DEPARTMENT OF HEALTH AND HUMAN SERVICES

PUBLIC HEALTH SERVICE

FOOD AND DRUG ADMINISTRATION

MILK LABORATORY EVALUATION FORM

LABORATORY		
LOCATION		LAB #
DATE	X = DEVIATION	U = UNDETERMINED
	O = NOT USED	NA = NOT APPLICABLE

PASTEURIZED MILK CONTAINERS [Unless otherwise stated all tolerances are ±5%]

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1.	Laboratory Requirements	b. Plastic capped containers (submitted with caps)
	a. Record time and date when samples received	1. Swab top of container with 70% alcohol when appropri-
	b. Record time and date when samples examined	ate
	DINOS METUOD	2. Add required amount of rinse solution by aseptically
	RINSE METHOD	removing cap, pouring in solution without touching the
	<u>APPARATUS</u>	top and replace cap
2.	See Cultural Procedures, items 1-23	3. Complete rinse procedure as described in 10a 3/9 above
3.	Adding Rinse Solution to Containers	c. Flexible-walled containers/bags
	a. Sterile hypodermic syringes (capacity 20 or 100 mL) and	1. Add 100 mL aseptically by swabbing an area of tube
	needles	adjacent to liner with 70% alcohol; introduce rinse by
	b. Or, sterile pipets	syringe and seal puncture with plastic tape
	c. Or, sterile automatic syringe	2. Place container/bag on smooth, clean, firm horizontal
	d. Or, sterile graduate cylinder	surface as flat as its construction permits
	e. Or, pre-dispensed dilution bottles or tubes with rinse	3. With hands or roller, move rinse solution back and forth
	solution, volumes checked as in CP item 28i	10 times, contacting all surfaces completely
	••	4. Lift liner and hang with "fill tube" down to permit rinse
	<u>Materials</u>	solution to collect for one minute
4.	See Cultural Procedures, items 24-32	5. Transfer rinse solution to sterile container by cutting "fill
5.	Rinse Solutions, see CP items 27 i-k	tube" with sterile scissors
6.	Ethyl Alcohol, 70%	11. Sample Measurements
7.	Plastic Tape	a. As described in SPC, item 6 and 7 except
	Docernupe	b. For Residual Bacterial Count (RBC), plate the following
	<u>Procedure</u>	amounts per container
8.		1. ½ pint: 5 mL in two plates
9.		2. Pint and larger: 2 mL in a single plate
	Samples	c. For Residual Coliform Count (RCC), distribute 10 mL of
	a. 20 mL for ½ pints (236 mL), pints (473 mL), and quarts (946	remaining rinse solution among three plates
	mL)	12. Plating (See SPC, item 13)
	b. 50 mL for ½ gallons (1892 mL)	13. Controls (See SPC, item 14)
	c. 100 mL for gallons (3784 mL) or larger	14. Incubation (See SPC, item 15)
	d. Containers less than 100 mL use swab method, see items 19 -	15. Counting Colonies (See SPC, items 16-18)
40	34	16. Confirmation Test (See SPC, item 18e)
10	Collection of Surface Rinse Samples	17. Recording Counts
	a. Firm walled paper containers, sealed on line	a. Count obtained from RBC plate recorded as colonies
	Swab top of containers with 70% alcohol Add required amount of rinse solution to each container	b. If no colonies on RBC plate, record as 0
	by injection and seal puncture with plastic tape	c. Count obtained from RCC plate recorded as colonies
	3. Vigorously shake container length-wise 10 times, holding	counted
	container horizontally	d. If no colonies on RCC plate, record as 0
	Each shake a complete back and forth movement of about	e. Values are recorded as number of colonies per container
	8 in	c. values are recorded as number of colonies per container
	5. Turn container 90° and repeat horizontal shaking treat-	<u>REPORTS</u>
	ment	18. Reporting Counts
	6. Turn container 90° twice more and repeat horizontal	a. Report computed count as Residual Bacterial Count (RBC)/
	shaking	specified container capacity in mL
	7. Grasp container and swirl 20 times in a small circle while	1. Containers rinsed with 20 mL
	upright (top up)	a. 5 mL plated for RBC, multiply colonies by 4
	8. Invert (top down) and repeat swirling of container 20	b. 2 mL plated for RBC, multiply colonies by 10
	times	2. Containers rinsed with 50 mL
	9. Stand upright and allow to drain for 1 min	a. 2 mL plated for RBC, multiply colonies by 25

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3. Containers rinsed with 100 mL		a. Aseptically remove sterile swab from container	
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		area (calculate or use sterile template) three times, reversing	
a. 10 mL plated for RCC, multiply colonies by 2		direction between successive strokes	
		d. Rinse swab in solution and press out excess	
a. 10 mL plated for RCC, multiply colonies by 5		e. Swab four additional 50 sq. cm areas	
a. 10 mL plated for RCC, multiply colonies by 10	_	and break stick, leaving swab head in vial	
c. If no colonies appear on plates, report as less than n/size of	27.	. Sample Measurement	
specified container in mL, substituting for n the number that		a. As described in SPC item 6 and 7 except	
would be reported if 1 colony had been counted from the		1. If calcium alginate wool is used, add 0.5 mL of sterile Na	
volume of rinse solution plated and multiply by appropriate		Hexa-metaphosphate solution to 4.5 mL rinse solution in	
factor	_	vial and shake until dissolved	
OWAR METUOR		2. For all other fibers	
		a. Shake swab container 50 times	
<u>APPARATUS</u>		b. Each shake a complete back and forth movement of 15	
		cm	
	_	c. Strike palm of hand at end of each cycle	
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MATERIALS		u. II IIO cololiles oli Noo piate, lecolu as o	
		REPORTS	
		Renorting Counts	
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<u>Procedure</u>			
Identify Plates (See SPC, item 4)			
	-		
	b. Report computed coliform count as Residual Coliform Count/container	a. 2 mL plated for RBC, multiply colonies by 50	a. 2 mL plated for RBC, multiply colonies by 50. Report computed coliform count as Residual Coliform Count/container 1. Containers rinsed with 20 mL. a. 10 mL plated for RCC, multiply colonies by 2. 2. Containers rinsed with 50 mL. a. 10 mL plated for RCC, multiply colonies by 5. a. 10 mL plated for RCC, multiply colonies by 5. b. Containers rinsed with 50 mL. a. 10 mL plated for RCC, multiply colonies by 5. d. Rinse swab in solution and press out excess. e. Swab four additional 50 Sq. cm areas. first rith rain rain and press out excess. e. Swab four additional 50 Sq. cm areas. first rith rain rain and press out excess. e. Swab four additional 50 Sq. cm areas. first rith rain rain and press out excess. e. Swab four additional 50 Sq. cm areas. first rith rain rain and press out excess. e. Swab four additional 50 Sq. cm areas. first rith rain rain as been swabbed on position swab head in vial and break stick, leaving swab head in vial and shake until dissolved. 2. For all other fibers a. 7 to 10 cm long to contain 5 mL. solution for non-sol