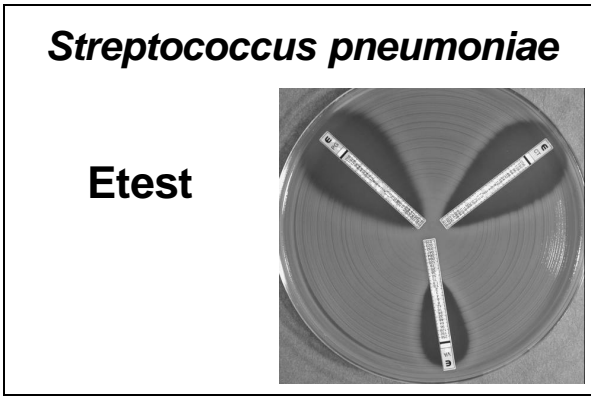
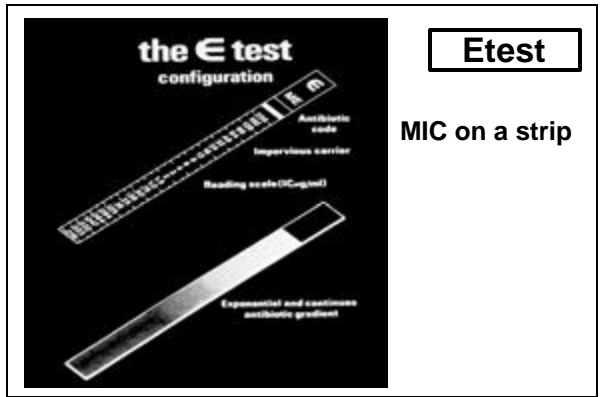
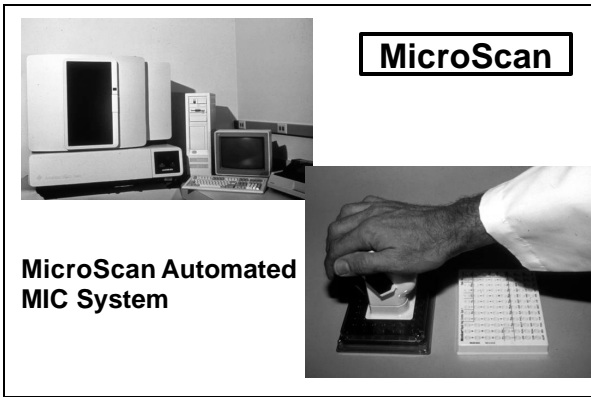
MICs



Vitek



Vitek Automated MIC System



NCCLS
MIC Interpretive Criteria (mg/ml)

Drug	Susc	Int	Res
cefazolin	£ 8	16	≥ 32
gentamicin	£ 4	8	≥ 16

NCCLS Table 1 NCCLS Vol. 22 No. 1

Table 1. Suggested Groupings of U.S. FDA-Approved Antimicrobial Agents That Should Be Considered for Routine Testing and Reporting on Nonfastidious Organisms by Clinical Microbiology Laboratories

GROUP A PRIMARY TEST AND REPORT	Enterobacteriaceae ^a	<i>Pseudomonas aeruginosa</i> and <i>Acinetobacter</i> spp. ¹	<i>Staphylococcus</i> spp.	<i>Enterococcus</i> spp. ^m
GROUP B ^b SELECTIVE TEST REPORT SELECTIVELY	Ampicillin ⁿ	Ceftazidime	Oxacillin ^k	Penicillin ^h or ampicillin
	Cefazolin ⁿ	Gentamicin	Penicillin ^h	
	Cephalexin ⁿ	Meropenem or ticarcillin		
	Gentamicin	Piperacillin		
GROUP B ^b SELECTIVE TEST REPORT SELECTIVELY	Amikacin	Amikacin	Azithromycin ^b or clarithromycin ^c or erythromycin ^d	Linezolid
	Amoxicillin-clavulanic acid or ampicillin-sulbactam	Artemisin		Quinupristin-dalfopristin ^l
	Piperacillin-tazobactam	Cefoperazone		Vancocycin ^o
	Ticarcillin-clavulanic acid		Clindamycin ^p	
	Cefamandole or cefonicid or cefuroxime		Linezolid	
	Cefepime	Cefepime	Trimethoprim-sulfamethoxazole	
	Cefmetazole	Ciprofloxacin	Vancocycin	
	Cefoperazone ^q			
	Cefotaxime ^{q, h, i} or ceftriaxone ^{q, h, i} or ceftazidime ^{q, h, i}	Imipenem or meropenem		
	Ciprofloxacin ^q or	Tobramycin		

Detecting and Reporting Antimicrobial Resistance

- ◆ *Staphylococcus aureus*
 - MRSA
 - CA-MRSA
 - VISA, VRSA
- ◆ Vancomycin-resistant enterococcus (VRE)
- ◆ Extended-spectrum β -lactamase producers (ESBLs)
- ◆ Others

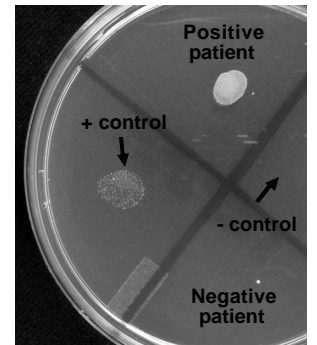
Staphylococcus aureus Testing Methods Reporting

S. aureus

S. aureus Testing

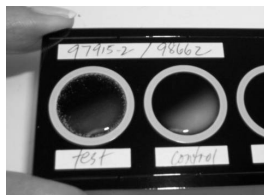
- ◆ Disk diffusion or MIC tests (e.g. Vitek, MicroScan)
- ◆ Oxacillin-salt agar screen plate
- ◆ Detection of gene or gene product
 - "Isolates of staphylococci that are shown to carry the *mecA* gene, or that produce PBP2a, the gene product, should be reported as oxacillin resistant" (NCCLS, 2003)

Oxacillin-salt agar
screen for *S. aureus*
(MHA + 4% NaCl + 6 ug/ml
oxacillin)



PBP2a rapid latex agglutination assay kit

- Perform on isolated colonies
- Extract PBP2a
- React with latex antibody to PBP2a



Report Example (following PBP2a assay): Leg Wound Culture

Gram Stain (day 1):
Many GPC clusters
Many WBCs

Preliminary Culture Report (day 2):
Many:
Staphylococcus aureus, oxacillin-resistant (MRSA)

"Additional susceptibility results to follow"

Staphylococcus aureus

clindamycin S
 erythromycin S
 oxacillin S
 penicillin R
 vancomycin S

“Cefazolin and other β -lactams (except amoxicillin, ampicillin, and penicillins) are active against oxacillin-S and penicillin-R staphylococci.”

Consider adding comment to report to further explain β -lactam results

Staphylococcus aureus

clindamycin R
 erythromycin R
 oxacillin R
 penicillin R
 vancomycin S

“Oxacillin-R staphylococci are resistant to all β -lactams. MRSA isolated, please check Infection Control policies.”

Staphylococcus aureus

cefazolin \odot R*
 clindamycin R
 erythromycin R
 oxacillin R
 penicillin R
 vancomycin S

**If any β -lactam is tested and tests “S”, do not report or change to “R” for MRSA*

Staphylococcus aureus

clindamycin S
 erythromycin R
 oxacillin R
 penicillin R
 vancomycin S

Historically, MRSA have been multiply resistant to other anti-staphylococcal agents. However, many community associated strains are not multiply resistant.

***Staphylococcus spp.* Erythromycin / Clindamycin**

Mechanism	Determinant	Erythro	Clinda
Efflux	<i>msrA</i>	R	S
Ribosome alteration	<i>erm</i>	R	S*
Ribosome alteration	<i>erm</i>	R	R constitutive

msrA = macrolide streptogramin resistance
erm = erythromycin ribosome methylase
 *requires induction to show resistance

Staphylococcus aureus

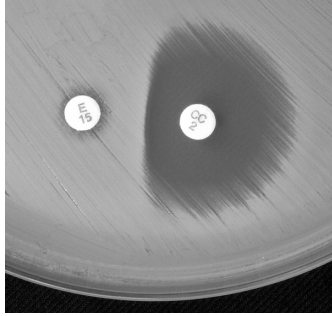
erythromycin R
 oxacillin R
 penicillin R
 vancomycin S

“Contact laboratory if clindamycin results needed”

If clindamycin-S, erythromycin-R, do not report as clindamycin-S without performance of “D Test”

“D Test”

Inducible
Clindamycin
Resistance
(*erm*-mediated)



“D Test” positive – forthcoming NCCLS
comment suggestion

Staphylococcus aureus

clindamycin	R
erythromycin	R
oxacillin	R
penicillin	R
vancomycin	S

“This *S. aureus* is presumed to be resistant based on detection of inducible clindamycin resistance. Clindamycin may still be effective in some patients.”

“D Test” negative

Staphylococcus aureus

clindamycin	S
erythromycin	R
oxacillin	R
penicillin	R
vancomycin	S

“This *S. aureus* DOES NOT demonstrate inducible clindamycin resistance in vitro.”

Staphylococcus spp. Vancomycin

	MIC ($\mu\text{g/ml}$)
Susceptible	≤ 4
Intermediate	8 - 16
Resistant	≥ 32

VISA = (4) - 16 $\mu\text{g/ml}$
VRSA = ≥ 32 $\mu\text{g/ml}$

NCCLS M100-S13; Table 2C

VISA

- ◆ 8 cases to date in USA
- ◆ Pts. previously had MRSA
- ◆ Pt. previously treated with vancomycin
- ◆ Most are MRSA

Fridkin et. al. 2001. Clin. Infect. Dis. 32:108.
Fridkin et. al. 2003. Clin. Infect. Dis. 36:429.

VRSA

- ◆ 2 cases to date
 - June 2002 – Michigan
 - September 2002 – Pennsylvania
- ◆ Both pts. previously had MRSA and VRE (*vanA*) and were treated with vancomycin
- ◆ VRSA had *mecA* and *vanA*
- ◆ “S” to other agents (chloramphenicol, linezolid, minocycline, quin-dalfo, TMP-SMZ)

MMWR. 2002; 51:565-7.
Chang et al. 2003. NEJM 348:1342

VISA / VRSA

Some test methods DO NOT detect
VISA or VRSA!!

If you suspect vancomycin problems in
treating *S. aureus* infection, contact
laboratory and suggest further testing!!

Enterococcus spp.

Testing Methods
Reporting

Enterococcus

Enterococcus spp. Testing

◆ AST of isolates from diagnostic
specimens:

- Disk diffusion or MIC tests (e.g. Vitek,
MicroScan)
- BHI-vancomycin screen plate

◆ VRE surveillance

- Culture media with vancomycin; sub primary
specimens (e.g. stool) onto this

E. faecalis (blood)

ampicillin	S
vancomycin	S
gent syn	R
strep syn	S

“Serious enterococcal infections need combination
therapy with amp, pen, or vancomycin plus an
aminoglycoside. Synergy occurs only when both
drugs are susceptible.”

E. faecium (blood)

ampicillin	R
vancomycin	R
gent syn	R
strep syn	R

“VRE isolated; Infectious Diseases
consult suggested”

E. faecium (blood)

MIC (µg/ml)

ampicillin	>64 R
chloramphenicol	8 S
ciprofloxacin	4 R
doxycycline	2 S
linezolid	1 S
quin/dalfo	0.5 S
rifampin	8 R
vancomycin	>32 R
gent synergy	R
strep synergy	R

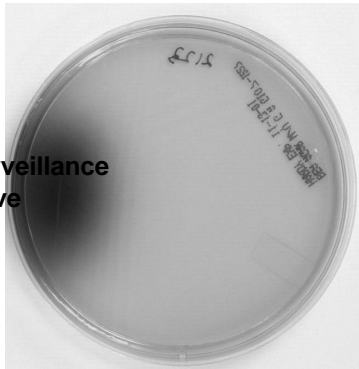
***E. faecium* (urine)**

ampicillin	R
ciprofloxacin	R
nitrofurantoin	S
tetracycline	R
vancomycin	R

VRE Stool Surveillance
Positive



VRE Stool Surveillance
Negative



Stool Surveillance Report

Many *Enterococcus faecium*

“VRE isolated; please check
Infection Control policy”

Extended-spectrum b-lactamases (ESBLs)*

*enzymes that destroy extended-spectrum
b-lactam drugs

ESBLs

ESBLs – Testing

- ◆ **Organisms:**
E. coli, *Klebsiella* spp., (others??)
- ◆ **Screen test:**
Look for decreased susceptibility (small zone or elevated MIC) to extended-spectrum β -lactam agents
- ◆ **Confirmatory test:**
 β -lactam activity restored by the β -lactamase inhibitor clavulanic acid
 - If positive, must change “S” result to “R” for any penicillin, cephalosporin, or aztreonam

Klebsiella pneumoniae ?ESBL

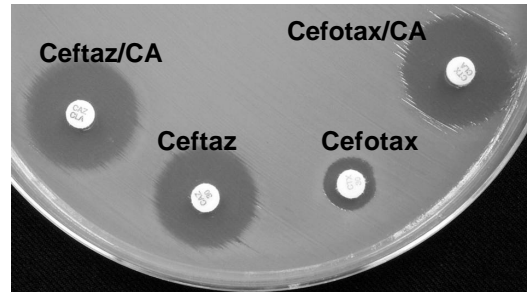
PRELIM:

	MIC (µg/ml)
cefoxitin	1 S
ciprofloxacin	0.5 S
imipenem	£ 0.25 S
pip-tazo	8 S
am, cfaz, gm, T-S	R

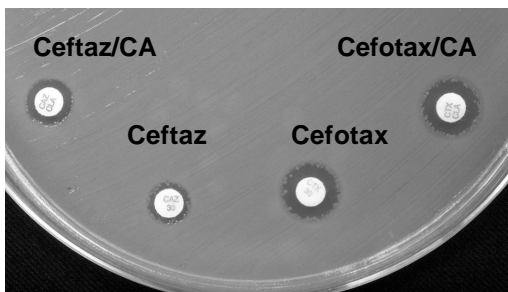
*hold cefepime, cefotaxime if "S"

"This *K. pneumoniae* is suspicious for extended-spectrum beta-lactamase (ESBL) production; confirmatory tests pending."

ESBL Confirmatory Test



ESBL Confirmatory Test



Klebsiella pneumoniae ESBL

FINAL:

	MIC (µg/ml)
cefepime	⊗ R
cefoxitin	1 S
cefotaxime	⊗ R
ciprofloxacin	0.5 S
imipenem	0.25 S
pip-tazo	8 S
am, cfaz, gent, T-S	R

"Confirmatory tests for this *K. pneumoniae* indicate unusual resistance [extended-spectrum beta-lactamase (ESBL) production]; Infectious Diseases consult suggested."

...More report examples

...More

Throat Culture

Many Group A Streptococcus

"Group A Streptococcus remains universally susceptible to penicillin"

Specimen: Anovaginal
Test: Prenatal screen

Group B Streptococcus

“Group B Streptococci are susceptible to ampicillin, penicillin and cefazolin, but may be resistant to erythromycin and/or clindamycin. Contact laboratory if erythromycin and/or clindamycin testing is needed.”

S. pneumoniae (blood)

	MIC (µg/ml)
ceftriaxone (meningitis)	1 I
ceftriaxone (non-meningitis)	1 S
erythro, trim-sulfa	R
levofloxacin	0.5 S
penicillin	1 I
vancomycin	0.5 S

“Use of cefotaxime or ceftriaxone in meningitis requires therapy with maximum doses. High dose IV pens (e.g. at least 2 ml U every 4 h in adults with normal renal function) or amp are effective in treating pneumococcal pneumonia due to strains in the penicillin “int” category.”

Blood Culture

“Coagulase-negative staphylococcus isolated from one culture only. Susceptibility testing is not routinely done. If testing is indicated, please contact lab within 24 h.”



Sputum Culture

GS:

- Many oral flora
- Many WBCs
- Many GNR

1 potential pathogen
Perform AST

Culture:

- Many Normal Flora
- Many *Klebsiella pneumoniae*

Leg Wound Culture

GS:

- Many GPC clusters
- Many pleomorphic GPR

Culture:

- Many coag-neg staphylococci*
- Many diphtheroids
- Few *E. coli**
- Few *Proteus mirabilis**

***Susceptibility results reported per Dr. Jones' request.”

Local Cumulative Antibigram Report

Antibiograms

Community Hospital Antibiogram 2002 – Exmp.

	% Susceptible						
	N	Am	Cf	Ctx	Cip	Gm	Pp T-S
<i>E. coli</i>	729	61	92	95	92	97	66 76
<i>E. cloacae</i>	144	-	-	71	95	88	65 84
<i>P. aerug</i>	221	-	-	10	76	88	91 -

NCCLS M39-A Guideline

“Analysis and Presentation of Cumulative Antimicrobial Susceptibility Test Data”

- ◆ May 2002
- ◆ Guide MDs on EMPIRIC THERAPY
- ◆ Additional reports may be needed for infection control



M-39A Recommendations

- ◆ Analyze/present data annually
 - Sufficient to GUIDE EMPIRIC THERAPY
- ◆ Include only species with [≥] 10 isolates
- ◆ Include diagnostic (not surveillance) isolates
- ◆ Exclude duplicates - include 1st isolate/patient, irrespective of
 - body site, antimicrobial pattern
- ◆ Include only drugs routinely tested

M-39A Recommendations (con't)

- ◆ Calculate %S (do not include %I)
- ◆ Special circumstances
 - *S. aureus* – all and MRSA subset
 - *S. pneumoniae* - %S and %I (penicillin)
 - *S. pneumoniae* – meningitis, non-meningitis (cefotaxime / ceftriaxone)
 - Viridans *streptococcus* - %S and %I (penicillin)

Antibiogram 2002 – Exmp.

	% Susceptible					
	N	Clin	Ery	Ox	Pen	T-S Van
all SA	1317	80	50	68	9	97 100
MRSA	449	44	4	0	0	94 100

Antibiogram 2002 – Exmp.

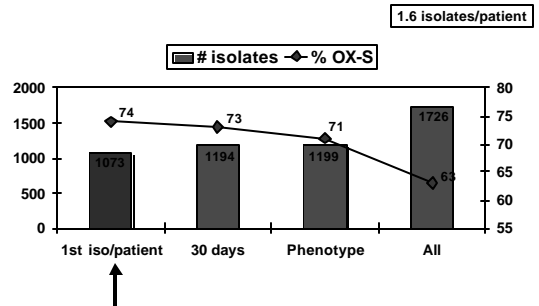
	% Susceptible					
	N	Cftrx	Ctx	Ery	Levo	Pen
<i>S. pneumoniae</i>	107	-	-	65	100	65*
meningitis		88	90	-	-	-
non-meningitis		97	96	-	-	-

*Susc = MIC ≤ 0.06 mg/ml
 Non-susceptible includes: 20% Int (MIC 0.12-1.0 mg/ml)
 15% high-level Res (MIC >1.0 mg/ml)

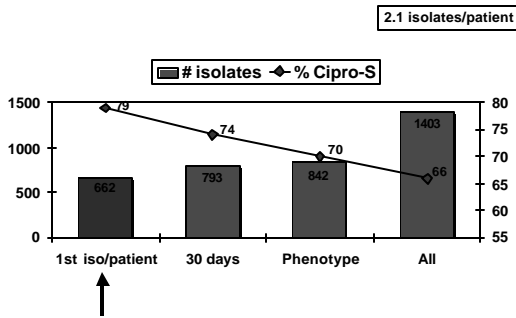
Limitations M39-A

- ◆ Will not identify very unusual phenotype if this is not the “first” isolate on the patient (e.g. VISA)
 - Daily QA checks should catch these
 - Option to additionally search entire database
- ◆ Significant Lab Computer limitations
 - Diagnostic test system (e.g. MicroScan, Vitek)
 - Laboratory Information System (LIS) (e.g. Cerner, Meditech, Mysis, etc.)
 - NCCLS is working with vendors to obtain assistance

S. aureus - Oxacillin



P. aeruginosa – Ciprofloxacin



P. aeruginosa – Ciprofloxacin

Variable results from analysis of single dataset

	N	%S	%I	%R
1st iso /pt	662	79*	7	14
All	1403	66	11	23

*M39-A recommendation

Problem: testing broad-spectrum agents only on isolates resistant to narrower-spectrum agents...

	N	<i>P. aeruginosa</i> % Susceptible			
		Cip	Ctaz	Gm	Impip
Hospital A	225	81	88	79	87
Hospital B	199	84	85	83	51*

*Imipenem only tested on ceftazidime-R isolates (n=29)

Problem: large number of urine isolates can dilute data...

	N	<i>E. coli</i> % Susceptible			
		Ctx	Cipro	Gm	T-S
All isolates	2993	99	92	93	76
Urine only	2799	99	93	93	77
Excl urine isolates	248	93	83	90	60

Problem: testing small numbers of isolates may reflect an intermittent resistant clone...

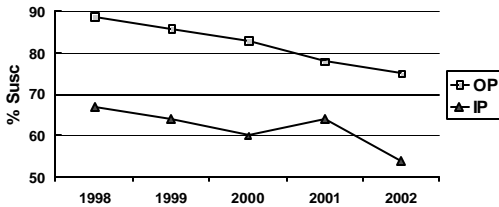
	<i>Morganella morganii</i>				
	% Susceptible				
	N	Ctx	Cipro	Gm	T-S
Hospital A	84	86	92	96	89
Hospital B*	22	10	100	10	10

*small hospital that had an outbreak of *M. morganii* in MICU during a 4 week period

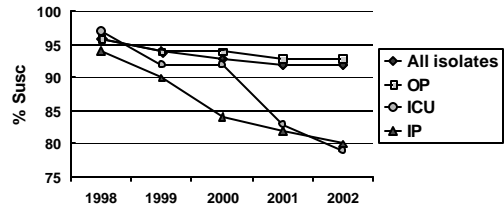
Problem: how to report VRE?? Not all enterococci are identified to species level...

	vancomycin	
	N	% Susc
<i>E. faecium</i> -all	226	9
<i>E. faecium</i> -bld	53	36
<i>Enterococcus</i> spp.- bld	111	71
<i>E. faecium</i> -stool	92	0
<i>Enterococcus</i> spp.-all	1144	79

**Trend Example 1998-2002
%S for *S. aureus* - Oxacillin
IP vs. OP**



**Trend Example 1998-2002
%S for *E. coli* - Ciprofloxacin
All and OP, ICU, IP**



So, is a 18% decrease from 2002 to 2003 in %S of gentamicin for *P. aeruginosa* due to:

- ◆ Antibigram calculation artifact?
- ◆ Infection control issue?
- ◆ Antibiotic utilization issue?
- ◆ Change in patient mix?
- ◆ Other?

95% Confidence Intervals for %S with Selected Sample Sizes

Sample size	50% S	90% S
10	19-81	55-100
100	40-60	82-95
1000	47-53	88-92

What's Next for M39 ??

- ◆ Expand suggestions for:
 - “Highlighting” certain data points
 - Specific presentation of data subsets (e.g., inpatient vs. outpatient)
 - Plotting data from year to year
 - Use of statistics (will provide table) to help determine significance of “x% change” in %S for “N isolates” from year to year

What Can You Do in Your Facility re: our Antimicrobial Resistance Dilemma?

- ◆ Interact with your laboratory
 - Exchange “scientific” information
 - Communicate your needs (e.g. immediate notification of select resistance results such as MRSA, VRE)
 - Develop policies as a team and “test” policies
- ◆ Interact with clinicians
 - Encourage culturing
 - Suggest use of AST reports and antibiograms to help in therapy decisions
- ◆ Maintain awareness of resources for information on antimicrobial resistance and share this with your microbiology laboratory, please!

Thank you!



Visit this site and order “FREE”: CD ROM

www.phppo.cdc.gov/dls/master/default.asp

Understanding Antimicrobial Resistance in Your Facility: the Patient Report and the Cumulative Antibiogram

Reference Material

Laboratory-oriented:

<http://www.phppo.cdc.gov/dls/master/default.asp>

MASTER (multilevel antimicrobial susceptibility educational resource) website!!!

-Includes case studies, Q&A, literature review and more

-describes how to obtain the new and "free" CDC CD-ROM on antimicrobial susceptibility testing

<http://www.cdc.gov/ncidod/hip/Lab/LAB.HTM>

CDC antimicrobial susceptibility testing fact sheets

Surveillance for Emerging Antimicrobial Resistance Connected to Healthcare (S.E.A.R.C.H.)
Confirmatory Reference Testing

Provides instructions for reporting VISA or VRSA to Public Health Departments.

<http://www.cdc.gov/ncidod/hip/ARESIST/search.htm>

<http://www.asm.org/division/c/index.htm>

ASM Division C (clinical microbiology) website includes "Askit" feature.. you can ask any question about clinical microbiology here!!!

Others:

<http://www.cdc.gov/drugresistance/>

CDC drug resistance

CDC Campaign to Prevent Resistance in Healthcare Settings

http://www.cdc.gov/ncidod/hip/ARESIST/visa_vrsa_guide.pdf

Investigating VISA and VRSA – A Guide for Health Departments and Infection Control Personnel

<http://www.idsociety.org/>

Infectious Diseases Society of America (IDSA)

<http://www.tufts.edu/med/apua/>

Alliance for Prudent Use of Antibiotics (APUA)

Text References

Gilbert, D. N., R. C. Moellering, and M. A. Sande (eds). 2003. *The Sanford Guide to Antimicrobial Therapy, 2003*. 33rd ed. Antimicrobial Therapy, Inc., Hyde Park, VT. (www.Sanfordguide.com)

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Notes: