

## **Abstract**

Henneberry, T.J., R.M. Faust, W.A. Jones, and T.M. Perring, eds. 2002. Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan (formerly Sweetpotato Whitefly, Strain B): Fourth Annual Review of the Second 5-Year Plan and Final Report for 1992–2002, held in San Diego, California, February 10–12, 2002. U.S. Department of Agriculture, Agricultural Research Service, 446 pp.

To ensure timely distribution, this report was reproduced essentially as supplied by the authors. It received no publications editing and design. The authors' views are their own and do not necessarily reflect those of the U.S. Department of Agriculture.

Mention of trade names, commercial products, or companies in this publication is solely for the purpose of providing specific information and does not imply recommendation or endorsement by the U.S. Department of Agriculture over others not mentioned.

This publication reports research involving pesticides. It does not contain recommendations for their use nor does it imply that uses discussed here have been registered. All uses of pesticides must be registered by appropriate state or Federal agencies or both before they can be recommended.

While supplies last, single copies of this publication may be obtained at no cost from T.J. Henneberry, U.S. Department of Agriculture, Agricultural Research Service, 4135 E. Broadway Rd., Phoenix, AZ 85040-8803.

Copies of this publication may be purchased from the National Technical Information Service, 5285 Port Royal Road, Springfield, VA 22161; telephone (703) 605-6000 or 1-800-553-6847.

The U.S. Department of Agriculture (USDA) prohibits discrimination in all its programs and activities on the basis of race, color, national origin, sex, religion, age, disability, political beliefs, sexual orientation, or marital or family status. (Not all prohibited bases apply to all programs.) Persons with disabilities who require alternative means for communication of program information (Braille, large print, audiotape, etc.) should contact USDA's TARGET Center at 202-720-2600 (voice and TDD).

To file a complaint of discrimination, write USDA, Office of Civil Rights, Room 326-W, Whitten Building, 1400 Independence Avenue, SW, Washington, DC 20250-9410 or call 202-720-5964 (voice and TDD). USDA is an equal opportunity provider and employer.

Issued June 2002

## Contents

Editors' Comments .....	iii
Progress Review, Research, Action Plan, and Technology Transfer Organizational Team and Acknowledgements .....	iii
Preface .....	iv
Executive Summary .....	iv
Introduction.....	v
I. Sweetpotato Whitefly (SPW) Strain B: Accomplishments of the National Research and Action Plans 1993 to 2002 and Future Goals .....	1
II. Plenary Session Keynote Address Summaries .....	44
Section A. Dale Gelman .....	44
Section B. Judy Brown and Robert Gilbertson .....	44
Section C. Nilima Prabhaker .....	46
Section D. Don Vacek.....	48
Section E. Greg Walker.....	49
Section F. Steve Castle.....	54
III. Reports of Research Progress for 2000 and 2001 .....	56
A. Biology, Ecology, and Population Dynamics .....	56
Research Abstracts .....	56
Research Summary .....	82
Table A.1 Summary of Research Progress 1997 Goals Statement .....	83
A.2 Summary of Research Progress 1998 Goals Statement .....	87
A.3 Summary of Research Progress 1999 Goals Statement .....	90
A.4 Summary of Research Progress 2000 and 2001 Goals Statement .....	93
B. Viruses, Epidemiology, and Virus-Vector Interactions .....	99
Research Abstracts .....	99
Research Summary .....	109
Table B.1 Summary of Research Progress 1997 Goals Statement.....	112
B.2 Summary of Research Progress 1998 Goals Statement .....	115
B.3 Summary of Research Progress 1999 Goals Statement .....	119
B.4 Summary of Research Progress 2000 and 2001 Goals Statement .....	124
C. Chemical Control, Biopesticides, Resistance Management, and Application Methods.....	128
Research Abstracts .....	128
Research Summary .....	147
Table C.1 Summary of Research Progress 1997 Goals Statement.....	148
C.2 Summary of Research Progress 1998 Goals Statement .....	150
C.3 Summary of Research Progress 1999 Goals Statement .....	152
C.4 Summary of Research Progress 2000 and 2001 Goals Statement .....	155

D.	Natural Enemy Ecology, and Biological Control.....	157
	Research Abstracts .....	157
	I. Research Summary .....	170
	Table D.1 Summary of Research Progress 1997 Goals Statement .....	173
	D.2 Summary of Research Progress 1998 Goals Statement .....	175
	D.3 Summary of Research Progress 1999 Goals Statement .....	177
	D.4 Summary of Research Progress 2000 and 2001 Goals Statement .....	179
E.	Host-Plant Resistance, Physiological Disorders, and Host-Plant Interactions.....	182
	Research Abstracts .....	182
	Research Summary .....	195
	Table E.1 Summary of Research Progress 1997 Goals Statement .....	198
	E.2 Summary of Research Progress 1998 Goals Statement .....	200
	E.3 Summary of Research Progress 1999 Goals Statement .....	202
	E.4 Summary of Research Progress 2000 and 2001 Goals Statement .....	205
F.	Integrated and Area-wide Pest Management Approaches, and Crop Management Systems .....	208
	Research Abstracts .....	208
	Research Summary .....	213
	Table F.1 Summary of Research Progress 1997 Goals Statement .....	216
	F.2 Summary of Research Progress 1998 Goals Statement .....	218
	F.3 Summary of Research Progress 1999 Goals Statement .....	221
	F.4 Summary of Research Progress 2000 and 2001 Goals Statement .....	223
IV.	Appendices .....	225
	Appendix A. Selected National and International <i>Bemisia</i> Research and Information Coordination .....	225
	Appendix B. Bibliography of <i>Bemisia tabaci</i> (Gennadius) & <i>Bemisia argentifolii</i> Bellows and Perring.....	227
	Appendix C. Meeting Agenda.....	416
	Appendix D. List of Registered Meeting Participants.....	420
	Appendix E. Minutes of the Program Planning Review Committee.....	425
	Appendix F. Five-Year National Research and Action Plan Priority Tables, Research Needs, and Yearly Goals: (1997-2001).....	426

## Editors' Comments

Henneberry, T. J., R. M. Faust, W. A. Jones and T. M. Perring (eds.) 2002 Sweetpotato (Strain B, Silverleaf) Whitefly Five-Year National Research and Action Plans; Fourth Annual Review of the Second Plan and Final Report for 1992 to 2002. The meeting was held in San Diego, CA, February 10-12, 2002.

This publication contains research, extension-education, industry and action agency reports of progress contributing to our knowledge of the whitefly complex and to the development of ecologically acceptable whitefly management systems. The multi-agency cooperative effort has, since 1992, provided a forum for information exchange, complementary, coordinated and cooperative research programs, avoidance of duplication of effort, and optimum return for expended research dollars. The result of the joint partnerships has been solutions and technology transfer to the stakeholders in the agricultural communities. These accomplishments have been achieved within a rapid timeframe, in large part, due to openness of communications, sharing of expertise and focus on common goals. The editors sincerely thank all those who participated in the annual reviews of the research and action plans. This year's publication supplement contains a final report for Whitefly National Research and Action Plans from 1992 to 2002. Although this year's activities terminate the formal functioning of the plan, communication guidelines and identification of other whitefly research and information exchange coordinating groups worldwide are provided in Appendix A.

## Progress Review Organizational Teams

### USDA Silverleaf Whitefly Research, Education and Implementation Coordinating Group:

R. M. Faust, Chair, ARS, Beltsville, MD  
R. Carruthers, Chair, ARS, Albany, CA  
J. R. Coppedge, Alternate, ARS, College Station, TX  
D. D. Kopp and R. Huettel (Alternate), CSREES, Washington, DC  
E. Delfosse (Alternate), APHIS, Riverdale, MD  
N. Toscano, State Agricultural Experiment Station representative, CA  
J. Brown, State Agric. Experiment Station representative, AZ

### Annual Review Program Chairs:

Walker Jones, USDA-ARS, Weslaco, TX  
Tom Perring, Department of Entomology, University of California, Riverside, CA

### Local and State Coordinators:

Amy Corley, University of California, Riverside, CA.

## Silverleaf Whitefly 2002 Program Planning and Review Committee

R. M. Faust, ARS, NPS, Beltsville, MD  
T. M. Perring and W. Jones, Program Chairs  
Amy Corley, Local arrangements 2002 review  
T. J. Henneberry and N. C. Toscano, Advisors  
Jackie Blackmer and David Byrne, Section A Chairs  
Bob Gilbertson and Judy Brown, Section B Chairs  
Shirley Taylor and John Palumbo, Section C Chairs  
Bill Roltsch and Greg Simmons, Section D Chairs  
Greg Walker and Cindy McKenzie, Section E Chairs  
Steve Castle and Peter Ellsworth, Section F Chairs

## Silverleaf Whitefly Working Group 2002:

Amy Corley, University of California, Riverside, CA

## J. Blackmer, USDA-ARS WCRL, Phoenix, AZ

J. Brown, University of Arizona, Tucson, AZ  
D. Byrne, University of Arizona, Tucson, AZ  
S. Castle, USDA-ARS WCRL, Phoenix, AZ  
P. Ellsworth, University of Arizona, Maricopa, AZ  
R. Faust, USDA-ARS-NPS, Beltsville, MD  
B. Gilbertson, University of California, Davis, CA  
T. Henneberry, USDA-ARS, WCRL, Phoenix, AZ  
C. McKenzie, USDA-ARS-USARL, FL  
J. Palumbo, University of Arizona, Yuma, AZ  
T. Perring, University of California, Riverside, CA  
B. Roltsch, CDFA, Brawley, CA  
G. Simmons, USDA-APHIS, Bakersfield, CA  
S. Taylor, Bayer Corporation, La Mesa, CA  
N. Toscano, University of California, Riverside, CA  
G. Walker, University of California, Riverside, CA  
I. Wedderspoon, Ian Industries, Inc., Miami, FL

## Acknowledgments:

The USDA Silverleaf Whitefly, Research, Education and Implementation Coordinating Group; Annual Review Program Chairs; Local and State Coordinators; Sweetpotato whitefly Program Planning and Review Committee; and the Silverleaf Whitefly Working Group sincerely appreciate the contributions of all the participants and those who have helped in organizing the 2002 and other meetings. Recognition is extended to the University of California Center for Exotic Pest Research for their support and cooperation in meeting site selection and program organization.

## Preface

The Sweetpotato Whitefly, *Bemisia tabaci* (Gennadius) Strain B (= Silverleaf Whitefly, *Bemisia argentifolii* Bellows and Perring), National Research and Action Plans (1992-1997 and 1997-2002) were developed by USDA agencies (ARS, APHIS, and CSREES), state agencies, state agricultural experimental stations, and representatives of the cotton, vegetable, ornamental, nursery crop and chemical industries. The objectives were to establish research priorities, avoid duplication of effort, and maximize the use of existing resources. Research needs, goals and objectives, and technology transfer to clientele (scientific community, legislators, regulators, the agricultural industry, and the public) were reviewed on an annual basis. The plan was flexible allowing responsiveness to changing needs and priorities with appropriate adjustments to terminate, redirect, or add priorities based on funding, current knowledge, and program needs. The results were the development of environmentally and socially acceptable areawide, community-based whitefly management methods.

The USDA Sweetpotato Whitefly Research, Education, and Implementation Coordinating Group, throughout the duration of the work, facilitated USDA interagency and partner state agricultural experiment stations activities. The Working Group was composed of members of participating agencies and met annually to maintain communication with the USDA Coordinating Group and the Sweetpotato Strain B (Silverleaf) Whitefly Program Planning and Review Committee.

Intensive studies outlined in the research and action plans result in development of short-term relief from the devastating impact of explosive *Bemisia* populations using chemical methods. Selected insecticides applied alone or in mixtures based on developed action thresholds and Insecticide Resistance Management (IRM) methods have been highly effective on a short-term basis, but continuing long-term insecticide based management efficacy cannot be relied on. The research conducted within the guidelines of the plan has also resulted in a large base of information on *Bemisia* biology, ecology, behavior, physiology, systematics and other information that has improved our understanding of the essentials necessary to develop long-term strategies for SPW management. Thus, building blocks are provided for continuing efforts to develop areawide, community-based *Bemisia* management systems that incorporate cultural, biological, and nonchemical methods into chemical control-IRM-based control methods.

T. J. Henneberry  
USDA-ARS, WCRL  
Phoenix, AZ

R. M. Faust  
USDA-ARS-NPS  
Beltsville, MD

W. A. Jones  
USDA-ARS, SARC  
Weslaco, TX

T. M. Perring  
University of California  
Riverside, CA

## Executive Summary

The Whitefly National Research and Action Plan Reviews from 1993 to 2002 provided annual updates of the federal, state, and industry cooperative efforts to develop efficient management of methods for *Bemisia* populations. The work was initiated in 1991 when federal and state agency, agricultural experiment stations, industry and commodity group representatives, identified priority research areas for development into a national research and action plan for control of sweetpotato whiteflies. Unacceptable losses in cotton and vegetable field crop production were being experienced in California, Arizona, Texas and Florida, as well as ornamental and vegetable losses in glasshouse production.

During the years 1992 to 1997, extensive research achievements were made that provided interim solutions and a better understanding of the whitefly problem. Some of the research accomplished as part of the plan has been implemented by growers in their management efforts. A complete published review of the program showed extensive progress in all priority research areas. Over 70 examples of technology transfer to growers and the scientific community were documented.

The second 5-year plan, "The Silverleaf Whitefly, (*Bemisia argentifolii* Bellows and Perring) Research, Action and Technology Transfer Plan" was finalized at the annual review meeting at San Diego, CA on January 28-30, 1997.

The SPW has been referred to in the literature as the cassava whitefly, tobacco whitefly, poinsettia whitefly and cotton whitefly, as well as *B. tabaci* and a number of other scientific names (synonyms). The taxonomic relationships and the synonymy of the species have been reviewed on several occasions and remain unresolved. Race and/or biotype designations have also often been recorded in past literature in relation to host affinities and virus transmission vector interactions. The possibility of biotype occurrence has received continuing interest with the increasing importance of the sweetpotato whitefly in crop production on a worldwide basis. The detection of differences in electrophoretic isozyme patterns, biology and extended host range have provided evidence for the existence of different biotypes as compared to the type

previously encountered in the desert Southwestern United States crop production areas. This biotype has been referred to in the literature as the sweetpotato whitefly strain B. Based on the lack of nomenclatural resolution, *B. tabaci* strain B and *B. argentifolii* names used interchangeably in these reports are considered the same species.

(Table 1), but all of the work and participants contributed to the success of the plans.

### **Introduction**

The SPW National Research and Action Plans were developed to serve as guidelines for multi-agency, federal, state, private industry and consumer programs to reduce excessive SPW-induced losses occurring in the cotton, vegetable, nursery and glasshouse crop production industries in the United States. The extremely diverse SPW host range that included cotton, field, vegetable, ornamental and nursery crops in commercial or home garden and recreational and landscape settings resulted in implementation of the accomplishments and technology transfer not only to commercial production, but an extension of the benefits to private homeowners, urban communities and the non-agriculturally involved public sector in general.

Coordinated, multi-partner projects have a number of advantages: 1) adoption of standardized data collection protocols that are appropriate for comparison over diverse areas; 2) periodic reviews to exchange information and reprioritize research areas; 3) increased efficiency by sharing knowledge, resources and expertise; 4) multiple commodity input; and 5) increased returns per invested research dollar as opposed to multiple efforts in an uncoordinated manner.

The development of the action plans stimulated by the economic, environmental and social impact of the SPW species complex worldwide resulted in development of extensive information on SPW biology, ecology, physiology, viruses, virus-vector interactions and systematics. Implementation of this information has resulted in highly acceptable management systems with improved cost-benefit ratios. An important contributing factor to the success of the plans has been participation of all members of agricultural communities, growers, industry, scientists, bankers, and others sharing and implementing information provided by researchers, extension, and educational systems to develop large-scale multidisciplinary approaches to SPW management. Additionally, the success of the plans benefited, far beyond our ability to measure, from the experiences and research inputs from scientists of more than 14 foreign countries that participated in cooperative research and the annual plan reviews.

In this final review, we recap the research highlights and accomplishments. It would be impossible to capture the specifics of every item of investigation since over 1,000 reports of progress were published during the reviews

## **I. Sweetpotato Whitefly Strain B (SPW): Accomplishments of the National Research and Action Plans 1993 to 2002 and Future Goals.**

T. J. Henneberry<sup>1</sup> and R. L. Nichols<sup>2</sup>

<sup>1</sup> USDA-ARS, Western Cotton Research Laboratory,  
Phoenix, AZ

<sup>2</sup> Cotton Incorporated, Cary, NC

### **Early History of the Sweetpotato Whitefly in the United States**

*Bemisia tabaci* (Gennadius) was first reported in the United States in Florida in 1894 (Russell 1957, Russell 1975). It was subsequently collected on various crop and weed hosts in Washington D.C., Arizona, Southern California, Texas and Georgia. Little information on the biology of the SPW was reported that could serve to compare the cause of outbreaks occurring in the mid 1950s and in the late 1970s with those that occurred in late the 1980s and early 1990s.

Before the 1980s, SPW was primary a minor pest, except that it was known to vector certain plant diseases. Cotton Leaf Crumple (CLC) disease was observed in California in 1948 (Dickson et al. 1954). CLC was subsequently determined to be transmitted exclusively by SPW (Laird and Dickson 1959). Substantial cotton losses occurred in California and Arizona (Erwin and Meyer 1961, Allen et al. 1960, van Schaik et al. 1962) establishing SPW as an economic pest in the United States. Stub (perennial) cotton was suspected as the virus carryover source and when culture of perennial cotton was eliminated CLC incidence declined (van Schaik et al. 1962).

SPW during the 1960s seldom reached economic levels in Southern California cotton except when insecticides were used excessively (Gerling 1967). In the Imperial Valley from 1964 through 1981, no SPW were collected in 1964, 1965 and 1966; only low numbers occurred in 1967; and none were collected from 1968 to 1974 (Johnson et al. 1982). Numbers began to increase in the Imperial Valley in 1975, were variable through 1980, and increased dramatically in 1981 (Johnson et al. 1982). In 1981, SPW was reported as a rapidly escalating cotton problem in Arizona and Southern California (Butler and Henneberry 1984, 1985).

Subsequently, several virus-like diseases were found affecting a number of important commercial crops, and the diseases apparently were coincident with the high SPW populations (Duffus and Flock 1982, Duffus et al. 1986). Losses in 1981 were reported to exceed 100 million dollars (Duffus and Flock 1982). The SPW remained a problem of varying intensity in Arizona and California through the mid and late 1980s (Butler and Henneberry 1984, 1985, Butler et al. 1986). A major epidemic of CLC occurred in Arizona in 1981 (Brown

and Nelson 1984) and was a problem annually thereafter through 1985 (Brown and Nelson 1986).

In 1986, SPW outbreaks in Florida resulted in losses in the tomato industry alone that were estimated to be over 140 million dollars (Costa et al. 1993). In 1991 losses for cotton and vegetables in Texas were estimated to be 24.0 and 29.0 millions of dollars, respectively (Riley and Sparks 1993). In southern California, crop losses of over 100 million dollars a year and a reduction in 3,000 agricultural jobs annually have been reported since 1992 (Henneberry et al. 1993, 1994, 1995, 1996, 1997, 1998, 1999, 2000). SPW losses for ornamentals in 1991 were reported to exceed 23 million dollars (Barr and Drees 1992). SPW has been considered a major pest in most greenhouse cultures in the Southeastern United States (Oetting and Buntin 1996), but reliable loss figures are not available. In addition several previously undescribed diseases were observed, including irregular ripening in tomato, silverleaf of curcurbits, and blanching of brocolli, that were coincident with infestations of SPW; while whitefly populations and virus diseases declined in lettuce (Bharathan et al. 1990, Byrne et al. 1990, Yokomi et al. 1990).

The serious nature of the escalating SPW problem in many areas of the United States and the economic impact on the agricultural community highlighted the need for a coordinated national effort to provide short and long-term solutions to the problem. During the same period it was noted that certain populations of *B. tabaci* differed in reproductive potential. Strains with the lower and higher fecundity were termed A and B, respectively (Bethke and Paine 1991).

### **Sweetpotato Whitefly: 5-Year Plans for Development of Management and Control Methodology**

#### **The First Five-Year Plan 1992-1996**

By the end of the 1991 growing season, it was apparent that unacceptable losses in the field grown cotton and vegetable production were being experienced in California, Arizona, Texas, and Florida, as well as losses to ornamental and vegetable crops in glasshouse cultures at various locations in the South and Northeastern United States. The need for immediate management recommendations was urgent. Working from a draft plan (Anonymous 1992), representatives from federal and state agencies, agricultural experiment stations, industry and commodity groups identified six priority areas for development into a SPW national research and action plan (USDA 1992). They also identified the need for an immediate plan of research action for the 1992 growing season. Standardized experimental procedures, data collection protocols, report preparation formats, and a system for exchanging results were established for a national testing program of promising chemicals, natural products, microbial insecticides and improved application technology for SPW control on all major crops (Akey

1992). Additionally, the framework for an insecticide-resistance management program (IRM) was established. Scientists in California, Texas, and Florida initiated laboratory studies monitoring insecticide responses of field populations to develop baseline information as a cornerstone for long-term SPW-IRM. Over 30 federal and state experiment station scientists were involved in the overall program in eight states. The results of the coordinated, national chemical control efficacy trials, with annual modification, improvement, and testing of additional materials have provided the basis for highly effective SPW chemical control and ongoing SPW-IRM.

The plan was designated “The Five-Year National Research and Action Plan for Development of Management and Control Methodology for the Sweetpotato Whitefly” (USDA 1992). The high priority research areas identified were: (1) Ecology, Population Dynamics, and Dispersal; (2) Fundamental Research - Behavior, Biochemistry, Biotypes, Morphology, Physiology, Systematics, Virus Diseases, and Virus Vector Interactions; (3) Chemical Control, Biorationals, and Pesticide Application Technology; (4) Biological Control; (5) Crop Management Systems and Host Plant Resistance; and (6) Integrated Techniques, Approaches, and Philosophies.

Annual workshops were held to review progress and to adjust the plan as needed (Henneberry et al. 1993, 1994, 1995, 1996, 1997, 1998, 1999, 2000). After the annual review meeting in Orlando, FL in 1994 the titles of the reports of progress for the 5-year plans were changed to recognize the revised description the SPW strain B as a new species, *B. argentifolii* Bellows and Perring (Bellow et al. 1994). Because of the continuing nomenclatural controversy, the reluctance of international researchers to accept the new disintegration, and to facilitate continuity in the literature, in this publication, we consider SPW Strain B and *B. argentifolii* Bellows and Perring as synonymous

During the years 1992 to 1997, many research achievements were made that provided interim solutions and a better understanding of the SPW problem. Some of the research results that have been accomplished through coordination with the plan have been implemented as practices by growers. A complete published review of the program showed extensive progress in all priority research areas (Henneberry et al. 1997). Over 70 examples of technology transfer to growers and the scientific community were documented. A complete management system for SPW is a primary goal for the future. The decision to develop a second 5-year plan was adopted when 85% of the individuals attending the 1996 annual review responded to a question and indicated that a sequel to the existing plan should be prepared to maintain continuity, and a continuing high level of coordinated research and technology transfer.

The Second Five-Year Plan 1997-2001

The second 5-year plan, “The Silverleaf Whitefly, (*Bemisia argentifolii* Bellows and Perring) Research, Action, and Technology Transfer Plan” (Henneberry et al. 1997, 1998, 1999, 2000), was finalized at the annual review meeting at San Diego, CA on January 28-30, 1997. The high priority research areas are: (A) biology, ecology, and population dynamics; (B) viruses, epidemiology, and virus-vector interactions; (C) chemical control, biopesticides, resistance management, and application methods; (D) natural enemy ecology and biological control; (E) host plant resistance, physiological disorders, and host-plant interactions; and (F) integrated and areawide pest management approaches, and crop management systems.

Over 1,000 abstracts of research progress were presented at reviews conducted from 1993 to 2002. Tables 2 and 3 are summaries of the research highlights for the research priority approaches for the 1992 to 1997 and the 1997 to 2002 national plans, respectively. Research credits can be identified in the original abstract sections for each for each year.

For the long-term, the extensive base of information on biology, ecology, physiology, systematics, virus, virus-vector interactions and other fundamental information (Tables 2 and 3) developed and reported by the two five-years plans will provide the basis for new technically driven, ecologically oriented and economically acceptable SPW management programs. The practical applications and incorporation of much of this information is directly or indirectly included in the following documented management approaches. Much additional implementation can be accomplished and will improve the efficacy, the environmental compatibility, and social acceptance of insect management.

### **Research Based Accomplishments in Sweetpotato Whitefly Management**

*Chemical control, action thresholds and resistance management.* In the low desert agricultural areas of the Southwest U.S. the SPW was a year-round pest, successively infesting green-leafy winter vegetables, spring melons, and cotton. Outbreaks in cotton were directly related to rapid population increases in spring melons, and availability of the winter vegetable crops to serve as a haven for the pest during the winter season. Epidemic SPW populations in California and Arizona that characterized the late 1980s through the mid-1990s have not occurred in the growing seasons of 1996 to 2001 as a result of implementing programs based on our increased knowledge of SPW and consequently our ability to develop improved control methods. The early search for stop-gap control methods identified certain chlorinated hydrocarbon, (e.g. endosulfan), organophosphate (e.g. Lorsban and others), pyrethroid (e.g. Capture, Danitol, Asana) and pyrethroid-organophosphate combinations



(e.g. Danitol-Orthene) that were effective. In 1993 imidicloprid, a systemic neonicotinoid insecticide became available for management of SPW in winter vegetables, and in 1994 for melons, as well. Beginning in 1993, programs of conventional insecticides were used to manage SPW, until in 1995, resistance to certain key insecticides, e.g. the pyrethroid plus organophosphate combinations used for whitefly management in cotton occurred (Dennehy et al. 1996a and b, Naranjo et al. 1998).

New systems for SPW were urgently needed for the 1996 season. Adult and immature SPW sampling methods and action thresholds were developed and validated for cotton and melons in the field, and for greenhouse crops. The action thresholds are an integral part of the overall management program and were particularly helpful in reducing superfluous insecticide use, reducing costs, and delaying insecticide resistance development (Naranjo and Flint, 1994, 1995, Naranjo et al. 1995, 1996, Tonhasca et al. 1994a and b). In Arizona, California and Texas cotton, the action thresholds were used with two previously unavailable insect growth regulators (buprofezin and pyriproxyfen). In combination with improvements in cultural control, a program of integrated use of insect growth regulators and insecticides in rotation has effectively managed SPW populations since 1996 (Ellsworth 1999). Because the propensity of SPW to rapidly develop resistance to control measures was recognized, the two newly-available insect growth regulators were introduced in the same year, and the use of each was restricted to one application per year to conserve their efficacy.

Chemical control efficacy has been protected by the implementation of integrated resistance management (IRM) programs (Prabhaker et al. 1996, Castle et al. 1996b, Dennehy et al. 1996). Yellow sticky card, vial and leaf dip techniques have been used in resistance monitoring (Dennehy et al. 1996, Prabhaker et al. 1996). Recommendations for SPW management in Arizona, and more generally in the Southwest, have been reviewed by entomologists from USDA, the cooperating universities, and representatives of the cotton and agrichemical industries at annual meetings since 1996 (Dennehy, unpublished, and Nichols, unpublished, respectively). SPW susceptibility levels to the various modes of action that have been measured differed among geographic locations and in some instances between years at the same location. Some level of tolerance to organophosphates, pyrethroid, carbamate, and chlorinated hydrocarbon insecticides may occur at various locations within the United States (Dennehy and Williams, 1997). It has been rare, however, to attribute control failures to insecticide resistance (Li et al. 2001).

The advances in improved SPW control were not attained without increasing control costs and agricultural community involvement (Henneberry et al. 2000). The

insect growth regulator treatments are costly (about 35 dollars per acre). However, they have been cost effective for cotton with resulting increased yield and quality, and they have replaced far greater numbers of insecticide treatment that were used, at times, ineffectively before their introduction. In Imperial Valley, California, cotton production was reduced from 15,000 acres in 1990-1991 to 4 to 5,000 acres in the early 1990s and has increased to about 8,000 acres in 1998 and 1999 because of the new technology. Similarly, imidicloprid has consistently provided excellent SPW control on melons and vegetables, but costs are \$65 dollars per acre. In California, Arizona and Texas almost all melon acres, of necessity, are treated with imidicloprid to protect them from SPW infestations. Recently buprofezin has been registered for SPW control on melons, thus cross commodity cooperation has been developed to avoid SPW exposure to excessive buprofezin selection pressure in the overlapping melon and cotton growing seasons. Costs associated with administrative coordination, resistance monitoring, and development and dissemination of information on resistance management programs also can be extensive.

In the Imperial Valley, as a result of whitefly populations, spring melon acres were severely reduced in the early 1990s, and continue to be below the average of 25 thousand acres that were grown in the pre-1990s. Melon acres in the Imperial Valley were about 14 thousand acres in 1999. Similarly, fall melons were grown on 10 thousand acres in 1990 and on only 2 thousand acres in 1998-1999. Indirect economic effects of SPW infestations have also occurred through reduced acres in other crops that may not be as obvious. Although new insecticidal chemistries have played a major role in providing SPW control in many of our major cropping systems, high costs may limit use of effective insecticides under some circumstances. For example, because of high prices of certain insecticides, growers of some specialty crops may not be able to take advantage of the new technologies. In more extreme cases, certain crops grown only on a relatively few acres may not have insecticides available to them because the expense of insecticide registration outweighs the anticipated returns. Often in these cases the crop is simply no longer grown commercially in the area where SPW is a problem. Extensive documentation of such cases does not exist, but termination of fall squash production in the Imperial Valley as related to SPW-transmitted squash leaf curl virus was a case in point (Agricultural Commissioner's Office, Imperial Valley, CA, Personal Communication).

In addition, to the development of new insecticide-based management systems, a number of biorational materials that include oils, soaps, and plant products have been shown effective for some crops and may be useful in insecticide rotations to avoid resistance (Stansly et al. 1995).

## Goals for Integrated Areawide Management

As described above, chemical control strategies, use of action thresholds, IRM programs, natural enemy conservation, with consideration for spatial and temporal crop sequencing, early harvest, and crop residue destruction have been highly effective within individual crops, with some benefit to adjacent and sequential crops due to SPW population reduction. Efforts are being made, and further efforts are needed, to expand SPW management systems to incorporate cultural, and biological methods into existing chemically based programs that in the future should include areawide, community efforts.

*Cultural control.* The crops grown, their temporal and spatial relationships, and their respective production inputs and practices are important considerations in SPW population dynamics. Extensive numbers of weed, ornamental, nursery stock, and cultivated crop-host-plants provide nutritional and reproductive host continuity to support SPW population growth. Inter- and intra-host SPW movement occurs during crop growing seasons (Blackmer and Byrne 1995, Byrne and Blackmer 1996). At present strategies for utilizing SPW dispersal in management are limited to grower's awareness of the importance of crop sequencing, planting times and wind direction on SPW dispersal. The proximity of other host crops, when establishing new plantings, is being considered more frequently than in the past because of increased awareness of host-pest relationships and movement. For example, spring melon to summer cotton to fall melon to winter cole crop planting cycles were modified in some areas of the West to eliminate fall melon plantings in order to break the SPW host cycle. An alternative may be to grow fall melons in areas that are distant from existing late-season cotton. Important in respect to crop sequencing is the strict adherence to early harvests of all host crops and destruction and plowdown of crop residues as soon as possible. In areas, where applicable, growers and urban community residents have participated in destruction of weed hosts to reduce another source of migrating SPW. Although, these efforts have not been quantified in relation to effects on SPW populations, they represent good farm practice, and growers have been encouraged to implement them.

*Avoidance - Water management and exclusion techniques.* Higher SPW populations occur in water-stressed compared to well-watered cotton (Flint et al. 1996). Irrigation schedules that effectively avoid water-stress have been recommended and are largely implemented in Arizona cotton. Also, overhead sprinkler irrigation in melon plantings appears to adversely affect SPW population development. However, overhead irrigation is not, at present, used commercially (Castle et al. 1996a). Row covers to protect plants from SPW, and other insect exclusion and reflective materials for repelling SPW have been adopted and are partially

effective in some cropping systems (Chu and Henneberry 1994).

*Natural enemies.* Biological control is expected to play important roles in long-term management of SPW populations. High levels of indigenous natural enemy activity have been identified in cotton, vegetable, and peanut ecosystems, suggesting that natural enemy augmentation and conservation approaches may be important avenues for exploitation in SPW management in the future (Hoelmer 1995, Nordlund and Legaspi 1996). Improvements in conservation have been achieved through use of selective chemistries in cotton. (Naranjo 2001). Part of the efficacy of insect growth regulator use in cotton is attributable to conservation of SPW predators (Ellsworth and Martinez-Carrillo 2001). Foreign explorations have resulted in collection of numerous exotic parasite and predator species that are being considered as potential biocontrol agents (Kirk et al. 2001). Release of some exotic parasitoid and predator species have been accomplished with variable results (Hoelmer 1995). Monitoring is continuing to identify and verify establishment and impact on SPW populations. For predators, a monoclonal antibody has been developed to quantify the role of predation on SPW population regulation as well as to determine candidate predator potential (Hagler and Naranjo 1994). Initial results show positive predation for several indigenous predator species. Natural enemy efficacy, ecosystem compatibility, habitat adaptability, and other factors are being studied to assure optimum utilization of indigenous and introduced parasites and predators. Progress in developing the potential of microbial control is also progressing with the identification of numerous indigenous and exotic pathogenic fungi attacking SLW (Lacey et al. 1996). Strains of *Paecilomyces fumosoroseus* and *Beauveria bassiana* have been identified as promising biological control agents (Akey and Henneberry 1994).

*Host Plant Resistance Factors.* Plant resistance to several SPW-vectored-virus diseases has been identified with resistance to tomato mottle (TMV) a prominent example. Germplasm resistance to SPW has been identified in alfalfa, peanut, melon, cotton, broccoli, collard and tomato. The hirsute leaf character has been recognized to support higher whitefly populations than smooth-leaf types (Mound 1965) and verified by many scientists. Host plant preferences for melons, cotton, broccoli and lettuce appear related to the amount of vascular tissue per unit of leaf area and the proximity of vascular bundles to leaf surfaces (Chu et al. 1995). Also, leaf surface morphology has been shown to play an important role in SPW nymph establishment (Cohen et al. 1996). Studies on plant-insect interactions, physiological disorders and their mechanisms of resistance and the modes of action of whitefly vectored and of disease resistance provide leads for scientists to identify resistant germplasm for incorporation into agronomic types.

### *Viruses and Virus Vectors*

New virus-induced plant diseases transmitted by *Bemisia*, have been observed to be on the increase. In Southern California a new SPW-transmitted geminivirus affecting cucurbits was found in 1998 and also recently detected in Arizona (E. Natwick, Univ. CA Farm Advisor, Holtville, CA; Personal Communication). The long-term impact is unknown, but past experience suggests that the situation must be monitored closely. SPW populations in Texas have also declined from epidemic levels that were experienced in the early to mid 1990s. However, cantaloupe and honeydew melons in the lower Rio Grande Valley have been identified as having SPW-transmitted Cucurbit Yellow Stunting Disorder Virus (CYSDV) disease (Liu et al. 2000). The occurrence of the closterovirus is of much concern to melon growers and its spread, persistence and economic impact will be followed closely. Economic losses have already been experienced. In Florida, the SPW virus vector activity is also being experienced (Polston 1999). Symptoms characteristic of Tomato Yellow Leaf Curl Virus (TYLCV) were first observed on a few tomato plants in the field and nurseries in 1997. The virus probably entered the U.S. in 1996 or 1997 and was rapidly distributed via retail garden centers around the state. Regulatory procedures, that are costly, as well as field management practices have been implemented to minimize the movement of the virus. Even so, TYLCV has spread throughout the state since its first identification. Yield losses have been experienced as well as increased production costs. Pesticide applications to minimize SPW populations have increased. New regulations have been imposed on transplant producers of known TYLCV host plants, which include lisianthus (*Eustoma grandiflorum*), tobacco (*Nicotiana tabacum*) and tomato, to minimize the occurrence of TYLCV in certified transplants. TYLCV has been demonstrated to have a devastating effect on tomato yield particularly when plants are infected in early stages of development. The long-term impact on tomato production in Florida remains unknown.

*Extension and education.* The role of information delivery systems, training and technology transfer cannot be overestimated in implementation of SPW management programs and their success (Kopp et al. 1995). Integration of risk assessment information, spatial analysis, geographic information systems, communications networking, ecological modeling, and extension programs are continually improving and could provide timely information to producers for implementation of improved practices with increasing efficiency. An invaluable service to the plans was the development of an extensive *Bemisia* spp. bibliography first published in the 1995 Supplement to the 5-Year National Research and Action Plan (Butler et al. 1995). The bibliography was updated in supplements thereafter (Naranjo et al. 1996, 1997, 1998, 1999, 2000, and 2002 [this issue]).

*Summary.* The goal of integrating chemical control with resistance management, crop sequencing and host-free periods, crop residue and weed destruction, SPW population and plant disease monitoring, and other cultural controls and management options is considered a high priority for future research. Additional tools such as descriptive models and geographical information systems are envisioned as components of large areawide programs. In most areas with SPW problems, steps are being taken to organize these approaches into coordinated community-action programs. Such programs have proven effective where they have been developed (Ellsworth 1998, 1999, Ellsworth and Naranjo 1999, Ellsworth et al. 1996). SPW and disease resistant crop plants, natural products and microbial insecticides, natural enemy introduction and augmentation have potential, and in some instances have been incorporated into SPW management systems when they become available.

Currently, effective SPW management has been accomplished with (1) selection of non-SPW preferred cultivars, (2) selected spatial and temporal modification of sequential crop systems, (3) intensive sampling and monitoring of whitefly populations, (4) chemical control focused on natural enemy conservation, established action thresholds, alternating chemical modes of action, and resistance monitoring, (5) optimum crop yield goals, allowing for early harvests and destruction of crop residues, and (6) active education and extension outreach to provide timely communication of new developments, SPW population dynamics, and other pertinent information to growers. Not all of the management components are applicable or used in all areas or for all crops, but are general principles that provide the agricultural community options for consideration in SPW management. Additionally, the systems remain open-ended and receptive to other compatible SPW management components.

### **References**

Akey, D.H. 1992. Protocols for ground application of chemical trials against sweetpotato whitefly (SPW) in the 1992 growing seasons, as established by the SPW workshop at San Antonio, TX, January 23-24, 1992, revised at SPW workshop at Houston, TX, February 18, 1992, March 12 and 23, 1992. In R. M. Faust [eds.] Conference report and 5-year national research and action plan for development of management and control methodology for the sweetpotato whitefly, pages 84-101. USDA-ARS, ARS-107, National Technical Information Service, Springfield, VA.

- Akey, D.H. and Henneberry, T.J. 1994. Sweetpotato whitefly control by *Naturalis*-L., the fungus *Beauveria bassiana* in furrow and sub-drip irrigated upland cotton. In D. J. Herber and D. A. Richter (editors) proceedings of the Beltwide Cotton Conferences, pages 1089-1091. National Cotton Council, Memphis, TN.
- Allen, R.M., H. Tucker, and R.A. Nelson. 1960. Leaf crumple disease of cotton in Arizona. Plant Disease Reporter 44: 246-250.
- Anonymous. 1992. Appendix F Sweetpotato Whitefly Ad Hoc Working Group Meeting, October 24-25, 1991, Atlanta, Georgia, pp. 132-134. In R.M. Faust [ed.].
- Barr, C.L. and B.M. Drees. 1992. The poinsettia strain of sweetpotato whitefly. Texas Nurseryman 23: 8-12.
- Bellows, T.S. Jr., T.M. Perring, R.J. Gill, and H. Headrick. 1994. Description of a Species of *Bemisia* (Homoptera: Aleyrodidae). Ann. Entomol. Soc. Of Am. 87 (2): 195-206.
- Bethke, J.A. and T.D. Paine. 1991. Comparative biology, morphometrics, and development of two populations of *Bemisia tabaci* (Homoptera: Aleyrodidae) on cotton and poinsettia. Ann. Entomol. Soc. Am. 84 (4): 407-411.
- Bharanthan, N. W.R. Graves, N.R. Narayanan, D.J. Schuster, H.H. Bryan, and R.T. McMillan Jr. 1990. Association of double-stranded RNA with whitefly-mediated silvering in squash. Plant Path. 39:530-538.
- Blackmer, J.L. and D.N. Byrne. 1995. Behavioral, morphological and physiological traits of migrating *Bemisia tabaci*. J. Insect Physiol. 8: 251-267.
- Brown, J.K. and M.R. Nelson. 1984. Geminate particles associated with cotton leaf crumple disease in Arizona. Phytopathol. 74: 987-990.
- Brown, J.K. and M.R. Nelson. 1986. Host range study of the cotton leaf crumple virus. AZ Agric. Exp. Stn., P 63: 171-176.
- Butler, G.D., Jr. and T.J. Henneberry. 1984. *Bemisia tabaci* as a cotton pest in the desert cotton-growing areas of the Southwestern United States. Proceedings Beltwide Cotton Production and Research Conference, 195-197.
- Butler, G.D., Jr. and T.J. Henneberry. 1985. *Bemisia tabaci* (Gennadius) a Pest of Cotton in the Southwestern United States. United States Department of Agriculture, Agricultural Research Service Tech. Bull. 1701. 19 pp.
- Butler, G.D., Jr., T.J. Henneberry, and W.D. Hutchinson. 1986. Biology, sampling, and population dynamics of *Bemisia tabaci*. In G. E. Russell (ed.) Agric. Zool. Rev. Intercept Ltd, pages 167-195. Pontland, Newcastle upon Tyne, England.
- Butler, G.D., Jr., S.E. Naranjo, T.J. Henneberry and J.K. Brown. 1995. Bibliography of *Bemisia tabaci*. In T.J. Henneberry, N.C. Toscano, R.M. Faust, J.R. Coppedge [eds.], 1995. Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-year National Research and Action Plan - Third Annual Review U.S. Dept. Agric., Agric. Res. Serv, 1995-2, 305 pp.
- Byrne, D.N., T.S. Bellows, and M.P. Parrella. 1990. Whiteflies in agricultural systems. pp. 227-261. In D. Gerling ed. Whiteflies: their bionomics, pest status, and management. Athenaeum Press, Newcastle upon Tyne. U.K. 348 pp.
- Byrne, D.N. and J.L. Blackmer. 1996. Examination of short-range migration by *Bemisia*. In D. Gerling and R.T. Mayer [eds.], *Bemisia*: 1995 Taxonomy, Biology, Damage, Control and Management. Intercept, U.K., pp 17-28.
- Castle, S.J., T.J. Henneberry, N. Prabhaker, and N.C. Toscano. 1996a. Trends in relative susceptibilities of whiteflies to insecticides through the cotton season in the Imperial Valley, CA. In P. Dugger and D. Richter [eds.], Proceedings Beltwide Cotton Conference. National Cotton Council, Memphis, TN, pp. 1032-1035.
- Castle, S.J., T.J. Henneberry, and N.C. Toscano. 1996b. Suppression of *Bemisia tabaci* (Homoptera: Aleyrodidae) infestations in cantaloupe and cotton with sprinkler irrigation. Crop Prot. 15(7): 657-663.
- Chu, C.C. and T.J. Henneberry. 1994. The effect of reflective plastic mulches and insecticides on silverleaf whitefly and broccoli production, pp.137-146. In R. Terry Jones (ed.) Proc. Am. Soc for Plasticulture, Univ. of Kentucky, Lexington, KY.
- Chu, C.C., T.J. Henneberry, and A.C. Cohen. 1995. *Bemisia argentifolii* (Homoptera: Aleyrodidae): Host preference and factors affecting oviposition and feeding site preference. Environ. Entomol. 24: 354-360.
- Cohen, A.C., C.C. Chu, T.J. Henneberry, T. Freeman, J. Buckner, and D. Nelson. 1996. Cotton leaf surface features serve as behavioral cues to silverleaf whiteflies. Southwest Entomol. 21: 377-385.
- Costa, H.S., D.E. Ullman, M.W. Johnson, B.E. Tabashnik. 1993. Squash silverleaf symptoms induced by immature, but not the adult, *Bemisia tabaci*. Phytopathology 83: 763-766.
- Dennehy, T.J., L. Williams, III, J.S. Russell, X. Li, M. Wigert. 1996a. Monitoring and management of whitefly resistance to insecticides in Arizona. In P. Dugger and D. Richter [eds.], Proceedings Beltwide Cotton Conference. National Cotton Council, Memphis, TN, pp. 743-748.

- Dennehy, T.J., P.C. Ellsworth, and R.L. Nichols. 1996b. The 1996 whitefly resistance management program for Arizona cotton. A strategy formulated and revised annually by the southwest whitefly resistance working group. Cooperative Extension, IPM Series 8, 16 pp. University of Arizona, College of Agriculture, Tucson, AZ.
- Dennehy, T.J. and L. Williams, III. 1997. Management of resistance in *Bemisia* in Arizona cotton. *Pest. Sci.* 51:398-406.
- Dickson, R.C., M. McD Johnson, and E.F. Laird., Jr. 1954. Leaf crumple, a virus disease of cotton. *Phytopathol.* 44: 479-480.
- Duffus, J.E. and R.A. Flock. 1982. Whitefly-transmitted disease complex of the desert southwest. *Calif. Agric.* 36: 4-6.
- Duffus, J.E., R.C. Larsen, and H.Y. Lieu. 1986. Lettuce infectious yellows virus -- A new type of whitefly-transmitted virus. *Phytopathol.* 76: 97-100.
- Ellsworth, P.C. 1998. Whitefly management in Arizona: Looking at whole systems. *In* P. Dugger and D. Richter [eds.], *Proceedings Beltwide Cotton Conferences, National Cotton Council, Memphis, TN*, pp. 65-68.
- Ellsworth, P.C. 1999. Whitefly management in Arizona cotton – status and needs, pp. 41-44. *In* P. Dugger and D. Richter [eds.], *Proc. Beltwide Cotton Production Res. Conference, National Cotton Council, Memphis, TN*.
- Ellsworth, P.C. and J. L. Martinez-Carrillo. 2001. IPM for *Bemisia tabaci*: A case study from North America *Crop Protection* 20: 853-869.
- Ellsworth, P.C. and S.E. Naranjo. 1999. Whitefly management with insect growth regulators and the influence of *Lygus* controls. *AZ Agric. Expt. Stn. P-116*: 339-354.
- Ellsworth, P.C., D.H. Akey, L. Williams, T.J. Dennehy, I.W. Kirk, J.B. Carlton, T.J. Henneberry, J.R. Coppedge, and J.W. Diehl. 1996. Understanding whitefly control: Threshold, insecticide rotation, & ground and air comparisons, pp 133-135. *In* P. Dugger, and D. Richter [eds.], *proceedings of the Beltwide Cotton Conference, National Cotton Council, Memphis, TN*.
- Erwin, D.C. and R. Meyer. 1961. Symptomology of the leaf crumple disease in several species and varieties of *Gossypium* and variations of the causal virus. *Phytopathol.* 51: 472-477.
- Flint, H. M., S. E. Naranjo, J. E. Leggett, and T. J. Henneberry. 1996. Cotton water stress, arthropod dynamics, and management of *Bemisia tabaci* (Homoptera: Aleyrodidae). *J. Econ. Entomo.* 89: 1288-1300.
- Gerling, D. 1967. Bionomics of the whitefly parasite complex associated with cotton in southern California (Homoptera: Aleyrodidae; Hymenoptera: Aphelinidae). *Ann. Entomol. Soc. Am.* 59: 142-144.
- Hagler, J.R. and S.E. Naranjo. 1994. Determining the frequency of Heteropteran predation on sweetpotato whitefly and pink bollworm using multiple ELISAs. *Entomol. Zexp. Appl.* 72: 59-66.
- Henneberry, T.J., N.C. Toscano, R.M. Faust, J.R. Coppedge. [eds.], 1993. Sweetpotato Whitefly: 1993 Supplement to the Five-year National Research and Action Plan - First Annual Review. U.S. Dept. Agric., Agric. Res. Serv. No. 112, 175 pp.
- Henneberry, T.J., N.C. Toscano, R.M. Faust, J.R. Coppedge [eds.], 1994. Silverleaf whitefly (formerly sweetpotato whitefly Strain B) 1994 Supplement to the 5-Year National Research and Action Plan - Second Annual Review U.S. Dept. Agric., Agric. Res. Serv., ARS-125, 224 pp.
- Henneberry, T.J., N.C. Toscano, R.M. Faust, J.R. Coppedge [eds.], 1995. Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-year National Research and Action Plan - Third Annual Review U.S. Dept. Agric., Agric. Res. Serv., 1995-2, 305 pp.
- Henneberry, T.J., N.C. Toscano, R.M. Faust, J.R. Coppedge. [eds.], 1996. Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-year National Research and Action Plan - Fourth Annual Review U.S. Dept. Agric., Agric. Res. Serv., 1996-01, 243 pp.
- Henneberry, T.J., N.C. Toscano, T.M. Perring, R.M. Faust [eds.], 1997. Silverleaf Whitefly, 1997 Supplement to the Five-year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001), Fifth Annual Review, U.S. Dept. Agric., Agric. Res. Serv., 1997-02, 272 pp.
- Henneberry, T.J., N.C. Toscano, T.M. Perring, R.M. Faust [eds.], 1998. Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001 (Formerly Sweetpotato Whitefly, Strain B): First Annual Review of the Second 5-Year Plan, U.S. Dept. Agric., Agric. Res. Serv., 1998-01, 187 pp.
- Henneberry, T.J., N.C. Toscano, T.M. Perring, R.M. Faust [eds.], 1999. Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan: Second Annual Review of the Second 5-Year Plan, U.S. Dept. Agric., Agric. Res. Serv., 1999-01, 185 pp.

- Henneberry, T.J., W.A. Jones, T.M. Perring, R.M. Faust [eds.], 2000. Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan (Formerly Sweetpotato Whitefly, Strain B): Third Annual Review of the Second 5-Year Plan, U.S. Dept. Agric., Agric. Res. Serv, July 2000, 209 pp.
- Hoelmer, K.A. 1995. Whitefly parasitoids for the control of field populations of *Bemisia*, pp. 451-476. In D. Gerling and R.T. Mayer [eds.], *Bemisia*: 1995 Taxonomy, Biology, Damage, Control and Management. Intercept, U.K.,
- Johnson, M.W., N.C. Toscano, H.T. Reynolds, E.S. Sylvester, K.R. Kido, and E.T. Natwick. 1982. Whiteflies cause problems for Southern California growers. *California Agric.* 9-10: 24-26.
- Kirk, A.A., L.A. Lacey, and J.A. Goolsby. 2001. Classical biological control of *Bemisia* and successful integration of management strategies in the United States, pp. 309-329. In K.E. Harris, O.P. Smith and J.E. Duffus [eds.]. *Virus-Insect-Plant Interactions*, Academic Press, New York, NY.
- Kopp, D.D., D.A. Nordlund and J. Norman. 1995. Section F: Integrated Techniques, Approaches and Philosophies. In T. J. Henneberry, N. C. Toscano, R. M. Faust, J. R. Coppedge [eds.], 1995. Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-year National Research and Action Plan - Third Annual Review U.S. Dept. Agric., Agric. Res. Serv, 1995-2, 305 pp.
- Lacey, L.A., J.J. Fransen, R. Carruthers. 1996. Global distribution of naturally occurring fungi of *Bemisia*, their biologies and use as biological control agents. In D. Gerling and R.T. Mayer [eds.], *Bemisia*: 1995 Taxonomy, Biology, Damage, Control and Management. Intercept, U.K., pp. 401-433.
- Laird, E.F., Jr. and R.C. Dickson. 1959. Insect transmission of the leaf-crumple virus of cotton. *Phytopathol.* 49: 366-376.
- Li, A.Y-S, T.J. Dennehy, S. X-H Li, M.E. Wigert and M. Zaborac. 2001. Sustaining Arizona's fragile success in whitefly resistance management, pp. 1108-1114. In P. Dugger and D. Richter [eds.], *Proc. Beltwide Cotton Conference*. National Cotton Council, Memphis, TN.
- Liu, T.-X., K. Crosby, M. Miller, L. Gregg and R. Hernandez. 2000. Cucurbit yellow starting disorder virus on melon in the Lower Rio Grande Valley of Texas: Blame to *Bemisia*? pp. 42. In T.J. Henneberry, W.A. Jones, T.M. Perring, and R.M. Faust [eds.] Silverleaf whitefly National Research, Action and Technology Transfer Plan 1997-2001: Third Annual Review of the Second 5-Year Plan. USDA Agric. Res. Serv. 2000, Nat'l Tech. Info. Serv. Springfield, VA.
- Mound, L.A. 1965. Effect of leaf hair on cotton whitefly populations in the Sudan Gezira. *Empire Cotton Growing Review* 42: 33-40.
- Naranjo, S.E. 2001. Conservation and evaluation of natural enemies in IPM systems for *Bemisia tabaci*. *Crop Protection* 20: 835-852.
- Naranjo, S.E., and H.M. Flint. 1994. Spatial distribution of preimaginal *Bemisia tabaci* (Homoptera: Aleyrodidae) in cotton and development of fixed-precision, sequential sampling plans. *Environ. Entomol.* 23: 254-266.
- Naranjo, S.E. and H.M. Flint. 1995. Spatial distribution of adult *Bemisia tabaci* (Homoptera: Aleyrodidae) in cotton and development and validation of fixed-precision, sequential sampling plans for estimating population density. *Environ. Entomol.* 24: 261-270.
- Naranjo, S.E., H.M. Flint, T.J. Henneberry. 1995. Comparative analysis of selected sampling methods for adult *Bemisia tabaci* (Homoptera: Aleyrodidae) in cotton. *J. Econ. Entomol.* 88, 1666-1678.
- Naranjo, S.E., G.D. Butler, Jr. and T.J. Henneberry. 1996. Bibliography of *Bemisia tabaci*, pp. 189-200. In T.J. Henneberry, N.C. Toscano, R.M. Faust, J.R. Coppedge, J.R. [eds.], 1996. Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-year National Research and Action Plan - Fourth Annual Review U.S. Dept. Agric., Agric. Res. Serv, 1996-01, 243 pp.
- Naranjo, S.E., H.M. Flint, T.J. Henneberry. 1996. Binomial sampling plans for estimating and classifying population density of adult *Bemisia tabaci* in cotton. *Entomol. Exp. Appl.* 80, 343-353.
- Naranjo, S.E., G.D. Butler, Jr. and T.J. Henneberry. 1997. Bibliography of *Bemisia tabaci*, pp. 220-238. In T.J. Henneberry, N.C. Toscano, T.M. Perring and R.M. Faust [eds.], 1997 Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): Supplement to the 5-year National Research and Action Plan - Fifth Annual Review U.S. Dept. Agric., Agric. Res. Serv, 1997-02, 272 pp.
- Naranjo, S.E., G.D. Butler, Jr. and T.J. Henneberry. 1998. Bibliography of *Bemisia tabaci*, pp. 119-137. In T.J. Henneberry, N.C. Toscano, T.M. Perring and R.M. Faust [eds.], 1998 Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): Supplement to the 5-year National Research and Action Plan - First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., 1998-02, 187 pp.
- Naranjo, S.E., G.D. Butler, Jr. and T.J. Henneberry. 1999. Bibliography of *Bemisia tabaci*, pp. 133-154. In T.J. Henneberry, N.C. Toscano, T.M. Perring and R.M. Faust [eds.], Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan: Second Annual Review of the Second 5-Year Plan, U.S. Dept. Agric., Agric. Res. Serv, 1999-01, 185 pp.

- Naranjo, S.E., G.D. Butler, Jr. and T.J. Henneberry. 2000. Bibliography of *Bemisia tabaci*, pp. 151-170. In T.J. Henneberry, N.C. Toscano, T.M. Perring and R.M. Faust [eds.], Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan (Formerly Sweetpotato Whitefly, Strain B): Third Annual Review of the Second 5-Year Plan, U.S. Dept. Agric., Agric. Res. Serv. July 2000, 209 pp.
- Naranjo, S. E., P.C. Ellsworth, C. C. Chu, T. J. Henneberry, D. G. Riley, T. F. Watson and R. L. Nichols. 1998. Action thresholds for management of *Bemisia tabaci* (Homoptera: Aleyrodidae) in cotton. J. Econ. Entomol. 91: 1415-1426.
- Nordlund, D.A. and J.C. Legaspi. 1996. Whitefly predators and their potential for use in biological control, pp. 499-513. In D. Gerling, and R. T. Mayer [eds.] "*Bemisia*: 1995 Taxonomy, Biology, Damage, Control and Management.", Intercept Ltd., Andover, Hants, UK.
- Oetting, R.D. and C.D. Buntin. 1996. *Bemisia* damage expressed in commercial greenhouse production, pp. 201-208. In D. Gerling and R.T. Mayer, [eds.], *Bemisia*: 1995 Taxonomy, Biology, Damage, Control and Management. Intercept, U.K.
- Polston, J.E. 1999. The appearance of tomato yellow leaf curl virus (Geminiviridae, Begomovirus) in Florida, pp. 2. In T.J. Henneberry, N.C. Toscano, T.M. Perring, R.M. Faust [eds.] Silverleaf Whitefly National Research Action and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. USDA Agric. Res. Serv. 1999-01. Nat'l Tech. Info. Serv., Springfield, VA.
- Prabhaker, N., N.C. Toscano, T.J. Henneberry, S.J. Castle, D. Weddle. 1996. Assessment of two bioassay techniques for resistance monitoring of silverleaf whitefly (Homoptera: Aleyrodidae) in California. J. Econ. Entomol. 89: 805-815.
- Riley, D.G. and A.N. Sparks, Jr. 1993. Management of the sweetpotato whitefly in the Lower Rio Grande Valley of Texas. Texas A & M Extension Bull. B-5082.
- Russell, L.M. 1957. Synonyms of *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae). Bull. of the Brooklyn Entomol. Soc. 52: 122-123.
- Russell, L.M. 1975. Collection records of *Bemisia tabaci* (Gennadius) in the United States (Homoptera: Homoptera: Aleyrodidae) USDA Coop. Econ. Insect Report 25: 229-230.
- Stansly, P.A., T-X. Lieu, D.J. Schuster, and D.E. Dean. 1995. Role of biorational insecticides in management of *Bemisia*. In D. Gerling and R. T. Mayer (editors) *Bemisia*: 1995 Taxonomy, Biology, Damage, Control and Management, pages 605-615. Intercept Lmtd., U.K.
- Tonhasca, A., J.C. Palumbo, D.N. Byrne. 1994a. Binomial sampling plans for estimating *Bemisia tabaci* populations in cantaloupes. Res. Popul. Ecol. 36: 159-164.
- Tonhasca, A., J.C. Palumbo, D.N. Byrne. 1994b. Distribution patterns of *Bemisia tabaci* (Homoptera: Aleyrodidae) in cantaloupe fields in Arizona. Environ. Entomol. 23: 949-954.
- USDA. 1992. Conference report and 5-year national research and action plan for development of management and control methodology for the silverleaf whitefly. USDA-ARS-107, 165 pp. National Tech. Info. Service, Springfield, VA
- van Schaik, P.A., D.C. Erwin, and M.J. Garber. 1962. Effects of time of symptom expression of the leaf-crumple virus on yield and quality of fiber of cotton. Crop Sci. 2: 275-277.
- Yokomi, R.K., K.A. Hoelmer, and L.S. Osborne. 1990. Relationship between the sweetpotato whitefly and the squash silverleaf disorder. Phytopathology 80:895-900.

**Table 1. Numbers of Research Abstracts published in 1993 to 2002 Sweetpotato Whitefly Strain B National Research and Action Plans Annual Progress Reviews.**

Agency <sup>c</sup> /State	Research Priorities <sup>a</sup>						Total
	A	B	C	D	E	F	
National Research and Action Plan for the Development of Management and Control Methodology for the Sweetpotato Whitefly, 1993-1997 <sup>a</sup>							
1993 Review, Tempe, AZ							
APHIS	-	-	1	1	-	1	3
ARS	7	11	16	13	6	-	53
AZ	2	4	3	1	-	2	12
CA	4	3	4	2	3	-	16
FL	2	3	2	2	2	-	11
GA	-	-	5	-	2	-	7
NY	1	-	1	1	-	-	3
OH	-	-	1	1	-	-	2
TX	1	1	2	-	2	3	9
OTHERS	-	-	1	-	1	-	2
TOTAL	17	22	36	21	16	6	118
1994 Review, Orlando, FL							
ADA	-	-	1	-	-	-	1
APHIS	-	-	-	3	-	-	3
ARS	7	14	13	10	5	1	50
AZ	7	4	5	4	2	3	25
CA	4	5	13	6	3	1	32
CDFFA	-	-	-	2	-	-	2
FL	-	3	5	3	2	2	15
GA	-	-	1	-	-	-	1
HI	1	1	-	-	-	-	2
SC	-	1	-	-	-	-	1
TX	1	-	1	2	1	-	5
WI	-	2	-	-	-	-	2
OTHERS	1	-	4	2	-	-	7
TOTAL	21	30	43	32	13	7	146
1995 Review, San Diego, CA							
APHIS	1	1	-	6	-	-	8
ARS	4	4	19	12	4	-	43
AZ, ASU	8	5	5	1	-	2	21
CA	3	7	7	7	2	4	30
CDFFA	-	-	-	2	-	-	2
FL	1	2	1	4	1	-	9
GA	-	-	1	-	-	-	1
HI	-	1	-	1	-	-	2
KY	-	-	-	-	1	-	1
NM	1	-	-	-	-	-	1
TX	1	-	2	1	1	1	6
WI	-	1	1	-	-	-	2
OTHERS	-	3	6	7	2	2	20
TOTAL	19	24	42	41	11	9	146



1996 Review, San Antonio, TX

APHIS	2	-	-	12	-	-	14
ARS	3	19	16	19	7	1	66
AZ, ASU	4	6	10	1	-	4	25
CA	3	2	12	1	-	-	18
CDFR	-	-	-	3	-	3	6
FL	-	1	2	5	5	1	14
GA	-	-	1	-	-	-	1
TX	-	3	-	1	2	-	6
OTHERS	-	-	6	8	1	1	16
TOTAL	12	31	47	50	15	10	166

Silverleaf Whitefly, National Research, Action and Technology Transfer Plan, 1997-2001<sup>b</sup>

1997 Review, San Diego, CA

APHIS	-	-	-	12	2	-	14
ARS	7	9	7	10	5	2	40
AZ	2	8	2	7	-	2	21
CA	3	7	7	2	1	1	21
CDFR	-	-	-	4	-	-	4
FL	2	-	3	5	1	1	12
GA	-	-	-	-	1	-	1
TX	-	1	1	1	-	-	3
OTHERS	1	-	4	2	1	1	9
TOTAL	15	25	24	43	11	7	125

1998 Review, Charleston, SC<sup>b</sup>

APHIS	-	-	-	11	-	-	13
ARS	12	-	6	13	6	1	38
AZ	-	-	1	-	-	2	3
CA	1	1	7	5	6	3	23
FL	1	1	-	-	1	-	3
GA	-	-	-	-	1	-	1
TX	-	-	-	2	-	-	2
OTHERS	3	-	-	1	1	1	6
TOTAL	17	2	14	32	15	9	89

1999 Review, Albuquerque, NM

APHIS	-	-	-	3	-	6	9
ARS	16	3	6	4	-	-	29
AZ	1	-	-	3	-	4	8
CA	1	1	5	5	6	-	18
FL	-	-	1	-	-	-	1
TX	1	4	5	-	-	-	10
OTHERS	5	1	1	3	3	2	15
TOTAL	24	5	17	23	9	12	90

2000 Review, San Diego, CA

APHIS	-	-	-	2	-	-	2
ARS	15	1	6	7	2	2	33
CA	2	1	5	5	7	1	21
FL	-	1	-	-	-	-	1
TX	-	1	1	1	1	1	5
OTHERS	3	2	-	9	-	3	17
TOTAL	20	6	12	24	10	7	79

2001/2002 Review, San Diego, CA

APHIS	-	-	-	3	-	-	3
ARS	14	3	3	5	3	3	31
AZ	1	2	-	-	-	-	3
CA	-	1	8	2	7	-	18
FL	-	1	4	-	2	1	8
TX	-	-	4	-	1	-	5
OTHERS	11	3	-	2	1	1	18
TOTAL	26	10	19	12	14	5	86

<sup>a</sup> For 1993-1997, A = Ecology, Population Dynamics and Dispersal; B = Fundamental Research, Behavior, Biochemistry, Biotype, Morphology, Physiology, Systematics, Virus Diseases and Vector Interactions; C = Chemical Control, Biorationals, and Pesticide Application; D = Biocontrol; E = Crop Management Systems and Host Plant Resistance, and F = Integrated Techniques, Approaches, and Philosophies.

<sup>b</sup> For 1997-2002, A = Biology, Ecology, and Population Dynamics; B = Viruses, Epidemiology and Virus-Vector Interactions; C = Chemical Control, Biopesticides, Resistance Management, and Application Methods; D = Natural Enemy Ecology, and Biological Control; E = Host-Plant Resistance, Physiological Disorders, and Host Plant Interactions; F = Integrated and Areawide Pest Management Approaches, and Crop Management Systems.

<sup>c</sup> APHIS = USDA, Animal and Plant Health Inspection Service; ARS = USDA, Agricultural Research Service.

**Table 2. Summary of Section A: Ecology, Population Dynamics and Dispersal Research Progress for the 5-Year National Research and Action Plan on Sweetpotato Whitefly Strain B (SPW) (1993-1997).**

Research Approaches	Research Result Highlights
<p><b>Define biology, phenology, and demography of SPW on greenhouse, field crop and wild host plants.</b></p>	<p>An extensive SPW host range of weeds, landscape vegetation and cultivated crops was identified in multistate surveys. Low winter temperatures reduced SPW survival and population development. Dispersal from cucurbits to cotton in the spring to alfalfa in the fall occurred in California and Arizona. In Texas, dispersal from cabbage to melons to cotton was common. SPW movement between sequentially planted crop hosts and naturally occurring weed hosts was evident. SPW population dynamics differed in different localities. Biology differed on different crops and also varied from year-to-year among and between geographical areas. Moisture deficit conditions increased SPW cotton densities in Arizona. Predators and severe wind/rain were important sources of mortality in life table studies. Significant correlations occurred between <i>Orius tristicolor</i> and <i>Chrysoperla carnea</i> and SPW population density. Parasitism was found to be extensive in some ecosystems in surveys in Arizona, California, Texas and Florida, but was not a limiting factor in population development.</p>
<p><b>Develop efficient SPW sampling plans for research and decision making.</b></p>	<p>Vertical population distribution of SPW adults and nymphs within host plants revealed that sampling in the top 20% of the plant provided good population estimates. Leaf disks from the 5th main stem nodes on cotton was the most efficient sample unit for eggs and nymphs. Based on this data, sampling plans for adults (5 to 10 adults/leaf turn) and nymphs (<math>0.3 \pm 0.05</math> nymphs <math>\text{cm}^2</math> of leaf area) were developed and evaluated for cotton and further refined for cantaloupes. The cotton sampling plan was validated in over 8000 grower acres and both plans were adopted by agriculturalists. The intraplant distribution of SPW eggs, nymphs, and pupae were also studied on tomato. Field sampling methods for honeydew and cotton stickiness showed that lint stickiness was uniformly to randomly distributed within cotton fields compared with the highly clumped distribution of SPW nymphs.</p>
<p><b>Develop economic thresholds for SPW in relation to feeding damage, honeydew production and virus transmission.</b></p>	<p>Sticky cotton as a result of honeydew contamination is a major concern in the cotton textile industry. Identification of the sugars in honeydew and subsequent determination of their relationships to whitefly population and stickiness determined by thermodeceptor analyses were major accomplishments. SPW densities were correlated to reduced cotton lint quality. The action threshold for minimizing honeydew contamination of lint and yield protection were found to be between 5 and 10 SPW adults per leaf. In melons, SPW population relationships to melon harvest weights, % sugars, and other crop performance characteristics were established. Action thresholds of 0.5 large nymph per 7.6 <math>\text{cm}^2</math> of leaf area were suggested. Action thresholds for adult whitefly have been incorporated in binomial sampling plans for cotton and melons. Rainfall and sampling variations for lint stickiness confound sampling method efforts.</p>
<p><b>Develop and test population models to describe and predict SPW dynamics.</b></p>	<p>Life process oriented models and more general spatial models were partially developed to identify influences of abiotic and biotic factors on site-specific and regional population dynamics. They can be valuable tools to identify research needs, assess priorities, and aid development of management systems. Model simulation results compared reasonably with trends observed in the sampling data, and were highly sensitive to natural enemy parameters. LandSat data were obtained and areawide models were developed and tested for the Imperial Valley. SPW population data for various crops were linked to cropping data and the effects of various cultural control strategies evaluated with the computer model.</p>

**Determine factors influencing  
SPW dispersal.**

SPW flight was shown to be sustained at temperatures up to 47° C and for 2 h in a small fraction of the population (males longer than females). Diet, sex and age (longest flights 3 to 6 days after adult eclosion) were shown to affect flight performance. Dispersal was influenced by plant phenology, plant amino acid content, abiotic factors (diurnal cycles, temperature, light, wind) and physiological and morphological conditions. Marked and released individuals moved as far as 4.8 km from the release site. Adult mark and recapture studies provided quantitative estimates of dispersal from melons and observational studies showed declining densities of SPW in cotton at progressively greater distances from source melons. Adult trapping studies in Texas indicated overall increases and declines in capture were correlated with increase and decline of the cotton crop.

---

Sources<sup>1</sup>

- Naranjo, S. and H. Browning. 1993. Section A Ecology, Population Dynamics and Dispersal, pp. 22-24. In: T. J. Henneberry, N. C. Toscano, R. M. Faust and J. R. Coppedge [eds.]. Sweetpotato Whitefly: 1993 Supplement to the Five-Year National Research and Action Plan-First Annual Review. U.S. Dept. Agric., Agric. Res. Serv., No. 112, 175 pp.
- Naranjo, S. E. and M. W. Johnson. 1994. Section A Ecology, Population Dynamics and Dispersal, pp. 33-35. In: T. J. Henneberry, N. C. Toscano, R. M. Faust and J. R. Coppedge [eds.]. Silverleaf Whitefly: 1994 Supplement to the Five-Year National Research and Action Plan-Second Annual Review. U.S. Dept. Agric., Agric. Res. Serv., No. 125, 228 pp.
- Johnson, M. W. and L. D. Godfrey. 1995. Section A Ecology, Population Dynamics and Dispersal, pp. 24-25. In: T. J. Henneberry, N. C. Toscano, R. M. Faust and J. R. Coppedge [eds.]. Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the Five-Year National Research and Action Plan-Third Annual Review. U.S. Dept. Agric., Agric. Res. Serv., 1995-02, 305 pp.
- Godfrey, L. D. and D. N. Byrne. 1996. Section A Ecology, Population Dynamics and Dispersal, pp. 14-15. In: T. J. Henneberry, N. C. Toscano, R. M. Faust and J. R. Coppedge [eds.]. Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the Five-Year National Research and Action Plan-Fourth Annual Review. U.S. Dept. Agric., 1996-01, 243 pp.
- Godfrey, L. 1997. Section A Ecology, Population Dynamics and Dispersal, pp. 40-42. In: T. J. Henneberry, N. C. Toscano, T. M. Perring and R. M. Faust [eds.]. Silverleaf Whitefly, 1997 Supplement to the Five-Year National Research and Action Plan-Fifth Annual Review. U.S. Dept. Agric., 1997-02, 272 pp.

<sup>1</sup> Summaries of research progress abstracts were prepared by the indicated authors. See individual abstracts for research credits and more detail.

**Table 2 (Continued): Summary of Research Progress for Section B: Fundamental Research - Behavior, Biochemistry, Biotypes, Morphology, Physiology, Systematics, Virus Diseases, and Virus Vector Interactions of the 5-Year National Research and Action Plan on Sweetpotato Whitefly Strain B (SPW) (1993-1997).**

Research Approaches	Research Result Highlights
<p><b>Studies of feeding behavior: sensory receptors, ultrastructure, morphology, digestive physiology; intra- and interspecific competition.</b></p>	<p>Extensive and detailed information was developed on SPW feeding, digestion, cuticular characterization and morphology. SPW honeydew from cotton contained at least 30 sugars created from sucrose in phloem sap. Trehalulose and sucrose were the stickiest lint contaminants. Treatment of sticky cotton lint with enzymes at harvest removed the stickiness more quickly and efficiently than water treatments alone. A and B biotype SPW wax glands were similar, surface cuticular lipids were characterized. Abundance of vascular bundles in leaf tissues of some crops appeared to affect SPW feeding preference. Microscopic examination of the SPW digestive system indicates that it serves as an organ of disease transmission and inferences can be made concerning food movement within the system, how water balance is regulated and in which tissues viral particles may accumulate. SPW egg stalks had walls of thick multiple layers of fibrous material; cytoplasm or yolk was not seen in the egg stalk. Observations on adults and nymphs indicate modes of stylet penetration, sheath formation, and feeding. Histological study of SPW stylet penetration has correlated host plant microstructure with success of whitefly colonization. Serine proteinase activities were undetectable in whole extracts of whiteflies. Results may expedite breeding of new cultivars of some crop plants. Nymphs target only minor vascular bundles, and stylets of surviving nymphs always reach bundles and phloem cells. Lower surfaces of the youngest leaves of most plant species tested were preferred.</p>
<p><b>Studies of biochemistry, physiology, nutrition, development and reproduction.</b></p>	<p>Rates of water loss (10%), and oxygen consumption for A and B SPW biotypes were similar. Thermal maximum thresholds were about 52°C. SPW survivorship was best on zucchini, intermediate on cabbage and lowest on sugar beet. Development times were longer on cabbage (30 d) than on zucchini (23.1 d). SPW females appear to prefer to oviposit on plants that enhance nymphal development. Host selection studies and host effects on tritrophic interactions have shown definite correlations between host plant and whitefly population levels. Artificial feeding systems were established to evaluate effects of chemical, toxins, and antibiotics on whitefly fecundity, development, and honeydew production, and to study feeding behavior, relative to intake of phloem sap and geminiviruses in phloem sap. One feeding chamber experiment tested semi-synthetic diet presented through a Parafilm membrane and demonstrated an ability to rear whiteflies successfully from egg to adult. Egg hatching rates were 2-50%, and specific dietary constituents were critical to development past third instar. Feeding behavior of immatures was studied using ingestion of fluorescent markers, and feeding structures active during ingestion and processing were video-recorded. Constituents of phloem sap ingested by whiteflies and composition of honeydew were studied; certain amino acids in phloem sap were in reduced forms, suggesting host amino acids are metabolized in whiteflies. Radiolabeled plant proteins were found to be taken up and converted by whiteflies, and radiolabel was found in new proteins and secreted into plants. Toxins from microorganisms were assayed against adult whiteflies using a new assay.</p>

**Studies to discover and analyze diagnostic characteristics of SPW, including component taxa, and to determine biological and genetic basis for development of biotypes, host races, and species, genetics and genetic diversity. Develop dsRNA and cDNA probe.**

DNA analyses showed differences between species and A and B SPW biotypes. Esterase analyses of SPW showed the B biotype is widely distributed worldwide. ACHE enzyme systems in A and B are different. rRNA analysis of A and B types suggest they are conspecific. Isoelectric focusing techniques used to analyze SPW from various locations in US, Egypt, Mexico and Spain identified A and B type. A, B and Hybrids have been reported. Unique dsRna was associated with the B-biotype." RAPDs analyses of DNA from a collection of *B. tabaci* indicates genetic variability between populations and that RAPDs were not useful markers for assigning phylogenetic associations. Direct genetic analysis of ribosomal (nuclear) and mitochondrial DNA of the *B. tabaci* complex was undertaken. The B biotype was proposed as a new species, *Bemisia argentifolii*, but key morphological characters to distinguish the A and B biotypes, or other biotypes are still lacking. Morphological plasticity and data from molecular studies indicate the most likely classification for the group is as a *B. tabaci* species complex. Some morphological plasticity stems from stimuli received by crawlers based in plant geography, hairiness, and crowded conditions on the leaf. Three whitefly complexes in the genus *Bemisia* have been defined based on classical taxonomic criteria and tentatively corroborated with molecular sequencing data. Molecular markers are under investigation for inference of phylogenies that can be compared to those predicted by morphometric and other methods. Successful diagnostics for natural enemy (especially parasitoid wasp) identification were identified. Many species were differentiated within the genera, *Encarsia* and *Eretmocerus*, using genetic fingerprinting by RAPD-PCR.

**Develop systematic analysis of the genus *Bemisia* utilizing various methods.**

Ultrastructural studies of SPW eggs showed that mycetomes and their symbionts form a prominent mass at the distal end of each egg as in aphids and other homopterans. The surface lipids and wax particles of SPW (biotypes A & B) were characterized and no differences were found. The major wax esters were C46 and the major acid and alcohol moieties in the esters were C20 and C26, respectively. Major components of the wax particles were a C34 aldehyde and a C34 alcohol plus small amounts of C32 aldehyde and alcohol. Morphological studies for taxonomic considerations of the A and B biotypes focus on the pupal vasiform orifice, and mapping of setae and pore arrangements. Widely scattered loci of the two strains could not be separated. The submarginal setae, ASMS4, present in the A biotype, in type material from Greece, and in type material from poinsettia is absent from the B biotype 95% of the time. Because the character is not present or absent 100% of the time, it was not useful as a taxonomic character for differentiating the A and B biotypes or sibling species. A 550 bp sequence of whitefly 16S ribosomal mt RNA gene showed promise as an informative molecular marker, based upon phylogenies inferred from several trial datasets. The B biotype, or silverleaf whitefly, clusters with other Old World whitefly sequences in a consensus tree derived from 100 bootstrap samples of rDNA sequences, suggesting the silverleaf whitefly is of Old World origin.

**Identify and define SPW toxicogenic effects.**

Plant growth regulator-induced silvering in cucurbits similar to SPW-induced silvering, but rootmass, chlorophyll and intercellular fluid expression was different. Gibberellic acid overcame PGR-induced symptoms and SPW-induced silvering, suggesting GA modification from SPW feeding. Feeding by biotype B nymphs causes squash silverleaf which is in part the result of the plant responses including autolysis of mesophyll cells at the feeding site. Other responses to B biotype feeding include the appearance of proteins at Mr 31,000 and 70,000 and the suppression of a Mr 60,000 protein in intercellular fluids of sweet sugar pumpkin. Other proteins that were suppressed were detected by 2-dimensional electrophoresis. While precise mechanisms of direct (toxicogenic) effects of whiteflies (especially B biotype) on host plants have not been discovered, some of the phenomena have now been histologically defined and nymphs identified as the source. New approaches to the molecular bases of plant syndromes such as tomato irregular ripening and squash silverleaf were considered. One such method that showed promise was the differential display of messenger RNA. Duration of feeding and distance from fruiting structures affected the severity of tomato irregular ripening. ( $\alpha$ -Tocopherol, squalene, and linolenic acid increased in squash (pumpkin) plants prior to expression of squash silverleaf. Direct toxicogenic effects of the B biotype were expressed as uneven ripening in tomato, silverleaf in squash, stem streaking and thickening in *Brassica*, and paleness of leaves in lettuce and other composites. Pathogenesis-related proteins are typically sequestered in vacuoles or transported out of cells into the intercellular fluid (IF). The IF from silvered leaves of B biotype-infested cucurbits contained induced proteins of 31K and 70K Mr, whereas a characteristically prominent constitutively expressed protein (60K) was shown to be absent.

**Characterize SPW endosymbiont (SPWe) influence on metabolism, host range, and biotype formation.**

Endosymbionts from A- and B-biotype SPW as well as several other whitefly species have been grouped according to ribosomal RNA sequences. Rod-shaped endosymbionts purified from adult SPW were morphologically similar to prokaryotes localized ultrastructurally in the mycetomes of third instar nymphs. Mycetomes in immature *B. tabaci* contain microorganisms which appear to be symbiotic to whiteflies and necessary for survival. Antibiotic treatment reduced mycetome size in offspring produced from treated adults and also reduced the immatures' ability to induce phytotoxic SSL. Ultrastructural studies by TEM indicated at least two morphologically different types of microorganisms. Microscopic analysis of endosymbionts has begun to define specific forms of symbiotic bacteria associated with the mycetomes of *Bemisia*, and to define differences among genetically defined populations. Artificial feeding systems were developed to rear whiteflies from egg to adult for the first time. Fundamental biological studies have also been conducted to evaluate the effects of insecticidal compounds on whiteflies and the transmission pathway of geminiviruses in adult whiteflies in relation to feeding behavior. Feeding chambers have also been utilized in the identification of sugars that contribute to stickiness due to whitefly honeydew. Mycetome-like organs containing prokaryotic-like and pleomorphic microorganisms were observed in immature instars, and at the distal tip of each egg. Transfer of endosymbionts in mycetocytes from the female to developing ova is a process that occurs continuously, prior to oviposition. Mycetocyte-containing bacteria were observed in oocytes inside a common membrane in the region that becomes the pedicel end of the egg. In the final stages of ovum development, the plasma of the oocyte completely surrounds the mycetocyte and the chorion thickens. Oocytes with mycetocytes were present in adult females 16 hr after emergence, and none were observed in individuals that were less than 2 hrs old. Antibiotics known to have a negative impact on prokaryotic processes and metabolism were detrimental to whiteflies, whereas whiteflies appeared to be unaffected by those specifically inhibiting prokaryotic cell membrane synthesis or affecting cell walls. Negative effects were attributed to mortality of the primary endosymbiont in whiteflies.

**Investigate etiology of diseases; biological and molecular characterization of causal agents; develop understanding of relationship; molecular probes for viral diseases; diagnostics and resistance; virus-vector specificity and interactions**

Several SPW-transmitted geminiviruses in Florida have been identified in bean, cabbage, tomato and weed species and compared at the nucleotide level at two sites in the genome. *B. tabaci* mouthparts and associated organs are similar in structure to those found in aphids. The salivary glands and the digestive system have been observed in detailed morphological studies. Diseases of vegetable and ornamental crops due to the whitefly-transmitted geminiviruses were also observed in Hawaii and Yucatan, Mexico. A PCR-based assay was developed to specifically and universally detect subgroup III geminiviruses. PCR primers target the core or central fragment of the coat protein (CP) gene and yield a 550 bp diagnostic product. The products are cloned and sequenced, and incorporated into the geminivirus CP gene sequence database. The approach has facilitated geminivirus identification. Whitefly-transmitted closteroviruses in lettuce was found that was vectored by the B biotype of *B. tabaci*. Artificial feeding chambers were used to study steps in the transmission process in which whiteflies ingest, acquire, and transmit geminiviruses. Geminivirus bNA was detected by PCR in saliva of the whitefly vector for the first time, whereas virus DNA was detectable in whole body extracts and honeydew of both vector and non-vector whiteflies. In Arizona, molecular cloning and characterization were the focus for isolates of cotton leaf crumple, squash leaf curl, and an uncharacterized geminivirus of Solanaceae. Studies were initiated to identify geminiviruses of cotton, kenaf, pepper, squash, tomato, tomatillo, watermelon, and weed species in Texas, and to determine the epidemiology of squash leaf curl in cucurbits. Studies in Florida concentrated on determining experimental and natural host ranges and identification of sources of inoculum for tomato mottle virus. TMoV infects species within the Solanaceae and Leguminosae, and that the most important source of TMoV inoculum is infected tomato fields. TMoV has been cloned and sequenced and has been shown to be related to other New World bipartite WFT geminiviruses. Breeding for virus resistance/tolerance in tomato was initiated with *Lycopersicon chilense* as a potential source of resistance gene(s). Engineered pathogen-derived resistance approaches have made promising strides using expression of *mutant CP, BCJ, and Rep*. An epidemic in bean caused by Macroptilium mosaic geminivirus were reported in Florida for the first time. a recently introduced strain of bean golden mosaic virus from the Caribbean Basin and is distinct from the BGMV -FL strain. A strain of TYLCV, nearly identical to TYLCV-Israel, was found for the first time in The Dominican Republic, Jamaica, and Cuba. A new WFT closterovirus, tomato infectious chlorosis (TICV) was discovered in coastal California tomato greenhouses and is transmitted by the greenhouse whitefly, but not *B. tabaci*. The virus was also identified in tomato greenhouses in Italy.



**Study epidemiological parameters: vector population dynamics; disease thresholds; identify sources of inoculum, distribution, severity, and prevalence of pathogens. Correlate efficiency of transmission with biotypes, diversity and parameters of cropping systems.**

Squash lines screened show variable degrees of susceptibility to SPW silvering. Hybridization profiles and partial host ranges have been generated for over 12 new viruses in tomato and pepper in the U.S. PCR-based methods are being developed to determine virus prevalence in existing cropping systems and to determine virus vector relations. New biotype specific viruses are being characterized (ToMoV, putative new geminivirus in crucifers) and vector efficiency is being evaluated with respect to previously characterized viruses (LIYV). Biotype A transmitted lettuce infectious yellows (LIYV) more effectively than B biotype. Result of reduced transmission of LIYV has been larger yields in certain crops because A biotype populations have been reduced. Whitefly-mediated transmission studies were undertaken to standardize experimental transmission methods for broad-scale use. A PCR technique was developed for universal detection of WFT geminiviruses. PCR primers target the core region (nts 494-1048) of the capsid protein gene, the most highly conserved gene among subgroup 111 viruses. PCR products of diagnostic 550bp size indicate positive infection. Fragments are cloned and DNA sequences obtained to verify identity with sequences from well characterized viruses. A collection of core coat protein gene sequences has been established in a database. This database will be useful for geminivirus identification, molecular epidemiological studies, and for making predictions about phylogenetic relationships between geminivirus isolates. Among isolates collected and analyzed from New World sites, all but one virus, now known to be introduced into the Caribbean Basin from the Eastern Hemisphere, cluster with New World viruses. This suggests that the majority of viruses problematic in the US, Mexico, and Caribbean Basin originated in the New World, and are not introduced viruses. Further, these data extends the hypothesis that recent epidemics in the New World are primarily the result of indigenous viruses that rarely reached epidemic status in the past due to mobilization by new whitefly biotypes and altered disease pressures. Cloning and sequence of a second key region of the genome, the common region, are underway for the same library of geminiviruses for identification and to provide a second molecular marker in a key region of viral genomes. Comparative analysis of core coat protein and common region sequences permitted delineation of (putative) genetically related subclusters within the WFT subgroup, and indicates a wider range of variability within the subgroup than previously thought. Ability to define WFT geminiviruses as constituents of particular genetic subclusters by key viral sequences will permit a better understanding of evolutionary relationships, permit early recognition of new and emerging viruses, and suggests a possible strategy for selecting a range of prototype isolates for classical breeding or engineered resistance efforts.

**Study mating and oviposition behavior.**

A and B biotype mating studies suggest reproductive isolation, but identified hybrid populations have occurred. Attempts to identify a SPW sex pheromone have been unsuccessful. Phosphorus deficiency cotton seedlings resulted in reduced oviposition on young leaves in greenhouse and growth chambers. Host acceptance was associated with low leaf sucrose concentrations (greenhouse), low total sugars and high leaf water (growth chambers) suggesting that factors affecting plant osmotic potential impact cotton susceptibility to SPW. Different whitefly species were shown to have different mating behaviors, and if extended to studies of *B. tabaci* populations has relevance to the (putative) reproductive isolation observed between biotypes. Studies of mating behavior and effects of mating on fecundity, viability, and population structure of whitefly on cotton indicate distinct behaviors for the B biotype and other whitefly genera. Viability of eggs can be much lower from an unfertilized female. Oviposition rate during the first days of adult life is not influenced by mating history. Mating behaviors for three whitefly species were captured on film using a video recorder attached to a binocular microscope. Leaf age and position on the plant influenced oviposition rates. Differences in behavioral patterns and temporal modulation of activities were documented during courtship and copulation. The basis for apparent reproductive isolation between certain populations of *B. tabaci* is not known, and has been another confounding factor in resolving the taxonomic status of *B. tabaci*.

**Determine factors influencing host plant selection and host acceptance.**

Sequential feeding periods on different hosts or exposure to different hosts did not affect SPW fecundity or survival, suggesting no relation to preference. B biotype immatures developed faster than A's on the same host leaves, different host leaves or different cucurbit plants. Host preference was positively correlated to the abundance/length of vascular bundles per unit leaf volume. The preferred habitat is the under side of the leaf and this characteristic may be related to the shorter distance to vascular bundles from the lower compared to the upper leaf surface. Although leaf surface pubescence is an important factor in ovipositional behavior which resulted in females choosing the lower leaves which have fewer leaf hairs for deposition of 90-95% of the eggs laid, negative geotropism was the overriding factor influencing oviposition on lower vs. upper leaves. Acceptability of host plant feeding sites, based on oviposition rates, was greatest on youngest leaves compared to older leaves or cotyledons. High osmotic potential of leaves was positively correlated with adult whitefly acceptability of the host plant. Surface and internal structures of leaves from different host plant species and cultivars were shown to affect immature whitefly morphology and ability to colonize (density of immatures) and feed. The latter is due to variation in density and proximity of available vascular bundles within leaves. SPW is strongly attracted to surfaces reflecting in yellow-green wavelengths at certain times and to blue/UV reflecting surfaces at others. Host acceptance or rejection seems to be largely due to taste response following short leaf probing in the mesophyll. Influence of host and host choice on whitefly fecundity was examined, and at times, eggs were deposited on plants suitable for adult feeding that were not conducive to progeny survival. AC feeding monitors were used to correlate ovipositional activity with adult feeding patterns, feeding patterns of females were apparently more indicative of ovipositional site preferences than adult nutrition.

**Identify plant nutritional and defensive responses to SPW and their effects on SPW and natural enemies**

A minimum of eight different proteins (four different Mr's) were isolated from plants colonized by the 'B' biotype. At least six other proteins of three different Mr's appear to be suppressed by SPW B biotype feeding on sweet sugar pumpkin. Measurements of chitinase activities show an inverse relationship with the length of SPW B biotype feeding time, i.e., the longer the plant is a host for SPW, and the more progressive the silverleaf symptom, the lower the amount of chitinase activity. SPW were shown to be unsuitable prey for some predators, due to nutritional deficiencies. In cucurbits, several natural products (alpha-tocopherol, squalene, and linolenic acid) were analyzed in developing leaves after whiteflies fed on lower leaves. Quantitative changes in these lipids may have nutritional consequences for the whitefly and/-physiological implications for the host plant. Linolenic acid and alpha-tocopherol (vitamin A) may induce host defensive responses and serve as vitamins for whiteflies, while squalene is a precursor of sterols which serve as critical structural components of plant cells and are also required by insects. Studies of protein expression in tomato indicated that several proteins were induced by adult and larval feeding. Though total leaf activities of chitinase, peroxidase, beta-1,3-glucanase, and chitosanase were greater than for controls; two proteins induced within 24 h were identified as chitinase and glucanase

---

**Sources<sup>1</sup>**

- Brown, J. K., D. R. Jimenez, and R. T. Mayer. 1993. Section B Fundamental Research, Behavior, Biochemistry, Biotypes, Morphology, Physiology, Systematics, Virus Diseases and Virus Vector Interactions, pp. 48-51. In: T. J. Henneberry, N. C. Toscano, R. M. Faust and J. R. Coppedge [eds.]. Sweetpotato Whitefly: 1993 Supplement to the Five-Year National Research and Action Plan-First Annual Review. U.S. Dept. Agric., Agric. Res. Serv., No. 112, 175 pp.
- Brown, J. K. and R. T. Mayer. 1994. Section B Fundamental Research, Behavior, Biochemistry, Biotypes, Morphology, Physiology, Systematics, Virus Diseases and Virus Vector Interactions, pp. 71-72. In: T. J. Henneberry, N. C. Toscano, R. M. Faust and J. R. Coppedge [eds.]. Silverleaf Whitefly: 1994 Supplement to the Five-Year National Research and Action Plan-Second Annual Review. U.S. Dept. Agric., Agric. Res. Serv., No. 125, 228 pp.
- Brown, J. K. and J. P. Shapiro. 1995. Section B Fundamental Research, Behavior, Biochemistry, Biotypes, Morphology, Physiology, Systematics, Virus Diseases and Virus Vector Interactions, pp. 52-54. In: T. J. Henneberry, N. C. Toscano, R. M. Faust and J. R. Coppedge [eds.]. Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the Five-Year National Research and Action Plan-Third Annual Review. U.S. Dept. Agric., Agric. Res. Serv., 1995-02, 305 pp.
- Brown, J. K. and J. P. Shapiro. 1996. Section B Fundamental Research, Behavior, Biochemistry, Biotypes, Morphology, Physiology, Systematics, Virus Diseases and Virus Vector Interactions, pp. 50-53. In: T. J. Henneberry, N. C. Toscano, R. M. Faust and J. R. Coppedge [eds.]. Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the Five-Year National Research and Action Plan-Fourth Annual Review. U.S. Dept. Agric., 1996-01, 243 pp.
- Brown, J. K. and J. P. Shapiro. 1997. Section B Fundamental Research, Behavior, Biochemistry, Biotypes, Morphology, Physiology, Systematics, Virus Diseases and Virus Vector Interactions, pp. 43-50. In: T. J. Henneberry, N. C. Toscano, T. M. Perring and R. M. Faust [eds.]. Silverleaf Whitefly, 1997 Supplement to the Five-Year National Research and Action Plan-Fifth Annual Review. U.S. Dept. Agric., 1997-02, 272 pp.

<sup>1</sup> Summaries of research progress abstracts were prepared by the indicated authors. See individual abstracts for research credits and more detail.

**Table 2 (Continued). Summary of Research Progress for Section C: Chemical Control, Biorationals and Pesticide Application Technology of the 5-year National Research and Action Plan for Sweetpotato Whitefly Strain B (SPW) (1993-1997).**

Research Approaches	Research Result Highlights
<b>Identify new chemicals and biopesticides for registration, that selectively control SLWF with minimal effects on beneficial organisms</b>	Greenhouse and field studies in Arizona, Georgia, California, Texas, Florida, Ohio, Maryland, New York and Mississippi identified conventional chemicals alone or in combination that had potential for SPW control on cotton, vegetables and ornamentals. Section 18's were obtained that allowed the emergency use of Admire, Capture, and Danitol in several states. The insect growth regulators, pyriproxyfen and buprofezin were highly effective for control of SPW as a foliar spray on cotton and melons. Their use in Arizona and California resulted in less damage and reduced tolerance to pyrethroids. Chemical control to maintain SPW populations below economic thresholds and avoid economic losses was demonstrated in many commercial crops. Yield responses to insecticidal control were demonstrated in virtually all host crops. Examination of plant derived oils, <i>Nicotiana</i> spp. extracts, neems, a "mycoinsecticide" <i>Beauveria bassiana</i> , and several soaps appeared as useful alternatives in combination with insecticides, in chemical rotational systems, and for special use purposes.
<b>Economic thresholds: Relate schedules and methods of application to economic thresholds.</b>	Tentative economic and action thresholds based on yield vs. SPW density relationships in cotton, melons, cole crops and lettuce were established. Relationships between SPW and sticky cotton were investigated in California, Arizona and Texas. Similarly, significant relationships between SPW adult and immature densities were established on cantaloupe yield and quality in Arizona and Texas. An action threshold between 5 and 10 adults per cotton leaf generate the best economic return. A 190-acre trial in AZ validated the SPW per leaf threshold.
<b>Insecticide resistance: Elucidate mechanisms, monitor whitefly populations and develop management systems</b>	Baseline SPW insecticide susceptibility data was obtained using several techniques in California, Arizona, Texas and Florida; and Sonoran and Baja California, Mexico. Arizona and California bioassays showed a trend toward increasing pyrethroid resistance. PCR-based molecular diagnostics were used to investigate cyclodiene resistance in SPW biotypes. Synergists were used to implicate esterase detoxification and oxidative metabolism in resistance to chlorpyrifos and bifenthrin. Resistance management experiments and/or programs in cotton, based on rotation of insecticide types have reduced or eliminated resistance.
<b>Application technology: Develop improved methods for application or delivery of materials to enhance control and conserve natural enemies..</b>	Aerial application investigations involving air speed, above crop height, and nozzle type suggest improved delivery of sprays to target leaf areas. Electrostatic, air assist and hydraulic, ground operated spray systems have not shown clear advantage for increased underleaf coverage, but optimizing pump pressure, spray volume and nozzle configuration of conventional hydraulic systems improved underleaf coverage.

**Integration with other control tactics: Integrate chemical control with biological and cultural control.**

Florida, Texas and California studies indicated that parasitoids and predators responded differently to biorationals, IGRs and conventional insecticides. Although progress was made in studying the impact of chemicals and biorationals on natural enemies, no significant progress has been made on evaluating application methodologies on the SLW and natural enemy complexes.

---

**Sources<sup>1</sup>**

Toscano, N. C. and J. Palumbo. 1993. Section C Chemical Control, Biorationals and Pesticide Application Technology, pp. 89-90. In: T. J. Henneberry, N. C. Toscano, R. M. Faust and J. R. Coppedge [eds.]. Sweetpotato Whitefly: 1993 Supplement to the Five-Year National Research and Action Plan-First Annual Review. U.S. Dept. Agric., Agric. Res. Serv., No. 112, 175 pp.

Palumbo, J.C. and N.C. Toscano. 1994. Section C Chemical Control, Biorationals and Pesticide Application Technology, pp 118-119. In: T. J. Henneberry, N. C. Toscano, R. M. Faust and J. R. Coppedge [eds.]. Silverleaf Whitefly: 1994 Supplement to the Five-Year National Research and Action Plan-Second Annual Review. U.S. Dept. Agric., Agric. Res. Serv., No. 125, 228 pp.

Palumbo, J.C. and P.A. Stansly. 1995. Section C Chemical Control, Biorationals and Pesticide Application Technology, pp. 99-100. In: T. J. Henneberry, N. C. Toscano, R. M. Faust and J. R. Coppedge [eds.]. Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the Five-Year National Research and Action Plan-Third Annual Review. U.S. Dept. Agric., Agric. Res. Serv., 1995-02, 305 pp.

Palumbo, J.C. and P.A. Stansly. 1996. Section C Chemical Control, Biorationals and Pesticide Application Technology, pp. 102-103. In: T. J. Henneberry, N. C. Toscano, R. M. Faust and J. R. Coppedge [eds.]. Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the Five-Year National Research and Action Plan-Fourth Annual Review. U.S. Dept. Agric., 1996-01, 243 pp.

Stansly, P.A. 1997. Section C Chemical Control, Biorationals and Pesticide Application Technology, pp. 51-52. In: T. J. Henneberry, N. C. Toscano, T. M. Perring and R. M. Faust [eds.]. Silverleaf Whitefly, 1997 Supplement to the Five-Year National Research and Action Plan-Fifth Annual Review. U.S. Dept. Agric., 1997-02, 272 pp.

<sup>1</sup> Summaries of research progress abstracts were prepared by the indicated authors. See individual abstracts for research credits and more detail.

**Table 2. Summary of Research Progress for Section D - Biological Control of the 5-year National Research and Action Plan for Sweetpotato Whitefly Strain B (SPW) (1993-1997).**

Research Approaches	Research Result Highlights
<b>Determine effects of indigenous natural enemies on regulating SPW populations.</b>	High levels of SPW parasitism occur in urban weed host habitats, but not cotton fields. <i>Drapetis</i> spp. flies were identified as a SPW adult predator. Predaceous coccinellids and chrysopids were identified as potential biocontrol agents. Natural enemy activity exists in most ecosystems and is responsible for some degree of population suppression. Surveys of parasites revealed at least one <i>Eretmocerus</i> sp. in Arizona, California, Texas and South Carolina. Up to 6 <i>Encarsia</i> spp. were found in southeastern U.S., 5 <i>Encarsia</i> spp. in Texas and 2 species in the southwestern desert regions. Parasitism rates vary seasonally and on different host plants from 0 to over 70% depending on crop or weed hosts. In the San Joaquin Valley of CA, the overall percentage parasitism was 1.5% during 1993-1994. Seven parasitoid species and five predator species were collected from the Lower Rio Grande Valley of TX during 1994; yet, this assemblage of 12 species was unable to control <i>Bemisia</i> populations below economic injury levels. An egg specific monoclonal antibody technology was developed and results suggest that <i>Orius tristicolor</i> , <i>Lygus hesperus</i> , <i>Chrysoperla rufilabris</i> and <i>Georcoris punctipes</i> were dominant predators of whitefly eggs in the West.
<b>Develop methods for enhancing habitats with refuge plantings to conserve natural enemies.</b>	Moderate to high SPW parasitism in urban and weed host areas suggest they may be potential refuges. Refuge plantings of sunflower, kale, collard and kenaf were established in Texas in association with unsprayed cotton plots. Rates of parasitization in the Texas refuge plantings indicates 3-4 parasite species are common seasonally. A native plant species, <i>Heterotheca grandifolia</i> , native to the desert southwest, and kenaf, a crop grown as a potential substitute for paper production harbor large numbers of whitefly parasitoids, ( <i>Eretmocerus</i> sp.). In the Imperial Valley, CA, refuge plantings have had limited impact on adjacent plantings of cantaloupe and cole crops due to the poor performance of the indigenous parasitoids in these crops. Annual plants as refuges are not suitable because of their poor sustainability and high costs, but perennials may be satisfactory. A new banker plant method using vegetable transplants is being tested in TX.
<b>Identify new natural enemies in areas of SPW origin; foreign exploration, importation and release.</b>	SPW natural enemies (parasites, predator, fungi) were collected in Egypt, Greece, India, Nepal, Pakistan and Spain and held in quarantine facilities. Availability of extensive exotic biological material was encouraging. At least 19 species of natural enemies including <i>Encarsia</i> and <i>Eretmocerus</i> parasite species, predators and several isolates of fungi were cultured in California, Florida, Texas, and France. Additional natural enemy collections from Argentina, Brazil, Cyprus, Italy, Israel, Malaysia, the Philippines, Taiwan and Thailand yielded three fungal pathogens ( <i>Paecilomyces fumosoroseus</i> , <i>Fusarium coccophilum</i> and <i>Beauveria bassiana</i> ), at least 6 species of <i>Eretmocerus</i> and <i>Encarsia</i> , and 1 predator ( <i>Illeis koebele</i> ). Quarantine procedures including identification of adaptation and hyperparasitism for the collected species were accomplished and permits issued for their release. Extensive release programs occurred in California, Arizona, and Texas with many different parasitoid strains released. Exotic parasitoid and predator species were reported in CA, AZ, TX and FL indicating establishment and impact on SPW populations in selected systems.
<b>Determine natural enemy host selection processes and mechanisms .</b>	Host-plant, host-whitefly, and natural enemy interactions associated with plant hairiness, glandular exudates, and plant nutritional quality were demonstrated to influence the efficiency of natural enemies. Peanut and soybean parasitoids had either different parasitization efficiencies or different microhabitat preferences that were based on foliar pubescence of the host plant. Behavioral studies examining the tritrophic relationships of SPW and <i>Delphastus pusillus</i> suggest that foraging efficiency was influenced by the phenotypes of both the whitefly and its host plant.

**Inoculate/augment parasite and predator populations through propagation and release.**

*Encarsia formosa* gave effective SPW control in greenhouses in Ohio but not in New York. *Eretmocerus californicus* releases were promising in field cage cotton studies. Various strains of *Encarsia formosa* failed to provide SPW control on eggplant in Hawaii and failed to establish within field cages in the Imperial Valley, CA. However, releases of 2 species of *Eretmocerus* into cotton in the San Joaquin Valley, CA provided significant reductions of whitefly densities. Artificial diets are being investigated to improve cost-effectiveness of mass-produced predators. Large scale field trials employing augmentation of parasitoids showed promising results in CA and TX. Augmentative biological control using the native *Eretmocerus* (AZ) is effective against whiteflies in the greenhouse.

**Determine effects of pathogens on regulating SPW populations**

Over 50 fungal pathogens were collected for SPW pathogenicity screening. A staining method was developed to identify spore viability. *Beauveria bassiana* results were promising for SPW control on cotton. A large number of *Beauveria bassiana* and *Paecilomyces fumosoroseus* isolates from diverse geographic regions of North America and Asia were found highly effective against immature whiteflies in the laboratory. Applications of oil-formulated *B. bassiana* and *P. fumosoroseus* produced highly variable levels of control in field and greenhouse trials. Trials with a commercial strain of *Beauveria bassiana* for control of SPW infesting greenhouse tomatoes and ornamentals, and for cotton in the southern US provided control similar to the best insecticide-based programs. Separate trials with 4 isolates of *Beauveria bassiana* provided consistently high levels of whitefly control in experimental plots of cantaloupe and cucumber during both the spring and fall seasons. Trials conducted in cotton during the summer and in spring-planted tomatoes yielded poor results, yet trials in fall-planted tomatoes produced > 90% reduction in whitefly populations.

**Evaluate compatibility of pesticides with SPW natural enemies.**

Residues of seven insecticides on cotton leaves were tested for effects on four species of adult parasitoids. Buprofezin was non-toxic to all parasitoids; amitraz showed intermediate effects. *Eretmocerus mundus* from Spain exhibited some tolerance to the four insecticides.

**Systematics of predators, parasites and pathogens**

Extensive biological material has been curated and identified. A protocol was established and agreed to by ARS, APHIS, and SAES scientists for vouchering of specimens from importations and a centralized database for all importations established for *Encarsia* and *Eretmocerus* of world. Molecular technique classification of *Eretmocerus* and *Encarsia* has been accomplished and an identification key of the North American *Eretmocerus* spp. published. Publications with keys to identify the various other parasitoid species are in preparation.

---

**Sources<sup>1</sup>**

- Osborne, L. and L. Wendel. 1993. Section D Biological Control, pp. 114-115. In: T. J. Henneberry, N. C. Toscano, R. M. Faust and J. R. Coppedge [eds.]. Sweetpotato Whitefly: 1993 Supplement to the Five-Year National Research and Action Plan-First Annual Review. U.S. Dept. Agric., Agric. Res. Serv., No. 112, 175 pp.
- Anonymous. 1994. Section D Biological Control, pp 156-157. In: T. J. Henneberry, N. C. Toscano, R. M. Faust and J. R. Coppedge [eds.]. Silverleaf Whitefly: 1994 Supplement to the Five-Year National Research and Action Plan-Second Annual Review. U.S. Dept. Agric., Agric. Res. Serv., No. 125, 228 pp.
- Heinz, K.M. and O. Minkenberg. 1995. Section D Biological Control, pp. 144-145. In: T. J. Henneberry, N. C. Toscano, R. M. Faust and J. R. Coppedge [eds.]. Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the Five-Year National Research and Action Plan-Third Annual Review. U.S. Dept. Agric., Agric. Res. Serv., 1995-02, 305 pp.
- Minkenberg, O. and K. Heinz. 1996. Section D Biological Control, pp. 155-156. In: T. J. Henneberry, N. C. Toscano, R. M. Faust and J. R. Coppedge [eds.]. Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the Five-Year National Research and Action Plan-Fourth Annual Review. U.S. Dept. Agric., 1996-01, 243 pp.
- Heinz, K. and O. Minkenberg. 1997. Section D Biological Control, pp. 53-55. In: T. J. Henneberry, N. C. Toscano, T. M. Perring and R. M. Faust [eds.]. Silverleaf Whitefly, 1997 Supplement to the Five-Year National Research and Action Plan-Fifth Annual Review. U.S. Dept. Agric., 1997-02, 272 pp.

<sup>1</sup> Summaries of research progress abstracts were prepared by the indicated authors. See individual abstracts for research credits and more detail.



**Table 2 (Continued). Summary of Section E: Crop Management Systems and Host Plant Resistance Research Progress of the 5-Year National Research and Action Plan on Sweetpotato Whitefly Strain B (SPW) (1993-1997).**

Research Approaches	Research Result Highlights
<b>Determine effect of traditional crop production inputs on SPW population development.</b>	Frequent cotton irrigations reduced SPW populations verifying that water stress facilitated population development. Weekly irrigated plants also required fewer insecticide application to maintain adults below treatment thresholds. Research conducted on evaluation of the effect of water stress on populations has been accepted and implemented by cotton growers. Integrating the use of reduced water stress by weekly irrigation and conventional insecticides has provided the best control. Water stressed plants had more SPW eggs than non-stressed plants. Sprinkler irrigation were found to reduce SPW populations compared to furrow irrigation in cotton. Whitefly flight behavior was reduced in irrigated plants as compared with water stressed plants. Populations of native parasitoids persisted in foliar applied imidacloprid plots, although their abundance was reduced compared with plots not receiving insecticide.
<b>Determine temporal and spatial effects of host plants on SPW populations and dispersion.</b>	SPW population differences within and between major cultivated crops were identified were apparently due to mechanism of resistance. The selection, crossing and genetic analysis to incorporate resistance characters into acceptable agronomic types has progressed with some favorable results. For example, smooth leaf cottons have shown less SPW numbers than hairy leaf varieties. The timing of SPW infestation occurrence on different crops in relation to spatial arrangements have allowed cultural practices such as planting times, harvest dates, crop residue destruction, etc., to be manipulated to reduce damage in some production areas. Large scale cropping patterns in SPW using LandSat data to map crops for modeling the spatiotemporal dynamics and SPW migrations have been integrated in areawide management systems. Areawide approaches focus on wind patterns, distance adults migrate, distribution of migratory SPW from point sources, and the ability of whiteflies to reproduce in relation to migration distances.
<b>Determine effect of colored mulches, trap crops, intercropping, row covers, and other innovative cultural practices as potential SPW control methods.</b>	The mode of action, disturbance, physical, mortality, etc. under sprinkler irrigation systems appears to be for reduced populations. Row covers were effective in limiting SPW cantaloupe infestations through the early season. Plastic mulches (white and silver reflective materials) repel SPW adults but have not been effective enough to justify their expense. Melons as a trap crop reduced the SLW infestations in cauliflower and interplanting melons in the system appear to increase efficacy. Fine-mesh screens exclude migrating SPW adults from greenhouses. Forced positive air flow through SPW-proof filters reduced the influx of SPW and incidence of virus transmission. Aluminum plastic mulch, from mesh screen row covers and living soil covers delayed the incidence of tomato mottle geminivirus on tomato by reducing early infestations in Florida.

**Develop reproducible evaluation techniques to isolate resistant germplasm**

Experimental and mesophyll leaf tissue morphological differences appear to be associated with SPW numbers on different cotton lines. Trichome exudate production in tomatoes was inversely related to SPW feeding and oviposition. Cotton trichome density was related to high SPW densities on cottons. Methods to assess host plant resistance were developed in: alfalfa, broccoli, collard, eggplant, lettuce, melon, pepper, soybean, and tomato. More SLW oviposition occurred on pubescent and hirsute near-isoline soybeans versus a glabrous genotype. In melon, there was a positive correlation between long-leaf trichomes and number of SPW adults and immatures. Type IV and VI trichomes of *Lycopersicon hirsutum* accessions were evaluated for their role in resistance against *Bemisia*. Differences were mainly associated with day length and accessions. Plants under 8 hour day length had the most dense type IV trichomes and received the least oviposition versus 12 and 16 hour day plants. Trichome density did not explain all behavioral differences by SPW among accessions. A simulation model for evaluating plant germplasm was developed based on antibiotic factors. A testing procedure was established to provide a repeatable protocol for evaluation of lettuce germplasm for SPW feeding. Also, procedures were established for evaluation of SPW feeding, honeydew contamination and sooty mold contamination in alfalfa. Leaf thickness was highly correlated with the depth of vascular bundles in cotton. Work on other resistance mechanisms in cotton includes reduced trichome numbers, okra-leaf shape, and red plant color. Two collard hybrids had fewer whiteflies than open-pollinated cultivars, but three glossy genotypes were the most resistant. Pubescence was a factor in host oviposition selection and vertical distribution of oviposition on soybean, but survival and development were effected. Leaf thickness was found to be highly correlated with the depth of vascular bundles in cotton and may be developed as a resistance mechanism.

**Identify resistant germplasm to SPW and associated viruses and plant disorders.**

SPW population differences among *Brassica oleracea* groups suggest plant size and color affected preference. Oviposition preference appears to be a factor in sweetpotato breeding line evaluations. Significant differences also occurred between peanut cultivars, melons, soybeans, cotton and tomatoes. High levels of resistance to SPW were not found in any crop species or cultivars tested. Evaluation of 15 Cotton cultivars for resistance to SPW resulted in no significant differences among cultivars. Differences were detected among 12 melon cultivars in ovipositional rate, nymphal numbers, adult survivorship in field trials, and greenhouse lifetable studies. Significant differences were detected among 12 broccoli cultivars for SPW colonization preference. Two zucchini lines (Sunseed 3 and A21-7) had resistance to SPW symptoms, but not to whitefly infestations.

**Conduct plant breeding studies to select SPW resistant plant germplasm.**

Tomato plant selection and crossing for tomato mottle resistance identified several sources of resistant germplasm. Breeding lines were investigated for both melons and tomatoes. Snakemelon was identified as a parental material that had low SPW counts/unit area. Several crosses with this material produced lines that had fewer SPW than snakemelon. Tomato lines were evaluated for stickiness and horticultural traits resulting in selection of 52 for low tomato mottle virus symptom expression, high stickiness and fruit-set. Breeding lines, half-sib "families" of alfalfa from a genetically diverse germ plasm pool, resulted in several crosses that had lint and well as fewer SPW immatures and less stickiness from honeydew. Alfalfa germplasm with whitefly resistance and other good qualities should soon (ca. 1998) be available for seed increases by industry. A glabrous type cotton with SPW resistance, good yield and other good qualities, was released and is commercially marketed as Texas 121. SPW on broccoli lines are being back crossed with commercial cultivars to improve agronomic characteristics.

---

**Sources<sup>1</sup>**

- Hardee, D.D. and G. G. Still. 1993. Section E Crop Management Systems and Host Plant Resistance, pp. 132-133. In: T. J. Henneberry, N. C. Toscano, R. M. Faust and J. R. Coppedge [eds.]. Sweetpotato Whitefly: 1993 Supplement to the Five-Year National Research and Action Plan-First Annual Review. U.S. Dept. Agric., Agric. Res. Serv., No. 112, 175 pp.
- Hardee, D.D. and R.I. Carruthers. 1994. Section E Crop Management Systems and Host Plant Resistance, pp 173-174. In: T. J. Henneberry, N. C. Toscano, R. M. Faust and J. R. Coppedge [eds.]. Silverleaf Whitefly: 1994 Supplement to the Five-Year National Research and Action Plan-Second Annual Review. U.S. Dept. Agric., Agric. Res. Serv., No. 125, 228 pp.
- Natwick, E.T. and A.M. Simmons. 1995. Section E Crop Management Systems and Host Plant Resistance, pp. 160-161. In: T. J. Henneberry, N. C. Toscano, R. M. Faust and J. R. Coppedge [eds.]. Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the Five-Year National Research and Action Plan-Third Annual Review. U.S. Dept. Agric., Agric. Res. Serv., 1995-02, 305 pp.
- Natwick, E.T. and A.M. Simmons. 1996. Section E Crop Management Systems and Host Plant Resistance, pp. 174. In: T. J. Henneberry, N. C. Toscano, R. M. Faust and J. R. Coppedge [eds.]. Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the Five-Year National Research and Action Plan-Fourth Annual Review. U.S. Dept. Agric., 1996-01, 243 pp.
- Simmons, A.M. and E.T. Natwick. 1997. Section E Crop Management Systems and Host Plant Resistance, pp. 56-58. In: T. J. Henneberry, N. C. Toscano, T. M. Perring and R. M. Faust [eds.]. Silverleaf Whitefly, 1997 Supplement to the Five-Year National Research and Action Plan-Fifth Annual Review. U.S. Dept. Agric., 1997-02, 272 pp.

<sup>1</sup> Summaries of research progress abstracts were prepared by the indicated authors. See individual abstracts for research credits and more detail.

**Table 2 (Continued). Summary of Research Progress for Section F Integrating Techniques, Approaches and Philosophies Research Progress of the 5-Year National Research and Action Plan on Sweetpotato Whitefly Strain B (SPW) (1993-1997).**

Research Approaches	Research Result Highlights
<b>Risk Assessment.</b>	<p>In Texas, large SPW nymph numbers provided the best index for relationships to yield and quality measurements. One large nymph/6.7 cm<sup>2</sup> of leaf at the 6th node or three adults/leaf at the 3rd node were initial thresholds. SPW population survey results were incorporated into GIS systems. GIS systems for predicting virus spread were used for tomato growers in northern Mexico. A broader understanding of virus-whitefly plant interaction, increased understanding of the role of different host plants in population development, increased understanding of SPW host plant resistance, and sampling techniques for monitoring population development are leading information for management development. There is an increased understanding of insecticide resistance management in whitefly management programs. Economic losses from California, Texas, and the Mexicali area of Mexico, showed a general downward trend to SPW infestations with new management approaches that included cultural practices such as destruction of old host crops and close attention to planting dates. Selected insecticides for vegetable crops grown prior to cotton production has reduced SPW in cotton early in early season. Fluctuation in populations occurred from season to season, but predictions are not possible. A more complete understanding of the role of different host plants in population development; mechanisms of host plant resistance; enhanced whitefly sampling methods and dispersal; and understanding of the genetic basis of insecticide resistance in whitefly populations are essential for areawide, community wide Integrated Management Basis or Systems.</p>
<b>Spatial Analysis and GIS.</b>	<p>Crop sequencing and spatial arrangement studies are on an area-wide basis in AZ, CA, and TX have been displayed on GIS systems to SPW areawide management is now generally accepted. An informal network of scientists interested in the remote sensing applications to SPW dynamics has formed. Satellite crop map information is being coupled with pest management issues related to spatial distribution. Methods for mapping crop systems from satellite (Landsat) image data were developed and applied to the San Joaquin, Imperial and Lower Rio Grande Valleys. Methods of interfacing satellite based crop maps with insect simulation models have been developed and used in simulations.</p>
<b>Ecosystem modeling.</b>	<p>Survey data from CA, TX, and FL have been used to develop simple models for SPW growth and movement. Work in the HERMES environment to model interactions between SPW and two of its natural enemies had begun. GIS technology has been developed and is bring applied to models to evaluate and predict SPW population development and dispersal within certain Agroecosystems. Field Sampling is being used to validate prediction capabilities of whitefly population models. New methods of modeling and visualizing spatial dynamics have been developed which use the fast Fourier transform for solving the spatial dispersal/reproduction process in a spatial model of whitefly dynamics making it possible to use of PCs for simulation of spatially extended systems. A satellite image analysis system (TNTmips) has now been installed at Kearney Agricultural Center in the San Joaquin Valley. A full Landsat scene (IOOxI15mi) of the Valley (July 3, [994) has been provided thus demonstrating the potential of this technology to producers. Field Sampling in Texas, Arizona, and California is being used to validate prediction capabilities of whitefly population models.</p>

## Networks.

The NBCI and SPW management committee bulletin board is on line and accessible through Internet. It has a SPW forum and is available for the dissemination of information and for communication. Pest alert database is available via gopher at the University of Florida and includes whitefly and virus information. A series of homepages and 1-800 hotlines for consumers on whiteflies have been put in place to date in Florida, California, and Arizona. A broad network of whitefly information is linked on the WWW with homepages in Arizona, California, Florida, and Texas.

## Integrated Extension Programs.

Area wide management programs have been initiated in Arizona, California and Texas. Publications and resource materials produced by scientists in Arizona, California, Florida and Texas are shared. Local leadership and team building, whitefly management committees and task forces have formed to address area wide management approaches. Training programs were developed and implemented in California and Arizona. Satellite crop map information is being coupled with pest management issues related to spatial distribution. This information is being targeted to IPM implementation programs as they relate to crop sequencing in areas wide management approaches. Whiteflies are an important component in cotton and vegetable IPM programs which have been implemented in Arizona, Mexico, California, and Texas. Growth and acceptance is visible in each program as they expand in geographic area and crops covered. A "train the trainer" program was developed and implemented in California and Arizona, which did much to strengthen the ties and cooperation between growers, extension, and research scientists. Satellite crop map information is being coupled with and related to pest population data, climatic data, whitefly population migration data, spatial distribution, and pest management data. This information is being targeted to IPM implementation programs as they relate to crop sequencing for community wide management approaches. A state regulatory agency (Arizona Dept. of Agriculture and a commodity group (Arizona Cotton Growers Association) took responsibility for stewarding a progressive Section 18 process which enabled uses of selective *Bemisia* growth regulators, namely Applaud and Knack. Cooperative Extension through industry collaboration developed the section 18 and implemented a proactive educational program.

---

## Sources<sup>1</sup>

- Armador, J.M. and D.A. Nordland. 1993. Section F Integrated Techniques, Approaches and Philosophies, pp. 141-142. In: T. J. Henneberry, N. C. Toscano, R. M. Faust and J. R. Coppedge [eds.]. Sweetpotato Whitefly: 1993 Supplement to the Five-Year National Research and Action Plan-First Annual Review. U.S. Dept. Agric., Agric. Res. Serv., No. 112, 175 pp.
- Armador, J.M., D.A. Nordland, and D.D. Kopp. 1994. Section F Integrated Techniques, Approaches and Philosophies, pp 184-185. In: T. J. Henneberry, N. C. Toscano, R. M. Faust and J. R. Coppedge [eds.]. Silverleaf Whitefly: 1994 Supplement to the Five-Year National Research and Action Plan-Second Annual Review. U.S. Dept. Agric., Agric. Res. Serv., No. 125, 228 pp.
- Kopp, D.D., D.A. Nordland, and J. Norman. 1995. Section F Integrated Techniques, Approaches and Philosophies, pp. 173-174. In: T. J. Henneberry, N. C. Toscano, R. M. Faust and J. R. Coppedge [eds.]. Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the Five-Year National Research and Action Plan-Third Annual Review. U.S. Dept. Agric., Agric. Res. Serv., 1995-02, 305 pp.
- Norman, J. Jr, D.D. Kupp and P. Goodell. 1996. Section F Integrated Techniques, Approaches and Philosophies, pp. 186-187. In: T. J. Henneberry, N. C. Toscano, R. M. Faust and J. R. Coppedge [eds.]. Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the Five-Year National Research and Action Plan-Fourth Annual Review. U.S. Dept. Agric., 1996-01, 243 pp.
- Norman, J. P. Goodell, P. Ellsworth, J. Allen, C. Brewster and D. Kopp. 1997. Section F Integrated Techniques, Approaches and Philosophies, pp. 59-62. In: T. J. Henneberry, N. C. Toscano, T. M. Perring and R. M. Faust [eds.]. Silverleaf Whitefly, 1997 Supplement to the Five-Year National Research and Action Plan-Fifth Annual Review. U.S. Dept. Agric., 1997-02, 272 pp.

<sup>1</sup> Summaries of research progress abstracts were prepared by the indicated authors. See individual abstracts for research credits and more detail.

**Table 3. Summary of Research Progress for Section A Biology, Ecology, and Population Dynamics of the 5-Year National Research, Action and Technology Transfer Plan on Sweetpotato Whitefly Strain B (SPW) (1997-2002).**

Research Approaches	Research Result Highlights
<b>Determine life cycle vulnerabilities (life tables), population development and natural mortality factors, natural enemies on major crops, urban plantings, weeds and predict overwintering potential.</b>	<i>Bemisia</i> life table analyses on cotton in Arizona showed that natural factors, including predation and physical dislodgment were major mortality factors. High winter mortality occurs. Parasitism was a minor source of mortality. Survivorship from egg to adult ranged from 0-18.2% over 6 generations in sprayed and unsprayed fields. Studies on wild hosts in Israel indicate that parasitoids may contribute to maintaining low levels of SPW on lantana.
<b>Develop sampling methodology, action and economic thresholds for all major crops. Sampling methods and thresholds modified in light of natural enemy levels and existing management strategies.</b>	Relationships between whitefly density and the occurrence of tomato irregular-ripening as well as preliminary sampling plans for SPW on tomato were developed.
<b>Develop population models to describe and predict whitefly population growth and spatial and temporal distribution. Develop simple day-degree sub-models for estimating phenology and temporal patterns of whitefly, natural enemies and host crops..</b>	Partial development of large-scale temporal and spatial models and temperature-dependent, site-specific population dynamics models was accomplished. Additional biological and ecological detail, as well as information on various aspects of pest management needs to be integrated into the models to make them useful as exploratory research tools. A temperature-dependent, site-specific population dynamics model in cotton and cantaloupe was completed. Additional refinements, enhancements and field validation are needed for predicting population dynamics under various management regimes and environmental conditions.
<b>Develop sampling methods for quality of cotton lint, vegetables and other commodities.</b>	Cotton lint stickiness was shown to be randomly distributed in cotton fields. Preliminary sampling plans were developed for estimating pre-harvest cotton lint stickiness. Comparative evaluations of manual and high speed cotton stickiness thermometers revealed differences in performance that have important implications for the development of measurement scales for stickiness and the number of samples that would need to be collected for the precise estimation of stickiness. Thermometer sticky cotton measurements in relation to amounts of honeydew and SPW populations were highly correlated. Plant stress can affect honeydew production.
<b>Quantify whitefly and natural enemy dispersals and contribution to population dynamics.</b>	Aerial distribution of SPW dispersing from cantaloupe fields to cotton and other hosts were documented. The SPW, <i>Eretmocerus eremicus</i> average 12 min. flight durations, respond to plant cues shortly after take off.
<b>Define mating behavior, reproductive isolation, species, biotypes.</b>	Surveys worldwide document the spread of SPW Strain B. Electrophoretic analyses demonstrated its presence in Australia and Brazil. Heterozygotes between <i>B. argentifolii</i> and the Australian <i>B. tabaci</i> corroborated previous laboratory results and support the existence taxonomic problems within the <i>Bemisia</i> species complex. Two SPW biotypes, cassava and sweetpotato, were identified in India.

<b>Validate <i>Bemisia</i> taxa morphology, genetic, biochemical, and biology characteristics.</b>	Comparative morphological analyses of <i>Bemisia</i> pupae from around the world show highly variable characters suggesting that pupal morphology should not be the sole criteria for classifying individuals within the <i>Bemisia</i> species complex.
<b>Define role of endosymbionts in metabolism, host adaptation, nutrition and survival.</b>	Several antibiotics that interfere with bacterial protein synthesis affected SPW nymph growth and development. None affected adult oviposition rates or sex ratios. <i>Wolbachia</i> endosymbiotic bacteria was discovered in SPW suggesting significant implications for development of control strategies manipulating reproductive biology.
<b>Characterize nutrient uptake and metabolism</b>	Sorbitol was suggested to function as a thermoprotectant in SPW that enables them to thrive in desert environments. The unique pathway of sorbitol synthesis and degradation in SPW may offer an opportunity to develop transgenic plants that disrupt sorbitol synthesis and compromise SPW ability to deal with heat stress. Nymphs and adult feeding, stylet lengths and feeding mechanism descriptions were adequately described – nymph stylets 110-200 µm long, adult up to 217µm long.
<b>Develop whitefly artificial diets and natural enemy mass-rearing.</b>	An artificial diet and feeding system for rearing SPW immatures was been developed. Rates of instar development were comparable to those reported on various host plants. Adult <i>Encarsia</i> , parasitoid wasps, have been successfully produced using the diet and feeding system for SPW hosts.
<b>**Pursue specific genetic and biological basis for variability in whitefly biotypes, strains, and species; determine impact of different genotypes/phenotypes on whitefly-mediated transmission and on the epidemiology of virus diseases.</b>	A cell line from the silverleaf whitefly, BtB-2.97-Hunter & Polston was tested for its ability to support replication of the insect iridescent virus 6 (IIV-6). Several lines of evidence indicated that a productive infection was achieved. The cells displayed cytopathic effects, virus particles accumulated in defined areas of the cytoplasm, the cell-associated virus titer was detected at three orders of magnitude higher than that released into the media, and western blot analysis indicated CIV structural proteins were being expressed. Virus was also detected in nymphs by PCR and electron microscopy, but the infections were not highly pathogenic. Infection by whiteflies by IIV-6 suggests that more pathogenic viral isolates may be found for this insect and that the virus has potential to be used as an intracellular probe.

---

#### Sources<sup>1</sup>

- Naranjo, S.E. 1998. Section A Biology, Ecology and Population Dynamics, pp. 23-24. In: T. J. Henneberry, N. C. Toscano, T. M. Perring and R. M. Faust [eds.]. Silverleaf Whitefly National Research, Action and Technology Transfer Plan, 1997-2001 (Formerly Sweetpotato Whitefly, Strain B: First Annual Review of the Second 5-year Plan. U. S. Dept. of Agric. 1998-01, 187 pp.
- Isaacs, R. and S.E. Naranjo. 1999 Section A Biology, Ecology and Population Dynamics, pp. 22-23. In: T. J. Henneberry and R. M. Faust [eds.]. Silverleaf Whitefly National Research, Action and Technology Transfer Plan, 1997-2001 (Formerly Sweetpotato Whitefly, Strain B: Second Annual Review of the Second 5-year Plan. U. S. Dept. of Agric. 1999-01, 195 pp.
- Salvucci, M. and T. Perring. 2000 Section A Biology, Ecology and Population Dynamics, pp. 29-30, In: T. J. Henneberry, R. M. Faust, W. A. Jones and T. M. Perring [eds.]. Silverleaf Whitefly National Research, Action and Technology Transfer Plan, 1997-2001 (Formerly Sweetpotato Whitefly, Strain B: Second Annual Review of the Second 5-year Plan. U. S. Dept. of Agric., Agric. Res. Ser., 209 pp.
- Byrne, D.N. and J.L. Blackmer. 2002 Section A Biology, Ecology and Population Dynamics, pp. 82. In: T. J. Henneberry, R. M. Faust, W. A. Jones and T. M. Perring [eds.]. Silverleaf Whitefly National Research, Action and Technology Transfer Plan, 1997-2001 (Formerly Sweetpotato Whitefly, Strain B: Fourth Annual Review of the Second 5-year Plan. U. S. Dept. of Agric., Agric. Res. Ser., 446 pp.

<sup>1</sup> Summaries of research progress abstracts or tables as listed were prepared by the indicated authors. See individual abstracts for research credits and more detail.

**Table 3 (Continued). Summary of Research Progress for Section B Epidemiology and Virus Vector Interactions of the 5-Year National Research, Action and Technology Transfer Plan on Sweetpotato Whitefly Strain B (SPW) (1997-2002).**

Research Approaches	Research Result Highlights
<b>Identification and characterization of new or emerging whitefly-transmitted viruses and strains.</b>	Tomato yellow leaf curl virus (TYLCV) is a monopartite virus within the 17 geminiviruses infecting tomato alone. The group is considered the most economically important. Various techniques (antibodies, blots, PCR, DNA sequencing) have been developed for virus identification. TYLCV introduction into Florida may have significant economic impact on Florida tomato production. A new cucurbit-infecting geminivirus was identified in Imperial Valley, CA. Cucurbit yellow sticky disorder virus (CYSDV) was identified in the Rio Grande Valley, TX. Characterization of several new geminiviruses is being accomplished. New technologies are developing with expanding needs.
<b>Molecular epidemiology: identification of economic viruses, host plants, and reservoirs, and determination of geographic distribution of viruses.</b>	PCR and DNA sequencing confirmed TYLCV in Florida. A number of symptomless weed hosts were found to have TYLCV. Host free periods effectively manage TYLCV. New systemic insecticides have been promising in some virus-disease problems and cultural practices such as time of planting are additional useful approaches to management.
<b>Virus-vector interactions, factors affecting virus transmission, and basis for virus-vector specificity; determination of endosymbiont involvement in whitefly-mediated transmission.</b>	Differences in feeding behavior of aphids and SPW adults relating to each of cell punctures early stylet probing was observed. Evidence for transovarial transmission of TYLCV in SPW and sexual transmission between males and females has been found.
<b>Strategies to reduce virus spread by management of cropping systems, reduced transmission frequencies, and other potentially effective approaches.</b>	Whitefly host (tomato, beans, melons and peppers) free periods of 3 months have been shown to reduce TYLCV incidence. Living covers (coriander and perennial peanuts) and plastic mulch were found to reduce TYLCV.



**Control of virus diseases:  
development of virus resistant  
germplasm through conventional and  
engineered/ molecular approaches.  
Define prospective strategies for  
selecting candidate viruses,  
identifying specific virus diseases to  
target, and prioritize specific crops  
and cultivars for protection  
approaches.**

Gene marking of viruses has been useful in studying virus movement and infection activity. Potentially resistant CLCrV have been identified. TYLCV resistant tomatoes have been made commercially available.

#### **Sources<sup>1</sup>**

- Huettel, R. N. 1998. Section B Viruses, Epidemiology and Virus Vector Interactions, pp. 32-32. In: T. J. Henneberry, N. C. Toscano, T. M. Perring and R. M. Faust [eds.]. Silverleaf Whitefly National Research, Action and Technology Transfer Plan, 1997-2001 (Formerly Sweetpotato Whitefly, Strain B: First Annual Review of the Second 5-year Plan. U. S. Dept. of Agric. 1998-01, 187 pp.
- Gilbertson, R. 1999 Section B Viruses, Epidemiology and Virus Vector Interactions, pp. 36-37. In: T. J. Henneberry and R. M. Faust [eds.]. Silverleaf Whitefly National Research, Action and Technology Transfer Plan, 1997-2001 (Formerly Sweetpotato Whitefly, Strain B: Second Annual Review of the Second 5-year Plan. U. S. Dept. of Agric. 1999-01, 195 pp.
- Gilbertson, R. K. and R. Huettel. 2000 Section B Viruses, Epidemiology and Virus Vector Interactions, pp. 46-47, In: T. J. Henneberry, R. M. Faust, W. A. Jones and T. M. Perring [eds.]. Silverleaf Whitefly National Research, Action and Technology Transfer Plan, 1997-2001 (Formerly Sweetpotato Whitefly, Strain B: Second Annual Review of the Second 5-year Plan. U. S. Dept. of Agric., Agric. Res. Ser., 209 pp.
- Gilbertson, R.K. and J.K. Brown. 2002 Section B Viruses, Epidemiology and Virus Vector Interactions, pp. 109-111. In T. J. Henneberry, R. M. Faust, W. A. Jones and T. M. Perring [eds.]. Silverleaf Whitefly National Research, Action and Technology Transfer Plan, 1997-2001 (Formerly Sweetpotato Whitefly, Strain B: Second Annual Review of the Second 5-year Plan. U. S. Dept. of Agric., Agric. Res. Ser., 446 pp.

<sup>1</sup> Summaries of research progress abstracts or tables as listed were prepared by the indicated authors. See individual abstracts for research credits and more detail.

**Table 3 (Continued). Summary of Research Progress for Section C Chemical Control, Biopesticides, Resistance Management, and Application Methods of the 5-Year National Research, Action and Technology Transfer Plan on Sweetpotato Whitefly Strain B (SPW) (1997-2002).**

Research Approaches	Research Results
<p><b>Improve insecticide efficacy: Develop, test, and assist in the registration of insecticides, biorationals, and natural products.</b></p>	<p>Numerous new chemistries, biorational plant extracts, potential biologicals and the like have been evaluated to expand conventional chemistry control options. Neonicotinoid systemics are effective for SPW control on numerous crops.</p>
<p><b>Develop improved methods of application including formulation and delivery of materials to improve control.</b></p>	<p>Higher volumes, increased pressure result in increased underleaf coverage. Conventional hydraulic spray systems have been as effective as air-assist, electrostatic systems.</p>
<p><b>Conserve insecticide efficacy: Relate action thresholds to insecticide usage patterns.</b></p>	<p>Action threshold implementation were demonstrated to reduce pesticide use. Also, they are important in resistance management and natural enemy conservation.</p>
<p><b>Elucidate the role of genetic, biochemical and ecological factors leading to insecticide resistance.</b></p>	<p>Modes of action, genetic and biochemical basis for resistance need further emphasis. Monitoring of resistance has not revealed resistance to new chemistries (IGRs, neonicotinoids, etc.).</p>
<p><b>Improve insecticide efficacy: Improve techniques for monitoring resistance.</b></p>	<p>Resistance management monitoring and implementation of action thresholds have been implemented in most cropping systems and their value in pest management demonstrated repeatedly. Uptake of systemics and relationships to mortalities appear to be a promising approach to improve efficacy and efficiency.</p>
<p><b>Develop, evaluate and refine resistance management systems.</b></p>	<p>Rotations, tank mixes, and new chemistries integrated into SPW management systems have resulted in conservation of natural enemies in large area programs and prevents delay and/or forego resistance development. Cost commodity approaches to conserving use patterns for multicrop IRM is essential.</p>

**Integrate chemical control with other tactics.** Increasing awareness of potentials for combining control strategies has stimulated implementation of control systems using new chemistry selections (IGR, neonicotinoids), biorationals, smooth leaf cottons and other resistant crops, and resistance management techniques. Conservation of natural enemies has become a reality with some of the new chemistries.

#### Sources<sup>1</sup>

Stansly, P. A. 1998. Section C Chemical Control, Biopesticides, Resistance Management and Application Methods, pp. 50-51. In T. J. Henneberry, N. C. Toscano, T. M. Perring and R. M. Faust [eds.]. Silverleaf Whitefly National Research, Action and Technology Transfer Plan, 1997-2001 (Formerly Sweetpotato Whitefly, Strain B: First Annual Review of the Second 5-year Plan. U. S. Dept. of Agric. 1998-01, 187 pp.

Brazzle, J. 1999 Section C Chemical Control, Biopesticides, Resistance Management and Application Methods, pp. 63. In T. J. Henneberry and R. M. Faust [eds.]. Silverleaf Whitefly National Research, Action and Technology Transfer Plan, 1997-2001 (Formerly Sweetpotato Whitefly, Strain B: Second Annual Review of the Second 5-year Plan. U.S. Dept. of Agric. 1999-01, 195 pp.

Brazzle, J. 2000 Section C Chemical Control, Biopesticides, Resistance Management and Application Methods, pp. 72, In T. J. Henneberry, R. M. Faust, W. A. Jones and T. M. Perring [eds.]. Silverleaf Whitefly National Research, Action and Technology Transfer Plan, 1997-2001 (Formerly Sweetpotato Whitefly, Strain B: Second Annual Review of the Second 5-year Plan. U.S. Dept. of Agric., Agric. Res. Ser., 209 pp.

Taylor, S. and J. Palumbo. 2002 Section C Chemical Control, Biopesticides, Resistance Management and Application Methods, pp. 147, In T. J. Henneberry, R. M. Faust, W. A. Jones and T. M. Perring [eds.]. Silverleaf Whitefly National Research, Action and Technology Transfer Plan, 1997-2001 (Formerly Sweetpotato Whitefly, Strain B: Second Annual Review of the Second 5-year Plan. U.S. Dept. of Agric., Agric. Res. Ser., 446 pp.

<sup>1</sup> Summaries of research progress abstracts or tables as listed were prepared by the indicated authors. See individual abstracts for research credits and more detail.

**Table 3 (Continued). Summary of Research Progress for Section D Natural Enemy Ecology and Biocontrol of the 5-Year National Research, Action and Technology Transfer Plan on Sweetpotato Whitefly (SPW) Strain B (1997-2002).**

Research Approaches	Research Results
<b>Develop natural enemy conservation practices to reduce mortality to indigenous and introduced natural enemies.</b>	New insect growth regulators tested well under field conditions, and reduced loss of natural enemies. A Life Table analysis was conducted on natural enemies in cotton. Role of predators in cotton identified; importance of narrow spectrum insecticides highlighted. Life history tables have been constructed comparing mortality factors of natural enemies in conventional vs IGR treated cotton. Functional response data available for several parasitoid species. Imidicloprid and IGR's are more compatible with natural enemies. Pathogenic fungi may be promising. Annual refugia plantings are of limited value, perennials are promising.
<b>Evaluate potential of alternate plants to act as in-field refuges or insectaries for natural enemies.</b>	Combinations of annuals and some perennials show promise as within field natural enemy refugia. They are attractive to parasites but support low numbers of whiteflies. Annuals served as outdoor insectaries when releasing exotic parasitoids. Perennial plants capable of growing in Imperial Valley identified, selected for a pilot project at a commercial organic farm. Research in the Imperial Valley has shown that perennial refuges support large numbers of whitefly and parasitoids that migrate to adjacent systems.
<b>Augmentation of natural enemies: Develop natural enemy mass-rearing systems.</b>	Foreign exploration exhaustive. Evidence for establishment following releases. Diets are being developed for generalist predators. Improvements have been made in rearing parasitoids, increasing rearing efficiency. Field studies have identified promising candidates for augmentative releases. Whitefly, parasitized by <i>Encarsia</i> , were grown on an artificial diet long enough for parasitoids to emerge as adults. First such report. Potential for research and commercial rearing. Mass rearing methods on SLWF has been accomplished. Artificial diets are still being researched, with economics undetermined. Parasite release monitoring suggest establishment in some areas. Several fungi appear promising for control.
<b>Develop release technologies to maximize the effectiveness of mass-reared natural enemies in the field.</b>	A novel "Banker Plant" field release strategy shows promise for increasing efficacy of releases. Releases of <i>Eretmocerus</i> into greenhouses controlled <i>Bemisia</i> attacking poinsettias when done at low pest densities. First year results show banker plants may prove more efficacious than releases of parasitoids by hand. Two species of coccinellids evaluated, compared for greenhouse use. Several release technologies have or are being tested. Banker plant technology appears to be very effective. Capsule delivery methods being tested. Cold storage of parasitoid pupae also being tested.
<b>Evaluate augmentative parasitoid, predator, or pathogen releases.</b>	Augmentative releases of parasitoids controlled <i>Bemisia</i> in large demonstration fields. These releases can be integrated with conventional pest management practices. Impact of <i>Beauveria bassiana</i> on generalist predators determined. Parasitoid dispersal was determined using new protein marking technique. Parasitoid release rates have been determined for major crops. Strategies for releasing/integrating parasitoid and predator in greenhouse crops have been determined. Significant information currently available on application of fungal pathogens in various crops.

**Importation biological control:  
Evaluate the ability of exotic natural  
enemies to suppress whitefly  
populations under field conditions.**

Combinations of annual plants that make excellent insectaries and can be farmed under local climatic conditions have been identified. Homeowners are being recruited to care for plants used for making releases. Several new exotics have persisted over several years and are multiplying and spreading in Texas and California. Whitefly suppression by exotic parasitoids determined by multiple researchers. Preliminary data suggests significant spread of established exotic parasitoids in some regions. Dispersal rates of natural enemies still under study. Book being prepared for publication.

**Systematics, ecology, and population dynamics of natural enemies:  
Clarify systematics of predators,  
parasitoids and pathogens.**

Taxonomic studies have been completed on the exotic *Eretmocerus* and a key to their identification is in press. PCR techniques have been developed to identify the purity of cultures and aid in identification of recovered parasites. Key on exotic *Eretmocerus* published. Program developed for curating, cataloging recovered parasitoids. Several taxonomic keys developed for imported parasitoid species. RAPD-PCR techniques proven as quick identification method. Preliminary Satellite DNA techniques proven, however, still under development.

**Determine *Bemisia*- natural enemy-  
host plant (Tritrophic) interactions.**

Controlled laboratory studies showed that *Bemisia* and aphelind oviposition rates varied depending on host plant. Parasitoid foraging, oviposition varied in response to different plants (crops) and host whitefly. Plants varied in color, architecture, and semiochemicals. Tri-trophic interactions determined for *B. bassiana* / SLWF / tomato. Some research completed on parasitoid / host / plant interactions.

**Identify the attributes of natural  
enemy biology and population level  
interactions to explain biological  
control successes and failures.**

The role of autoparasitism in native populations of *Encarsia* and its impact on native *Eretmocerus* has been evaluated. Results from one study show no adverse affect of *Encarsia* on overall parasitism of SLWF. No interference competition measured, with respect to whitefly control, when mixing primary parasitoids and autoparasitoids. Some life history data collected on parasitoid and predator populations in cotton. BioControl-Parasite simulation model available for testing/ validation. Some Laboratory data available for testing theoretical predictions of field level performance.

---

**Sources<sup>1</sup>**

- Pickett, C. H. 1998. Section D Natural Enemy, Ecology and Biological Control pp. 85. In: T. J. Henneberry, N. C. Toscano, T. M. Perring and R. M. Faust [eds.]. Silverleaf Whitefly National Research, Action and Technology Transfer Plan, 1997-2001 (Formerly Sweetpotato Whitefly, Strain B: First Annual Review of the Second 5-year Plan. U. S. Dept. of Agric. 1998-01, 187 pp.
- Pickett, C. H. and J. R. Hagler. 1999 Section D Natural Enemy, Ecology and Biological Control, pp. 8991. In: T. J. Henneberry and R. M. Faust [eds.]. Silverleaf Whitefly National Research, Action and Technology Transfer Plan, 1997-2001 (Formerly Sweetpotato Whitefly, Strain B: Second Annual Review of the Second 5-year Plan. U. S. Dept. of Agric. 1999-01, 195 pp.
- Hagler, J. R. and M. Ciomperlik. 2000 Section D Natural Enemy, Ecology and Biological Control, pp. 104-106, In: T. J. Henneberry, R. M. Faust, W. A. Jones and T. M. Perring [eds.]. Silverleaf Whitefly National Research, Action and Technology Transfer Plan, 1997-2001 (Formerly Sweetpotato Whitefly, Strain B: Second Annual Review of the Second 5-year Plan. U.S. Dept. of Agric., Agric. Res. Ser., 209 pp.
- Roltsch, B. and G. Simmons. 2002. Section D Enemy, Ecology and Biological Control, pp. 170-172. In: T. J. Henneberry, R. M. Faust, W. A. Jones and T. M. Perring [eds.]. Silverleaf Whitefly National Research, Action and Technology Transfer Plan, 1997-2001 (Formerly Sweetpotato Whitefly, Strain B: Fourth Annual Review of the Second 5-year Plan. U.S. Dept. of Agric., Agric. Res. Ser., 446 pp.

<sup>1</sup> Summaries of research progress abstracts or tables as listed were prepared by the indicated authors. See individual abstracts for research credits and more detail.

**Table 3 (Continued). Summary of Research Progress for Section E Host-Plant Resistance, Physiological Disorders, and Host-Plant Interactions of the 5-Year National Research, Action and Technology Transfer Plan on Sweetpotato Whitefly (SPW) Strain B (1997-2002).**

Research Approaches	Research Results
<b>Characterize resistance mechanisms and identify chemical/ morphological components, and study effects of insect adaptation.</b>	Potential sources of germplasm for SPW resistance in alfalfa, cotton, melon, cole crops, and cucurbits; and resistance to virus symptoms and silverleaf disorder in cotton and cucurbits have been identified. Selection for SPW resistant alfalfa is close to completion. Partially SPW-resistant crops including cotton, collard, and melons have been identified. Varieties of cotton and tomato with resistance or partial resistance to transmitted viruses also have been identified. In collards, the glossy leaf trait, and in cotton, the okra-leaf trait have been suggested as mechanisms of whitefly resistance. Increased levels of phenolics and peroxidase in response to plant stress have been associated with decreased whitefly performance in tomato. In <i>Datura wrightii</i> , glandular trichomes were demonstrated to be a very effective resistance mechanism to whiteflies. SPW fecundity on alfalfa was higher for alfalfa-reared whiteflies than for cotton-reared whiteflies. In cotton, glabrous-leaf trait has been demonstrated as a mechanisms of partial SPW resistance against whiteflies. The mechanism of resistance against whiteflies in a tomato variety carrying the <i>Mi</i> gene has been shown to be due to factors encountered by whiteflies before they penetrate sieve elements rather than factors in the phloem sap. During initial contact with the plant on good host to poor hosts detected little difference in occurring SPW behavior and oviposition occurred on poor hosts. SPW probably will oviposit on resistant crops and mechanisms of plant resistance will be continuously challenged.
<b>Develop molecular level techniques to produce resistant germplasm.</b>	Characterization of plant genomone was demonstrated in tomato and squash. Pathogenesis related mRNAs accumulated in response to SPW feeding on tomato leaves. SPW probing behavior indicates that host evaluation phase is dominated by probing. The natural plant products, neem seed extract, azadiractin, and extract of bitterwood, were shown to be effective insecticides against silverleaf whitefly. The natural plant products, azadiractin, was shown to be an effective insecticide against silverleaf whitefly. Tomato transformation successful for ToMoV, appeared resistant to TYLCV..
<b>Determine influence of host plant morphology, physiology and phenology on feeding behavior and competition.</b>	Morphological plant traits such as okra-leaf and large distance from leaf surface to vascular bundles in cotton, and glandular trichomes in <i>Datura wrightii</i> have been suggested as resistant characteristics. Fluctuations in amino acid concentrations over the lifespan of melon leaves appeared correlated with whitefly performance. Also in melons, group feeding by whiteflies was shown to create a nutrient sink in the plant, and thus provide the whiteflies with improved amino acid nutrition. Senescence in poinsettia reduces host plant quality for SPW. In cotton, decreased nitrogen fertilization decreases populations. In tomato, plant stress caused by fertilizer and/or water deficiency reduced host plant quality. In cotton, the okra-leaf trait and glabrous-leaf trait have been confirmed as a mechanisms of partial resistance. Factors encountered by whiteflies during their stylet penetration to vascular bundles has been shown to confer partial resistance in a tomato variety with the <i>Mi</i> gene. Phloem sap factors do not appear to play a role in this resistance. The known host plant range of SPW has been expanded to include some medicinal plants and weed species.

**Define whitefly feeding and oviposition behavior and investigate approaches for interrupting whitefly feeding and digestion.<sup>c</sup>**

SPW host evaluation was shown to be dominated by probing. Time spent in a particular behavior was affected by imidacloprid when the whitefly came into contact with the chemical in its diet rather than on the leaf surface. Intercropping of resistant within susceptible cole crops did not lessen the abundance of whiteflies. Improvements have been made in a system for rearing whiteflies on an artificial liquid medium. This will allow direct experimentation on the role of specific plant nutrients and allelochemicals on whitefly feeding and performance. Stylet contact with minor vascular bundles is essential for successful whitefly feeding on cotton. The fine structure of whitefly eggs and their attachment to host leaves has been studied with electron microscopy, and the distal end of the egg petiole that is inserted into the host leaves possesses morphological structures that suggest a role in water uptake from the host leaf which is a very important process for egg survival. Morphological studies on whitefly stylets indicate that they are sufficiently long to reach minor vascular bundles (the major feeding site) from virtually any place on the abaxial leaf surface of cotton. This makes variation in vascular bundle depth an unlikely mechanism of resistance to whiteflies in cotton. Variation in nitrogen fertilization has been shown to decrease amino acid concentrations in phloem sap and thus affects nutrition available to whiteflies. Whitefly feeding differentially induces pathogenesis-related (PR) proteins in two cucurbit species, cantaloupe and watermelon, and apparently is not affected by treatment with plant growth-promoting rhizobacteria (PGPR).

**Study whitefly toxicogenic plant reactions.**

Research on tomato identified a gene that is specifically induced by whitefly feeding. Four classes of genes were identified in inducing squash leaf silvering. These genes were further characterized by hybridization, sequence analysis and complementation studies. Genes specifically induced by whitefly feeding have been identified in tomato and in squash. These genes may play a role in the plant's defensive response to the whitefly and/or the plant's toxicogenic reaction such as irregular ripening in tomato and silverleaf symptom in squash. Two genes, one of which appears to be a general plant defense, have been shown to be differentially induced in squash by silverleaf and sweetpotato whiteflies. This may be related to the different toxicogenic effects of these two whitefly species on squash. The activation of these genes is systemic. In tomatoes, feeding by both silverleaf whitefly and greenhouse whitefly induced pathogenesis related genes, but not genes regulated by the octadecanoid pathway. These studies indicate that tomato plants perceive phloem-feeding silverleaf and greenhouse whiteflies in a manner distinct from that of chewing insects.

---

**Sources<sup>1</sup>**

- Natwick, E. 1998. Section E Host Plant Resistance, Physiological Disorders, and Host Plant Interactions, pp. 103. In. T.J. Henneberry, N. C. Toscano, T.M. Perring and R. M. Faust [eds.]. Silverleaf Whitefly National Research, Action and Technology Transfer Plan, 1997-2001 (Formerly Sweetpotato Whitefly, Strain B: First Annual Review of the Second 5-year Plan. U.S. Dept. of Agric. 1998-01, 187 pp.
- Walker, G. 1999. Section E Host Plant Resistance, Physiological Disorders, and Host Plant Interactions, pp. 109-110. In. T.J. Henneberry and R.M. Faust [eds.]. Silverleaf Whitefly National Research, Action and Technology Transfer Plan, 1997-2001 (Formerly Sweetpotato Whitefly, Strain B: Second Annual Review of the Second 5-year Plan. U.S. Dept. of Agric. 1999-01, 195 pp.
- Walker G. and C. McKenzie. 2000. Section E Host Plant Resistance, Physiological Disorders, and Host Plant Interactions, pp. 123-126. In. T.J. Henneberry, R.M. Faust, W.A. Jones and T.M. Perring [eds.]. Silverleaf Whitefly National Research, Action and Technology Transfer Plan, 1997-2001 (Formerly Sweetpotato Whitefly, Strain B: Second Annual Review of the Second 5-year Plan. U.S. Dept. of Agric., Agric. Res. Ser., 209 pp.
- McKenzie, C. and G. Walker. 2002. Section E Host Plant Resistance, Physiological Disorders, and Host Plant Interactions, pp. 195-197. In. T.J. Henneberry, R.M. Faust, W.A. Jones and T.M. Perring [eds.]. Silverleaf Whitefly National Research, Action and Technology Transfer Plan, 1997-2001 (Formerly Sweetpotato Whitefly, Strain B: Second Annual Review of the Second 5-year Plan. U.S. Dept. of Agric., Agric. Res. Ser., 446 pp.

<sup>1</sup> Summaries of research progress abstracts or tables as listed were prepared by the indicated authors. See individual abstracts for research credits and more detail.

**Table 3 (Continued). Summary of Research Progress for Section F Integrated and Areawide Pest Management Approaches, and Crop Management Systems. of the 5-Year National Research, Action and Technology Transfer Plan on Sweetpotato Whitefly (SPW) Strain B (1997-2002).**

Research Approaches	Research Results
<p><b>Development:</b>  <b>Study whitefly-crop interactions as cultural components that affect population dynamics, e.g., water, nutrients, plant population, planting/termination/harvest dates, other farm practices, intercrop relationships.</b></p>	<p>The roles of soil fertility (nitrogen), water-stress &amp; some other agronomic factors on <i>Bemisia</i> population dynamics have been identified. Cross-commodity integration of pesticides used across multi-crop ecosystems and potential impact on whitefly management were developed. Progress was made with studies in Texas on seasonal dynamics of <i>Bemisia</i> on spring collards, and the impact of cotton defoliants on <i>Bemisia</i> and parasitoid populations. Economic study in Arizona showed &gt; 80% reduction in pink bollworm-SPW IPM systems. Crop-free was demonstrated to break the tomato yellows leaf curl virus cycle.</p>
<p><b>Develop behavioral barriers to whitefly colonization and population development, e.g., mulches, trap crops, inter-cropping, row covers, etc.</b></p>	<p>Row covers and screens as physical barriers, mulches and oils as behavioral barriers, living mulches as behavioral barriers have proven useful in some situations. Investigations on intercropping took place in both desert and tropical environments. Approaches have also been identified as having potential.</p>
<p><b>Integration:</b>  <b>Develop Integrated Pest Management systems using dual or multiple control tactics, e.g., cultural, biological, chemical, host plant resistance, etc.</b></p>	<p>1) Insect Growth Regulators &amp; biological control in cotton (conservation), 2) imidacloprid &amp; other chemical control tactics &amp; various forms of biological control, especially in vegetables, 3) studies of direct &amp; indirect effects of chemical control on bio-control agents. A number of field studies employed multiple tactics directed against whitefly populations. Biorational insecticides were examined in combination with IGRs and other biopesticidal agents such as <i>Beauveria bassiana</i> for control efficacy of silverleaf whitefly. There was an indication of inhibitory action by <i>B. bassiana</i> when used in combination with imidacloprid as well as deleterious effects to predators contacted by <i>B. bassiana</i> treatments. Neem products were used to reduce whitefly populations and incidence of yellow mosaic virus in India. A melon trap crop was integrated with chemical control to focus potentially disrupting treatments into a limited area while preserving natural mortality factors in cotton as the principle crop. Much progress with integrating control tactics: 1) Life table evaluation of both natural and insecticide-based mortalities, 2) Compatibility of IGR's for whitefly control in greenhouses, 3) IPM development in cotton for <i>Bemisia</i> and other cotton pests, 4) Augmentative biocontrol using crop transplants inoculated with parasitoids in Admire<sup>®</sup>-treated fields.</p>
<p><b>Integrate sampling with other key components of IPM systems, e.g., thresholds, economics, decision-making, biological control, etc.</b></p>	<p>1) <i>Bemisia</i> distributions have been examined in tomato, 2) new binomial sampling system for large nymphs in cotton, &amp; integration with thresholds for IGR decisions, 3) sampling &amp; IGR re-treatment decisions tested in cotton. Analysis of types and patterns of chemical treatments made on a large number of cotton fields in central Arizona over a 4 year period revealed extraordinary differences in the number of treatments and amount of time that whiteflies exceeded threshold levels prior to and following the advent of the IGRs buprofezin and pyriproxyfen. The proactive initiative taken by Arizona growers to pursue chemical use harmonization across commodities required consideration of all aspects of pest and crop management. A similar whole system appraisal was made in the San Joaquin Valley with an emphasis on integrating multiple practices with diverse insecticide classes as part of an insecticide resistance management program. Fourth consecutive year of monitoring <i>Bemisia</i> populations in the Imperial Valley using the CC trap. Sampling-based refinement of action thresholds for IGR's in Arizona cotton show better conservation of predators with high threshold (10 vs. 5).</p>



**Delivery and Implementation:**

**Elevate single field/farm practices to areawide community-based contexts; develop methodology for installing and evaluating areawide control technologies and their impact.**

Areas dominated by cotton production were identified in AZ & CA for implementation of cooperative programs. Areas of melon and vegetable production were identified in TX for potential area-wide programs. Area-wide sampling, & decision-making was the main focus of most programs; however, coordinated natural enemy releases were also conducted. Large areas in the San Joaquin Valley observed specific guidelines for IPM and IRM in cotton with evaluations continuing on the benefits attained over areas that did not observe these guidelines. Community wide evaluations were made on quality of whitefly management according to chemical control practices. The successful IPM and IRM programs practiced in Arizona cotton continued for a third consecutive year. Further cross-commodity development of these programs is under way. Economic analysis of the use of IGR's in Arizona cotton.

**Implement and deliver Integrated Pest Management and Integrated Crop Management systems or system components to clientele.**

IPM recommendations were distributed in AZ, CA, Mexico & FL. Bilateral discussions between Brazil & U.S. took place. A 'best agricultural practices' demonstration project was conducted on 50.5 acres (Univ. of AZ Maricopa Agricultural Center) with inputs from agronomy, entomology, irrigation management, weed sciences and plant pathology. SPW management was fully integrated with management of other insect pests and required only a single application of pyriproxyfen. Lint yields of 2.81 bales/acre were higher than the historical as well as the 1998 farm-wide average. An integrated areawide management program involving the cooperation of growers, PCAs, ginners and state and university researchers was expanded during a second year in the San Joaquin Valley. Sticky cotton bulletin published by University of Arizona and Cotton Incorporated. International development of IPM for managing whiteflies and geminiviruses. Development of *Bemisia*-resistant alfalfa cultivars. Crop and pest management demonstration project on cotton in Arizona.

---

**Sources<sup>1</sup>**

- Ellsworth, P. C. 1998. Section F Integrated and Areawide Pest Management Approaches and Crop Management Systems, pp. 115-116. In. T. J. Henneberry, N. C. Toscano, T. M. Perring and R. M. Faust [eds.]. Silverleaf Whitefly National Research, Action and Technology Transfer Plan, 1997-2001 (Formerly Sweetpotato Whitefly, Strain B: First Annual Review of the Second 5-year Plan. U.S. Dept. of Agric. 1998-01, 187 pp.
- Castle, S. 1999. Section F Integrated and Areawide Pest Management Approaches and Crop Management Systems, pp. 125-127. In. T. J. Henneberry and R. M. Faust [eds.]. Silverleaf Whitefly National Research, Action and Technology Transfer Plan, 1997-2001 (Formerly Sweetpotato Whitefly, Strain B: Second Annual Review of the Second 5-year Plan. U.S. Dept. of Agric. 1999-01, 195 pp.
- Castle, S. and B. Roltsch. 2000. Section F Integrated and Areawide Pest Management Approaches and Crop Management Systems, pp. 141-142. In. T. J. Henneberry, R. M. Faust, W. A. Jones and T. M. Perring [eds.]. Silverleaf Whitefly National Research, Action and Technology Transfer Plan, 1997-2001 (Formerly Sweetpotato Whitefly, Strain B: Second Annual Review of the Second 5-year Plan. U.S. Dept. of Agric., Agric. Res. Ser., 209 pp.
- Castle, S and P.C. Ellsworth. 2002. Section F Integrated and Areawide Pest Management Approaches and Crop Management Systems, pp. 213-215. In. T. J. Henneberry, R. M. Faust, W. A. Jones and T. M. Perring [eds.]. Silverleaf Whitefly National Research, Action and Technology Transfer Plan, 1997-2001 (Formerly Sweetpotato Whitefly, Strain B: Fourth Annual Review of the Second 5-year Plan. U.S. Dept. of Agric., Agric. Res. Ser., 446 pp.

<sup>1</sup> Summaries of research progress abstracts or tables as listed were prepared by the indicated authors. See individual abstracts for research credits and more detail.

## II. Plenary Session Keynote Address Summaries

### Section A: Plenary Session Summary

Dale B. Gelman, Michael B. Blackburn, Jing S. Hu and  
\*D. Gerling

USDA, ARS, Insect Biocontrol Laboratory, Beltsville,  
MD, USA; \*Department of Entomology, Tel Aviv  
University, Ramat Aviv, 69978, Israel

#### **Whitefly Metamorphosis—Timing, Regulation and Influences on the Development of its Parasitoid, *Encarsia formosa***

A staging system based on body depth (thickness) and maturation of the adult eye was used to track development and identify physiologically synchronous 4<sup>th</sup> instar/pharate adult silverleaf (SLWF) and greenhouse (GHWF) whiteflies. The median depth of Stages 1, 2, 3 and 4 is 0.1, 0.15, 0.2 and 0.25 mm, respectively, and the depth of Stage 5 is  $\geq 0.27$  mm. As the whitefly progresses through Stages 6, 7, 8, and 9, the eye changes from a small, intense red dot (Stages 1 – 5) to a diffuse dot (Stage 6), to a light red adult eye (Stage 7), to a medium bipartite red (Stage 8) and finally dark red or red-black bipartite adult eye (Stage 9). An examination of histological sections of last instar SLWFs revealed that adult eye and wing development were initiated in Stage 6, the stage in which external examination showed that the eye had just begun to undergo pigment diffusion. In contrast, adult eye and wing development of GHWFs was first observed in Stages 4 or 5, a time prior to the initiation of eye pigment diffusion. Wings had become deeply folded by Stages 6 and 7 in GHWF and SLWF, respectively. Although adult development was initiated later in the SLWF than in the GHWF, metamorphosis occurred at a very rapid rate in both species of whitefly. Within approximately 24 h, the simple bi-layered wing bud developed into a wing of nearly adult proportions, and within an additional 12–24 h, the nymphal eye and wing bud had been replaced by the well-differentiated eye and wing of the whitefly adult. Ecdysteroid titers, as measured by a very sensitive ELISA (range = 500 – 40,000 fgs), peaked at Stages 4 - early 6 in the SLWF and at Stages 4 - 5 in the GHWF, a time just prior to or at the onset of adult development. It is noteworthy that at their peak, ecdysteroid titers were three to four times greater in the SLWF than in the GHWF, although for both species, titers were 100 to 1000 times less than those reported for species from other orders of insects. The identity of the host plant did not significantly affect the magnitude or timing of the ecdysteroid peak in SLWF last instars. However, leaf pubescence did impact the length, width and depth dimensions of 4<sup>th</sup> instar/pharate adults. When reared on the pubescent leaves of green beans and tomatoes the period of whitefly development prior to becoming pharate adults could be divided into five stages with depth reaching a maximum of 0.25 – 0.3 mm during

stages 4 – 5. In contrast, when reared on the glabrous leaves of cotton, collard or sweet potato, whitefly body thickness rarely surpassed 0.17 mm, a thickness equivalent to Stage 2. However, whitefly nymphs on glabrous plants were significantly longer and wider than those developing on pubescent leaves. Thus, it appears that Stages 3 – 5 of nymphs that are reared on pubescent leaves are developmentally similar to Stage-2 nymphs that are grown on glabrous leaves.

The critical feeding period (CFP), the time after which 50% of whiteflies can be removed from the leaf and complete development, i.e., emerge, was also determined. Stages 1 through 9 were removed from the leaf, maintained under high humidity and monitored for percent adult emergence. It was surprising that although adult development was initiated earlier in GHWFs than in SLWFs, the CFP occurred at a later stage in the GHWF. At 25°C and L : D 16 : 8, more than 50 % of test GHWFs and SLWFs successfully completed development if removed from the leaf was performed on or after whiteflies reached Stages 4 and 3, respectively. When removed at Stage 2, adult emergence was observed in 40% of SLWFs but in no GHWFs.

It was confirmed that the developmental rates of *Encarsia formosa* differed significantly based upon the whitefly instar parasitized. Development was the most rapid and synchronous when 3<sup>rd</sup> and young 4<sup>th</sup> instar whiteflies were parasitized. In addition, adult longevity was greatest and emergence was most synchronous when these older instars were selected for oviposition. It is significant that our results indicate that no matter which GHWF instar is parasitized, the wasp does not molt to its last instar until the host has reached Stage 4/5 of its last instar. It appears, then, that a condition(s) associated with host adult formation may be required for the parasitoid's final molt.

### Section B: Plenary Session Summary

Judith K. Brown, Department of Plant Sciences,  
The University of Arizona, Tucson, AZ 85721

#### **Diversity of Begomoviruses and the Whitefly Vector in the US Sunbelt States: An Overview**

Recent upsurges in populations of the whitefly *Bemisia tabaci* (Genn.) have drawn much attention to its worldwide importance as an insect pest and as the vector of emergent begomoviruses (Family: *Geminiviridae*; Genus: *Begomovirus*). Several begomoviruses that are considered 'new' and others previously regarded as minor pathogens have been linked to recent epidemics. Recent studies have revealed much variation in begomoviruses, despite the view that DNA-containing viruses do not rapidly accumulate mutations. Also, certain *B. tabaci* 'variants' are known that more effectively or

selectively transmit certain begomoviruses and exhibit biotic differences that may influence their spread. Patterns of distribution and dissemination of begomoviruses transmitted by *B. tabaci* are poorly understood because standardized molecular-based tracking methods have not been available. Understanding virus/whitefly vector/host plant interrelationships in the context of emerging problems can be achieved only by linking predicted evolutionary histories with epidemiology using molecular phylogenetic approaches. To address etiology and investigate the molecular epidemiology and ecology of begomoviruses, we have employed a molecular marker approach targeting key viral sequences that enable provisional virus identification. Polymerase chain reaction (PCR) and degenerate primers have been designed to amplify the 'core' region of the viral coat protein gene (core CP) of virtually all begomoviruses (Genus: *Begomovirus*; Family: Geminiviridae) (Wyatt and Brown, 1996). Subsequent 'positives' are subjected to additional PCR and sequence analysis using a suite of degenerate primers to target additional informative sequences. We applied this approach to identify and map the global distribution of begomoviruses. Analysis of the begomoviral CP sequence reveals an Old and New World clade. Old World begomoviruses are further separated based on major geographical barriers. In contrast, New World begomoviruses cluster as a single, large polytomy containing certain distinctive lineages that are somewhat representative of geographical boundaries in the region; however, definitive barriers are less apparent than for Old World exemplars. Within-cluster divergence for New World viruses is somewhat less than for Old World viruses, suggesting that extant begomoviruses have evolved from fewer founder events than occurred for the Old World viruses, or that only a few lineages survived despite multiple founder events. All begomoviruses are transmitted by a single species of whitefly, *Bemisia tabaci* (Genn.) (Aleyrodidae: Hemiptera). A species complex has been proposed to encompass whiteflies identifiable as *B. tabaci*, which is rich in biological variants that lack distinguishing morphological characteristics. We have previously shown that the mitochondria cytochrome oxidase I (mtCOI) gene is useful for studying genetic variation within this group. A fragment (~800 bp) of the mtCOI gene was amplified using polymerase chain reaction and degenerate primers and sequence analysis reveal numerous clusters (or clades) with a basis in geographical origin. New World *B. tabaci* cluster as two groups: (1) North/Central America & the Caribbean region and (2) South America, which exhibited <2-6% divergence, while eight or more Old World clades are well-supported at ~8-12+% divergence from other Old clusters and from the two New World clades as well. These results provide no

overwhelming rationale for erecting the B biotype as a distinct *Bemisia* species, in light of the extensive, but as yet, poorly understood basis for the diversity evident in this complex. Also, begomoviral CP and whitefly vector mt COI trees are generally concordant with respect to extant geographical origin, suggesting that markers are also reliable predictors of present-day distributions of viral or whitefly genotypes. Interestingly, virus-vector relationships corroborate a tight evolutionary relationship between all *B. tabaci* and the Genus: *Begomovirus* that is thought to have a basis in virus-vector specificity. Indeed, the CP gene is the most conserved among begomoviral ORFs. This observation, together with the demonstrated congruence between vector and virus trees, reflects a co-evolved relationship between the whitefly vector and viral capsid protein and confers properties necessary for vector-mediated transmission.

Robert L. Gilbertson  
University of California, Davis

#### **Tomato yellow leaf curl virus in the Dominican Republic-A case study**

Tomato yellow leaf curl disease is one of the most devastating diseases of tomato (*Lycopersicon esculentum*). Infected plants show stunted and erect or upright growth and leaves with upward curling, crumpling and interveinal chlorosis. Yield losses of 100% are common, particularly if plants are infected at an early stage in development. The disease is caused by the whitefly-transmitted geminivirus (Genus *Begomovirus*, Family *Geminiviridae*), *Tomato yellow leaf curl virus* (TYLCV). TYLCV is transmitted by *Bemisia tabaci* and *B. argentifolii* (*B. tabaci* biotype B) in a persistent manner. It is a phloem-limited virus that is not transmitted via seed or by sap-inoculation. TYLCV was first described in Israel in 1931 and has been reported from many countries around the Mediterranean Basin and Africa. It is a limiting factor for tomato production in countries such as Spain, Cyprus, Jordan, Israel, Egypt, Senegal, Mali, Thailand, and India. Moreover, it has been established that tomato yellow leaf curl disease is caused by more than one begomovirus species, depending on the geographical location. The name TYLCV now refers to the monopartite begomovirus that causes TYLCV in Israel.

TYLCV had not been reported from the Western Hemisphere but, in the early 1990s, tomatoes in the Caribbean island, the Dominican Republic (DO), developed disease symptoms that were similar to those caused by TYLCV. Using the polymerase chain reaction (PCR) and a pair of TYLCV-specific primers designed to direct the amplification of a full-length TYLCV DNA fragment, the expected size DNA fragment (~2.8 kb) was amplified from tomato tissues showing TYLCV

symptoms from the DO. This DNA was cloned and sequenced. DNA sequence analyses revealed that the cloned DNA was nearly identical (97%) to TYLCV and that the genome organization of the Dominican Republic TYLCV (TYLCV-DO) was identical to that of TYLCV. Using squash blot hybridization analysis and the cloned TYLCV-DO as a probe, it was established that TYLCV had spread throughout the major tomato growing areas of the Dominican Republic and that the primary host was tomato. Thus, TYLCV had been inadvertently introduced into the DO and it had spread rapidly, presumably via already existing *B. tabaci*/*B. argentifolii*. TYLCV was not detected in weeds with or without symptoms, with the exception of *Datura stramonium* plants that showed leaf curl and interveinal yellowing.

As part of a TYLCV management strategy, a whitefly host-free period was instituted in the two major processing tomato growing areas (one in the North and one in the South) in the DR. To assess the efficacy of this approach, a PCR-based method for assessing the incidence of TYLCV in whiteflies was used to monitor TYLCV incidence over time. By the end of the tomato growing season, TYLCV was detected in nearly all of the whiteflies collected from tomato fields in the North and the South. However, during the host-free period, the incidence of TYLCV decreased dramatically in whiteflies (now collected from weeds around the fields). Thus, the whitefly host free period resulted in a considerable decrease in the incidence of TYLCV in the DO. More importantly, the host free period provided a lag period of 6-8 weeks before newly planted tomatoes developed TYLCV symptoms.

Thus, the whitefly host-free period resulted in a dramatic decrease in the incidence of the virus, but it was able to persist during the host free period. Using squash blot hybridization analysis of numerous crop and weed plants in and around TYLCV-infected tomatoes, very few potential TYLCV host were identified. Interestingly, while pepper plants commonly showed symptoms of virus infection in the DO, few were positive for TYLCV, even in cases where pepper fields were adjacent to heavily infected tomato fields. An exception to this was a pepper field in the North in January 2000 that had high whitefly populations and numerous plants with stunted growth and upcurled and yellowed leaves; TYLCV was detected in these plants by PCR. Thus, TYLCV can infect pepper in the DO; the low frequency may relate to host preference of the whitefly vector. TYLCV infection of common bean in the DR was demonstrated by planting seed of cv. Topcrop near a tomato field with a high incidence of TYLCV infection. Many bean plants showed stunted growth, curled and crumpled leaves and chlorosis; these plants were infected with TYLCV based on PCR analysis. Interestingly, it appears that the smaller seeded beans (e.g., pinto bean, cv. Othello) may develop symptomless infections. Thus, beans are also a host for TYLCV in the DR. These results demonstrate that it is important to

include these crops as well as tomato in the host-free period.

Squash blot hybridization failed to show TYLCV infection in most weeds. However, TYLCV was detected in weeds using the more sensitive PCR approach. The virus was detected in a number of common weeds, including *Croton lobatus*, *Cleome viscosa* and *Solanum nigrum*. Thus, TYLCV may persist in weeds as a symptomless infections; the inefficient nature of these infections may explain why it takes weeks before newly planted tomatoes develop TYLCV infections via inoculum from weeds.

Management of TYLCV in the DR involves an integrated approach using the whitefly host-free period, whitefly control (e.g., imidicloprid), and the use of early maturing hybrids and resistant varieties. Using this approach, levels of tomato production in the DR have returned to pre-TYLCV levels.

### Section C: Plenary Session Summary

Nilima Prabhaker  
Department of Entomology, University of California,  
Riverside

#### Insecticide resistance in *Bemisia tabaci*: Past expectations and present reality

Pesticide resistance is considered to be one of the limiting problems in agriculture and human health. It is therefore critical that we understand the nature of resistance and the steps that can be taken to avoid its occurrence. This is especially true for a handful of recidivist organisms that repeatedly develop resistance to novel chemistries in agricultural systems and environments all over the world. *Bemisia tabaci* is one such organism that has a well-documented legacy of resistance to all major classes of insecticides. Moreover, populations of *B. tabaci* in some parts of the world have already developed high levels of resistance to novel compounds that have proven so effective in the southwestern USA. Because insecticides remain a primary control option against *B. tabaci* in the USA and much of the world, there remains a high potential for further cases of resistance development. However, the rate at which resistance develops will depend upon genetic, biological/ecological and operational factors. The importance of some of these factors will be considered with reference to the case of whiteflies in California and other regions of the world. The current status of insecticide resistance in *B. tabaci* to various insecticides in various regions of the world will be discussed. More importantly, the differences among regions will be compared and contrasted to ascertain the principle factors that determine whether resistance development will be a significant or trivial occurrence.

Resistance to organophosphates and pyrethroids in *B. tabaci* was reported during the early 1980s in Sudan and California. Additional examples of resistance to OPs and pyrethroids were identified in the following years at a time when choice of chemistry was much more limited than at the present. Less effective, more broad-spectrum chemistry used repeatedly as the only recourse to increasing whitefly populations would certainly represent an untenable situation from a resistance management standpoint. It is little surprise that the resistance examples of the 1980s were all related to cotton production, a long-season crop that relies heavily on insecticides. In the California example, the Type A strain of *B. tabaci* was principally a cotton pest in that its population growth cycle occurred in cotton and not on a series of crops prior to and in addition to cotton, as was to become the case for the Type B strain. Consequently, refuge areas for preserving susceptible genes were not part of the set of ecological factors that would become so important for the Type B strain during the 1990s in California.

The most significant change that took place in the 1990s was the invasion of North America by the Type B strain of *B. tabaci*. Although this had been initiated as early as 1985 in Florida, it wasn't until the late 1980s that signs of the Type B strain were apparent in Arizona and California. After simmering for some period of months and perhaps years in the desert agricultural valleys, *B. tabaci* Type B erupted in 1991 in the Imperial Valley in a sensational display of biotic force. What had been mostly a late season cotton pest and vector of viruses to fall vegetables in the form of Type A was now seemingly transmuted to a year round pest of cotton, fall and winter vegetables, and spring melons in the form of Type B. This meant heavier use of insecticides on multiple crops, and from what people assumed based on unmanageable numbers of whiteflies, high levels of insecticide resistance.

A resistance-monitoring program was initiated in the Imperial Valley in 1992 to determine relative susceptibilities of whiteflies to the various insecticides that were being used on crops year round. Additionally, cross-resistance patterns between chemistries and the influence of management strategies to maintain long-term viability of products were also studied in the laboratory and the field. Results of four years of whitefly monitoring data to bifenthrin, endosulfan, chlorpyrifos and methomyl showed no progression towards higher resistance in spite of heavy dependence on these compounds. Instead, whiteflies were less sensitive in 1993 compared to the following 2-3 years as indicated by lower LC<sub>50</sub>s, thereby demonstrating reversion rather than progression of resistance. This was not the case for all whitefly treatments used in the Imperial Valley. A comparison of whitefly responses to a mixture of danitol®+orthene® from 1994 to 1999 showed some loss of susceptibility between 1994 and 1997, but stabilized through 1999. However, the modest shift towards

reduced susceptibility observed in the Imperial Valley contrasts sharply with the high resistance levels to danitol+orthene observed in central Arizona. A year-by-year comparison between 1995 and 1998 consistently showed a flatter response to increasing concentrations of danitol+orthene for Maricopa whiteflies compared to whiteflies collected in the Imperial Valley, even though whiteflies from both regions were collected from cotton that received a similar number of danitol+orthene treatments. The fact that whiteflies from different regions, but same crop, responded so differently to similar exposures of insecticides argued strongly for the importance of the different ecological conditions under which each population was exposed. Insecticide selection pressure for Maricopa whiteflies in terms of the proportion of the regional population exposed to repeated treatments of danitol+orthene was almost certainly much higher than that proportion of the Imperial Valley regional population exposed to danitol+orthene. This is due to the proportionally low acreage of cotton grown in the Imperial Valley relative to other crops such as alfalfa as well as ornamentals grown at homes and businesses in the more densely populated Imperial Valley. In contrast, cotton is the highest acreage summer crop grown in central Arizona, and this region is sparsely populated by humans. Thus, whiteflies would be concentrated in cotton and would therefore receive a greater exposure quota of insecticide treatments as a function of the total regional population. It is somewhat analogous to the situation in the Imperial Valley in the 1980s where the concentration of the A type in cotton led to greater proportional exposure of the whole population with little refuge or conservation of susceptible genotypes.

The neonicotinoid insecticides have become an indispensable group of compounds for combating whiteflies on crops grown worldwide. Monitoring data of 3 neonicotinoids, acetamiprid, imidacloprid and thiamethoxam in 1999 and 2000 showed variations from field to field, however, the LC<sub>50</sub>s were extremely low (<10 ppm) and gave no indication of reduced sensitivity. An interesting trend was evident in our study which showed highest survivorship to imidacloprid occurring with whiteflies collected from fall melons. The highest susceptibility to imidacloprid was during early fall when insects were collected from cotton compared to whiteflies collected later in the fall on broccoli plants. These results confirm the interesting phenomenon of shifting levels of susceptibility depending on crop, type of treatments applied, and other ecological events that may interact to maintain an overall high level of susceptibility.

Comparative bioassay results with imidacloprid on whiteflies from Guatemala collected on melon crops compared to the Imperial valley insects indicated a plateauing of mortality at the higher concentrations with <80% mortality at 100 ppm suggesting that they may be resistant to this compound. Similarly insects from Almeria appear to be resistant to imidacloprid in addition

to some other chemistries. A comparison from the literature on whiteflies from India also report resistance to imidacloprid and pyrethroids. A number of factors that may have influenced the resistance pattern to insecticides as evident by the differences in responses of whiteflies from various parts of the world are compared and discussed with emphasis on agro-ecosystem, crop rotation and insecticide use patterns as well as the refugia concept.

An analysis of chemical control has shown the importance of research and development of resistance management programs' for continuing use of insecticides for whitefly control during the last decade. An evaluation of the benefit of experience gained over the last decade with this species has proven that whitefly specialists are better able to assess the resistance risks based on the ecological conditions of the agro-environment with help of new chemistries with novel modes of action. Now we have proven formulas for managing resistance based on examples from Arizona and Israel.

#### **Section D: Plenary Session Summary**

Don C. Vacek

USDA-APHIS-PPQ Mission Plant Protection Center, P.O.  
Box 2140, Mission, TX 78573-2140

#### **Application of Genetic Diagnostics to Biological Control of Silverleaf Whitefly**

Genetic technology has had a significant impact on the history and delivery of the biological control program against silverleaf whitefly. Early challenges in identification of the imported natural enemies were met with a specific genetic diagnostic, RAPD-PCR. Later taxonomic studies corroborated the genetic data. Genetic patterns assigned to natural enemy collections from all over the world set a precedent for facilitating mass rearing, quality control, ecological studies, and field evaluation of natural enemies. MPPC imported and cultured over 56 populations of *Encarsia* spp. and *Eretmocerus* spp., several of which were new species. Parasitoids in quarantine were categorized with RAPD-PCR and morphologically based systematics. Integration of the two techniques proved to be useful in capturing the maximum amount of species diversity with a minimum amount of duplication in cultures. Rearing personnel needs were reduced by 11 service years over a five-year period and insect quarantine space was used effectively. In field evaluation efforts, the two methods were integrated for identifying indigenous and imported parasitoids. Cooperators have initiated an additional technique to identify specific satellite DNA probes for development into a squash blot kit that could alleviate the cost and time constraints of RAPD assays.

*Eretmocerus* and *Encarsia* were separated into distinct groups using the morphology of the pupae and adult females. Individuals from each unique accession were

immediately characterized at the MPPC Genetics Diagnostic Laboratory using RAPD-PCR with primers C04 and A10 from Operon Technologies. Cohorts of the original parental material were sent to cooperating systematists. Information from the collaborating systematists and geneticists allowed for characterization of quarantine material while the original parental cohort was still alive. Typically, material was characterized using both methods within two to three days after acceptance into quarantine. Based on RAPD patterns, unique parasitoid accessions (possible cryptic species) were set up in pure cultures and reared on the local SLWF. The integration of the morphology-based systematics and the molecular techniques has optimized the efficient characterization of natural enemies in quarantine. The integration has allowed each new cryptic species to be fully evaluated and maintained as a pure culture while cooperating systematists continue to access the genetically unique accessions. Apart from differences in RAPD patterns, new species were only distinguishable from each other by minute differences in the first funicular antennal segment of the female. Screening with RAPD assays was really the only method available for distinguishing such cryptic species at the time of culture initiation and efficacy studies. Without RAPD assays, multiple cultures of widely distributed species, such as *E. mundus* would have used most of the quarantine resources.

The techniques were also equally valuable in field-testing. It was possible to test multiple species simultaneously in the field and sort out species in the recovery samples by their DNA profile. Combining the use of RAPD-PCR and classical systematics allowed for an increase in the numbers of individuals that could be characterized to species in the evaluation effort. Ultimately this led to the best possible determination of which species showed the greatest efficacy in the field. The molecular methods were only valuable when combined with classical systematics. Properly identified and curated specimens will provide the best permanent record of the species released and established. Representatives of all the accessions were cryogenically stored at MPPC and vouchered at the Texas A&M University, Department of Entomology Collection, College Station, Texas and the USDA-ARS, Systematic Entomology Laboratory, Washington, D.C.

## Section E: Session Summary

G. P. Walker

Department of Entomology, Univ. CA, Riverside

### Overview of Whitefly Feeding Behavior: What We Know and What We Need to Know

#### Mechanical Movement of the Stylets

The mouthparts of whiteflies are typical of the order Homoptera. The mandibles and maxillae are modified as very thin and elongate stylets that pierce into the plant tissue. The paired maxillary stylets are tightly joined together along their length with an interlocking tongue-and-groove coupling. The tongue-and-groove design provides a very tight coupling while allowing each maxillary stylet to move forward and back independent of one another. The paired maxillary stylets enclose a food canal and salivary canal, and their primary function is ingestion and salivation. The mandibular stylets fit on either side of the joined maxillary stylets, and enclose the maxillary stylets on their lateral aspect. The mandibular stylets are serrated at their tip, and their main function is to penetrate through the plant tissue. The maxillary stylets, with their food and salivary canals, penetrate the plant tissue along with the mandibular stylets, but it is primarily the mandibular stylets that do the mechanical work cutting and penetrating through the plant tissue.

Each stylet is operated by a pair of antagonistic muscles in the head: a protractor muscle that pushes the stylet deeper into the plant tissue, and a retractor muscle that pulls the stylet back. The four stylets can be moved independently of one another by their individual protractor and retractor muscles. The protractor and retractor muscles are short, and thus each muscle contraction can advance or retract their respective stylet only a short distance. Consequently, stylet advancement is incremental and consists of a repeating series of short distance advancements of the stylets. First, the mandibular stylets are advanced a short distance, followed by the maxillary stylets. Stylets also can be withdrawn from the plant tissue in an incremental manner similar to stylet advancement, or they can be withdrawn more rapidly by a mechanism described later. The mandibular stylets are naturally curved inward (medially) which facilitates turning of the stylets in the plant tissue. Because of their medial curvature, the left mandibular stylet tends to turn to the right in the plant tissue while the right mandibular stylet tends to turn left. If the left and right mandibular stylets advance simultaneously, their tendency to curve in opposite directions counteracts each other, and the stylets advance straight into the plant tissue. If the left and right mandibular stylets do not advance simultaneously, the stylets will turn in the plant tissue. For example, if the left mandibular stylet advances by itself, it will turn right in the plant tissue; then the right mandibular stylet advances, and is forced to turn to the right by following the pathway previously cut by the left

mandibular stylet. A turn to the left is accomplished by a similar, but vice versa manner. The stylets are capable of turning only left and right. To allow 360° of movement of the stylets, the whitefly has to pivot its body around the point of stylet insertion. This rotates the stylets in the plant tissue so that the left-right plane of the stylets is rotated into any position, thus allowing the stylets to turn in any direction. The stylets are remarkable flexible, and microscopic examination of stylets in plant tissue reveals that they twist and turn in virtually every direction, making the stylet pathway usually very tortuous.

In adult whiteflies, the labium is elongate, four-segmented, and has a longitudinal groove that encloses the stylets when the insect is not feeding. When not feeding, the stylets extend the entire length of the labial groove. During stylet penetration, the apex of the labium is pressed against the plant surface and the protractor muscles begin to advance the stylets into the plant tissue. As the stylets are advanced, the head is lowered closer to the plant surface and the labium shortens, allowing the stylets to extend past the tip of the labium and into the plant tissue. The labium does not penetrate the plant tissue. The length of the stylets in the plant tissue equals the length that the labium shortens, so the length of stylet penetration can be indirectly measured by measuring the length of labial shortening. Note that since the pathway of the stylets in the plant tissue is usually very tortuous rather than linear, the length of stylet penetration is greater than the depth of stylet penetration (depth = distance perpendicular from the plant surface to the apex of the stylets).

A long stout apophysis, called the crumena, extends internally from the apical labial segment into the head capsule. Powerful crumena protractor muscles attach to the base of the crumena in the head capsule. This allows very rapid withdrawal of the stylets from the plant tissue as follows. When the stylets are inserted into the plant tissue, the labium shortens, as described above. When the crumena protractor muscle contracts, it pushes the apical labial segment away from the head, rapidly extending the labium to its full length. The labium lengthening forces the stylets to pull out of the plant tissue and back into the labial groove.

#### Sucking Pump and Precibarial Valve

The food canal at the base of the maxillary stylets opens to a narrow canal, called the precibarium, that is enclosed anteriorly by the epipharynx and posteriorly by the hypopharynx. The precibarium widens proximally into the cibarium which also is enclosed by the epipharynx and hypopharynx. There is a valve, the precibarial valve, between the precibarium and cibarium. The proximal end of the cibarium opens to the anterior-most segment of the alimentary canal, the pharynx. The cibarium functions as the sucking pump. A series of muscles extends from the midline of the inner clypeal wall to the midline of the epipharyngeal wall of the cibarium and to the

epipharyngeal wall of the precibarial valve which is immediately distal to the cibarium. When the muscles are at rest, the epipharyngeal wall presses tightly against the hypopharyngeal wall due to its natural elasticity. This closes the valve. Also, when the muscles are at rest, the epipharyngeal wall of the cibarium rests close to the hypopharyngeal wall (again due to natural elasticity), and the volume of the cibarium is at a minimum. When these muscles contract, the epipharyngeal wall of both the precibarial valve and the cibarium is pulled away from the hypopharyngeal wall. This has two effects. First, it opens the precibarial valve, allowing fluid to flow from the maxillary food canal into the cibarium; and second, it dilates the cibarium, creating the suction needed to pull fluid from the maxillary food canal into the cibarium. When the muscles relax, the epipharyngeal wall of the cibarium returns to its resting position, constricting the volume of the cibarium, and forcing fluid out of the cibarium. At the same time, relaxation of the muscles cause the precibarial valve to close, preventing backflow of fluid from the cibarium into the maxillary food canal. Since the precibarial valve is closed when the cibarium constricts, there is only one direction the fluid can leave the cibarium: into the pharynx.

It should be noted that the primary food source, phloem sap, is under very high turgor pressure. Consequently, suction may not be always necessary to ingest phloem sap. The high turgor pressure of the phloem sap will force sap up through the food canal as soon as the maxillary stylets penetrate a sieve element, in a manner analogous to an artesian well. In fact, if the base of the stylets are severed above the plant surface when the stylet tips are in a phloem sieve element, the turgor pressure of the phloem sap forces sap up through the severed stylets where it can be collected for chemical analysis. Consequently, ingestion rate of phloem sap may be regulated more by the precibarial valve than by the cibarial pump. The pump would play a more important role when the whiteflies occasionally ingest xylem sap (which is under negative pressure) or when the stylets get clogged with one of two sealing mechanisms in phloem sieve elements: coagulation of P-protein or deposition of callose.

### **Salivary Glands and Salivary Pump**

The salivary glands and salivary pump has been described only in the adult stage of whiteflies. Adult whiteflies have two pairs of salivary gland, the primary glands and accessory glands. The ducts from each gland eventually join to form a common salivary duct that enters the salivary pump. The salivary pump functions as a syringe and connects the common salivary duct to the salivary canal enclosed by the maxillary stylets. Muscles attaching to the dorsal wall of the pump pull the dorsal wall away from the ventral wall, creating suction that draws fluid into the pump. When the muscles relax, the dorsal wall snaps back against the ventral wall (due to natural elasticity), expelling fluid from the pump. Opening and closing of valves on the afferent and efferent

ends of the pump in synchrony with contraction and relaxation of the syringe muscles ensure a unidirectional flow of saliva from the salivary glands to the maxillary salivary canal.

### **Saliva**

Whiteflies, as well as all other homopterans that have been studied, produce two kinds of salivary secretions: sheath saliva which gels upon secretion to form the salivary sheath surrounding the stylets in the plant tissue, and watery saliva which contains salivary enzymes and metabolites. At least in aphids, the pH of sheath saliva is slightly acidic while the pH of watery saliva is alkaline with values up to 9.

*Sheath saliva.* As the stylets penetrate into plant tissue, saliva is secreted intermittently from the stylet tips, and this saliva gels soon after secretion, forming the "salivary sheath" or "stylet sheath" which characteristically encases the stylets in a continuous sheath from the initial point of penetration on the plant surface to the tips of the stylets in the plant tissue. The advancing stylets produce the salivary sheath in a sequence of repeating events in synchrony with the incremental advancement of the stylets that was described previously. First, a small volume of sheath saliva, as it is called, is secreted from the stylet tips. It is initially liquid, but gels soon after secretion, encasing the stylet tips in a semi-solid matrix. The stylets then advance a few microns until the tips just break through the recently gelled sheath saliva. Then another small increment of sheath saliva is secreted from the stylet tips, encasing them again in a solid matrix. The stylets then advance a few more microns, breaking through the recently gelled sheath saliva, and the process is repeated over and over as the stylets advance deeper into the plant tissue. Each increment of sheath saliva adheres to the increment secreted previously, so the increments form a continuous sheath encasing the entire stylet length in the plant.

The incremental production of sheath saliva as the stylets advance can be detected by electrical penetration graphs (EPGs) as alternating periods of low electrical resistance, when the sheath saliva is secreted, and high electrical resistance, in between the incremental secretions. Since the rate of increments of salivary sheath production is a function of the rate of stylets advancement, we can detect different rates of stylet advancement through plant tissue by measuring the rate of resistance fluctuations in the EPGs. EPG studies have thus revealed that when silverleaf whiteflies are feeding on non-host plants or are having a first encounter with a novel host, the whitefly spends a much larger proportion of its time with the stylets stationary in the plant tissue without advancing in comparison to when it is feeding on a familiar host plant species.

The production of sheath saliva during stylet penetration appears to be characteristic of the entire order Homoptera and many Hemiptera. Studies on aphids, pentatomids,



and lygaeids indicate that the salivary sheath is mainly protein and contains about 10% phospholipids and probably some conjugated carbohydrate. It is mainly hydrogen-bonded but is stabilized with disulfide linkages. The precursors of the sheath material are secreted by specialized lobes of the salivary gland. If the contents of these lobes are mixed together *in vitro*, the sheath material does not solidify as long as oxygen is excluded. Within the glands, the sheath precursors are maintained in solution by reducing conditions. The precursors also are mixed with amino acids that are presumed to prevent the formation of hydrogen bonds by maintaining a high dielectric constant. Once the precursors are secreted into the plant, the amino acids diffuse out, allowing formation of hydrogen bonds. Reducing components also diffuse out, allowing exposure to oxygen in the plant tissue to initiate disulfide bonding.

While the production of salivary sheaths by homopterans has been known for a very long time, the function of the salivary sheaths is uncertain. Several hypothetical functions proposed in the literature include: (1) the salivary sheath may assist in keeping the stylet bundle (the two maxillary stylets and 2 mandibular stylets) cohesive, preventing them from separating; (2) the salivary sheath may provide a relatively frictionless sheath in which the stylets can easily advance, retract, or rotate; (3) the salivary sheath may seal punctured sieve elements (the primary feeding site - see below) to prevent loss of sap which may trigger a plant defensive response that shuts down the flow of sap to damaged sieve elements; (4) the salivary sheath may provide a relatively inert barrier between the stylets and the plant tissue to minimize the plant wound response to piercing.

Stylet sheaths remain in the plant even after the stylets are withdrawn, and they stain very readily so they can be easily distinguished from the surrounding plant tissue. Consequently, for research purposes, they conveniently provide a detectable record of all places in the plant tissue where the whitefly has inserted its stylets.

*Watery saliva.* In contrast to sheath saliva, watery saliva does not gel upon secretion. Watery saliva contains salivary enzymes and metabolites. Enzymes detected in whitefly salivary glands and/or salivary secretions that presumably are from watery saliva include: amylase, invertase, and alkaline phosphatase. The role of these enzymes in the feeding biology is unknown. At one time, the presence of amylase had been the basis of a hypothesis that whiteflies sometimes feed on mesophyll cells, but more recent evidence indicates that whiteflies do not obtain significant nutrition from mesophyll cells.

### **Stylet Pathway in the Plant**

The target site of stylet penetration for whiteflies is the sap conductive cells of the phloem, the sieve elements. The pathway of the stylets from the plant surface to the sieve element is primarily intercellular, and the stylets weave their way between and around plant cells prior to

puncturing a sieve element. Their great flexibility and ability to twist and turn facilitate their tortuous pathway between and around cells. There is conflicting evidence of whether the pathway of the stylets through the epidermis is directly through the epidermal cells or between adjacent cells; but once past the epidermis, evidence from different laboratories is consistent that the stylet pathway is primarily intercellular through the mesophyll.

The nature of the physical contact between the stylets and the punctured sieve element has not been reported for whiteflies. However, the physical contact between the stylets and the punctured sieve elements has been examined with transmission electron microscopy for aphids, and until proven otherwise, the most parsimonious assumption is that whiteflies behave similarly. In aphids, the mandibular stylets stop at the cell wall of the sieve element, and only the maxillary stylets penetrate a short distance into the sieve element. Sheath saliva also stops at the cell wall and does not extend into the sieve element.

Occasionally, during the primarily intercellular stylet penetration, the stylets penetrate into cells, a phenomenon that can be readily detected in EPG recordings. These intracellular penetrations are usually very brief (< 30 s), and the whitefly then withdraws its stylets from the cell (or pushes them all the way through) back into intercellular space. Aphids also have a primarily intercellular stylet pathway and make brief intracellular punctures along the way from the plant surface to the sieve elements. However, there are some interesting quantitative and qualitative differences between the brief intracellular punctures of aphids and whiteflies. Most aphids that have been studied produce many brief intercellular punctures along the way to the sieve elements, sometimes up to 50 or more between the plant surface and the sieve element. Whiteflies produce far fewer; on average only 3-6 between the plant surface and the sieve element. Most aphids also begin making brief intracellular punctures almost as soon as stylet penetration begins, frequently within the first 30 s of stylet penetration. Whiteflies, on the other hand, do not make intercellular punctures until much later in the probe, generally 8 - 30 min after initiation of stylet penetration. Also, EPG recordings strongly suggest that during the brief intracellular punctures, the behavior of the insect differs between aphids and whiteflies. These differences between whiteflies and aphids likely explain why non-persistent viruses are transmitted primarily by aphids and very few are transmitted by whiteflies (of 211 known non-persistent viruses, 208 are transmitted by aphids and only 3 are transmitted by whiteflies). Non-persistent viruses are transmitted during brief intracellular punctures, especially intracellular punctures produced early in the probe; thus the production of relatively few intracellular punctures by whiteflies, and the virtual absence of intracellular punctures early during probes, are not conducive to transmission of non-persistent viruses by whiteflies. Furthermore, EPGs detect specific behaviors

during brief intracellular punctures by aphids that are associated with inoculation and acquisition of non-persistent viruses, and these behaviors are not detectable in EPGs of whiteflies.

The differences between brief intracellular punctures of aphids versus whiteflies suggests different functions for intracellular punctures in these two families of phloem-feeders. As noted previously, aphids begin making intracellular punctures very early in a probe (usually within the first 30 s) while whiteflies make brief intracellular punctures only late during probes. If the function of the brief intracellular punctures is to sample cell contents to discriminate host from nonhost plants, it would be logical that the intracellular punctures should occur early during probing rather than later after considerable time and energy has been expended penetrating the plant tissue. Thus, I hypothesize that one of the functions of intracellular punctures for aphids but not for whiteflies is host discrimination. Whitefly intracellular punctures generally occur shortly before a phloem sieve element is penetrated and phloem sap ingestion occurs. Thus, I further hypothesize that in whiteflies, brief intracellular punctures represent trial-and-error attempts to locate sieve elements once the stylets are deep within the plant tissue. This may or may not be an additional function of brief intracellular punctures in aphids. These hypotheses need testing.

### **Sensory Organs Involved in Feeding**

Whiteflies have several olfactory receptors on their antennae, but their role in host selection and feeding is unknown. There are only a few experiments reported in the literature to determine if whiteflies are attracted to host plants by their odor. There is no strong evidence for olfactory attraction to host plants. Two papers report some degree of attraction to host plant odor, but the methodology and analysis were so poorly described, that the reader has no way of evaluating their validity. The fact that both male and female whitefly antennae are well equipped with olfactory organs suggest an important role of odor detection in their biology. While there is evidence that greenhouse whitefly females produce a pheromone that attracts males, the presence of well-developed olfactory organs on both sexes, as well as in parthenogenic species like *Parabemisia myricae*, make it tempting to deduce that the olfactory organs also may be used to detect host plant odor. This is a hypothesis that needs future investigation.

As do many homopterans, whiteflies generally rub the tip of their labium on the plant surface prior to inserting their stylets. Whiteflies have seven pairs of sensory organs on the tip of their labium, four of which are mechanoreceptors and three of which are compound mechano/chemoreceptors. In aphids, which have only mechanoreceptors at the tip of their labia, the mechanoreceptors are thought to assist the aphids in locating intercellular grooves on the plant surface to

facilitate stylet penetration between adjacent epidermal cells. Whiteflies also tend to initiate probes in intercellular grooves, so the function of the mechanoreceptors at the tip of the labium of whiteflies may be the same as that postulated for aphids. The role of the chemoreceptors at the tip of the whitefly labium is unknown. Several possible functions are postulated. They may detect relevant chemicals on the plant surface simply by rubbing the tip of the labium over the plant surface or by salivating on the plant surface and pressing the tip of the labium into the saliva where the chemoreceptors could detect plant surface chemical dissolved in the saliva. Alternatively, it is possible that the apical labial chemoreceptors detect chemicals *inside* the plant tissue. While the labium never penetrates the plant surface, it is still at least theoretically possible that the apical chemoreceptors can detect internal fluids in the plant. As discussed previously, whiteflies produce a salivary sheath that extends into the plant as the insect advances its stylets. The salivary sheath actual begins on the plant surface, and during stylet penetration, the tip of the labium with its chemoreceptors is pressed into the salivary sheath on the plant surface. Since the sheath is continuous from the surface to the internal tissues of the plant, it is possible that the apical labial chemoreceptors could detect internal plant chemicals that diffuse up the salivary sheath and come in contact with the chemoreceptors pressed into the surface part of the salivary sheath. Research is needed to test these hypotheses and determine the role of the apical labial chemoreceptors in feeding and host selection.

Despite the known chemoreceptors on the tip of the labium and the well documented behavior of rubbing the tip of the labium on the plant surface prior to inserting the stylets, it is not known if whiteflies taste the surface of the plant and whether or not plant surface chemistry plays a role in host selection by whiteflies. There is some circumstantial evidence that plant surface cues are used by whiteflies: *Parabemisia myricae* discriminates between suitable and unsuitable leaves prior to probing. While plant surface chemistry is an obvious candidate as an explanation for this pre-probing discrimination, other explanations can be postulated. Research is needed to determine what, if any, role plant surface chemistry plays in whitefly host selection and feeding behavior.

The mandibular stylets of whiteflies are hollow through most of their length and each mandibular stylet is traversed by a pair of dendrites. These are believed to be mechanosensory and provide the whitefly with sensory feedback about the movement of the stylets. The maxillary stylets are not associated with any dendrites.

The precibarium, formed by the opposing hypopharynx and the epipharyngeal wall of the labrum provides the pathway of fluid from the maxillary food canal to the cibarial pump. The precibarial valve, as described previously, regulates the flow of fluid through the

precibarium. Close to the precibarial valve are 10 chemoreceptors distal to the valve and 8 chemoreceptors proximal to the valve. These sensillae presumably taste the ingested fluid passing through the precibarium. The role of these sensillae in feeding and host selection are not known, but their location and the lack of any other known chemoreceptors in the stylets or anterior alimentary canal appear to make them the primary taste organs for ingested fluid. Thus, they probably are used to taste plant sap as it is being ingested. At least in some hemipterans, they apparently are also used to taste the plant surface as follows: the insect secretes a small amount of saliva on the plant surface which dissolves surface chemicals, and then ingests the saliva where the precibarial sensillae can detect the dissolved plant surface chemicals. Whether or not this occurs in whiteflies is unknown.

### **Sieve Element Contact and Feeding**

The primary feeding site of whitefly adults and nymphs is the phloem sieve element. However, fewer than 10% of probes by whiteflies succeed in reaching a phloem sieve element. This suggests that locating a sieve element during a probe is not an easy task. How whiteflies and other phloem feeders locate the sieve elements has intrigued researchers for nearly a century, but there is still no conclusive data indicating how they accomplish this. Perhaps it may be simply a trial and error process, randomly piercing cells after the stylets are deep in the plant tissue, as hypothesized previously. The time from the beginning of a successful probe to initial penetration of a sieve element has been examined for several combinations of whitefly species and plant species. The average times for these different combinations range from 16 - 66 min.

Once a sieve element is pierced, whether this is the result of trial and error or result of a more direct method, sieve elements have several physiochemical characteristics that could provide the insect with the information that a sieve element rather than a non-sieve element has been penetrated. These characteristics include very high turgor pressure, very high sugar concentration, and slightly alkaline pH. Which, if any, of these cues is used by whiteflies or other phloem feeders to identify sieve elements is not known for certain. However, the phagostimulatory effect of sucrose in artificial diets fed to whiteflies and other phloem-feeders suggests that the high sugar concentration of sieve elements may be at least one cue used by the insects to identify them.

When an adult whitefly first penetrates a sieve element, it begins salivating into the sieve element. The salivary phase lasts several minutes, and it is during the salivary phase that persistent viruses, such as geminiviruses, are inoculated. After the salivary phase, the whitefly enters an ingestion phase where phloem sap is ingested. The salivary phase may appear again interspersed with periods of ingestion. The function of the salivary phase is unknown. After a variable period of "phloem phase" (the

combined term for salivary + ingestion phase), which for *Bemisia* adults generally lasts several minutes to several hours, the whitefly withdraws its stylets from the sieve element, terminating phloem phase, and may continue probing and eventually initiate another phloem phase, or it may terminate the probe altogether. Whitefly nymphs, on the other hand, tend to stay in the same sieve element for much longer periods of time (measured in days) and have very regular alternating periods of salivation and ingestion. Again, the function of the salivation phase is unknown but it may be involved in conditioning the sieve elements for continued ingestion, and very likely is when the whitefly nymph introduces factors responsible for the systemic physiological disorders inflicted on some host plants by silverleaf whitefly nymphs.

### **Ingestion from Non-Phloem Tissues**

As noted previously, the presence of amylase had been the basis of a hypothesis that whiteflies sometimes feed on mesophyll cells, but this hypothesis has recently been rejected. EPG studies suggest that whiteflies penetrate cells other than sieve elements, but these intracellular penetrations are too brief (usually < 30 s) to be a significant source of ingested nutrients. It is likely that the intracellular punctures serve to sample plant sap with their precibarial sensillae either to aid in host discrimination or to locate sieve elements.

Whiteflies (at least adult whiteflies) occasionally ingest xylem sap, but this is much, much less frequent than phloem sap ingestion. Xylem sap is extremely dilute with very low concentrations of nutrients. This, combined with the infrequency of xylem sap ingestion, indicates that xylem sap is not a significant source of nutrition for whiteflies. Then why ingest xylem sap? Evidence in aphids, which also are primarily phloem feeders that occasionally ingest xylem sap, indicates that xylem sap ingestion is the equivalent of drinking water, and occurs much more frequently if the aphid is dehydrated. Phloem sap, with its very high osmolarity, would not be ideal for addressing a water deficit in the insect.

### **Critical Questions Needing Answers**

In addition to the questions posed in the above text, the following are some important questions. Do whiteflies get gustatory information from the plant surface? Do whiteflies get gustatory information from the apoplast (intercellular space and fluid) during intercellular penetration? What behaviors occur during brief intercellular punctures? How is host acceptance/rejection decided? What is the composition of whitefly saliva? Does the composition of saliva differ between nymphs and adults? Does the composition of saliva differ during different phases of feeding? What are the roles of the different salivary components? How do whiteflies circumvent phloem sealing responses?

## Section F: Plenary Session Summary

Steve Castle  
USDA-ARS, Western Cotton Research Laboratory,  
Phoenix, AZ

### Outbreak Occurrences: Factors that Contribute and Tactics that Suppress

The status of *Bemisia tabaci* as a major pest species of agriculture throughout the world has intensified in recent decades. With its expansion into previously uninfested production systems and increased activity in regions where it has long been established, significant losses have occurred on a global scale irrespective of pest management capabilities. An unprecedented interest in *B. tabaci* biology and management has arisen in recent years in response to the serious challenges posed to agriculture.

Explanations for the upsurge in *B. tabaci* outbreaks have often centered on failures of management. In particular, insecticides have been extensively incriminated for causing large-scale disruptions in control. For example, in the Sudan Gezira during the late 1970s, destructive outbreaks of *B. tabaci* were attributed either to the direct effects of insecticide use, i.e. resistance development and fertility-stimulating effects of hormoligosis, or the indirect effects of insecticide use, i.e. depredations of natural enemy populations. Thereafter, anywhere in the world that a *B. tabaci* outbreak occurred, an insecticide-based explanation would generally be a central element of the post-outbreak head scratching.

While many regions in the world have been impacted by severe infestations of *B. tabaci*, these represent only the latest in a series of *B. tabaci* outbreaks world-wide dating back to the 1920s. Accounts of historical *B. tabaci* infestations provide an essential perspective on present day outbreaks by demonstrating serious pest potential under a range of agricultural and management conditions. The set of observations, historical and modern, allows for a broad consideration of factors that determine the size of *B. tabaci* populations in agricultural systems. When contrasted to agricultural regions with only endemic populations of *B. tabaci*, an even more complete picture of manifold factors becomes possible.

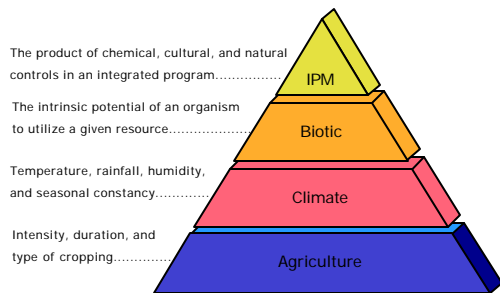
Two approaches have been taken for comparing *B. tabaci* infestations in various systems. One has been the comparison of modern outbreaks with historical outbreaks by examining published accounts. Historical documentation of *B. tabaci* outbreaks prior to 1940 establish their capacity as serious agricultural pests prior to the advent of synthetic organic insecticides. The second approach has been to examine outbreak vs. non-outbreak regions in present times to deduce principle factors contributing to outbreaks. *B. tabaci* occurs in agricultural zones around the world, but does not reach outbreak status in all. It is instructive to examine various

regions where infestations of varying magnitude occur and contrast their physical and cultural features for clues concerning the central factors governing infestation levels. The diversity of California agriculture including cropping variations and climatic differences has served well for deducing important factors contributing to *B. tabaci* populations. In particular, comparisons among Imperial, Coachella, and San Joaquin Valleys have helped to illustrate the role of temperature and cropping system in the buildup, or lack thereof, of *B. tabaci* populations. Although other elements of *B. tabaci* population dynamics are clearly recognized, there has been no synthesis of the fundamental factors that contribute to *B. tabaci* outbreaks. The Outbreak Pyramid represents a conceptual synthesis of 4 general factors that determine the size of *B. tabaci* infestations. The foundation of the Outbreak Pyramid is represented by the Agriculture layer, which forms the resource base subject to exploitation by whiteflies and other pest populations. The quality of the Agriculture layer from a resource standpoint depends on the types of crops grown, the area over which they are grown, or intensity, as well as the length of season, or duration, that they are available for exploitation. The next layer of the pyramid is represented by Climate that stands as the most unalterable component. The Climate layer will dictate to some degree the composition of the Agriculture layer, but only in defining the limits of what can be grown. The particular character of agriculture in any specific region will be more a product of market forces, historical precedent, and physico-environmental limitations such as water availability and soil characteristics. But whatever the composition of the Agriculture layer, climate will exert a major influence on the dynamics of insect populations. Climate factors such as temperature, rainfall, and humidity impinge on many facets of the ecology of *B. tabaci* including growth rates, survival or demise on a host plant during intense weather, and dispersal between crops. In particular, temperature is a driving force in generation time and how fast *B. tabaci* populations grow, whereas low rainfall favors greater survivorship. The Biotic layer represents the intrinsic capacity of *B. tabaci* to exploit the resources available as determined by the agricultural makeup and within the constraints of the physical environment as defined by the climate. At the pinnacle of the pyramid is the management or IPM layer, representing the product of chemical, cultural and natural controls, and is the layer subject to the most manipulation by humans.

The essence of the Outbreak Pyramid is that in circumstances where the basal 2 layers of the pyramid are "broad" with agro-ecological potential, the biotic characteristics of *B. tabaci* permit exploitation of the resource base in excess of the various types of control, both natural and manipulated. Conceptually, the volume of the IPM layer at the top of the pyramid is dwarfed in comparison to the combined volume of the three layers that form the pyramid base. In practice, management efforts that are targeted at individual fields, even if they

provide superior results, have been overwhelmed by region-wide population increases of *B. tabaci* on crop and non-crop vegetation. It is not a failure of management per se to control regional populations of *B. tabaci*, but rather a function of the combined potential contained within the climate, agriculture and biotic levels of the pyramid that eventually exceed the capacity of local crop protection.

The Outbreak Pyramid



Regions with year-round crop production such as the desert southwest USA have experienced incremental buildups of *B. tabaci* populations on a succession of crops. Management of whitefly infestations within crops grown early in the annual cycle has often proved adequate through harvest time. But with each crop transition, and with an increasingly favorable environment as summer temperatures arrive, whiteflies break away from local control and eventually saturate an entire region. In some instances, as crops near harvest, local field controls are relaxed because of the access required for hand picking. After the harvest, there is often considerable delay in plowing the field that serves only to generate large numbers of whiteflies ready to disperse to neighboring crops. These are examples of vulnerability in crop production that whiteflies rapidly exploit. The tools of management cannot always be used fight back whitefly onslaughts because of regulatory issues, e.g. pre-harvest intervals, as well as economic issues such as the cost of insecticide applications on low value crops. The broad polyphagy of *B. tabaci* allows for field, vegetable, and tree crops to all serve as hosts. Moreover, some regions harbor a large biomass of non-crop hosts in the form of both wild and ornamental hosts, and these often contribute significantly to whitefly numbers.

In regions where *B. tabaci* outbreaks have occurred chronically, modifications targeted at 1 or more levels of the Outbreak Pyramid have been pivotal in tempering further outbreaks. Changes can only be made at the Agriculture level or at the IPM level as the macro climate and the intrinsic biotic characteristics of the organism are unmanipulable. The IPM level is the most flexible level in terms of modifying control practices or instituting new information-gathering techniques for improving the decision-making process. It is the level that must be expanded in volume in order to be able to suppress the

explosive potential represented by the expansion of the lower 3 levels. Expanded IPM happens through the development of sampling programs, determination of action thresholds, conservation of natural enemies, and the integration of chemical, cultural and natural controls that becomes possible through research and information gathering. When true integration of control tactics occurs as opposed to haphazard and uncoordinated implementation, the result can be spectacularly successful as it has been in Arizona from 1996 to the present. Expanded IPM in Arizona and California has also occurred with the arrival of superior insecticides that target multiple stages of *B. tabaci*, are residually active, and have relatively low impact on natural enemy populations. Modification of the Agriculture layer has played an important role in curbing *B. tabaci* outbreaks in California's Imperial Valley. Relatively few acres of fall melons are grown now compared to pre-1991 levels, and spring melon and fall vegetable acreages have also dropped. The Agriculture layer of the Outbreak Pyramid has contracted in the Imperial Valley, and in combination with the expansion of the IPM layer, *B. tabaci* outbreaks have been brought under control.

### III. Reports of Research Progress – 2001 & 2002

#### Reports of Research Progress

##### Section A: Biology, Ecology, and Population Dynamics

Co-Chairs: Jackie Blackmer and David Byrne

**Investigator's Name(s):** Ma. del Rosario Avila, P. Cano, U. Nava and E. López

**Affiliation & Location:** Comité Regional de Sanidad Vegetal. Calz. Cuauhtemoc 1540 Sur, Torreón, Coahuila

**Research & Implementation Area:** Section A: Biology, Ecology, and Population Dynamics

**Dates Covered by Report:** 2000

##### Identification of the Whitefly Species and their Plant Hosts in the Comarca Lagunera, Mexico, 2000

In the Comarca Lagunera area from 1994 to 1996, three species of whiteflies were identified, which were: sweet potato whitefly (SPWF), *Bemisia tabaci* Gennadius, the silverleaf whitefly (SLWF), *Bemisia argentifolii* Bellows & Perring and the greenhouse whitefly, *Trialeurodes vaporariorum* Westwood. During 1997 four additional whitefly species were identified: the banded wing whitefly, *Trialeurodes abutilonea* Haldeman, the woolly whitefly, *Aleurothrixus floccosus* Maskell, the acacia whitefly, *Tetraleurodes acaciae* Quaintance and the citrus blackfly, *Aleurocanthus woglumi* Ashby.

At the beginning of the sampling on the cultivated fields during 1994 the whitefly species identified were: the SPWF *B. tabaci* and the SLWF *B. argentifolii*. However, the more recent (1996-97) whitefly samples indicated that dominant species in the cultivated fields was *B. argentifolii* and that the *B. tabaci* species has been practically displaced.

Additionally, 108 plant species were identified as SLWF, *B. argentifolii* plant hosts. The main crop hosts were: *Brassica oleracea* L. vars. *botrytis* and *capitata*, *Cucumis melo* L., *Cucumis sativus* L., *Cucurbita pepo* L., *Citrullus lanatus* (Thunb.) Mansf. and *Gossypium hirsutum* L. It was found that ten weed had high SLWF infestation, but, because of their higher distribution in the crop fields the most important ones were: *Convolvulus arvensis* L., *Flaveria trinervia* (Spreng.) Mohr, *Ipomoea* spp, *Solanum elaeagnifolium* Cav. and *Xanthium strumarium* L. In the urban areas 38.7% of the plant hosts had from medium to high SLWF infestation. The plant species with the highest SLWF infestation were: *Acacia berlandieri* Benth., *Acalypha wilkesiana* Muell., *Althaea rosea* Cav., *Bahuinia divaricata* L., *Camellia japonica* L., *Sedum* spp, *Ipomoea* spp, *Jasminum* spp, *Lantana camara* L. and *Oenothera* spp.

**Investigator's Name(s):** Jacquelyn L. Blackmer<sup>1</sup> and Dale Cross<sup>2</sup>

**Affiliation & Location:** <sup>1</sup>USDA-ARS, Western Cotton Research Laboratory, 4135 E. Broadway Rd., Phoenix, AZ 85040, USA; <sup>2</sup> Department of Entomology, The University of Arizona, Tucson, AZ, 85721

**Research & Implementation Area:** Section A: Biology, Ecology, and Population Dynamics

**Dates Covered by the Report:** 2001

**Response of *Eretmocerus eremicus* (Hymenoptera: Aphelinidae) to Skylight and Plant Cues in a Vertical Flight Chamber**

In the southwestern United States, *Eretmocerus eremicus* (Hymenoptera: Aphelinidae) is a native parasitoid of the *Bemisia* complex (Homoptera: Aleyrodidae). However, little information currently exists on its potential as a biological control agent of whiteflies or on the factors that influence its tendency to disperse. Information on this parasitoid's dispersal propensity might alter the release strategy employed. Low dispersal rates could mean that multiple release sites might be necessary for optimal distributions relative to its host. Our study examined the host-habitat/host location phase(s) of host-finding behavior for *E. eremicus*. In this study, we examined the flight behavior of male and female *E. eremicus* in response to skylight (here simulated by a mercury-vapor lamp) and plant cues (a 550-nm filter) in a vertical flight chamber. The visual plant cue was presented to the parasitoid in the absence of whitefly stimuli. Approximately 90% of the parasitoids took off in response to the skylight cue. Both sexes were capable of sustained flights in excess of 60 min; however, average flight durations were  $15.6 \pm 3.8$  min for males and  $7.8 \pm 2.6$  min for females. When a plant cue was presented during the parasitoid's phototactic flight, four relatively distinct responses were observed. Fifty-one percent of the individuals responded to the plant cue throughout their flight by flying toward or by landing on the cue. The majority of these parasitoids were females. Approximately 12% of the wasps exhibited an intermittent, positive response to the plant cue. Twenty percent exhibited a 'migratory' response. These parasitoids, which were predominantly males, failed to respond to the plant cue until they had flown for a considerable period. Finally, 17% failed to respond to the target during their flight. Approximately 37% of the individuals that showed a positive response to the plant cue actually landed on it and the majority of these were female. However, in the absence of additional whitefly stimuli, tenure time on the visual plant cue was relatively short. The differential response to the plant cue by male and female parasitoids could be, in part, because females are driven to locate hosts in which to oviposit, and males are driven to find mates. Additional studies need to determine how biotic and abiotic factors influence the dispersal tendencies of *E. eremicus*, the role of visual and olfactory information in the host-finding process, and whether the vertical flight chamber might be a useful tool for screening the flight propensity of potential biological control agents.

**Investigator's Name(s):** James S. Buckner and Marcia M. Hagen

**Affiliation & Location:** USDA-ARS, Insect Genetics and Biochemistry Research Unit, Red River Valley Agricultural Research Center, Fargo, ND

**Research & Implementation Area:** Section A: Biology, Ecology, and Population Dynamics

**Dates Covered by the Report:** 2001

### **The Triacylglycerol Composition of Silverleaf Whitefly Adults**

*Bemisia argentifolii* Bellows and Perring (Homoptera: Aleyrodidae) adults were collected from a whitefly colony maintained on hibiscus plants, weighed and stored at -80°C. Groups of adults (30-50 mg) were placed into a 20-ml glass vial containing 8-10 ml of CHCl<sub>3</sub>/methanol (2:1) and covered with a Teflon-lined cap. To homogenize (sonicate) the tissues, the vial was placed in the water bath of a Fisher Scientific, Model FS 60, ultrasonic cleaner for 15-30 min. The homogenate was filtered through glass wool with CHCl<sub>3</sub> and the CHCl<sub>3</sub> washed with water. The lipid extract in CHCl<sub>3</sub> was spotted on 250-µm silica gel TLC plates and the triacylglycerol (TAG) fraction was separated from other lipid fractions by developing the TLC plates in hexane/ethyl ether/formic acid (80:20:1). The purified TAG fraction was visualized with iodine vapors, scraped from the TLC plates and eluted from the silica gel with CHCl<sub>3</sub>.

High performance liquid chromatography (HPLC) was used to separate the TLC-purified mixture of TAG using a ChromSpher 5 Lipid column (Chrompack, 5 µm, 4.6 X 250 mm). Mass detection of TAG constituents was performed using a Sedex (Model 55) evaporative light scattering detector (ELSD). Peak areas from the ELSD were used to determine quantities of resolved TAG components and structural identifications were made by HPLC-MS. Individual TAG components were identified by comparing their column retention times to those of authentic TAG standards and by comparative mass spectral analysis using the Waters Model 2690 Separations Module linked to a Waters Thermabeam Mass Detector.

For the mixture of triacylglycerols from *B. argentifolii* adults, the major fatty acid constituents were identified as oleic acid (18:1), palmitic acid (16:0), stearic acid (18:0) and linoleic acid (18:2), in descending order of abundance. Lesser amounts of palmitoleic acid (16:1) and arachidonic acid (20:0) were detected, as well as trace amounts of myristic acid (14:0). The major intact TAG in decreasing order of abundance, as analyzed by HPLC-MS, were 1-palmitoyl-2,3-dioleoylglycerol (POO), 1,2-dipalmitoyl-3-oleoylglycerol (PPO), 1-palmitoyl-2-stearoyl-3-oleoylglycerol (PSO), 1,2,3-trioleoylglycerol (OOO), 1-stearoyl-2,3-dioleoylglycerol (SOO), 1-palmitoyl-2-oleoyl-3-linoleoylglycerol (POL) and 1-palmitoyl-2-linoleoyl-3-stearoylglycerol (PLS). The fatty acids of the TAG from *B. argentifolii* adults were also analyzed as their methyl ester derivatives following hydrolysis with 5% KOH in methanol and subsequent reaction of the free fatty acids with 10% HCl in methanol to form methyl esters. The distribution of fatty acids was similar to that indicated above for intact TAG: approximately 58% as 18:1, 29% as 16:0, and lesser amounts of 18:0, 18:2, 18:3 and 16:1. These methods developed for characterization of TAG from adults will be used to determine the TAG composition of *B. argentifolii* nymphs and to provide useful information in regard to the lipid nutrient reward for whitefly predators and parasitoids.



**Investigator's Name(s):** James S. Buckner and Marcia M. Hagen

**Affiliation & Location:** USDA-ARS, Insect Genetics and Biochemistry Research Unit, Red River Valley Agricultural Research Center, Fargo, ND

**Research & Implementation Area:** Section A: Biology, Ecology, and Population Dynamics

**Dates Covered by the Report:** 2001

### **The Fatty Acid Composition of the Internal Lipids from Silverleaf Whitefly Nymphs and Adults**

The cuticular lipids of *Bemisia argentifolii* Bellows and Perring (Homoptera: Aleyrodidae) nymphs (1) and adults (2, 3) have been characterized, but the identification and quantification of the internal lipids of whiteflies have not been determined. The assessment of the internal lipids as a source of nutrient reward is especially relevant to specific parasitoids of whiteflies since they consume their host from within. The identification and composition of the fatty acids associated with the major lipid classes (triacylglycerols, free fatty acids and phospholipids) within *B. argentifolii* nymphs were determined. Comparisons were made to the fatty acids present in the internal lipids of adult *B. argentifolii*.

Feeding silverleaf whitefly nymphs were raised on young cantaloupe plants and adult whiteflies were maintained on hibiscus. Groups of 200-300 third and fourth instar nymphs were carefully removed from leaves, placed in glass vials, and their surface lipids were removed and discarded using hexane. The surface lipids from groups of pre-chilled adults were also removed with hexane. The internal lipids were then extracted by sonication for 45 min in 7-8 ml of chloroform/MeOH (2:1). The chloroform/MeOH homogenate was filtered through glass wool and partitioned twice between chloroform and water. The internal lipids of whiteflies were separated into neutral lipids, free fatty acids and phospholipids using a column of 80–180 mg of Porasil Prep silica (Waters Corp., Milford, MA). The separation was achieved using a modification of a novel solvent system containing tertiarybutylmethylether (*t*-BME) (4). Lipid samples were applied to the column in hexane and lipid fractions eluted using the following solvents: hydrocarbons, wax esters, aldehydes (99.5:0.5 hexane/*t*-BME); triacylglycerols (TAG), alcohols (96:4 hexane/*t*-BME); free fatty acids (FFA) (100:2 hexane/acetic acid); phosphatidylethanolamine (PE) (20:4:1 *t*-BME/MeOH/0.001M ammonium acetate, pH 8.6) and phosphatidylcholine (PC) (5:4:1 *t*-BME/MeOH/0.001M ammonium acetate, pH 8.6). The purity of lipid fractions was verified by silica HPTLC. Fatty acid methyl esters (FAME) were formed by alkaline hydrolysis of the triacylglycerol and phospholipid fractions followed by treatment of the free fatty acids with methanolic HCl. The FAME derivatives of fatty acids were separated by capillary gas chromatography (CGC) and analyzed using splitless injection at 200°C, a Supelco SP-2380 capillary column (30m x 0.25mm id.) in a Hewlett-Packard Model 5890 GC. The quantities of FAME were determined by converting integrated peak area data from the FID response using nonlinear regression slope data for increasing amounts (0.78-200 ng) of the authentic methyl esters of 14:0, 16:0, 16:1, 18:0, 18:1, 18:2, 18:3 and 20:0 fatty acids.

All lipid classes contained variable distributions of 8 fatty acids: the saturated fatty acids, myristic acid (14:0), palmitic acid (16:0), stearic acid (18:0), arachidonic acid (20:0); the monounsaturated fatty acids, palmitoleic acid (16:1), oleic acid (18:1); the polyunsaturated fatty acids, linoleic acid (18:2), linolenic acid (18:3). Fourth instar nymphs had 5-10 times the quantities of fatty acids as compared to third instar nymphs and 1-3 times the quantities from adults. The quantity differences of fatty acids between fourth and third instar nymphs related to their size and weight differences: fresh weights of 19-28 mg/nymph and 5-9 mg/nymph, respectively. For nymphs and adults, TAG lipids were the major source of fatty acids, with oleic (18:1) and palmitic (16:0) acids as major components. For nymphs and adults, the majority of the polyunsaturated fatty acids, linoleic (18:2) and linolenic (18:3) were present in the phospholipid fractions (PE + PC).

#### References

1. Buckner et al. 1999. *Comp. Biochem. Physiol.* 124B: 201-207.
2. Buckner et al. 1994. *Insect Biochem. Mol. Biol.* 24: 977-87.
3. Nelson et al. 1994. *Comp. Biochem. Biophys.* 109B: 293-303.
4. Hamilton and Comai. 1988. *Lipids* 23: 1146-1149.

**Investigator's Names:** L. Canas<sup>1</sup>, S. E. Naranjo<sup>2</sup> & P. C. Ellsworth<sup>1</sup>

**Affiliation & Location:** <sup>1</sup>Department of Entomology, University of Arizona, Maricopa Agricultural Center, 37860 W. Smith-Enke Road, Maricopa, AZ 85239 and <sup>2</sup>USDA-ARS, Western Cotton Research Laboratory, Phoenix, AZ 85040

**Research & Implementation Area:** Section A: Biology, Ecology and Population Dynamics

**Dates Covered by the Report:** November 2000-December 2001

**Seasonal Ecology of *Bemisia tabaci* in Arizona: Low Temperature and Host Plant Effects on Field Populations and Associated Mortality Factors**

The current ongoing study has examined seasonality and mortality patterns of *B. tabaci* on different hosts during the year. Plots of six representative host plants (broccoli, cantaloupe, cotton, alfalfa, Lantana and various weeds) were established at the Yuma, Maricopa and Marana Agricultural Centers in Arizona. These sites represent the range of geographic and climatic areas of the state for cotton production. Here we report findings from the first year of the study. Two host plants, cantaloupe and Lantana, were observed to provide a good environment for the buildup of *B. tabaci* populations during the fall. *B. tabaci* populations declined steadily with decreasing temperatures during the winter. Low temperatures affected the host plants, with Lantana plants losing all foliage due to freezing temperatures. However, a significant number of Lantana plants produced new foliage during the spring and summer. After the decline during the winter, populations survived in broccoli and *Malva parviflora* (cheeseweed), and built up in broccoli, cantaloupe and Lantana. Populations of *B. tabaci* were extremely high in Lantana and cantaloupe, which allowed for significant densities in cotton once this crop was established. In addition, during the middle of the summer, ground cherry plants hosted large densities of *B. tabaci*. The prevalence of different weed species through the year also allowed for increased populations. From the three different regions, Yuma showed earlier increases in *B. tabaci* populations, followed by Maricopa and Marana. Yuma populations were in general smaller than those of Maricopa and Marana. *B. tabaci* populations in alfalfa were low at all sites. Life table analyses demonstrated differences in survivorship patterns on the different hosts. In general, the highest survivorship was observed in cantaloupe at all sites. The highest *B. tabaci* survival at Marana occurred in cantaloupe (about 47%) followed by Lantana (15%), weeds (15%) cotton (4%), alfalfa (3%) and broccoli (2%). Yuma had the highest survival in cantaloupe (45%) followed by broccoli (31%), Lantana (19%), weeds (17%) and cotton (7%). Maricopa had the highest survival in cantaloupe (45%) followed by broccoli (45%), Lantana (33%), alfalfa (28%), weeds (15%) and cotton (4%). From the different mortality factors observed, both predation and dislodgment accounted for a significant portion of the mortality at all locations. Dislodgment and desiccation were important in some host plants and probably were influenced by low temperatures. For instance, broccoli cohorts suffered high mortality due to desiccation and dislodgment. Rates of predation were relatively high in all host plants but more so in cotton across the different locations. Parasitism by *Encarsia* and *Eretmocerus* varied from host to host and from location to location. Parasitism was highest in Lantana, especially at Maricopa (32%). Despite high levels of *B. tabaci* mortality at the three locations, it was insufficient to prevent population outbreaks in most plant hosts. The examination of the biodemographic characteristics of *B. tabaci* during the year, especially during the winter months will provide important information relative to the cold hardiness and survival of this species. Such information will aid the prediction, and possibly the prevention, of outbreak populations in the late spring and summer

**Investigator's Name :** S. J. Castle

**Affiliation & Location:** USDA, ARS, Western Cotton Research Lab, Phoenix, AZ

**Research & Implementation Area:** Section A: Biology, Ecology, and Population Dynamics

**Dates Covered by the Report:** 2000-2001

### **Differential Host Acceptance of Cotton and Melon by *Bemisia tabaci***

A series of greenhouse experiments were conducted to evaluate host acceptance of melons relative to cotton by *Bemisia tabaci*. The objectives were to quantify the differences between melons and cotton in terms of the number of adult whiteflies that settled on either plant, then determine the levels of oviposition that occurred over the course of the experiment. Two types of choice assays were used to examine adult settlement and oviposition. For the first, acetate cylinders 0.6 m high by 0.28 m diameter were fashioned with 3 equidistant slots cut into the cylinder wall at a height of 0.46 m, large enough for fully expanded melon and cotton leaves to be inserted into the interior space of each cylinder. The tops of the cylinders were covered with nylon organdy to retain whiteflies within the cylinder space. The leaves projecting into the cylinder interiors remained attached to their plants. A foam-rubber collar was sandwiched around each leaf petiole and then snugly fit into the rectangular slots in the cylinder walls so that no whiteflies would be able to escape from the interiors. Once all cylinders were fitted with their complement of leaves and all openings sealed, they were ready to be infested with adult whiteflies. Adult whiteflies were aspirated from colonies maintained on both melon and cotton plants. Approximately 120 adults were collected into an aspiration tube, then transferred through a sleeve opening to be released inside each cylinder cage. As many as 18 cylinders were set up at one time and infested with adult whiteflies. A series of 5 counts were made beginning at 18:00 on the day of released, then followed by 2 days of counts in the morning and at dusk. Each leaf projecting into the cylinder spaces was closely viewed from outside the cylinder walls. A total of 10 cylinder cage experiments were conducted.

For the 2<sup>nd</sup> type of assays, experiments were conducted in 2 small greenhouses with 16 plants of each type, melon and cotton, arranged into a randomized complete block design with 4 blocks consisting of 8 plants each. The 4 blocks were arranged on a bench on one side of each greenhouse. Whiteflies were collected separately from cotton and melon cultures. Approximately 1200 adults were released into each of the 2 greenhouses. Leaf counts of adults were made over the next 2.5 days, morning and late afternoon. Following the 5<sup>th</sup> and final count, all leaves were collected from each test plant for egg counts. Whitefly adult and egg counts were expressed as the mean number ( $\pm$ SEM) of whiteflies per leaf at each leaf node.

In both the cylinder cage and greenhouse experiments, *Bemisia tabaci* consistently demonstrated a much higher affinity for melons compared to cotton. Adult settling and oviposition rates were greater on melon in the greenhouse experiments that used whole plants and for which whiteflies were free to move from plant to plant. A similar pattern was observed in the cylinder cage tests, but the differential in adult settling and oviposition between melons and cotton, although highly significant, was not as great as the greenhouse test. Overall, a ratio of 2:1 or greater was observed for mean numbers of adults on the melon leaf relative to either one of the two cotton leaves. In some cases, i.e. experiment 1, the differential was approximately 5:1 in favor of melon leaves. The relative number of whitefly eggs on either host tended to follow the same pattern observed for adults.

The differential between cotton and melon plants was more pronounced in the 2 greenhouse experiments in terms of mean whitefly adult and egg densities. The mean numbers of whitefly adults on cotton leaves compared to melon leaves ranged between 8-31-fold greater on melon leaves. Egg densities on melon leaves exceeded those on cotton leaves between 8-56-fold. There was little difference in results with respect to the origin of the whiteflies used in the greenhouse experiments. However, there was a marked tendency for more eggs overall to be deposited on leaves if the whiteflies originated from the melon colony.

**Investigator's Name(s):** C. C. Chu, P. Alexander, C. G. Jackson, T. J. Henneberry

**Affiliation & Locations:** USDA-ARS, Western Cotton Research Laboratory, Phoenix, AZ

**Research & Implementation Area:** Section A: Biology, Ecology, and Population Dynamics

**Dates Covered by the Report:** 1998-2000

#### **Modification of CC whitefly traps**

Modifications of CC whitefly traps are in progress to improve their potential for adult whitefly control in greenhouses. Adult catches in the modified CC traps have been increased by 50% by coating trap tops with Tanglefoot and removing the deflector plates. In laboratory studies, installation of a lime green LED light in the trap top resulted in catches of 281 adults/trap/24-h compared with catches of 18 and 12 adults/trap/24-h for traps with white LED light and no light, respectively. Studies are in progress to test the effects of the modified CC traps on catches of whitefly parasitic wasps.

**Investigator's Name(s):** <sup>1</sup>C. C. Chu, <sup>2</sup>J. S. Buckner, <sup>2</sup>D. R. Nelson, <sup>3</sup>T. P. Freeman, <sup>1</sup>T. J. Henneberry

**Affiliation & Locations:** <sup>1</sup>USDA, ARS, Western Cotton Research Laboratory, Phoenix, AZ; <sup>2</sup>USDA, ARS, Biosciences Research Laboratory, Fargo, ND, <sup>3</sup>Electronic Microscopy Center, North Dakota State University, Fargo, ND

**Research & Implementation Area:** Section A: Biology, Ecology, and Population Dynamics

**Dates Covered by the Report:** 2000

### **Preliminary Study of the Morphological Development of Silverleaf Whitefly Stages in the Field**

Leaf samples were taken from field grown cotton and cantaloupe in Phoenix, AZ in 2000. The morphological development of silverleaf whitefly stages were determined at Fargo, ND. Nymphs on cotton leaves were longer and wider compared with nymphs on cantaloupe leaves. From cotton, the length of the four nymph stages were 257, 355, 589 and 724  $\mu\text{m}$ , respectively, and widths 140, 217, 395 and 498  $\mu\text{m}$ , respectively. For cantaloupe nymphs, the lengths were 244, 350, 456, and 601  $\mu\text{m}$  and widths were 129, 192, 267 and 411  $\mu\text{m}$ , respectively.

With nymphs placed on their sides, dorso-ventral measurements for nymphs on cantaloupe leaves were greater except for the third instars compared with dorso-ventral measurements for nymphs on cotton leaves. The values for nymphs from cotton and cantaloupe leaves were 63 vs. 66  $\mu\text{m}$ , 78 vs. 98  $\mu\text{m}$ , 139 vs. 124  $\mu\text{m}$  and 214 vs. 259  $\mu\text{m}$  for the first, second, third and fourth instars, respectively. As nymphs developed from first to fourth instars, the ventral half of the body increased more compared with the dorsal half of the body. The ratios of ventral and dorsal halves of bodies were 0.53, 0.57, 0.63 and 0.70 for first, second, third and fourth instars, respectively. On the average, adult body length from body lengths from head to tail were 953 and 1127  $\mu\text{m}$  and body weights were 17 and 39  $\mu\text{g}$  for males and females, respectively. Egg weight was 0.8  $\mu\text{g}$  per egg. Exuvia weighed 0.8 and 5.4  $\mu\text{g}$  for non-parasitized and parasitized from fourth instar nymph's exuvia, respectively. The significances of the differences for each stage of development on the two plant species are being analyzed.

**Investigator's Name(s):** C. C. Chu<sup>1</sup>, J. S. Buckner<sup>2</sup>, D. R. Nelson<sup>2</sup>, T. P. Freeman<sup>3</sup>, and T. J. Henneberry<sup>1</sup>

**Affiliation & Locations:** <sup>1</sup>USDA, ARS, Western Cotton Research Laboratory, Phoenix, AZ; <sup>2</sup>USDA, ARS, Biosciences Research Laboratory, Fargo, ND, <sup>3</sup>Electronic Microscopy Center, and North Dakota State University, Fargo, ND

**Research & Implementation Area:** Section A: Biology, Ecology, and Population Dynamics

**Dates Covered by the Report:** 2001-2002

### **Preliminary Study of the Morphological Development of Sweetpotato Whitefly Stages in the Fields**

Leaf samples were taken from field grown cotton and cantaloupe in Phoenix, AZ in 2000 and 2001. Measurements from eggs to nymphs and exuvia for the morphological development of whitefly stages were made at Fargo, ND. The 4<sup>th</sup> instars on cotton leaves were longer and wider compared with that on cantaloupe leaves. From cotton, the length of the four nymph stages were 250, 355, 434 and 724  $\mu\text{m}$ , respectively, and widths 135, 214, 270 and 498  $\mu\text{m}$ , respectively. For cantaloupe nymphs, the lengths were 244, 350, 456, and 603  $\mu\text{m}$  and widths were 129, 192, 267 and 411  $\mu\text{m}$ , respectively. With nymphs placed on their sides, dorso-ventral measurements for nymphs on cantaloupe leaves were greater compared with nymphs on cotton leaves. The values for nymphs from cotton and cantaloupe leaves were 57 vs. 66  $\mu\text{m}$ , 76 vs. 98  $\mu\text{m}$ , 71 vs. 123  $\mu\text{m}$  and 214 vs. 258  $\mu\text{m}$  for the first, second, third and fourth instars, respectively. As nymphs developed from first to fourth instars, the ventral half of the body increased more compared with the dorsal half of the body. The ratios of the ventral and dorsal halves of the bodies were 0.53, 0.57, 0.61 and 0.70 for first, second, third and fourth instars, respectively. On average, adult body length from head to tail were 807 and 884  $\mu\text{m}$  and body weights were 17 and 39  $\mu\text{g}$  for males and females, respectively. Eggs weighed 0.8  $\mu\text{g}$  per egg. Exuvia weighed 1.2 and 3.6  $\mu\text{g}$  for non-parasitized and parasitized exuvia, respectively. The significances of the differences for each stage of development on the two plant species are being analyzed.

**Investigator's Name(s):** T. P. Freeman<sup>1</sup>, J. S. Buckner<sup>2</sup>, D. R. Nelson<sup>2</sup>, C. C. Chu<sup>3</sup>, and T. J. Henneberry<sup>3</sup>

**Affiliation & Location:** <sup>1</sup>Electron Microscopy Center, Plant Pathology Department, North Dakota State University, Fargo, ND; <sup>2</sup>USDA-ARS Bioscience Research Laboratory, Fargo, ND; <sup>3</sup>USDA-ARS Western Cotton Research Laboratory, Phoenix, AZ

**Research & Implementation Area:** Section A: Biology, Ecology and Population Dynamics

**Dates Covered by the Report:** 2000

### **The Mechanics of Stylet Penetration by *Bemisia argentifolii*.**

Most adult silverleaf whitefly probe sites occurred along the margins of the abaxial epidermal cells. Even though the penetration sites were in the area of the cell margins, we found that the stylets passed directly through the cytoplasm of the epidermal cell and not through the common radial wall between epidermal cells. Adult stylets averaged 217  $\mu\text{m}$  long. During non-feeding periods the stylet is completely enclosed within the labium, which consists of four distinct segments. The stylet bundle enters between the first and second labial segment and extends to the tip of the labium. After the tip of the labium is anchored to the epidermal surface the adult whitefly lowers its head pushing the stylet bundle down the labial groove into the host plant. The entire length of the stylet can be inserted into the leaf in this manner. The depth of penetration can be determined by examining the position of the labrum along the labial groove.

**Investigator's Names:** T. P. Freeman<sup>1</sup>, J. S. Buckner<sup>2</sup>, D. R. Nelson<sup>2</sup>

**Affiliation & Location:** <sup>1</sup>Electron Microscopy Center, Plant Pathology Department, North Dakota State University, Fargo, ND; <sup>2</sup>USDA-ARS Bioscience Research Laboratory, Fargo, ND

**Research & Implementation Area:** Section A: Biology, Ecology and Population Dynamics

**Dates Covered by the Report:** 2000-2001

### **What I Know About Silverleaf Whitefly Nymph Feeding and What I Would Like to Know**

Nymphs feed during all four stages of development. Once they begin to feed, they settle down and become flattened against the leaf surface. The rostrum of the nymph is short and has sensillia on the tip similar to those found on the labium of the adult. The rostrum also has a groove through which the stylet bundle passes similar to the labial groove of adults. The tip of the mandibular stylets are ridged or serrated and used to penetrate the host leaf tissue. Stylets range from 110  $\mu\text{m}$  in first instar to over 200  $\mu\text{m}$  in fourth instar. The stylets can be at least partially withdrawn and reinserted during the feeding process. The nymphs increase in length and width with each molt and the newly formed stylets are also longer with each molt. However, the stylets do not increase in length proportional to the increase in size of the nymphs.

What are the mechanisms involved in the insertion and withdrawal of the nymphal stylet? Do the stylets of subsequent molts use the salivary sheaths developed by an earlier instar? What is the function of the sensillia on the tip of the rostrum in sedentary nymphs? How does the nymph stylet find the phloem tissue in host leaves? Does a salivary sheath form within the sieve tube element and do the stylets move from one sieve tube element to another?



**Investigator's Names:** Thomas P. Freeman<sup>1</sup>, James S. Buckner<sup>2</sup>, Dennis R. Nelson<sup>2</sup>

**Affiliation & Location:** <sup>1</sup>Electron Microscopy Center, Plant Pathology Department, North Dakota State University, Fargo, ND; <sup>2</sup>USDA-ARS Bioscience Research Laboratory, Fargo, ND

**Research & Implementation Area:** Section A: Biology, Ecology and Population Dynamics

**Dates Covered by the Report:** 2001

### **Silverleaf Whitefly Nymph Stylets and Feeding Characteristics**

After emerging from the egg, silverleaf whitefly (*Bemisia argentifolii* Bellows & Perring) nymphs move across the surface of the leaf in search of a feeding site. The crawler is the only nymphal stage that is capable of locomotion. The crawler must begin feeding quickly if it is to survive. Prior to the onset of feeding the crawler's stylet is folded within the body of the nymph. When the crawler begins to feed it becomes flattened against the leaf surface. Like the adult whitefly, the nymph produces a flange like material that appears to cement the tip of the rostrum to the surface of the host leaf. Staining techniques can be used to determine both probe and feeding sites. The nymph stylet bundle consists of a pair of mandibular stylets serrated at the tip and a pair of maxillary stylets locked together to form the food canal. The ultrastructure of the nymph and adult stylets are similar. The nymph stylet is extended through a groove in the rostrum. After penetrating the epidermal layer of the leaf the stylet moves through the apoplast of mesophyll all the way to the phloem tissue. The stylet path can be studied by examining the salivary sheath which remains in the leaf even after the stylet has been removed. The salivary sheaths are often highly branched, demonstrating that the stylet can be at least partially withdrawn and then extended in a different direction. Prior to each molt, the stylets are withdrawn into the body of the nymph and left behind in the cast exuviae. With each successive molt the stylet becomes longer and in the fourth instar may reach lengths greater than 200  $\mu\text{m}$ .

**Investigator's Name(s):** C. Joel Funk<sup>1</sup>, Elizabeth W. Davidson<sup>2</sup>, and Wayne Hunter<sup>3</sup>

**Affiliation & Location:** <sup>1</sup>USDA, ARS, Western Cotton Research Laboratory, Phoenix, AZ 85040; <sup>2</sup>Department of Biology, Arizona State University, Tempe, AZ 85287-1501; <sup>3</sup>USDA, ARS, USHRL, 2001 South Rock Rd., Fort Pierce, FL 34945

**Research & Implementation Area:** Section A: Biology, Ecology and Population Dynamics

**Dates Covered by the Report::** 1999-2000

### **Replication of an Iridovirus In Whitefly Cells and Whitefly Nymphs**

A cell line from the silverleaf whitefly, BtB-2.97-Hunter & Polston, was tested for its ability to support replication of the insect iridescent virus 6 (IIV-6). Several lines of evidence indicated that a productive infection was achieved. The cells displayed cytopathic effects (CPE) that included a change in cell shape and the appearance of cytoplasmic inclusions. Virus particles accumulated in defined areas of the cytoplasm. The cell-associated virus titer was detected at three orders of magnitude higher than that released into the media ( $1.3 \times 10^8$  vs.  $1.1 \times 10^5$ ). Fluorescent antibody staining was used to visualize the virogenic stroma that initially was localized to areas adjacent to the nucleus in the cytoplasm of infected cells. The accumulation of viral DNA and proteins in the virogenic stroma led to the formation of the cytoplasmic inclusions that were observed as CPE. In addition, western blot analysis indicated CIV structural proteins were being expressed. Each of these lines of evidence indicated that whitefly cells could support a productive CIV infection. Nymphs were infected by feeding the virus in diet on an artificial rearing system. Virus was detected in nymphs by PCR and electron microscopy, but the infections were not highly pathogenic. Infection of whiteflies by IIV-6 suggests that more pathogenic viral isolates may be found for this insect and that the virus has potential to be used as an intracellular probe.

**Investigator's Names:** Dan Gerling and Dale B. Gelman

**Affiliation & Location:** Department of Zoology, Tel Aviv University, Ramat Aviv, e-mail: [dangr@post.tau.ac.il](mailto:dangr@post.tau.ac.il); Insect Biocontrol Laboratory, Bldg. 011A, Rm. 214, BARC West Beltsville, MD 20705, e-mail: [gelmand@ba.ars.usda.gov](mailto:gelman@ba.ars.usda.gov)

**Research & Implementation Area:** Section A: Biology, Ecology and Population Dynamics.

**Dates Covered by the Report:** Sept 2000 - July 2001

#### **Physiological Considerations in the Penetration of *Eretmocerus mundus* into *Bemisia* Nymphs**

*Eretmocerus mundus* oviposits under its host, *Bemisia tabaci*. Egg hatch occurs within ca. 3 days of oviposition. The first instar larva penetrates the host and development proceeds within a capsule of epidermal origin. Recent studies have shown that no matter under which host instar *E. mundus* eggs are laid, penetration into the host proceeds only during the host's fourth instar, typically sometime prior to the transition to adult characteristics. Thus, it is understandable that while *E. mundus* will oviposit under any of its host's four instars, the parasitoid prefers to lay an egg under either a second or third instar. Since approximately three days are required for parasitoid egg development, oviposition under a second or third instar whitefly will result in hatch occurring under a third or young fourth instar host; the period of time during which first instar parasitoid will be forced to remain outside its host will be minimal as will be its chances to miss the window during which the whitefly is vulnerable to penetration.

Epidermal capsule formation accompanies parasitoid penetration. The capsule surrounds/engulfs the parasitoid and must contribute to parasitoid development. Although the function(s) of the very large glands that are apparent in the first instar parasitoid larva is not known, it is possible that they influence the production of the capsule and that they may be essential for facilitating *E. mundus* penetration and/or development. The question as to whether host ecdysteroid levels are altered upon parasitization also was examined. Results showed that the level of ecdysteroids was always lower in a parasitized whitefly into which a first instar parasitoid had just begun to penetrate than in a healthy whitefly of the same developmental stage. Therefore, it does not appear that a parasitoid-induced increase in host ecdysteroid levels is involved in the stimulation of capsule formation. Rather, other regulatory molecules such as growth factors may be involved.

**Investigator's Name(s):** T. J. Henneberry, L. Forlow Jech and D. L. Hendrix

**Affiliation & Location:** USDA-ARS, Western Cotton Research Laboratory, Phoenix, AZ

**Research & Implementation Area:** Section A: Biology, Ecology, and Population Dynamics

**Dates Covered by the Report:** 2000

### **Sweetpotato Whiteflies, Cotton Aphids, and Sticky Cotton**

We compared the sugars extracted and cotton lint exposed to SPW and CA under laboratory conditions; also their effects on cotton lint stickiness as measured by thermodetector counts. Sweetpotato whitefly (SPW), *Bemisia tabaci* (Gennadius) Strain B (= *B. argentifolii*) and cotton aphid (CA), *Aphis gossypii* Glover, are the two most common honeydew producing insect species that occur on cotton. Honeydew contaminated lint is a serious problem in lint processing at the textile mill. It can also be difficult to harvest and gin. The major sugar components of the honeydew of both insect species are glucose, fructose, sucrose, trehalulose, and melezitose. Trehalulose and melezitose are insect-produced sugars. SPWs produce more trehalulose in relation to melezitose and the opposite is true for CAs. We exposed clean cotton lint to SPW or CA in the laboratory. The total sugar contents of water extracts of honeydew-contaminated lint after exposure to the insects were significantly correlated to increasing thermodetector counts (a measure of stickiness) that occurred as a result of increasing durations (days) of exposure. Higher concentrations of total sugars measured in these extracts occurred following exposures to SPW compared with CA. However, numbers of SPW and CA were different and the results are therefore not directly comparable. Research is continuing to further define differences and similarities in cotton lint stickiness as a result of honeydew lint contamination by SPW or CA.

**Investigator's Name(s):** T. J. Henneberry and L. Forlow Jech

**Affiliation & Location:** USDA-ARS, Western Cotton Research Laboratory, Phoenix, AZ

**Research & Implementation Area:** Section A: Biology, Ecology and Population Dynamics

**Dates Covered by the Report:** 2000

### **Open Cotton Boll Exposure and Development of Sticky Cotton in Whitefly Infested Fields**

We used thermodetector analysis to determine the effect on lint stickiness of different numbers of days of exposure of open cotton bolls to whitefly, *Bemisia tabaci* (Gennadius) Strain B (= *Bemisia argentifolii*) populations. Thermodetector lint stickiness measurement is accomplished by manipulating a 2.5 g sample of cotton lint into a fine mat, which is placed between two sheets of aluminum foil and heated under pressure. The foil sheets are separated and the number of sticky spots counted. Thermodetector counts do not distinguish between the contributions of the different sugars in honeydew or plant physiological sugars. However, the counts are an overall assessment of cotton lint stickiness. We also studied the effect of rainfall on sticky cotton.

Trehalulose and melezitose produced by *Bemisia* and thermodetector counts in cotton lint increased with increasing numbers of days of exposure of open cotton bolls in infested cotton plots. Thermodetector counts were significantly correlated to amounts of trehalulose and melezitose. Rainfall of 0.5 inch reduced trehalulose and melezitose in cotton lint within 5 h following the rain. The results suggest dissolution of the sugars followed by runoff as opposed to microbial degradation.

**Investigator's Name(s):** T. J. Henneberry, L. Forlow Jech, D. L. Hendrix, T. de la Torre

**Affiliation & Location:** USDA-ARS, Western Cotton Research Laboratory, Phoenix, AZ

**Research & Implementation Area:** Section A: Biology, Ecology and Population Dynamics

**Dates Covered by Report:** 2000-2001

**Cotton Water Stress Effects on *Bemisia tabaci* Strain B (Homoptera: Aleyrodidae) on Honeydew Production**

Comparisons of honeydew production by sweetpotato whitefly (SPW), *Bemisia tabaci* (Gennadius) Strain B while feeding on water-stressed or non-water-stressed cotton showed that more honeydew sugars were produced on non water-stressed leaves of cotton plants (four days after irrigation) compared to those feeding on leaves of cotton plants 7 or 13 days after irrigation (water stress). Leaf water potentials, as a measure of water stress, decreased with increasing numbers of days following irrigations. Leaf water potentials of furrow irrigated cotton and furrow irrigated plus supplementary drip irrigated cotton (1 h per day) showed the same decreasing leaf water potential patterns. The decreases, however, were less in furrow irrigated plus supplementary drip irrigation compared with furrow irrigation alone. SPW feeding on plants in the field with furrow plus supplementary drip irrigation and in the greenhouse on non-stressed cotton plants produced more micrograms of honeydew sugars per gram of honeydew compared to SPW feeding on plants with furrow irrigation alone in the field and on water-stressed plants in the greenhouse.

**Investigator's name:** Kamil Karut<sup>1</sup>, Erdal Sekeroglu<sup>1</sup>

**Affiliation & Location:** <sup>1</sup>Cukurova University, Agricultural Faculty, Department of Plant Protection, 01330, Adana, Turkey

**Research & Implementation area:** Section A: Biology, Ecology and Population Dynamics

**Dates Covered by the Report:** 1999

**Distribution of Immature, Adult and Immature Parasitoids of *Bemisia tabaci* Genn. (Homoptera: Aleyrodidae) in Cotton, Cukurova, Turkey**

Cotton is one of the important cultivated plants in Cukurova, Turkey. The climate in Cukurova region is subtropical, with hot dry summers and mild winters. The cotton growing season lasts from the end of March to the beginning of October. Many pests are attacked cotton in this period. *Bemisia tabaci* Genn has been one of the most important pest of cotton under irrigated conditions. Although its populations vary from year to year, it is still the key pest in Cukurova, causing economic damage and yield reduction. However, there is no efficient sampling plan to estimate or classify populations of whitefly for management purposes. The first step in developing a sampling plan is to define the sampling unit and random variable of spatial distribution.

Studies were conducted to determine the sampling unit and spatial distribution of immature and adult *B. tabaci* and immature parasites. For this purpose 20 plants were examined at five different dates for immature *B. tabaci* and parasites. Adults were counted by leaf turn method on a randomly selected leaf from top, middle and bottom part of the cotton plants at nine different dates. Also adults counted on individual mainstem leaf from node 2 through 8 (terminal:node 1) at four different date. All studies were conducted in 1998-1999, but only data from 1999 was used to determine the distribution pattern.

We looked at 3 parameters to describe the distribution of whitefly and parasitoid life-stages among various leaf positions as follows: (1) the leaf position with the highest percentage of a particular stage (2) leaf position where insect counts were best correlated with counts on the total plant (3) the coefficient of variation. The pattern of aggregation for immature, adult *B. tabaci* and immature parasites was measured by Taylor's power law. All 3 distribution parameters changed throughout the season. Across the entire season, the 3 distribution parameters for whitefly eggs were associated with leaf positions 2, 3 and 4; for nymphs with leaf positions 5, 6 and 7; with pupae (red-eye) leaf positions 6, 7, 8, 9 and 10; and for immature parasitoids with leaf positions 8, 9, 10 and 11. Based on consideration of Taylor's power law and precision, it was most efficient to sample leaf 3 for eggs, leaf 5 for nymphs, leaf 10 for pupae and immature parasitoids.

Adult were consistently more abundant on leaves from the top than leaves from the middle and bottom strata. According to the 3 parameters, it was most efficient to sample mainstem leaf 4 for whitefly adult stage.

**Investigator's Names:** Steven E. Naranjo & Thomas J. Henneberry

**Affiliation & Location:** USDA-ARS, Western Cotton Research Laboratory, Phoenix

**Research & Implementation Area:** Section A: Biology, Ecology and Population Dynamics

**Dates Covered by the Report:** January 2000-December 2001

**Development of Sampling Methodology for Estimating Cotton Lint Stickiness  
due to Infestations of *Bemisia tabaci***

Sticky cotton continues to be a worldwide problem that may presently be considered by the cotton industry as the most serious factor affecting cotton quality. The nature and causes of lint stickiness are complex. Plant-produced sugars have been implicated in the sticky cotton problem; however, a high percentage of the documented lint stickiness problems have been associated with the occurrence of honeydew-producing insects such as aphids and whiteflies. Honeydew induced stickiness is problematic at many post-harvest phases of lint processing including ginning, carding, and particularly spinning. The sticky cotton problem in the U.S. has increased with recent population outbreaks of the whitefly *Bemisia tabaci* and research is currently addressing an array of issues from characterization of sugars to management of plant and insect sources.

The accurate determination of cotton lint stickiness in the field could greatly aid management and research efforts. Field sampling of stickiness could augment pest monitoring techniques and improve overall decision-making for pest suppression. Field sampling could also identify unacceptable levels of stickiness at harvest that may require some remedial action such as the use of enzyme treatment systems that are currently being investigated. Our research targets the development of standard methodologies for collecting lint samples from fields to ensure statistically precise estimates of stickiness. Two assay systems were investigated, the manual thermodetector (SCT) and the high speed (automated) sticky cotton thermodetector (H2SD). The development of sampling plans involves four inter-related components: 1) description of the sampling distribution, 2) selection of the optimal sample unit, 3) partitioning of variance components, and 4) determination of the number and allocation of sample units.

The newest H2SD system consistently detects more thermodetector spots than the SCT and the relationship between the platforms appears to be nonlinear, at least for field-collected samples encompassing a very wide range of stickiness levels. The distribution of sticky lint in the field is random regardless of which platform is used to assay the lint. Time and cost considerations are critical to any sampling procedure. Examination of many different sizes of field sample units suggest that smaller sample units (e.g. all lint from 1 plant or lint from 20 bolls collected at random) are more cost-efficient than larger sample units. Further analyses of variance components suggests that only a single assay for each sample unit should be conducted with the SCT, but that 3 assays should be conducted for each sample unit on the H2SD. This analysis takes into account the relative costs of sample collection in the field and the time required to complete a single assay with each platform.

The assay system (SCT or H2SD) used is critical to the development of a final field sampling plan; however, there is considerable flux in the industry relative to a standard platform for stickiness testing. Given this uncertainty, we have developed sampling plans for both platforms. Reasonably precise estimates of stickiness can be achieved with a sample size of about 15 on either platform using the 1-plant sample unit. Even fewer samples would be required using a 20-boll unit. Sample sizes as high as 41 or 82 would be required for estimates with very high precision on the H2SD or SCT platform, respectively. Overall sampling costs are much lower for the H2SD because of the speed with which samples can be assayed. Sampling efficiency could be further improved with sequential sampling strategies or with sample plans focused on categorizing (non-sticky vs. sticky) rather than estimating actual levels of stickiness. These sampling applications await further research to define critical levels of stickiness and/or the implementation of wide-scale testing where any time savings would be at a premium.



**Investigator's Names:** Steven E. Naranjo & Rebecca Burke

**Affiliation & Location:** USDA-ARS, Western Cotton Research Laboratory, Phoenix

**Research & Implementation Area:** Section A: Biology, Ecology and Population Dynamics

**Dates Covered by the Report:** January 2000-December 2001

### **Movement and Mortality of *Bemisia tabaci* Crawlers on Cotton**

Greenhouse studies were conducted to examine movement and estimate rates of mortality of crawler stage *Bemisia tabaci* on cotton. Five female whiteflies were allowed to oviposit for 24 h in clip cages on the lower surface of a single leaf (generally the 2<sup>nd</sup> to 3<sup>rd</sup> leaf from the terminal) of cotton plants that had 6-8 expanded leaves. After removal of adults, plants were then placed in organandy cages to prevent further oviposition. One to two weeks after oviposition plants were destructively sampled, and the location (leaf node, upper or lower leaf surface) and number of settled nymphs were recorded. The petiole lengths of all leaves and internode distances were measured and recorded. The number of hatched and unhatched eggs on the oviposition leaf also were recorded. The experiment was replicated 20 times in still air and 20 times with plants exposed to a table-top fan blowing air at a velocity of approximately 2.2 meters/second. This latter condition was used to simulate the effects of a moderate breeze on crawler movement and mortality.

The survival of crawlers did not differ statistically between still-air and wind treatments and averaged 85.5% overall. Survival ranged from 50 to 100% on individual plants. Over 99% of all surviving crawlers settled on the lower surface of leaves and 99.9% settled on the leaf where eggs were oviposited. Over both treatments only 3 individuals (out of 2700 surviving nymphs) moved from the leaf of origin and they all moved to leaves lower on the plant. Assuming that these individuals crawled rather than fell, the mean distance moved was 200 mm. These results suggest that crawlers move very little and that survival is high on young cotton plants in the absence of other natural factors such as natural enemies. Further study is needed in the field over a wider range of conditions to evaluate natural levels of mortality for life table analyses.

**Investigator's Name(s):** Dennis R. Nelson, James S. Buckner and Thomas P. Freeman

**Affiliation & Location:** Biosciences Research Laboratory, USDA-ARS, Fargo, ND and Electron Microscopy Center, Dept. of Plant Pathology, North Dakota State University, Fargo, ND

**Research & Implementation Area:** Section A: Biology, Ecology and Population Dynamics

**Dates Covered by the Report:** 2000 and 2001

### **Comparison of the Feeding Process of Nymph and Adult Whiteflies**

With scanning electron microscopy we have examined the feeding processes of nymph and adult whiteflies. The *Bemisia argentifolii* adult feeding apparatus consists of a four-segmented labium in which is contained a slender stylet bundle (1.5-2 micrometers in diameter), consisting of two mandibles and two maxillary stylets. The stylet bundle of the adult is completely contained within the groove in the labium when not feeding. To penetrate the leaf, the adult places the tip of the labium against the leaf and then lowers its head in order to slide the stylet bundle down the labial groove and push the tip of the stylet bundle into the leaf. The length of the stylet bundle, averaging 217 micrometers, is sufficient to allow the adult to reach phloem tissue from any position on the leaf, from either abaxial or adaxial surfaces.

The total length of the adult stylet bundle is determined by measuring the distance from the junction of labial segments 1 and 2 to the tip of the labium of non-feeding adults. The length that the stylet bundle has penetrated the leaf can be determined by measuring the distance from the junction of labial segments 1 and 2 to the position of the head (where the stylet bundle enters the labial groove) along the labial groove. This distance varied from 43 to 151 micrometers indicating that the adult uses less than 70% of the length of its stylet to reach a phloem bundle in cotton. The distance from the abaxial epidermal surface to the phloem in the cotton leaves we examined ranged from 53 to 127 micrometers.

The stylet bundle of the immature whitefly, *Bemisia argentifolii*, appears to be looped or coiled within the body. The end of the bundle enters a groove in the rostrum and extends beyond the rostrum tip. In order to penetrate the leaf, the nymph extends the stylet bundle beyond the tip of the rostrum in an unknown manner. The length of the stylet bundle ranges from a minimum of 110 micrometers in the crawler to a maximum of 200 micrometers in the 4th instar. The length of the stylet bundle is not proportional to the length of the nymph. From the crawler to the 4th instar the length of the bundle only increased 39% while the length of the nymph increased 168%. The length of the nymph stylet bundle appears to be of sufficient length to reach phloem from any position on the leaf surface.

**Investigator's Name(s):** M.S.Palaniswami, Binu Antony and Lisha Vijayan. S

**Affiliation & Location:** Division of Crop Protection, Central Tuber Crops Research Institute, (Indian Council of Agricultural Research), Trivandrum 695 017, India

**Research & Implementation Area:** Section A: Biology, Ecology and Population Dynamics

**Dates Covered By The Report:** January 2000 - December 2000

#### **Life Table Studies of *Bemisia tabaci* Genn. on Cassava**

An investigation was undertaken to study the age specific mortality of individuals in the whitefly population on cassava. Cassava plants (cv. M4 & H226) were raised in mud pots (21' diameter) placed in five groups of ten plants each. When cohorts of settled first instar nymphs were established from natural population; three leaves were selected from each plant and their location was marked with a non-toxic indelible marker pen. The fate of the each individual was then tracked by visual observation using a pocket microscope (10-60x magnification), every third day to estimate mortality due to predation, parasitisation and missing/unknown factors.

During the first observational period (18 January - 20 February, 2000 ) first instar nymphs selected were 17.2/leaf, and pupae and adults emerged were 2.0 and 2.5/leaf, respectively. Predation was the prime source of mortality (43.6%) followed by missing/unknown (28.3%) and parasitisation (15.2%). During the second observational period (2 February - 22 February 2000), first instar nymphs mean of 17.4/leaf was observed and emergence of pupae and adults averaged 2.9 and 0.9/leaf, respectively. In this period, 50.4% of mortality was due to missing/unknown factors and 26.8% and 22.4% due to predation and parasitisation, respectively. During the third (9 - 31 March), fourth (4 -30 April) and fifth observational period (9 - 26 May), more adults emerged. Out of 11.4, 17.6 and 13.8 nymphs/leaf during March, April and May, respectively; 5.2 pupae and 4.4 adults during March, and 9.7 pupae and 8.2 adults during April, and 8.0 pupae and 7.1 adults emerged during May. This may be probable due to the favourable climatic/weather factors. In the last three observational periods, missing / unknown factors contributed to maximum mortality (36.8% / leaf, March, 22.9%, April and 19.7% May). While predator induced mortality was 15.9, 11.6 and 19.3 % during March, April and May, respectively. During April, parasitisation (13.8%) outnumbered predation (11.6%).

Natural enemies observation were aphelinid parasitoids, predatory coccinellids, dolichopodids, mites and spiders. Adult and larval stage of the coccinellids were found to feed on all the stages of the host. Usually adult predatory mites sucked on the body fluid of the host. Missing/unknown factors were a significant source of mortality in most of the experiments, followed by predation and parasitism. However, predation and parasitism contributed to high mortality to immature stages.

**Investigator's Name(s):** M.S.Palaniswami, Lisha Vijayan, .S and Binu Antony

**Affiliation & Location:** Division of Crop Protection, Central Tuber Crops Research Institute (Indian Council of Agricultural Research), Trivandrum 695 017, India

**Research & Implementation:** Section A: Biology, Ecology and Population Dynamics

**Dates Covered by the Report:** January -December 2000

### **Presence of Biotypes in *Bemisia tabaci* Genn. in India**

Studies were undertaken to investigate the host associated variation in *B. tabaci*. Host transfer studies using clip cages and host preference studies using host plants, cassava, sweetpotato, egg plant, tobacco and cotton showed two host associated strains of *B. tabaci* in India. The two strains were identified as cassava biotype and sweet potato biotype. The whitefly population from sweet potato did not colonize on cassava, similarly whiteflies from cassava showed limited colonization on cotton but failed to complete its development on sweet potato. Egg plant and tobacco proved to be the common host for both. The cassava biotype showed 96.7% adult emergence on cassava compared to 70.0% on egg plant followed negligible on cotton and nil on sweet potato. Whereas with sweet potato biotype showed 90.7% adult emergence on sweet potato followed by 78.8% on cotton, 55.2% on egg plant and no adult emergence on cassava.

The duration or the life cycle of sweet potato whitefly was maximum on brinjal (20.2 days). The cassava whitefly showed the highest total developmental duration on cassava (19.2 days). The longevity of cassava whitefly on sweet potato and cotton and that of sweet potato whitefly on cassava, and cotton were also studied. The longevity of the cassava whitefly ranged from 4-10 on cotton with a mean of 7.8 days and 4-6 on sweet potato with mean of 4.7 days. The longevity of the sweet potato whitefly ranged from 2-7 days on cassava and 14-29 days on cotton, with their means 4.2 and 17.7 days respectively.

The studies on esterase profile corroborated the presence of two different biotypes. There was clear difference in the esterase banding pattern between the two biotypes.

Cassava mosaic virus transmission studies conducted using seedlings of cassava and cassava whitefly biotype could make successful transmission of CMD from infected to healthy seedlings. Symptoms developed within 9-30 days and were confirmed by ELISA.

**Investigator's Name(s):** M.S.Palaniswami, Binu Antony and Lisha Vijayan. S

**Affiliation & Location:** Division of Crop Protection Central Tuber Crops Research Institute (Indian Council of Agricultural Research), Trivandrum 695 017, India

**Research & Implementation:** Section A: Biology, Ecology and Population Dynamics

**Dates Covered by the Report:** January 1999- December 2001

### **Life Table Studies of *Bemisia Tabaci* Genn. on Cassava and Laboratory Evaluation of Potentiality of Predators**

An investigation was undertaken to study the age specific mortality of individuals in the whitefly population on cassava. Cassava plants (cv. *M4 & H226*) were raised in mud pots (21' diameter) placed in five groups of ten plants each. When cohorts of settled first instar nymphs were established from natural population; three leaves were selected from each plant and their location was marked with a non-toxic indelible marker pen. The fate of the each individual was then tracked by visual observation using a pocket microscope (10-60x magnification), every third day to estimate mortality due to predation, parasitisation and missing/unknown factors.

The observations were grouped into four observational periods. During the first observational period adult survivorship (11.3%) was very low due to increased predation followed by unknown factors. The predator mortality was highest in the first instar to pupal stage. During the second observational period, adult survivorship (4.72%) was lowest among the four observational periods. Mortality from parasitisation was maximum in the pupal to adult stage. Mortality for missing/unknown factors was highest during the third observational period. The percentage of adult survival was 46.6%. Adult survivor (55%) was highest due to decreased predation and parasitisation.

The overall survival from the first instar to the adult ranged from 4.72-55% during these periods. Parasitisation was the dominant mortality factor from pupae to adult emergence.

Natural enemies noticed during observations included aphelinid parasitoids, predatory coccinellids, mites and spiders. Adult and larval stages of the coccinellids were found to feed on all stages of the host. Predatory mites were found to suck the body fluid of the host, and this was the most dominant mode of predation. Missing/unknown factors were a significant source of mortality in most of the experiments, followed by predation and parasitism. However, predation and parasitism together contributed to high mortality of immature stages.

The predation efficiency of five coccinellid predators and predatory mite were studied in the laboratory. *Serangium paracesetosum* ( $91.45 \pm 5.32$ ) was found to be dominant over the other five predators. *Anegleis cardoni* was less effective than *S. paracesetosum* but superior to the other four species. The predation efficiency of the four other predators was at par and showed no significant differences.

**Investigator's Name(s):** M.S.Palaniswami, Lisha Vijayan S and Binu Antony

**Affiliation & Location:** Division of Crop Protection Central Tuber Crops Research Institute (Indian Council of Agricultural Research) Trivandrum 695 017, India.

**Research & Implementation Area:** Section A: Biology, Ecology and Population Dynamics

**Dates Covered by the Report:** January 1999 - December 2001

**Isozyme, Vector Capability and Cross Breeding Studies to Confirm the B biotypes in *Bemisia tabaci* (Genn.) in India**

Host associated variation studies revealed the presence of two strains of *Bemisia tabaci*- Cassava whitefly and Sweet potato whitefly, which showed clear-cut host preferences. Additional confirmations were carried out by analysing the variations in the isozyme patterns, vectoring capabilities and by doing cross-breeding studies.

Two enzyme systems studied under isozyme analysis includes Esterase and Malate dehydrogenase (MDH). The esterase-banding pattern in sweet potato whitefly differed from the cassava whitefly in having an additional fast moving esterase band (Rf value 0.293). The migration distances of the two slow moving bands, common to both were also different (0.150 and 0.204 in CWF and 0.108 and 0.165 in SPWF). Banding patterns using the MDH system (Rf values 0.104 and 0.162) was identical in both biotypes.

Investigations on the role cassava biotype as a vector of Indian Cassava Mosaic Virus (ICMV) was confirmed by ELISA using monoclonal antibodies raised (MAbs) to ICMV isolates. Cassava biotypes could successfully transmit ICMV, whereas sweet potato biotype failed to transmit. A single whitefly could transmit ICMV with 10h AAFP & IAAP. Maximum percentage of transmission was obtained when AAFP & IAAP were 48h. This fact corroborated the indication of biotypes of *B. tabaci* in this region.

The biological consequences of mating interaction between the two biotypes of *B. tabaci* were studied by cross breeding female SPWF and male CWF, and female CWF and male SPWF. This showed an increased proportion of male progeny indicating that cassava and sweet potato are two biotypes and not separate species.

Thus biological and electrophoretic studies and vectoring efficiency have very clearly revealed the presence of two distinct biotypes of *B. tabaci* – a cassava biotype and a sweet potato biotype in India. This is the first report on a scientific basis.

**Investigators Name:** Bahram Tafaghodinia

**Affiliation & Location:** Iranian Research Organization of Science & Technology, Tehran, Iran

**Research & Implementation Area:** Section A: Biology, Ecology and Population Dynamics

**Dates Covered By The Report:** Summer, 2001

### **Engineering-Based Computer Simulation for Modeling Greenhouse Whitefly Population**

Development of a mathematical model is a suitable technique for understanding insect population dynamics. It offers a possibility to bring together the results of the extensive research conducted and recognizes possible gaps in our knowledge. Once the model has been verified and validated, it can be used to obtain qualitative and quantitative information on the importance of certain relations or parameters and it may serve to predict the reaction of a system on a change of the values of parameters, variables etc.

It was decided to restrict the model to one host-plant species, from which the most experimental data were available, so tomato was used as a host-plant in this research. A simulation model was constructed since dynamic model was desired. SLAM II is used as an engineering-based computer simulation language. This simulator was originally developed for modeling industrial manufacturing processes. This software was used to simulate the population growth of the greenhouse whitefly. Temperature was used as one of the abiotic parameters. Whitefly females were then given a value for daily fecundity equal to 5. At this point, the female was checked and if its lifetime was terminated it was routed to a death or termination node. If it was not terminated, it was allowed to lay its complement of eggs and then routed back to the fecundity node. It continued in this cycle until death. The eggs were each assigned the current simulation time as their "birth day". Next 6.1% of eggs were routed to termination node corresponding to egg mortality. The remaining eggs were sent to another assignment node that was assigned egg development time. The model continued in this manner until all eggs developed to pupae. The total population was obtained by summing these five activity counts. The program of the whitefly model was ran (with TRACE option).

To validate the part of the model that described the developmental stages, there were two data sets available for comparison. The data came from an experiment that was carried out in the greenhouse. In this experiment the numbers of developmental stages of two sets of 106 and 115 whitefly eggs on tomato leaves were followed and recorded daily at average 18 C° and %65±5 RH. The average developmental period from egg to adult took 52 days, while it lasted 56 days in the simulation. The average developmental period for the second data set took 52 days, while it lasted 58 days in the simulation. The mortality was lower than simulated particularly during egg and second larval stages. The average of mortality in greenhouse was 21.2% and in model was 24%.

To validate the part of model that described the population growth, there were two data sets available for comparison. The data came from an experiment carried out in greenhouse conditions. In this experiment, the numbers of empty pupae of two sets of 106 whitefly eggs on tomato leaves were followed and recorded weekly at 18±2 C° and %65 RH in autumn and 100 whitefly eggs in average 24 C° and 65%±5 RH in spring.

The greenhouse data was above the curve of the model. This may be related to temperature conditions in the model and greenhouse. The development of immature stages of greenhouse whitefly can be predicted with this model. But sensitive analysis shows the developmental duration has the most effect on population growth.

## Section A Research Summary

Compiled by D. N. Byrne and J. L. Blackmer

Recent greenhouse studies examined dispersal by *B. tabaci* crawlers on cotton. Crawlers were found to move as far as 200 mm. Ongoing studies examined seasonality and mortality patterns of *B. tabaci* on a variety of crop plants and weeds. Owing to their polyphagy, a large number of plant species support whitefly populations at various times of the year. Predation and dislodgment were found to account for a significant portion of *Bemisia* mortality. Age specific mortality was investigated on cassava plants. Adult survivorship depended on time of year. On cassava, parasitism was the dominant mortality factor from pupal to adult emergence. In laboratory studies, *Serangium paracastosum* was the most important predator. Studies were conducted to determine the most appropriate sampling unit as well as spatial distribution of immature and adult *Bemisia*. Similar factors were examined for immatures and parasitoids. Units and distribution were stadia-dependent. As a result of two related studies, improvements have been made in the CC trap. Two assay systems for detecting sticky cotton were investigated, the manual thermodetector and the high speed (automated) sticky cotton thermodetector. Sampling costs are much lower for the automated system because samples can be assayed more quickly. Field and laboratory studies in the Southwest indicated that *Eretmocerus eremicus* may have a more limited ability to disperse than *Bemisia*. This may limit their use as biological control agents in southwestern agriculture. In India, studies revealed the presence of two strains of *Bemisia tabaci* - the cassava whitefly and the sweet potato whitefly. The ability of the cassava biotype to vector Indian Cassava Mosaic Virus was confirmed. A system based on body depth and maturation of the adult eye was used to track development and identify physiological synchronous 4<sup>th</sup> instar/parhate adult *Bemisia*. An examination of histological sections of the last instar revealed that adult eye and wing development were initiated in Stage 6. Wings were deeply folded by Stages 6 and 7. Ecdysteroid titers peaked at Stages 4-early 6, just before the onset of adult development. In Mexico, numerous whitefly species have been identified including *Bemisia*, *Trialeurodes vaporariorum*, *T. abutilonea*, *Aleurothrixus floccosus* (Maskell), *Tetraleurodes acaciae* and *Tetraleurodes acaciae*. The identification and composition of the fatty acids associated with the major lipid classes within *Bemisia* nymphs were determined. All lipid

classes contained variable distributions of eight fatty acids: myristic acid, palmitic acid, stearic acid, arachidonic acid, palmitoleic acid, oleic acid, linoleic acid, and linolenic acid. The triacylglycerol composition of *Bemisia* adults was characterized, and the major fatty acid constituents were identified as oleic acid, palmitic acid, stearic acid, and linoleic acid. Several different types of bacteria were cultured from surface-sterilized *Bemisia* adults and nymphs, including *Bacillus* spp. *Enterobacter cloacae*, was found within the gut cells of adults. The rostrum of the *Bemisia* nymphs was found to be short and with sensilla on the tip. Stylets range from 110  $\mu\text{m}$  in the first instar to over 200  $\mu\text{m}$  in the fourth instar. Prior to the onset of feeding, the crawlers stylet is folded within the body of the nymph. A newly developed artificial rearing system was used to determine how biotic and abiotic factors influenced egg hatch, crawler establishment and development of *Bemisia*. Egg age, day length, light intensity and density of eggs placed on the artificial membrane influenced hatch rates, establishment of crawlers, and to a certain extent subsequent development of nymphs. Recent studies indicated that no matter on which host instar *E. mundus* eggs are laid, penetration into the host proceeds only during the host's fourth instar. A cell line from the *Bemisia*, BtB-2 tested for its ability to support replication of the insect iridescent virus 6 (IIV-6).



**Table A. Biology, Ecology, and Population Dynamics.**

Research Approaches	Year 1 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Determine life cycle vulnerabilities (life tables)<sup>a</sup>, population development and natural mortality factors, natural enemies on major crops, urban plantings, weeds and predict overwintering potential.</b>	Whitefly and natural enemy sampling in cultivated crops, urban planting and weed hosts.	X		Partial life table analyses have been completed for <i>B. argentifolii</i> on cotton in Arizona. Natural forces, including predation and dislodgment are major mortality factors; parasitism was a minor source of mortality. Survivorship from egg to adult ranged from 0-8.5% over 4 generations in sprayed and unsprayed fields. Studies on wild host crops in Israel indicate that parasitoids may contribute to low levels of whitefly on lantana. Whitefly and natural enemy populations were monitored in cropping systems in the Imperial and San Joaquin Valleys of California, Maricopa, Arizona and the Rio Grande Valley of Texas. The spread of <i>B. argentifolii</i> is being documented in Brazil. Life table studies provide valuable quantitative information on sources of whitefly mortality; surveys define the temporal and spatial dynamics of pest and natural enemy populations. This information is critical in developing and refining more biologically-based management systems.
<b>Develop sampling methodology, action and<sup>b,c</sup> economic thresholds for all major crops. Sampling methods and thresholds modified in light of natural enemy levels and existing management strategies.</b>	Initiate whitefly to identify spatial and temporal distributions in major cultivated crops.	X		Relationships between whitefly density and the occurrence of tomato irregular-ripening as well as preliminary sampling plans for whitefly on tomato have been developed. Evaluations of a reusable trap for surveying adult whiteflies in various crops are continuing. Studies of the effects of various insecticides on whitefly natural enemies are ongoing. Sampling plans and action thresholds are still needed for a number of affected crops.

**Table A. Biology, Ecology, and Population Dynamics. (Continued)**

Research Approaches	Year 1 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Develop population models to describe and predict whitefly population growth and spatial and temporal distribution. Develop simple day-degree sub-models for estimating phenology and temporal patterns of whitefly, natural enemies and host crops.</b>	Summarize whitefly biology, ecology and plant phenology to identify whitefly host plant interfaces.	X		Development of large-scale temporal and spatial models and temperature-dependent, site-specific population dynamics models continues. Such models have the potential to encapsulate our current knowledge and provide a framework for developing more efficient management systems. However, considerable biological and ecological detail, as well as information on various aspects of pest management is available and needs to be integrated into these models to make them most useful as exploratory tools.
<b>Develop sampling methods for quality of cotton lint, vegetables and other commodities.</b>	Initiate sampling of seed cotton in the field during the season, at harvest, after picking, moduling and ginning.	X		Research has characterized the temporal distribution of honeydew deposition by <i>B. argentifolii</i> in cotton, improved our understanding of the relationship between lint stickiness and whitefly abundance and compared the production of trehalulose and melezitose between nymphs and adults. Studies reveal that cotton lint stickiness is randomly distributed in cotton fields. Preliminary sampling plans have been developed for estimating pre-harvest cotton lint stickiness. Stickiness constitutes one of the most important problems currently facing the cotton industry.
<b>Quantify whitefly and natural enemy dispersals and contribution to population dynamics.</b>	Review and analyze existing knowledge of whitefly dispersal.	X		Studies have characterized the aerial distribution of whiteflies dispersing from cantaloupe fields and have examined the trade-offs between oogenesis and flight activity. Studies on whitefly parasitoid dispersal are ongoing. Understanding and predicting the timing and extent of the movement of whiteflies and their natural enemies is an important component in developing areawide management systems.

**Table A. Biology, Ecology, and Population Dynamics. (Continued)**

Research Approaches	Year 1 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Define mating behavior, reproductive isolation, species, biotypes.</b>	Initiate studies on mating, oviposition and other behavior.	X		Surveys worldwide continue to document the spread of <i>B. argentifolii</i> . Electrophoretic analyses demonstrate the presence and extent of this pest in throughout Australia and Brazil. <i>B. argentifolii</i> appears to be displacing <i>B. tabaci</i> Biotype A in Brazil and is having a large impact on agricultural production through direct feeding and geminivirus transmission. Reports of heterozygotes between <i>B. argentifolii</i> and the extant Australian type of <i>B. tabaci</i> corroborates previous laboratory and highlight the taxonomic challenges within the <i>Bemisia</i> species complex.
<b>Validate <i>Bemisia</i> taxa morphology, genetic, biochemical, and biology characteristics.</b>	Continue examination of <i>Bemisia</i> sp. for distinct morphological character differences.	X		Comparative morphological analyses have been completed on <i>Bemisia</i> pupae from around the world. Several of these characters are highly variable among populations suggesting that pupal morphology should not represent the sole criteria for classifying individuals within the <i>Bemisia</i> species complex.
<b>Define role of endosymbionts in metabolism, host adaptation, nutrition and survival.</b>	Identify endosymbionts in whitefly.	X		The effects of antibiotics on the biology of <i>B. argentifolii</i> have been examined. Several antibiotics that interfere with bacterial protein synthesis affected growth and development of immatures, but none affected oviposition rates or sex ratio. Results have important implications for the use of antibiotics to disrupt the function of whitefly endosymbionts and other associated microbes as potential control methods.
<b>Characterize nutrient uptake and metabolism</b>	Determine the process of uptake and metabolism of carbohydrates, amino acids and other nutrients.	X		High levels of a polyol, sorbitol, were associated with elevated ambient temperatures. Sorbitol may function as a thermoprotectant in whiteflies that enables them to thrive in desert environments. The pathway of sorbitol synthesis and degradation in <i>B. argentifolii</i> is unique and may offer an avenue to develop transgenic plants which could disrupt sorbitol synthesis and compromise the whiteflies ability to deal with heat stress.

**Table A. Biology, Ecology, and Population Dynamics. (Continued)**

Research Approaches	Year 1 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Develop whitefly artificial diets and natural enemy mass-rearing.</b>	Identify whitefly nutritional components in plant tissue.	X		An artificial diet and feeding system for rearing immatures of <i>B. argentifolii</i> has been developed. Rates of development of individual instars were comparable to those estimated on various host plants. The feeding system has proven to be a useful bioassays for examining diet components and for studies of primary metabolism based on defined diets, and has the potential to provide a means of mass rearing whitefly parasitoids.

<sup>a</sup> Natural enemy research complements from Section D, see Table D.

<sup>b</sup> Action and economic thresholds also apply in Section C, see Table C.

<sup>c</sup> Sampling technology applicable to all other sections, see Tables B to F.

**Table A. Biology, Ecology, and Population Dynamics.**

Research Approaches	Year 2 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Determine life cycle vulnerabilities (life tables)<sup>a</sup>, population development and natural mortality factors, natural enemies on major crops, urban plantings, weeds and predict overwintering potential.</b>	Determine potential of intercrop weed host & urban planting, movement of whiteflies and natural enemies.	X		Life table studies continued to characterize and quantify mortality factors for immatures of <i>B. argentifolii</i> on cotton. Predation and dislodgment accounted for much of the mortality in untreated fields and survivorship from egg to adult ranged from 0-18.2% over 6 generations. Several perennial plants species show potential to serve as refugia for exotic and native parasitoids. Life history and reproductive potential has been studied on various crop and weed hosts in the US and Italy. Whitefly population dynamics and virus incidence has been examined in cropping systems in Costa Rica, India and Guadeloupe. These ecological and biological studies form the foundation of effective pest management strategies.
<b>Develop sampling methodology, action and<sup>b,c</sup> economic thresholds for all major crops. Sampling methods and thresholds modified in light of natural enemy levels and existing management strategies.</b>	Analysis and identification of needed additional sampling research to develop appropriate sampling protocol.	X		A multistate study determined action thresholds for cotton in Arizona and California. Evaluations of a reusable trap for surveying adult whiteflies in various crops are continuing. Studies of the effects of various insecticides on whitefly natural enemies are ongoing. Sampling plans and action thresholds are still needed for a number of affected crops.
<b>Develop population models to describe and predict whitefly population growth and spatial and temporal distribution. Develop simple day-degree sub-models for estimating phenology and temporal patterns of whitefly, natural enemies and host crops.</b>	Begin model development to include all biological and plant phenology data in simulation development.	X		The first version of a temperature-dependent, site-specific population dynamics model of <i>B. argentifolii</i> in cotton and cantaloupe was completed. Additional refinements, enhancements and field validation are needed to improve the utility of the model for predicting whitefly population dynamics under various management regimes and environmental conditions. In general, considerable biological and ecological data are available and need to be integrated into these models to make them most useful as exploratory tools.

**Table A. Biology, Ecology, and Population Dynamics. (Continued)**

Research Approaches	Year 2 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Develop sampling methods for quality of cotton lint, vegetables and other commodities.</b>	Based on year 1 results, expand and repeat sampling protocols as described.	X		Comparative evaluations of manual and high speed cotton stickiness thermodetector revealed differences in performance that have important implications for the development of measurement scales for stickiness and the number of samples that would need to be collected for the precise estimation of stickiness. Research on quality-related problems in other affected crops is needed.
<b>Quantify whitefly and natural enemy dispersals and contribution to population dynamics.</b>	Validate times of whitefly dispersal, environmental factors and identify modifying factors.	X		Studies on whitefly and parasitoid dispersal are ongoing in the desert southwest. Understanding and predicting the timing and extent of the movement of whiteflies and their natural enemies is an important component in developing areawide management systems.
<b>Define mating behavior, reproductive isolation, species, biotypes.</b>	Define interspecies interbiotype mating interactions.	X		Research continues on the role of reproductive isolation in the formation of species and biotypes, using insects from around the globe. There has been little detailed study of mating behavior <i>per se</i> , and its relevance for mating incompatibility.
<b>Validate <i>Bemisia</i> taxa morphology, genetic, biochemical, and biology characteristics.</b>	Develop genetic molecular level and acceptable species level separation.	X		Molecular characterization of the global whitefly complex is ongoing to clarify the taxonomic relationships between <i>Bemisia</i> whitefly populations. The whitefly karyotype has been determined and is an important development in our understanding of whitefly reproduction.
<b>Define role of endosymbionts in metabolism, host adaptation, nutrition and survival.</b>	Determine role of endosymbionts in whitefly biological functioning.	X		The discovery of <i>Wolbachia</i> endosymbiotic bacteria in whiteflies is a new development that has significant implications for development of control strategies targeting the reproductive biology of whiteflies.

**Table A. Biology, Ecology, and Population Dynamics. (Continued)**

Research Approaches	Year 2 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Characterize nutrient uptake and metabolism</b>	Determine the biochemical pathways for metabolism of compounds essential for whitefly development.	X		Fundamental questions about the nutritional physiology of whiteflies are being answered with the aid of artificial diets. Biochemical pathways for carbohydrate metabolism and polyol synthesis have been determined. Metabolism of plant toxins is being studied to assess the ability of <i>Bemisia</i> to detoxify plant deterrent compounds. The role of nitrogen fertilization in whitefly-cotton interactions was determined in field trials.
<b>Develop whitefly artificial diets and natural enemy mass-rearing.</b>	Develop whitefly artificial feeding systems.	X		Development of an artificial feeder for whiteflies that will support development from egg to adults has been successful, and improvements continue to increase the proportion of <i>Bemisia</i> adults produced. This system has been tested for its effectiveness at supporting parasitoid wasp development, and adult <i>Encarsia</i> have been successfully produced in this system. Further research is needed to optimize the system for both whitefly and parasitoid development.

<sup>a</sup> Natural enemy research complements from Section D, see Table D.

<sup>b</sup> Action and economic thresholds also apply in Section C, see Table C.

<sup>c</sup> Sampling technology applicable to all other sections, see Tables B to F.

**Table A. Biology, Ecology, and Population Dynamics.**

Research Approaches	Year 3 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Determine life cycle vulnerabilities (life tables)<sup>a</sup>, population development and natural mortality factors, natural enemies on major crops, urban plantings, weeds and predict overwintering potential.</b>	Identify potential low population manipulation on vital host links for survival	Y		Low survivorship was documented both on cotton during peak season and on citrus during overwintering. Mortality factors were identified and shown to have overlapping influence. Seasonal distribution on various host species was determined.
<b>Develop sampling methodology, action and<sup>b,c</sup> economic thresholds for all major crops. Sampling methods and thresholds modified in light of natural enemy levels and existing management strategies.</b>	Validate and refine sampling methods.		X	
<b>Develop population models to describe and predict whitefly population growth and spatial and temporal distribution. Develop simple day-degree sub-models for estimating phenology and temporal patterns of whitefly, natural enemies and host crops.</b>	Provide model simulation of whitefly populations and multiple cropping systems.		X	
<b>Develop sampling methods for quality of cotton lint, vegetables and other commodities.</b>	Develop sampling protocol for field and harvest and processing sampling and determine interrelationships.	X		Correlations established between thermodeceptor measurements of cotton stickiness and amounts of honeydew on the fiber.
<b>Quantify whitefly and natural enemy dispersals and contribution to population dynamics.</b>	Determine proportion of whitefly population that are migratory and their reproductive potential.	X		Trapping studies showed low dispersal for October to early June and evidence for sudden dispersal activity
<b>Define mating behavior, reproductive isolation, species, biotypes.</b>	Define factors involved in mating, cues, feedback mechanisms, etc.		X	
<b>Validate <i>Bemisia</i> taxa morphology, genetic, biochemical, and biology characteristics.</b>	Discuss results, plan additional research, arrive at a consensus decision.	X		Difference in ecdysteroid titers and waxy particle length, width and morphology documented between <i>Bemisia</i> and other whiteflies.



**Table A. Biology, Ecology, and Population Dynamics. (Continued)**

Research Approaches	Year 3 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Define role of endosymbionts in metabolism, host adaptation, nutrition and survival.</b>	Determine potential for manipulating, interfering with or inhibiting endosymbiont function.		X	
<b>Characterize nutrient uptake and metabolism</b>	Determine the physical and biochemical processes involved in uptake of carbohydrates, amino acids and other essential nutrients.	X		Length of stylet determined and correlated with vascular bundle depth. Basic metabolic rate and minimum carbohydrate content to support this rate were determined. Preliminary analysis of amino acid metabolism was conducted.
<b>Develop whitefly artificial diets and natural enemy mass-rearing.</b>	Conduct addition, deletion studies to identify essential nutritional needs.	X		Optimal conditions of pH and egg load were established. Hatching and survivorship was evaluated on various artificial diets.
<b>**Pursue specific genetic and biological basis for variability in whitefly biotypes, strains, and species; determine impact of different genotypes/phenotypes on whitefly-mediated transmission and on the epidemiology of virus diseases.</b>	Continue with work from previous years. Study impact of biotypes, strains, and species differences in the disease spread, crop damage, and specific control measures to reduce whitefly vector populations. Linkages with biological and chemical control sections.		X	This research approach transferred to section A

<sup>a</sup> Natural enemy research complements from Section D, see Table D.

<sup>b</sup> Action and economic thresholds also apply in Section C, see Table C.

<sup>c</sup> Sampling technology applicable to all other sections, see Tables B to F.

\*\* Transferred from Table B 3/31/2000.

**Table A. Biology, Ecology, and Population Dynamics, 2001-2002.**

Research Approaches	Year 4/5 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Determine life cycle vulnerabilities (life tables)<sup>a</sup>, population development and natural mortality factors, natural enemies on major crops, urban plantings, weeds and predict overwintering potential.</b>	Initiate studies to manipulate host sequences to determine potential influence on whitefly population.	X		Greenhouse studies examined movement and mortality rates of <i>B. tabaci</i> crawlers on cotton. Airflow did not influence crawler survival, which averaged 86%. Crawlers moved as far as 200 mm, and over 99.9% settled on the lower surface of the same leaf where eggs were oviposited. Ongoing studies examined seasonality and mortality patterns of <i>B. tabaci</i> on broccoli, cantaloupe, cotton, alfalfa, <i>Lantana</i> and various weeds in Yuma, Maricopa and Marana, AZ. These sites represented a range of geographic and climatic areas in the state. Cantaloupe and <i>Lantana</i> served as good hosts for <i>B. tabaci</i> population buildups in the fall. In the winter, populations steadily declined. Whiteflies survived the winter best in broccoli and cheeseweed, and shortly thereafter built up in broccoli, cantaloupe and <i>Lantana</i> . Large populations in cantaloupe and <i>Lantana</i> were probably responsible for the initial buildups in cotton. During the summer, ground cherry and other weeds also played a role. Yuma showed the earliest increase in whitefly numbers in the spring, but had lower numbers throughout the season when compared to Maricopa and Marana. Whiteflies survived best on cantaloupe at all sites. Predation and dislodgment accounted for a significant portion of the mortality at all locations. Parasitism was highest in <i>Lantana</i> and at the Maricopa site. In India, age specific mortality factors were investigated for whiteflies reared on potted cassava plants. Adult survivorship depended on time of year. Predation, parasitism and unknown factors accounted for variation in survivorship. Overall survivorship from first instar to adult ranged from 5-55%. Parasitism was the dominant mortality factor from pupal to adult emergence. Natural enemies included aphelinid parasitoids, predatory coccinellids, mites and spiders. Missing and unknown factors were the dominant mortality factor in most experiments, followed by predation and parasitism. In laboratory studies, <i>Serangium paracestosum</i> was the most important predator.
	Continue 4 and finalize analysis of the potential of habitat modification as a management tool.	X		

**Table A. Biology, Ecology, and Population Dynamics, 2001-2002. (Continued)**

Research Approaches	Year 4/5 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Develop sampling methodology, action and<sup>b,c</sup> economic thresholds for all major crops. Sampling methods and thresholds modified in light of natural enemy levels and existing management strategies.</b>	Implement sampling protocols through cooperative extension outlets and other technology transfer methods	X		In Cukurova, Turkey, studies were conducted to determine the most appropriate sampling unit and spatial distribution of immature and adult <i>B. tabaci</i> , and immature parasitoids. Whitefly eggs were most often associated with cotton leaf positions 2-4; nymphs with positions 5-7; pupae with positions 6-10; and immature parasitoids with positions 8-11. Based on Taylor's power law it was most efficient to sample leaf 3 for eggs, leaf 5 for nymphs, leaf 10 for pupae and immature parasitoids, and leaf 4 for adults. The CC trap was modified by coating the trap tops with Tanglefoot. This resulted in a 50% increase in trap catches. The addition of a lime green LED (light-emitting diode) to the trap resulted in a 15-23 fold increase in whiteflies trapped relative to traps with a white LED or no light.
	Finalization, implementation and use in IPM systems.	X		
<b>Develop population models to describe and predict whitefly population growth and spatial and temporal distribution. Develop simple day-degree sub-models for estimating phenology and temporal patterns of whitefly, natural enemies and host crops.</b>	Identify weak points and needed information to improve model simulations.	X		In Iran, an engineer-based computer simulation model was used to predict greenhouse whitefly development. Developmental time from egg to adult was longer and mortality was higher particularly during the egg and second nymphal stages with the computer simulation relative to greenhouse data. Incorporation of additional biotic and abiotic factors into the model may improve predictability.
	Validate and expand effort to provide predictive models capabilities for whitefly population development and crop interfaces..	X		

**Table A. Biology, Ecology, and Population Dynamics, 2001-2002. (Continued)**

Research Approaches	Year 4/5 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Develop sampling methods for quality of cotton lint, vegetables and other commodities.</b>	Extend sampling protocols to textile mill and verify field findings in relation to mill problems.	X		Two assay systems for detecting sticky cotton were investigated, the manual thermodetector (SCT) and the high speed (automated) sticky cotton thermodetector (H2SD). The newest H2SD system consistently detected more thermodetector spots than the SCT and the relationship between the platforms appeared to be nonlinear. Examination of many different sizes of field sample units suggested that smaller sample units were more cost-efficient than larger sample units. Further analyses of variance components suggested that only a single assay for each sample unit should be conducted with the SCT, but that 3 assays should be conducted for each sample unit on the H2SD. Reasonably precise estimates of stickiness can be achieved with a sample size of about 15 on either platform using the 1-plant sample unit. Even fewer samples would be required using a 20-boll unit. Samples sizes as high as 41 or 82 would be required for estimates with very high precision on the H2SD or SCT platform, respectively. Sampling costs are much lower for the H2SD because samples can be assayed faster. Sweet potato whitefly feeding on plants in the field with furrow plus supplementary drip irrigation and in the greenhouse on non-stressed cotton plants produced more micrograms of honeydew sugars per gram of honeydew compared to whiteflies feeding on plants with furrow irrigation alone in the field and on water-stressed plants in the greenhouse. Thermodetector analysis was used to determine the effect of days of exposure to <i>B. tabaci</i> and lint stickiness. The effect of rainfall on lint stickiness was also examined. Trehalulose and melezitose produced by <i>Bemisia</i> and thermodetector counts in cotton lint increased with increasing numbers of days of exposure of open cotton bolls in infested cotton plants. Thermodetector counts were significantly correlated to amounts of trehalulose and melezitose. Rainfall of 0.5 inches reduced trehalulose and melezitose in cotton lint within 5 h. Sweet potato whitefly and cotton aphid honeydew sugars were compared, and their effect on cotton lint stickiness was measured by thermodetector counts. The major sugar components of both insects were glucose, fructose, sucrose, trehalulose, and melezitose. SPW produced more trehalulose relative to melezitose, while the cotton aphid produced more melezitose relative to trehalulose. There was a direct relationship between cotton lint stickiness, as detected by thermodetector counts, and increasing duration of exposure to both insects.
	Modify, refine and complete sticky cotton sampling protocols from the field to the mill..	X		

**Table A. Biology, Ecology, and Population Dynamics, 2001-2002. (Continued)**

Research Approaches	Year 4/5 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Quantify whitefly and natural enemy dispersals and contribution to population dynamics.</b>	Quantify the role of dispersal in population dynamics on different crop systems.	X		The flight behavior of male and female <i>Eretmocerus eremicus</i> in response to skylight and plant cues was examined in a vertical flight chamber. Both male and female parasitoids were capable of flying in excess of 60 min, but averaged 12 min in duration. When plant cues were presented during the parasitoid's phototactic flight, 63% of the wasps responded to the plant cue shortly after takeoff, and most of these wasps were females. Another 37% exhibited a response consistent with migration. They failed to respond to the plant cue until they had flown for an extended time. Most of these wasps were males. A separate study, also found that <i>E. eremicus</i> could fly for an extended period in the vertical flight chamber. Virgin females made the longest flights. This work was validated in field studies where it was found that female flight was wind-aided and directional. Males flew for shorter distances, mostly within the boundary layer
	Formulate theory for manipulating and/or using dispersal as a tool in IPM.	X		
<b>Define mating behavior, reproductive isolation, species, biotypes.</b>	Develop potential methods of utilizing behavioral information in management strategies	X		In India, host-associated variation studies revealed the presence of two strains of <i>Bemisia tabaci</i> – the cassava whitefly and the sweet potato whitefly. Two enzyme systems were used to study variations in the isozyme patterns: Esterase and Malate dehydrogenase. The esterase-banding pattern in sweet potato whitefly differed from the cassava whitefly in having an additional fast moving esterase band (Rf value 0.293). The migration distances of the two slow moving bands, common to both biotypes were also different (0.15 and 0.204 in CWF and 0.108 and 0.165 in SPWF). Banding patterns using the Malate dehydrogenase system was identical in both biotypes. The ability of the cassava biotype to vector Indian Cassava Mosaic Virus was confirmed by ELISA. The sweet potato biotype was unable to transmit ICMV. In crossbreeding studies an increased proportion of male progeny indicated that cassava and sweet potato are two biotypes and are not separate species. Host-associated variation of <i>B. tabaci</i> in terms of development and survival also were examined. Cassava, sweet potato, egg plant, tobacco, and cotton were used for these studies. Whitefly populations from sweet potato could not colonize cassava and whiteflies from cassava could not colonize sweet potatoes .
	Review, summarize and propose additional needed research.	X		

**Table A. Biology, Ecology, and Population Dynamics, 2001-2002. (Continued)**

Research Approaches	Year 4/5 Goals Statement		Progress Achieved		Significance
			Yes	No	
<b>Validate <i>Bemisia</i> taxa morphology, genetic, biochemical, and biology characteristics.</b>	Publish verification of new species or other appropriate taxa -----.	X			<p>A staging system based on body depth (thickness) and maturation of the adult eye was used to track development and identify physiological synchronous 4<sup>th</sup> instar/pharate adult silverleaf and greenhouse whiteflies. An examination of histological sections of last instar SLWF revealed that adult eye and wing development were initiated in Stage 6. Wings were deeply folded by Stages 6 and 7. Ecdysteroid titers peaked at Stages 4-early 6, just before the onset of adult development.</p> <p>Leaf samples were taken from field-grown cotton and cantaloupe, and developmental differences in whitefly length and width were determined. Fourth instar nymphs on cotton were longer and wider than nymphs on cantaloupe leaves. In Mexico, numerous whitefly species have been identified including SPWF, SLWF, the greenhouse whitefly, banded-wing whitefly, woolly whitefly, acacia whitefly and citrus whitefly. Between 1994 surveys and 1996-97 surveys, SPWF has nearly been displaced by SLWF. More than 100 plant species have been identified as hosts for SLWF. The identification and composition of the fatty acids associated with the major lipid classes within <i>B. argentifolii</i> nymphs were determined, and comparisons with the internal lipids of adult whiteflies were made. All lipid classes contained variable distributions of 8 fatty acids: myristic acid, palmitic acid, stearic acid, arachidonic acid, palmitoleic acid, oleic acid, linoleic acid, and linolenic acid. Fourth instar nymphs had 5-10 times the quantities of fatty acids as compared to third instar nymphs and 1-3 times the quantities from adults. The quantity differences of fatty acids between fourth and third instar nymphs related to size and weight differences. The triacylglycerol composition of silverleaf whitefly adults was characterized, and the major fatty acid constituents were identified as oleic acid, palmitic acid, stearic acid, and linoleic acid. This information may be useful in regard to the lipid nutrient rewards for whitefly predators and parasitoids.</p>
<b>Define role of endosymbionts in metabolism, host adaptation, nutrition and survival.</b>	Determine associated enzymes and/or other endosymbionts and whitefly relationships  Summarize and implement findings with suggestion for additional research.	X			<p>Several different types of bacteria were cultured from surface-sterilized <i>Bemisia argentifolii</i> adults and nymphs, including <i>Bacillus</i> spp. <i>Enterobacter cloacae</i>, was found within the gut cells of adult whiteflies and was mildly pathogenic.</p>

**Table A. Biology, Ecology, and Population Dynamics, 2001-2002. (Continued)**

Research Approaches	Year 4/5 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Characterize nutrient uptake and metabolism</b>	Determine the potential for blocking key steps in nutrient uptake and/or metabolism	X		It was found that whitefly nymphs feed during all four stages of development. The rostrum of the nymphs is short and has sensilla on the tip, which are similar to those found on the labium of the adult. Stylets range from 110 $\mu\text{m}$ in first instar to over 200 $\mu\text{m}$ in the fourth instar. Prior to the onset of feeding the crawlers stylet is folded within the body of the nymph. Like the adult whitefly, the nymph produces a flange like material that appears to cement the tip of the rostrum to the surface of the host leaf. Prior to each molt, the stylets are withdrawn into the body of the nymph and left behind in the cast exuviae. Most adult whitefly probe sites occurred along the margins of the abaxial epidermal cells. Stylets passed directly through the cytoplasm of the epidermal cell and not through the common radial wall between epidermal cells. Stylets averaged 217 $\mu\text{m}$ long. The adult whitefly uses less than 70% of the length of its stylet to reach a phloem bundle in cotton.
	Implement findings by developing inhibitors of nutrient uptake and/or metabolism.		X	
<b>Develop whitefly artificial diets and natural enemy mass-rearing.</b>	Evaluate developed diets on whitefly fecundity/longevity biology, behavioral characteristics.	X		A newly developed artificial rearing system to determine how biotic and abiotic factors influenced egg hatch, crawler establishment and development of SLWF. Egg age significantly influenced hatch rates, and to a lesser extent survival and development of nymphs reared on the artificial diet. There were negative associations between the number of eggs placed on the membranes and both hatch rate and establishment of crawlers. Eggs oviposited on and then subsequently removed from plants held under long-day conditions (14:10 L:D) or high light intensity had higher hatch rates than eggs oviposited under short-day conditions or low light intensity. Long-day conditions during oviposition also significantly enhanced survival of nymphs through Day 20 and developmental rate for Day 6 counts. Light intensity, at least for the range tested, did not significantly affect development or survival of whitefly nymphs. Recent studies indicated that no matter which host instar <i>E. mundus</i> eggs are laid, penetration into the host proceeds only during the host's fourth instar, typically sometime prior to the transition to adult characteristics. Since approximately three days are required for parasitoid egg development, oviposition under a second or third instar whitefly will result in hatch occurring under a third or young fourth instar host. Epidermal capsule formation accompanied parasitoid penetration. The capsule surrounds/engulfs the parasitoid and must contribute to parasitoid development. The question as to whether host ecdysteroid levels are altered upon parasitized whitefly into which a first instar parasitoid had just begun to penetrate than in a healthy whitefly of the same developmental stage.
	Develop whitefly rearing system and adapt for production of natural enemies.		X	

**Table A. Biology, Ecology, and Population Dynamics, 2001-2002. (Continued)**

Research Approaches	Year 4/5 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>**Pursue specific genetic and biological basis for variability in whitefly biotypes, strains, and species; determine impact of different genotypes/phenotypes on whitefly-mediated transmission and on the epidemiology of virus diseases.</b>	<p>Identify potential factors related to specific genetic and biological variability that may be manipulated to reduce disease spread.</p> <p>Develop molecular approaches to track biotypes, strains, and species relative to disease spread, based on differential molecular markers.</p> <p>Summarize results, identify new research needs and make recommendations for implementation or expansion of research</p>	X		A cell line from the silverleaf whitefly, BtB-2.97-Hunter & Polston was tested for its ability to support replication of the insect iridescent virus 6 (IIV-6). Several lines of evidence indicated that a productive infection was achieved. The cells displayed cytopathic effects, virus particles accumulated in defined areas of the cytoplasm, the cell-associated virus titer was detected at three orders of magnitude higher than that released into the media, and western blot analysis indicated CIV structural proteins were being expressed. Virus was also detected in nymphs by PCR and electron microscopy, but the infections were not highly pathogenic. Infection by whiteflies by IIV-6 suggests that more pathogenic viral isolates may be found for this insect and that the virus has potential to be used as an intracellular probe.

<sup>a</sup> Natural enemy research complements from Section D, see Table D.

<sup>b</sup> Action and economic thresholds also apply in Section C, see Table C.

<sup>c</sup> Sampling technology applicable to all other sections, see Tables B to F.

\*\* Transferred from Table B 3/31/2000



## Reports of Research Progress

### Section B: Viruses, Epidemiology, and Virus-Vector Interactions

Co-Chairs: Bob Gilbertson and Judy Brown

**Investigator's Name(s):** J.K. Brown<sup>1</sup>, A.M. Idris<sup>1</sup>, and J. Bird<sup>2</sup>

**Affiliation & Location:** <sup>1</sup>Univ of Arizona, Plant Sciences Dept; Tucson AZ 85721/<sup>2</sup>Univ of Puerto Rico, Rio Piedras, PR 00928

**Research & Implementation Area:** Section B: Viruses, Epidemiology, and Virus-Vector Interactions.

**Dates Covered by the Report:** 2001

#### **Cloning and Molecular Characterization of the A and B Components for Macroptilium Mosaic Virus from Puerto Rico.**

Macroptilium mosaic virus (MaMV) is a begomovirus that infects common bean and *Macroptilium lathyroides* in Puerto Rico. Leaf samples were collected from *M. lathyroides* plants exhibiting typical bright yellow mosaic symptoms. Total nucleic acids were extracted and aliquots were incubated with selected restriction enzymes. Digestion products were analyzed by Southern hybridization using PCR-amplified of MaMV A and B component-specific probes. As Cla I digestion linearized dsDNA viral forms, it was used to clone A and B viral components. Recombinant plasmids containing full-length viral inserts were selected and their identity was confirmed by Cla I digestion. DNA sequence comparisons revealed that MaMV shared 66.3 and 59.8 % nt sequence identity with the A and B components, respectively, of its closest relative, *Bean golden mosaic virus PR*. MaMV is a new bean-infecting begomoviral species, and only the second begomovirus identified in the BGMV-Caribbean cluster to date.

**Investigator's Name(s):** J.K. Brown and A.M. Idris

**Affiliation & Location:** Univ of Arizona, Plant Sciences Dept; Tucson AZ 85721

**Research & Implementation Area:** Section B: Viruses, Epidemiology, and Virus-Vector Interactions

**Dates Covered by the Report:** 2001

**Cloning and Sequencing of *Cotton Leaf Crumple Virus* –a Begomovirus Infecting Cotton in the Sonoran Desert**

Cotton leaf crumple virus (CLCrV) is a bipartite, whitefly-transmitted geminivirus from the southwestern US and Sonora, Mexico that has been known to infect cotton since the 1950's. The CLCrV DNA A and DNA B components for isolates from Arizona and Sonora were cloned and the nucleotide sequence was determined. Sequence comparisons indicated that the DNA A component (GB Accession AF480940) shared the highest nucleotide sequence identities with members of the SLCV group, while the closest relatives for the CLCrV DNA-B component (GB Accession AF480941) were begomoviruses from the Caribbean, Central America, and Mexico. Parsimony and maximum likelihood analyses indicated that CLCrV is the first member of a previously undiscovered begomovirus group from the New World. The Rep binding element within the common region of CLCrV, GGAGT-CT-GGAGT, is 100% conserved for both DNA-A and DNA-B components, indicating they are cognate for the same virus. Lack of phylogenetic congruence between CLCrV DNA A and DNA B indicate that they have evolved along different paths, and/or that recombination may have occurred.

**Investigator's Name(s):** C.L. McKenzie, Robert G. Shatters, Jr., and R.T. Mayer.

**Affiliation & Location:** USDA-ARS, Horticultural Research Lab, Ft. Pierce, FL

**Research & Implementation Area:** Section B: Viruses, Epidemiology, and Virus-Vector Interactions

**Dates Covered by the Report:** 2000

**Differential Plant Response and Whitefly Performance in Comparisons of Whitefly and Whitefly-Tomato Mottle Virus (ToMoV) Complex Challenged Tomato.**

Whitefly challenges of tomato are known to induce the accumulation of a class of proteins termed pathogenesis related or PR-proteins. It is also known that in general different types of plant challenges can induce different sets of PR-proteins. The present work was to determine how activity and buildup of a group of PR-proteins responded to an insect (whitefly) versus an insect-virus complex (whitefly-ToMoV) challenge.

Tomato PR-proteins (chitinase,  $\beta$ -1,3-glucanase, peroxidase, P2 and P4) responses were measured over time in plants divided into three treatments: untreated controls, plants infested with healthy whiteflies, and plants infested with ToMoV carrying whiteflies. Six- to seven-leaf plants were infested with ~5 adult whitefly per leaf. Subsequent plant samples were pulled prior to whitefly infestation and at 14, 28, 42, and 56 days post-infestation for protein and enzyme analyses. By 56 days, there were 2.5- and 4.5-fold more whitefly eggs and nymphs respectively on the plants infested with viruliferous whiteflies than on the healthy whitefly infested plants. A significant increase in the enzymatic activity of all measured PR-proteins, as compared to control plants, was only seen in viruliferous whitefly infested plants. There was no significant difference observed in enzyme activities between uninfested and healthy whitefly infested plants; however, a trend for healthy whitefly induced increases in PR-protein activities was observed. Foliar protein (protein g/leaf (mg)) in tomato did not differ significantly among treatments for any of the sample dates indicating that any significant changes in enzyme levels was a result of differences in the specific measured enzymes. At 56 days post-infestation, virus infected plants exhibited severe virus symptoms and PR enzyme activities declined. The greatest differences for all PR-proteins assayed was observed 42 days after treatment initiation. Western blot analyses showed that the differences in PR-protein activities among the treatments were due to changes in specific enzyme levels within the plant and were associated with concomitant increases in levels of P2 and P4 PR-proteins; however, the level of viral induction of PRs was variable among virus infected plants. Under our experimental conditions, it is clear that the whitefly-ToMoV complex is a much stronger inducer of tomato PR-proteins response than whiteflies alone. Because of the increased egg and nymph production by the viruliferous whiteflies, the cause of the increased PR-protein response when the virus is present may be the result of either increased feeding pressure of viruliferous whiteflies or direct interactions of virus and plant.

**Investigator's Name(s):** C.L. McKenzie

**Affiliation & Location:** USDA-ARS, Horticultural Research Lab, 2001 South Rock Road, Ft. Pierce, FL

**Research & Implementation Area:** Section B: Viruses, Epidemiology, and Virus-Vector Interactions

**Dates Covered by the Report:** 2000-2001

### **Tomato Mottle Virus (ToMoV) Effects Whitefly Oviposition and Adult Survivorship on Healthy Tomato**

The effect of plant viruses on the reproductive potential of the vector is key to understanding geminivirus epidemiology and developing effective control measures. The objective of this study was to determine the effect of tomato mottle virus (ToMoV) on whitefly oviposition and survival rates on healthy tomato.

Adult *B. tabaci* biotype B were obtained from laboratory colonies maintained by the U.S. Horticultural Research Laboratory, Ft. Pierce, FL. Whiteflies used in these experiments were originally obtained from Dr. Lance Osborne, University of Florida, Apopka, FL and have been maintained on dwarf cherry tomato (*Lycopersicon esculentum* cv. Florida Lanai) since 1996 by serial transfer. In 1997, a ToMoV whitefly colony was established by obtaining tomato plants infected with ToMoV from Dr. Philip Stansly, University of Florida, Immokalee, FL and infesting with whiteflies from the healthy colony. Whitefly biotyping was based on RAPD PCR analysis using primers developed by De Barro and Driver (1997). Nonviruliferous and viruliferous whitefly colonies were housed separately in screened Plexiglass cages located in separate growth chambers at  $25 \pm 1^\circ\text{C}$  and a 16:8 L:D photoperiod. Whiteflies from the viruliferous colony were confirmed to be infected with ToMoV prior to infestation by PCR analysis.

In each experiment, one male and one female whitefly of unknown age from healthy or ToMoV-infected whitefly colonies were confined in clip cages attached to the terminal leaf of the 3<sup>rd</sup> fully expanded leaflet of a healthy cv. Florida Lanai plant. Clip cages were made from clear plastic cups (PC100 30 ml cups, Jet Plastica, Harrisburg, PA) fitted with a foam seal on the bottom and an organdy window on top for ventilation. The foam bottom was backed with a thin square of balsa wood, and an aluminum hair clip was glued to the balsa wood and cup portion. Whiteflies were introduced through a small hole in the side of the cup. After a 48-hr access period, adult whiteflies were removed and eggs were counted. Treatments were maintained separately at  $25 \pm 1^\circ\text{C}$  and a photoperiod of 16:8 L:D. For each experiment, 20 test plants were typically used for each treatment; however, the final number of replicates (=clip cages) per treatment varied when leaves of test plants died or were severed during the experiment. A minimum of 8 replicates was used for all treatments. Experiments were repeated five times. There were no significant interactions between experiment\*treatment ( $F=2.0$ ;  $df=4,80$ ;  $P=0.10$ ) or treatment\*clip cage ( $F=0.66$ ;  $df=19,80$ ;  $P=0.85$ ) so results were pooled over experiments for mean comparison. In the last two experiments, progeny from cohorts used in the oviposition clip cage experiments ( $n = 15$ ) were used to include survival to adult emergence which was evaluated 30 days after egg lay to ensure that all viable whiteflies had emerged.

Whiteflies infected with ToMoV deposited significantly more eggs ( $F=19.51$ ;  $df=1,80$ ;  $P < 0.01$ ) on healthy tomato leaves than nonviruliferous whiteflies (~40%). There was no significant difference between virus-infected and nonviruliferous whiteflies for the number of adults emerged or the proportion of those adults surviving from the egg stage. There was no significant correlation between the number of eggs deposited per female and progeny survival rates on healthy tomato for whitefly infected with or without the virus.

In our experiments, whiteflies were well adapted to the host plant, either with or without ToMoV. High survival of both healthy and ToMoV-infected whitefly reflect this host-plant adaptation. Plants from the virus treatment exhibited characteristic ToMoV symptoms 30 days after clip cages were removed. This suggests adaptation to the host plant and virus by the vector could override any adverse effect the virus had on host plant physiology. Insect adaptation to the host-plant is a critical factor that should be considered on a host-by-host basis when evaluating insect biology and vector-host-plant interactions for polyphagous insect species.

**Investigator's Names:** R.T. McMillan, Jr., M. J. Davis, and Z. Ying

**Affiliation & Location:** University of Florida, TREC, Homestead, FL

**Research & Implementation Area:** Section B : Viruses, Epidemiology, and Virus-Vector Interactions

**Dates Covered by the Report:** 1997-2001

### **Tomato yellow leaf curl geminivirus Management in South Florida**

Tomato yellow leaf curl geminivirus (TYLCV) was identified in the United States for the first time in late July 1997 in a field planting at a commercial breeding facility in Florida. Shortly thereafter, infected tomato plants were also found in several retail garden outlets in Florida (personal communication J. Polston). The source of these plants was traced back to two commercial nurseries near Homestead in south Florida that had shipped tomato plants to retail outlets in Florida and other states and out of the country. Subsequently, infected tomatoes from retail garden outlets have been found in Virginia (confirmed by laboratory tests) and possibly other locations in the southern United States supporting the possibility that TYLCV will become a regional problem.

Due, apparently, to the introduction and spread of the silver leaf whitefly, there has been a recent emergence of seventeen whitefly-transmitted geminiviruses in tomato in the Western Hemisphere. Of these, only tomato mottle virus was present in Florida before TYLCV. We used polymerase chain reaction (PCR) and DNA sequence analyses to confirm the recent introduction of TYLCV into Florida. Degenerate primers for geminiviruses were used to amplify a fragment of the viral genome containing part of the coat protein gene. The partial gene sequence had greater than 98% homology with that of an Israel strain of the TYLCV. These results suggest that the virus is an Eastern Mediterranean strain and possibly the same strain previously introduced into the Dominican Republic from Israel in 1991 and subsequently found in Jamaica and Cuba. We have designed non-degenerate primers for PCR detection of TYLCV based on our DNA sequence data for the virus. Detection of TYLCV in tomato plants with these primers has been 10-100 times more sensitive than with the degenerate primers, which even with the degeneracy have mismatched bases.

Tomatoes are a winter crop in south Florida, and TYLCV was detected in October, 1997, in newly established commercial plantings. Tomatoes are still being planted in south Florida at the present time. The incidence of TYLCV-infected plants initially appeared to be due mostly to primary spread into the fields. Sources of inoculum are presumed to be weeds outside of the tomato fields.

Several studies have been conducted at TREC to manage SLW in tomatoes using insecticides of various classes. Most of these studies are yet to be completed. Several generalist predators (minute pirate bug, lady bug, lacewing) were evaluated in the laboratory for control of SLW, but none of them provided satisfactory control. Compatibility of these predators with various chemical and biological insecticides were also determined. All foliar sprays significantly (90-100%) affected survival of these predators. Soil drench of imidacloprid did not cause any mortality of SLW predators. Various fungal insecticides were also evaluated in the laboratory and greenhouse situations to control SLW. A commercial formulation of *Beauveria bassiana* (Mycotrol®) was consistently more effective than *Paecilomyces fumosoroseus* (PFR-97®). The performance of *Acromonium* and *Verticillium lecanii* was inconsistent. Compatibility of these fungal insecticides with fungicides was also determined. Control of SLW with both *B. bassiana* and imidacloprid offers promising possibilities and needs to be evaluated for management of TYLCV.

Whitefly population densities have been generally low, and even less in tomatoes due to the extensive application of the systemic insecticide, imidacloprid, to transplants. However, the occurrence and incidence of TYLCV in cultivated tomatoes have steadily increased during the growing season. Secondary spread within fields has become more prevalent and whitefly population densities are on the increase. TYLCV appears to be firmly established in Florida, and will continue to be a major problem in the region in years to come.

Florida Agricultural Experiment Station, Journal Series No. N-02188.

**Investigator's Name(s):** Thomas M. Perring and Charles A. Farrar

**Affiliation & Location:** Department of Entomology, University of California, Riverside, CA. 92521

**Research & Implementation Area:** Section B: Viruses, Epidemiology and Virus-Vector Interactions

**Dates Covered by the Report:** January 1, 2001 - December 31, 2001

**Whitefly-borne Virus Epidemiology: Lessons Learned from the Greenhouse Whitefly and Tomato Infectious Chlorosis virus**

We sampled 57 weed species and 13 crop species in Orange County, California in order to describe the host range and seasonality of greenhouse whitefly (GHWF). This survey identified 21 weed hosts and 9 crop hosts of GHWF. The agricultural areas in the survey region are farmed intensively; most land carries 2 to 3 crops per year. Strawberries are planted in September and October and harvested from the following winter into summer. Zucchini squash and cucumbers are grown year-round; young plants are grown under plastic tunnels in late winter, while old plantings are harvested into early and mid-winter as long as the plants survive. Beans generally are present except in midwinter. Tomatoes are planted from winter, in tunnels until summer, and are harvested from spring into December. Thus crop plants are available throughout the year as GHWF hosts.

The most consistent aspect of GHWF infestation of these crops was that most fields surveyed had very few whiteflies. Very few fields that had low numbers of GHWF were found to be associated with a GHWF source, while fields with a maximum GHWF count of greater than 100 were, in almost every case, near a source of GHWF. Closer examination of fields typically revealed that there was a sharp decrease in GHWF numbers with increased distance from the whitefly source. GHWF outbreaks over the past several years were not regional but local in nature. Growers can minimize their own GHWF infestations by distancing new plantings from sources of infestation, by interrupting serial plantings which perpetuate infestations, and by eliminating infested crop residues as soon as possible after harvest. In our survey we found that some fields separated by as little as 0.1 mile from GHWF sources escaped even moderate infestation.

The ability to control GHWF levels is most important for tomato growers, because those numbers affect tomato infectious virus (TICV) infection. In epidemiological studies, we found that disease levels were low and seldom occurred before the onset of harvest. Tomato fields categorized according to their maximum GHWF levels revealed that few lightly infested fields became infected with TICV, while over half of the fields with high GHWF numbers were found to have virus. Additional data from individual sites supported the connection between GHWF numbers and rates of TICV infection, particularly when the whitefly source was tomato. The presence and proximity of GHWF sources strongly influenced subsequent infestations and local TICV epidemics.

**Investigator's Name(s):** Yash Pal S. Rathi

**Affiliation & Location:** Centre of Advanced Studies in Plant Pathology, G.B. Pant University of Agri. & Tech. PANTNAGAR-263145 (India)

**Research & Implementation Area :** Section B: Viruses, Epidemiology, and Virus-Vector Interactions

**Dates Covered by the Report :** 1998-1999, 1999-2000

### **Frenchbean Crinkle Stunt: Epidemiology and Management**

Crinkle stunt, a recently identified disease has become a potential threat to the cultivation of frenchbean (*Phaseolus vulgaris* L.) particularly in the *Tarai* and Hill regions of Uttaranchal and Uttar Pradesh states of India. The disease incidence was higher in Rabi as compared to spring planting season. The diseased plants remain stunted showing characteristic symptoms of leaf rolling, crinkling, rugosity and vein chlorosis with dark green colour of the leaf lamina. Diseased plants show complete sterility or bear a very few pods. Yield losses may go even up to 100 per cent in case of a susceptible genotype if infected at an early stage of crop growth. Transmission studies showed that the frenchbean crinkle stunt virus (FbCSV) was not seed, soil, sap or dodder transmissible. It is vectored only by whitefly (*Bemisia tabaci* Genn.). For host range studies, 87 varieties of 50 host species belonging to 10 families, were inoculated with viruliferous whiteflies, however, the virus remained restricted only to frenchbean.

The virus (FbCSV)-vector (*Bemisia tabaci*) relationship studies were conducted in glasshouse. The minimum acquisition and inoculation feeding periods were 6 and 4 hrs, respectively. Three hrs starvation before acquisition and inoculation accesses increased the transmission efficiency of the whitefly adults. An incubation period of 12 hrs was observed in the vector. Singly whitefly adult was able to transmit the virus. However, increased number of viruliferous whitefly adults per test plant, increased percentage of transmission. The viruliferous adults were infective for a maximum period of 7 days indicating that virus is the persistent (circulative). Female whitefly adults were more efficient vectors as compared to male adults.

Delayed planting (Oct- 30), narrow plant spacing (30 x 10 cm) & increased levels of N alone or in combination with P not only significantly decreased the disease incidence but also increased the yield. Two insecticidal (Monocrotophos 0.1% + Endosulfan 0.1%) sprays at 10 day interval decreased the population of whiteflies considerably. Out of 50 germplasm lines 15 were found resistant after artificial screening in glasshouse. Since none of the individual control measure is effective, an integrated management strategy has to be developed.

**Investigator's Name(s):** Y.P.S. Rathi

**Affiliation & Location:** Centre of Advanced Studies in Plant Pathology, G. B. Pant University of Agriculture & Technology, Pantnagar-263145 (India)

**Research & Implementation Area:** Section B: Viruses, Epidemiology, and Virus-Vector Interactions

**Dates Covered by the Report:** Last 4 *Kharif* seasons

### **Mungbean Yellow Mosaic Virus: Epidemiology and Management**

Mungbean yellow mosaic virus (MYMV) also called as a "yellow plague" of *Kharif* pulses is vectored by whitefly (*Bemisia tabaci* Genn.) in a persistent circulative manner. Currently, it is a number one problem causing substantial yield losses in urdbean (*Vigna mungo*), mungbean (*V. radiata*), mothbean (*V. aconitifolia*) and soybean (*Glycine max*) in India. The host crop exhibits general yellowing and the diseased fields can be recognized from far distance.

Studies on epidemiology in *Tarai* region of Uttaranchal State revealed that besides the population of whiteflies, weather parameters like temperature, rainfall and humidity play important role in the development of the disease epidemic. Whitefly population increased with increase in temperature. High relative humidity, heavy showers and strong winds in rainy season were found detrimental to whitefly adults. Further studies on host range revealed that ratoon crop of pigeonpea and/or some other weeds might be serving as primary source of inoculum.

Experiments were conducted at Crop Research Centre of Pantnagar University to contain this disease through agronomic practices (manipulation in planting dates and inter-cropping with barrier (non-host) crops, insecticides (foliar and soil application, seed treatment) and host resistance. Low disease incidence was recorded in early (June) and late (August) plantings as well as in low plant spacing (5 cm). Seed dressing with carbofuran 3-G or phorate 10-G with two foliar sprays of monocrotophos (0.1%) + endosulphan (0.1%) also reduced the disease incidence. Spray of 2% mineral oil + 0.5% Neem oil + 5% detergent + 0.1% monocrotophos was equally effective. Inter-cropping of non host crops with mungbean, soybean and urdbean were not found very effective barriers. Some genotypes viz., Pant U 19, 30, 35 and PU-1 GD (urdbean), PM 1, 2, 3 & 4 (mungbean) and PK 416, PK 564, PK 1029, SL 142 (Soybean) resistant to MYMV have been identified.

None of the individual management approach was found highly effective against the vector/disease due to high vector population and wide range of host plants serving as initial foci of the whitefly vector and the virus through out the year. Moreover a single whitefly adult is sufficient to initiate the infection. The only promising way of effective management of this yellow plague (MYMV) appears to be the development of integrated management strategy having tolerant varieties in core.



**Investigator's Name(s):** Satya Vir

**Affiliation & Location:** Central Arid Zone Research Institute, Jodhpur-342003, India

**Research & Implementation Area:** Section B: Viruses, Epidemiology, and Virus-Vector Interactions

**Dates Covered By The Report:** January 2000 - December 2000

### **Integrated Management of *Bemisia tabaci* Genn. and Yellow Mosaic Virus in Arid Legumes**

Mothbean, Mungbean and cowpea (*Vigna aconitifolia*, *V. unguiculata* and *V. radiata*) are the important legumes commonly grown in the arid region of India. But the productivity of these crops is extremely poor (125-250 kg /ha). The incidence of Whitefly, *Bemisia tabaci* Genn. and the heavy incidence of yellow mosaic virus (YMV) are the major factors affecting the yield of these important arid legumes. Field studies conducted at Central Arid Zone Research Institute, Jodhpur have revealed that both whitefly and YMV are responsible for the yield loss of 15.23 to 73.15 percent in different cultivars of these crops. Four hundred lines of these crops were screened for their susceptibility to the incidence of whitefly and YMV. Seven cultivars of mothbean viz. IPCMO-943, IPCMO-1035, T-16, T-2, Jadia, PLMO-240 and PLMO-216 and four of cowpea i.e. JC5, JC10, HFG42-1 and FS-68 and two cultivars of mungbean V7 and V8 were isolated to be least susceptible to the attack of whitefly and YMV. Recommendation of early sowing preferably in the 1<sup>st</sup> or 2<sup>nd</sup> week of July and cultivation of above listed seed material is most profitable in these arid legumes. In the early sowing of these cultivars, the loss due to whitefly and YMV is minimised (14%) as compared to the late sown local cultivars where the loss can extend up to 35 to 54 percent.

Neem oil, quality Neem seed preparations and the commercial available Neem products were tested both under laboratory and field conditions against whitefly and YMV. The quality Neem seeds selected and processed were from the candidate plus trees which contain 48.23 percent oil and 0.892 percent Azadirachtin. These plant extracts were tested either alone or as mixture with a synthetic insecticide (monocrotophos) for the efficacy in the control of whitefly and YMV. Whitefly population per plant varied significantly in different treatments. Subsequently, there was significant reduction in YMV in all the three arid legumes. Use of monocrotophos was superior to all treatments followed by the treatment of Neem oil and Neem seed extracts. The use of Neem oil was more effective when used as mixture with the synthetic insecticide than when applied alone. The treatment of Neem oil mixture with synthetic insecticide gave the highest yield of 875 kg /ha and was at par with Neem oil treatment (846kg/ha) followed by NSKE with monocrotophos (823 kg/ha) in mungbean crop. The data reported also involve the utilization of different management strategies in a compatible manner and designed to suppress the pest population and disease in these arid legumes. The use of Neem products either alone or in combination with synthetic insecticide will help to minimise the use of insecticide and conserve the beneficial organisms. This will also help to maintain the ecological situations with long-term benefits.

**Investigator's Name(s):** William M. Wintermantel and Arturo A. Cortez

**Affiliation & Location:** USDA-ARS, Salinas, CA

**Research & Implementation Area:** Section B: Viruses, Epidemiology, and Virus Vector Interactions

**Dates Covered by Report:** 2001

**Complementation for Transmission by Non-vector Whiteflies  
among Tomato-infecting Criniviruses**

*Tomato chlorosis crinivirus* (ToCV) and *Tomato infectious chlorosis crinivirus* (TICV) appear to have largely distinct geographical distributions, but have been found together in field-grown tomato. TICV is transmitted only by *Trialeurodes vaporariorum*, while ToCV is transmitted by *T. vaporariorum*, *T. abutilonea* and *Bemisia* species. Both viruses have similar genome size and organization, suggesting the potential exists for transmission by non-vector whiteflies from mixed infections. We established *Physalis wrightii* source plants, containing either TICV alone, ToCV alone, or both viruses together, confirmed by northern blot to virus specific probes. *T. vaporariorum* and *T. abutilonea* were allowed to feed separately on all virus sources, as well as virus-free plants for 24 hours, then were transferred to young host plants. Symptomatic plants were tested by northern blots as before, and transmission of TICV by *T. abutilonea* from mixed infection was confirmed in two of sixty plants. Separate experiments in which *T. vaporariorum*, *T. abutilonea*, and *B. argentifolii* were allowed to feed initially on ToCV source plants, followed by TICV source plants, then transferred to test plants did not result in transmission of TICV by non-vector whiteflies. Although cross-transmission does not appear to occur frequently, this rate is substantial considering the high whitefly populations that can occur in the field. Cross-transmission also suggests that genetic compatibility between these viruses may be possible, and could lead to evolution of TICV strains with expanded vector transmissibility.

## Section B. Summary

Judy Brown and Bob Gilbertson

### Begomoviruses.

**Cotton leaf crumple virus (CLCrV).** CLCrV is a bipartite, whitefly-transmitted geminivirus from the southwestern US and Sonora, Mexico that has been known to infect cotton since the 1950's. The CLCrV DNA A and DNA B components for isolates from Arizona and Sonora were cloned and the nucleotide sequence was determined. Sequence comparisons indicated that the DNA A component (GB Accession AF480940) shared the highest nucleotide sequence identities with members of the SLCV group, while the closest relatives for the CLCrV DNA-B component (GB Accession AF480941) were begomoviruses from the Caribbean, Central America, and Mexico. Parsimony and maximum likelihood analyses indicated that CLCrV is the first member of a previously undiscovered begomovirus group from the New World. The Rep binding element within the common region of CLCrV, GGAGT-CT-GGAGT, is 100% conserved for both DNA-A and DNA-B components, indicating they are cognate for the same virus. Lack of phylogenetic congruence between CLCrV DNA A and DNA B indicate that they have evolved along different paths, and/or that recombination may have occurred.

**Crinkle stunt disease (CLD).** CLD is a new whitefly-transmitted viral disease of frenchbean (*Phaseolus vulgaris* L.) in the Tarai and Hill regions of Uttaranchal and Uttar Pradesh states, India. Diseased plants are stunted and exhibit leaf rolling, crinkling, veinal chlorosis, and a dark green leaf lamina, and yields are severely reduced. Transmission studies showed that the Frenchbean crinkle stunt virus (FbCSV) is vectored by whitefly (*Bemisia tabaci* Genn.) and is not seedborne. Host range studies revealed that of 50 species in 10 families examined, only frenchbean is a virus host. The minimum acquisition and inoculation feeding periods were determined as 6 and 4 hrs, respectively. A 12 hr latent period was observed and single whiteflies transmitted the virus. Viruliferous adults transmitted virus for 7 days. Females were more efficient than males as vectors. Delayed planting, narrow spacing (30 x 10 cm, and increased N applications, alone or in with P, decreased disease incidence and increased yields. Two insecticidal (Monocrotophos 0.1% + Endosulfan 0.1%) treatments at 10 day intervals reduced whitefly populations. Fifteen of 50 germplasm lines examined under glasshouse conditions exhibited virus resistance.

**Macroptilium mosaic virus (MaMV).** MaMV is a begomovirus that infects common bean and *Macroptilium lathyroides* in Puerto Rico. Total nucleic acids were extracted and aliquots were incubated with selected

restriction enzymes. Digestion products analyzed by Southern hybridization using PCR-amplified of MaMV A and B component-specific probes indicated Cla I digestion linearized dsDNA viral forms, thus it was used to clone A and B viral components. Recombinant plasmids containing full-length viral inserts were selected and their identity was confirmed by Cla I digestion. DNA sequence comparisons revealed that MaMV shared 66.3 and 59.8% nt sequence identity with the A and B components, respectively, of its closest relative, *Bean golden mosaic virus PR*. MaMV is a new bean-infecting begomovirus species, and only the second begomovirus identified in the BGMV-Caribbean cluster to date.

**Mungbean yellow mosaic virus (MYMV).** MBYMV causes significant loss of urdbean (*Vigna mungo*), mungbean (*V. radiata*), mothbean (*V. aconitifolia*) and soybean (*Glycine max*) crops in India. Studies on disease epidemiology in Tarai region of Uttaranchal State revealed that whitefly pressures and weather parameters (temperature, rainfall and humidity) influence epidemic development. Agronomic practices to reduce disease pressures were examined, including manipulation of planting dates, inter-cropping with barrier (non-host) crops, insecticide treatments (foliar and soil application, seed treatment), and planting of disease resistant varieties. Disease incidence was low in early (June) and late (August) plantings and in low plant spacing (5 cm). Seed dressing with carbofuran 3-G or phorate 10-G with monocrotophos (0.1%) + endosulphan (0.1%) treatment also reduced disease incidence. A treatment of 2% mineral oil + 0.5% Neem oil + 5% detergent + 0.1% monocrotophos was as effective. Non-host crops intercropped with mungbean, soybean, and urdbean were not effective virus or vector barriers. No management approach alone was highly effective against the vector/disease due to high vector population pressures and the availability of a broad range of plants that are virus and/or whitefly vector hosts, indicating that disease management requires an integrated approach that includes resistant varieties.

At the Central Arid Zone Research Institute, Jodhpur, whitefly and yellow mosaic disease (caused by MBYMV) are responsible for 15.23 to 73.15 percent losses in cowpea, mothbean, and mungbean. Four hundred lines were screened for susceptibility to whitefly and YMV. Seven cultivars of mothbean: IPCMO-943, IPCMO-1035, T-16, T-2, Jadia, PLMO-240 and PLMO-216, four of cowpea: JC5, JC10, HFG42-1 and FS-68, and two cultivars of mungbean: V7 and V8 were least susceptible whitefly and YMV pressures. Early sowing (preferably in the 1<sup>st</sup> or 2<sup>nd</sup> week of July) of tolerant germplasm is recommended, based on only 14% losses, compared to the 35-54% losses when local cultivars are planted later in the season. Neem oil, quality Neem seed preparations, and commercially available Neem products were tested under

laboratory and field conditions against whitefly and YMV. The quality Neem seeds selected and processed were from the candidate trees which contained 48.23 % oil and 0.892% Azadirachtin. These extracts were tested alone or as mixture with a synthetic insecticide (monocrotophos) for whitefly and YMV control. Results indicated a significant reduction in virus damage in all three legume crops. Monocrotophos was superior to all treatments, followed by Neem oil, and Neem seed extract. Neem oil was more effective when mixed with the synthetic insecticide than when applied alone, resulting in the highest yields (875 kg /ha), and was at par with Neem oil treatment (846kg/ha), followed by NSKE with monocrotophos (823 kg/ha). Uses of Neem products either alone or in combination with synthetic insecticides will also aid in protection of beneficial organisms.

**Tomato mottle virus (ToMoV).** Experiments were conducted to determine the effect of ToMoV on whitefly oviposition rates and adult survivorship. In 3 of 6 experiments, significantly more eggs were oviposited by viruliferous whiteflies compared to non-viruliferous controls. Although there was no significant difference between treatments for half of the experiments, a trend for higher oviposition by viruliferous whiteflies was observed. Data combined for all dates revealed a significant difference in whitefly oviposition. Adult survivorship was significantly greater for viruliferous whiteflies for one replicate, while no significant difference was seen in two others. When data were combined, the percent eggs developing into adults was the same (~90%), but the number of adults surviving between treatments was significant. Virus assays revealed that not all whiteflies harbored ToMoV, suggesting an explanation for somewhat inconsistent results.

The effect of ToMoV on whitefly oviposition and survival rates on healthy tomato was investigated. Whiteflies exposed to ToMoV deposited more eggs on healthy tomato than did nonviruliferous whiteflies. There was no significant difference between virus-infected and nonviruliferous whiteflies in number of adults emerged or proportion of adult survival. There was no significant correlation between the number of eggs deposited and progeny survival rates on virus-free tomato for viruliferous versus non-viruliferous whitefly. High survival of whiteflies colonizing both virus-free and ToMoV-infected plants reflected adaptation of whiteflies to their host. Virus-infected plants exhibited characteristic ToMoV symptoms 30 days after inoculation, suggesting that host adaptation by the whitefly may counter the otherwise adverse effects that virus infection may have on host physiology. Host-adaptation of the whitefly vector to the host plant is an important consideration to vector-host-plant studies.

A study was carried out to evaluate PR-proteins in plants exposed to whitefly compared to insect-virus complex (whitefly-ToMoV) challenge. Tomato PR-proteins (chitinase,  $\beta$ -1, 3-glucanase, peroxidase, P2 and P4)

responses were measured over time in plants for three treatments: untreated control, whitefly-infested, whitefly exposed to ToMoV infected tomato. At 56 days after infestation, 2.5- and 4.5-fold more whitefly eggs and nymphs, respectively were found on plants infested with viruliferous whiteflies than for the other two treatments, together with a significant increase in enzymatic activity for PR-proteins. Amount of leaf protein (gm/mg) did not differ significantly among treatments indicating that significant changes in enzyme levels were due to differences in specific enzymes. At 56 days post-infestation, virus infected plants exhibited severe disease symptoms, concomitant with a decline in PR enzyme activities. Western blot analyses indicated that differences in PR-protein activities were due to changes in levels of specific enzymes, and were associated with an increase in P2 and P4 PR-proteins. Thus, the whitefly-ToMoV complex more strongly induces PR-proteins in tomato than does whiteflies alone. PR-protein induction may be due to whitefly feeding or to interactions between virus and host plant.

Several generalist predators (minute pirate bug, ladybug, and lacewing) were evaluated in the laboratory for control of SLW to reduce ToMoV incidence, but none of them provided satisfactory control. Compatibility of these predators with various chemical and biological insecticides was examined. Foliar sprays significantly (90-100%) affected survival of predators, whereas, a soil drench of Imidacloprid did not cause predator mortality. Various fungal insecticides were also evaluated in the laboratory and greenhouse situations to control the whitefly vector. A commercial formulation of *Beauveria bassiana* (Mycotrol®) was more effective than *Paecilomyces fumosoroseus* (PFR-97®). Performance of *Acromonium* and *Verticillium lecanii* treatments was inconsistent for both. Whitefly control using *B. bassiana* plus Imidacloprid offered promising possibilities for virus disease management by reducing vector populations.

**Tomato yellow leaf curl virus (TYLCV).** TYLCV was introduced into Florida in 1997 and infects vegetable and nursery crops. The source of these plants has been traced to two commercial nurseries near Homestead in south Florida that shipped tomato plants to retail outlets in Florida, other locations in the US, and as exports abroad. Subsequently, infected tomatoes from retail garden outlets were documented in Virginia and possibly other locations in the southern US, suggesting that TYLCV-IS will become a regional problem. Polymerase chain reaction (PCR) using degenerate primers for a fragment of the coat protein gene (CP) and DNA sequencing of amplicons were employed to identify the specific viral strain or species. The CP sequence for the Florida isolate shared greater than 98% identity with an Israel strain of the TYLCV (TYLCV-IS) and is probably the same as that introduced into the Dominican Republic in 1991, and subsequently in Jamaica and Cuba. Non-degenerate primers for TYLCV were designed for specific detection

of TYLCV-IS by PCR and were 10-100 times more sensitive than degenerate CP primers.

#### **Criniviruses.**

**Tomato infectious chlorosis virus (TICV).** A survey of 57 weed species and 13 crop species was carried out in Orange County, California to identify the host range and seasonal occurrence of the greenhouse whitefly (GHWF). Results indicated 21 weed hosts and 9 crop hosts, including strawberries, squash, tomato, and cucumber, some of which are cultivated year-round. Most fields had very few whiteflies and outbreaks were localized, not regional. Thus, growers can minimize GHWF infestations by distancing new plantings from sources of infestation, interrupting serial plantings, and eliminating infested crop residues after harvest. Some fields separated by as little as 0.1 mile from GHWF sources escaped moderate infestation. Ability to control GHWF is most important for tomato growers, because this whitefly is a vector of TICV in tomato. Disease incidence was low and infection seldom occurred before the onset of tomato harvest. Tomato fields categorized according to their maximum GHWF levels revealed that few lightly infested fields were infected with TICV, while over half of the fields with high GHWF numbers also harbored virus. Thus, presence and proximity of GHWF sources strongly influences local TICV epidemics.

**Tomato chlorosis virus (ToCV) and TICV.** ToCV and TICV have largely distinct geographical distributions, but may occur together in field-grown tomato. TICV is transmitted by *Trialeurodes vaporariorum*, while *T. vaporariorum*, *T. abutilonea* and the B biotype of *Bemisia tabaci* (Genn) transmit ToCV. (syn. *B. argentifolii*). Both viruses infect tomato, suggesting the potential for transmission by a non-vector whitefly when viruses occur in a mixed infection. Experiments were conducted in which *T. vaporariorum*, *T. abutilonea*, and *B. tabaci* B biotype were allowed to feed on ToCV source plants, followed by TICV source plants, prior to transfer to test plants, revealing that TICV was not transmitted by non-vector whiteflies. However, transmission of TICV by *T. abutilonea* from a mixed virus infection was confirmed in two of sixty plants. Although such cross-transmission was infrequent, the frequency could be substantial under high whitefly vector pressures. Cross-transmission also suggests the possibility for genetic compatibility between these viruses that could facilitate evolution of TICV strains that are transmissible by additional vector species.

**Table B. Viruses, Epidemiology and Virus Vector Interactions.**

Research Approaches	Year 1 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Identification and characterization of new or emerging whitefly-transmitted viruses and strains.</b>	Monitor crops for presence of whitefly-transmitted diseases, and determine relative disease incidence. Begin virus identification and strain differentiation.		X	Rapid techniques are available for identification and characterization of geminiviruses through sequencing of PCR-amplified viral DNA fragments. This approach was used to show 98% sequence identity between the tomato yellow leaf curl gemini virus (TYLCV) from the Dominican Republic and an Eastern Mediterranean virus strain indicating that the virus was probably introduced on tomato transplants from the Eastern Mediterranean area. The use of such sequences in comparisons of viruses are important in establishing their relatedness and origin. Several other assays are available for rapid detection of geminiviruses such as dot blot and squash blot hybridization analysis.
<b>Molecular epidemiology: identification of economic viruses, host plants, and reservoirs, and determination of geographic distribution of viruses.</b>	Monitor and identify host plants, virus reservoirs in affected areas. Linkages to diagnostic methods for virus ID and tracking.		X	The use of squash blot analysis using a TYLCV-specific DNA probe to assess the role of weeds as hosts in the Dominican Republic showed that they were not infected with TYLCV and not significant molecular sources for the virus. TYLCV newly discovered in Florida was also 98% identical to the Dominican Republic strain. Geminiviruses, are known throughout the world and distinct viruses are known to occur in many countries. For instance, tomato mottle virus (ToMoV) was first detected in Florida in 1989 and is thought to have originated from that state.

**Table B. Viruses, Epidemiology and Virus Vector Interactions. (Continued)**

Research Approaches	Year 1 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Virus-vector interactions, factors affecting virus transmission, and basis for virus-vector specificity; determination of endosymbiont involvement in whitefly-mediated transmission.</b>	Initiate studies on virus-vector interactions and on basis for the specificity of whitefly-mediated geminivirus transmission.		X	Studies on feeding duration and position has demonstrated differences in aphids and whiteflies. These differences may determine why some geminiviruses are transmitted by one group and not the other. The use of the autofluorescent GFP gene, in tracking the virus movement and replication in plants indicated that a cell to cell movement of the virus occurred and the virus was not phloem limited. Understanding the movement of the virus in terms of insect feeding behavior may play a role in developing resistant varieties.
<b>Strategies to reduce virus spread by management of cropping systems, reduced transmission frequencies, and other potentially effective approaches.</b>	Develop approaches to managing cropping systems to reduce vector densities to decrease transmission frequency and inoculum sources, taking into account weed and crop reservoirs in disease incidence and distribution.		X	Host-free practices used in the Dominican Republic for TYLCV have been successful in reducing the incidence of this disease. In Florida, management of whiteflies with insecticides, field sanitation, and clean transplants has reduced the incidence of ToMoV. In whitefly reduction studies using biological control based IPM, there was a 10% reduction in geminiviruses in squash (See Table D).
<b>Control of virus diseases: development of virus resistant germplasm through conventional and engineered/molecular approaches. Define prospective strategies for selecting candidate viruses, identifying specific virus diseases to target, and prioritize specific crops and cultivars for protection approaches.</b>	Define strategies for resistance efforts. Identify target viruses. Identify germplasm with virus resistance. Initiate efforts toward defining prospective engineered resistance strategies. Identify candidate crops and recipient cultivars.			Resistance to the geminivirus, bean dwarf mosaic virus (BDMV), was found in Pinto bean variety, Othello. Using the GFP gene as a marker, virus infection in this variety was compared with that in a susceptible variety. In the resistant variety, there was a collapse of tissue at the infection site and continuing necrosis in the vascular areas indicating a hypersensitive reaction to the virus. The gene(s) involved in this response may be a source of resistance to this virus either through conventional breeding efforts or by identifying the gene(s) involved. In cotton, some resistance to the cotton leaf crumple virus was reported (See Table F).

**Table B. Viruses, Epidemiology and Virus Vector Interactions. (Continued)**

Research Approaches	Year 1 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Pursue specific genetic and biological basis for variability in whitefly biotypes, strains, and species; determine impact of different genotypes/phenotypes on whitefly-mediated transmission and on the epidemiology of virus diseases.</b>	Identify differences in species, strains and biotypes with respect to transmission, host range, mating compatabilities, molecular variability, and map the biogeographic distribution of distinct types within the <i>B. tabaci</i> species complex.			No reports in this area.



**Table B. Viruses, Epidemiology and Virus Vector Interactions.**

Research Approaches	Year 2 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Identification and characterization of new or emerging whitefly-transmitted viruses and strains.</b>	Virus identification and characterization. Develop methods for identifying causal agents and for tracking viruses and strains using molecular methods.		X	<p>1. Significant progress has been made in the detection and characterization of tomato yellow leaf curl geminivirus in Florida. A comprehensive survey of the incidence and distribution of TYLCV has been made.</p> <p>2. Evidence has been obtained of a synergistic interaction among three geminivirus DNA components associated with chino del tomate disease of tomato (pepper huasteco geminivirus [PHV] DNA-A and DNA-B and another distinct DNA-A component, chino-A). Here, the disease symptoms induced in three hosts (<i>Nicotiana benthamiana</i>, tomato, and pepper) by PHV plus the chino-A are much more severe than symptoms induced by PHV alone. These results establish that (i) chino del tomate disease may be caused by a complex of geminivirus components, (ii) that complexes of geminivirus components can dramatically influence disease symptom expression and (iii) that identification of geminiviruses based on disease symptoms alone is difficult.</p> <p>3. Tomato geminivirus diseases in Guadeloupe are caused, at least in part, by a strain of potato yellow mosaic geminivirus (PYMV).</p>

**Table B. Viruses, Epidemiology and Virus Vector Interactions.**

Research Approaches	Year 2 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Molecular epidemiology: identification of economic viruses, host plants, and reservoirs, and determination of geographic distribution of viruses.</b>	Continue field studies. Determine economic input of diseases on crop production and associated losses.	X		<p>1. The spread of TYLCV in Florida has been extensively documented. The virus has been disseminated throughout the state, including some northern counties. The highest incidences of TYLCV have been correlated with high populations of whiteflies. Extensive host range studies are being conducted with TYLCV in Florida, and TYLCV has been found to infect and cause disease in petunia and common bean. Detection in petunia could have serious implications in terms of exporting this ornamental plant.</p> <p>2. Efforts are being conducted to understand how TYLCV survives in the Dominican Republic during the three-month whitefly host-free period. Using a polymerase chain reaction test to determine the relative contamination of whiteflies with TYLCV, it was found that by the end of the tomato-growing season, TYLCV was readily detected in whiteflies collected from all tomato fields tested. However, within one month of the host-free period, the amount of virus detected in whiteflies collected from plants surrounding tomato fields decreased tremendously. By the end of the host free period, little or no virus could be detected in whiteflies. These results suggest that whiteflies themselves are not likely to be the primary way in which the virus survives during the host-free period. Weeds and other plants in and around fields during the host-free period were then collected and tested for TYLCV using PCR. A number of weeds were found to be symptomless carriers of TYLCV. These results suggest that such symptomless hosts may be the primary way that the virus survives during the host-free period.</p>

**Table B. Viruses, Epidemiology and Virus Vector Interactions. (Continued)**

Research Approaches	Year 2 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Virus-vector interactions, factors affecting virus transmission, and basis for virus-vector specificity; determination of endosymbiont involvement in whitefly-mediated transmission.</b>	Determine specific cellular and molecular factors involved in virus transmission. Study role of endosymbionts in virus acquisition and transmission.		X	Strategies to reduce virus spread by management of cropping systems, reduced transmission frequencies, and other potentially effective approaches. Continue studies of management approaches for disease abatement. Interdisciplinary studies in conjunction with whitefly control methods in Sections B and C.
<b>Strategies to reduce virus spread by management of cropping systems, reduced transmission frequencies, and other potentially effective approaches.</b>	Continue studies of management approaches for disease abatement. interdisciplinary studies in conjunction with whitefly control methods in Sections B and C.	X		In Costa Rica, experiments conducted using living covers (such as coriander and perennial peanuts) and silver plastic mulch demonstrated that these strategies reduced the incidence of geminivirus infection of tomato under moderate whitefly/ geminivirus pressure, but not under high pressure. Thus, living covers and/or silver plastic represent a promising management tool, but one that needs to be used in combination with other practices that lead to reduced inoculum pressure. In the Dominican Republic, the mandatory whitefly host-free period continues to provide an effective management tool for TYLCV. There is a lag period of approximately one-month after planting tomatoes before TYLCV appears and this lag period allows for early-planted tomatoes to provide good yields. This strategy, together with the use of insecticides and tolerant varieties for late season planting, have allowed for the almost complete recovery of the processing tomato industry in the Dominican Republic.

**Table B. Viruses, Epidemiology and Virus Vector Interactions. (Continued)**

Research Approaches	Year 2 Goals Statement	Progress Achieved		Significance
		Yes	No	
<p><b>Control of virus diseases: development of virus resistant germplasm through conventional and engineered/molecular approaches. Define prospective strategies for selecting candidate viruses, identifying specific virus diseases to target, and prioritize specific crops and cultivars for protection approaches.</b></p>	<p>Continue to define suitable strategies for determining target viruses. Isolate and characterize virus-resistant germplasm. Continue work toward engineered resistance in target crops and selected viruses.</p>	X		<p>1. Cotton varieties have been screened under field conditions in the Imperial Valley of California for resistance to cotton leaf crumple geminivirus (CLCrV). A number of lines looked promising, particularly C95-387, which showed no symptoms of infection and in which no virus was detected. Two other lines, C95483 &amp; C95383 also showed potential resistance to CLCrV.</p> <p>2. Efforts are underway to identify tomato germplasm that is resistant to TYLCV as well as to develop genetically engineered tomatoes with resistance to TYLCV.</p>
<p><b>Pursue specific genetic and biological basis for variability in whitefly biotypes, strains, and species; determine impact of different genotypes/phenotypes on whitefly-mediated transmission and on the epidemiology of virus diseases.</b></p>	<p>Continue to study differences in species/strains/biotypes with respect to transmission, host range, mating compatibilities, molecular variability. Determine molecular basis of observed variability in biological, molecular &amp; genetic terms. Infer molecular phylogenies from molecular markers.</p>		X	

**Table B. Viruses, Epidemiology and Virus Vector Interactions.**

Research Approaches	Year 3 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Identification and characterization of new or emerging whitefly-transmitted viruses and strains.</b>	Continue etiological studies and virus characterization. Apply molecular diagnostics to virus identification and evaluation of disease incidence and virus distribution.	X		<p>1.A new cucurbit-infecting geminivirus, <i>Cucurbit leaf crumple virus</i> (CuLCrV) was identified in the Imperial Valley of California. This virus causes leaf crumpling and yellowing in watermelon, cantaloupe, and muskmelon, but no symptoms were observed in honeydew melons. Very high incidences of the virus were detected in fall melons in the Imperial Valley and the virus was detected in melons with leaf crumple symptoms from Blythe, CA and Yuma, AZ. Significant progress has been made in the characterization of this virus (e.g., much of the DNA sequence of the virus has been elucidated) and tools are in hand for the monitoring for the virus in the spring and fall melon crops in 2000.</p> <p>2.<i>Cucurbit yellow stunting disorder virus</i> (CYSDV), a closterovirus in the Genus <i>Crinivirus</i>, was identified for the first time in the United States in the Rio Grande Valley of Texas. CYSDV is transmitted by the silverleaf whitefly. Previously, CYSDV was only known to occur in Europe and the Middle East. Like most of the so-called yellowing viruses, this virus causes yellowing symptoms in older leaves of infected melons. These symptoms can resemble nutritional deficiencies. Thus, infections by these viruses can be hard to diagnose. Molecular tools are available for detection of CYSDV and other related closteroviruses, particularly a degenerate PCR primer pair designed based upon the heat shock protein-like gene that is found in all of these viruses. It will be important to carefully monitor the spread of this virus in Texas and to look for it in other melon growing areas.</p>

**Table B. Viruses, Epidemiology and Virus Vector Interactions.**

Research Approaches	Year 3 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Molecular epidemiology: identification of economic viruses, host plants, and reservoirs, and determination of geographic distribution of viruses.</b>	Establish geographic distribution of viruses and identify sources of inoculum. Assess role of alternative host virus reservoirs on spread of diseases.		X	<p>1.The distribution of CuLCrV in the Imperial Valley of California in 1999 was extensively documented using PCR and degenerate primers for whitefly-transmitted geminiviruses and using squash blot hybridization with a general probe for these viruses. The results of these analyses revealed that the virus had spread extensively by late fall 1999 and that it was infecting muskmelons throughout the Imperial Valley. It will be important to use these tools to monitor for CuLCrV in melons in spring and fall 2000. Infectious DNA clones of CuLCrV are being generated and will be used to determine the host range of this virus.</p> <p>2.Molecular tools are now in place to study the distribution of whitefly-transmitted geminiviruses in Central America and South America. This information will be important in order to develop resistance strategies.</p> <p>3.Many criniviruses have the capacity to infect weed and other hosts. Thus, it will be important to carefully monitor areas in which CYSDV has become established to assess the potential for weed and alternate hosts to contribute to the epidemiology of this disease. Findings from this work will impact management strategies.</p>

**Table B. Viruses, Epidemiology and Virus Vector Interactions. (Continued)**

Research Approaches	Year 3 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Virus-vector interactions, factors affecting virus transmission, and basis for virus-vector specificity; determination of endosymbiont involvement in whitefly-mediated transmission.</b>	Continue studies in progress to determine specific factors involved in virus transmission, and the role of endosymbionts in virus acquisition and transmission.	X		Important advances have been made in understanding the interaction of <i>Tomato yellow leaf curl virus</i> and <i>B. tabaci</i> . There is now evidence for transovarial transmission (rates ranging from 0-10%) and sexual transmission between male and female insects. How these finding impact the epidemiology of the virus and disease management remain to be determined. It was also reported that a chaperonin protein produced in whiteflies by endosymbionts called GroEL may be involved in the protection of the virus in the insect body during it's journey from the gut to the salivary glands. This may involve an interaction between the GroEL chaperonin protein and the viral capsid protein.
<b>Strategies to reduce virus spread by management of cropping systems, reduced transmission frequencies, and other potentially effective approaches.</b>	Continue studies of management approaches for disease abatement. Focus on interdisciplinary studies in conjunction with whitefly control methods in Sections B and C.		X	It was reported that living ground covers and silver plastic reflective mulches can reduce virus spread in Costa Rica and Florida, but only under conditions of low to moderate virus pressure. Of the living ground covers tested, perennial peanuts seemed to be the best. Overall, silver plastic reflective mulch was the best for slowing spread of virus.

**Table B. Viruses, Epidemiology and Virus Vector Interactions. (Continued)**

Research Approaches	Year 3 Goals Statement	Progress Achieved		Significance
		Yes	No	
<p><b>Control of virus diseases: development of virus resistant germplasm through conventional and engineered/molecular approaches. Define prospective strategies for selecting candidate viruses, identifying specific virus diseases to target, and prioritize specific crops and cultivars for protection approaches.</b></p>	<p>Further identification of resistant germplasm and develop new methods of incorporating resistance into crop plants. Evaluate resistance strategies with respect to broad spectrum or virus-specific protection.</p>	X		<p>1. Tomato varieties resistant to TYLCV are now commercially available and some of these provide high levels of resistance. Research in Israel suggests that use of highly resistant varieties (e.g., TY172) will reduce the rate of virus transmission compared to susceptible cultivars. However, moderately resistant cultivars (e.g., Fiona) may enhance virus transmission and actually enhance epidemics of TYLCV because they provide better sources of inoculum over longer periods of time compared with susceptible cultivars, which become severely diseased or die thereby making them poor sources of inoculum. These findings may influence how management strategies are developed.</p> <p>2. Cotton varieties continue to be screened under field conditions in the Imperial Valley of California for resistance to <i>Cotton leaf crumple virus</i> (CLCrV). A number of lines continue to look promising, particularly line C95-387, which showed no symptoms of infection and in which no virus was detected. Two other lines, C95 483 and C95 383 also continue to show potential resistance to CLCrV. It will be very important to screen these materials in India or Pakistan to see if they have any resistance to the devastating cotton leaf curl virus, another whitefly-transmitted geminivirus that infects cotton.</p> <p>3. Progress has been made in the generation of transgenic crops that are resistant to geminivirus infection. The major strategy that has been pursued to date is that of pathogen-derived resistance in which a wild-type or mutated virus gene or sequenced is introduced into the crop plant of choice. The idea is that expression of the viral sequence/ protein will interfere with the normal life cycle of the virus and, thus, interfere with the viral infection process. A number of viral genes have been evaluated to date including: capsid protein, replication-associated protein and movement protein. Promising levels of resistance have been obtained, though no plants have been reported to be highly resistant (i.e., immune).</p>



**Table B. Viruses, Epidemiology and Virus Vector Interactions. (Continued)**

Research Approaches	Year 3 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>**Pursue specific genetic and biological basis for variability in whitefly biotypes, strains, and species; determine impact of different genotypes/phenotypes on whitefly-mediated transmission and on the epidemiology of virus diseases.</b>	Continue with work from previous years. Study impact of biotypes, strains, and species differences in the disease spread, crop damage, and specific control measures to reduce whitefly vector populations. Linkages with biological and chemical control sections.		X	This research approach should be transferred to section A

\*\* Transfer to Table A 3/31/2000

**Table B. Viruses, Epidemiology and Virus Vector Interactions, 2001-2002.**

Research Approaches	Year 4/5 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Identification and characterization of new or emerging whitefly-transmitted viruses and strains.</b>	<p>Continue etiological studies and virus characterization efforts. Apply molecular diagnostics to virus identification and evaluation of disease incidence and virus distribution</p> <p>Summarize and review results. Determine areas of new research.</p>	X		<p>1. Continued characterization of the new cucurbit-infecting geminivirus, <i>Cucurbit leaf crumple virus</i> (CuLCrV)/<i>Cucurbit leaf curl virus</i> (CuLCV), has occurred. Infectious clones of isolates from Arizona and California have been obtained and sequenced completely. Results support the concept that it is a new virus species that is closely related to <i>Squash leaf curl virus</i> (SLCV). 2. Two viruses responsible for causing golden mosaic symptoms in <i>Macroptilium lathryoides</i> in Florida have been characterized. One virus is <i>Macroptilium mosaic virus</i> (MaMV) and the other is <i>Macroptilium golden mosaic virus</i> (MAYMV). Infectious clones have been generated and sequence and other analyses suggest that these are distinct begomoviruses. They are infectious in common bean plants, though their role in bean golden mosaic disease in Florida remains to be established. 3. A squash blot hybridization assay has been commercialized by AgDia Co. (Elkhart, IN) and is available as a general tool for detection of geminiviruses in plants. 4. A dot blot assay has been developed for detection of criniviruses in plants.</p> <p><b>New areas of research:</b></p> <p>1. Continue characterization of new emerging whitefly-transmitted begomoviruses, criniviruses and potyviruses. 2. Continue to refine detection and identification methods, e.g., develop virus-specific detection tools. 3. Improve our understanding of virus distribution: regional and global</p>

**Table B. Viruses, Epidemiology and Virus Vector Interactions, 2001-2002 (Continued)**

Research Approaches	Year 4/5 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Molecular epidemiology: identification of economic viruses, host plants, and reservoirs, and determination of geographic distribution of viruses.</b>	Identify and characterize virus involvement in disease establishment and spread. Assess potential methods of reducing virus reservoirs as a method of reducing disease  Review and make recommendations for further research and potential implementation of results.	X		1. It was established that Tomato yellow leaf curl virus could be effectively managed in the Dominican Republic through the use of a whitefly host-free period. However, the virus reappears after the host free period. Using PCR, TYLCV infections were detected in a number of symptomless weeds hosts. It is hypothesized that the virus overwinters in such hosts, but that these hosts are poor inoculum sources, thereby resulting in the slow build-up of the virus after the host free period. 2. In Florida, effective management of TYLCV has been accomplished through the use of imidacloprid and insect growth regulators (IGRs). In addition the use of the fungal insecticide, <i>Beauveria bassiana</i> has also helped. Other factors influencing disease development include time and location of planting. However, TYLCV appears to be established in Florida and it is spreading to other states in the U.S. In the case of Bean golden yellow mosaic virus, the main reservoir appears to be common bean. Thus, time of planting is very important; i.e., do not establish young plants near older infected plantings. 3. Good progress has been made on understanding the epidemiology of <i>Tomato infectious chlorosis virus</i> (TICV) in Southern California fresh market tomato fields. TICV is vectored by the greenhouse whitefly and is common in southern California coastal production areas and greenhouses. The virus occurs in a number of weed hosts as well as tomato, but tomato seems to be the key host in terms of viral epidemiology. Thus, efforts for disease management need to focus on effective whitefly management and regional planning in terms of establishing tomato plantings. <b>New areas of research:</b> 1. Identify CuLCV/CuLCrV reservoirs and distribution 2. Continue efforts to define reservoir hosts for criniviruses (e.g., TICV, ToCV and <i>Cucurbit yellow stunt disorder virus</i> )

**Table B. Viruses, Epidemiology and Virus Vector Interactions, 2001-2002 (Continued)**

Research Approaches	Year 4/5 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Virus-vector interactions, factors affecting virus transmission, and basis for virus-vector specificity; determination of endosymbiont involvement in whitefly-mediated transmission.</b>	Continue virus-vector interactions studies toward the development of approaches for disease control.	X		1. Progress has been made in understanding the nature of whitefly transmission of TYLCV and Tomato mottle virus in Florida. In contrast to reports on TYLCV in Israel, neither TYLCV in Florida nor ToMoV was found to be transovarially transmitted. This has important implications for disease epidemiology. <b>New areas of research:</b> 1. More studies need to be conducted on transovarial transmission (or not) with different begomoviruses. 2. Need more information on endosymbiont/whitefly interactions..
	Summarize findings and suggest new research needs; implementation of existing knowledge			
<b>Strategies to reduce virus spread by management of cropping systems, reduced transmission frequencies, and other potentially effective approaches.</b>	Evaluate strategies for crop management and impact on disease epidemiology.	X		Success in managing whitefly-transmitted geminivirus diseases was reported for TYLCV in the Dominican Republic based upon implementation of a whitefly host free period. The success was based upon the implementation of a regional approach, in which the host free period was enforced in the major tomato growing regions of the island. In addition, efforts were made to plow under crop debris after harvest. The recovery of the tomato industry from devastating losses caused by TYLCV has resulted in the acceptance of this regional approach by growers and others in the Dominican Republic. Similarly, effective control of BGMV in Florida has been achieved by better planting strategies and the use of a BGMV-resistant variety. Management of TICV and <i>Tomato chlorosis virus</i> (ToCV) has been achieved through the use of crop rotation and other approaches that reduce populations of the greenhouse whitefly. <b>New areas of research:</b> 1. Continue to optimize and modify cropping systems for effective disease management.
	Evaluate approaches and identify areas of future research for disease control by management of cropping systems. Linkages with IPM approaches			

**Table B. Viruses, Epidemiology and Virus Vector Interactions, 2001-2002 (Continued)**

Research Approaches	Year 4/5 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Control of virus diseases: development of virus resistant germplasm through conventional and engineered/ molecular approaches. Define prospective strategies for selecting candidate viruses, identifying specific virus diseases to target, and prioritize specific crops and cultivars for protection approaches.</b>	Continue development of resistant varieties. Evaluate resistance strategies with respect to broad spectrum or virus-specific protection. Define mechanisms of resistance.	X		Commercially available tomato varieties with TYLCV resistance are now being used in the Dominican Republic and allowing for relatively high yields even under high virus pressure. Together with the implementation of the whitefly host free period, the use of these varieties has helped allow the tomato industry in the Dominican Republic recover to per-TYLCV levels. Two cotton lines were identified continue to show very high levels of resistance to Cotton leaf crumple virus (CLCrV). It will be very important to screen these materials in India or Pakistan to see if they have any resistance to the devastating cotton leaf curl virus, another whitefly-transmitted geminivirus that infects cotton. Promising levels of resistance to TICV have been identified in wild tomato species. This could result in the introgression of this resistance into <i>L. esculentum</i> germ plasm. An agroinoculation procedure has been developed that allows for the screening of cucurbit germ plasm for resistance to CLCuV/CuLCrV. Progress has been made in the development of bush beans with resistance to Bean golden mosaic virus. This has been accomplished through a traditional breeding approach. These varieties have been released for commercial use in Florida and are helping in the management of this disease. <b>New areas of research:</b> Continue to evaluate strategies for generating genetically modified crops with resistance to begomoviruses. <b>Technology transfer:</b> The squash blot hybridization method has now been made into a commercial kit for the detection of whitefly-transmitted geminiviruses. A crinivirus dot blot assay may also be commercialized in the near future. TYLCV-resistant tomato cultivars are now available and these allow for respectable yields in area with high incidences of TYLCV, and BGMV-resistant bean varieties also have been developed. There is a need to evaluate the CLCrV-resistant lines in Pakistan and India for resistance to <i>Cotton leaf curl virus</i> . An agroinoculation method has been developed for CLCuV/CuLCrV and it is being evaluated for screening cucurbits for resistance to this new begomovirus.
	Evaluate resistant plants in greenhouse and field experimentation, and identify additional research. Molecular-based monitoring of transgenes in environment	X		

**Reports of Research Progress**  
**Section C: Chemical Control, Biopesticides, Resistance Management, and Application Methods**  
Co-Chairs: Shirley Taylor and John Palumbo

**Investigator's Name:** S. J. Castle<sup>1</sup>, F. J. Byrne<sup>2</sup>, N. Prabhaker<sup>2</sup>, & N. C. Toscano<sup>2</sup>

**Affiliation & Location:** <sup>1</sup>USDA, ARS, Western Cotton Research Lab, Phoenix, AZ; <sup>2</sup>Department of Entomology, University of California, Riverside

**Research & Implementation Area:** Section C: Chemical Control, Biopesticides, Resistance Management, and Application Methods

**Dates Covered by the Report:** 2000-2001

**Sources of Variation in Systemic Uptake Bioassays of Neonicotinoid Insecticides**

Interpretation of insecticide bioassays is predicated on the assumption that all test subjects exposed to a dosage of insecticide actually received that dosage and not one greater or lesser. Mortality at a series of insecticide dosages can then be analyzed statistically and the pertinent statistics for different populations compared to assess relative differences in susceptibility to a particular insecticide. Most insecticides are effective as contact poisons that act either by direct exposure with a contact spray, or by indirect exposure to a residue deposited on a plant or other surface within the insect's environment. Accordingly, bioassays designed to test the susceptibility of a sample population to a contact insecticide have relied upon various devices to ensure equal exposure of test subjects either by topical application or by uniform deposition of a residue. Precise quantities of known concentrations of an insecticide could then be deposited upon a test insect or surface to assess the susceptibility of the sample subjects. Treatment replication helps to compensate for minor variability in test conditions and reduce experimental error.

Neonicotinoid insecticides are active both as contact and as ingested insecticides. However, bioassays performed worldwide on imidacloprid, the longest available and most widely used of the neonicotinoids, have relied principally upon systemic uptake of imidacloprid solutions of known concentrations by severed plants or leaves. Interpretation of bioassay results have been made on the basis of the concentration of the imidacloprid solutions only, without regard for how much solution is actually taken up by the sample leaf or small plant. In many laboratories, a disk is punched from the uptake leaf and placed on agar within a petri dish or vial, to which test subjects are added and subsequently evaluated for mortality. In addition to substantial variation in uptake both within and between treatment concentrations of imidacloprid by the severed leaves or plants, variability in the distribution of imidacloprid within leaves can also affect mortality assessment of the test insects. We have measured uptake by detached leaves over a series of concentrations of imidacloprid and thiamethoxam ranging between 1 and 1000 ppm and found that volume uptake by leaves progressively declines with increasing concentrations. This is especially true for imidacloprid, a less soluble compound than thiamethoxam, at concentrations above c. 300 ppm. Variation in volume uptake within a concentration among individual detached leaves can vary by as much as 4-fold. This degree of variability among leaves often is carried over into variable mortality among replicates within a treatment concentration. However, volume uptake by individual leaves tends to be a poor predictor of mortality, suggesting that within-leaf distribution of active ingredients is non-uniform.

**Investigator's Name(s):** T. J. Henneberry, L. Forlow Jech, D. L. Hendrix, T. de la Torre, and J. Maurer

**Affiliation & Location:** USDA-ARS, Western Cotton Research Laboratory, Phoenix, AZ

**Research & Implementation Area:** Section C: Chemical Control, Biopesticides, Resistance Management, and Application Methods

**Dates Covered by Report:** 2000-2001

**Effects of Applaud and Knack on Sweetpotato Whitefly (SPW) Nymph Mortality and Adult and Nymph Honeydew Production**

Sweetpotato whiteflies, *Bemisia tabaci* (Gennadius) produce honeydew that results in cotton lint contamination causing reduced lint processing efficiency. The insect growth regulators, Applaud® and Knack®, provide effective control of SPW on cotton by interfering with their reproduction and development. We investigated their effect on SPW honeydew production. Amounts of the major sugar components of honeydew produced by adults and nymphs collected on day six following Applaud or Knack applications to cotton field plots were not significantly different compared to amounts produced by those collected from untreated plots. Similar results occurred with SPW adults confined for 48 h on Applaud or Knack < 1 to 48 h-old residues in the laboratory following cotton leaf dips. Also in the laboratory, mortality of adults was not affected by nebulizer applied contact spray applications and honeydew sugars were not significantly reduced. In contrast, mortality of first and second instar SPW nymphs on leaves was higher on day six following leaf dips in Applaud solutions compared with leaf dips in Knack or water solutions. Nymph mortality on day six following leaf-dips in Knack solutions was higher than mortality of nymphs following leaf-dips in water. Honeydew collected from nymphs during two to 50 h intervals after leaf dip treatment had reduced amounts of glucose, fructose and trehalulose, but not sucrose and melezitose per nymph compared with honeydew from nymphs on leaves dipped in water.

**Investigator's Names:** Tong-Xian Liu<sup>1</sup>, A. N. Sparks<sup>1</sup>, Jr., S. M. Greenberg<sup>2</sup>, and Ge -Mei Liang<sup>1</sup>

**Affiliation & Location:** <sup>1</sup>Texas Agricultural Research and Extension Center, Texas A&M University, 2415 E. Highway 83, Weslaco, TX 78596, and <sup>2</sup>Integrated Farming & Natural Resources Research Unit, USDA-ARS, 2401 E. Highway 83, Weslaco, TX 78596

**Research & Implementation Area:** Section C: Chemical Control, Biopesticides, Resistance Management, and Application Methods

**Dates Covered by the Report:** 2000

### Effects of Defoliants Alone and in Combination with Insecticides on Silverleaf Whitefly

Silverleaf whitefly, *Bemisia argentifolii* Bellows & Perring, is one of the most important pests of cotton, vegetables and other crops in the US. The spring-summer cotton and fall-winter-spring vegetable cropping system in south Texas is unique, this sequencing of crop production allows whiteflies to thrive year-round. Although defoliants are used in every season in south Texas, the role of these defoliants on *B. argentifolii* and their natural enemies are unknown. The objective of this study was to determine the effects of defoliants on survival, migration and dispersal of *B. argentifolii* and their parasitoids after cotton plants were defoliated. Def 6 (S,S,S-tributylphosphorotrithioate); Karate (lambda-cyhalothrin), and Guthion (azinphosmethyl), were used. Filed rates were: full rate of Def at (2 pts/ac), full rate of Dropp at (0.2 lb/ac), full rate of Karate at (0.3 lb [AI]/ac), half rate of Guthion (0.25 lb [AI]/ac), full rate of Def + full rate of Karate, full rate of Def + half rate of Guthion. Untreated plots were used as controls. Each plot was 6 rows of cotton and 45 m long. Each treatments had three replications. Cotton leaves, 4th or fifth leaf from each plant, were collected directly from the treated plants 1 day after the application of chemicals. Twenty living individuals of each developmental stage were marked with an ink pen. The immatures were checked daily for survival until they developed to the next stage or died. Yellow sticky cards were used to trap whiteflies and parasitoids in the field. Five cards were placed in the middle of the plot, about 8 m apart. The cards were collected and replaced with new ones at 1-3 d intervals after the chemical applications. All arthropods trapped on the cards were counted in the laboratory.

Def and in combination with insecticides did significantly affect the survival rates of young nymphs but not eggs, older nymphs and pupae. Significant lower survival rates (13.3-30.0%) were found in Def alone and in combination with insecticides, compared with 70% survival rate in untreated control. Generally, the cotton leaves treated with Def alone had higher survival rates in all immatures stages than those treated with combinations of Def+insecticides. Karate and Guthion alone did not have significant effects on *B. argentifolii*, although lower survival rates were found. However, we do not know if Guthion and Karate, in combination with Def, added any synergistic effects on *B. argentifolii* to Def. Although numbers of *B. argentifolii* adults caught on yellow sticky cards varied greatly from date to date among the treatments, there were no clear picture showing significant effects among the treatments. Generally, more whitefly adults were caught on yellow sticky cards in the plots treated with Karate and Guthion, and slightly fewer in the plots treated with Def and untreated control. It seems that fewer whitefly adults took off and fly high in the untreated plots because the plants were green, and abundant young and nutritious leaves were available. However, we do not know exactly why fewer whitefly adults were caught in the plots treated with Def or Def+insecticide. Numbers of parasitoids, *Eretmocerus* spp. and *Encarsia* spp., caught on the yellow sticky cards are shown in Figure 2. Generally, among the treatments, yellow sticky cards placed in Def-treated plots caught similar numbers of parasitoids to those in untreated plots, but had significantly fewer parasitoids than from other chemical-treated plots. Numbers of parasitoids caught on yellow sticky cards placed in the Karate-treated plots was the greatest, followed by those placed in Guthion-and Def+Karate-treated plots. Results from this study indicate that Karate had no significant effects on the parasitoids. Although the yellow sticky cards in Guthion-treated plots than those in Karate-treated plots caught fewer parasitoids, the effects might not be directly related to Karate and Guthion themselves. Whereas we do not know the reason why the yellow sticky cards caught similar numbers of parasitoids in untreated and Def-treated plots, possible explanations might be that in the Def-treated plots, application of Def might affect the emergence or migration of parasitoids, whereas in untreated plots, parasitoids might not likely fly high or disperse with the presence of hosts (live whitefly nymphs), and therefore, fewer were caught on the yellow sticky cards. Because Karate and Guthion had no effects on leaf defoliation, we do not know why more parasitoids were caught on the yellow sticky cards. Perhaps, the odors or other chemical cues stimulated the parasitoids to take off, fly high and be caught. Many species of insects and spiders were caught on yellow sticky cards.



**Investigator's Name:** Tong-Xian Liu

**Affiliation & Location:** Texas Agricultural Experiment Station, Texas A&M University System 2415 E. Highway 83, Weslaco, TX 78596-8399

**Research & Implementation Area:** Section C: Chemical Control, Biopesticides, Resistance Management, and Application Methods

**Dates Covered by the Report:** 2000

### **Interactions of Planting Dates & Applications of Imidacloprid for Management of Silverleaf Whitefly on Spring Melon in South Texas**

The silverleaf whitefly, *Bemisia argentifolii* Bellows & Perring, continues to threaten the melon production in south Texas. Melon was severely damaged by the whitefly if Admire or other measure is not applied. In south Texas, the silverleaf whitefly infests melons from late March, and the whitefly population gradually increase in April, and reach the peak in late April or May. Melons planted late usually are more vulnerable to whitefly attack and have more severe damage than the melons planted earlier. The objective of this study was to determine the relationships and interactions between planting dates and applications of Admire (imidacloprid) for management of the silverleaf whitefly on spring melons in south Texas. There were 16 treatments, and each will had 4 replications. Each plot was 30 ft long with two separate rows 80 in wide, and 10-20 plants each. The plots were arranged in a randomized complete block design. Melons were planted in four dates, 13 and 27 Jan., and 10 and 24 Feb., 2 weeks apart. In each planting date, there were four treatments, Admire were applied in full rate (16 oz/ac) at transplanting, full rate in the mid-season, half rate in the mid-season, and an untreated control. Whitefly adults were sampled when whitefly population increase significantly. Thereafter, plants were sampled in 7-day intervals. Adults was counted by leaf turn method. When plants have <6 leaves, adults on the oldest leaf were counted, and when plants have >6 leaves, adults on the 4<sup>th</sup> or 5<sup>th</sup> leaf from the proximal were counted. Red-eyed nymphs (pupae), empty pupal cases per 4 leaf-discs (1 cm diameter) per leaf were counted. Virus incidence, leaf area covered by sooty mold, yield and sugar contents were recorded.

Interactions of planting dates and insecticide application were significant, indicating that numbers of adults on the plants with different planting dates and insecticide application were significantly different. Results also indicated that the later the plants were planted, the more whitefly adults were found, with 5.7, 4.5, 3.1 and 2.7 adults per leaf on the plant that were planted on 24, 10 Feb., 27 and 13 Jan., respectively. On the plants planted on 13 Jan., fewest adults were found on the plants treated with Admire in full rate at transplanting, followed by the plants treated Admire in full rate in mid-season. On the plants planted on 27 Jan., numbers of whitefly adults on the plants treated with Admire whether applied at transplanting, mid-season, full rate or half rate, were significantly reduced before mid-April, and were not significantly different in the last 4-5 weeks. Numbers of adults on the plants planted on 10 and 24 Feb. were very low until mid-April., and increased rapidly in late April, and peaked in early May. Plants treated with Admire (all treatments) had fewer adults on the plants planted on 10 Feb. compared with untreated plants. However, numbers of adults on the plants planted on 24 Feb. were significantly different among the treatments, with significantly fewer adults on the plant treated with Admire applied in full rate in mid-season, followed by the plants treated Admire applied in full rate at transplanting. Few red-eyed nymphs were found on the plants that planted in all four different planting dates until late March. Numbers of red-eyed nymphs increased gradually until early April, and then increased rapidly from late April until the end of the season. Generally, numbers of whitefly nymphs on the plants planted on different dates were significantly different. The plants planted early on 10 Jan. had the fewest numbers of nymphs, followed by the plants planted on 27 Jan. and on 24 Feb., the plants planted on 10 Feb. had the most. Numbers of red-eyed nymphs on the plants were also significantly different among the plants applied Admire in different rates and on different dates. The plants treated with Admire in full rate at transplanting had the fewest nymphs, followed by the plants with Admire applied full rate in mid-season, and then the plants with Admire in half rate in mid-season. Untreated plants had the most number of nymphs. On the plants planted on 13 Jan., numbers of nymphs on the plants treated with Admire at full rates at transplanting or mid-season were significantly fewer than those on the plants treated with Admire applied in half rate and those on untreated plants. In contrast, numbers of nymphs on the plants planted on 27 Jan. were not significantly different among the three treatments with Admire applied in full rates at transplanting, in mid-season, and half rate in mid-season. On the plants planted on 10 Feb., numbers of nymphs on the plants treated Admire in full rates at transplanting or in mid-season were significantly fewer than those on other treatments, especially in the late season. Numbers of nymphs on the plants planted on 27 Feb. among the four treatments were similar to those on the plants planted in 13 Jan., with fewest nymphs on the plants treated Admire applied in full rate at transplanting, followed by Admire applied in full rate in mid-season. Number of whitefly nymphs on the plants treated with Admire applied in half rate in mid-season was not significantly different from those on untreated plants. The numbers of melons harvested from plots of different treatments differ significantly. Total numbers of melons harvested from each plots among the treatments of different planting dates and insecticide application were significantly different. Among the planting dates, more melons were harvested from the plants planted on 13 Jan. and on 24 Feb. Numbers of melons harvested from the plants among the treatments of Admire applied in different rates and on different date were not significantly different. However, the size and weights of the melons among the treatments were not significantly different.

**Investigator's Name:** Tong-Xian Liu

**Affiliation & Location:** Texas Agricultural Experiment Station, Texas A&M University System, 2415 E. Highway 83, Weslaco, TX 78596-8399

**Research & Implementation Area:** Section C Chemical Control, Biopesticides, Resistance Management, and Application Methods

**Dates Covered By The Report:** 2000

### **Effects of Planting Dates and Applications of Imidacloprid For Management of Silverleaf Whitefly in Melons**

The silverleaf whitefly, *Bemisia argentifolii* Bellows & Perring, continues to threaten the melon production in south Texas. Melon was severely damaged by the whitefly if Admire or other measure is not applied. The objective of this study was to determine the relationships and interactions between planting dates and applications of Admire (imidacloprid) for management of the silverleaf whitefly on spring melons in south Texas. There were 16 treatments, and each will had 4 replications. Each plot was 30 ft long with two separate rows 80 in wide, and 10-20 plants each. The plots were arranged in a randomized complete block design. Melons were planted in four dates, 13 and 27 Jan., and 10 and 24 Feb., 2 weeks apart. In each planting date, there were four treatments, Admire were applied in full rate (16 oz/ac) at transplanting, full rate in the mid-season, half rate in the mid-season, and an untreated control. Whitefly adults were sampled when whitefly population increase significantly. Thereafter, plants were sampled in 7-day intervals. Adults were counted by leaf turn method. When plants have <6 leaves, adults on the oldest leaf were counted, and when plants have >6 leaves, adults on the 4<sup>th</sup> or 5<sup>th</sup> leaf from the proximal were counted. Red-eyed nymphs (pupae), empty pupal cases per 4 leaf-discs (1 cm diameter) per leaf were counted.

Interactions of planting dates and insecticide application were significant. Results also indicated that the later the plants were planted, the more whitefly adults were found, with 5.7, 4.5, 3.1 and 2.7 adults per leaf on the plant that were planted on 24, 10 Feb., 27 and 13 Jan., respectively. On the plants planted on 13 Jan., fewest adults were found on the plants treated with Admire in full rate at transplanting, followed by the plants treated Admire in full rate in mid-season. On the plants planted on 27 Jan., numbers of whitefly adults on the plants treated with Admire whether applied at transplanting, mid-season, full rate or half rate, were significantly reduced before mid-April, and were not significantly different in the last 4-5 weeks. Numbers of adults on the plants planted on 10 and 24 Feb. were very low until mid-April., and increased rapidly in late April, and peaked in early May. Plants treated with Admire (all treatments) had fewer adults on the plants planted on 10 Feb. compared with untreated plants. However, numbers of adults on the plants planted on 24 Feb. were significantly different among the treatments, with significantly fewer adults on the plant treated with Admire applied in full rate in mid-season, followed by the plants treated Admire applied in full rate at transplanting. Few red-eyed nymphs were found on the plants that planted in all four different planting dates until late March. Numbers of red-eyed nymphs increased gradually until early April, and then increased rapidly from late April until the end of the season. Generally, numbers of whitefly nymphs on the plants planted on different dates were significantly different. The plants planted early on 10 Jan. had the fewest numbers of nymphs, followed by the plants planted on 27 Jan. and on 24 Feb., the plants planted on 10 Feb. had the most. Numbers of red-eyed nymphs on the plants were also significantly different among the plants applied Admire in different rates and on different dates. The plants treated with Admire in full rate at transplanting had the fewest nymphs, followed by the plants with Admire applied full rate in mid-season, and then the plants with Admire in half rate in mid-season. Untreated plants had the most number of nymphs. On the plants planted on 13 Jan., numbers of nymphs on the plants treated with Admire at full rates at transplanting or mid-season were significantly fewer than those on the plants treated with Admire applied in half rate and those on untreated plants. In contrast, numbers of nymphs on the plants planted on 27 Jan. were not significantly different among the three treatments with Admire applied in full rates at transplanting, in mid-season, and half rate in mid-season. On the plants planted on 10 Feb., numbers of nymphs on the plants treated Admire in full rates at transplanting or in mid-season were significantly fewer than those on other treatments, especially in the late season. Numbers of nymphs on the plants planted on 27 Feb. among the four treatments were similar to those on the plants planted in 13 Jan., with fewest nymphs on the plants treated Admire applied in full rate at transplanting, followed by Admire applied in full rate in mid-season. Number of whitefly nymphs on the plants treated with Admire applied in half rate in mid-season was not significantly different from those on untreated plants.

**Investigator's Names:** Tong-Xian Liu<sup>1</sup>, S. M. Greenberg<sup>2</sup>, and Robert R. Saldana<sup>1</sup>

**Affiliation & Location:** <sup>1</sup>Texas Agricultural Research and Extension Center, Texas A&M University, 2415 E. Highway 83, Weslaco, TX 78596, and <sup>2</sup>Integrated Farming & natural Resources Research Unit, USDA-ARS, 2401 E Highway 83, Weslaco, TX 78596

**Research & Implementation Area:** Section C: Chemical Control, Biopesticides, Resistance Management, and Application Methods

**Dates Covered by the Report:** 2001

#### **Effects of Defoliants Alone and in Combination with Insecticides on Silverleaf Whitefly and Parasitoids, 2001**

Effects of Def and Dropp alone and in combination with two insecticides, Karate (a pyrethroid) and Guthion (an organophosphate) on silverleaf whitefly, *Bemisia argentifolii* Bellows & Perring were determined in 2001. Two defoliants, Def 6 (S,S,S-tributylphosphorotrithioate, Bayer, Kansas City, MO); Dropp (50% thidiazuron, Aventis [AgrEvo], Wilmington, DE), and two insecticides, a pyrethroid, Karate 2.08CS (lambda-cyhalothrin, Zeneca, Wilmington, DE), and an organophosphate, Guthion 2L (azinphosmethyl, Bayer, Kansas City, MO), were used in this study. There were eight treatments with different combinations of defoliants and insecticides at different rates: 1. Def (2 pts/ac)+Dropp (0.2lb/ac) + Guthion (0.25 lb/ac); 2. Def (1 pt/ac) + Dropp (0.1 lb/ac); 3. Dropp (0.2 lb/ac) + Guthion (0.5 lb/ac); 4. Def (2 pt/ac) + Guthion (0.25 lb/ac); 5. Def (2 pt/ac) + Karate (0.03 lb AI/ac); 6. Guthion (0.5 lb AI/ac); 7. Karate (0.03 lb AI/ac), and 8. untreated control. To test the effect of applied chemicals on adult emergence for *B. argentifolii*, the third, and fifth and seventh leaf from the terminal was collected after chemical applications. The leaves were placed in paper bags and held in the laboratory for 3-4 weeks. Numbers of whitefly and parasitoid adults emerged from each bag were examined.

Numbers of whiteflies were significantly reduced on the leaves sampled on 26 July treated with defoliants and their combination with Karate and Guthion except for the cotton leaves treated Karate and Def + Dropp at 0.5X rate on which number of whiteflies was not significantly different from those on untreated leaves. Defoliants and their combinations with insecticides also significantly reduced the number of silverleaf whitefly parasitized. Again, untreated leaves had the most parasitized whiteflies, followed by Karate-treated leaves, and then other treated leaves. Cotton leaves that treated with Def + Dropp + Guthion at 0.5X rate and Def + Guthion at 1.0X rate had the least number of parasitized whiteflies. Few whiteflies and parasitoids were found on the cotton leaves sampled on 16 August when leaves were almost defoliated. Therefore, there were no significant differences in numbers of whiteflies and parasitoids on sampled leaves. Numbers of *B. argentifolii* adults emerged from treated leaves after treatment. Cotton leaves treated with defoliants alone or in combination with Karate and Guthion reduced number of whitefly adults emerged in the samples on 26 July, although numbers of adults emerged in the treatments of Guthion and Karate alone and Def + Guthion at 1.0X rate were not significantly different from that in untreated control. There were no significant differences for both whitefly adult and parasitoids emerged for the samples in 16 August.

In conclusion, defoliants, Def and Dropp, and their combinations with Guthion and Karate significantly affected the infection and survival of both silverleaf whiteflies and their parasitoids, *Encarsia* spp. and *Eretmocerus* spp, although the effects varied greatly among the treatments. Karate had no significant effects on silverleaf whitefly and its parasitoids. Combining defoliants and insecticides increased the effectiveness on whiteflies and parasitoids on cotton.

**Investigator's Name(s):** C.L. McKenzie<sup>1</sup> and Gary J. Puterka<sup>2</sup>

**Affiliation & Location:** <sup>1</sup>USDA-ARS, Horticultural Research Lab, 2001 South Rock Road, Ft. Pierce, FL; <sup>2</sup>USDA-ARS, Appalachian Fruit Research Station, 45 Wiltshire Rd., Kearneysville, WV

**Research & Implementation Area:** Section C: Chemical Control, Biopesticides, Resistance Management, and Application Methods

**Dates Covered by the Report:** 2000-2001

### **Efficacy of Sucrose Octanoate to Whitefly Using a Plant-based Bioassay**

The objectives of this study were to determine the insecticidal activity of a synthetic analogue of natural sugar esters (sucrose octanoate (SO)) found in leaf trichomes of wild tobacco to egg, nymph and adult sweetpotato whitefly, *Bemisia tabaci* biotype B, (=silverleaf whitefly, *Bemisia argentifolii*), vector of Begomoviruses.

A plant-based petri dish bioassay system was developed to hold single tomato leaves infested with whitefly for their entire life cycle (egg to adult). A hole was cut in the lid of a Polystyrene petri dish (100x20 mm) to facilitate ventilation and ultra-fine screen mesh was used to cover the hole to prevent whitefly escapes. Polystyrene conical tubes (15 ml, 17x120 mm) were filled with a plant tissue culture water agar and fertilizer (9-45-15) mixture. A hole was cut at the base of the petri dish to hold the conical tube so that it fit tightly. Tomato leaves were cut so that the terminal leaf of the leaflet would fit in the petri dish. The remaining leaves of the leaflet were cut off and the stem was placed as far into the agar as possible with the abaxial leaf surface facing the screen to facilitate counting. A small hole was drilled into top of the conical tube above the agar line so that H<sub>2</sub>O could be added as needed (leaves will sprout roots and last > 28 d).

The SO solutions were applied using an ultra-low volume spray device that consisted of a spray platform that holds a pressurizable spray bottle at the proper distance and angle. Measured amounts (200 µl) of SO solution were placed in a test tube (12 x 75 mm). The spray bottle siphon tube was placed in the test tube so that the test tube fit into the nozzle -pump body. Parafilm was wrapped around the top of the test tube to insure a tight fit and the spray bottle was pressurized to ~10 psi w/ 20 strokes of the pump mechanism to deliver a fine spray for each application. Both the abaxial and adaxial sides of the leaf were sprayed with 200 µl of SO solution.

Whiteflies were obtained from a laboratory colony maintained at USHRL, Ft. Pierce, FL. Sucrose octanoate solutions were prepared in concentrations ranging from 125 to 48,000 ppm in ddH<sub>2</sub>O plus a ddH<sub>2</sub>O control. Rates varied depending on the lifestage evaluated and were applied to eggs, nymphs or adults using an ultra-low volume spray device. Freshly laid whitefly eggs (24-48 hrs old) were sprayed and evaluated 7, 14, 21 and 28 DAT (until adult emergence). Whitefly nymphs were sprayed at the crawler/2nd instar stage and evaluated at 1, 3, 7 and 14 DAT (until adult emergence). Whitefly adults were sprayed and evaluated 3, 6, and 24 hours after treatment (HAT). Each concentration was replicated 5 times.

Whitefly egg and nymph mortality increased over time whereas whitefly adult mortality virtually remained the same. Freshly laid eggs sprayed with the higher concentrations (12,000 - 48,000) had significantly fewer adults emerge. Lower concentrations applied to freshly laid eggs had little effect on adult emergence. Toxicity of SO to whitefly nymphs ranged from LC<sup>90</sup> values of 55,827 (20,162 - 774,817) ppm to 2,225 (1,517 - 11,255) ppm at 1 and 14 DAT (=adults emerged), respectively. Adult whiteflies were killed immediately if they were going to die and LC<sup>90</sup> values calculated 1 DAT were 5,174 (4,345-6,602) ppm. Preliminary results indicate SO could be an effective tool for nymph and adult whitefly control to levels of >90% at higher concentrations of SO. Good coverage is key to efficacy.

**Investigators Name(s):** Eric T. Natwick and Keith S. Mayberry

**Affiliation & Location:** UC Cooperative Extension, UC Desert Research and Extension Center, Holtville, CA

**Research & Implementation Area:** Section C: Chemical Control, Biopesticides, Resistance Management, and Application Methods

**Dates Covered by Report:** September 2000 through December 2000

### **Neonicotinoid Insecticide Efficacy for Silverleaf Whitefly Control In Cabbage, 2000**

Cabbage var. Charmant was sown at UC Desert Research & Extension Center 4 October 2000. Six insecticide treatments and a non-treated control were replicated five times in a randomized complete design experiment. Insecticide treatments were as follows: Admire 2F at 0.25 lb ai/acre and Platinum 2SC at 0.141 lb ai/acre injected 3 inches below the seed-line on 15 September, and Assail 70 WP at 0.0375 lb ai/acre, Assail 70 WP at 0.05 lb ai/acre, Assail 70 WP at 0.075 lb ai/acre, and Actara 25 WG at 0.047 lb ai /acre were applied as foliar spray treatments on 24 October, 8 and 16 November. Silverleaf whitefly, *Bemisia argentifolii*, were sampled by counting adults via leaf turn of basal leaves on ten plants at random from each plot and nymphs were counted on 1.65 cm<sup>2</sup> of leaf surface from basal leaves of ten plants at random from each plot on 16, 23 & 30 October, 6, 13, 20 & 27 November, 4, 11, & 18 December.

The seasonal silverleaf whitefly adult per leaf mean for the non-treated control (4.2) was greater than the seasonal means for Admire 2F (3.0), Assail 70 WP at 0.05 lb ai/acre (3.3) and Assail 70 WP at 0.075 lb ai/acre (3.5), but the non-treated seasonal adult mean was not greater than seasonal means for Platinum 2SC (3.8), Actara 25 WG (3.7), nor Assail 70WP at 0.0375 lb ai/acre (4.1); P#0.05. The seasonal silverleaf whitefly nymphs per cm<sup>2</sup> of leaf mean for the non-treated control (11.1) was greater than the seasonal mean for Admire 2F (4.7) but was not greater than any of the other treatments.

**Investigators Name(s):** Eric T. Natwick

**Affiliation & Location:** University of California Cooperative Extension, University of California Desert Research and Extension Center, 1050 E. Holton Road, Holtville, CA 92250

**Research & Implementation Area:** Section C: Chemical Control, Biopesticides, Resistance Management, and Application Methods

**Dates Covered by Report:** March 2000 through July 2000

### **Insecticides Efficacy against Silverleaf Whitefly In Spring Planted Cantaloupe Melons, 2000.**

A stand of Cantaloupe melons, var. Topmark, was established at UC Desert Research & Extension Center 23 March 2000. Nine insecticide treatments and a non-treated control were replicated four times in a randomized complete design experiment. Insecticide treatments were as follows: Platinum 2 SC applied via drip irrigation at rates of 0.14 and 0.17 lb ai/acre and Admire 2 F was applied through the drip irrigation at 0.25 lb ai/acre, but was not followed by foliar sprays, Admire 2 F was applied through the drip irrigation at 0.25 lb ai/acre followed by foliar sprays treatments of LQ 215 (insecticidal soap) at 0.4% solution, foliar sprays of LQ 215 at 0.4%, foliar sprays of Capture 2 EC at 0.1 lb ai/acre, foliar sprays of Capture 2 EC at 0.1 lb ai/acre plus LQ 215 at 0.4% 3 EC, foliar sprays of Capture 2 EC at 0.1 lb ai/acre alternating with LQ 215 at 0.4% 3 EC, and foliar sprays of Actara 25 WG at 0.063 lb ai/acre. Drip irrigation insecticides treatments were applied 29 March. Foliar spray insecticide treatments were applied 19 & 26 April, 3 & 10 May. Silverleaf whitefly, *Bemisia argentifolii*, were sampled by counting adults on the fourth leaf from the terminal of the main stem cane from ten plants at random in each plot via the leaf turn method and whitefly nymphs were counted on 1.65 cm<sup>2</sup> leaf disks from ten crown leaves extracted from randomly selected melon plants in each plot. Adult silverleaf whitefly and nymphs were sampled on the following dates: 11, 17 & 24 April, 1, 8, 15, 22 & 31 May, 2000.

Adult whitefly population levels were suppressed by both rates of Platinum 2 SC and Admire 2F for 5 weeks following drip irrigation application. Adult whitefly population levels were suppressed by all foliar insecticide spray applications except LQ 215 used alone. Silverleaf whitefly nymphal population levels were suppressed by Platinum 2 SC treatments and by Admire 2 F for 9 weeks following drip irrigation application. Silverleaf whitefly nymphal population levels were suppressed by all foliar treatments except LQ 215 used alone and LQ 215 alternating with Capture 2 EC.

**Investigators Name(s):** Eric T. Natwick<sup>1</sup> and T. J. Henneberry<sup>2</sup>

**Affiliation & Location:** <sup>1</sup> UC Cooperative Extension, UC Desert Research and Extension Center, Holtville, CA and USDA-ARS, <sup>2</sup> Western Cotton Research Laboratory, Phoenix, AZ

**Research & Implementation Area:** Section C: Chemical Control, Biopesticides, Resistance Management, and Application Methods

**Dates Covered By Report:** March 2000 through December 2000

### **Insecticides Efficacy Against Silverleaf Whitefly In Cotton, 2000**

A stand of cotton, var. DPL 5415, was established at UC Desert Research & Extension Center 27 March 2000. Seven insecticide treatments and a non-treated control were replicated four times in a randomized complete design. Insecticide treatments were as follows: Assail 70 WP (acetamiprid) at 0.075 and 0.1 lb ai/acre, Actara 25 WP (thiamethoxam) at 0.05 and 0.06 lb ai/acre, Provado 1.6 F (imidacloprid) at 0.05 lb ai/acre, Danitol 2.4EC (fenpropathrin) + Orthene 97S (acephate) at 0.2 and 0.5 lb ai/acre, and Thiodan 3 EC (endosulfan) at 1.0 lb ai/acre. Silverleaf whitefly adults were sampled from ten plants at random in each plot via the leaf turn method using the fifth main stem leaf from the terminal. Silverleaf whitefly nymphs were counted on 1.65 cm<sup>2</sup> leaf disks from 5th position, main-stem terminal leaves extracted from ten randomly selected plants in each plot. Whitefly adults and nymphs were sampled on 6, 13, 20 & 27 June, 4, 11, 18 & 25 July, 1, 8 & 15. Seed cotton was hand picked from 0.002 acre per plot and seed cotton yield data were recorded. Seed cotton samples were ginned at the USDA-ARS Western Cotton Research Laboratory in Phoenix, AZ and lint samples were sent to the USDA/ARS Cotton Quality Research Station in Clemson, SC for stickiness and sugar analysis.

There were no differences among the treatments for silverleaf whitefly means on any of the sampling dates except 25 July when Thiodan 3 EC at 1.0 lb ai/acre, Assail 70 WP at 0.1 lb ai/acre and Danitol 2.4 EC + Orthene 97S had adult means lower than the non-treated control, P# 0.05. There were no differences among the treatments for silverleaf whitefly seasonal means. Assail 70 WP at 0.1 lb ai/acre gave the best control of silverleaf whitefly nymphs through the season followed by Assail 70 WP at 0.075 and Danitol 2.4 EC + Orthene 97S and all three of these treatments had silverleaf whitefly seasonal means lower than the non-treated control. The seasonal means of silverleaf whitefly nymphs for both rates of Actara 25 WP, for Provado 1.6 F and for Thiodan 3 EC were not different from the non-treated control.

**Investigators Name(s):** Eric T. Natwick<sup>1</sup>, T. J. Henneberry<sup>2</sup> and Brian D. Deeter<sup>3</sup>

**Affiliation & Location:** <sup>1</sup> UC Cooperative Extension, UC Desert Research and Extension Center, Holtville, CA; USDA-ARS, <sup>2</sup> Western Cotton Research Laboratory, Phoenix, AZ; and <sup>3</sup>Aventis Crop Science, Auberry, CA

**Research & Implementation Area:** Section C: Chemical Control, Biopesticides, Resistance Management, and Application Methods

**Dates Covered By Report:** March 1997 through December 2000

### **Comparison of Neonicotinoid Insecticides with Silverleaf Whitefly Insecticide Standards for Cotton**

Silverleaf whitefly insecticide efficacy research trials were conducted during the cotton seasons of 1997 through 2000 at the University of California Desert Research and Extension Center in the Imperial Valley, CA to evaluate neonicotinoid insecticides and standard insecticides for control of silverleaf whitefly in cotton. Neonicotinoid insecticidal compounds, acetamiprid (Assail®), imidacloprid (Provado®), and thiamethoxam (Actara®) were compared to the standard whitefly insecticide fenpropathrin (Danitol®) in a tank mixture with an organophosphate, acephate (Orthene®) and compared to a cyclodiene compound endosulfan (Thiodan® or Phaser®) for control efficacy of whitefly adults, eggs and nymphs. Cotton stands, var. DPL 5415, were established at UC Desert Research and Extension Center in March for each year of study for the establishment of silverleaf whitefly insecticide efficacy trials. Each year the insecticides treatments and non-treated controls were replicated four times in randomized complete design experiments. Plots measured 15 m long and 8 m wide.

During four years of study, lint yields were not often different between the pyrethroid standard treatment (Danitol® 2.4 EC + Orthene® 90S) and the neonicotinoid treatments (Assail®, Actara® and Provado®). Treatments resulting in lower numbers of whitefly adults, eggs, and nymphs generally produced higher values of seed cotton pounds per acre and lint pounds per acre. In these experiments other factors that could influence yield included relative susceptibility of western flower thrips, cotton leafperforator and *Empoasca* sp. leafhoppers to the various insecticides. Insecticides in these studies vary in their spectra of activity. Danitol®, Orthene® 90S and endosulfan are active against a broad range of cotton insect pests and Assail®, Actara®, and Provado® have narrower ranges of activity.

The neonicotinoid insecticides provide silverleaf whitefly control in cotton at levels similar to the pyrethroid plus organophosphate standard Danitol® + Orthene®. The 0.06 lb ai/acre rate of Actara® and the 0.01 lb ai/acre and 0.075 lb ai/acre rates of Assail® 70 WP, and Danitol® + Orthene® maintain similar levels of silverleaf whitefly adult, egg, and nymph throughout the cotton season. The neonicotinoid insecticide, Assail®, and Danitol® plus Orthene® treatments provided the highest levels of control for silverleaf whitefly.



**Investigators Name(s):** Eric T. Natwick

**Affiliation & Location:** University of California Cooperative Extension, University of California Desert Research and Extension Center, 1050 E. Holton Road, Holtville, CA 92250

**Research & Implementation Area:** Section C: Chemical Control, Biopesticides, Resistance Management, and Application Methods

**Dates Covered by Report:** March 2001 through July 2001

### **Insecticides Efficacy against Silverleaf Whitefly in Cantaloupe Melons, 2001**

A stand of Cantaloupe melons, var. Topmark, was established at UC Desert Research & Extension Center 29 March 2001. Seven insecticide treatments and an untreated control were replicated four times in a randomized complete design experiment. The following insecticide treatments were applied as foliar sprays: Calypso 4 SC (clothianidin) at 0.094 lb ai/acre, Assail 70 WP (acetamiprid) at 0.05 lb ai/acre, Actura 25 WG (thiamethoxam) at 0.047 lb ai/acre and at 0.63 lb ai/acre, Danitol 2.4EC (fenpropathrin) at 0.2 lb ai/acre plus Thiodan 3 EC (endosulfan) at 0.5 lb ai/acre, Capture 2 EC (bifenthrin) at 0.08 lb ai/acre plus Thiodan 3 EC at 0.5 lb ai/acre, and Knack 0.86 EC (pyriproxyfen) at 0.054 lb ai/acre. Foliar insecticide spray treatments were applied 18 May, 2001. Silverleaf whitefly, *Bemisia argentifolii*, were sampled by counting adults on the fourth leaf from the terminal of the main stem cane from ten plants at random in each plot via the leaf turn method and whitefly nymphs were counted on 1.65 cm<sup>2</sup> leaf disks from ten crown leaves extracted from randomly selected melon plants in each plot. Adult silverleaf whitefly and nymphs were sampled on the following dates: 14, 21, 24, 29, 31 May, and 4 June 2001.

The adult whitefly mean for the untreated control was significantly greater than the means for all other treatment on 24 May, but the adult whitefly mean for the untreated control was not different from other treatments on all other sampling dates;  $P \leq 0.05$ . The whitefly nymph means for all insecticide treatments were significantly lower than the untreated control on 21, 24 and 29 May. Seasonal means for whitefly nymphs were significantly lower for all insecticide treatments compared to the untreated control.

**Investigator's Name:** Eric T. Natwick

**Affiliation & Location:** UC Cooperative Extension, UC Desert Research and Extension Center, Holtville, CA

**Research & Implementation Area:** Section C: Chemical Control, Biopesticides, Resistance Management, and Application Methods

**Dates Covered By Report:** September 2001 through January 2002

### **Insecticide Efficacy for Silverleaf Whitefly Control In Cauliflower, 2001**

Cauliflower var. Minuteman was transplanted into plots at UC Desert Research & Extension Center 21 September 2001. Five insecticide treatments and an untreated control were replicated five times in a randomized complete design experiment. The insecticide treatments Actara 25 WG (thiamethoxam) at 0.05 lb and 0.06 lb ai/acre, Fulfill 50 WG (pymetrozine) at 0.086 lb ai/acre, Thiodan 3 EC (endosulfan) at 0.75 lb ai/acre, and Provado 1.6 F (imidacloprid) at 0.05 lb ai/acre were applied as foliar sprays on 16 and 26 October and on 9 November 2001. Silverleaf whitefly, *Bemisia argentifolii*, were sampled by counting adults via leaf turn of basal leaves on ten plants at random from each plot and nymphs were counted on 1.65 cm<sup>2</sup> of leaf surface from basal leaves of ten plants at random from each plot on 2, 9, 16, 23, 30 October, 6, 14, 20 November 2001. Yield data as numbers of marketable cauliflower heads per 0.001 acres and weight as pounds of marketable cauliflower heads per 0.001 acres were collected 15 January 2002.

The post treatment mean for silverleaf whitefly adults for the untreated control (52.5) was significantly greater ( $P < 0.05$ ) than the seasonal means for all insecticide treatments except the mean for Fulfill (48.7). The post treatment adult whitefly mean for Provado 1.6 F (27.1) was significantly greater than the means for Actara 25 WG at 0.05 lb ai/acre (21.9) and 0.06 lb ai/acre (21.5). The post treatment mean for silverleaf whitefly nymphs per cm<sup>2</sup> of leaf for the untreated control (24.9) was significantly greater than the seasonal means for all insecticide treatments except the mean for Fulfill (20.7). The post treatment mean for silverleaf whitefly nymphs per cm<sup>2</sup> of leaf for Fulfill was significantly greater than all other insecticide treatments with post treatment means for whitefly nymphs ranging from 15.8 for Actara 25 WG at 0.05 lb ai/acre to 12.8 for Thiodan 3 EC at 0.75 lb ai/acre. There were no significant differences among treatment means for numbers of marketable cauliflower heads per 0.001 acre (10.5) nor for pounds marketable cauliflower heads per 0.001 acre.

**Investigator's Name:** Eric T. Natwick

**Affiliation & Location:** UC Cooperative Extension, UC Desert Research and Extension Center, Holtville, CA

**Research & Implementation Area:** Section C: Chemical Control, Biopesticides, Resistance Management, and Application Methods

**Dates Covered By Report:** September 2001 through January 2002

### **Neonicotinoid Insecticide Efficacy for Silverleaf Whitefly Control In Cauliflower, 2001**

Cauliflower var. Minuteman was transplanted into plots at UC Desert Research & Extension Center 21 September 2001. Seven insecticide treatments and an untreated control were replicated four times in a randomized complete design experiment. The insecticide treatments Admire 2F (imidacloprid) at 0.156 lb and 0.25 lb ai/acre, and Platinum 2 SC (thiamethoxam) at 0.071 lb, 0.11 lb, 0.125 lb, 0.141 lb, and 0.172 lb ai/acre were injected 3 inches below the seed-line on 20 September 2001. Silverleaf whitefly, *Bemisia argentifolii*, were sampled by counting adults via leaf turn of basal leaves on ten plants at random from each plot and nymphs were counted on 1.65 cm<sup>2</sup> of leaf surface from basal leaves of ten plants at random from each plot on 2, 9, 16, 23, 30 October, 6, 14, 20 November 2001. The plant height (cm) was measured 13 December and yield data as numbers of marketable cauliflower heads per 0.001 acres and weight (lb) of marketable cauliflower heads per 0.001 acres were collected 15 January 2002.

The seasonal silverleaf whitefly adult mean for the untreated control (48.9) was significantly greater ( $P = 0.05$ ) than the seasonal means for all insecticide treatments. Admire 2F at 0.25 lb ai/acre had a seasonal mean for adult whitefly (12.2) that was significantly lower than all other insecticide treatments except Admire 2F at 0.156 lb (14.5) ai/acre and Platinum 2 SC at 0.172 lb ai/acre (13.5). The seasonal mean for silverleaf whitefly nymphs per cm<sup>2</sup> of leaf for the untreated control (11.1) was significantly greater ( $P = 0.05$ ) than the seasonal means for all insecticide treatments. Admire 2F at 0.25 lb ai/acre had a seasonal mean for whitefly nymphs (1.9) that was significantly lower than all other insecticide treatments except Admire 2F at 0.156 lb (3.0) ai/acre and Platinum 2 SC at 0.172 lb ai/acre (2.0).

The untreated control plants mean height in cm (66.7) was significantly lower than the means for all insecticide treatments. The mean plant height (72.2 cm) for Platinum 2 SC at 0.071 lb ai/acre was significantly lower than the means for all other insecticide treatments, except Platinum 2 SC at 0.125 lb ai/acre with a plant height mean of 74.5 cm. The plant mean heights for all other insecticide treatment did not differ significantly and ranged from 76.6 cm for Platinum at 0.11 lb ai/acre to 78.2 cm for Admire 2F at 0.25 lb ai/acre.

The mean number of marketable cauliflower heads per 0.001 acre (10.5) was significantly lower than the mean numbers of marketable heads for Platinum 2 SC at 0.141 lb ai/acre (18.0) and for Platinum 2 SC at 0.172 lb ai/acre (19.0). The mean pounds marketable cauliflower heads per 0.001 acre (22.0) was significantly lower than the mean pounds of marketable heads for all insecticide treatments except Platinum 2 SC at 0.071 lb ai/acre (28.6) and for Platinum 2 SC at 0.125 lb ai/acre (25.8). The means for marketable cauliflower heads for all other insecticide treatment did not differ significantly and ranged from a mean of 36.6 lb per 0.001 acres for Platinum 2 SC at 0.11 lb ai/acre to 45.6 lb per 0.001 acres for Platinum 2 SC at 0.141 lb ai/acre.

**Investigators Name(s):** Eric T. Natwick<sup>1</sup> and T. J. Henneberry<sup>2</sup>

**Affiliation & Location:** <sup>1</sup> UC Cooperative Extension, UC Desert Research and Extension Center, Holtville, CA and USDA-ARS, <sup>2</sup> Western Cotton Research Laboratory, Phoenix, AZ

**Research & Implementation Area:** Section C: Chemical Control, Biopesticides, Resistance Management, and Application Methods

**Dates Covered by Report:** March 2001 through December 2001

### **Insecticides Efficacy Against Silverleaf Whitefly In Cotton, 2001**

A stand of cotton, var. DPL 5415, was established at UC Desert Research & Extension Center 21 March 2001. Seven insecticide treatments and an untreated control were replicated four times in a randomized complete design. The following insecticide treatments were applied as foliar sprays: Assail 70 WP (acetamiprid) at 0.075 and 0.1 lb ai/acre, Danitol 2.4EC (fenpropathrin) at 0.2 lb ai/acre plus Orthene 97 (acephate) at 0.5 lb ai/acre, Danitol 2.4EC at 0.2 lb ai/acre plus Thiodan 3 EC (endosulfan) at 0.5 lb ai/acre, Knack 0.86 EC (pyriproxyfen) at 0.054 lb ai/acre, and Novaluron 0.83 EC (benzoylurea) at 0.013 lb ai/acre and at 0.026 lb ai/acre. Foliar insecticide spray treatments were applied 11 July, 7, 21, 30 August 2001. Silverleaf whitefly adults were sampled from ten plants at random in each plot via the leaf turn method using the fifth main stem leaf from the terminal. Silverleaf whitefly nymphs were counted on 1.65 cm<sup>2</sup> leaf disks from fifth position, main-stem terminal leaves extracted from ten randomly selected plants in each plot. Whitefly adults and nymphs were sampled on 10, 16, 20, 23, 26, 31 July, 6, 13, 20, 27 August, 4, 10, 17 September 2001. Seed cotton was hand picked from 0.002 acre per plot and seed cotton yield data were recorded. Seed cotton samples were ginned at the USDA-ARS Western Cotton Research Laboratory in Phoenix, AZ and lint samples were sent to the USDA/ARS Cotton Quality Research Station in Clemson, SC for stickiness and sugar analysis.

Seasonal means for whitefly adults for all of the insecticide treatments were significantly lower than the untreated control except the Knack 0.86 EC treatment,  $P \leq 0.05$ . The Assail 70 WP treatments and the Danitol 2.4 EC + Orthene 97S treatment had seasonal means for whitefly adults that were significantly lower than the means for the Novaluron insecticide treatments. Seasonal means for whitefly nymphs for all of the insecticide treatments were significantly lower than the untreated control treatment. The Assail 70 WP treatments had seasonal means for whitefly nymphs that were significantly lower than the means for all other insecticide treatments. The seasonal mean for whitefly nymphs was significantly lower for the Knack 0.86 EC treatment compared to the means for all other insecticide treatments except the Assail 70 WP treatments. The Danitol 2.4EC plus Orthene 97 treatment had a seasonal mean for whitefly nymphs that was significantly lower than the means for Danitol 2.4EC plus Thiodan 3 EC and the Novaluron 0.83 treatments. There were no differences among the treatments for pounds of seed cotton per acre.

**Investigator's Name(s):** D. R. Seal, M. J. Davis and R. T. McMillan, Jr.

**Affiliation & Location:** University of Florida-IFAS, Tropical Research and Education Center, Homestead, FL 33031.

**Research & Implementation Area:** Section C: Chemical Control, Biopesticides, Resistance Management, and Application Methods

**Date Covered by the Report:** 2000

**Effectiveness of Neemix in Managing Silverleaf Whitefly (SLW) and Tomato Yellow Leaf Curl Virus in Tomatoes, 2000**

Tomato seedlings were planted in a Rockdale fine gravelly loam with pH 7.0 – 7.3 on 12 May 2000 at Homestead, Florida. Seedlings were placed 18 in. apart within rows and 6 ft. between rows and were drip irrigated. Plots were arranged in a randomized complete block design with four replications. Treatments evaluated were: 1) Admire at 8 ozs. as soil drench at planting followed by weekly foliar application of Neemix at 4 ozs./A; in this treatment plants were treated with one application of Neemix at 4 ozs./A one week before planting the plants. 2) Admire at 8 ozs. as soil drench at planting followed by weekly foliar application of Neemix at 4 ozs./A; 3) Admire at 8 ozs. as soil drench at planting followed by two weekly foliar application of Neemix at 8 ozs./A and then followed by weekly foliar application of Neemix at 4 ozs./A ; 4) Admire 8 ozs./A as a soil drench at planting followed by Knack at 8 ozs. in rotation with Applaud at 8 ozs.; 5) Weekly foliar application of Knack at 8 ozs. followed by Applaud at 8 ozs.; 6) Admire at 16 ozs. as soil drench at planting; 7) Admire at 8 ozs. as soil drench at planting; 8) Neemix at 8 ozs./A as weekly foliar application; 9) A nontreated check.

Application of foliar treatments were made by using a backpack sprayer with two nozzles/row at 30 psi delivering 70 gpa. Weekly application of treatments were made on four dates -21, 28, May 2000, 4 and 12 June, 2000. Application of insecticides was terminated when about 80% of the plants in all treatment plots showed TYLCV symptoms. Evaluation of treatments was made 24 h after each application by counting SLW adults on one leaf/plant of randomly selected five plants per treatment plot. Finally, treatments were evaluated by harvesting all marketable fruits from all plants in a 25 ft. long middle row of each treatment plot.

Silverleaf whitefly adults were significantly fewer in all plants treated with various insecticides when compared with the nontreated control. This reduction in the numbers of SLW was consistent in all sampling dates during the present study. However Treatment 3, where Admire at the rate of 8 ozs. was drenched at planting followed by two weekly applications of Neemix at the rate of 8 ozs./A and then followed by weekly application of Neemix at 4 ozs./A, provided highest reduction of SLW. This reduction in the number of SLW was significantly different from Admire at the rate of 8 ozs./A. Marketable fruits were significantly greater in plants treated with the Admire-Knack-Applaud combination which did not differ from Admire alone and Admire-Neemix treated plants. In Admire-Neemix treated plants where Neemix at 8 ozs./A was applied two times followed by Neemix at the rate of 4 ozs./A, provided the second highest marketable yield next to Admire-Knack-Applaud treatment.

This study demonstrates that Neemix in combination with Admire provides significant reduction of SLW and its transmitted Tomato Yellow Leaf Curl Virus.

**Investigator's Name(s):** D. R. Seal and R. T. McMillan, Jr.

**Affiliation & Location:** University of Florida-IFAS, Tropical Research and Education Center, Homestead, FL 33031.

**Research & Implementation Area:** Section C: Chemical Control, Biopesticides, Resistance Management, and Application Methods

**Date Covered by the Report:** 2000

### **Control of the Silverleaf Whitefly (SLW) and its Associated Bean Golden Mosaic Virus in Beans, 2000**

Snap bean was planted on 15 March, 2000 into a Rockdale fine gravelly loam with pH 7.0-7.3 pH at the Tropical Research and Education Center research plot. Treatment plots consisted of three 30-ft. long rows which were separated by 30-in space. Eight treatments were arranged in a randomized complete block design with four replications. A 5-ft. planted area separated the replicates. Treatments evaluated were: 1) Actara at 9 & 11 ozs./A as soil drench at planting; 2) Admire was drenched at planting at the rate of 24 ozs./A followed by four weekly applications of Knack at 8 & 10 ozs./A; 3) Admire was drenched at planting at the rate of 24 ozs./A followed by weekly foliar application of Knack at 8 ozs. in rotation with Applaud at 8 ozs; 4) weekly foliar application of Provado at 3.75 ozs/A; 5) weekly foliar application of Rimon at 6 ozs./A, and 6) a nontreated control. Evaluations of insecticides against the silverleaf whitefly were made by randomly collecting 5 leaves, one leaf/plant, 24 h after each application. Numbers of silverleaf whitefly eggs and nymphs were recorded on randomly selected 3 one cm. square areas per leaf. Also, numbers of silverleaf whitefly adults were recorded 24 h after each foliar application by counting all adults on a leaf/plant of five randomly selected plants. Effects of insecticide treatments on the management of BGMV were determined by counting all healthy plants in the center row. All plants from 10-foot area in the center of the middle row were collected and weighed to determine effect of insecticide treatments on plant quality. Finally, 10 plants were collected from the center row of each treatment plot and number of fruits and weight of those fruits were recorded.

Silverleaf whitefly adults were significantly fewer on plants when treated with Actara at 11 ozs./A than on plants treated with other insecticide treatments. Silverleaf whitefly eggs were significantly fewer in all treatments when compared with the nontreated control. Silverleaf whitefly nymphs were significantly fewer on plants treated with all insecticides except in Rimon - treated plants. Lowest number of nymphs was observed in plants treated with Actara at 11 ozs./A.

Percentage healthy plants, free from BGMV, were significantly higher when they had been treated with Actara at 11 ozs./A. Provado and Rimon did not provide any control of Bean Golden Mosaic Virus on beans. Treatments did not show pronounced effects on weights and heights of the treated plants. Numbers of marketable fruits and fruit-weights were significantly higher when plants were treated with Actara or Admire followed by Knack and Applaud. Provado and Rimon did not increase the numbers of fruits or fruit weight. Although numbers of fruits increased on plants treated with Admire at 24 ozs. and Knack at 8 ozs., fruit weights did not increase. This inconsistency in fruit weights was caused by variable drying of fruits in the field environment before harvest.

Overall, Actara and Admire applied as soil drenches provided significant reduction of Bean Golden Mosaic Virus incidence. However, Admire in soil does not provide significant control of silverleaf whitefly for more than 4 weeks. After this critical period, Admire application should be followed by Knack to suppress both silverleaf whitefly and Bean Golden Mosaic Virus

**Investigator's Name(s):** D. R. Seal

**Affiliation & Location:** University of Florida-IFAS, Tropical Research and Education Center, Homestead, FL 33031.

**Research & Implementation Area:** Section C: Chemical Control, Biopesticides, Resistance Management, and Application Methods

**Date Covered by the Report:** 1998

### **Effectiveness of Mycotrol in Controlling Silverleaf Whitefly**

'Pod Squad' beans were planted in a Rockdale fine gravelly loam with pH 7.0-7.3. Plant spacing was 2.5 cm in the row and 0.9 m between rows. Atrazine was broadcast in the furrow at planting at the rate of 0.5 kg/ha to control weeds in the rows, and the field was cultivated biweekly to clean weeds between the rows. Fertilizer was applied in kg/ha at 366 N, 732 P, 732 K on two dates at planting and 2 weeks after planting. The plots were irrigated by overhead sprinkler system at weekly intervals at approximately 5 cm. depth. Plots were four rows each 30 ft long. Six treatments were arranged in a randomized complete blocks with 4 replications. A nontreated 3 m wide section separated the plots. Treatments evaluated were: 1) Fipronil at 0.05 and 0.075 lb. a.i./A; 2) Provado 1.6F at 0.05 lb. a.i./A; 2) Mycotrol at 2 qts./A; 3) Provado at 0.025 lb. a.i./A in combination with 2 qts./A of Mycotrol; and 4) a nontreated control. Treatments were applied weekly using a backpack sprayer with two nozzles per row. During the spray of insecticides, the backpack sprayer was maintained at 30 psi. delivering 70 to 100 gallons/A. Treatments were evaluated by recording *Bemisia argentifolii* adults on randomly selected 10 leaves, one leaf/plant, per treatment plot. This was conducted by gently turning the leaves. In addition, *B. argentifolii* eggs and nymphs were recorded on randomly selected ten leaves per plant, one leaf/plant. These leaves were incised and placed in a plastic bag to transport them to the laboratory. Leaves were checked using a binocular microscope (10X) to record number of eggs and nymphs.

All treatments significantly reduced *B. argentifolii* eggs, nymphs and adults when compared with the nontreated control. Fipronil and Provado significantly reduced *B. argentifolii* adults and eggs after the first application. This reduction in the numbers of *B. argentifolii* eggs and nymphs was consistent in the subsequent weeks of study. Pupal populations did not differ among treatments after the first application. Pupal population on Fipronil and Provado treated plants decreased significantly when compared with the nontreated control. Among treatments, Mycotrol treated plants had more whiteflies than the others; although significantly different from nontreated control. Addition of Mycotrol with Provado did not improve management of silverleaf whitefly on beans

**Investigator's Name(s):** D. R. Seal

**Affiliation & Location:** University of Florida-IFAS, Tropical Research and Education Center, Homestead, FL 33031.

**Research & Implementation Area:** Section C: Chemical Control, Biopesticides, Resistance Management, and Application Methods

**Date Covered by the Report:** 1998

#### **Effectiveness of Fipronil in Controlling Silverleaf Whitefly (SLW) on Beans**

'Pod Squad' beans were planted on 2 January 1998. Crop was maintained following recommended cultural practices. Plots were 2 rows by 30 feet long. Treatment plots were arranged in a randomized complete block design with four replications. Treatments evaluated in this study were: 1) Admire 2F (32 ozs./A as soil drench); 2) Provado 1.6F (3.75 ozs./A as foliar); 3) three rates of Fipronil 70WP (0.044, 0.05 & 0.075 lb. [a.i.]/A; and 4) a non-treated control. Admire was drenched in a 1 in.-deep furrow 4 in. apart from the seed row 3 weeks after planting. Provado and Fipronil were applied on foliage on 1/12, 1/19, 1/29 and 2/9/98 depending on the abundance of *Bemisia argentifolii* on the treated plants. Foliar application of insecticides was made by using a backpack sprayer with two nozzles per row at 30 psi. delivering 70 gpa. Prespray samples were collected once on 8 January and postspray samples were collected on 14, 20, 22, 27, 30 January and 24 Feb. 1998. Treatments were evaluated by counting numbers of *Bemisia argentifolii* adults and nymphs on randomly 5 plants, one leaf/plant, from the two center rows of each treatment plot on each sampling date. Admire, Provado and Fipronil significantly reduced SLW adults and nymphs on beans when compared with the nontreated control plants. Mean numbers of SLW nymphs and adults did not differ among rates. Use of Admire, Provado and Fipronil in rotation in a management program may provide an excellent control of SLW.



## **Section C Research Summary - Chemical Control, Biopesticides, Resistance Management and Application Methods.**

John Palumbo

### **Current Research Activities**

Chemical control continues to be a very important part of silverleaf whitefly management. Area-wide suppression and management of whiteflies has occurred largely due to the successful deployment of various chemical control approaches. Development and registration of new insecticides chemistries, insect growth regulators and biopesticides continues to be important and will continue to provide producers with a number of pest management options. In addition, industry, government and university scientists continue to collaboratively develop and refine integrated and resistance management programs within a number of cropping systems. Over the past 10 years, these efforts have resulted in a reduction and harmonization of chemical use, and a return to economic crop production in the United States.

Progress was made on the evaluation of systemic neonicotinoid insecticides, insect growth regulators (IGRs) and biopesticides. A significant amount of work was reported on the development of use patterns for the neonicotinoid class of chemistry. Formulations of thiamethoxam and acetamiprid applied as sprays to foliage and injected into the soil appear promising and may provide alternatives to prophylactic uses of imidacloprid in vegetables and melons crops. Use of IGRs remains an important chemical control approach as illustrated by their continued success in desert cotton crops and melons, and the development of action thresholds in tomatoes. Over the past several years, the uses of systemic insecticides and insect growth regulators have led to a lessening of crop losses due to whitefly while opening the door to a greater role for biological control. A considerable amount of work was conducted across disciplines to examine the impacts of several IGRs on whitefly parasitoids and predators which has led to the understanding of "bioresidual". Workers continued to evaluate several biopesticides, azadirachtin, entomopathogenic fungi, oils, and sucrose esters for whitefly control and compatibility with natural enemies in integrated management programs.

Because growers rely heavily on insecticides for whitefly control, monitoring and management of insecticide resistance continues to be a primary focus. Studies focusing on our basic understanding of resistance have examined the genetics and biochemistry of resistance and cross-resistance. Baselines continue to be established for insect growth regulators and imidacloprid across many growing regions. Continued work on new chemistries is important in detecting shifts early in a products use. The

establishment of resistant colonies to various classes of insecticides has increased providing resistant specimens for use in resistance studies. In particular, genetic and biochemical studies have been focused on the role of MFO-based metabolism and esterases in resistance to imidacloprid. The metabolic role of synergists DEF and piperonyl butoxide to enhance the toxicity of imidacloprid was also investigated. A significant effort has been made by several laboratories to examine the cross-resistance patterns between three neonicotinoids, acetamiprid, thiamethoxam and imidacloprid. Although no definitive cross-resistance patterns were observed in a number of whitefly populations, the development of cross-resistance with extensive use is a possibility because of similarity in structure. Consequently, in order to avoid high selection pressure on any one chemical from this group, there is an urgent need for integrating the neonicotinoids into a diversified management program.

The development of resistance management programs at the grower level has been a priority in the desert growing regions of the southwestern U.S. Significant progress was made in Arizona to implement a cross commodity IRM approach for the shared use of buprofezin and neonicotinoids among commodities such as cotton, melons and vegetables. It is too early to determine the success of the Arizona program, but workers continue to collect bioassay data on insecticide susceptibility and providing this information to producers on a regional basis. Within the next few years, scientists and extension workers are confident that based on the above science, integrated programs in numerous cropping systems will be in place that optimizes neonicotinoid, IGR and biopesticide efficacy while minimizing their impacts on biological control efforts.

**Table C. Chemical Control, Biopesticides, Resistance Management, and Application Methods.**

Research Approaches <sup>a</sup>	Year 1 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Improve insecticide efficacy:</b>				
<b>Develop, test, and assist in the registration of insecticides, biorationals, and natural products.</b>	Develop new chemistries and natural products. Develop improved techniques for evaluating efficacy of insecticides. Support registration of desirable new products by providing information to regulatory agencies.	X		New studies reported in this area in 1997 = 39. New biopesticides like <i>Petunia</i> extract and <i>Melia</i> extract tested. New biorationals tested or reported on included benzyl phenal urea naphaphenol and antibiotics (to act against symbiotic bacteria).
<b>Develop improved methods of application including formulation and delivery of materials to improve control.</b>	Develop spray systems for better underleaf coverage. Evaluate rates, timing, placement in relation to efficacy. Consider formulation, UV protectants, and other means to improve efficacy. Develop improved methods to evaluate application efficacy. Field test under commercial conditions for technology transfer.	X		New studies = 10. Thermal fogger evaluated for greenhouse use. However, a comparison of five-sprayers in the field trials showed no significant differences between hydraulic, air-assist and electrostatic technology.
<b>Conserve insecticide efficacy:</b>				
<b>Relate action thresholds to insecticide usage patterns.</b>	Refine action thresholds based on insecticide efficacy and input from other control strategies.	X		New studies = 8. Cost-benefit study of IPM system in cotton. Life table approach to evaluate impact of mortality factors initiated. Training effort to extend threshold information to growers in Arizona.
<b>Elucidate the role of genetic, biochemical and ecological factors leading to insecticide resistance.</b>	Establish whitefly strains resistant and susceptible to various classes of insecticide. Conduct studies to determine the genetics and biochemistry of resistance and cross resistance to different classes of insecticide.	X		New studies = 4. Imidacloprid binding site elucidated. Studies completed on stability of resistance in <i>Bemisia</i> including agricultural and ecological factors.

**Table C. Chemical Control, Biopesticides, Resistance Management, and Application Methods. (Continued)**

Research Approaches <sup>a</sup>	Year 1 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Improve insecticide efficacy:</b>				
<b>Improve techniques for monitoring resistance.</b>	Establish baseline data on toxogenic responses of whitefly populations to new insecticides.	X		New studies = 9. Bioassays developed for testing sensitivity to imidacloprid. Baseline data obtained on sensitivity to imidacloprid and IGRs pyriproxyfen and buprofazin.
<b>Develop, evaluate and refine resistance management systems.</b>	Evaluate the effects of mixtures and rotations of new and old chemistries to mitigate selection for resistance.	X		New studies = 14. Area-wide plans for management of resistance refined in Arizona and California. Large-scale trials of resistance management strategies conducted.
<b>Integrate chemical control with other tactics.</b>	Evaluate selectivity of synthetic insecticides and natural products to key whitefly natural enemies.	X		New studies = 10, including laboratory and field studies on compatibility with whitefly natural enemies. Also a study on effects of pyrethroids on antibiotic factors bred into crops.

<sup>a</sup> See Table A for complementary research on thresholds.

<sup>a</sup> See Table B for complementary research on virus/vector interactions.

<sup>a</sup> See Table D for complementary research on biological control.

<sup>b</sup> See Table E and F for complementary research on systems management.

**Table C. Chemical Control, Biopesticides, Resistance Management, and Application Methods.**

Research Approaches <sup>a</sup>	Year 2 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Improve insecticide efficacy:</b>				
<b>Develop, test, and assist in the registration of insecticides, biorationals, and natural products.</b>	Determine new modes of action of effective materials. Elucidate biochemical pathways of synthesis and degradation of natural products.	X		(1) Section 3 registration of IGRs. (2) Section 18's supported, acetamiprid summer '99. (3) progress evaluating soil applied modes of action, sugar esters and entomopathic fungi., integration of biorationals and conventional chemistries. Need to evaluate future impact of FQPA. References 40, 69, 71, 78, 81, 103, 104, 108, 145, 165, 166, 167, 168, 180, 186, 187, 188, 212, 220, 262, 263, 273, 274, 297
<b>Develop improved methods of application including formulation and delivery of materials to improve control.</b>	Develop spray systems for better underleaf coverage. Evaluate rates, timing, placement in relation to efficacy. Consider formulation, UV protectants, and other means to improve efficacy. Develop improved methods to evaluate application efficacy. Field test under commercial conditions for technology transfer.	X		Akey's work with high PSI systems, increasing stability for azadiractin and utilizing digital photographs to evaluate application efficacy. References 47, 131, 139
<b>Conserve insecticide efficacy:</b>				
<b>Relate action thresholds to insecticide usage patterns.</b>	Refine action thresholds based on insecticide efficacy and input from other control strategies.	X		Mint sampling plan/thresholds, distribution patterns validated, References 35, 125, 212, 317
<b>Elucidate the role of genetic, biochemical and ecological factors leading to insecticide resistance.</b>	Establish whitefly strains resistant and susceptible to various classes of insecticide. Conduct studies to determine the genetics and biochemistry of resistance and cross resistance to different classes of insecticide. Evaluate the role of refuge habitats (weeds, tolerant crops, urban areas) to assure input of susceptible genes in whitefly population.	X		Resistant colonies exist to endosulfan, chlorpyrifos, imidacloprid, bifenthrin; genetic and biochemistry studies are concentrated on acetylcholinesterase (Byrne) and nicotinyls; cross resistance being studied between nicotinyls and neonicotinyls. Impact of ecological factors such as nutrition, host plant response, local cropping patterns are being studied. Role of alfalfa as a refuge has been evaluated. References 18, 22, 46,

**Table C. Chemical Control, Biopesticides, Resistance Management, and Application Methods. (Continued)**

Research Approaches <sup>a</sup>	Year 2 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Improve insecticide efficacy:</b>				
<b>Improve techniques for monitoring resistance.</b>	Establish baseline data on toxogenic responses of whitefly populations to new insecticides. Expand comparative studies of resistance levels in diverse agro-ecosystems. Evaluate relationship between monitoring results and field efficacy.	X		Baselines have and are being established for IGRs and systemic insecticides. Work in Cal. & Arizona ongoing to evaluate regional resistance management techniques which include four distinct agro-ecosystems. Work in ornamentals is increasing. Relationships between monitoring results and field failure are primarily anecdotal at this time. References 140, 227, 286, 314
<b>Develop, evaluate and refine resistance management systems.</b>	Evaluate the effects of mixtures and rotations of new and old chemistries to mitigate selection for resistance. Develop methods to evaluate and augment the beneficial influence of refuges as sources of susceptible genes to the population pool.	X		Prabhaker et al. in press. Studies to increase horizontal integration of resistance management programs are addressing influence of refuges in diverse agro-ecosystems. References 20, 47, 76, 77, 235, 245, 284, 310
<b>Integrate chemical control with other tactics.</b>	Evaluate selectivity of synthetic insecticides and natural products to key whitefly natural enemies. Test compatibility of biological control with selective synthetic or natural product insecticides as required.	X		Most efficacy trials include a compatibility evaluation, selectivity evaluations include systemics, entomopathic fungi; life tables are contributing to our understanding here. References 7, 9, 10, 11, 13, 24, 25, 27, 31, 32, 37, 38, 49, 70, 89, 90, 93, 96, 97, 98, 105, 110, 129, 147, 149, 153, 160, 169, 174, 194, 200, 247

<sup>a</sup> See Table A for complementary research on thresholds.

<sup>a</sup> See Table B for complementary research on virus/vector interactions.

<sup>a</sup> See Table D for complementary research on biological control.

<sup>b</sup> See Table E and F for complementary research on systems management.

**Table C. Chemical Control, Biopesticides, Resistance Management, and Application Methods.**

Research Approaches <sup>a</sup>	Year 3 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Improve insecticide efficacy:</b>				
<b>Develop, test, and assist in the registration of insecticides, biorationals, and natural products.</b>	Develop new chemistries and natural products. Develop improved techniques for evaluating efficacy of insecticides. Support registration of desirable new products by providing information to regulatory agencies. Determine new modes of action of effective materials. Elucidate biochemical pathways of synthesis and degradation of natural products. Same as Year 2. Evaluate the potential for transforming plants with natural product genes.	X		Section 3 registration of IGRs & Admire in additional crops, Section 18's supported, work with Rimon a new IGR, continued work with neonicotinyls as soil and foliar applications, sugar esters, azadirachtins and entomopathic fungi, toxicity of abamectin, tebufenozide, chlorfenpyr & pymetrozine evaluated, integration of biorationals and conventional chemistries, work on products that induce resistance has some promise, sampling plan in cantaloupes, life table studies, evaluated the impacts of FQPA References: 59, 60, 83, 145, 164, 167, 183, 184, 189, 192, 210, 214, 252 Tomato transformations – Florida.
<b>Develop improved methods of application including formulation and delivery of materials to improve control.</b>	Develop spray systems for better underleaf coverage. Evaluate rates, timing, placement in relation to efficacy. Consider formulation, UV protectants, and other means to improve efficacy. Develop improved methods to evaluate application efficacy. Field test under commercial conditions for technology transfer. Same as Year 2	X		Additional work with swivel nozzles, several commercial studies examining drip and at planting methods of application, ultrasonic fogging devices for greenhouses, application methods to improve efficacy of entomopathic fungi, the use of DEF & PBO to enhance efficacy and overcome resistance. References: 76, 77, 244,
<b>Conserve insecticide efficacy:</b>				
<b>Relate action thresholds to insecticide usage patterns.</b>	Refine action thresholds based on insecticide efficacy and input from other control strategies  Same as Year 2	X		Continued work on sticky cotton sampling plan, development of IGR specific action thresholds in melons, Yield, sticky cotton – whitefly relationships studied. References: 18, 95, 182

**Table C. Chemical Control, Biopesticides, Resistance Management, and Application Methods. (Continued)**

Research Approaches <sup>a</sup>	Year 3 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Elucidate the role of genetic, biochemical and ecological factors leading to insecticide resistance.</b>	Conduct studies to determine the genetics and biochemistry of resistance and cross resistance to different classes of insecticide. Evaluate the role of refuge habitats (weeds, tolerant crops, urban areas) to assure input of susceptible genes in whitefly population. Evaluate the influence of host plant on susceptibility to insecticides.	X		Genetic and biochemistry studies are concentrated on the role of MFO-based metabolism and esterases in resistance to nicotinyls and bifenthrin; cross resistance being studied with nicotinyls and neonicotinyls. Impact of ecological factors such as nutrition, host plant response, local cropping patterns are being studied. Role of alfalfa as a refuge has been evaluated. Host plant influence on efficacy has been evaluated based on seasonality, external & internal uptake, changes in metabolites and several compounds screened on various host plants. References: 12, 128, 185, 220
<b>Improve insecticide efficacy:</b>				
<b>Improve techniques for monitoring resistance.</b>	Establish baseline data on toxogenic responses of whitefly populations to new insecticides. Same as Year 1. Expand comparative studies of resistance levels in diverse agro-ecosystems. Evaluate relationship between monitoring results and field efficacy. Same as Year 2. Summarize, analyze, and produce standardized comparable monitoring systems.	X		Baselines have and are being established for IGRs and systemic insecticides. Work in Cal. & Arizona are evaluating regional resistance management techniques which include four distinct agro-ecosystems. Work in ornamentals is increasing. Relationships between monitoring results and field failure are primarily anecdotal at this time. References: 73, 74, 75

**Table C. Chemical Control, Biopesticides, Resistance Management, and Application Methods. (Continued)**

Research Approaches <sup>a</sup>	Year 3 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Develop, evaluate and refine resistance management systems.</b>	Evaluate the effects of mixtures and rotations of new and old chemistries to mitigate selection for resistance. Same as Year 1. Develop methods to evaluate and augment the beneficial influence of refuges as sources of susceptible genes to the population pool. Same as Year 2. Develop criteria for integration of successful strategies in agricultural systems. Field test resistance management systems as long range components of successful IPM.	X		Prabhaker et al. in press. Studies to increase horizontal integration of resistance management programs are addressing influence of refuges in diverse agro-ecosystems. Areawide resistance management programs in place in Cal. and Arizona.
<b>Integrate chemical control with other tactics.</b>	Evaluate selectivity of synthetic insecticides and natural products to key whitefly natural enemies. Same as Year 1. Test compatibility of biological control with selective synthetic or natural product insecticides as required. Same as Year 2. Integrate systems with host plant resistance and cultural controls	X		Most efficacy trials include a compatibility evaluation, selectivity evaluations include IGR's, neonicotinyls, entomopathic fungi; life tables are contributing to our understanding here. Integration of smooth leaf varieties of cotton with chemical control being practiced.

<sup>a</sup> See Table A for complementary research on thresholds.

<sup>a</sup> See Table B for complementary research on virus/vector interactions.

<sup>a</sup> See Table D for complementary research on biological control.

<sup>b</sup> See Table E and F for complementary research on systems management.



**Table C. Chemical Control, Biopesticides, Resistance Management, and Application Methods.**

Research Approaches <sup>a</sup>	Year 4/5 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Improve insecticide efficacy: Develop, test, and assist in the registration of insecticides, biorationals, and natural products.</b>	Evaluate the potential for transforming plants with natural product genes.	X		Several new registrations were granted for a number of products including: sucrose esters, neem based products, Applaud (Courier), Esteem and Platinum, Actara and Assail. Work to determine efficacy and chemical use patterns continued with neonicotinyls as soil and foliar applications, sugar esters, azadirachtins and entomopathic fungi, and pymetrozine. No progress was achieved in evaluating the potential for transforming plants with natural product genes
<b>Develop improved methods of application including formulation and delivery of materials to improve control.</b>	Develop spray systems for better underleaf coverage. Evaluate rates, timing, placement in relation to efficacy. Consider formulation, UV protectants, and other means to improve efficacy. Develop improved methods to evaluate application efficacy. Field test under commercial conditions for technology transfer		X	No new advancements in foliar spray delivery systems or assessment techniques were reported in 2002. A considerable amount of progress was made in evaluating efficacy relative to insecticide rates, timing, and placement. Progress in this area is highlighted in several abstracts.
<b>Conserve insecticide efficacy: Relate action thresholds to insecticide usage patterns.</b>	Refine action thresholds based on insecticide efficacy and input from other control strategies. Summarize and recommend in IPM systems.		X	Very little new work on action thresholds was reported in 2002. The most significant work was done in tomatoes to develop AT for IGR use (D.J. Schuster). See section F for technology transfer.
	Summarize and recommend in IPM systems.			
<b>Elucidate the role of genetic, biochemical and ecological factors leading to insecticide resistance.</b>	Conduct studies to determine the genetics and biochemistry of resistance and cross resistance to different classes of insecticide. Evaluate the role of refuge habitats (weeds, tolerant crops, urban areas) to assure input of susceptible genes in whitefly population. Evaluate the influence of host plant on susceptibility to insecticides.	X		Progress was made in characterizing cross resistance among neonicotinoid insecticides. Monitoring of whitefly susceptibility to IGR, neonicotinoids and other chemistries continued in AZ, CA, and FL with no major shifts in susceptibility measured.

**Table C. Chemical Control, Biopesticides, Resistance Management, and Application Methods. (Continued)**

Research Approaches <sup>a</sup>	Year 4/5 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Improve insecticide efficacy: Improve techniques for monitoring resistance.</b>	Summarize, analyze, and produce standardized comparable monitoring systems. Develop standard systems for general use including user friendly techniques to assist growers and extension agents to evaluate susceptibility of whitefly populations to commonly used insecticides.	X		Significant progress was made in determining the parameters of chemical uptake of imidacloprid through petioles in systemic bioassays. This information will allow for the improvement of adult bioassays for systemic compounds based on concentration taken up by leaves.
<b>Develop, evaluate and refine resistance management systems.</b>	Develop criteria for integration of successful strategies in agricultural systems. Field test resistance management systems as long range components of successful IPM. Technology transfer.	X		Progress was made in AZ to expand the AZ Whitefly IRM/IPM program to implement recommendations for the shared use of buprofezin and neonicotinoids among commodities such as cotton, melons and vegetables.
<b>Integrate chemical control with other tactics.</b>	Test compatibility of biological control with selective synthetic or natural product insecticides as required. Integrate systems with host plant resistance and cultural controls  Integrate systems with host plant resistance and cultural controls. Summarization and technology transfer	X		Significant progress was made in determining the effect of IGR and other chemistries in conservation of natural enemies and additive effects to chemical control – “Bioresidual “

<sup>a</sup> See Table A for complementary research on thresholds.

<sup>a</sup> See Table B for complementary research on virus/vector interactions.

<sup>a</sup> See Table D for complementary research on biological control.

<sup>b</sup> See Table E and F for complementary research on systems management.

## Reports of Research Progress

### Section D: Natural Enemy Ecology and Biological Control

Co-Chairs: Bill Roltsch and Greg Simmons

**Investigator's Name(s):** Earl Andress<sup>1</sup>, Julie Gould<sup>2</sup>, Larry Heilman<sup>3</sup>, Arland Oleson<sup>3</sup>, and Mark Quinn<sup>4</sup>

**Affiliation & Location:** 1. USDA-APHIS, Imperial Valley Research Station, Brawley CA., 2. USDA-APHIS-PPQ, Otis Protection Center, Otis MA, 3. Department of Biochemistry, North Dakota State University, Fargo ND, 4. Department of Crop and Soil Sciences, Washington State University, Pullman WA

**Research & Implementation Area:** Section D: Natural Enemy Ecology and Biological Control

**Dates Covered by the Report:** January 1, 2001 - December 31, 2001

#### Exotic Parasitoids in Four Crops in the Imperial Valley

Exotic parasitoid species introduced into the Imperial Valley of California were recovered throughout the agricultural system in 2000 and 2001. No parasitoids were released at all in 2001, and the last of the mass releases were made during the summer of 1999. Samples were collected from four crops in 2000 and 2001 and tested for parasitism using the Squash blot methods developed by (Heilman et al. *ibid*). Percentages of whiteflies parasitized by exotic parasitoid species were as high as 32, 28, 52, and 28 in broccoli, cantaloupe, alfalfa, and cotton respectively, and were as low as 0. The high variability in parasitism by exotics observed here may suggest that although populations of exotic introduced species of Aphelinid parasitoids are well established in the Imperial Valley, their influence on whiteflies has the potential to continue to grow in coming years. In some cases exotic parasitoids even outnumbered indigenous parasitoid. The broad range of parasitism levels observed confirms that factors affecting parasitism of whiteflies are highly variable within the Imperial Valley agricultural system. Factors that will be quantified and analyzed for their effect on both whitefly and parasitoid populations observed during the course of this study will include distance to the edge of the agriculture areas, coverage of host crops, coverage of non-host crops, proximity to urban areas, insecticide use, and predation.

**Investigator's Name(s):** James S. Buckner<sup>1</sup> and Walker A. Jones<sup>2</sup>

**Affiliation & Location:** <sup>1</sup>USDA-ARS, Insect Genetics and Biochemistry Research Unit, Red River Valley Agricultural Research Center, Fargo, ND; <sup>2</sup>USDA-ARS, Beneficial Insects Research Unit, Subtropical Agricultural Research Center, Weslaco, TX.

**Research & Implementation Area:** Section D: Natural Enemy Ecology and Biological Control

**Dates Covered by the Report:** 2001

**Methyl-Branched Alkanes as Possible Marking Pheromones for Female *Eretmocerus mundus*,  
a Parasitoid of *Bemisia argentifolii***

Previous studies with *Bemisia argentifolii* Bellows and Perring (Homoptera: Aleyrodidae) nymphs have shown that wax esters were the major components of the cuticular lipids with lesser amounts of hydrocarbons, long-chain aldehydes and long-chain alcohols (1). The hydrocarbon fraction was identified as a homologous series of odd-carbon-number *n*-alkanes, C<sub>31</sub> (40%), C<sub>33</sub> (31%), C<sub>35</sub> (18%) and C<sub>29</sub> (11%) and trace amounts of even-carbon-number *n*-alkanes (C<sub>28</sub>-C<sub>34</sub>). In a more recent study, the cuticular lipids from *B. argentifolii* nymphs parasitized by the whitefly parasitoid *Eretmocerus mundus* Mercet contained measurable quantities of two additional components in their hydrocarbon fraction (2). Capillary gas chromatography (CGC) and CGC-mass spectrometry (CGC-MS) analyses and comparisons with an authentic standard indicated that the two hydrocarbons were the monomethyl-branched alkanes, 2-methyltriacontane (31 carbons) and 2-methyldotriacontane (33 carbons). In parallel experiments, no appreciable changes in lipid composition were observed for the cuticular lipids of whiteflies parasitized by another parasitoid, *Encarsia pergandiella*. In contrast to *Encarsia* spp., *Eretmocerus* spp. oviposit externally (under the nymph), yet still readily avoid superparasitism suggesting that an external discrimination cue must be present (2). Previous behavioral observations with *E. mundus* showed that, following oviposition, females often make apparent "host marking movements" that may leave a chemical that enables the searching female to distinguish parasitized from unparasitized hosts (3). Those comparative findings suggest a possible function of 2-methylalkanes as marking pheromones for ovipositing female *E. mundus*.

The following set of experiments decreased the possibility that the extra 2-methylalkanes on cuticular surfaces of *E. mundus*-parasitized nymphs arose from biosynthesis and surface deposition during growth and development of the parasitoid, and supported the possibility for transfer of these compounds onto the surface of the host (nymphs) by searching/ovipositing *E. mundus* females. *B. argentifolii* nymphs were exposed to *E. mundus* females for approximately 8 hr and then removed from leaves after 48 hr. Only those nymphs with visible signs of an oviposited egg beneath each nymph were removed and pooled for subsequent analysis of their cuticular lipids. The hydrocarbon fraction of nymph samples that had been exposed to ovipositing *E. mundus* females showed the same 2-methylalkanes that were also present on those parasitized *B. argentifolii* nymphs that had been held 10 days after exposure to *E. mundus* females. In addition to 2-methylalkanes, small quantities of 31- and 33-carbon dimethyl-branched-alkanes were also identified in the hydrocarbon fraction from those nymphs exposed to *E. mundus* for 48 hr. Preliminary CGC-MS analyses of the cuticular lipids from *E. mundus* females have revealed major quantities of the same 2-methylalkanes and dimethyl-branched-alkanes that were on the cuticular surfaces of *E. mundus*-exposed *B. argentifolii* nymphs. These findings clearly demonstrate that the origin of the 'extra lipids' on the surfaces of *E. mundus*-parasitized nymphs is from the ovipositing female and that these lipids could be playing a role as host marking chemicals. Current studies are focused on defining the process for transfer of these methyl-branched lipids from the ovipositing parasitoid to its host.

**References**

1. Buckner et al. 1999. Comp. Biochem. Physiol. B124: 201-207.
2. Buckner et al. 2000. Arch. Insect Biochem. Physiol. 44: 82-89.
3. Foltyn and Gering 1985. Entomol. Exp. Appl. 38: 255-260.

**Investigators Name(s):** Matthew A. Ciomperlik & John A. Goolsby<sup>2</sup>

**Affiliation & Location:** USDA-APHIS-PPQ Mission Plant Protection Center, P.O. Box 2140, Mission, TX 78573-2140;  
<sup>2</sup>USDA-ARS, Australian Biological Control Laboratory, Private Mailbag #3, Indooroopilly, Queensland, Australia, 4068

**Research & Implementation Area:** Section D: Natural Enemy Ecology and Biological Control

**Dates Covered by the Report:** Sep 1999 -Dec. 2000

### **Establishment of Introduced Parasitoids of Silverleaf Whitefly in the Lower Rio Grande Valley of Texas**

Since 1993, a large number of exotic *Eretmocerus* and *Encarsia* parasitoid species/strains have been evaluated in quarantine, laboratory, and field studies to determine promising species for release in subtropical agroecosystems in the lower Rio Grande Valley (LRGV) of Texas. Those evaluations suggested that *Eretmocerus hayati* Rose & Zolnerowich (M95012, Pakistan), *Eretmocerus* nr *emiratus* (M96076, Ethiopia), and *Eretmocerus mundus* (M92014, Spain) might become established on whitefly in crops grown in the region, because they attacked significantly greater numbers of hosts than the native parasitoid species. Prior to exotic parasitoid releases, the native parasitoid complex attacking silverleaf whitefly in the LRGV was comprised of six parasitoid species. Those species included; *Eretmocerus tejanus* Rose & Zolnerowich, *Encarsia pergandiella* Howard, *E. meritoria*, *E. nr strenua*, and *E. formosa*. Of those, *Eretmocerus tejanus* and *Encarsia pergandiella* were the most abundant species collected from all crop types and weed host plants sampled (Riley & Ciomperlik, 1997). These two parasitoid species were widely distributed across the region and collected from almost all crop and weed host plant types.

Field releases of exotic parasitoid species/strains occurred from 1993 to 1998 across multiple release sites, as either direct releases of small to large numbers of parasitoid adults onto hosts within the crop, or as large scale evaluations of parasitoid release methods (Goolsby & Ciomperlik 1996). Establishment evaluations during June of 1997 showed that the native parasitoids comprised approximately 98% of all parasitoids collected. Subsequent monthly evaluations in 1998 showed increasing numbers of exotics becoming established through time (Goolsby et al. 1999). Those evaluations during 1998 showed that *Eretmocerus hayati* and *Eretmocerus mundus* were at times as abundant as the native *Eretmocerus tejanus*.

Field sampling to determine parasitoid establishment, abundance trends, and distribution were conducted in 1999 and 2000. Three sampling methods were routinely used to collect parasitoid samples from economically important crops and weed host plants within the LRGV. The three sampling methods were; 'recovery samples' where release sites were re-visited either 1 or 2 years after initial releases were made, 'random samples' where crops were sampled at multiple random locations within the same county as where the parasitoids were released, and 'triplicate sentinel' host plants infested with second instar silverleaf whitefly placed in large acreage crop production areas. In the case of the recovery and random samples, leaf samples containing whitefly pupae were returned to the laboratory, where individual parasitized pupae were isolated in ¼ dram glass vials for emergence. In the case of the triplicate sentinel plant method, three seedling plants including cotton, cantaloupe, and cucumber infested with 2<sup>nd</sup> instar whitefly were placed within the crop canopy at each sample site. During the winter months, cabbage seedlings were substituted for the cucumber seedlings. After three days of exposure, the three seedling sentinel plants were returned to the laboratory and held for parasitoid development and handled as described above. All of the emerging parasitoids adults were identified to genus and species where possible using morphological characters. All suspect identifications were confirmed by RAPD-PCR analysis.

Analysis of all samples indicate that *Eretmocerus hayati* (Pakistan) is ubiquitously distributed amongst all the crop and weed host plants sampled in all counties with the exception of Starr County. This parasitoid accounted for 96.5% of the *Eretmocerus*, and 88% of all species of parasitoids collected in the establishment evaluations (n=866 parasitoids). *Eretmocerus mundus* comprised about 3%, while *E. nr emiratus* comprised about 0.6% of all *Eretmocerus* individuals collected. *Eretmocerus tejanus* and *Encarsia pergandiella* were infrequently collected, comprising 0.6% and 4.8% of all parasitoids collected. Attack rate patterns by the various native and exotic parasitoids are still under analysis at this time. The low numbers of recoveries of native parasitoids may suggest that exotic and native populations have not reached equilibrium densities, therefore establishment evaluations will continue in 2001.

**Investigator's Names:** Larry J. Heilmann<sup>1</sup>, Jianzhong Zhang<sup>1</sup>, Earl R. Andress<sup>2</sup>, and Arland E. Oleson<sup>1</sup>, Dennis R. Nelson<sup>3</sup>, Juli R. Gould<sup>4,6</sup>, Matthew W. Ciomperlik<sup>5</sup>, John A. Goolsby<sup>5,7</sup>, and Don C. Vacek<sup>5</sup>

**Affiliation & Location:** <sup>1</sup>Department of Biochemistry and Molecular Biology, North Dakota State University, Fargo, ND 58105, <sup>2</sup>USDA-APHIS-PPQ-PPPC, Brawley, CA 92227, <sup>3</sup>USDA-ARS-RRVARC Biosciences Research Laboratory, Fargo, ND 58105, <sup>4</sup>USDA-APHIS Phoenix PPC, Phoenix AZ 85040, <sup>5</sup>USDA-APHIS Mission PPC, Mission, TX 78572, <sup>6</sup>Current Address: USDA-APHIS-PPQ Otis Plant Protection Center, Otis NGB, Otis MA 02542, <sup>7</sup>Current Address: USDA-ARS Australian Biological Control Laboratory, Indooroopilly, Queensland, Australia

**Research & Implementation Area:** Section D: Natural Enemy Ecology and Biological Control

**Dates Covered by the Report:** 1996-2001

### DNA Based Species Identification Test for Parasitic Wasps of the Whitefly

A significant problem in using parasitic wasp species in biological control of the whitefly is identification of the individual wasp species parasitizing the nymph. This is particularly important in areas where imported foreign species are released. It is necessary to follow these releases to determine which species establish and reproduce, if they effectively parasitize local whitefly populations and if they displace native parasitoids. Current taxonomic tests for species identification are time consuming and require experienced and trained personnel.

We have developed a simple and relatively quick DNA based species identification test that can be applied to squashes of adult, pupal, and larval stage *Eretmocerus* and *Encarsia* wasps parasitizing whiteflies. The test utilizes highly repetitive but species-specific DNA sequences to identify the parasitizing wasp in one to two days. We currently have cloned DNA probes for *Eretmocerus mundus* (Spain), *E.sp.* (Ethiopia), *E. hayati*, and *E. eremicus* as well as *Encarsia formosa* and *Encarsia sophia* (transvena). The probes range from highly species-specific to only being selective for old world vs new world species. Each of the cloned probe DNAs has been sequenced and their distribution in relevant parasitoid species quantified.

In a major field test of the procedure over 26,000 whitefly larvae from the Imperial Valley of California were squashed on filters and hybridized with the probes. These larvae were from multiple locations, multiple crops and different times of the year. Approximately 20% of these larvae were parasitized by the native California species, *E. eremicus*, with individual field variations ranging from 0 to 77%. Imported species from Europe, Africa and Asia parasitized only 3-4% of the larvae. Results on this field test will be presented.

**Investigator's Name:** Jesusa C. Legaspi

**Affiliation & Location:** USDA -ARS-Center for Biological Control, Tallahassee, FL

**Research & Implementation Area:** Section D: Natural Enemy Ecology and Biological Control

**Dates Covered by the Report:** Summer-fall 2001

### **Parasitism of the Silverleaf Whitefly in North Florida**

The objective of this study is to assess the population densities of the silverleaf whitefly (SLWF), *Bemisia argentifolii*, and the degree of parasitism (from native and exotic parasites) in various vegetables and alternate plant hosts in north Florida. Surveys for silverleaf whitefly were conducted during the summer of 2001 in vegetables, including tomatoes, squash, cabbage, watermelon, and okra and in ornamentals, such as poinsettias and hibiscus. During September to October 2001, a minimum of 10 leaves were collected from soybean (*Glycine max*), poinsettia (*Euphorbia pulcherrima*), and lantana (*Lantana camara*) in Leon and Gadsden Co. FL and returned to the laboratory for data processing. Counts of the different stages and numbers of whiteflies per leaf were made and recorded using a stereomicroscope. The leaf samples were kept in emergence cages (plastic petri dishes or plastic boxes) that were painted black with a glass vial attached to the side of the cage. Each leaf petiole was inserted into a small plastic vial filled with water to keep the leaves from drying inside the cage. The emergence cages were kept in an environmental growth chamber at 28-30 °C and 14L:10D photoperiod. Parasites that emerged were collected in the vial and placed in 70% alcohol. The parasite specimens were sent to local taxonomists for identification of species.

Surveys for silverleaf whitefly populations during the summer of 2001 indicated a very low population of SLWF. Preliminary results indicated that the mean numbers of whitefly per leaf were highest in lantana (53.3) followed by poinsettia (31.50) and soybean (14.50). Average percentage of parasitism of SLWF was highest in soybean (12.12%), followed by poinsettia (7.89%) and lantana (3.81%). The parasite:whitefly ratio was calculated as 7.25, 11.67 and 25.26 in soybean, poinsettia and lantana, respectively. The main parasites of SLWF that emerged from poinsettia were of the genus *Encarsia* sp. while those from lantana and soybean were of the genus *Eretmocerus* sp. (both Hymenoptera: Aphelinidae). Further species identification will be confirmed by local taxonomists.

**Investigator's Name(s):** C.L. McKenzie, W.B. Hunter, S.L. Lapointe and P. Dang

**Affiliation & Location:** USDA-ARS, Horticultural Research Lab, 2001 South Rock Road, Ft. Pierce, FL

**Research & Implementation Area:** Section D: Natural Enemy Ecology and Biological Control

**Dates Covered by the Report:** 2000-2001

### **Iridovirus: Potential Entomopathogenic Biocontrol Agent for Whitefly**

Few microbial biocontrol agents and no known viral agents are commercially available for use on whiteflies. A newly described entomopathogenic virus was recently discovered in south Florida whitefly populations and was determined to be an iridovirus by DNA analysis. Invertebrate iridescent virus 6 (IIV-6) is known to be pathogenic to insects including whiteflies. Demonstrated modes of transmission for iridovirus in other insect systems have been shown to be through oral ingestion, cuticular wounding (abrasions) and occasionally sexually transmitted. Traditionally, many biological agents are applied by foliar spray application; therefore, we attempted to determine if this would be a feasible method of application of iridovirus to control whiteflies on tomatoes.

A plant-based petri dish bioassay system was developed to hold single tomato leaves infested with whitefly for their entire life cycle (egg to adult). A hole was cut in the lid of a Polystyrene petri dish (100x20 mm) to facilitate ventilation and ultra-fine screen mesh was used to cover the hole to prevent whitefly escapes. Polystyrene conical tubes (15 ml, 17x120 mm) were filled with a plant tissue culture water agar and fertilizer (9-45-15) mixture. A hole was cut at the base of the petri dish to hold the conical tube so that it fit tightly. Tomato leaves were cut so that the terminal leaf of the leaflet would fit in the petri dish. The remaining leaves of the leaflet were cut off and the stem was placed as far into the agar as possible with the abaxial leaf surface facing the screen to facilitate counting. A small hole was drilled into top of the conical tube above the agar line so that H<sub>2</sub>O could be added as needed (leaves will sprout roots and last > 28 d).

The iridovirus solutions were applied using an ultra-low volume spray device that consisted of a spray platform that holds a pressurizable spray bottle at the proper distance and angle. Measured amounts (200 or 400 µl) of iridovirus solution were placed in a test tube (12 x 75 mm). The spray bottle siphon tube was placed in the test tube so that the test tube fit into the nozzle-pump body. Parafilm was wrapped around the top of the test tube to insure a tight fit and the spray bottle was pressurized to ~10 psi w/ 20 strokes of the pump mechanism to deliver a fine spray for each application.

Uniform infestations of whitefly were allowed to acclimate on tomato leaves in the petri dish bioassay system. Abaxial and adaxial leaf surfaces were sprayed with 200 µl of crude virus preparation in a cold room. Controls were sprayed with PBS buffer only. Petri dishes were placed upright in a Percival Scientific incubator at 22 ± 1° C under a L/D, 16:8 photoperiod. Whitefly were harvested at 10, 14 and 18 days post spray and were subjected to PCR analysis for presence of IIV6. The experiment was repeated using the purified virus preparation and only the adaxial leaf surfaces were sprayed with 400 µl of iridovirus solution.

We confirmed adult whitefly infection with IIV-6 through oral ingestion via sucrose parafilm feeding chamber experiments and PCR analysis. Whiteflies were fed a 15% sucrose solution with crude or purified virus preparation for 72 hr and transferred to tomato leaves in the petri dish bioassay system for 14 days to allow the virus to replicate. Controls were fed sucrose only. Whiteflies were analyzed via PCR for presence of IIV6.

Although we could demonstrate infection in whiteflies *per os*, spray applications were inconclusive. When virus solutions were sprayed onto the insects and plant tissues no apparent virus infection was detected. This may be due low virus titer in our spray solutions; whiteflies did not ingest surface droplets when feeding, and/or the inability of the virus to remain viable on the leaf surface. Experiments are underway to try and address these concerns.



**Investigator's Names:** Steven E. Naranjo<sup>1</sup> & Nilima Prabhaker<sup>2</sup>

**Affiliation & Location:** <sup>1</sup>USDA-ARS, Western Cotton Research Laboratory, Phoenix, AZ and <sup>2</sup>Department of Entomology, University of California, Riverside, CA

**Research & Implementation Area:** Section D: Natural Enemy Ecology and Biological Control

**Dates Covered by the Report:** January 2000-December 2001

**Toxicological Studies of Two Insect Growth Regulators on *Geocoris punctipes*,  
*Orius insidiosus* and *Collops vittatus***

Laboratory experiments were continued to evaluate the direct effects of two insect growth regulators (IGR), buprofezin (chitin synthesis inhibitor) and pyriproxyfen (JH analog), on survival and reproduction of *Geocoris punctipes*, *Orius insidiosus* and *Collops vittatus*, 3 common generalist predators found in cotton. Both topical and contact residue assays were performed on adults and terminal-stage nymphs at recommended field rates of the materials. Buprofezin was found benign to all species and stages of exposure in terms of survivorship and reproduction. This is consistent with previous dose-response studies with *G. punctipes* where it was impossible to estimate lethal concentrations. Pyriproxyfen applied to adults had no effect on survival or reproduction of *G. punctipes* or *O. insidiosus*. However, egg viability, but not survival, of *C. vittatus* appeared to be affected by pyriproxyfen. Pyriproxyfen applied to terminal nymphal stages of *G. punctipes* and *O. insidiosus* caused some mortality and adult deformities compared to the control. Pyriproxyfen did not affect egg viability of treated adults or adults arising from treated nymphs of either of these species.

**Investigator's Names:** Steven E. Naranjo

**Affiliation & Location:** USDA-ARS, Western Cotton Research Laboratory, Phoenix, AZ

**Research & Implementation Area:** Section D: Natural Enemy Ecology and Biological Control

**Dates Covered by the Report:** January 2000-December 2001

### **Comparison of Natural Enemy Fauna and Function in Transgenic Bt and Non-Bt cotton**

Field studies were conducted in 1999-2001 to evaluate the combined direct and indirect effects of transgenic-Bt cotton on populations of natural enemies. Paired non-Bt and Bt fields of the same background were established and monitored weekly for foliar dwelling natural enemies. In 2001, each treatment was further split to contrast completed unsprayed plots to those receiving insecticides as needed for whitefly, *Lygus* and lepidopteran pests. Analyses are incomplete, but preliminary results indicate no consistent differences in predator populations between Bt and non-Bt cotton. Parasitism by aphelinids attacking *Bemisia tabaci* was very low and unaffected by Bt cotton. Additional studies were conducted in 2001 to examine effects on natural enemy function. Sentinel pink egg masses were placed in plots to estimate rates of predation and parasitism on pink bollworm eggs. Cohort-based life table studies were conducted to estimate rates of predation and parasitism on immature *B. tabaci*. Preliminary results indicate that rates of natural enemy -induced mortality on these two key pests were the same in Bt and non-Bt cotton.

**Investigators Name(s):** Alberto Pantoja<sup>1</sup>, Matthew A. Ciomperlik<sup>2</sup>, Norberto Gabriel<sup>3</sup>, Pedro Vazquez<sup>3</sup>, Leyinska Wiscovitch<sup>3</sup>, and Wilfredo Robles<sup>1</sup>

**Affiliation & Location:** <sup>1</sup>Department of Crop Protection, Puerto Rico Agricultural Experiment Station, P.O. Box 21360, Rio Piedras, Puerto Rico 00928. <sup>2</sup>USDA-APHIS-PPQ Mission Plant Protection Center, Moore Air Base, Bldg. S-6414, Route 3 Box 1014, Edinburg, TX. <sup>3</sup>USDA-APHIS-PPQ, San Juan, Puerto Rico

**Research & Implementation Area:** Section D: Natural Enemy Ecology and Biological Control

**Dates Covered by the Report:** 1999-2001

### **Evaluation of the Establishment and Distribution of Exotic Biological Control Agents of Silverleaf Whitefly in Puerto Rico**

Silverleaf whitefly (SLWF) is a key pest of many broadleaf vegetables, field crops, and ornamentals in tropical and subtropical regions. Losses caused by plant debilitation, sooty mold growth, physiological disorders and transmission of geminiviruses by whiteflies, and control costs, have impacted vegetable production systems in Puerto Rico significantly. In efforts to combat these losses, a biological control program was cooperatively implemented by USDA/APHIS/PPQ and the University of Puerto Rico to control SLWF. The biological control program included both classical and augmentative approaches. From 1996 to 2000 the USDA/APHIS/PPQ laboratory in Mission, Texas and the local USDA/APHIS/PPQ office released 1,262,536 parasitoids in 20 locations throughout Puerto Rico. The species released include *Encarsia formosa*, *Encarsia sophia* (*transvena*), *Eretmocerus mundus*, *E. hayati*, *E. emiratus*, *Eretmocerus sp. nr emiratus* and a predatory coccinellid *Serangium parcesetosum*.

Recovery studies were initiated during 1999 to determine establishment and distribution of the parasitoids released in the collaborative effort. Five hundred and thirty specimens have been recovered from twenty locations and shipped to the USDA-APHIS-PPQ Mission Biological Control Center (MBCC) in Texas for genetic identification. The specimens were subjected to RAPD-PCR analysis using the OPC-04 primer at the MPPC. *Eretmocerus mundus*, *E. tejanus*, and *E. haytii* represents 16, 0.6, and 1% respectively of the specimens recovered.

Reconciliation of data records indicating the initial release and the recovery site indicates that parasitoids can be recovered three years after the initial liberation. This suggests that the parasitoids have established on the island. The data also suggest that several native *Encarsia* species might be present. The native parasitoid fauna of Puerto Rico has not been well established, however, the predominant parasitoids collected during pre-release field sampling indicated that several *Eretmocerus* species are present but are less numerous than the native species of *Encarsia transvena* (M. Ciomperlik, Unpublished Data). From pre-release evaluations, it was clear that the native parasitoid complex was not well suited to utilize *Bemisia argentifolii* as a host. After the release of the exotic *Encarsia* and *Eretmocerus* species, growers have indicated that whitefly pressures have been less severe, that yields have increased, and the use of insecticide reduced. After two years the savings associated to the reduction of insecticide use by augmenting natural enemies is estimated to exceed \$1200 /acre (Pantoja unpublished data). The data indicates that the exotic parasitoids have become established and are regulating whitefly populations.

**Investigator's Name(s):** C. H. Pickett, J. Brown, G. Simmons<sup>1</sup>, J. Goolsby<sup>2</sup>, and Bill Abel<sup>3</sup>

**Affiliations & Locations:** California Department of Food & Agriculture, Biological Control Prog., Sacramento, CA;

<sup>1</sup>USDA-APHIS PPQ, Oswell St. Biological Control Facility, Bakersfield, CA; <sup>2</sup>USDA-ARS, Australian Biological Control Laboratory, Indooroopilly, Queensland, Australia; <sup>3</sup>USDA-APHIS PPQ, Shafter, CA

**Research & Implementation Area:** Section D: Natural Enemy Ecology and Biological Control

**Dates Covered by the Report:** August 1997 to November 2001

### **Releases of Exotic Parasitoids for Permanent Establishment in Central California**

The silverleaf whitefly, *Bemisia argentifolii*, was an increasingly important pest of cotton in the San Joaquin Valley from 1994 through 1997 when this project was initiated. Field studies suggested that citrus had become an important overwintering site for this whitefly. Cotton has the highest incidence of whitefly infestations in areas of the Valley with a matrix of citrus orchards and cotton fields. We report on large scale releases of *Eretmocerus emiratus* (M95104, U.A.E.), *Eretmocerus* nr. *emiratus* (M96076, Ethiopia), *E. mundus* (M92014, Spain), *E. hayti* (M95012, Pakistan), and trace numbers of *Encarsia sophia* (= *transvena*; M95107, Pakistan) into four citrus orchards. The study had two goals: (1) to determine if exotic parasites released into citrus during the fall will overwinter in this habitat and move into cotton the following spring; and (2) to permanently establish new populations of exotic parasites specific for the silverleaf whitefly.

Three study sites were identified initially, one each in Fresno, Tulare, and Kern counties. A fourth was added because one of the original growers stopped farming cotton (Kern Co.). Sites consisted of citrus and cotton acreage managed by the same owner. Cotton is grown directly adjacent to the citrus, and growers have had a history of silverleaf whitefly problems. We began releasing parasites in early August or September 1997, 1998, 1999, and 2000 when migrating whitefly nymphs were first recorded from citrus leaves. Typically, over 100,000 parasites were released weekly at each location and a total of 4.05 million were released in 1997, over 10 million in fall 1998, 3.2 million in 1999, and 124,000 in 2000. The dispersal of the released parasites was recorded using sticky cards with identification based on the adult males since they could be readily distinguished from native *Eretmocerus* while on the sticky cards.

Parasitism of silverleaf whitefly on citrus was generally quite low, averaging 28% overall. However this value is quite high with respect to an earlier survey in which less than 1.5% of nymphs examined on cotton from the same region were found parasitized. Whitefly densities on citrus remained very low, usually less than 0.1 nymphs per cm<sup>2</sup> leaves. During years in which exotics were being released, most of the parasitoids recovered from weed samples taken within 1 mile of citrus orchards were exotic, 81% to 95%. Two years after the last releases of exotic *Eretmocerus* spp., fall 2001, the proportion of all parasitoids sampled from weeds that were exotic dropped to 11%. Although primarily *Eret. emiratus* (M95104 + M96076) was released, *Eret. mundus* (M92014) was the dominate species recovered in fall 2001. The density of silverleaf whitefly on weeds has varied about the same from 1998 to 2001, most samples from 0.1 to less than 20 per gram dry weight.

**Investigator's Name(s):** William. J. Roltsch

**Affiliation & Location:** California Department of Food & Agriculture, Biological Control Program, 3288 Meadowview Rd., Sacramento, CA 95832

**Research & Implementation Area:** Section D: Natural Enemy Ecology and Biological Control

**Dates Covered by the Report:** August 1994 – Nov. 2001

### **Establishment of Introduced Parasitoids of the Silverleaf Whitefly in Imperial Valley, CA**

Several species and strains of exotic *Eretmocerus* and *Encarsia* were evaluated in field cages, and released in large numbers in commercial fields, refuge nursery plots and urban yards in Imperial Valley by State, Federal and University scientists. The most promising *Eretmocerus* for this desert region include *E. emiratus* Zolnerowich & Rose, *E. sp. nr. emiratus* (from Ethiopia) and *E. mundus* Mercet. *Encarsia sophia* (= *Encarsia transvena*) from Malan, Pakistan appears promising as well. Identification to species was accomplished using recently published morphological keys and by DNA analysis (RAPD-PCR) by the USDA-APHIS, Plant Protection Center, Mission, TX. This report is a summary of silverleaf whitefly (SLW), *Bemisia argentifolii* Bellows & Perring, parasitoid population development in long-term refuge nursery plots from 1994 to 2001, and a three-year, late season survey of exotic parasitoids in commercial cotton within the Valley.

From 1994 through 1997, exotic parasitoids were released into long-term refuge field plots on multiple occasions each year. Plots (1/2 to 1 acre) were located at the Imperial Valley Research Center near Brawley, CA, and at an organic farm at the south end of the county. During the warm season, the plots consisted of okra and basil. During the cool season, cole crops (esp. collard) and sunflower were present. Kenaf, roselle and eggplant were also periodically present (1994-1996) along with adjacent plantings of cotton and spring cantaloupe. Leaf samples were taken approximately 6 times during each year to determine the status of parasitoid population increase and persistence. Neither *E. tejanus* nor *E. stauferi* (i.e., *Eretmocerus* spp. from Texas) have been recovered following their release in 1994. During 1995, *E. melanoscutus* was released in large numbers but recoveries of this parasitoid were rare. Releases of *E. mundus*, *E. hayati* and *E. emiratus* began in April of 1996. Numbers of exotic parasitoids compared to natives were high during early summer; however, the proportion of the sample consisting of exotic species dropped markedly by late July, indicating poor performance (population increase and persistence) during this very warm summer period. During 1997, *E. emiratus* (origin: United Arab Emirates) and *E. sp. nr. emiratus* (origin: Ethiopia) were released. The relative performance of exotics was considerably better than in 1996. Several samples from May to September consisted of up to 90% exotic *Eretmocerus* spp. versus native *Eretmocerus eremicus*. The proportion of exotic *Eretmocerus* relative to native *Eretmocerus eremicus* declined once again during the fall, however, not to the same extent. During 1998, none of the long-term refuge plots was inoculated with exotic whitefly parasitoids. This made possible the assessment of populations released in previous years at these sites, in terms of their ability to overwinter and compete with native species of SLW parasitoids. Overwintering on cole crops was confirmed albeit in low numbers. During the summer of 1998, 1999, & 2000 exotic *Eretmocerus* spp. were dominant over native *E. eremicus* on okra, basil and adjacent cotton in nearly all samples taken. The order of dominance of exotic *Eretmocerus* species is *E. sp. nr. emiratus*, *E. emiratus* and *E. mundus*. *Encarsia sophia* reached high densities during the summer and fall of 1998 through 2000 in several of the refuge field plots as well. During 2001 SLW densities on most plant species were very low. From a considerably smaller number of samples compared to previous years, it was indicated that exotic *Eretmocerus* spp. were not nearly as dominant as in the past three years, resulting in most samples containing well over 50% native *E. eremicus*.

Further evidence regarding the extent of exotic parasitoid establishment was provided by surveying cotton fields from 1998 to 2000. Leaf samples were obtained from three edges of conventionally managed cotton fields in Imperial Valley during each year during September and October. Exotic *Eretmocerus* were detected in 10 of the 23 fields (i.e., 43%) sampled in the fall of 1998, 31 of 42 fields (i.e., 74%) sampled in the fall of 1999 and 23 of 24 (i.e., 96%) sampled in 2000. In those fields where exotic *Eretmocerus* were detected, 4% of the *Eretmocerus* were exotics in 1998 and 21% were exotic in 1999 and 48% were exotic in 2000. Similarly, an increase in *Encarsia sophia* was noted as well from 1998 to 2000. *Encarsia sophia* was detected in only one of 23 cotton fields (i.e., 4%) in 1998. However, *E. sophia* was detected in 27 of 42 cotton fields (64%) in 1999, and 24 of 32 (75%) fields in 2000.

In summary, up to four exotic species of silverleaf whitefly parasitoids are well established in Imperial Valley. Compared to native *Eretmocerus eremicus*, their relative yearlong abundance increased consistently from 1998 through 2000. Evidence in 2001 suggested that the level of activity by exotic parasitoid species may have declined, however, it is likely that this represents a simple year-to-year fluctuation in activity.

**Investigator's Name(s):** Gregory Simmons<sup>1</sup>, Kim Hoelmer<sup>2</sup>, Charles Pickett<sup>3</sup>, Eric Natwick<sup>4</sup>, & Earl Andress<sup>1</sup>.

**Affiliation & Location:** <sup>1</sup>USDA-APHIS-PPQ-CPHST, Decision Support and Pest Management Systems Lab, Phoenix AZ;

<sup>2</sup>USDA-ARS, European Biological Control Lab, Montpellier France; <sup>3</sup>California Department of Food & Agriculture, Biological Control Prog., Sacramento, CA; <sup>4</sup>University of California Coop. Extension, Holtville, CA.

**Research & Implementation Area:** Section D: Natural Enemy Ecology and Biological Control

**Dates Covered by the Report:** January 1, 1994 - December 31, 2000.

### **Augmentative Biological Control of Silverleaf Whitefly Infesting Melons with Releases of Exotic *Eretmocerus* Species: a Review of the Progress.**

The primary goal of this project was to increase biological control of the silverleaf whitefly in spring melons by rearing and releasing several species of whitefly parasites. Rates of parasitism by native parasites of whitefly (*Eretmocerus eremicus* & *Encarsia* spp.) are generally low in the spring melon crop in desert production areas of California and Arizona, which is where whitefly populations first start to rapidly increase leading to high regional populations that go on to infest cotton and alfalfa after the melon harvest. If parasitoids are introduced directly into the crop, they can provide control within the crop as well as lead to lower rates of whitefly migration from melons to cotton and alfalfa, reducing whitefly population densities on these crops. Furthermore, releases into spring melons will lead to higher regional populations of parasitoids, that can then go on to attack whitefly in successive crops. Seven years of field investigations of augmentative biological control strategies of whitefly infesting spring melons have demonstrated that parasitism can be increased and whitefly populations suppressed by making open field releases of exotic *Eretmocerus* spp. in commercial melon field plots ranging from ½ to 28 ha in size. Releases of *Eretmocerus mundus* (ex Spain, M92014), *E. emiratus* (ex United Arab Emirates, M 95104) and *E. nr emiratus* (ex Ethiopia, M96076) into both non-treated and Admire<sup>™</sup> (imidacloprid) treated melons have all proven to be about equally effective and increased parasitism greater than releases of the native *E. eremicus*. Mean rates of parasitism ranged from 25-43%. These rates of parasitism were significantly higher than the 7-11% parasitism by naturally occurring native *Eretmocerus* observed in no-release plots. Rates of parasitism in individual plots were as high as 85% indicating that high rates of parasitism are possible. Whitefly densities were reduced by as much as 59% in release plots and were significantly lower than whitefly densities in the no-release control plots. In another study, parasitoid release into Admire treated fields was compared to the standard control measure of pyrethroid applications that growers often used to control late season populations of whitefly that develop after the Admire application is no longer effective. The Admire + parasitoid release treatment achieved levels of whitefly control equal to the Admire + pyrethroid treatment and whitefly densities were about half that of the Admire only treatment, suggesting that parasitoid release could be substituted for pyrethroid treatment. Several techniques for releasing parasitoid pupae were tested including placing pupae in the field in paper cups, using melon transplants inoculated with whitefly nymphs and parasitoids, and releasing pupae with tractor driven release systems using gravity release with dry carriers and sprayer technology with liquid adhesive carriers. The two best release techniques were using transplants, which resulted in the highest levels of control with the fewest number of released parasitoids, followed by releases in paper cups. Parasitoid release using the tractor driven release system with the dry carrier was less effective as wasp emergence rates were reduced (range of 43-78%) relative to the transplant and the paper cup techniques (range 85-100%). It appears that the dry carrier (vermiculite) may have caused desiccation of pupae and that the mechanized gravity drop system itself did not affect emergence. Because the tractor delivery system can increase control by providing a more uniform distribution of released parasitoids within a field, lower labor costs, and increases grower acceptance of the use of augmentative biological control, it would be worthwhile to continue to work with these systems to improve emergence rates. The liquid adhesive release systems resulted in very poor emergence rates as *Eretmocerus* spp. pupae are unable to reorient within the whitefly pupa and if the whitefly pupa is stuck to the leaf surface with the dorsal side down, the wasp is unable to emerge. This results in an average reduction in emergence of 50%. The use of this kind of release system is not recommended for *Eretmocerus* spp. though should be explored for release of *Encarsia* spp. pupae as they are able to reorient within the whitefly and emerge normally. Release rate studies showed that a range of releases of 98-198 thousand parasitoid pupae/ha were the most effective in controlling whitefly. Releasing parasitoids at this rate would cost an estimated \$592 - \$1,186/ha, which is about \$100 - \$686/ha greater than melon growers spent on whitefly control during the worst years of the whitefly outbreak. It is possible that improved release techniques and lower rearing costs could further reduce release rates making augmentative parasitoid releases cheaper. In recent years, regional population levels of whitefly are lower, which means that release rates may be further reduced. Finally, in the high value spring melon crop, using augmentative releases may be helpful to growers by eliminating the need to observe cumbersome worker safety regulations and lengthy reentry intervals associated with restricted use pesticide applications. Future research needs include developing economic thresholds for the use of augmentative releases in melons, refining release methods, and the development of predictive population models to determine the best timing and frequency of parasitoid release. Some additional benefits of this program included: the release of more than 60 million exotic species of whitefly parasitoids into the Imperial valley, which helped establish several new species; the transfer of rearing technology and new more effective whitefly parasitoids to the beneficial insect industry; and cooperation with industry to improve the use of mechanized beneficial insect delivery systems.

**Investigator's Name(s):** Jianzhong Zhang<sup>1</sup>, Arland E. Oleson<sup>1</sup>, Larry J. Heilmann<sup>2</sup>, and Dennis R. Nelson<sup>2</sup>.

**Affiliation & Location:** Biochemistry Dept., North Dakota State University<sup>1</sup>, Fargo, ND 58105; USDA-ARS, Red River Valley Research Center<sup>2</sup>, Fargo, ND 58105.

**Research & Implementation Area:** Section D. Natural Enemy Ecology and Biological Control

**Dates Covered by the Report:** Jan. 1-Dec. 31, 2000

**Microhymenopteran Insect Species as Biocontrol Agents for Silverleaf Whitefly: Differentiation by Use of Hybridization Probes Directed at Highly Repeated Satellite DNA Sequences.**

Highly repetitive satellite DNAs have been employed as hybridization targets for identification of various species of insects. Previously, we have described sequences from selected species of *Encarsia* and *Eretmocerus*, two genera that are being used in the biological control of whiteflies in greenhouse and field environments. The present communication describes efforts to extend the array of probes available for differentiation of various *Encarsia* and *Eretmocerus* species and provides additional information on the distribution and properties of the selected sequences.

The development of these probe sequences entails a multistep process: (1) isolation of genomic DNA from the parasitoid wasp species under study, (2) preparation of a library of cloned DNA restriction fragments in a suitable vector, (3) selection of a subset of the library representing highly repeated sequences by probing with radiolabeled genomic DNA from the insect under study, and (4) examination of the distribution of the subset of cloned repetitive sequences by probing with radiolabeled genomic DNA preparations from other parasitoid wasp species. With several wasp species, clones of unique repeated sequences were recovered. In contrast, some wasp species only yielded cloned sequences that were present in other species as well. Fortunately, the copy numbers of some of these shared repeated sequences vary substantially from one species to another, and many of these sequences are also of use in differentiation of parasitoid wasp species.

Our efforts have been directed at five *Eretmocerus* species (*E. hayati*, Pakistan; *E. emiratus*, United Arab Emirates; *E. mundus*, Spain; *E. sp.*, Ethiopia; and *E. eremicus*, USA) and three *Encarsia* species (*E. formosa*, Egypt; *E. sophia*, Thailand; and *E. pergandiella*, USA). These organisms represent the major species that are indigenous or have been released in whitefly biocontrol efforts in the Southwestern United States. Species-specific probes were obtained for *E. eremicus*, *E. formosa*, and *E. sophia*. These sequences have repeat sizes of 143, 33, and 120 bp, respectively, and they comprise approximately 2%, 7%, and 1% of the wasp genome, respectively. A cloned 260-bp sequence from *E. hayati* comprises 1.5% of the genome in this organism; this sequence cross reacts at a very low level (3%) only with *E. emiratus*. A 172-bp sequence from *E. mundus* has applications as a non-native selective probe. No hybridization signal was displayed by *E. eremicus*, but all other *Eretmocerus* species gave strong signals. This sequence, which represents 2.5% of the *E. mundus* genome, was also the most common clone recovered from the *E. sp.* (Ethiopian) library. The latter library also yielded a clone of a 185-bp moderately repeated element (0.15% of genome) that hybridized strongly with *E. sp.* (Ethiopian) and *E. emiratus*, weakly (6% relative signal) with *E. mundus* and *E. hayati*, and not at all with *E. eremicus*. Another *E. sophia* clone was characterized that provided a weak signal with *E. sophia*, but a much stronger signal with *E. pergandiella*. Hybridization of insect squash blots with one or more of these probes allowed differentiation of all of the tested parasitoids.

Some of these repeated sequences have been tested for their utility as fluorescent, nonradioactive probes of squash blots of parasitoid wasps. In general, oligonucleotide sequences bearing a single fluorescent tag at the 5'-terminus did not produce sufficient signal to allow detection of the cognate wasp species, but larger polynucleotide structures containing many reporter groups appear to provide signals that permit detection of the parasitoids in insect squash blots.

## Section D: Natural Enemy Ecology and Biological Control

Compiled by W. J. Roltsch and G. S. Simmons

Biological control continued to play a prominent role in the Silverleaf Whitefly National Research, Action, and Technology Transfer Plan and was a primary component in work on basic biology, population ecology, host-plant interactions, chemical control technology, and the development of integrated pest management programs. Progress on biological control of *Bemisia* over the last decade has been recently reviewed (Faria and Wraight, 2001. Crop Protection 20, 767-778; Gerling et al., 2001. Crop Protection 20, 779-799; and Naranjo, 2001. Crop Protection 20, 835-852). This research summary covers several specific topics including: classical biological control, augmentation, conservation, natural enemy biology and technology transfer.

Biological control is reliant on sound systematics and the identification of natural enemies and their whitefly host. This topic of key importance is briefly reviewed.

**Foreign Exploration and Classical Biological Control.** The foreign exploration for new biological control agents was completed by 1997. Over 56 populations of *Eretmocerus* and *Encarsia* parasitoids were collected worldwide and cultured in quarantine, in addition to several predator species. Most accessions underwent laboratory and field evaluation. This was followed by the mass-rearing and release of the most promising species and strains. Mass-rearing was conducted in Texas, California, and Arizona through cooperative work plans with Federal, State, County, Universities, and private industry. Because the silverleaf whitefly cycles among numerous short-lived annual crops and urban environments in the harsh arid climates of the southwest United States, an emphasis was placed on making large inoculative releases to increase the probability of establishment of new species. Several release strategies were used including: urban releases, field releases, and refuges near commercial fields. In addition, the establishment effort was aided by extensive releases made in agriculture field plots as part of the evaluation of augmentative biological control using the most promising exotic species.

In California, the silverleaf whitefly severely affected the desert region of Imperial Valley and to a lesser extent the San Joaquin Valley. By 2000, collections from field samples showed that several exotic species were well established. *Eretmocerus* sp. nr. *emiratus* (ex Ethiopia) was the dominant exotic species present. A late season survey in cotton indicated that nearly half of the *Eretmocerus* in commercial cotton fields were *E. sp. nr. emiratus* or *E. emiratus* (ex United Arab Emirates). Small numbers of *Eretmocerus mundus* were also collected.

These samples were composed of approximately 50% native *Eretmocerus eremicus*. The exotic species *Encarsia sophia* was also found to be present in most of the cotton fields. During 2001, sampling efforts across all cropping systems, including a multivariate valley-wide survey effort, suggested that exotic parasitoid activity was less than in 2000. It is unlikely that sufficient time has passed for exotic and native species to achieve a state of equilibrium and year-to-year fluctuations are expected. In the San Joaquin Valley, *Eretmocerus mundus* is the predominant exotic parasitoid established and has been collected from common weeds near citrus and in adjacent cotton fields.

In the lower Rio Grande Valley (LRGV) of Texas, field releases of exotic parasitoids ended in 1998. Parasitoid establishment has been monitored by collecting field samples from numerous host plant species at different releases sites, from various host plant locations and by using a sentinel plant technique. Results from 2000 indicate that *Eretmocerus hayati* has become the primary exotic parasitoid established in the LRGV representing about 95% of the parasitoid species recovered on weed species and commercial crops. *Eretmocerus mundus* and *E. sp. nr. emiratus* represented 3% and 0.6% respectively of the species composition of collected material.

Surveys from 2000 in Arizona showed that, *E. sp. nr. emiratus* is the most common exotic species. A survey of *Bemisia* parasitoids in Florida, the Caribbean, and Central and South America was conducted. Evaluation of parasitoid species composition in northern Florida has recently begun.

### Augmentation

Continuing progress has been made on the development of pathogens for whitefly control in both greenhouse and field crops and was recently reviewed (Faria and Wraight, 2001. Crop Protection 20, 767-778). Advances in production, formulation, and application technology have improved the efficacy of fungal pathogens from strains of *Verticillium lecanii*, and led to the development and registration of several new products based on strains of *Beauveria bassiana* and *Paecilomyces fumosoroseus*. Efficacy of fungal pathogens has been improved by making applications early (before whitefly populations increase), targeting specific stages of *Bemisia* most susceptible to fungal infection, making applications when environmental conditions are favorable to spore survival and growth, and by identifying and avoiding the use of incompatible fungicides. Economic advantages of using conventional insecticides limits wider adaptation of these materials, but continued problems with chemical insecticide resistance, increased regulation of broad spectrum pesticides, and continuing consumer interest in pesticide-free foods should increase the use and acceptance of these materials. A recently discovered entomopathogenic virus was isolated from silverleaf



whitefly and has been identified as an iridovirus. Bioassay systems for this virus have been developed and investigations on its potential for use as an insecticide are underway.

Several studies on augmentative releases of *Eretmocerus* spp. in melon crops in California, Arizona, and Texas have shown that parasitoid releases can increase parasitism and decrease whitefly densities. Releases of *Eretmocerus* spp. using melon transplants inoculated with parasitoid pupae lowered release rates and achieved increases in rates of parasitism and decreases in whitefly density similar to releases by other methods requiring higher release rates. The use of tractor driven mechanized release systems with dry and liquid carriers were explored to learn if more uniform and efficacious control could be achieved by providing more uniform release of parasitoid pupae within the melon crop. Emergence rates were lower relative to other release methods. Because of the potential for better control with more even distributions of released parasitoids, lower labor costs, and increased grower acceptance, continued research is recommended to improve the use of these systems.

Further research on augmentative release strategies is needed for refinement of release techniques, to develop economic thresholds for the use of augmentative biological control in melons, and to develop predictive population models to determine the best timing and frequency of parasitoid releases.

### **Conservation**

Several biorational insecticides including insect growth regulators (IGRs) and juvenile hormone analogs (JH analogs) have been integrated into the pest management of cotton production and vegetable crops. These selective materials are considered to have less impact on natural enemies (predators and parasitoids) of silverleaf whitefly and other crop pests than conventional broad-spectrum insecticides. Natural enemy data from field tests with IGRs indicate that differences in predator numbers in treated and untreated plots were low to moderate for a range of species. Reductions in treated plots typically ranged from 15-35 %. However, reduction in population densities as great as 80% of the ladybug species *Hippodamia convergens* was observed. From these data, it could not be determined whether reductions in ladybug densities were a direct effect of the treatment or an indirect effect of reduced host availability. Laboratory studies have shown that certain IGRs and JH analog products have negligible to low impact on survivorship and reproductive potential of several common predators (*Geocoris punctipes*, *Orius insidiosus* and *Collops vittatus*).

The entomopathogenic fungi *Verticillium lecanii*, *Beauveria bassiana* and *Paecilomyces* are effective against whiteflies in many greenhouse and field crops.

Though non-target effects on some insect natural enemy species have been noted, the use of fungi should improve the conservation of biological control agents in whitefly affected crops (see Faria and Wraight, 2001. Crop Protection 20, 767-778).

Refuges have been used very effectively to provide a good environment for releasing and establishing exotic parasitoid species during the summer and fall period. This included the use of basil and okra (both annual plant species) that were inoculated in early summer with exotic parasitoids and grown into late fall as a parasitoid nursery site for the specific purpose of achieving regional establishment of new species. In contrast, utilizing refuge plantings consisting of annual species in a site-specific pest management mode in Imperial Valley failed. That is, unsatisfactory parasitoid to whitefly ratios occurred when planting a sequence of annual plant species (collard, sunflower, basil and okra) as a yearlong habitat to provide a continual source of parasitoids (native or exotic) to control whitefly in adjacent commercial crops. It is possible that refuge systems composed of perennial plant species could fill such a need. Silverleaf whitefly perennial host plants that have been identified as potentially valuable refuge plants, host low to moderate densities of whitefly for extended periods; whereas several annual plant species (esp. collard) commonly exhibit extreme fluctuations in silverleaf whitefly densities. This needs further investigation.

### **Natural Enemy Biology**

Several studies on dispersal and flight characteristics of both native and exotic species of *Eretmocerus* were conducted in laboratory flight chambers and in open field releases in melons and cotton. In flight chamber studies, flight duration of the native *Eretmocerus eremicus* was affected by mating status with both unmated females and males flying longer than when mated. In field releases in cotton and melons, most *E. eremicus* were shown to move only a few meters from the release point, though some movements as far as 82 m were observed. Field releases of exotic *E. emiratus* in cotton and into surrounding okra and melon plots showed that the majority of recaptures of this species were made within a few meters of the release point. A pattern of more active flight activity in the morning hours was noted as well as sex-based differences in recapture rates, with more males recaptured than females. Information from these studies should be useful for designing more effective augmentative release strategies. Studies on long distance movement of parasitoids and predators, which may be dependent on specific conditions or time of the year, are still needed to better understand establishment processes of new species. Such information could be used to better implement classical as well as augmentative and conservation biological control efforts.

Several years of detailed life-table studies in both treated and untreated cotton has shown that mortality by

conserved natural enemies can be as high as 40%. This information has led to the development of improved IPM for whitefly in cotton.

Field cage studies have provided little evidence that interspecific interactions among parasitoids can significantly disrupt the suppression of whiteflies. Silverleaf whitefly suppression was not reduced (from the highest level of suppression observed by any one parasitoid) when *Eretmocerus eremicus* and *Encarsia sophia* were tested separately or in combination. When *Eretmocerus mundus*, *Encarsia formosa* and *Encarsia pergandiella* were evaluated in all combinations, some reduction in suppression was observed in only one of the combinations that included the two *Encarsia* species. When all three were present, no reduction in suppression was observed.

Studies on parasitoid marking pheromones have been conducted for *Eretmocerus* and *Encarsia* species. This information may play a role in understanding the fundamental differences among species or species group performance at the population level. Research has also uncovered an iridovirus that affects silverleaf whitefly. This is a significant discovery as little is known about viruses of sucking insects.

The effects of leaf surface characteristics (trichomes, exudates etc.) on host plant attractiveness to parasitoids, and parasitoid searching efficiency can be considerable. *Eretmocerus* tenure on melon leaves (having a high density of trichomes) is far less than on cotton (having a considerably smoother leaf surface). Furthermore, trichome exudates of certain plants have the potential of entrapping searching parasitoids and inflicting a considerable degree of mortality. Such studies demonstrate the wide range of influences that silverleaf whitefly host plants may have on parasitoid performance.

### **Systematics and Species Identification**

The ability to identify natural enemy species and populations is imperative to prevent costly duplication and misdirected work. Because of small size and remarkably little variation in morphology among species, the aphelinid Hymenoptera are difficult to identify. Several morphological keys were developed for native and recently introduced species of *Eretmocerus* and *Encarsia*. Because morphological identification of some species is problematic, and body structures with subtle character differences can be easily destroyed, the development of genetic diagnostic techniques has been invaluable. RAPD-PCR and DNA probe technology have been developed for identifying many of the *Eretmocerus* and *Encarsia* species. RAPD-PCR was used for identifying field specimens and testing the purity of cultures. The more recent development of satellite DNA probes allow for the identification of large numbers of field collected immature whitefly specimens.

### **Technology Transfer**

Exotic parasitoid species and rearing technology were transferred to several commercial insectaries. Cooperative research with the beneficial insect industry to develop the use of mechanized parasitoid delivery systems for melon crops was conducted. Life-table studies along with toxicology studies on selective insecticides led to the development of more effective IPM in the cotton system in Arizona and elsewhere. Biochemical and molecular studies were developed to identify new and native species of whitefly parasitoids. An artificial diet for lacewings was developed and transferred to industry. The use of the new diet dramatically decreased costs of producing lacewings and made the use of lacewings to control whitefly in greenhouse crops more feasible. The new diet should also allow research on lacewing releases in field crops.

**Table D. Natural Enemy Ecology and Biocontrol.**

Research Approaches <sup>a</sup>	Year 1 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Natural control and conservation:</b>				
<b>Develop natural enemy conservation practices to reduce mortality to indigenous and introduced natural enemies.</b>	Conduct life table analyses of indigenous and introduced natural enemies to identify key mortality factors of natural enemy populations.	X		New insect growth regulators tested well under field conditions, and reduced loss of natural enemies. A Life Table analysis was conducted on natural enemies in cotton.
<b>Evaluate potential of alternate plants to act as in-field refuges or insectaries for natural enemies.</b>	Identify potential plants for natural enemy population development and assess risks of these plants to foster additional pest problems.	X		Combinations of annuals and some perennials show promise as within field natural enemy refugia. They are attractive to parasites but support low numbers of whiteflies. Annuals served as outdoor insectaries when releasing exotic parasitoids.
<b>Assess cues used by natural enemies to locate whitefly and to identify potential methods for enhancing natural enemy activity.</b>	Conduct laboratory tests to identify cues used by natural enemies to locate and attack whitefly.		X	Some research has been initiated but was not reported at this meeting.
<b>Augmentation of natural enemies:</b>				
<b>Develop natural enemy mass-rearing systems.</b>	Identify natural enemies with the highest potential for controlling whitefly in key cropping systems.	X		Diets are being developed for generalist predators. Improvements have been made in rearing parasitoids, increasing rearing efficiency. Field studies have identified promising candidates for augmentative releases
<b>Develop release technologies to maximize the effectiveness of mass-reared natural enemies in the field.</b>	Identify natural enemies with the highest potential for controlling whitefly in key cropping systems and that may be economically mass produced.	X		A novel “Banker Plant” field release strategy shows promise for increasing efficacy of releases. Releases of <i>Eretmocerus</i> into greenhouses controlled <i>Bemisia</i> attacking poinsettias when done at low pest densities.

**Table D. Natural Enemy Ecology and Biocontrol. (continued)**

Research Approaches <sup>a</sup>	Year 1 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Evaluate augmentative parasitoid, predator, or pathogen releases.</b>	Initiate studies on natural enemy augmentation with identified high potential natural enemies.	X		Augmentative releases of parasitoids controlled <i>Bemisia</i> in large demonstration fields. These releases can be integrated with conventional pest management practices
<b>Importation biological control:</b>				
<b>Evaluate the ability of exotic natural enemies to suppress whitefly populations under field conditions.</b>	Identify sites suitable for the release and subsequent evaluation of each candidate natural enemy. Conduct inoculative releases of natural enemies.	X		Combinations of annual plants that make excellent insectaries and can be farmed under local climatic conditions have been identified. Homeowners are being recruited to care for plants used for making releases
<b>Systematics, ecology, and population dynamics of natural enemies:<sup>b</sup></b>				
<b>Clarify sytematics of predators, parasitoids and pathogens.</b>	Conduct taxonomic studies of species within targeted releases sites. Verify taxonomic purity of mass-reared natural enemies. Complete taxonomic work on poorly characterized but important groups. Assist in determining most suitable natural enemies for release through biogeographical analysis.	X		Taxonomic studies have been completed on the exotic <i>Eretmocerus</i> and a key to their identification is in press. PCR techniques have been developed to identify the purity of cultures and aid in identification of recovered parasites.
<b>Determine <i>Bemisia</i>- natural enemy-host plant (Tritrophic) interactions.</b>	Initiate studies to identify mechanisms involved in <i>Bemisia</i> - and natural enemy plant attraction.	X		Controlled laboratory studies showed that <i>Bemisia</i> and aphelind oviposition rates varied depending on host plant.
<b>Identify the attributes of natural enemy biology and population level interactions to explain biological control successes and failures.</b>	Assess the value of the <i>Bemisia</i> biological control research to evaluate key issues to the science of biological control.	X		The role of autoparasitism in native populations of <i>Encarsia</i> and its impact on native <i>Eretmocerus</i> has been evaluated. Results from one study show no adverse affect of <i>Encarsia</i> on overall parasitism of SLWF

<sup>a</sup> See Table C for complementary research.

<sup>b</sup> See Table A for complementary research.

**Table D. Natural Enemy Ecology and Biological control.**

Research Approaches <sup>a</sup>	Year 2 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Natural control and conservation:</b>				
<b>Develop natural enemy conservation practices to reduce mortality to indigenous and introduced natural enemies.</b>	Evaluate predator gut contents. Conduct life table analysis.	X		Role of predators in cotton identified; importance of narrow spectrum insecticides highlighted.
<b>Evaluate potential of alternate plants to act as in-field refuges or insectaries for natural enemies.</b>	Determine refugia plant phenology in relation to cultivated crop phenology.	X		Perennial plants capable of growing in Imperial Valley identified, selected for a pilot project at a commercial organic farm.
<b>Assess cues used by natural enemies to locate whitefly and to identify potential methods for enhancing natural enemy activity.</b>	Determine potential methods for manipulating cues as part of a whitefly management program.		X	No work reported.
<b>Augmentation of natural enemies:</b>				
<b>Develop natural enemy mass-rearing systems.</b>	Determine nutritional, physiological, and ecological requirements for mass-rearing.	X		Whitefly, parasitized by <i>Encarsia</i> , were grown on an artificial diet long enough for parasitoids to emerge as adults. First such report. Potential for research and commercial rearing.
<b>Develop release technologies to maximize the effectiveness of mass-reared natural enemies in the field.</b>	Evaluate the fate of natural enemy life stages under field conditions to identify the appropriate developmental stage to be released.	X		First year results show banker plants may prove more efficacious than releases of parasitoids by hand. Two species of coccinellids evaluated, compared for greenhouse use.
<b>Evaluate augmentative parasitoid, predator, or pathogen releases.</b>	Conduct releases on selected crop systems at various rates of release.	X		Impact of <i>Beauveria bassiana</i> on generalist predators determined. Parasitoid dispersal was determined using new protein marking technique

**Table D. Natural Enemy Ecology and Biological Control. (continued)**

Research Approaches <sup>a</sup>	Year 2 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Importation biological control:</b>				
<b>Evaluate the ability of exotic natural enemies to suppress whitefly populations under field conditions.</b>	Evaluate establishment of exotic natural enemies within target release area. Determine if additional releases are necessary.	X		Several new exotics have persisted over several years and are multiplying and spreading in Texas and California.
<b>Systematics, ecology, and population dynamics of natural enemies:<sup>b</sup></b>				
<b>Clarify sytematics of predators, parasitoids and pathogens.</b>	Provide taxonomic support for importation and mass-rearing programs. Publish keys to assist in species identifications.	X		Key on exotic <i>Eretmocerus</i> published. Program developed for curating, cataloging recovered parasitoids.
<b>Determine <i>Bemisia</i>- natural enemy-host plant (Tritrophic) interactions.</b>	Study plant characteristics mediating whitefly and natural enemy population densities.	X		Parasitoid foraging, oviposition varied in response to different plants (crops) and host whitefly. Plants varied in color, architecture, and semiochemicals.
<b>Identify the attributes of natural enemy biology and population level interactions to explain biological control successes and failures.</b>	In conjunction with field evaluations, validate predictions made by behavioral and population models important to biological control.	X		No interference competition measured, with respect to whitefly control, when mixing primary parasitoids and autoparasitoids.

<sup>a</sup> See Table C for complementary research.

<sup>b</sup> See Table A for complementary research.

**Table D. Natural Enemy Ecology and Biological control.**

Research Approaches <sup>a</sup>	Year 3 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Natural control and conservation:</b>				
<b>Develop natural enemy conservation practices to reduce mortality to indigenous and introduced natural enemies.</b>	Conduct manipulative experiments to evaluate the impact of each natural enemy mortality agent on whitefly suppression.	X		Life history tables have been constructed comparing mortality factors of natural enemies in conventional vs IGR treated cotton. Functional response data available for several parasitoid species.
<b>Evaluate potential of alternate plants to act as in-field refuges or insectaries for natural enemies.</b>	Conduct field tests to assess whether refuges act of natural enemy and whitefly sinks or sources to adjacent cropping systems.	X		Research in the Imperial Valley has shown that perennial refuges support large numbers of whitefly and parasitoids that migrate to adjacent systems.
<b>Assess cues used by natural enemies to locate whitefly and to identify potential methods for enhancing natural enemy activity.</b>	Conduct small scale trials to enhance whitefly suppression by manipulating natural enemy location and attack of whitefly .		X	
<b>Augmentation of natural enemies:</b>				
<b>Develop natural enemy mass-rearing systems.</b>	Develop rearing systems on selected hosts and on artificial diets. Determine economic feasibility of the procedure.	X		Mass rearing methods on SLWF has been accomplished. Artificial diets are still being researched, with economics undetermined.
<b>Develop release technologies to maximize the effectiveness of mass-reared natural enemies in the field.</b>	Develop necessary technology for release of the appropriate natural enemy life stage.	X		Several release technologies have or are being tested. Banker plant technology appears to be very effective. Capsule delivery methods being tested. Cold storage of parasitoid pupae also being tested
<b>Evaluate augmentative parasitoid, predator, or pathogen releases.</b>	Identify optimal release strategies for key cropping systems.	X		Parasitoid release rates have been determined for major crops. Strategies for releasing/integrating parasitoid and predator in greenhouse crops have been determined. Significant information currently available on application of fungal pathogens in various crops.

**Table D. Natural Enemy Ecology and Biological Control. (continued)**

Research Approaches <sup>a</sup>	Year 3 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Importation biological control:</b>				
<b>Evaluate the ability of exotic natural enemies to suppress whitefly populations under field conditions.</b>	Assess spread of established natural enemies and their ability to suppress whitefly populations	X		Whitefly suppression by exotic parasitoids determined by multiple researchers. Preliminary data suggests significant spread of established exotic parasitoids in some regions. Dispersal rates of natural enemies still under study.
<b>Systematics, ecology, and population dynamics of natural enemies:<sup>b</sup></b>				
<b>Clarify sytematics of predators, parasitoids and pathogens.</b>	Provide taxonomic support for importation and mass-rearing programs.	X		Several taxonomic keys developed for imported parasitoid species. RAPD-PCR techniques proven as quick identification method. Preliminary Satellite DNA techniques proven, however, still under development.
<b>Determine <i>Bemisia</i>- natural enemy-host plant (Tritrophic) interactions.</b>	Study compatibility of characteristics of plant traits mediating whitefly populations with the abilities of natural enemies to suppress whitefly populations.	X		Tri-trophic interactions determined for <i>B. bassiana</i> / SLWF / tomato. Some research completed on parasitoid / host / plant interactions.
<b>Identify the attributes of natural enemy biology and population level interactions to explain biological control successes and failures.</b>	Assess deviations between theoretical predictions and field data.	X		Some life history data collected on parasitoid and predator populations in cotton. BioControl-Parasite simulation model available for testing / validation. Some Laboratory data available for testing theoretical predictions of field level performance.

<sup>a</sup> See Table C for complementary research.

<sup>b</sup> See Table A for complementary research.



**Table D. Natural Enemy Ecology and Biological control, 2001-2002.**

Research Approaches <sup>a</sup>	Year 4/5 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Natural control and conservation: Develop natural enemy conservation practices to reduce mortality to indigenous and introduced natural enemies.</b>	Conduct a feasibility study and economic assessment of altered crop management practices that may enhance the impact of indigenous natural enemies.	X		IGR's compared to conventional insecticides reduce negative impacts on natural enemies. Pathogenic fungi have been shown to be of use. Annual refuge plantings are of limited value, however, perennial plant refuges are promising as a source of parasitoids while providing minimal source populations of SLW.
	Develop and evaluate area wide programs to facilitate full implementation.	X		
<b>Evaluate potential of alternate plants to act as in-field refuges or insectaries for natural enemies.</b>	Conduct field tests to evaluate spacing of refuges necessary to achieve satisfactory whitefly suppression.	X		Spacing studies have not been conducted, however, other field studies indirectly provide information pertaining to this topic, including several parasitoid movement studies.
	Conduct a feasibility study and economic assessment of alternate plantings in terms of an entire crop management program.		X	
<b>Assess cues used by natural enemies to locate whitefly and to identify potential methods for enhancing natural enemy activity.</b>	Conduct large scale field trials and evaluate product development for commercial investment as necessary.		X	No, however, recent work has been done on chemically-based intraspecific communication cues between parasitoid individuals.
	Transfer technology (as needed) to commercial interests for full implementation.		X	
<b>Augmentation of natural enemies: Develop natural enemy mass-rearing systems.</b>	Evaluate rearing system effects on natural enemy life history characteristics, behavior, and ability to suppress whitefly populations.	X		Parasitoid development on <i>Bemisia</i> produced on artificial diet has been examined. Predator fitness when produced on artificial diets has been studied. Additional work in this area, including large scale production using plant reared <i>Bemisia</i> to rear parasitoids is in need of further study.
	Facilitate transfer of mass-rearing technology to commercial interests as necessary.	X		

**Table D. Natural Enemy Ecology and Biological Control, 2001-2002. (continued)**

Research Approaches <sup>a</sup>	Year 4/5 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Develop release technologies to maximize the effectiveness of mass-reared natural enemies in the field.</b>	Evaluate re lease technology effects on natural enemy life history characteristics, behavior, and ability to suppress whitefly populations.	X		Tractor pulled, parasitoid pupal drop box release method needs further refinement. Technique developed using “Banker plants” with parasitoid pupae, to inoculate crops. Several container based release methods for pupae were evaluated and the best methods identified.
	Facilitate transfer of mass-rearing technology to commercial interests as necessary.	X		Contractual agreements made to provide commercial insectaries with several promising parasitoid species for field and greenhouse augmentation.
	<b>Evaluate augmentative parasitoid, predator, or pathogen releases.</b>	Continue evaluation of releases, determine need for additional releases. Compare results in different cropping systems and environments.	X	
	Analyze information and make recommendation regarding need for expansion of the approach.	X		Studies in greenhouse crops completed and analyzed. Recommendations have been developed. Extensive evaluation of augmentative biocontrol in spring cantaloupe fields using parasitoids has occurred. Analysis nearly completed.
<b>Importation biological control: Evaluate the ability of exotic natural enemies to suppress whitefly populations under field conditions.</b>	Continue to assess the spread of established natural enemies and their ability to suppress whitefly populations. Evaluate program progress and determine if additional strategies are necessary	X		Extensive assessment conducted in CA., TX, AZ, & FL. Long term field plots, sentinel plants, area wide field surveys, and life table methods used. One to three parasitoid species established in CA , TX and AZ. Establishment process is ongoing, probably taking several more years. Results to date look promising in several regions.
	Complete program analysis. Publish program assessment and conduct an economic assessment.	X		Book in preparation, K. Hoelmer and J. Gould editors.

**Table D. Natural Enemy Ecology and Biological Control, 2001-2002. (continued)**

Research Approaches <sup>a</sup>	Year 4/5 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Systematics, ecology, and population dynamics of natural enemies:<sup>b</sup></b>				
<b>Clarify systematics of predators, parasitoids and pathogens.</b>	Provide taxonomic support for importation and mass-rearing programs.	X		Numerous publications developed for <i>Eretmocerus</i> and <i>Encarsia</i> spp. identification. Extensive development and availability of molecular based identification methodology.
<b>Determine <i>Bemisia</i>- natural enemy-host plant (Tritrophic) interactions.</b>	Assess the implementability of favorable tritrophic interactions within the context of an whitefly management program.	X		Relationships among leaf surface characteristics, SLW and parasitoid performance identified in several instances. Associations identified between plant species and parasitoid activity.
	Implement and evaluate large scale crop management programs for suppression of whitefly populations.	X		Crop vulnerability windows identified.
<b>Identify the attributes of natural enemy biology and population level interactions to explain biological control successes and failures.</b>	Evaluate behavioral or population level parameters that may explain observed deviations	X		Life table analysis, laboratory and field cage studies of interspecific parasitoid interactions, field cage and in-field sex ratio studies, and studies of parasitoid indosymbionts were conducted.
	Quantify the impact of basic research on the development of feasible biological control programs for <i>Bemisia</i> and the advancement of the field as a science.		X	Artificial diets developed for mass rearing several predators. Although an artificial diet for SLW is unlikely to support mass rearing of parasitoids, its development has been very useful for conducting detailed biological studies. Parasitoid movement studies have provided a basis for developing augmentative release approaches and interpreting exotic parasitoid establishment data.

<sup>a</sup> See Table C for complementary research.

<sup>b</sup> See Table A for complementary research.

## Reports of Research Progress

### Section E: Host-Plant Resistance, Physiological Disorders, and Host-Plant Interactions

Co-Chairs: Greg Walker and Cindy McKenzie

**Investigator's Name(s):** Shaaban Abd-Rabou<sup>1</sup> & Alvin M. Simmons<sup>2</sup>

**Affiliation & Location:** <sup>1</sup>Germain Foundation for International Development, Food and Agricultural Development Center, Leipzig, Federal Republic of Germany and <sup>2</sup>USDA-ARS, U.S. Vegetable Laboratory, Charleston, SC

**Research & Implementation Area:** Section E: Host Plant Resistance, Physiological Disorders, and Host-Plant Interactions

**Dates Covered by the Report:** 2001

#### Parasitism of *Bemisia tabaci* on Numerous Species of Host Plants

The influence of numerous vegetable and other agronomic plant species on incidence of parasitism of the B-biotype sweetpotato whitefly, *Bemisia tabaci* (Gennadius), by augmentation with parasitoids was determined in field plots. Tests were conducted on 16 taxonomically diversified plant species (*Beta vulgaris* L., *Brassica oleracea* var *botrytis* L., *Brassica oleracea* var *capitata* L., *Citrullus lanatus* (Thunberg) Matsum. & Nakai ssp. *lanatus*., *Cucumis melo* L., *Cucumis sativus* L., *Glycine max* (L.) Merrill, *Gossypium barbadense* L., *Helianthus annuus* L., *Ipomea batatas* L., *Lantana camara* L., *Lycopersicon esculentum* Miller, *Phaseols vulgaris* L., *Solanum melongena* L., *Solanum tuberosum* L., and *Vigna sinensis* L.). Parasitism in plots with feral infestations by *B. tabaci* was evaluated through augmentations with *Eretmocerus mundus* Mercet from a laboratory colony, and comparisons were made with check plots in which no parasitoid releases were made. Plots were set up at five locations in Egypt (Beihera, Beni-Suef, Kafr ElShikh, Minufiya, and Qalyubiya). Each plot (0.13 hectares) contained a single plant species. Adult *E. mundus* were released during each of 15 weeks in treatment plots, and parasitism data were collected weekly over 15 weeks. The release rate was 5-12 parasitoids per plant and the releases were done using vials of parasitoids that were attached to the plants. Parasitism was enhanced in all plots where augmentations were made. In some plots, e.g., both species of *Brassica*, *B. vulgaris*, and *G. max*, overall parasitism was enhanced at a relatively high rate while in other plots, e.g., *C. lanatus*, *S. melongena*, and *L. esculentum*, the enhancement of overall parasitism was relatively low. In both treated and untreated plots for all plant species, parasitism peaked 7-12 weeks after the first augmentation date. Results from this study describe the relative seasonal abundance and relative degree of augmentative enhancement of parasitism of *Bemisia* among numerous plant species of economic importance.

**Investigators' Name(s):** Jiang Chen, Heather J. McAuslane, R. Bruce Carle, and Susan E. Webb

**Affiliation & Location:** Dept. of Entomology & Nematology, Univ. of Florida, Gainesville, FL

**Research & Implementation Area:** Section E: Host Plant Resistance, Physiological Disorders, and Host-Plant Interactions

**Dates Covered by the Report:** Jan. – Dec. 2000

### **Effect of *Bemisia argentifolii* and Squash Silverleaf Disorder on Zucchini Plant Growth and Yield**

Squash silverleaf disorder is a serious systemic physiological disorder of *Cucurbita* spp. Previous studies have shown that large whitefly populations can reduce plant vigor, growth and yield. However, we had no idea of the *Bemisia argentifolii* infestation levels that would cause damage to zucchini plants. Heavy whitefly feeding and expression of squash silverleaf disorder are two factors that may cause zucchini plants to become stunted, weak and lose yield. Two closely-related zucchini breeding lines were selected to separate these two factors; ZUC61 is silverleaf susceptible, whereas ZUC76-SLR is silverleaf resistant. Experiments to test the effects of whitefly infestation levels on zucchini seedling development and mature plant fruit yield were conducted in the greenhouse and outdoor screened rooms, respectively, in the spring and fall of 2000. In the greenhouse experiments, zucchini seedlings were infested with 0, 15, 30 or 60 pairs of *B. argentifolii* adults at the 2-leaf stage in the spring (Jan. 28 – Mar. 20), and with 0, 30, 60 or 90 pairs in the fall (Aug. 24 – Oct. 6). Whiteflies were confined on plants and allowed to feed and oviposit for 72 h. Whitefly eggs remained on plants to develop to nymphs. All whitefly-infested leaves, usually the 3-4 oldest ones, were removed immediately before whitefly adults emerged. Plant dry weight and height, and petiole length of silvered leaves were measured to evaluate zucchini plant growth 14 d after whitefly infested leaves were removed. Our results indicated that the progeny of 60 pairs of whiteflies significantly stunted ZUC61 and ZUC76-SLR zucchini seedling growth in the spring (13% dry weight reduction), whereas as few as 30 pairs and their progeny did so in the fall (9 % dry weight reduction). Plant height and the petiole length of silvered leaves of ZUC61 and ZUC76-SLR were also shortened after whitefly infestation. The effect of whitefly nymphal feeding continued at least 2 weeks after whitefly-infested leaves were removed. The silverleaf severity of ZUC61 did not differ significantly among the three treatment infestation levels in both spring and fall experiments. However, significant dry weight reduction only appeared at the highest infestation level 60 in the spring experiment. In the fall experiment, significant plant dry weight reduction appeared at all treatment infestation levels. Plant dry weight was reduced more at infestation level 90 (16%) than at infestation level 30 (9%). Although silverleaf symptoms were nonexistent or slight (grade 1) on ZUC76-SLR leaves, this breeding line experienced the same reduction in seedling growth as ZUC61.

In the screen room experiments, we used four whitefly infestation levels of 0, 30, 60 and 120 pairs of adults in both spring (Apr. 8 – June 3) and fall (Aug. 23 – Oct. 24) seasons. Whiteflies were released onto zucchini plants at the 2-leaf stage once a week for 3 weeks. Silverleaf severity of the 4<sup>th</sup> leaf (length > 8 cm) from the shoot apex and the leaves immediately above and below it was graded once a week until the end of the experiments. Female flowers were pollinated manually. Fruits were harvested at 16-17 cm in length in the spring, and at 96 h after pollination in the fall. The harvest was continued for 4 weeks from the beginning of fruit production in each season. Serious silverleaf symptoms of grade 4-5 were expressed on all whitefly-infested ZUC61 plants, whereas slight silverleaf symptoms of grade 1-3 were expressed on ZUC76-SLR plants. However, the silverleaf severity of each zucchini breeding line did not differ significantly among treatment infestation levels in both spring and fall experiments. Zucchini fruit number was significantly decreased at whitefly infestation level 60 for ZUC61 plants and at infestation level 120 for ZUC76-SLR plants in the spring. Fruits at infestation levels 60 and 120 developed more slowly and took approximately 1 more day to reach the same length as fruits from control plants. Fruit length from plants under these two whitefly infestation levels was shorter than that of control fruits on the 3<sup>rd</sup> d after pollination. However, fruits from all infestation levels weighed almost the same when they reached the same harvestable size. All plants at infestation levels 60 and 120 died toward the end of the experiment in the fall, but no plants died at any whitefly infestation levels in the spring. No fruit was produced on plants at infestation level 120, and only five fruits were harvested from 14 plants at infestation level 60 in the fall. Significant fruit yield reduction appeared at infestation level 30 for both ZUC61 and ZUC76-SLR breeding lines.

Based on these studies, we concluded that the stunting of zucchini seedling growth and yield reduction were mainly induced by the feeding of high whitefly populations, not by expression of silverleaf symptoms.

**Investigator's Name(s):** Yun-Shu Chen, Wilhelmina T.G. van de Ven, and Linda L. Walling

**Affiliation & Location:** Department of Botany and Plant Science, University of California, Riverside

**Research & Implementation Area:** Section E. Host Plant Resistance, Physiological Disorders and Host-plant Interaction

**Dates Covered by the Report:** January 2000-2002

### **Expression of *SLW3* (silverleaf whitefly-induced) Gene in the Transgenic Plants**

Two silverleaf white-induced genes, *SLW1* and *SLW3*, were isolated by differential RNA display. The transcripts of *SLW* genes accumulated differentially by silverleaf whitefly (SLWF) and sweetpotato whitefly (SPWF). After SLWF nymph feeding, *SLW1* and *SLW3* RNAs accumulate in the apical, non-infested leaves. In contrast, neither gene was induced by SPWF in apical leaves. Both *SLW* transcripts were accumulated in the local, infested leaves of SLWF-infested plants; however, only *SLW3* was induced in the local leaves of SPWF-infested leaves. The expression of *SLW1* was detected during flower and fruit development, while *SLW3* RNAs were not. The regulation of *SLW1* was involved in the two plant defense signals, jasmonic acid and ethylene, but the defense signals that regulate *SLW3* are still unknown. Both *SLW* genes were induced by the water deficit but not by other stresses such as wounding and pathogen infection. Database searches indicated that *SLW1* and *SLW3* encode a M20B-like peptidase and  $\beta$ -glucosidase, respectively; however their roles in the plant defense response to the SLWF infestation are still not understood. The reverse genetic approach has been used for investigating the expression and function of *SLW* genes in the plant response to SLWF feeding. The *SLW3:GUS* transgenic tomato and *Arabidopsis* have been generated and the *SLW3* expression studies under abiotic and biotic stresses and during development are ongoing. In addition, transgenic plant lines ectopically expression *SLW3* have been constructed.

**Investigator's Name(s):** <sup>1</sup>C. C. Chu, <sup>2</sup>E. T. Natwick, <sup>3</sup>J. S. Buckner, <sup>3</sup>D. R. Nelson, <sup>4</sup>T. P. Freeman, <sup>1</sup>T. J. Henneberry

**Affiliation & Locations:** <sup>1</sup>USDA, ARS, Western Cotton Research Laboratory, Phoenix, AZ; <sup>2</sup>University of California Coop. Extension, Holtville, CA, <sup>3</sup>USDA, ARS, Biosciences Research Laboratory, Fargo, ND, and <sup>4</sup>Electron Microscopy Center, North Dakota State University, Fargo, ND

**Research & Implementation Area:** Section E: Host-Plant Resistance, Physiological Disorders, and Host-Plant Interactions

**Dates Covered by the Report:** 2000-2001

#### **Silverleaf Whitefly Populations and Trichome Densities of Upland Cottons**

The study was conducted in 2000 at the University of Arizona Agricultural Research Center, at Maricopa, AZ. Cotton cultivars were five normal-leaf cotton cultivars (Deltapine [DPL] 20B, DPL 50 B, DPL 90B, NuCOTN 33B, and Stoneville [ST] 474) and four okra-leaf cultivars (E0223, E0798, E1028, and Siokra L-23). All entries were smooth leaf cultivars except for the hairy -leaf ST 474. On average, ST 474 had 71.3 stellate trichomes/cm<sup>2</sup> of leaf disk and other cultivars fewer than 1.0 stellate trichomes/cm<sup>2</sup>. ST 474 had 12.9 adults per leaf-turn count and other cultivars had fewer than 6.2 adults. ST 474 had 36.0 eggs/cm<sup>2</sup> and 13.3 nymphs/cm<sup>2</sup> leaf disk. Other cultivars had fewer than 15.4 eggs/cm<sup>2</sup> and 6.0 nymphs/cm<sup>2</sup> leaf disk. On average of all cultivars studied, leaves on the top main stem leaf node (#1) had the highest number of stellate trichomes but the lowest numbers of eggs, nymphs and adults compared with leaves on nodes #2, #3, #4, #5 or #7.

**Investigator's Name:** Tong-Xian Liu

**Affiliation & Location:** Texas Agricultural Experiment Station, Texas A&M University System  
2415 E. Highway 83, Weslaco, TX 78596-8399

**Research & Implementation Area:** Section E: Host Plant Resistance, Physiological Disorders, and Host-Plant Interactions

**Dates Covered by the Report:** 2000

### **Field Evaluations of the Six Selected Cantaloupe Varieties for Silverleaf Whitefly Resistance**

Silverleaf whitefly, *Bemisia argentifolii* Bellows & Perring, continues to be the most important pest insect for cucurbits, in south Texas. In the past few years, many varieties, lines, and PIs had been studied under field conditions for whitefly resistance in south Texas. It has been reported that the glabrous leafed varieties (lines) are highly resistant to whitefly. Unfortunately, the yield and quality of these smooth leafed varieties or lines are not commercially desirable. Results from the laboratory experiment for the six selected varieties indicated that among the six varieties, Tam Sun, Tam Sun x gl., and Hymark show some degree of resistance to the silverleaf whitefly, whereas other varieties did not show significant differences. The resistance of these varieties to the silverleaf whitefly under the field condition will confirm and verify the results from the laboratory study. The objective of this study was to determine the resistance of six selected varieties to silverleaf whitefly under field conditions to verify the data obtained from the laboratory study. Six varieties and lines were used in this study: Hymark, Tam Sun x gl, Explorer, Tam Sun, Primo, and Perlita. The seedlings (10-11 cm high) were transplanted in the field in middle or late January. Each plot was 30 ft (12 m) long with two separate rows 80 in (2 m) wide, and 10-20 plants each. The plots were arranged in a randomized complete block design with 4 replications. Adult sampling was conducted when whitefly population increase significantly. Thereafter, plants were sampled in 7-day intervals. Adults were counted by leaf turn method. When plants had <6 leaves, adults on the oldest leaf were counted, and when plants had >6 leaves, adults on the 4<sup>th</sup> or 5<sup>th</sup> leaf from the proximal were counted. Pupae (re-eyed nymphs), empty pupal cases per 4 leaf-discs per leaf were counted. Yield, sugar content and foliage damage (by sooty molts) were also evaluated.

Whitefly adult population was relatively low on the plants in all 6 varieties before late March, and increased gradually to mid April. Numbers of adults increased rapidly, and reached the peak in late April and early May. Among the six varieties, numbers of adults per leaf were significantly different. The variety that had the least numbers of whitefly adults throughout the season was TP45, a Weslaco line, followed by Tam Sun, a TAES variety. The variety that had the greatest number of whitefly adults was Primo, as many as 660 adults per leaf were found on 3 May. Number of adults on Impact and Hymark were also relatively high, with >300 adults per leaf in late April and early May. Significant differences were found for both the adults and nymphs among the six varieties. Primo had the most and TP45 had the fewest whitefly adults, and difference between the two varieties was >8 folds. Primo, Hymark and Impact had more nymphs than other varieties, and again, TP45 had the fewest number of nymphs among the six varieties. Tam Sun was the only variety that has fewer melons than other varieties; whereas the total numbers of melon harvested from other five varieties were not significantly different. Impact and TP45 had more larger-melons compared with other varieties; whereas Tam Sun x gl. and Impact had the most numbers of small melons. TP45 not only had the most numbers of melons, but also had the most large-melons (Table 4). Impact also had relatively greater weight, which, however, was not significantly different from Hymark, Tam Sun x gl., and Primo. Total weight for Tam Sun was the least, although it was not significantly different from Hymark and Tam Sun x gl. Results from this study indicated that Significant differences in responses to silverleaf whitefly infestation were found among the six cantaloupe varieties. TP45, a Weslaco line, had the fewest whiteflies (adults and red-eyed-nymphs or pupae) compared to other five varieties. TP45 also had the highest yield (more and larger melons). However, melons of TP45 had the lowest sugar contents (3.80-3.86%) compared 7.6-9.5% in other varieties. Other varieties which had fewer silverleaf whiteflies were Tam Sun and Tam Sun x gl. Primo, Impact and Hymark had more silverleaf whitefly than others. However, those varieties had higher yield and sugar contents than others. Similar results were found in the laboratory experiment that Tam Sun, Tam Sun x gl., and Hymark show some degree of resistance to the silverleaf whitefly.



**Investigator's Name(s):** Heather J. McAuslane

**Affiliation & Location:** Dept. of Entomology & Nematology, University of Florida, Gainesville, FL

**Research & Implementation Area:** Section E: Host Plant Resistance, Physiological Disorders, and Host-Plant Interactions

**Dates Covered by the Report:** 1995-2000

### **Mechanisms and Genetics of Resistance to Squash Silverleaf Disorder in *Cucurbita* spp. Squash**

Squash silverleaf disorder (SSL) is a serious physiological disorder of commercial *Cucurbita* species associated with the feeding of nymphal *Bemisia argentifolii*. Screening of the National Plant Germplasm System repository of *C. moschata*, *C. pepo*, and *C. maxima* yielded accessions that showed no SSL symptoms in field trials in Mayaguez, Puerto Rico and Leesburg, FL, where other accessions around them silvered heavily. Field and greenhouse studies of the mechanisms of resistance indicated that although SSL-resistant germplasm did not silver, plants received similar levels of oviposition by whiteflies and supported similar whitefly survival and rates of development as did SSL-susceptible accessions. The mechanism of resistance in two *C. pepo* SSL-resistant lines (ZUC76-SLR and ZUC33-SLR/PMR) and two *C. moschata* lines (UP9606-13-4 and UP9706-3-1) was tolerance to the effects of whitefly feeding. Controlled infestation of SSL-resistant and susceptible germplasm with similar whitefly populations resulted in no silvering in resistant lines and grade 5 SSL symptoms in susceptible lines.

Silvering of leaves begins at the apex of young leaves of *C. pepo* and proceeds basipetally to cover the whole leaf. Chloroplasts are smaller in silvered leaves even before macroscopically visible symptoms develop. Silvering results from an abnormal air space between the upper epidermis and the upper palisade layer of cells. Leaf cross-sections reveal that the development of the silvering occurs as normal air space formation is occurring. Cell separation between epidermis and palisade cells is not a consequence of differential cell division or expansion between the two layers because the number of cells per mm in each layer over time is the same in whitefly-infested plants and controls. Air space formation is more likely a result of increased cell wall degradation during normal air space formation. Leaves of SSL-resistant ZUC76-SLR showed no anatomical response to whitefly feeding but did have lighter green and shorter petioles as did susceptible plants. In grafts of the resistant plants onto susceptible plants challenged with whitefly nymphs, the resistant plants did not develop leaf silvering. Susceptible scions grafted onto resistant rootstocks did exhibit leaf silvering. These experiments indicate that the site of SSL resistance is at the young developing leaves and not at the mature leaf where insect feeding occurs. Susceptible and resistant *C. pepo* germplasm responded systemically to application of chlormequat chloride on mature leaves or as soil drenches with shortened internodes and silverleaf-like symptoms on new foliage. Petioles and new leaves of ZUC61 and ZUC76-SLR showed similar reductions in chlorophyll content after whitefly feeding on mature leaves.

Greenhouse and outdoor screen room studies indicated that zucchini seedlings were stunted when infested with whiteflies and infested plants yielded less fruit than uninfested plants. To separate the effects of whitefly feeding versus expression of SSL symptoms on growth and yield reduction, two closely related breeding lines were infested with several levels of adult whiteflies; ZUC61 was SSL-susceptible and ZUC76-SLR was resistant (see abstract by Chen et al., section E). Both ZUC61 and ZUC76-SLR seedlings were stunted by feeding of 30-60 pairs of adult whiteflies and their progeny for 2 weeks, showing ca. 10% reduction in dry weight. ZUC61 plants showed grade 5 SSL symptoms at these infestation levels whereas ZUC76-SLR had no or mild (grade 1) symptoms. Season-long feeding by 60-120 pairs of whiteflies and their progeny caused significant yield and quality reduction in fruit for both ZUC61 and ZUC76-SLR. Seedling stunting and yield reduction appear to be more related to intensity of whitefly feeding than to susceptibility to SSL. The genetics of resistance to SSL have been investigated in *C. moschata* and *C. pepo* germplasm. In *C. moschata*, SSL resistance appears to be related to the presence of natural leaf mottling. The recessive gene for silverleaf resistance appears to work only in combination with the lack of natural leaf mottling. Leaf mottling is conditioned by a single partially dominant gene (M), thus only those genotypes with *ssmm* are resistant to SSL. In *C. pepo*, SSL resistance appears to be conditioned by two genes, one recessive and one incompletely dominant. In both *C. moschata* and *C. pepo*, the severity of SSL symptoms can be modified by whitefly infestation level and environmental conditions, such as light intensity and temperature. I gratefully acknowledge the important contributions of graduate students, Yasmin Cardoza and Jiang Chen, and collaborators, Drs. Judy Schmalstig, Bruce Carle, Linda Wessel-Beaver, and Susan Webb. Research was supported by USDA Special Research Topic grants FLA-ENY-03443 and -03721.

**Investigators Name(s):** E. T. Natwick<sup>1</sup>, C. G. Cook<sup>2</sup>, R. L. Gilbertson<sup>3</sup>, Young-Su Seo<sup>3</sup> and T. Turini<sup>1</sup>

**Affiliation & Location:** <sup>1</sup>University of California Coop. Ext., Holtville, CA, <sup>2</sup>Syngenta, Santa Rosa, TX, and <sup>3</sup>University of California Davis, Davis, CA

**Research & Implementation Area:** Section E: Host Plant Resistance, Physiological Disorders, and Host-Plant Interactions

**Dates Covered by Report:** March 2000 through December 2000

### **Resistance To Whitefly Transmitted Cotton Leaf Crumple Disease**

Ten cotton, *Gossypium hirsutum* L., cultivars or breeding-lines were evaluated in the field for resistance to the cotton leaf crumple (CLCr) disease caused by (Genus *Begomovirus*, Family *Geminiviridae*) *Cotton leaf crumple virus* (CLCrV) transmitted by silverleaf whitefly, *Bemisia argentifolii* Bellows and Perring. The cultivars Texas 121, AP 4103, AP 6101 and Stoneville 474 and the breeding-lines were NK 2165C, NK 2108SS, NK 2387C, NKX C429-93-2ct, NKX 2907, and NKX 2207 were in Imperial Valley, CA. Cotton entries were rated for severity of CLCr disease symptoms and the presence of CLCrV in leaves of selected plants of each cultivar/breeding-line was determined by dot blot hybridization with a CLCrV DNA probe and PCR analysis with degenerate geminivirus primers. DNA sequencing of geminivirus DNA-A and DNA-B fragments, amplified from symptomatic cotton plants, was used to confirm geminivirus infection and partial characterize CLCrV. Results showed differences in whitefly infestation levels and virus disease symptoms among cotton entries. The cultivar AP 4103 had a higher CLCr disease rating than other entries except AP 6101. The breeding-line NK 2387C, with Cedix parentage, had a lower CLCr disease rating than other entries except Stoneville 474 and NK X2207. There were visible CLCr disease symptoms in Stoneville 474 and NKX 2207, but KN 2387C did not display visible CLCr disease symptoms nor was viral DNA detected in this line.

**Investigators Name(s):** E. T. Natwick<sup>1</sup>, G. Walker<sup>2</sup>, C. C. Chu<sup>3</sup>, T. J. Henneberry<sup>3</sup>, D. Brushwood<sup>4</sup>, and G. Constable<sup>5</sup>

**Affiliation & Location:** <sup>1</sup>University of California Cooperative Extension, University of California Desert Research and Extension Center, 1050 E. Holton Road, Holtville, CA 92250, <sup>2</sup>University of California, Riverside, CA, <sup>3</sup>USDA-ARS, Western Cotton Research Laboratory, Phoenix, AZ, <sup>4</sup>USDA-ARS Cotton Quality Research Station, Clemson, SC and <sup>5</sup>CSIRO, Narrabri, NSW Australia

**Research & Implementation Area:** Section E: Host Plant Resistance, Physiological Disorders, and Host-Plant Interactions

**Dates Covered by Report:** January 2000 through December 2000

#### **Susceptibility of Upland Cotton Cultivars to Infestation by Silverleaf Whitefly.**

Sixteen upland cotton, *Gossypium hirsutum* L., cultivars and experimental breeding-lines were evaluated in the field for susceptibility to silverleaf whitefly, *Bemisia argentifolii* Bellows and Perring, sown at the UC Desert Research & Extension Center, Imperial Valley, CA, into plots of a randomized complete block design experiment replicated four times, and irrigated 24 March 2000. The normal leaf cultivars were DP 20, DP 50, DP 90, DP 5415, DP 5432, DP 5461, DP 5557, HCR 9257, HCR 9240, and Stoneville 474 and the okra-leaf cultivars and experimental breeding-lines were Siokra L23, FiberMax 832, FiberMax 819, E1028, E0798 and E0223. Individual plots measured 14 m in length with 8-beds on 1 m centers or 8m wide. Silverleaf whitefly adults were sampled from ten plants at random in each plot via the leaf tum method using the 5th main stem leaf from the terminal. Silverleaf whitefly nymphs were counted on 1.65 cm<sup>2</sup> leaf disks of from ten 5th position leaves down from the terminal extracted from randomly selected plants in each plot. Adults and nymphs were sampled on 23 & 29 May, 5, 12, 19, & 26 June, 3, 10, 17, 24 & 31 July, 7, & 14 August, 2000. on 30 June, 7, 14, 20, 28 July, 4, 11, 18, & 25 August. Seed cotton was hand picked from 0.002 acre per plot and yield data were recorded. The okra-leaf entries as a group had fewer silverleaf whitefly adults and nymphs than the normal leaf cotton entries. The okra-leaf experimental breeding-lines had the lowest numbers of silverleaf whitefly adults and nymphs among the okra-leaf entries. Stoneville 474, a hirsute leafed cotton, had the greatest numbers of silverleaf whitefly adults and nymphs among the normal leaf cottons. ?There were no differences in seed cotton yield among the entries, p = 0.05, SNK?.

**Investigator's Name(s):** E. T. Natwick<sup>1</sup> & G. P Walker<sup>2</sup>

**Affiliation & Location:** <sup>1</sup>University of California Coop. Ext., Holtville, CA, <sup>2</sup>Department of Entomology, University of California, Riverside, CA

**Research & Implementation Area:** Section E: Host-Plant Resistance, Physiological Disorders, and Host Plant Resistance

**Dates Covered by Report:** May 2000 through September 2001

### **High Level of Resistance to Silverleaf Whitefly in the Cotton Relative, *Gossypium thurberi***

In 1993, F. D. Wilson et al. from USDA's Western Cotton Research Laboratory reported a high degree of resistance to silverleaf whitefly, *Bemisia argentifolii*, in the cotton relative, *Gossypium thurberi*. *G. thurberi* has been successfully crossed with commercial cotton in the past, and therefore could serve as a source of whitefly resistance genes that could be introduced into commercial cotton. The study presented here was initiated to verify the results of Wilson et al. (1993) and to determine the degree of resistance of *G. thurberi* to silverleaf whitefly.

#### **Materials & Methods**

Two field studies (one in 2000, the other in 2001) with a randomized complete block design were conducted at the University of California Desert Research and Extension Center in Holtville, CA. The preliminary test in 2000 had two entries: *G. thurberi* and DP 5415 of Delta and Pine Seeds. The 2001 test had four entries: *G. thurberi*, DP 5415 of Delta and Pine Seeds, Siokra L23 of Cotton Seed Distributors Ltd. of Australia, and Stoneville 474 of Stoneville Pedigree Seed Company. On each sample date, whitefly adults (*Bemisia argentifolii*) were sampled using the leaf turn method. Samples of ten 5<sup>th</sup> node leaves per plot were excised from plants at random on the same dates adult whitefly counts were taken. Whitefly eggs and nymphs were counted on single leaf disks of 1.65 cm<sup>2</sup> taken from the lower left quadrant from each leaf.

In an additional test, greenhouse grown plants were used for experiments comparing nymphal survival and development time of *Bemisia argentifolii* on *G. thurberi* and DP 5415. Plants were young seedlings at the 2-4 leaf stage.

#### **Results and Discussion**

The 2000 field experiment demonstrated that *G. thurberi* had a high level of resistance against silverleaf whitefly when compared to DP 5415. *G. thurberi* has okra leaf shape and DP 5415 has normal shaped leaves. Okra leaf shape has sometimes been associated with whitefly resistance; consequently the 2000 field test was unable to determine whether the resistance in *G. thurberi* was due to okra leaf shape or due to a novel factor. Both *G. thurberi* and DP 5415 have smooth leaves, a trait associated with reduced whitefly susceptibility.

The 2001 field experiment included three commercial cotton cultivars including the okra-leaf Siokra L23. *G. thurberi* exhibited a high level of resistance against silverleaf whitefly when compared to all three commercial cultivars, including the okra-leaf Siokra L23. Siokra L23 and DP 5415 also are smooth-leaf varieties while Stoneville 474 has hairy leaves. The resistance of *G. thurberi* relative to the commercial smooth-leaf and okra-leaf cultivars suggests that *G. thurberi* has a novel mechanism of resistance against silverleaf whitefly.

The greenhouse results starkly contrast the field results. Silverleaf whitefly performance, measured as nymphal survivorship and developmental time, was slightly better on *G. thurberi* than on DP 5415. We suspect that the contrasting greenhouse and field study results are due to the use of greenhouse grown seedling plants that had not hardened off. We plan to test this hypothesis in the oncoming season.

Despite the anomaly of the greenhouse experiment, the data strongly suggest that *G. thurberi* has a very high level of resistance against *Bemisia argentifolii* in the field, and that the mechanism of resistance is something not previously known.

**Investigator's Name(s):** Laura Petro, Rick Redak, Jim Bethke, and Thomas M. Perring

**Affiliation & Location:** Department of Entomology, University of California, Riverside, CA. 92521

**Research & Implementation Area:** Section E: Host Plant Resistance, Physiological Disorders, and Host-Plant Interactions

**Dates Covered by the Report:** January 1, 2001 - December 31, 2001

### **Preference and Performance of Silverleaf Whitefly on Selected Poinsettia Cultivars**

Studies showed that silverleaf whitefly females clearly showed a preference for ovipositing on the poinsettia cultivar Peterstar. The cultivars Pepride and Red Velvet were the least preferred of 7 cultivars tested. The development of immatures that eclosed from these eggs were consistent with the numbers of eggs laid on the cultivar (good development on Peterstar, and poor development on Pepride and Red Velvet). Females laid significantly fewer eggs on another cultivar, Petoy, than they laid than Peterstar, however the development of immatures was highly successful on Petoy. Further work with these 4 cultivars showed a very high intrinsic rate of increase ( $R_m=0.434$ ) on Petoy and a low value ( $R_m=0.190$ ) on Pepride for the  $F_0$  generation. This is interesting, since Petoy was not the most preferred host for oviposition. On the other hand, Pepride was among the least preferred oviposition host and this cultivar had the lowest intrinsic rate of increase. These results are not conclusive that the most preferred cultivars are those on which the whitefly will perform the best.

From a practical standpoint, the difference in  $R_m$  values for the cultivars can significantly alter the management of silverleaf whitefly on poinsettia. Small differences in  $R_m$  values result in remarkable differences in expected population growth over time. Our results indicate that the predicted density of whiteflies on the cultivar 'Petoy' would be ten times higher than on the relatively resistant cultivar 'Red Velvet' (189,094 vs. 17,676 respectively) after just 30 days. Using these empirical curves, and an arbitrary treatment threshold of 100 whiteflies, we compared the hypothetical number of treatments required across a 90 day time frame for the four varieties evaluated in chapter 3. 'Petoy' would require 7 pesticide treatments within 90 days, while 'Red Velvet' would require only 6 pesticide treatments within 90 days. Thus, the attributes of a single cultivar may result in fewer pesticide applications and possible overall savings in the costs associated with chemical control for growers. Growers should utilize this information on Pepride and incorporate this cultivar into their breeding programs.

**Investigator's Name(s):** Alvin M. Simmons and Amnon Levi

**Affiliation & Location:** USDA-ARS, U.S. Vegetable Laboratory, Charleston, SC 29414

**Research & Implementation Area:** Section E: Host Plant Resistance, Physiological Disorders, and Host Plant Interaction

**Dates Covered by the Report:** 2000

### **Watermelon Germplasm for Resistance to *Bemisia***

Like many vegetable crops, the cultivated watermelon, *Citrullus lanatus* var. *lanatus*, is plagued by *Bemisia*. *Citrullus* contains four known diploid species. However, there have not been any reports on screening *Citrullus* germplasm for resistance to whiteflies. This study was conducted to identify potential germplasm sources that could be used in the improvement of watermelon for whitefly resistance. The study was conducted on 42 *Citrullus* genotypes. Seven watermelon cultivars, a triploid line, and 16 U.S. Plant Introduction accessions (PIs) of *C. lanatus* var. *lanatus*; 10 PIs of *C. lanatus* var. *citroides*; and 8 PIs of *C. colocynthis* were evaluated for resistance to B-biotype *B. tabaci*. Bioassays were conducted on non-preference, and on ovipositional and survival performance of the whiteflies. Adults and eggs were observed on all genotypes studied, but the abundance of the insects and the survival of the nymphs varied greatly among genotypes. Most of the watermelon cultivars and *C. lanatus* PIs tested were highly susceptible to whitefly infestation, while the *C. colocynthis* PIs exhibited whitefly resistance. Trichome density on the resistant genotypes (*C. colocynthis* PIs) were elevated compared with the susceptible genotypes. However, we believe that the observed resistance was primarily related to effects of plant nutrition on the whiteflies.

This study identified useful sources of germplasm (such as PI 386015, PI 386018, and PI 386024, all of *C. colocynthis*) which can be used for the improvement of watermelon for resistance to whiteflies. Currently, F<sub>2</sub> populations (Charleston Gray x PI 386024) are being constructed and will be used to determine the mode of inheritance of whitefly resistance in watermelon.

**Investigator's Name(s):** Alvin M. Simmons

**Affiliation & Location:** USDA-ARS, U.S. Vegetable Laboratory, Charleston, SC

**Research & Implementation Area:** Section E: Host Plant Resistance, Physiological Disorders, and Host-Plant Interactions

**Dates Covered by the Report:** 2001

### **Movement of Whitefly Crawlers on Several Types of Vegetable Plants**

The nymphal stage of whitefly pests is important in terms of its relationship to plant damage, plant disorders, and pest management measures. It is well known that the B-biotype sweetpotato whitefly, *Bemisia tabaci* (Gennadius), feeds on and damages numerous species of vegetable crops. The crawler, i.e., active first instar, is the only mobile form of the immature whitefly. A study was conducted to determine any influence of vegetable plant species and temperature on net distance moved (between the egg site and final resting site) by crawlers of the B-biotype *B. tabaci*. Tests were conducted in the greenhouse as well as under controlled laboratory conditions on five vegetable hosts: cantaloupe, *Cucumis melo* L.; collard, *Brassica oleracea* ssp. *acephala* de Condolle; cowpea, *Vigna unguiculata* (L.) Walpers ssp. *unguiculata*; pepper, *Capsicum annuum* L. ssp. *annuum*; and tomato, *Lycopersicon esculentum* Miller. On the different host species, the average net distance that crawlers moved ranged from 2-15 mm with the shortest distance on collard. Individuals on pepper and cowpea ceased traveling 50 and 62 mm, respectively, from the site of hatching. Observations on intact collard leaves in the laboratory indicate that the crawlers traveled an average of 21 minutes before developing to the 2<sup>nd</sup> instar. No effect of constant temperature (16-34°C) was detected on the net travel distance of the crawler. The data suggest that among the plant species in this study, collard is highly attractive for feeding and/or it offers suitable feeding sites that are easy to locate by the crawler. The results of this study help define the behavior of crawlers on several species of vegetable plants and help in ongoing research on host plant resistance.

**Investigator's Name(s):** Wilhelmina T.G. van de Ven, Yun-Shu Chen, Linda L. Walling

**Affiliation & Location:** Department of Botany and Plant Sciences, University of California Riverside, Riverside CA 92521

**Research & Implementation Area:** Section E: Host Plant Resistance, Physiological Disorders, and Host-Plant Interactions

**Dates Covered by the Report:** January-2000-December 2000

### **Silverleaf whitefly-induced (SLW) Genes: Species-Specific Induction of Novel Signaling Pathways**

The squash (*Cucurbita pepo*) genes *SLW1* and *SLW3* were differentially expressed in response to silverleaf whitefly (*Bemisia argentifolii*) and sweetpotato whitefly (*B. tabaci* Type A) nymphs (van de Ven., 2000). *SLW1* and *SLW3* RNAs accumulated systemically in apical, silvered leaves after silverleaf whitefly feeding, but not sweetpotato whitefly feeding. Both RNAs accumulated, although at different levels, in leaves on which silverleaf and sweetpotato whitefly nymphs were feeding. The expression of *SLW1* and *SLW3* in response to aphids, caterpillars, nematodes and a bacterial pathogen were evaluated. Consistent with the lack of *SLW1* and *SLW3* expression after mechanical wounding, *SLW1* and *SLW3* transcripts did not accumulate in response to tobacco hornworm (*Manduca sexta*) larvae. Surprisingly, other animals that feed on the vasculature, such as the cotton aphid (*Aphis gossypii*) or root-knot nematodes (*Meloidogyne incognita*), did not cause local accumulation of *SLW1* and *SLW3* RNAs in squash leaves or roots, respectively, or systemic RNA accumulation in non-infested leaves. Infiltration of leaves with *Pseudomonas syringae* pv *syringae* did not induce *SLW1* or *SLW3*. Collectively, these data suggest that *SLW1* and *SLW3* are not typical pathogen- or pest-response genes, since they were preferentially expressed by a single whitefly species.

To investigate the signaling pathways important for *SLW* gene expression, squash plants were treated with a variety of defense and wound signals. *SLW1* (a M20B peptidase gene) was regulated by both JA (jasmonic acid) and ethylene and therefore is activated by an established JA/ethylene-dependent pathway. *SLW3* (a  $\beta$ -glucosidase gene) RNAs did not accumulate in response to JA, ethylene, salicylic acid, abscisic acid, nitric oxide, hydrogen peroxide or a combination of these defense signals. These data indicate that *SLW3* is regulated by a novel, defense-signaling pathway not previously recognized in plants. *SLW1* and *SLW3* RNAs were abundant after water-deficit stress suggesting that the signals generated after silverleaf whitefly feeding and water-deficit stress may overlap or *SLW* genes may respond to multiple signaling pathways. A homolog of *SLW1*, *DRIP-1*, was recently found in watermelon and was also induced after water-deficit stress (Kawasaki et al, 2000).

To understand the tissue specificity of *SLW* gene expression and to evaluate the impact of *SLW1* and *SLW3* proteins on plant defense, we are using the tools of reverse genetics. *SLW1* and *SLW3* promoters were isolated, sequenced, and fused to the  $\beta$ -glucuronidase (GUS) reporter gene in a *Ti*-plasmid binary vector. These chimeric constructs have been used in *Agrobacterium tumefaciens*-mediated transformation. Transgenic tomato and *Arabidopsis* plants expressing *SLW1:GUS* or *SLW3:GUS* transgenes are being regenerated and characterized. These plants will aid in unraveling the similarities and differences between the signals that induce *SLW1* and *SLW3*. Transgenic tomatoes and *Arabidopsis* over-expressing *SLW3* and *SLW1* are also being constructed and will allow us to determine the role of *SLW* genes on whitefly infestation and leaf-silvering.



## Section E Research Summary

Compiled by C. L. McKenzie and Greg Walker

### Host-Plant Resistance:

Advancements continue to be made in breeding and screening plants for resistance to whitefly and whitefly-transmitted viruses and include studies with cultivars, breeding-lines and germplasm of cantaloupe, watermelon, poinsettia, and cotton.

Six cantaloupe varieties were evaluated under field conditions for whitefly resistance in south Texas: Hymark, Impact, Tam Sun x gl, Tam Sun, Primo, and TP45. TP45, a Weslaco line, had the fewest whiteflies (adults and red-eyed-nymphs or pupae) throughout the season, followed by Tam Sun, a TAES variety. Although Tam Sun had the second lowest whitefly numbers, it also had the fewest and lowest total weight of melons. Primo had the greatest number of adult whiteflies (660 / leaf) followed by Hymark and Impact (>300 / leaf). Number of adult whiteflies was > 8-fold higher on Primo compared to TP45. TP45 and Impact had more large melons compared with other varieties whereas Tam Sun x gl. and Primo had more small melons. TP45 also had the highest yield, but unfortunately sugar content was the lowest (~3.8%) compared to other varieties (7.6 - 9.5%). Results from this study indicated that significant differences in yield responses to silverleaf whitefly infestation were found among the six cantaloupe varieties and confirm earlier laboratory findings.

In South Carolina, watermelon germplasm (42 *Citrullus* genotypes) was screened for resistance to whitefly infestation to identify potential germplasm sources that could be used in the improvement of cultivated watermelon varieties. Seven watermelon cultivars, a triploid line, 16 U.S. Plant Introduction accessions (PIs) of *C. lanatus* var. *lanatus*, 10 PIs of *C. lanatus* var. *citroides*, and 8 PIs of *C. colocynthis* were evaluated for whitefly resistance. Bioassays were conducted on whitefly non-preference, oviposition and survival. Most of the watermelon cultivars and *C. lanatus* PIs tested were highly susceptible to whitefly infestation, while the *C. colocynthis* PIs exhibited whitefly resistance. This study identified useful sources of germplasm (PI 386015, PI 386018, and PI 386024, all of *C. colocynthis*) which can be used for the improvement of watermelon for resistance to whiteflies.

Seven poinsettia cultivars were evaluated at U CA in Riverside for preference and performance of silverleaf whitefly. Peterstar was the most preferred cultivar while Pepride and Red Velvet were the least preferred. Development of immature stages that eclosed from these eggs were consistent with the numbers of eggs laid on the cultivar (good development on Peterstar, and poor development on Pepride and Red Velvet). Females laid

significantly fewer eggs on Peto compared to Peterstar, however the development of nymphs was highly successful on Peto ( $R_m=0.434$ ). In contrast, Pepride was among the least preferred oviposition host plus it had the lowest intrinsic rate of increase ( $R_m=0.190$ ). Results are not conclusive that the most preferred cultivars are those on which the whitefly will perform the best. Growers should utilize this information on Pepride by incorporating this cultivar into their breeding programs.

Sixteen upland cotton types were evaluated in the field for susceptibility to silverleaf whitefly infestations in California's Imperial Valley and included 10 normal leaf and three okra-leaf cultivars plus three experimental okra-leaf breeding-lines. The okra-leaf entries as a group had fewer whitefly adults and nymphs than the normal leaf cotton entries. The okra-leaf experimental breeding-lines had the lowest numbers of whitefly adults and nymphs among the okra-leaf entries. Stoneville 474 (hirsute leaf) had the greatest numbers of whitefly adults and nymphs among the normal leaf cottons. However, no significant differences in seed cotton yield among the entries were detected.

Ten cotton cultivars or breeding-lines were evaluated in Imperial Valley, CA for resistance to cotton leaf crumple (CLCr) disease caused by cotton leaf crumple virus (CLCrV) and transmitted by the silverleaf whitefly. Results showed differences in whitefly infestation levels and virus disease symptoms among cotton entries in terms of severity of CLCr disease symptoms and the presence of CLCrV. The cultivar AP 4103 had a higher CLCr disease rating than other entries except AP 6101. The breeding-line NK 2387C, with Cedix parentage, had a lower CLCr disease rating than other entries except Stoneville 474 and NKX 2207. Visible CLCr disease symptoms were observed in Stoneville 474 and NKX 2207 entries, but NK 2387C did not display visible CLCr disease symptoms nor was viral DNA detected implicating this line as a potential source for improvement of cotton resistance to CLCrV and disease development.

### Physiological Disorders:

Great strides have been made in understanding the plant molecular mechanisms and genetics of squash silverleaf (SSL) disorder, a serious physiological disorder of commercial *Cucurbita* species associated with the feeding of nymphal *Bemisia argentifolii*, and how that translates into reduction in plant growth and yield observed in the field.

At the U CA (Riverside) researchers discovered that silverleaf whitefly-induced (*SLW*) genes utilize novel signaling pathways that appear to be species-specific. The squash (*Cucurbita pepo*) genes (*SLW1* and *SLW3*) were differentially expressed in response to silverleaf whitefly (*SLWF*) and sweetpotato whitefly (*SPWF*) nymphs. Both *SLW* genes accumulate systemically in

apical, non-infested, silvered leaves after SLWF feeding, but not after SPWF feeding. Both *SLW* transcripts accumulate in the local, infested leaves of SLWF infested plants, however only *SLW3* was induced in the local leaves of SPWF infested leaves. The expression of *SLWI* was detected during flower and fruit development, while *SLW3* RNAs were not. In contrast, neither gene transcript accumulated in response to insects, nematodes or a bacterial pathogen. Collectively, these data suggest that *SLWI* and *SLW3* are not typical pathogen- or pest-response genes, since they were preferentially expressed by a single whitefly species.

*SLWI* (a M20B peptidase gene) is regulated/activated by an established jasmonic acid/ethylene-dependent pathway. However the signaling pathway for *SLW3* (a  $\beta$ -glucosidase gene) appears to be regulated by a novel, defense-signaling pathway not previously recognized in plants because plant RNA transcripts did not accumulate in response to known defense and wound signals. In addition, both genes were abundant after water-deficit stress suggesting that the signals generated after SLWF feeding and water-deficit stress may overlap or *SLW* genes may respond to multiple signaling pathways.

The reverse genetic approach is being used to investigate the tissue specificity of *SLW* gene expression and function of *SLW* genes in the plant response to SLWF feeding and evaluate the impact of *SLW* proteins on plant defense. *SLW3: GUS* transgenic tomato and *Arabidopsis* have been generated and *SLW3* expression studies under abiotic and biotic stresses and during plant development are ongoing. In addition, transgenic plant lines ectopically expressing *SLW3* have been constructed and will aid in future studies to determine the role of *SLW* genes on whitefly infestation and leaf silvering.

Silvering results from an abnormal air space between the upper epidermis and the upper palisade layer of cells, beginning at the tip of young leaves and proceeding downward to cover the entire leaf. The number of cells in each layer over time is the same in whitefly-infested plants and controls indicating cell separation between epidermis and palisade cells is not a result of differential cell division or expansion between the two layers. Silvering develops simultaneously as normal air space formation is occurring and is more likely a result of increased cell wall degradation. No anatomical response to whitefly feeding was observed in SSL-resistant zucchini, but plants were lighter green with shorter petioles and comparable to SSL-susceptible plants. In grafts of resistant plants onto susceptible plants challenged with whitefly nymphs, the resistant plants did not develop leaf silvering. However, susceptible scions grafted onto resistant rootstocks did exhibit leaf silvering indicating that the site of SSL resistance is at the young developing leaves and not at the mature leaf where whitefly feeding occurs.

Companion field trials conducted in Mayaguez, Puerto Rico and Leesburg, FL screened *Cucurbita* spp. (squash) accessions obtained from the National Plant Germplasm System repository and identified several resistant lines to SSL disorder. Researchers found that even though SSL-resistant germplasm did not silver, plants received similar levels of oviposition and supported similar rates of development and survival as did SSL-susceptible accessions indicating that the mechanism of resistance was tolerance to the effects of whitefly feeding. The genes responsible for SSL resistance appear to be related to the absence of natural leaf mottling and are conditioned by one or two genes depending on the accession. Environmental conditions and whitefly infestation level can modify the severity of SSL symptoms.

Studies to determine the effect of whitefly and squash silverleaf disorder (SSL) on zucchini plant growth and yield were conducted at the U FL, Gainesville in greenhouse and outdoor screened room experiments using two closely-related zucchini breeding-lines, ZUC61 (silverleaf susceptible) and ZUC76-SLR (silverleaf resistant), and four infestation levels. Greenhouse results indicated seedlings for both zucchini breeding-lines were stunted, had shortened plant height and petiole length of silvered leaves, and showed ~ 10% reduction in dry weight after infestation of 30-60 pairs of adult whiteflies and their progeny over 2 weeks. Susceptible plants exhibited severe (grade 5) SSL symptoms whereas ZUC76-SLR had no or mild (grade 1) symptoms, yet the same reduction in seedling growth was observed. In outdoor screened room experiments, serious silverleaf symptoms (grade 4-5) were expressed on all susceptible plants and slight silverleaf symptoms (grade 1-3) were expressed on resistant plants, but silverleaf severity of each zucchini breeding-line did not differ significantly among whitefly infestation levels (spring or fall). Significant fruit yield reduction appeared at infestation level 30 for both breeding-lines. Season-long feeding by higher infestation levels (60-120 pairs) caused significant yield and quality reduction in fruit for both breeding-lines and even resulted in plant death in fall experiments. Zucchini seedling stunting and yield reduction appears to be more related to intensity of whitefly feeding than to susceptibility to SSL and expression of silverleaf symptoms.

#### **Host-Plant Interactions:**

In Egypt, parasitism of whitefly on 16 taxonomically diverse host plants by augmentation with *Eretmocerus mundus* Mercet was determined in field plots across five locations. Vegetable and other agronomic host plants included bean, broccoli, cabbage, cantaloupe, cotton, cucumber, egg plant, lantana, potato, southern pea, soybean, sugar beet, sunflower, sweet potato, tomato, and watermelon. Parasitism was enhanced in all plots where augmentations were made, but rates differed. In some plots, e.g., cabbage, broccoli, bean, and soybean, overall parasitism was enhanced at a relatively high rate while in

other plots, e.g., watermelon, egg plant, and tomato, the enhancement of overall parasitism was relatively low.

Researchers in South Carolina conducted greenhouse and laboratory experiments to determine the influence of five vegetable hosts (cantaloupe, collard, cowpea, pepper and tomato) and temperature on net distance traveled by whitefly crawlers. No effect of constant temperature (16-34°C) was detected on the net travel distance of the crawler. The average net distance that crawlers moved ranged from 2-15 mm on the different hosts, but crawlers on pepper and cowpea ceased traveling 50 and 62 mm, respectively, from the site of hatching. Crawlers moved the shortest distance on collards and traveled an average of 21 minutes before developing to the 2<sup>nd</sup> instar. The data suggest that among the plant species in this study, collard is highly attractive for feeding and/or it offers suitable feeding sites that are easy to locate by the crawler.

Nine upland cotton cultivars were evaluated for whitefly populations in relation to trichome densities in field trials over two growing seasons at the U AZ Agricultural Research Center, at Maricopa, AZ. Entries included five normal-leaf and four okra-leaf cotton cultivars with all but the hairy-leaf ST 474 being smooth-leaf. ST 474 had 71-fold and > 6-fold more stellate trichomes/cm<sup>2</sup> of leaf disk compared to smooth-leaf cultivars in 2000 and 2001, respectively. ST 474 had > 2-fold more eggs/cm<sup>2</sup> and nymphs/cm<sup>2</sup> leaf disk in 2000 and >2-fold more adults per leaf-turn count compared to smooth-leaf cultivars in both years. In 2001, ST 474 had > 4-fold more eggs/cm<sup>2</sup> and nymphs/cm<sup>2</sup> leaf disk compared to smooth-leaf cultivars. On average of all cultivars studied for both years, leaves on the top main stem leaf node (#1) had the highest number of stellate trichomes, but the lowest numbers of eggs, nymphs and adults compared with leaves lower in the canopy.

**Table E. Host-Plant Resistance, Physiological Disorders, and Host-Plant Interactions.**

Research approaches	Year 1 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Characterize resistance mechanisms and identify chemical/morphological components, and study effects of insect adaptation.</b>	Identify potential sources of germplasm for disease, plant disorders and whitefly resistance. <sup>a</sup>	X		Research was conducted on identifying potential sources of germplasm for whitefly resistance in alfalfa, cotton, melon, cole crops, and cucurbits; and resistance to virus symptoms and silverleaf disorder in cotton and cucurbits, respectively. These studies included research on plant tolerance, antibiosis, and antixenosis. Antixenosis was found not to be responsible for resistance to squash silverleaf in two zucchini lines.
<b>Develop molecular level techniques to produce resistant germplasm.</b>	Identify physiological processes of whiteflies to target for inhibition.	X		Characterization of plant genome was demonstrated in tomato and squash. Pathogenesis related mRNAs accumulated in response to whitefly feeding on tomato leaves. Data on whitefly probing behavior indicates that host evaluation phase of <i>Bemisia</i> -host interaction is dominated by probing.
<b>Incorporate resistance traits into commercial genotypes.</b>	Identify and isolate genetic sources of resistance for transformation and/or breeding.	X		From promising genetic materials, inbreds, F <sub>1</sub> and F <sub>2</sub> progenies, and assorted cultivars were studied for whitefly resistance (in alfalfa, cotton, melon and squash), and susceptibility to diseases (in cotton) and plant disorders (in squash). Including plant geneticists and other specialists on the research team has been an asset.
<b>Determine influence of host plant morphology, physiology and phenology on feeding behavior and competition.<sup>b</sup></b>	Characterize nutritional and other preference properties of various host plants.	X		Research was studied on the acceptability of cotton and vegetable hosts on whitefly feeding behavior. Work was conducted on distance from abaxial surface to minor veins, and feeding response on abaxial and adaxial surfaces of different hosts.
<b>Define whitefly feeding and oviposition behavior and investigate approaches for interrupting whitefly feeding and digestion.<sup>c</sup></b>	Investigate approaches for interruption of feeding, assimilation, development and reproduction.	X		The host evaluation phase of <i>Bemisia</i> -host interactions was shown to dominate by probing, and the time spent in a particular behavior was affected by imidacloprid when the whitefly came into contact with the chemical in its diet rather than on the leaf surface. Intercropping of resistant within susceptible cole crops did not lessen the abundance of whiteflies.

**Table E. Host-Plant Resistance, Physiological Disorders, and Host-Plant Interactions. (Continued)**

Research approaches	Year 1 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Study whitefly toxicogenic plant reactions.</b>	Determine effects of whitefly feeding on host plant physiology, morphology and anatomy.	X		Research on tomato identified a gene that is specifically induced by whitefly feeding. Four classes of genes were identified in inducing squash leaf silvering. These genes were further characterized by hybridization, sequence analysis and complementation studies.

<sup>a</sup> See Table B for additional plant disease resistance research.

<sup>b</sup> See Section A.

<sup>c</sup> See Section A, approach #9.

**Table E. Host-Plant Resistance, Physiological Disorders, and Host-Plant Interactions.**

Research approaches	Year 2 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Characterize resistance mechanisms and identify chemical/morphological components, and study effects of insect adaptation.</b>	Determine physiological and/or morphological basis for resistance, & effects of host-plant history and insect adaptation on plant resistance to whiteflies. Continue to identify resistant germplasm.	X		Selection for a whitefly resistant variety of alfalfa is close to completion; release of a commercial variety is expected within a few years. Whitefly-resistant or partially whitefly-resistant varieties of a number of crops have been identified, including cotton, collard, and melons. Varieties of cotton and tomato with resistance or partial resistance to whitefly-transmitted viruses also have been identified. In collards, the glossy leaf trait, and in cotton, the okra-leaf trait and large leaf surface to vascular bundle depth have been implicated as mechanisms of whitefly resistance in plants. Increased levels of phenolics and peroxidase in response to plant stress have been associated with decreased whitefly performance in tomato. In <i>Datura wrightii</i> , glandular trichomes were demonstrated to be a very effective mechanism of resistance to whiteflies.
<b>Develop molecular level techniques to produce resistant germplasm.</b>	Identify natural products for inhibiting processes.	X		The natural plant products, neem seed extract, azadiractin, and extract of bitterwood, were shown to be effective insecticides against silverleaf whitefly.
<b>Incorporate resistance traits into commercial genotypes.</b>	Insert genes into plants <sup>b</sup> via plant transformation.	X		Resistant commercial lines of alfalfa are close to release and commercial varieties of collard have been shown to exhibit whitefly resistance. Also, lines of cotton and melon have been identified with partial whitefly resistance. No progress has been made in the specific year 2 goal of inserting whitefly resistance genes into plants via transformation.

**Table E. Host-Plant Resistance, Physiological Disorders, and Host-Plant Interactions.**

Research approaches	Year 2 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Determine influence of host plant morphology, physiology and phenology on feeding behavior and competition.<sup>b</sup></b>	Determine the biochemical mechanism regulating adaptation to host plants.	X		Morphological plant traits such as okra-leaf and large distance from leaf surface to vascular bundles in cotton, and glandular trichomes in <i>Datura wrightii</i> have been shown to provide partial or complete whitefly resistance. Fluctuations in amino acid concentrations over the lifespan of melon leaves were correlated with whitefly performance. Also in melons, group feeding by whiteflies was shown to create a nutrient sink in the plant, and thus provide the whiteflies with improved amino acid nutrition. Senescence in poinsettia reduces host plant quality for silverleaf whitefly. In cotton, decreased nitrogen fertilization decreases whitefly populations. In tomato, plant stress caused by fertilizer and/or water deficiency reduces host plant quality for silverleaf whitefly.
<b>Define whitefly feeding and oviposition behavior and investigate approaches for interrupting whitefly feeding and digestion.<sup>c</sup></b>	Identify physiological and morphological mechanisms regulating processes.	X		Improvements have been made in a system for rearing whiteflies on an artificial liquid medium. This will allow direct experimentation on the role of specific plant nutrients and allelochemicals on whitefly feeding and performance. Stylet contact with minor vascular bundles is essential for successful whitefly feeding on cotton. The fine structure of whitefly eggs and their attachment to host leaves has been studied with electron microscopy, and the distal end of the egg petiole that is inserted into the host leaves possesses morphological structures that suggest a role in water uptake from the host leaf which is a very important process for egg survival.
<b>Study whitefly toxicogenic plant reactions.</b>	Determine biochemical basis for physiological response of plant.	X		Genes specifically induced by whitefly feeding have been identified in tomato and in squash. These genes may play a role in the plant's defensive response to the whitefly and/or the plant's toxicogenic reaction such as irregular ripening in tomato and silverleaf symptom in squash.

<sup>a</sup> See Table B for additional plant disease resistance research.

<sup>b</sup> See Section A.

<sup>c</sup> See Section A, approach #9.

**Table E. Host-Plant Resistance, Physiological Disorders, and Host-Plant Interactions.**

Research approaches	Year 3 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Characterize resistance mechanisms and identify chemical/morphological components, and study effects of insect adaptation.</b>	Elucidate biochemical and molecular basis for resistance. Continue to identify resistant germplasm.	X		Selection for a whitefly resistant variety of alfalfa is close to completion; release of a commercial variety is expected within a year. Whitefly fecundity tests on clonal propagules of alfalfa plants chosen in the field for whitefly resistance indicate that the field-selection criteria reflect actual resistance. Fecundity of whiteflies on alfalfa was higher for alfalfa-reared whiteflies than for cotton-reared whiteflies. This suggests whitefly adaptation to a crop species. In cotton, the okra-leaf trait and glabrous-leaf trait have been again demonstrated as a mechanisms of partial resistance against whiteflies; however, closer scrutiny of an earlier report that a large leaf surface to vascular bundle depth confers whitefly resistance has been discredited. This is useful information so that resources can be focused on examining effective mechanisms of resistance and avoid wasting resources on unlikely mechanisms. The mechanism of resistance against whiteflies in a tomato variety carrying the <i>Mi</i> gene has been shown to be due to factors encountered by whiteflies before they penetrate sieve elements rather than factors in the phloem sap. Varieties of cotton, cantaloupe, and poinsettia with resistance or partial resistance to whiteflies have been identified; and varieties of cotton and tomato with resistance or partial resistance to whitefly-transmitted viruses have been identified. Comparison of whitefly behavior on different plant species that range in suitability from good host to poor host detected little difference in whitefly behavior during initial contact with the plant. Whiteflies oviposited even on poor hosts, indicating non-selectivity on the part of the ovipositing female. This suggests that whiteflies will readily oviposit on resistant crops, and consequently mechanisms of plant resistance will be continuously challenged by migrating whiteflies.



**Table E. Host-Plant Resistance, Physiological Disorders, and Host-Plant Interactions.**

Research approaches	Year 3 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Develop molecular level techniques to produce resistant germplasm.</b>	Isolate the relevant biosynthetic enzymes that encode for natural products inhibiting processes.	X		The natural plant products, azadiractin, was shown to be an effective insecticide against silverleaf whitefly. Presently, there are no attempts to insert the genes for this plant product into crop plants.
<b>Incorporate resistance traits into commercial genotypes.</b>	Evaluate potential of newly transformed germplasm.	X		Whitefly-resistant commercial lines of alfalfa are close to release. Commercially available varieties of cotton, cantaloupe, and poinsettia that are resistant or partially resistant against whiteflies or whitefly-transmitted viruses have been identified. No genetically transformed germplasm has yet been evaluated for whitefly resistance.
<b>Determine influence of host plant morphology, physiology and phenology on feeding behavior and competition.<sup>b</sup></b>	Determine changes in whitefly gene expression in response to host manipulation.	X		In cotton, the okra-leaf trait and glabrous-leaf trait have been confirmed as a mechanisms of partial resistance against whiteflies; however, closer scrutiny of an earlier report that a large leaf surface to vascular bundle depth confers whitefly resistance has been discredited. Factors encountered by whiteflies during their stylet penetration to vascular bundles has been shown to confer partial resistance in a tomato variety with the <i>Mi</i> gene. Phloem sap factors do not appear to play a role in this resistance. The known host plant range of silverleaf whitefly has been expanded to include some medicinal plants and weed species. An abundance of host plant species suitable for overwintering in California's San Joaquin Valley have been identified; thus a strategy of host-free periods for whitefly management is not very promising in the San Joaquin Valley.

**Table E. Host-Plant Resistance, Physiological Disorders, and Host-Plant Interactions.**

Research approaches	Year 3 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Define whitefly feeding and oviposition behavior and investigate approaches for interrupting whitefly feeding and digestion.<sup>c</sup></b>	Determine biochemical and molecular basis for inhibiting processes.	X		Morphological studies on whitefly stylets indicate that they are sufficiently long to reach minor vascular bundles (the major feeding site) from virtually any place on the abaxial leaf surface of cotton. This makes variation in vascular bundle depth an unlikely mechanism of resistance to whiteflies in cotton. Variation in nitrogen fertilization has been shown to decrease amino acid concentrations in phloem sap and thus affects nutrition available to whiteflies. Whitefly feeding differentially induces pathogenesis-related (PR) proteins in two cucurbit species, cantaloupe and watermelon, and apparently is not affected by treatment with plant growth-promoting rhizobacteria (PGPR).
<b>Study whitefly toxicogenic plant reactions.</b>	Elucidate changes in plant gene expression.	X		Two genes, one of which appears to be a general plant defense, have been shown to be differentially induced in squash by silverleaf and sweetpotato whiteflies. This may be related to the different toxicogenic effects of these two whitefly species on squash. The activation of these genes is systemic. In tomatoes, feeding by both silverleaf whitefly and greenhouse whitefly induced pathogenesis related genes, but not genes regulated by the octadecanoid pathway. These studies indicate that tomato plants perceive phloem-feeding silverleaf and greenhouse whiteflies in a manner distinct from that of chewing insects.

<sup>a</sup> See Table B for additional plant disease resistance research.

<sup>b</sup> See Section A.

<sup>c</sup> See Section A, approach #9.

**Table E. Host-Plant Resistance, Physiological Disorders, and Host-Plant Interactions.**

Research approaches	Year 4/5 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Characterize resistance mechanisms and identify chemical/morphological components, and study effects of insect adaptation.</b>	Determine potential for transfer of resistance traits.	X		Natwick & Walker's work identified high levels of whitefly resistance in the wild cotton species, <i>Gossypium thurberi</i> , a <i>Gossypium</i> species that in the past had been successfully used to introduce new genes into commercial cotton. Chu and Natwick's work on Siokra cotton added to this body of work. Heather McAuslane's lab (U FL) determined that the genes responsible for SSL disorder appear to be related to the absence of natural leaf mottling and are conditioned by one or two genes depending on the accession.
	Evaluate potential for incorporating <i>Bemisia</i> , plant disorder and disease resistance into acceptable plant type. <sup>a</sup>	X		Larry Tueber (UC Davis) developed and released a commercial whitefly resistant alfalfa variety (UC-Impalo WF) for desert SW alfalfa growing regions. Tong-Xian Liu (TAES) evaluated cantaloupe varieties for whitefly resistance in south Texas and found significant differences in yield responses to whitefly pressure. R.T. McMillan (U FL) identified whitefly transmitted virus resistant beans and released a commercially available bean cultivar (Genuine) for resistance to Bean Golden Mosaic virus in south Florida. Natwick evaluated cotton for resistance to CLCrV and identified NK 2387C as a potential source for improvement of cotton resistance to CLCrV and disease development. McAuslane's lab identified several squash accessions from the National Plant Germplasm System repository that were resistant to SSL disorder.
<b>Develop molecular level techniques to produce resistant germplasm.</b>	Insert genes into plants via plant transformation. <sup>b</sup>	X		Jane Polston (U FL) successfully transformed tomato for resistance to ToMoV and found that plants resistant to ToMoV also appear to convey resistance to TYLCV. Linda Walling's lab (UC Riverside) has generated <i>SLW3:GUS</i> transgenic tomato and <i>Arabidopsis</i> for expression studies.
	Evaluate potential of newly transformed germplasm. <sup>b</sup>	X		Polston's lab is currently evaluating transformed tomato for field performance and resistance to TYLCV.

**Table E. Host-Plant Resistance, Physiological Disorders, and Host-Plant Interactions (Continued)**

Research approaches	Year 4/5 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Incorporate resistance traits into commercial genotypes.</b>	Continue to refine resistance factors to improve resistance in newly transformed germplasm.	X		Through conventional breeding, Teuber and Walker are working on improving whitefly resistance in alfalfa and are making progress identifying the mechanism of resistance.
	Incorporate other desirable plant characteristics for crop production	X		Teuber's continued breeding to incorporate more desirable traits into the whitefly-resistant alfalfa that he developed.
<b>Determine influence of host plant morphology, physiology and phenology on feeding behavior and competition.<sup>b</sup></b>	Relate changes in gene expression to whitefly physiology	X		Walling's lab determined that silverleaf whitefly-induced genes utilize novel signaling pathways that appear to be species-specific. These pathways appear to overlap with signals generated by water-deficit stress or <i>SLW</i> genes may be responding to multiple signaling pathways.
	Summarize and disseminate results	X but limited		Manuscripts are in preparation.
<b>Define whitefly feeding and oviposition behavior and investigate approaches for interrupting whitefly feeding and digestion.<sup>c</sup></b>	Determine potential for transfer of resistance traits.	X		Perring's lab (UC Riverside) looked at whitefly preference and the intrinsic rate of increase between poinsettia cultivars and found Pepride to be a promising cultivar for improving breeding programs. Simmons evaluated the effect of temperature and five vegetable hosts on net distance traveled by crawlers and found no effect of constant temperature (16-34°C) with crawlers traveling the shortest distance on collards < cowpea < pepper before developing into the 2 <sup>nd</sup> instar. Chu consistently found whitefly to prefer hairy-leaf cotton (ST 474) to normal-leaf and okra-leaf cultivars evaluated with > 2 to 4-fold more eggs, nymphs and adult whiteflies observed. Researchers in Egypt found parasitism to be enhanced by augmentation with <i>Eretmocerus mundus</i> Mercot in all of the 16 taxonomically diverse host plants studied, but rates differed from relatively high (cole and legume hosts) to relatively low (crucifer and solanaceous crops).
	Insert genes into plants <sup>a</sup> via plant transformation	X		See notes above regarding Polston and Walling's work.

**Table E. Host-Plant Resistance, Physiological Disorders, and Host-Plant Interactions.**

Research approaches	Year 4/5 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Study whitefly toxicogenic plant reactions.</b>	Identify resistance germplasm.	X		See notes above regarding Natwick & Walker's work on <i>Gossypium thurberi</i> and Chu & Natwick's work on Siokra cotton. See R. T. McMillan, Tong-Xian Liu, McAuslane above. Also, Walker has identified several genotypes of alfalfa highly resistant to <i>Bemisia</i> . Simmons (USDA-ARS) screened watermelon germplasm and identified <i>Citrullus colocynthis</i> accessions and other PIs, which are being used to improve cultivated watermelon varieties for resistance to whiteflies.
	Evaluate potential for transferring new germplasm.	X		McMillan and Tueber have released commercially available resistant bean and alfalfa varieties, respectively. Other programs with tomato, melon, squash, and cotton are in various stages of getting resistant cultivars to the grower.

<sup>a</sup> See Table B for additional plant disease resistance research.

<sup>b</sup> See Section A.

<sup>c</sup> See Section A, approach #9.

Reports of Research Progress

**Section F: Integrated and Areawide Pest Management Approaches, and Crop Management Systems**

Co-Chairs: Steve Castle and Peter Ellsworth

**Investigator's Name(s):** C. C. Chu, P. Alexander, C. G. Jackson, and T. J. Henneberry

**Affiliation & Locations:** USDA-ARS, Western Cotton Research Laboratory, Phoenix, AZ

**Research & Implementation Area:** Section F: Integrated and Areawide Pest Management Approaches, and Crop Management Systems

**Dates Covered by the Report:** 1998-2001

**A Light-Emitting Diode Equipped CC Trap**

Trap catches of adult *Bemisia tabaci* (Gennadius), biotype-B, and their *Eretmocer* spp. and *Encarsia* spp. parasitoids were compared in cage studies in the greenhouse. Average catches of adult *B. tabaci* in CC traps were 41% of the numbers caught on 100 cm<sup>2</sup> yellow sticky card (YC) traps. Coating the insides of CC trap tops with Tanglefoot® (TCC trap) and removing the deflector plate, increased adult *B. tabaci* catches to 65% of the number caught on YC traps. Equipping TCC traps with light-emitting diodes (LED-CC trap) increased adult *B. tabaci* catches by 614% compared with TCC traps. Few *Eretmocer* spp. and *Encarsia* spp., were caught in LED-CC traps. The LED-CC traps may have potential in greenhouse crop production systems where YC traps are used to supplement parasite releases for *Bemisia* control.

**Investigator's Name(s):** C. C. Chu<sup>1</sup>, A. M. Simmons<sup>2</sup>, P. J. Alexander<sup>1</sup>, and T. J. Henneberry<sup>1</sup>

**Affiliation & Locations:** <sup>1</sup>USDA-ARS, Western Cotton Research Laboratory, Phoenix, AZ, and <sup>2</sup>USDA-ARS, U.S. Vegetable Laboratory, Charleston, SC 29414

**Research & Implementation Area:** Section F: Integrated and Areawide Pest Management Approaches, and Crop Management Systems

**Dates Covered by the Report:** 2000-2001

#### **A Light-Emitting Diode Equipped Yellow Sticky Card Trap**

Yellow sticky card (YC) traps were modified with 530 nm lime green light-emitting diodes (LED-YC) to increase the efficacy of capture of *Bemisia tabaci* (Gennadius) and other greenhouse insects as compared with catches on standard YC traps. More adult *B. tabaci* (biotype-B) and fungal gnats, *Bradysia coprophila* (Lintner), were caught on LED-YC traps compared with standard YC traps. Likewise, more *Delphastus pusillus* (LeConte) (coccinellid predators), were caught on the LED-YC traps compared with standard YC traps in a greenhouse test with collard plants, but not in a test with melon plants. However, in tests on both crops, the LED-YC traps did not catch more *Eretmocerus* spp. (parasitoids of whiteflies) than the standard YC traps. Moreover, the capture of western flower thrips, *Franklinella occidentalis* (Pergrande) was not affected by the LED-YC trap compared with the standard YC trap. These results demonstrate the potential of using LED-YC traps in greenhouses for insect detection, monitoring, and control.

**Investigator's Name(s):** Robert T. McMillan, Jr., M. J. Davis, Z. Ying and Dakshina Seal

**Affiliation & Location:** University of Florida, IFAS, TREC 18905 SW 280<sup>th</sup> Street Homestead, FL 33031

**Research & Implementation Area:** Section F. Integrated and Area-wide Pest Management Approaches, and Crop Management Systems

**Dates Covered by Report:** 1999-2000 Crop Season

### **Tomato Yellow Leaf Curl Gemini Virus Management in South Florida**

Tomato Yellow Leaf Curl Geminivirus (TYLCV) was identified in the United States for the first time in late July 1997 in a field planting at a commercial breeding facility in Florida. Shortly thereafter, infected tomato plants were also found in several retail garden outlets in Florida (personal communication J. Polston). The source of these plants was traced back to two commercial nurseries near Homestead in south Florida that had shipped tomato plants to retail outlets in Florida and other states and out of the country. Subsequently, infected tomatoes from retail garden outlets have been found in Virginia (confirmed by laboratory tests) and possibly other locations in the southern United States supporting the possibility that TYLCV will become a regional problem.

Due, apparently, to the introduction and spread of the silver leaf whitefly, there has been a recent emergence of seventeen whitefly-transmitted geminiviruses in tomato in the Western Hemisphere. Of these, only tomato mottle virus was present in Florida before TYLCV. Polymerase chain reaction (PCR) and DNA sequence analyses were used to confirm the recent introduction of TYLCV into Florida. Degenerate primers for geminiviruses were used to amplify a fragment of the viral genome containing part of the coat protein gene. The partial gene sequence had greater than 98% homology with that of an Israel strain of the TYLCV. These results suggest that the virus is an Eastern Mediterranean strain and possibly the same strain previously introduced into the Dominican Republic from Israel in 1991 and subsequently found in Jamaica and Cuba. We have designed non-degenerate primers for PCR detection of TYLCV based on our DNA sequence data for the virus. Detection of TYLCV in tomato plants with these primers has been 10-100 times more sensitive than with the degenerate primers, which even with the degeneracy have mismatched bases.

Tomatoes are a winter crop in south Florida, and TYLCV was detected in October 1997, in newly established commercial plantings. Tomatoes are still being planted in south Florida at the present time. The incidence of TYLCV-infected plants initially appeared to be due mostly to primary spread into the fields. Sources of inoculum are presumed to be weeds outside of the tomato fields. Silverleaf whitefly (*Bemisia argentifolii*) population densities have been generally low, and even less in tomatoes due to the extensive application of the systemic insecticide, imidacloprid (Admire), to transplants. However, the occurrence and incidence of TYLCV in cultivated tomatoes have steadily increased during the growing season. Secondary spread of TYLCV within fields has become more prevalent and Silverleaf whitefly population densities are on the increase. TYLCV appears to be firmly established in Florida, and will likely become a major problem in the region in years to come.

Recent field studies with combinations of Admire at 8 oz. and Neemix at 4 oz. have reduced the whitefly populations, as well as the incidence of TYLCV and improved tomato yields. Also, the combination of Admire at 8 oz. plus Knack at 8 oz. plus Applaud at 8 oz., reduced whitefly populations, incidence of TYLCV and improved tomato yields.



**Investigator's Names:** Steven E. Naranjo<sup>1</sup>, Peter C. Ellsworth<sup>2</sup>, C. C. Chu<sup>1</sup> & Thomas J. Henneberry<sup>1</sup>

**Affiliation & Location:** <sup>1</sup>USDA-ARS, Western Cotton Research Laboratory, Phoenix, AZ and <sup>2</sup>Department of Entomology, University of Arizona, Maricopa Agricultural Center, 37860 W. Smith-Enke Road, Maricopa, AZ 85239

**Research & Implementation Area:** Section F: Integrated and Areawide Pest Management Approaches, and Crop Management Systems

**Dates Covered by the Report:** January 2000- December 2001

#### **Conservation of Predatory Arthropods in Cotton: Role of Action Thresholds for *Bemisia tabaci***

Studies were conducted in 1994 and 1995 to measure the effects of prescriptive insecticide use for *Bemisia tabaci* in cotton on populations of arthropod predators in Imperial Valley, California and Maricopa, Arizona. Replicated plots were sprayed with a pyrethroid + organophosphate mixture when densities of adult *B. tabaci* exceeded 2.5, 5, 10, or 20 per leaf. Untreated plots served as controls. Application of insecticides significantly reduced population densities of spiders, *Geocoris punctipes* (Say), *G. pallens* (Stål), *Orius tristicolor* (White), *Nabis alternatus* Parshley, *Zelus renardii* Kolenati, *Hippodamia convergens* Guérin-Méneville, *Spanogonicus albofasciatus* (Reuter), *Drapetis* sp., and *Chrysoperla carnea* Stephens in one or both years and sites compared to untreated controls. Use of higher *B. tabaci* thresholds conserved some species and groups relative to lower thresholds. Stepwise regression analyses indicated that reductions in predator populations were generally influenced more strongly by the timing of the first insecticide application than by the total number of sprays necessary to maintain suppression of the pest below any given action threshold. A predation index, which weights the importance of predator species based on their known frequency of predation on *B. tabaci* and another key pest, *Pectinophora gossypiella* (Saunders), was developed and analyzed. General patterns were similar to results based on changes in abundance alone, but the index generally revealed less severe effects of insecticides on overall predator function. The current action threshold for conventional insecticidal control of *B. tabaci* in Arizona and southern California is 5 adults per leaf. Results here suggest that predator conservation may be enhanced by raising the initial threshold to delay the first application or initially using more selective materials such as insect growth regulators.

**Investigator's Name(s):** Yash Pal S. Rathi

**Affiliation & Location:** Centre of Advanced Studies in Plant Pathology, G.B. Pant Univ. of Agri. & Tech. Pantnagar PIN: 263 145 (India)

**Research & Implementation Area:** Section F. Integrated and Areawide Pest Management Approaches, and Crop Management Systems

**Dates Covered by Report:** 1999-2000

### **Management Of Mungbean Yellow Mosaic Virus, A Yellow Plague Of *Kharif* Grain Legumes**

Mungbean yellow mosaic virus (MYMV) also called as a “yellow plague” of *Kharif* grain legumes is transmitted by whitefly (*Bemisia tabaci*) in persistent circular manner. Currently, it is a number one problem causing substantial yield losses in urdbean (*Vigna mungo*), mungbean (*V. radiata*), mothbean (*V. aconitifolia*), french bean (*Phaseolus vulgaris*), soybean (*Glycine max*) and pigeonpea (*Cajanus cajan*) in India. The host crop exhibit general yellowing and the diseased fields can be recognized from far distance.

Experiments were conducted at Crop Research Centre of Pantnagar University to contain this disease through agronomic practices (manipulation in planting dates and inter-cropping with barrier (non-host) crops, insecticides (foliar and soil application, seed treatment) and host resistance. Low disease incidence was recorded in early (June) and late (August) plantings as well as in low plant spacing (5 cm). Seed dressing with Carbofuran 3G or Phorate 10G with two foliar sprays of Monocrotophos (0.1%) + Endosulphan (0.1%) also reduced the disease incidence. Use of barrier crops used as inter-cropping with mungbean, soybean and urdbean were not very effective. Some genotypes viz., Pant U19, 30 and 35, (urdbean) PM 1, 2, 3 and 4 (mungbeari) and PK 416, PK564, SL 142 (soybean) resistant to MYMV have been identified.

As none of the individual control measure was found highly effective against the disease due to high vector population and wide range of host plants serving as initial foci of the whitefly and the MYMV in this area, integrated management modules have been developed keeping the tolerant varieties as core.

**Section F Research Summary**  
**Integrated and Areawide Pest Management**  
**Approaches**  
**And Crop Management Systems**

Compiled by Steve Castle

Ten years of whitefly meetings and what do you have to show for it? *A well-managed pest.*

The story should end so simply, unfettered by dreary pontification about advances in knowledge and changes in practices that brought about quantum improvements in how we coexist with *Bemisia tabaci*, *Bemisia argentifolii*, or just plain *Bemisia*. But if simple statements don't suffice in a world obsessed with post facto analysis, then let it be stated that significant achievements were realized in each of the 6 sections that comprised the 5-Year Plans. The crowning achievement was that an outbreak pest has now been subdued through multi-disciplined contributions distilled and synthesized into viable control programs. Every abstract and presentation, coffee-break conversation and skeptical thought, every right idea and wrong direction contributed to the process of working out a solution to a devastating problem. But to focus only on the practical successes on the farm would be an injustice to the countless successes attained at more fundamental levels of investigation. Significant advancements were made in our understanding of *B. tabaci* as an organism with a complex genetics that affect host range, virus transmission, insecticide resistance and undoubtedly many other interactions with its biotic and abiotic environment. We gained valuable knowledge of how *B. tabaci* adapts physiologically to a hostile thermal environment, how developmental morphology proceeds temporally, how mouthpart anatomy influences feeding biology, how feeding biology influences virus transmission, and how virus transmission can be influenced by plant host and cultivar differences. The listing could go on at great length, but this would be redundant as all abstracts have been included in each annual edition of the 5-Year Plan Reviews. The combined set of Reviews serves as a testimonial to the breadth of work initiated in response to the outbreak crises, as well as to the power of an organized and collaborative approach to solving monumental pest management problems.

Unfortunately, some problems proved more solvable than others, and universal agreement over taxonomic identification happens to be among the others. But so far as Section F is concerned, correct taxonomy will always be more of an academic issue than a practical problem. What is done in the field to control whitefly populations will likely remain unchanged whether the target population is comprised of Type A, B, or Q, or a combination of all 3. Some might argue that the

particular biotype involved may have unique characteristics such as a particular insecticide resistance profile that should be factored into the choice of chemical treatment. Or perhaps there might be a host biotype preference among parasitoids as some considered to be a possibility in the shift from Type A to B that occurred in North America. Although differences among biotypes with respect to insecticide susceptibilities may exist, they also exist between populations within a biotype. Insecticide resistance profiles would unlikely ever become so specific and universal within a biotype as to be recognized as a characteristic and a basis of decision-making. On the other hand, biotype preferences could very well exist in certain parasitoids principally due to the interaction of the primary and tertiary trophic levels. Foraging parasitoids may be less likely to search and/or accept a whitefly host on one particular plant species compared to another. Therefore, variations in host plant ranges among *B. tabaci* biotypes could alter the efficiency of parasitoids that exhibit putative host plant preferences. However, biocontrol workers have a long way to go to first demonstrate that such a scenario occurs, then validate that it has any practical significance to the quality of management. Meanwhile, sound IPM practices tailored to a particular crop system that produce sterling results against 1 biotype will almost certainly produce the same results against a different biotype.

Every growing region across North America that struggled with whitefly control in the late 1980s and early 1990s benefited by the addition of far more effective chemical tools beginning in the early 1990s. While the potency, residual activity, and in many cases the specificity of these new tools are unparalleled, so too was the collective research that revealed not only their practical utility, but how and when the newer tools should be applied and incorporated into a coordinated control program. Sampling plans for whitefly in cotton and melons were developed and joined with action thresholds to produce highly effective control, especially in places where their implementation was preceded by extensive communication to, and education of, grower clientele. Nowhere else was the sampling and threshold foundation along with product label and IRM guidelines better developed than in Arizona. Industry representatives worked closely with Arizona scientists to construct product labels that limited per field use of pyriproxyfen and buprofezin to 1 application each per cotton season and mandated that purchasers of either product be certified through attendance at a training session. Nowhere else were education and training sessions in addition to cohesive guidelines, web site, and printed reference materials more skillfully delivered to clientele and subsequently implemented than in Arizona. The result of this expert blending of research and transfer has been an extraordinarily successful suppression of whitefly populations ever since 1996. That was the first year that the IGRs became available commercially and the strategy

for their implementation was formalized into a pest management framework. The overall effect of this combined action has been progressive decreases in whitefly pressure each year and a concomitant drop in insecticide applications that target whiteflies.

In addition to the aforementioned factors that contributed to the success of the Arizona IPM program for cotton, there are other intangible factors that have possibly played significant roles in the decline of whitefly numbers.

Certainly among these would be to what extent buprofezin and pyriproxyfen treatments act across developmental stages of whiteflies to produce their full effects on a population. Product descriptions may actually underestimate the full impact that each compound has when various stages are exposed in the field, including a number of sub-lethal effects on adults that occur beyond the well-recognized sterility effect produced in females by pyriproxyfen. Another factor that has begun to be quantified is the degree to which natural enemy populations are left intact following treatment with either of the IGRs, and what level of bio-residual activity (*sensu* Ellsworth and Naranjo) continues to act against whiteflies and other pest populations. Still another intangible is what role education and training sessions of growers and advisors played in leading to more effective area-wide control. More synchronous insecticide applications over wide areas may have been an outcome of the emphasis placed on timing the first IGR application according to whether or not the prescribed action threshold had been reached. The upshot of the thorough research that produced sampling plans and action thresholds, in addition to the outstanding communication and transfer of this information to the grower clientele, is that what was essentially delivered as an individualized, grower-by-grower program with the ultimate purpose of protecting each one's livelihood, soon transcended into a larger phenomenon that saw individual growers benefiting not only by their own knowledge-driven actions, but by the synchronous actions of all of their peers acting on the same knowledge. Rather than disjunct applications made in a patchwork manner over a region prior to 1996 (not having had the benefit of disciplined decision-making instilled by a coherent IPM framework), treatments against whiteflies beginning in 1996 instead became a de facto area-wide program of synchronous applications that yielded blanket-like suppression of whitefly populations.

The transformation of pest management practices that took place in Arizona beginning in 1996 makes an interesting study of how a combination of crisis and novelty can sublimate ordinary pest management into something extraordinary. Pest management as it was being practiced in Arizona prior to 1996 was anything but ordinary because sampling plans, action thresholds, and a cotton IRM had been under development for years and were all in place as a cohesive set of recommendations to growers by 1995. However, the impact these recommendations had on whitefly management in 1995

was still deficient as control of late-season generations was lost, resulting in poor cotton quality and yields. Among the repercussions of the 1995 whitefly crisis was a pervasive realization that another way had to be found to manage whiteflies in cotton. Fortunately, years of research by Arizona and Florida scientists had identified buprofezin and pyriproxyfen as highly effective compounds with unique modes of action. The volumes of data accumulated made it possible to rapidly put together a strong case for an emergency registration of both compounds. The package submitted for registration also benefited by the example in Israel where both compounds had been used effectively to help control whitefly populations. A tough resistance management component that precluded use of either compound more than once per season and mandated at least a 14-day separation between use of the IGRs was built into the package as well.

Where the novelty aspect enters into the formula that transformed whitefly pest management is in the nature of each compound's activity against whitefly populations and the special instructions required to maximize each product's potential. The unique, slow-acting properties of each product could well be deceptive to novice users otherwise accustomed to rapid knockdown by conventional insecticides. Increasing grower awareness at seminars and training sessions held at multiple locations throughout Arizona was an essential component to the successful use of the IGRs. During these sessions, emphasis was also placed on the critical timing required to produce the best possible results, and how sampling was the key to correct timing. The crisis of the previous year produced a sense of urgency to the process of adapting to novel products, and the novelty of the products demanded their deployment under the best IPM practices. Had some other pair of insecticides with more conventional activities been introduced, it is possible that growers would not have been mandated to attend training sessions in order to become certified to use the products. Their expertise would already have been in the use of conventional-acting products, and would therefore not have required the specialized training accompanied by an emphasis on using them with accurate observance of action thresholds. Or if they had, the same important message of employing the recommended sampling program and treating at the right threshold would have been given, as it had been previously in 1995, but it would have fallen too well within their comfort zone of operation so long as they had only to be concerned about new products that acted the same as all previous products. I submit that the novelty of the IGRs altered the spray-and-knockdown culture to such a degree that, in combination with the crisis mode still pervasive from the previous year's disaster, growers and PCAs became more receptive to the teachings of IPM than ever before. The product of this grower education, mandated collaboratively by company representatives for both products and by University of Arizona scientists that recognized the need for grower training through their own

research experience, was a sensational turnaround in the first year that has been sustained ever since.

While other regions in North America that had previously suffered under whitefly attack also benefited by collective advances in whitefly management, none developed coordinated programs as sophisticated as the one in Arizona. Early on, whitefly outbreaks had flourished from one region to another, but there were no satisfactory management solutions because the necessary tools were as yet underdeveloped. A few more years of experience in whitefly management, some more time to develop the basic tools of IPM, and perhaps the next outbreak situation would be suppressed. That opportunity arose in 1995 in Arizona at a time when all of the components of good IPM were in place and in practice, yet still were inadequate against late-season whiteflies in cotton. It was this latter-day crisis, however, that spawned the search for a reliable solution to the one-too-many outbreaks at the propitious time that groundwork for a novel solution had already been completed. The combination of events resulted in the Arizona cotton IPM for whitefly that has few equals in the realm of IPM solutions in terms of the quality of research, education, and implementation that have contributed to its sustaining success.

#### **Development, Integration, and Delivery and Implementation**

Progress was made in each one of the research areas that subdivided Section F (see Table F synopsis). Development in whitefly-crop interactions has continued to advance with life history studies being conducted throughout the year in various crops and ornamental hosts. The cumulative data from these studies and ones conducted previously in cotton will provide enormously valuable information on the population dynamics of *B. tabaci* as well as on the relative efficiencies of various management tactics. Development of heuristic frameworks describing the interacting strategies and components of IPM in one case, and in another the factors that contribute to whitefly outbreaks, helped synthesize existing and new information into conceptual models that advanced understanding of why outbreaks occur and how management can most effectively respond. Most impressively, many of the concepts contained in the IPM Pyramid have been validated by real world examples where implementation has brought about management successes. In addition to the case already described for Arizona cotton IPM, a coordinated program was instituted in the Dominican Republic to counter epidemics of tomato yellow leaf curl virus (TYLCV) in production tomato fields. By implementing a mandatory crop free period to reduce inoculum potential, incidence of TYLCV in tomatoes has greatly declined while production has fully recovered. This is an example of avoiding a pest or disease problem by manipulating crop plantings and seasons as spelled out in the Avoidance layer of the IPM Pyramid.

Further refinement in the implementation of pest management will continue to improve existing practices. For example, reassessment of the data that contributed to setting the action threshold at 5 adults/leaf showed that predator conservation increased with higher thresholds or if the date of first application was delayed. Improved integration of natural and chemical controls will occur as a more complete picture develops over the consequences of applying inputs to agricultural systems. In this case, slight restraint on the timing of the first application may pay dividends in the form of greater natural control and reduced pesticide use overall. On a wider scale, integration of chemical classes within and across crop seasons is a goal of cross-commodity efforts to conserve insecticide efficacies. This becomes more of a challenge as insecticides gain registrations on additional crops. The most pressing concern is that multiple uses in multiple crops will add up to too many uses on a pest that moves sequentially from one crop to another. Resistance selection pressure can be expected to increase if exposure levels increase with multiple-crop usage of a particular insecticide or insecticide class. However, much progress is being made in Arizona to harmonize insecticide use across commodities so as to limit maximum exposures to any one product or insecticide class.

**Table F. Integrated and Areawide Pest Management Approaches, and Crop Management Systems.**

Research Approaches <sup>a</sup>	Year 1 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Development:</b>				
<b>Study whitefly-crop interactions<sup>b</sup> as cultural components that affect population dynamics, e.g., water, nutrients, plant population, planting/termination/harvest dates, other farm practices, intercrop relationships.</b>	Identify potential beneficial or exacerbating farm practices or inputs for testing.	X but limited		Only minor progress has been made on this approach (since last 5-yr review), & mainly in area-wide programs. This work is correlative, & little experimental work has been planned for or reported. Past work identified the potential or described the role of fertility status, water-stress & some other agronomic factors on <i>Bemisia</i> population dynamics. Conceptual discussion was presented on the role of pesticidal & non-pesticidal factors on <i>Bemisia</i> outbreaks.
<b>Develop behavioral barriers<sup>b</sup> to whitefly colonization and population development, e.g., mulches, trap crops, intercropping, row covers, etc.</b>	Review potential behavioral disrupters and evaluate as potential IPM components.	X		Progress has been made in several areas: <ul style="list-style-type: none"> <li>• row covers and screens as physical barriers,</li> <li>• mulches and oils as behavioral barriers,</li> <li>• living mulches as behavioral barriers.</li> </ul>
<b>Integration:</b>				
<b>Develop Integrated Pest Management<sup>c</sup> systems using dual or multiple control tactics, e.g., cultural, biological, chemical, host plant resistance, etc.</b>	Identify candidate dual or multiple control tactic systems, e.g., IGRs and natural enemy conservation.	X		Significant activity on this goal has occurred: <ul style="list-style-type: none"> <li>• Insect Growth Regulators &amp; biological control in cotton (conservation)</li> <li>• imidacloprid &amp; other chemical control tactics &amp; various forms of biological control, especially in vegetables</li> <li>• studies of direct &amp; indirect effects of chemical control on bio-control agents.</li> </ul>
<b>Integrate sampling with other key components of IPM systems, e.g., thresholds, economics, decision-making, biological control, etc.</b>	Develop or modify sampling systems for new crops; integrate with thresholds and decision-making.	X		Limited progress has been made in this area: <ul style="list-style-type: none"> <li>• <i>Bemisia</i> distributions have been examined in tomato,</li> <li>• new binomial sampling system for large nymphs in cotton, &amp; integration with thresholds for IGR decisions</li> <li>• sampling &amp; IGR re-treatment decisions tested in cotton.</li> </ul>

**Table F. Integrated and Areawide Pest Management Approaches, and Crop Management Systems. (Continued)**

Research Approaches <sup>a</sup>	Year 1 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Delivery and Implementation:</b>				
<b>Elevate single field/farm practices to areawide community-based contexts; develop methodology for installing and evaluating areawide control technologies and their impact.</b>	Identify agricultural communities amenable to areawide management; conduct thorough pre-implementation evaluation.	X		<p>Significant progress was made in this area mainly in cotton:</p> <ul style="list-style-type: none"> <li>• areas dominated by cotton were identified in AZ &amp; CA for implementation of cooperative programs.</li> <li>• areas of melon and vegetable production were identified in TX for potential area-wide programs.</li> <li>• area-wide sampling, &amp; decision-making was the main focus of most programs; however, coordinated natural enemy releases were also conducted.</li> </ul>
<b>Implement and deliver Integrated Pest Management and Integrated Crop Management systems or system components to clientele.</b>	Develop and distribute provisional IPM & ICM recommendations.	X		<p>Continued progress was made in this area:</p> <ul style="list-style-type: none"> <li>• IPM recommendations were distributed AZ, CA, Mexico &amp; FL; bilateral discussions between Brazil &amp; U.S. took place.</li> <li>• IPM &amp; ICM guidelines were coordinated in AZ cotton.</li> </ul>

<sup>a</sup> See Tables A to E for additional complementary research.

<sup>b</sup> See Tables A for additional complementary research.

<sup>c</sup> See Tables E for additional complementary research.

**Table F. Integrated and Areawide Pest Management Approaches, and Crop Management Systems.**

Research Approaches <sup>a</sup>	Year 2 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Development:</b>				
<b>Study whitefly-crop interactions<sup>b</sup> as cultural components that affect population dynamics, e.g., water, nutrients, plant population, planting/termination/harvest dates, other farm practices, intercrop relationships.</b>	Determine nature and character of relationship between interaction and whitefly population dynamics.	X		Nitrogen fertilization at different rates in cotton and its impact on whitefly population densities and honeydew deposition was studied. Considerable development occurred on cross-commodity integration of pesticides used in multi-cropped situations and in conceptualization of the multiple levels and factors upon which whitefly management depends.
<b>Develop behavioral barriers<sup>b</sup> to whitefly colonization and population development, e.g., mulches, trap crops, intercropping, row covers, etc.</b>	Conduct field-level trials; quantify impact to crop and whitefly dynamics	X		Investigations on intercropping took place in both desert and tropical environments. Although reductions in whitefly densities were observed in both systems, further experimentation is required to establish the effectiveness of the trap crops relative to more conventional management techniques.
<b>Integration:</b>				
<b>Develop Integrated Pest Management<sup>c</sup> systems using dual or multiple control tactics, e.g., cultural, biological, chemical, host plant resistance, etc.</b>	Initiate field testing of candidate systems.	X		A number of field studies employed multiple tactics directed against whitefly populations. Biorational insecticides were examined in combination with IGRs and other biopesticidal agents such as <i>Beauveria bassiana</i> for control efficacy of silverleaf whitefly. There was an indication of inhibitory action by <i>B. bassiana</i> when used in combination with imidacloprid as well as deleterious effects to predators contacted by <i>B. bassiana</i> treatments. Neem products were used to reduce whitefly populations and incidence of yellow mosaic virus in India. A melon trap crop was integrated with chemical control to focus potentially disrupting treatments into a limited area while preserving natural mortality factors in cotton as the principle crop.



**Table F. Integrated and Areawide Pest Management Approaches, and Crop Management Systems. (Continued)**

Research Approaches <sup>a</sup>	Year 2 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Integrate sampling with other key components of IPM systems, e.g., thresholds, economics, decision-making, biological control, etc.</b>	Establish practical utility of system through economic analyses; field efficiencies and costs.	X		Analysis of types and patterns of chemical treatments made on a large number of cotton fields in central Arizona over a 4 year period revealed extraordinary differences in the number of treatments and amount of time that whiteflies exceeded threshold levels prior to and following the advent of the IGRs buprofezin and pyriproxyfen. The proactive initiative taken by Arizona growers to pursue chemical use harmonization across commodities required consideration of all aspects of pest and crop management. A similar whole system appraisal was made in the San Joaquin Valley with an emphasis on integrating multiple practices with diverse insecticide classes as part of an insecticide resistance management program.
<b>Delivery and Implementation:</b>				
<b>Elevate single field/farm practices to areawide community-based contexts; develop methodology for installing and evaluating areawide control technologies and their impact.</b>	Install control technologies into community; develop systems for evaluation.	X		Large areas in the San Joaquin Valley observed specific guidelines for IPM and IRM in cotton with evaluations continuing on the benefits attained over areas that did not observe these guidelines. Community wide evaluations were made on quality of whitefly management according to chemical control practices. The successful IPM and IRM programs practiced in Arizona cotton continued for a third consecutive year. Further cross-commodity development of these programs is under way.

**Table F. Integrated and Areawide Pest Management Approaches, and Crop Management Systems. (Continued)**

Research Approaches <sup>a</sup>	Year 2 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Implement and deliver Integrated Pest Management and Integrated Crop Management systems or system components to clientele.</b>	Conduct whole farm/operation demonstrations of IPM systems.	X		A 'best agricultural practices' demonstration project was conducted on 50.5 acres at the University of Arizona Maricopa Agricultural Center that included inputs from extension specialists in agronomy, entomology, irrigation management, weed sciences and plant pathology according to university recommendations. Whitefly management was fully integrated with management of other insect pests and required only a single application of pyriproxyfen. Lint yields of 2.81 bales/acre were higher than the historical as well as the 1998 farm-wide average. An integrated areawide management program involving the cooperation of growers, PCAs, ginners and state and university researchers was expanded during a second year in the San Joaquin Valley.

<sup>a</sup> See Tables A to E for additional complementary research.

<sup>b</sup> See Tables A for additional complementary research.

<sup>c</sup> See Tables E for additional complementary research.

**Table F. Integrated and Areawide Pest Management Approaches, and Crop Management Systems.**

Research Approaches <sup>a</sup>	Year 3 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Development:</b>				
<b>Study whitefly-crop interactions<sup>b</sup> as cultural components that affect population dynamics, e.g., water, nutrients, plant population, planting/termination/harvest dates, other farm practices, intercrop relationships.</b>	Identify mechanisms governing relationship and alter or manipulate factors that suppress whitefly dynamics.	X		Progress was made with studies in Texas on seasonal dynamics of <i>Bemisia</i> on spring collards, and the impact of cotton defoliants on <i>Bemisia</i> and parasitoid populations. Work continued in California on the affect of various nitrogen levels in cotton with <i>Bemisia</i> population densities
<b>Develop behavioral barriers<sup>b</sup> to whitefly colonization and population development, e.g., mulches, trap crops, intercropping, row covers, etc.</b>	Apply promising technologies to high-value crop systems; field test and evaluate	X		Further progress with research on: <ul style="list-style-type: none"> <li>• Living ground covers for managing whitefly-transmitted gemini viruses</li> <li>• Behavioral disruptance by UV-blocking barriers</li> <li>• Melons as a trap crop for cotton</li> </ul>
<b>Integration:</b>				
<b>Develop Integrated Pest Management<sup>c</sup> systems using dual or multiple control tactics, e.g., cultural, biological, chemical, host plant resistance, etc.</b>	Continue field testing & evaluate feasibility of large scale testing; add components as necessary.	X		Much progress with integrating control tactics: <ul style="list-style-type: none"> <li>• Life table evaluation of both natural and insecticide-based mortalities</li> <li>• Compatibility of IGR's for whitefly control in greenhouses</li> <li>• IPM development in cotton for <i>Bemisia</i> and other cotton pests</li> <li>• Augmentative biocontrol using crop transplants inoculated with parasitoids in Admire<sup>®</sup>-treated fields</li> </ul>
<b>Integrate sampling with other key components of IPM systems, e.g., thresholds, economics, decision-making, biological control, etc.</b>	Integrate additional control components into sampling, threshold & decision-making systems	X		<ul style="list-style-type: none"> <li>• Fourth consecutive year of monitoring <i>Bemisia</i> populations in the Imperial Valley using the CC trap.</li> <li>• Sampling-based refinement of action thresholds for IGR's in Arizona cotton</li> </ul>

**Table F. Integrated and Areawide Pest Management Approaches, and Crop Management Systems. (Continued)**

Research Approaches <sup>a</sup>	Year 3 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Delivery and Implementation:</b>				
<b>Elevate single field/farm practices to areawide community-based contexts; develop methodology for installing and evaluating areawide control technologies and their impact.</b>	Identify additional IPM/ICM compatible components. Re -assess and adapt program. Conduct areawide economic analyses.	X		<ul style="list-style-type: none"> <li>• Economic analysis of the use of IGR's in Arizona cotton</li> <li>• Further development of cross-commodity planning and cooperation</li> <li>• Multivariate techniques used to assess parasitoid establishment in the Imperial Valley based on crop and land use</li> </ul>
<b>Implement and deliver Integrated Pest Management and Integrated Crop Management systems or system components to clientele.</b>	Expand sites of testing with grower cooperators; conduct validation studies.	X		<ul style="list-style-type: none"> <li>• Sticky cotton bulletin published by University of Arizona and Cotton Incorporated</li> <li>• International development of IPM for managing whiteflies and geminiviruses</li> <li>• Development of <i>Bemisia</i>-resistant alfalfa cultivars</li> <li>• Crop and pest management demonstration project on cotton in Arizona</li> </ul>

<sup>a</sup> See Tables A to E for additional complementary research.

<sup>b</sup> See Tables A for additional complementary research.

<sup>c</sup> See Tables E for additional complementary research.

**Table F. Integrated and Areawide Pest Management Approaches, and Crop Management Systems , 2001-2002.**

Research Approaches <sup>a</sup>	Year 4-5 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Development:</b> <b>Study whitefly-crop interactions<sup>b</sup> as cultural components that affect population dynamics, e.g., water, nutrients, plant population, planting/termination/harvest dates, other farm practices, intercrop relationships.</b>	Refine system, add other compatible components, evaluate economic impact; conduct field testing and evaluations.	X		Significant progress made in the realm of intercrop relationships with comparative life history studies of <i>B. tabaci</i> on multiple crop and non-crop hosts. An economic impact analysis was developed for IGR use in Arizona as well as for pesticide use in general.
	Conduct economic analyses and determine next level of IPM/ICM systems evaluation. Develop recommendations of best management practices.			The IPM Pyramid was advanced as a conceptual framework for understanding the interacting components in an IPM program for whiteflies. The Outbreak Pyramid was advanced as a conceptual device for understanding causes of outbreaks. Avoidance as a major component of IPM along with conservation of natural enemies through selective chemistry were advanced as topics and supported by research. A crop-free period was pivotal in breaking epidemics of TYLCV in the Dominican Republic was elucidated, but so too the role of symptomless weeds in perpetuating the virus.
<b>Develop behavioral barriers<sup>b</sup> to whitefly colonization and population development, e.g., mulches, trap crops, intercropping, row covers, etc.</b>	Refine system, add other components, and conduct economic feasibility analyses.	X		Synthesis of field exclusion techniques published as a review article. Cost-benefit analysis of insect-exclusion screens used on greenhouses in Israel demonstrating benefit for the grower and consumer.
	Summarize and evaluate results; prepare crop systems-specific recommendations.			
<b>Integration:</b> <b>Develop Integrated Pest Management<sup>c</sup> systems using dual or multiple control tactics, e.g., cultural, biological, chemical, host plant resistance, etc.</b>	Initiate large-scale experiments; incorporate economic evaluation.	X		Large plot studies conducted in Arizona demonstrated marginal mortalities associated with different control tactics or through various natural sources. Evaluation of parasitoid releases in concert with imidacloprid treatment of spring melons.
	Evaluate multiple component system as potential deliverable; prepare recommendations.			Recommendations prepared and delivered in Mexico's Yaqui Valley and in the Dominican Republic.

**Table F. Integrated and Areawide Pest Management Approaches, and Crop Management Systems, 2001-2002. (Continued)**

Research Approaches <sup>a</sup>	Year 5 Goals Statement	Progress Achieved		Significance
		Yes	No	
<b>Integrate sampling with other key components of IPM systems, e.g., thresholds, economics, decision-making, biological control, etc.</b>	Evaluate in whole field systems. Identify weaknesses; target improvements.	X		Further analysis of sampling/action threshold database that demonstrates greater conservation of predator species when the initial treatment threshold of 10 adults/leaf is used rather than the standard of 5 adults/leaf.
	Evaluate redesigned decision systems; continue field testing and economic analyses			
<b>Delivery and Implementation: Elevate single field/farm practices to areawide community-based contexts; develop methodology for installing and evaluating areawide control technologies and their impact.</b>	Formulate clientele surveys; develop & begin to implement protocols for evaluating areawide technologies.	X		Pesticide use information in Arizona as a validation of improved pest management; in California, pesticide use incorporated into GIS analyses to evaluate whitefly distributions.  Cross-commodity efforts to harmonize pesticide use and to identify weaknesses in cross registration of neonicotinoids and IGRs.
	Refine, reevaluate and identify weaknesses. Formulate recommendations for future areawide management systems. Conduct surveys.			
<b>Implement and deliver Integrated Pest Management and Integrated Crop Management systems or system components to clientele.</b>	Incorporate new information and economics into recommendations.	X		Development of web sites as whitefly management resources for clientele to access and download.  Crop free period in Dominican Republic and recommendations for dealing with symptomless weed hosts, beans and peppers.
	Validate new components; finalize recommendations; expand to new crops.			

<sup>a</sup> See Tables A to E for additional complementary research.

<sup>b</sup> See Tables A for additional complementary research.

<sup>c</sup> See Tables E for additional complementary research.

## Appendix A: Selected National and International *Bemisia* Research and Information Coordination

### Selected National and International *Bemisia* Research Projects and Information Coordination

Several national and international cooperative *Bemisia* projects have been implemented worldwide to facilitate communication, research, technology transfer, and rapid response to problem solving (Appendix IX, Table 1). Additional information on these efforts is available on the listed websites. The list is not all inclusive, and these are numerous local groups with commodity-orientation that meet informally to exchange information and experiences, and discuss current control efforts. It does, however, serve as a starting point for identifying sources of whitefly information and contacts to gain additional information.

These international, regional and national networks for *B. tabaci* and *B. tabaci*-transmitted plant viruses and other whitefly species essentially function as information exchanges. The active international and national collaborative research projects have improved operational efficiency, by optimizing returns from financial, physical and human resources. Open lines of communication and discussion identified priority research areas and focused efforts that increased returns for each invested research dollar compared to numerous small projects utilizing diverse approaches. The economic, environmental and social impact of research coordination and information exchange groups for the *B. tabaci* species complex worldwide have stimulated much new and useful research resulting in advanced knowledge of biology, ecology, physiology, and virus-vector interactions of the species complex.

Table 1. Selected national and international *Bemisia tabaci* research and information exchange projects.

Project	Year Created	Accomplishments
International <i>Bemisia</i> Newsletter	1984	Information exchange, meeting announcements, electronic conference, Whitefly-L. Coordinators: D. Gerling <a href="mailto:dangr@ccsg.tam.ac.il">dangr@ccsg.tam.ac.il</a> , W. Jones <a href="mailto:wjones@weslaco.ars.usda.gov">wjones@weslaco.ars.usda.gov</a>
Sticky Cotton Action Team, Cotton Incorporated (CI), Raleigh, NC	1990	Provided communication between the cotton industry and researchers on sticky cotton research. CI and state support committees provided \$1.9 million for research. Coordinated multi-federal, -state, -university, and -industry agencies efforts that resulted in efficient whitefly management and reduced sticky cotton problems. Financed research also contributed to implementation of insect growth regulator use patterns and resistance management. Also, stimulated improved sticky cotton lint sampling methods, demonstration of the relationships of whiteflies and aphids to sticky cotton, improved sticky cotton measurement instrumentation and enzyme approaches to eliminate honeydew sugars from contaminated lint. Coordinators: R.L. Nichols <a href="mailto:BNichols@cottoninc.com">BNichols@cottoninc.com</a> , W. Lalor <a href="mailto:BLalor@cottoninc.com">BLalor@cottoninc.com</a>
Latin America and Caribbean Whitefly and Geminivirus Network	1992	Quarterly newsletter, Mosca Blanca al Dia, <a href="http://www.catie.ac.cr/cooperacion/boletin.htm">www.catie.ac.cr/cooperacion/boletin.htm</a> Promotes research, diagnostic activities, and IPM technology transfer in 21 countries including Spain. Annual meeting. Network host: Tropical Agricultural Research and Higher Education Center (CATIE) Costa Rica. Coordinator: L. Hilje, <a href="mailto:lhilje@catie.ac.cr">lhilje@catie.ac.cr</a>
Integrated Management of Whitefly in Northwestern Mexico	1992	Identified soybeans as the key host and eliminated it in the cropping system to break the reproductive host continuity. Published eight scientific summaries of 262 articles. Covered 4 states, Baja California Norte, Baja California Sur, Sonora and Sinola. Participating agencies are the National Research Institute of Forestry, Agriculture and Animal Husbandry (INIFAP), Regional Centers for Agricultural Research and Mexicali Experiment Station. Coordinators: J. L. Martinez Carrillo <a href="mailto:jlmc@cirno.inifap.conacyt.mx">jlmc@cirno.inifap.conacyt.mx</a> , J. Jose Pacheco <a href="mailto:Pacheco@cirno.inifap.conacyt.mx">Pacheco@cirno.inifap.conacyt.mx</a>

Five-Year National Research and Action Plan for Development of Management and Control Methodology for the Sweetpotato Whitefly. Revised as Five-Year Silverleaf Whitefly Research, Action and Technology Transfer Plan, in 1997.	1992-1996 1997-2001	National and international cooperative programs that led to action thresholds, effective chemical control, new chemistry and resistance management principles and agricultural community management systems. Extensive basic biology, physiology, ecology for long-term suppression systems (see text). Annual meetings. Nine annual publications with over 1000 abstracts of research progress. A bibliography of more than 4,800 entries: <a href="http://www.wcrl.ars.usda.gov">http://www.wcrl.ars.usda.gov</a> Coordinators: T. J. Henneberry <a href="mailto:thenneberry@wcrl.ars.usda.gov">thenneberry@wcrl.ars.usda.gov</a> , N. Toscano <a href="mailto:ntoscano@ucr.ac1.ucr.edu">ntoscano@ucr.ac1.ucr.edu</a> , W. Jones, T. Perring <a href="mailto:perring@ucr.ac1.ucr.edu">perring@ucr.ac1.ucr.edu</a> , R. Faust <a href="mailto:rmf@ars.usda.gov">rmf@ars.usda.gov</a>
The Safe and Effective Use of Pesticides Project	1995	USAID-funded project carried out under the umbrella of the Middle East Peace Process. Annual meetings of IPM and public health experts in Egypt, Jordan, Israel and Palestine for information exchange. Pest monitoring, training of farm workers, blood testing, reports on safety progress. Information <a href="http://www.mercph.org">www.mercph.org</a> Coordinating Center: National Institute of Health (NIH), Bethesda, MD. Coordinator: K. Abdo <a href="mailto:abdok@niehs.nih.gov">abdok@niehs.nih.gov</a>
CGIAR Whitefly IPM Project	1996	Consultative Group for International Agricultural Research (CGIAR) project, under the umbrella of the Systemwide Program for Integrated Pest Management (see text). Coordinating Center: International Center for Tropical Agriculture, Cali, Colombia. Coordinator: P. Anderson <a href="mailto:p.anderson@cgiar.org">p.anderson@cgiar.org</a>
European Whitefly Studies Network	1997	Gathered and disseminated whitefly information. Working group meetings. Established website: <a href="http://www.jic.bbsrc.ac.uk/hosting/eu/ewsn">www.jic.bbsrc.ac.uk/hosting/eu/ewsn</a> Network host: John Innes Center, United Kingdom. Coordinator: Ian Bedford <a href="mailto:ian.Bedford@bbsrc.ac.uk">ian.Bedford@bbsrc.ac.uk</a>
Brazilian Agricultural Research Enterprise (EMBRAPA) Project (1998-2000)	1998	National Whitefly Commission by EMBRAPA (3 year project). Identified priorities, coordinated efforts of the ministry, federal, state, agricultural agencies and growers. Identified infested areas, identified effective insecticides, transferred technology from other countries. Coordinator: MRV Oliveira <a href="mailto:vilarin@cenargen.embrapa.br">vilarin@cenargen.embrapa.br</a>
Brazilian virology project to characterize geminiviruses tomatoes	1998	Characterized 8 new tomato-infecting geminiviruses in Brazil. Developed standardized virus survey protocols, shared results, initiated varietal resistance efforts. Coordinators: F. Murilo Zerbini <a href="mailto:zerbini@mail.ufv.br">zerbini@mail.ufv.br</a> , S.G. Ribeiro <a href="mailto:simone@cenargen.embrapa.br">simone@cenargen.embrapa.br</a>

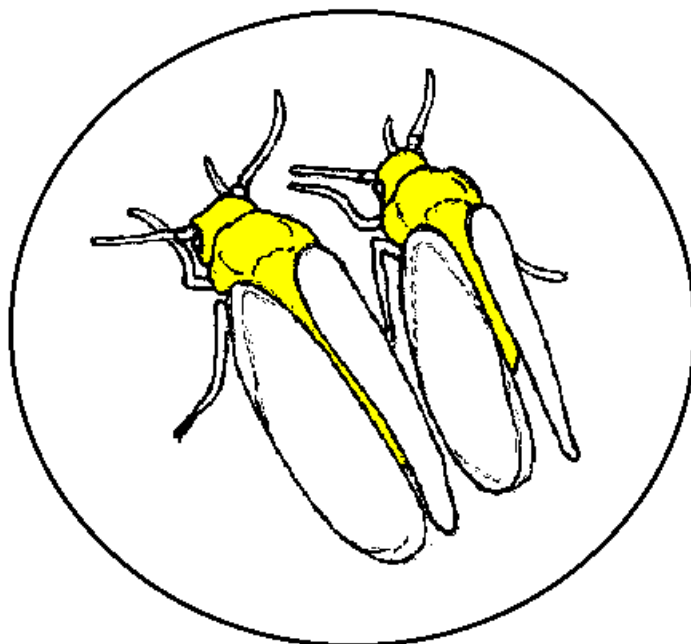
<sup>a</sup> This table was reproduced, with the permission of the author from the publication "History, Current Status and Collaborative Research Projects for *Bemisia tabaci* by Oliveira, M.R.V., T. J. Henneberry and P. Anderson. 2001. Crop Protection 20: 709-723.



Appendix B. Bibliography of *Bemisia tabaci* (Gennadius) & *Bemisia argentifolii* Bellows and Perring

Complete Bibliography of

***Bemisia tabaci* (Gennadius)  
&  
*Bemisia argentifolii* Bellows and Perring**



**Steven E. Naranjo  
George D. Butler, Jr.  
Thomas J. Henneberry**

**February 2002**

## Bibliography of *Bemisia tabaci* (Gennadius) and *Bemisia argentifolii* Bellows & Perring

February 2002

Steven E. Naranjo, George D. Butler, Jr., and Thomas J. Henneberry

*Bemisia tabaci* (Gennadius) was described over 100 years ago as a tobacco pest in Greece and has since become one of the most important pests of food, fiber and ornamental crops in the world. The recent geographic expansion of this pest has been closely associated with a new and more virulent biotype, known widely as biotype B, that may represent a new species (*Bemisia argentifolii* Bellows and Perring). The taxonomy and systematics of this pest remains confused and controversial. Recent analyses, using a wide array of morphological and molecular tools, suggest that *B. tabaci* may represent a species complex. Regardless, this insect has had an extraordinary impact on agricultural and horticultural production systems worldwide. In addition to direct feeding damage, this insect vectors a number of devastating plant viruses, causes debilitating plant disorders of unknown etiology and, by the excretion of honeydew, reduces the quality of harvested products. We face monumental challenges in the development of economically-efficient and environmentally-sound management systems for *Bemisia*. The insect has a reported host-range of well over 500 plant species, a high reproductive rate, the ability to readily disperse among hosts and breed year-round, and a propensity to develop resistance to insecticides. Previous distributions of this insect were limited to regions between the 30<sup>th</sup> parallels. However, in the past two decades, the pest has invaded every continent in the world except Antarctica, and commercial trade has facilitated the regular occurrence of populations in temperate greenhouse production systems in many parts of the world.

In 1992 we began to catalog the world literature on *Bemisia tabaci* and *Bemisia argentifolii* as an aid to researchers, educators, extension personnel, agricultural producers, industry, and governmental administrators. In 1995 we published a bibliography that attempted to cover the world literature through the end of 1994 (Butler et al. 1995). Since that time, addenda to this bibliography have been published annually (Naranjo et al. 1996, 1997, 1998, 1999, 2000, 2001). Here we present the entire bibliography based on literature cataloged through the end of February 2002. This bibliography attempts to cover all of the scientific literature on *B. tabaci* and *B. argentifolii* (peer-reviewed journal articles; books and book chapters; symposia and workshop proceeding and abstracts; and governmental, university, and industry reports). It also attempts to cover much of the literature on viruses vectored by *Bemisia* and that dealing with issues related to honeydew contamination.

This bibliography has been compiled from a number of sources. The Current Awareness Literature Service of the National Agricultural Library (NAL) has been instrumental in conducting searches of abstracting databases, such as AGRICOLA (NAL), AGRIS (United Nations, Food & Agriculture Organization), Biological Abstracts (BIOSIS), Commonwealth Agricultural Bureaux Abstracts (CABI), and Zoological Records (BIOSIS). We have also searched Current Contents (Institute for Scientific Information), Dissertation Abstracts International (University Microfilms International), and our personal reprint collections. Finally, this bibliography includes citations from the two annotated bibliographies of Cock (1986, 1993), and the proceedings of various international conferences, workshops and symposia. We have noted throughout the bibliography those references which have been cited by Cock. His bibliographies provide abstracts for many of the papers and because many references may be difficult to obtain we thought researchers might find it useful to check Cock's abstracts before beginning the arduous task of locating original papers.

The current bibliography and future annual addenda can be downloaded electronically from the Western Cotton Research Laboratory web site at ([www.wcrl.ars.usda.gov](http://www.wcrl.ars.usda.gov)) in ProCite (2.0 for DOS; 4.0 for Windows), MS Word (2000), and ASCII text formats. The bibliography also can be searched directly on this web site. Finally, a runtime version of the ProCite software and the databases can be provided on CD (contact SEN at [snaranjo@wcrl.ars.usda.gov](mailto:snaranjo@wcrl.ars.usda.gov)).

1. Abak, K., Yilmaz, M.A. and Kesiei, S. 1991. Problems caused by TYLCV in Turkey and means to fight the disease. pp. 28-30. *In* H. Laterrot and C. Trousse (Eds.), Proceedings EEC Seminar: Resistance of the tomato to TYLCV, Avignon, France.
2. Abbass, A.K., Al-Hitty, A.A., Ali, A.A. and Hassan, N.M. 1988. [Evaluation of various control practices and their time of application against the whitefly (*Bemisia tabaci* Genn.) and some other pests on fall cucumber]. *J. Agric. Water Resources Res.* 7(1): 123-142. [Cock (1993), Arabic, English summary]
3. Abboud, R. and Ahmad, M. 1998. Effect of temperature and prey-species on development of the immature stages of the coccinellid, *Serangium parcesetosum* Sicard. [Arabic, English summary]. *Arab J. Plant Prot.* 16(2): 90-93.
4. Abd-Allah, Y.E.Y. 1997. Damage and damage loss assessments of certain insect pests attacking cotton in Egypt. Ph.D. Dissertation, Ain Shams University, Cairo, Egypt, 192 pp.
5. Abd-El-Kareim, A. 1998. Searching rate and potential of some natural enemies as bio-agent against the cotton whitefly, *Bemisia tabaci* Genn. (Hom., Aleyrodidae). *J. Appl. Entomol.* 122(8): 487-492.
6. Abd-Rabou, S. 1996. Egyptian Aleyrodidae. *Acta Phytopathol. Entomol. Hungarica* 31(3-4): 275-285.
7. Abd-Rabou, S. 1997. Hosts, distribution and vernacular names of whiteflies (Homoptera: Aleyrodidae) in Egypt. *Moshtohor Ann. Agric. Sci. (Egypt)* 35(2): 1029-1048.
8. Abd-Rabou, S. 1998. Inundative releases of *Encarsia formosa* Gahan (Hymenoptera: Aphelinidae) for the control of *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae) on outdoor crops. *Acta Phytopathol. Entomol. Hungarica* 33(3-4): 389-394.
9. Abd-Rabou, S. 1998. Parasitoids attacking *Bemisia tabaci* (Genn.) (Hom. Aleyrodidae) in Egypt. *Bull. Lab. Entomol. 'Agraria Filippo Silvestri' Portici* 54: 11-16.
10. Abd-Rabou, S. 1999. Biological control of the cotton whitefly, *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae), in Egypt. *Shashpa* 6(1): 53-57.
11. Abd-Rabou, S. 1999. The efficacy of indigenous and imported predators utilized in the biological control of *Bemisia tabaci* biotype "B" (Homoptera: Aleyrodidae) in greenhouse. *Acta Phytopathol. Entomol. Hungarica* 34(4): 333-339.
12. Abd-Rabou, S. 1999. Follow up of imported and indigenous natural enemies released to control whiteflies (Homoptera: Aleyrodidae) in Egypt. *Acta Phytopathol. Entomol. Hungarica* 34(3): 241-244.
13. Abd-Rabou, S. 1999. New records on whiteflies in Egypt. *Egyptian J. Agric. Res.* 77(3): 1143-1146.
14. Abd-Rabou, S. 2000. Role of *Encarsia inaron* (Walker) (Hymenoptera: Aphelinidae) in biological control of some whitefly species (Homoptera: Aleyrodidae) in Egypt. *Shashpa* 7(2): 187-188.
15. Abdel-Baky, N.F. and Abdel-Salam, A.H. 2000. Relative abundance of the silverleaf whitefly, *Bemisia argentifolii* Bellows and Perring, on squash plants in Egypt. *Pakistan J. Biol. Sci.* 3(2): 223-230.
16. Abdel-Fattah, M.I., Hendi, A. and El-Said, A. 1986. Ecological studies on parasites of the cotton whitefly, *Bemisia tabaci* (Genn.) in Egypt. *Bull. Entomol. Soc. Egypt Econ. Ser.* 14: 95-105. [Cock (1993)]
17. Abdel-Fattah, M.I., Hendi, A. and El-Said, A. 1987. Abundance of *Bemisia tabaci* (Genn.) associated with common weeds in tomato fields at Shebin El-kom region, Egypt. (Homoptera: Aleyrodidae). *Bull. Entomol. Soc. Egypt* 65: 109-117. [Cock (1993)]
18. Abdel-Fattah, M.I., Hendi, A., Kolaib, M.O. and El-Said, A. 1985. Studies on *Prospatella lutea* Masi, a primary parasite of the cotton whitefly, *Bemisia tabaci* (Genn.) in Egypt. (Hymenoptera: Aphelinidae). *Bull. Entomol. Soc. Egypt* 65: 119-129. [Cock (1993)]
19. Abdel-Fattah, S.A.S., Sharaf, I.M.R. and El-Sebae, A. 1983. [Performance of flucythrinate (Cybolt) against a wide spectrum of agricultural pests in Egypt. *Arab J. Plant Prot.* 1(2): 74-78. [Cock (1986), Arabic, English summary]
20. Abdel-Gawaad, A.A., El-Sayed, A.M., Shalaby, F.F. and Abo-El-Ghar, M.R. 1990. Natural enemies of *Bemisia tabaci* Genn. and their role in suppressing the population density of the pest. [Arabic summary]. *Agric. Res. Rev.* 68: 185-195. [Cock (1993)]
21. Abdel-Megeed, M.I., Hegab, M.F., Hegazy, G.M. and Kamel, M.H. 1998. Association of certain weather factors with population dynamics of the cotton whitefly *Bemisia tabaci* Genn. on tomato plants. *Ain-Shams Univ. Ann. Agric. Sci. (Egypt)* 43: 161-176.
22. Abdel-Megeed, M.I., Hegazy, G.M., Hegab, M.F. and Kamel, M.H. 1998. Non-traditional approaches for controlling the cotton whitefly, *Bemisia tabaci* Genn. infesting tomato plants. *Ain-Shams Univ. Ann. Agric. Sci. (Egypt)* 43: 177-189.
23. Abdel-Megeed, M.I., Zidan, Z.H., Dahroug, S.A., Salem, M. and Daoud, M. 1994. Factors influencing the performance of yellow sticky traps for monitoring whitefly, *Bemisia tabaci*, on cucumber in Egypt. *Ain-Shams Univ. Ann. Agric. Sci. (Egypt)* 39(2): 823-828.
24. Abdel-Salam, A.M., Assem, M.A. and Abdel-Shaheed, G.A. 1971. Experimental studies on tomato pests. I. Effect of some pesticides on tomato pests in the U.A.R. *Z. Angew. Entomol.* 69: 55-59. [Cock (1986)]
25. Abdel-Salam, A.M., Assem, M.A., Abdel-Shaheed, G.A., Hammad, S.M. and Ragab, F.Y. 1972. Chemical control of some squash pests in U.A.R. *Z. Angew. Entomol.* 70: 169-174. [Cock (1986)]
26. Abdel-Salam, A.M., Assem, M.A., Hammad, S.M. and Eid, G.H. 1972. Studies on potato pests in U.A.R. II. Susceptibility of some potato varieties to insect infestation in the field and in the storage. *Z. Angew. Entomol.* 70: 76-82. [Cock (1986)]
27. Abdel-Salam, A.M., Kaiery, E., Abbasy, A.M. and Assem, M.A. 1972. Effects of granular insecticides on some pests of horse bean and peas as well as on plant growth and root nodulation. *Z. Angew. Entomol.* 70: 408-413. [Cock (1986)]
28. Abdel-Salam, A.M. and Thomas, P.E. 1994. Comparative strategies for the purification of a whitefly-transmitted geminivirus infecting tomato fields in Washington and Oregon states. *Egyptian J. Phytopathol.* 22(1): 107-123.
29. Abdel-Salam, A.M.M., Hassan, A.A., Merghany, M.M., Abdel-Ati, K.A. and Ahmed, Y.M. 1997. The involvement of a geminivirus, a closterovirus, and a spherical virus in the interveinal mottling and yellows disease of cucurbits in Egypt. *Bull. Faculty Agric. (Univ. Cairo)* 48(4): 707-722.
30. Abdel-Shaheed, G.A., Abdel-Salam, A.M., Assem, M.A. and Amin, S.M. 1972. Control of the pests of snake-cucumber (*Cucumis melo* L. var *flexuosus* L.) and cucumber (*C. sativus* L.) in Arab Republic of Egypt. *Indian J. Agric. Sci.* 42: 95-99. [Cock (1986)]
31. Abdel-Wahhab, I.S. 1997. Strategy of using integrated pest management (IPM) programs in adjustment and arrangement of the population of major cotton sucking insect pests. Ph.D. Dissertation, Cairo University, Egypt, 230 pp.
32. Abdeldaffie, E.Y., El-Hagh, E.A. and Bashir, N.H.H. 1987. Resistance in cotton whitefly, *Bemisia tabaci* (Genn.), to insecticide recently introduced in to Sudan Gezira. [French summary]. *Trop. Pest Manage.* 33(4): 283-286. [Cock (1993)]
33. Abdelrahman, A.A. 1980. Cotton Spraying experiments. pp. 122-130. *In* Annu. Rep. Gezira Res. Stn. Substns. 1973/1974. Agric. Res. Corp., Sudan [Cock (1993)]
34. Abdelrahman, A.A. 1985. Cotton spraying experiments. pp. 112-116. *In* Annu. Rep. Gezira Res. Stn. Substns. 1977/1978. Agric. Res. Corp., Sudan [Cock (1993)]
35. Abdelrahman, A.A. 1985. Large scale cotton spraying experiment. pp. 116-118. *In* Annu. Rep. Gezira Res. Stn. Substns. 1977/1978. Agric. Res. Corp., Sudan [Cock (1993)]
36. Abdelrahman, A.A. 1985. Studies on whitefly sampling. p. 118. *In* Annu. Rep. Gezira Res. Stn. Substns. 1977/1978. Agric. Res. Corp., Sudan [Cock (1993)]
37. Abdelrahman, A.A. 1986. The potential of natural enemies of the cotton whitefly in Sudan Gezira. *Insect Sci. Appl.* 7(1): 69-73. [Cock (1986, 1993)]

38. Abdelrahman, A.A., Alsaffar, A., Munir, B. and Stam, P.A. 1998. Cotton integrated pest management in central Sudan. *Arab J. Plant Prot.* 16(1): 54-52.
39. Abdelrahman, A.A. and Munir, B. 1989. Sudanese experience in integrated pest management of cotton. [French summary]. *Insect Sci. Appl.* 10(6): 787-794. [Cock (1993)]
40. Abdelrahman, A.A. and Saleem, M.B.A. 1977. Effect of different levels of nitrogen and plant density on the population and life cycle of the cotton whitefly *Bemisia tabaci* (Genn.). pp. 119-120. *In Annu. Rep. Gezira Res. Stn. Substns. 1976/1977. Agric. Res. Corp., Sudan*
41. Abisgold, J.D. and Fishpool, L.D.C. 1990. A method for estimating population sizes of whitefly nymphs (*Bemisia tabaci* Genn.) on cassava. *Trop. Pest Manage.* 36(3): 287-292. [Cock (1993)]
42. Abo-El-Ghar, M.R., Eisa, A.A., El-Nabawi, A., Attia, M.B. and Ammar, I.M.A. 1984. Effect of certain insecticides on the population of some cotton pests and their predators. *Minufiya J. Agric. Res. (Egypt)* 9: 489-499.
43. Abou-Aiana, R. and Draz, K.A.A. 1993. The preferable hosts of cotton white fly, *Bemisia tabaci* (Genn.) on certain vegetable crops together with the influence of relative humidity and temperature on its population. *Tanta J. Agric. Res. (Egypt)* 19(2): 502-507.
44. Abou-Awad, B.A., El-Sherif, A.A., Hassan, M.F. and Abou-Eleila, M.M. 1998. Studies on development, longevity, fecundity and predation of *Amblyseius olivi* Nasr and Abou-Awad (Acari: Phytoseiidae) on various kinds of prey and diets. *Zeitschrift für pflanzenkrankheiten und Pflanzenschutz* 105(5): 538-544.
45. Abou-Awad, B.A. and Elsayi, S.A. 1993. Biology and life table of the predaceous mite, *Agistemus exsertus* Gonz. (Acari: Stigmaeidae). [German summary]. *Anz. Schadlingskunde, Pflanzenschutz, Umweltschutz* 66(5): 101-103.
46. Abou-Elhagag, G.H. 1998. Effect of spraying cotton plants during the early season against cotton aphid on cotton pests, natural enemies and some crop characters in Southern Egypt. *Assiut J. Agric. Sci. (Egypt)* 29(4): 91-100.
47. Abou-Elhagag, G.H. 1998. Seasonal abundance of certain cotton pest and their associated natural enemies in Southern Egypt. *Assiut J. Agric. Sci. (Egypt)* 29(3): 253-267.
48. Abou-Fakhr Hammad, E.M., Nemer, N.M., Hawi, Z.K. and Hanna, L.T. 2000. Responses of the sweetpotato whitefly, *Bemisia tabaci*, to the chinaberry tree (*Melia azedarach* L.) and its extracts. *Ann. Appl. Biol.* 137(2): 79-88.
49. Abou-Fakhr Hammad, E.M., Zournajian, H. and Talhouk, S. 2001. Efficacy of extracts of *Melia azedarach* L. callus, leaves and fruits against adults of the sweetpotato whitefly *Bemisia tabaci* (Hom., Aleyrodidae). *J. Appl. Entomol.* 125(8): 483-488.
50. Abou-Jawdah, Y., Maalouf, R., Shebaro, W., Soubra, K. and Abou-Jawdah, Y. 1999. Comparison of the reaction of tomato lines to infection by tomato yellow leaf curl begomovirus in Lebanon. *Plant Pathol.* 48(6): 727-734.
51. Abou-Jawdah, Y., Sobh, H., Fayad, A., Lecoq, H., Delecolle, B. and Trad, F.J. 2000. Cucurbit yellow stunting disorder virus: A new threat to cucurbits in Lebanon. *J. Plant Pathol.* 82(1): 55-60.
52. Aboul-Ata, A.E., Awad, M.A.E., Abdel-Aziz, S., Peters, D., Megahed, H. and Sabik, A. 2000. Epidemiology of tomato yellow leaf curl begomovirus in the Fayium area, Egypt. *Bull. OEPP* 30(2): 297-300.
53. Abouzid, A. and Jeske, H. 1986. The purification and characterization of gemini particles from Abutilon mosaic virus infected Malvaceae. *J. Phytopathol.* 115: 344-353. [Cock (1986)]
54. Abouzid, A.M., Frischmuth, T. and Jeske, H.A. 1988. A putative replicative form of the Abutilon mosaic virus (gemini group) in a chromatin-like structure. *Mol. Gen. Genet.* 212: 252-258.
55. Abouzid, A.M., Polston, J.E. and Hiebert, E. 1992. The nucleotide sequence of tomato mottle virus a new geminivirus isolated from tomatoes in Florida. *J. Gen. Virol.* 73: 3225-3229.
56. Abreu, R.A., Peña, C.E. and Gálvez, J.E. 1979. Control del virus de mosaico dorado del frijol (*Phaseolus vulgaris* L.) por resistencia varietal y por control químico del insecto vector *Bemisia tabaci* Genn. [Control of the golden mosaic virus of beans (*Phaseolus vulgaris* L.) with varietal resistance and chemical control of the insect vector *Bemisia tabaci* Genn]. *Reunion Anual del PCCMCA* 25(3): L14.1-L14.3. [CATIE (1992), Spanish]
57. Abtali, Y. and Espahbody, A. 1991. A preliminary study of Cotton white fly (*Bemisia tabaci* Gennadius) in Mazandaran. p. 4. *In Proceedings 10th Plant Protection Congress Iran, 1-5-Sep. 1991, Kerman, Iran.* [Persian, English summary]
58. Abu-Gharbieh, W.I., Makkouk, K.M. and Saghir, A.R. 1978. Response of different tomato cultivars to the root-knot nematode, tomato yellow leaf curl virus and orobanche in Jordan. *Plant Dis. Rep.* 62: 263-266.
59. Abu-Yaman, I.K. 1971. Outbreaks and new records. *FAO Plant Prot. Bull.* 19(6): 140-141. [Cock (1986)]
60. Abul-Nasr, S. and El-Nahal, A.K.M. 1969. Seasonal population of Hemiptera-Homoptera infesting cotton plants in Egypt. *Bull. Entomol. Soc. Egypt* 52(1968): 371-389. [Cock (1986)]
61. Accotto, G.P., Donson, J. and Mullineaux, P.M. 1989. Mapping of digitaria streak virus transcripts reveals different RNA species from the same transcription unit. *EMBO J.* 8(4): 1033-1039.
62. Adam, K.M. 1997. Relative susceptibility of eight tomato cultivars to infestations with *Bemisia tabaci* (Genn.) with special reference to the percentage of virus infection and total yield. *Moshtohor Ann. Agric. Sci. (Egypt)* 35(2): 1013-1019.
63. Adam, K.M., Bachatly, M.A. and Doss, S.A. 1997. Populations of the whitefly *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae) and its parasitoid *Eretmocerus mundus* Mercet (Hymenoptera: Aphelinidae) in protected cucumber cultivations. *Egyptian J. Agric. Res.* 75(4): 939-950.
64. Adam, K.M., El-Hamaky, M.A. and Doss, S.A. 1992. A comparative study of the effect of host plants on *Bemisia tabaci*, Genn, Aleyrodidae: Hemiptera. *Minia J. Agric. Res. Devel. (Egypt)* 14(3): 799-808.
65. Adames-Mora, C., Beaver, J.S. and Diaz, O. 1996. Una metodología para evaluar en el invernadero el virus del mosaico dorado de la habichuela. [A greenhouse screening technique for bean golden mosaic virus]. *J. Agric. Univ. Puerto Rico* 80(1-2): 65-72. [Spanish, English summary]
66. Adashkevich, B.P. and Kadyrov, A.K. 1990. Biological means of whitefly control. *Zashchita Rastenii (Moskva)* 11: 37-38. [Russian]
67. Adashkevich, B.P. and Saidova, Z.K.H. 1990. *Encarsia* Conservation. *Zashchita Rastenii (Moskva)* 12: 15-16. [Russian]
68. Adejag, G. and Coutts, R. 1982. Ultrastructural studies on *Nicotiana benthamiana* tissue following infection with a virus transmitted from mosaic-diseased Nigerian cassava. *Phytopathol. Z.* 103(1): 87-92.
69. Adipala, E., Byabakama, B.A., Ogenga-Latigo, M.W. and Otim-Nape, G.W. 1998. Effect of planting date and varietal resistance on the development of cassava mosaic virus disease in Uganda. *African Plant Prot.* 4(2): 71-79.
70. Aerts, D., Coremans-Pelseneer, J., van de Veire, M., Sterk, G. and Degheele, D. 1997. Side-effects of pesticides on the development of the entomopathogenic fungus, *Paecilomyces fumosoroseus* (Wize) Brown and Smith, strain Apopka 97. *Med. Fac. Landbouww. Univ. Gent* 62(2b): 581-587.
71. Afifi, F.M.L., Haydar, M.F. and Omar, H.I.H. 1990. Effect of different intercropping systems on tomato infestation with major insect pests; *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae), *Myzus persicae* Sulzer (Homoptera: Aphididae) and *Phthorimaea operculella* Zeller (Lepidoptera: Gelechiidae). [Arabic summary]. *Bull. Faculty Agric. (Univ. Cairo)* 41(3): 885-900. [Cock (1993)]
72. Afzal, M., Farooq, M., Razaq, M. and Ayub, S. 2000. Determination of effective and economic dose of two carbamate insecticides against cotton whitefly *Bemisia tabaci* (Genn.) on Niab karisham cotton at Faisalabad (Pakistan). *Pakistan J. Biol. Sci.* 3(8): 1283-1284.

73. Afzal, M. and Khan, M.R. 1978. Life history and feeding behaviour of green lacewing, *Chrysopa carnea* Stephens (Neuroptera, Chrysopidae). Pakistan J. Zool. 10(1): 83-90. [Cock (1986)]
74. Agas, C.N. 1991. Survey of insect pests attacking chrysanthemum and anthurium. Philippine J. Plant Industry 44/56(1/3): 32-55.
75. Agnew, G.K., Frisvold, G.B. and Baker, P. 2000. Adoption of insect growth regulators in Arizona cotton: determinants and economic implications. pp. 361-364. In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
76. Agnew, G.K., Frisvold, G.B. and Baker, P. 2000. Use of insect growth regulators and changing whitefly control costs in Arizona cotton. pp. 307-314. In Cotton. A College of Agriculture Report, Series P-121. University of Arizona, Tucson.
77. Agrawal, H.S., Gupta, N.K., Prasad, V.K. and Vishwakarma, S. 1979. Chemical control of yellow mosaic of moong. Pesticides 13(5): 44-47. [Cock (1986)]
78. Aharonson, N. 1982. Soil-applied systemic insecticides for the control of *Bemisia tabaci*. Phytoparasitica 10: 297-298.
79. Aharonson, N., Magal, Z., Muszkat, L., Tepperman, D., Goren, E. and Tadmor, U. 1986. Application of aldicarb in a drip-irrigated cotton field for the control of the tobacco whitefly (*Bemisia tabaci*). Phytoparasitica 14(1): 87-91. [Cock (1993)]
80. Aharonson, N., Muszkat, L. and Neubauer, I. 1984. Accumulation pattern and insecticidal effect of aldicarb in cotton following soil treatment for the control of the tobacco whitefly (*Bemisia tabaci*). Phytoparasitica 12: 127-134. [Cock (1986)]
81. Aheer, G.M., Hussain, A.M., Iqbal, Z., Ali, A. and Ramzan, M. 1997. Role of detergents in increasing the efficacy of insecticides against whitefly (*Bemisia tabaci* Genn.), the vector of cotton leaf curl virus. J. Agric. Res. (Pakistan) 35(4): 253-256.
82. Ahlawat, Y.S. and Srivastava, K.P. 1998. *Ocimum sanctum*: A natural host of a geminivirus and its vector, *Bemisia tabaci* during off-season. Indian J. Entomol. 60(3): 213-215.
83. Ahmad, F., Khan, F.R. and Khan, M.R. 1995. Comparative efficacy of some traditional and non-traditional insecticides against sucking insect pests of cotton. Sarhad J. Agric. (Pakistan) 11(6): 733-739.
84. Ahmad, F. and Mohsin, M.D. 1969. Control of cotton bollworm *Heliothis armigera* (Hb.) by air in Multan District of West Pakistan. Int. Pest Control 11(6): 14-15. [Cock (1986)]
85. Ahmad, F.U. and Shafi, M.M. 1966. Concentrates against cotton pests. World Crops 18(4): 60-62. [Cock (1986)]
86. Ahmad, M. 1978. Whitefly (*Bemisia tabaci*) transmission of a yellow mosaic disease of cowpea (*Vigna unguiculata*). Plant Dis. Rep. 62(3): 224-226. [Cock (1986)]
87. Ahmad, M. 1978. A whitefly -vectored yellow mosaic of jute. FAO Plant Prot. Bull. 26(4): 169-171. [Cock (1986)]
88. Ahmad, M. 1979. Studies with a whitefly -transmitted yellow vein mosaic of *Digera alternifolius*. Phytopathol. Z. 96: 21-24. [Cock (1986)]
89. Ahmad, M., Arif, M.I. and Ahmad, Z. 1999. Insecticide resistance in *Helicoverpa armigera* and *Bemisia tabaci*, its mechanisms and management in Pakistan. pp. 143-150. In Proceedings ICAC-CCRI Regional Consultation, Insecticide Resistance Management in Cotton. Central Cotton Research Institute, Multan, Pakistan.
90. Ahmad, M., Arif, M.I. and Ahmad, Z. 2000. Resistance of cotton whitefly, *Bemisia tabaci* to cypermethrin, alphacypermethrin and zetacypermethrin in Pakistan. pp. 1015-1017. In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
91. Ahmad, M., Arif, M.I. and Ahmad, Z. 2001. Reversion of susceptibility to methamidophos in the Pakistani populations of cotton whitefly, *Bemisia tabaci*. pp. 874-876. In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
92. Ahmad, M., Arif, M.I., Ahmad, Z. and Denholm, I. 2002. Cotton whitefly (*Bemisia tabaci*) resistance to organophosphate and pyrethroid insecticides in Pakistan. Pest Manage. Sci. 58(2): 203-208.
93. Ahmad, M. and Harwood, R.F. 1973. Studies on a whitefly -transmitted yellow mosaic of urd bean (*Phaseolus mungo*). Plant Dis. Rep. 57: 800-802. [Cock (1986)]
94. Ahmad, M., Khan, M.R., Akram, M. and Yousuf, R.M. 1985. Studies on varietal resistance in cotton to insect pests. Pakistan Entomol. 7(1/2): 65-70.
95. Ahmad, N. and Manzoor, H. 1981. Some studies on resistance in cotton against jassid, *Amrasca devastans* (Distant) and whitefly, *Bemisia tabaci* (Genn.). Pakistan Entomol. 3(1/2): 29-34.
96. Ahmad, R. and Varma, S. 1984. Residues of dimethoate and phosphamidon in brinjal fruits and determination of waiting period. Indian J. Entomol. 46(4): 398-401.
97. Ahmad, R.D., Naqvi, S.M.S.H., Hussain, T., Rustamani, M.A. and Khan, M.M. 1994. Relative toxicity of different insecticides against whitefly, *Bemisia tabaci* Genn., on cotton. Proc. Pakistan Congress Entomol. 14: 383-386.
98. Ahmad, S.A.H. 1999. Entomological studies on phaseolus, Mung bean, *Vigna radiata*. Ph.D. Dissertation, Zagazig University, Moshthor, Egypt, 203 pp.
99. Ahmad, Z. 1999. Pest problems of cotton - a regional perspective. pp. 5-20. In Proceedings ICAC-CCRI Regional Consultation, Insecticide Resistance Management in Cotton. Central Cotton Research Institute, Multan, Pakistan.
100. Ahmed, A.H.M., El-Hagh, E.A. and Bashir, N.H.H. 1987. Insecticide resistance in cotton whitefly (*Bemisia tabaci* Genn.) in the Sudan Gezira. [French summary]. Trop. Pest Manage. 33(1): 67-72, 103, 107. [Cock (1993)]
101. Ahmed, K., Khalique, F., Afzal, M. and Malik, B. 1986. Studies on population of white flies, *Bemisia tabaci* infesting mashbean. Food Legume Improvement Programme. Pulses Entomology Report June 1985 - May 1986. Pakistan Agricultural Research Council, Islamabad, 53 pp.
102. Ahmed, K., Khalique, F. and Bashir, M. 1987. Insect pests of pulse crops and their control. pp. 240-252. In C. Inayatullah (Ed.), Integrated Pest Management, Islamabad, Pakistan, 22 Nov-3 Dec 1987. Pakistan Agricultural Research Council, Islamabad.
103. Ahmed, M.A. 1994. Differences in susceptibility of six cucumber cultivars to infestation by *Aphis gossypii* Glov., *Tetranychus urticae* and *Bemisia tabaci* as correlated to protein and amino acid contents of leaves. Moshthor Ann. Agric. Sci. (Egypt) 32(4): 2189-2194.
104. Ahmed, N.E., Kanan, H.O., Sugimoto, Y., Ma, Y.Q. and Inanaga, S. 2001. Effect of imidacloprid on incidence of Tomato yellow leaf curl virus. Plant Dis. 85(1): 84-87.
105. Ahmed, R. and Baig, M.M.H. 1987. Observations on efficacy and economics of cotton pest control with deltamethrin alone and its combinations with monocrotophos and DDT. Pakistan J. Sci. Indus. Res. 30(7): 517-519. [Cock (1993)]
106. Ahmed, R. and Muzaffar, N. 1977. Studies on cotton pests and their natural enemies with reference to effects of three insecticides of different persistencies. Agric. Pakistan 28: 193-203. [Cock (1986)]
107. Ahohuendo, B.C. 1993. Das afrikanische Maniok-Mosaik-Virus (ACMV): Untersuchungen zur Verbreitung der Virose im Mischbaupflanzensystem in der Republik Benin und ueber die Eigenschaften des Virus. [The African maniok-mosaic-virus (ACMV): investigations on occurrence of the virosis in mixed cropping system in the republic of Benin and on properties of the virus]. Ph.D. Dissertation, Hohenheim University, Stuttgart, Germany, 104 pp.
108. Ahohuendo, B.C. and Sarkar, S. 1995. Partial control of the spread of African cassava mosaic virus in Benin by intercropping. Zeitschrift für pflanzenkrankheiten und Pflanzenschutz 102(3): 249-256.

109. Akey, D.H., Chu, C.C. and Henneberry, T.J. 1992. Application equipment and under-leaf coverage of cotton with cotton seed oils, soap, and fenpropathrin/acephate against the sweetpotato whitefly, *Bemisia tabaci*. pp. 701-702. In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
110. Akey, D.H. and Henneberry, T.J. 1994. Sweetpotato whitefly control by Naturalis-L, the fungus *Beauveria bassiana*, in furrow and sub-drip irrigated upland cotton. pp. 1089-1091. In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
111. Akey, D.H. and Henneberry, T.J. 1998. Control of silverleaf whitefly with the entomopathogenic fungi, *Paecilomyces fumosoroseus* and *Beauveria bassiana* in upland cotton in Arizona. pp. 1073-1077. In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
112. Akey, D.H. and Henneberry, T.J. 1999. Control of the silverleaf whitefly with the neem product azadirachtin as BOLLWHIP in upland cotton in Arizona. pp. 914-918. In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
113. Akey, D.H., Henneberry, T.J. and Chu, C.C. 1993. Control studies on field populations of the sweetpotato whitefly, *Bemisia tabaci* in Arizona upland and Pima cotton. pp. 675-679. In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
114. Akey, D.H., Henneberry, T.J., Dennehy, T.J. and Ellsworth, P.C. 1996. Large scale whitefly management and trials using insecticide rotations to develop IPM strategies for Arizona upland cotton. pp. 817-818. In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
115. Akey, D.H., Henneberry, T.J., Williams, L.H., III, Dennehy, T.J. and Ellsworth, P.C. 1997. Strategies for insecticide resistance management of silverleaf whitefly populations in two upland cottons: ground applications of insect growth regulators and pyrethroids. pp. 918-921. In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
116. Akey, D.H., Henneberry, T.J. and Wuertz, D.A. 1994. Whole season rotational pesticide system for integrated pest management for control of sweetpotato whitefly in cotton. pp. 344-345. In Cotton, A College of Agriculture Report, Series P-96. University of Arizona, Tucson.
117. Al-Abdulmohsin, A.M. 1997. Saudi Arabia. pp. 55-56. In N. Ioannou (Ed.), Management of the whitefly-virus complex. FAO Plant Production and Protection Paper 143, Rome, Italy.
118. Al-Ani, R.A., Samir, S.H. and Jarjees, M.M. 1987. [Identification and control of tobacco leaf curl virus]. Arab J. Plant Prot. 5(2): 70-73. [Cock (1993), Arabic, English summary]
119. Al-Bagham, S.H. and Salim, A. 1997. United Arab Emirates. pp. 81-83. In N. Ioannou (Ed.), Management of the whitefly-virus complex. FAO Plant Production and Protection Paper 143, Rome, Italy.
120. Al-Hitty, A., Abbass, A.K. and Ali, A.S.A. 1988. Development and survival of *Deraecoris pallens* Reut. (Heteroptera, Miridae) on *Bemisia tabaci* Gen. and *Aphis gossypii* Glv. [Arabic summary]. J. Agric. Water Resources Res. (Iraq) 7(2): 115-123. [Cock (1993)]
121. Al-Hitty, A. and Sharif, H.L. 1987. [Studies on host plant preference of *Bemisia tabaci* (Genn.) on some crops and effect of using host on the trap spread of tomato yellow leaf curl virus to tomato in the plastic house]. Arab J. Plant Prot. 5(1): 19-23. [Cock (1993), Arabic]
122. Al-Mansoor, H. and Ahmadi, A.A. 1993. Natural enemies of cotton whitefly, *Bemisia tabaci* (Gennadius) in Fars province. p. 106. In Proceedings 11th Plant Protection Congress of Iran, 28-Aug - 2 Sep. 1993, Rasht, Iran. [Persian]
123. Al-Mansoor, H. and Ahmadi, A.A. 1995. Host- parasitoid interaction between *Eretmocerus mundus* Mercet (Hym.: Aphelinidae), a parasitoid of cotton whitefly nymphs *Bemisia tabaci* (Genn.), in the Fars province. p. 101. In Proceedings 12th Iranian Plant Protection Congress, 2-7 September 1995, Karadj, Iran. [Persian]
124. Al-Musa, A. 1982. Incidence, economic importance, and control of tomato yellow leaf curl in Jordan. Plant Dis. 66: 561-563.
125. Al-Musa, A.M. 1986. Tomato yellow leaf curl virus in Jordan: Epidemiology and control. Dirasat 8: 199-208.
126. Al-Musa, A.M., Nazer, I.K. and Sharaf, N.S. 1983. Muslin and plastic tunnels; effect on incidence of tomato yellow leaf curl early blight and various growth characteristics of tomatoes. Dept. Plant Prot., Jordan Univ., Amman, 21 pp.
127. Al-Musa, A.M., Nazer, I.K. and Sharaf, N.S. 1987. Effect of certain combined agricultural treatments on whitefly population and incidence of tomato yellow leaf curl virus. Dirasat 14(11): 127-134.
128. Al-Musa, A.M., Nazer, I.K., Sharaf, N.S. and Mansour, A.N. 1985. Muslin and plastic tunnels; effect on incidence of tomato yellow leaf curl, early blight and various growth characteristics of tomatoes. [Arabic summary]. Dirasat 12(6): 101-109. [Cock (1993)]
129. Al-Musa, A.M. and Sharaf, N.S. 1983. Role of cucumbers in delaying the onset of tomato yellow leaf curl in Jordan. Dept. Plant Prot., Jordan Univ., Amman, 13 pp.
130. Al-Samariee, A.I., Al-Majeed, K.A. and Al-Bassomy, M. 1987. Pirimphos-methyl residues on the cucumber cultivated in commercial greenhouses. [Arabic summary]. J. Biol. Sci. Res. 18(2): 89-99. [Cock (1993)]
131. Al-Sayed, M.E.M. and Othmani, A. 1997. Syria. pp. 67-70. In N. Ioannou (Ed.), Management of the whitefly-virus complex. FAO Plant Production and Protection Paper 143, Rome, Italy.
132. Al-Shahwan, I.M., Abdalla, O.A. and Al-Saleh, M.A. 2001. Detection of Tomato yellow leaf curl virus (TYLCV) in Saudi Arabia. J. Plant Dis. Prot. 108(4): 407-412.
133. Al-Shayji, Y., Shaheen, N., Saleem, M. and Ibrahim, M. 1998. The efficacy of some *Bacillus thuringiensis* formulations against the whitefly *Bemisia tabaci* (Homoptera: Aleyrodidae). Kuwait J. Sci. Eng. 25(1): 223-229.
134. Al-Zidjali, T.S. 1997. Sultanate of Oman. pp. 45-54. In N. Ioannou (Ed.), Management of the whitefly-virus complex. FAO Plant Production and Protection Paper 143, Rome, Italy.
135. Alam, M.S. 1989. Whitefly (Hemiptera: Aleyrodidae) -a potential pest of rice in West Africa. International Rice Res. Newsletter (Philippines) 14(3): 38-39.
136. Alam, M.Z., Ali, M., Akanda, A.M., Choudhury, D.A.M., Haque, N.M.M., Hossain, M.M. and Ogata, K. 1994. Grafting technology: an integrated pest management component for eggplant and tomato. Bull. Inst. Trop. Agric. (Kyushu Univ.) 17: 85-91. [English, Japanese summary]
137. Alam, M.Z., Hossain, M.M., Choudhury, D.A.M., Uddin, M.J. and Haque, N.M.M. 1995. Effect of grafting technology on suppression of whitefly (*Bemisia tabaci* Gennadius) disseminating virus diseases in tomato. Ann. Bangladesh Agric. 5(2): 91-98.
138. Alao C. R. and Misari, S.M. 1992. Some studies of identity and relationships of leafcurl diseases on some solanaceous crop species in Northern Nigeria. Samaru J. Agric. Res. 9: 35-48.
139. Alaux, J.P. and Fauquet, C. (Eds.). 1990. African Cassava Mosaic Disease: from Knowledge to Control. p. 263pp. In Summary Report of the International Seminar The African Cassava Mosaic Disease and Its Control, Yamoussoukro, Ivory Coast, 4-8 May 1987. UNIVERSA, Wetteren, Belgium, [Cock (1993)]
140. Albajes, R. and Alomar, O. 1999. Current and potential use of polyphagous predators. pp. 265-275. In R. Albajes, M.L. Gullino, J.C. van Lenteren and Y. Elad (Eds.), Integrated Pest Management and Disease Management in Greenhouse Crops. Kluwer, Dordrecht, The Netherlands.

141. Albajes, R., Alomar, O., Riudavets, J., Castane, C., Arno, J., Gabarra, R. and van Lenteren, J.C. 1996. The mirid bug *Dicyphus tamaninii*: an effective predator for vegetable crops. Bull. OILB-SROP (IOBC/WPRS) 19(1): 1-4.
142. Albert, R. 1990. Experiences with biological control measures in glasshouses in southwest Germany. Bull. OILB/SROP (IOBC/WPRS) 13: 1-5. [ DeBarro (1995)]
143. Albert, R. 1990. [White flies in vegetables and ornamental cultures in greenhouses]. Gärtnerbörse und Gartenwelt (Germany) 90(4): 677-681. [ German]
144. Albert, R. and Sautter, H. 1989. [Parasitoids protect christmas stars from whiteflies]. Deutscher Gartenbau 43(27): 1671-1673. [ Cock (1993), German]
145. Albert, R., Sautter, H. and Schneller, H. 1990. [Biological control in poinsettias. A good beginning found for the use of beneficials in ornamental plant crops]. Gärtnerbörse und Gartenwelt (Germany) 90(15): 734-736. [ Cock (1993), German]
146. Albert, R. and Schneller, H. 1989. [Biological control in ornamental plants. I. Poinsettias (*Euphorbia pulcherrima* Willd. ex. Klotzsch)]. Med. Fac. Landbouww. Univ. Gent 54(3a): 873-882. [ Cock (1993), German, English summary]
147. Albert, R. and Schneller, H. 1989. [Successful biological control in ornamental plants - 1. Poinsettias (*Euphorbia pulcherrima* Willd. ex Klotzsch)]. Gesunde Pflanzen 41(11): 389-395. [ Cock (1993), German, English summary]
148. Albert, R. and Schneller, H. 1991. [Biological pest control on ornamental plants. I. Experiences with *Trialeurodes vaporariorum* and *Bemisia tabaci*]. Gärtnerbörse und Gartenwelt (Germany) 91(1): 10-15. [ Cock (1993), German]
149. Albert, R. and Schneller, H. 1992. [Plant protection from cuttings to mother plants in poinsettia]. Gärtnerbörse und Gartenwelt (Germany) 92(6): 301-305. [ German]
150. Albert, R. and Schneller, H. 1994. *Eretmocerus californicus*, ein weiterer Gegenspieler der Weissen Fliege. [*Eretmocerus californicus*, a further antagonist of white flies]. Taspo Gartenbaumagazin (Germany) 3(5): 44-45. [ German]
151. Albert, R., Schneller, H. and Sautter, H. 1993. Development of biological control in ornamentals. Bull. OILB/SROP (IOBC/WPRS) 16(8): 1-5.
152. Aldana de Leon, L.F., Masaya-Sanchez, P. and Yoshii, K. 1982. La tolerancia al mosaico dorado del frijol [*Phaseolus vulgaris*] comun y el combate quimico del vector, *Bemisia tabaci*, Genn como medio de control quimico. [Tolerance of common kidney bean [*Phaseolus vulgaris*] to golden mosaic virus and chemical control of the vector, *Bemisia tabaci*, Genn]. Ciencia Tecnologia Agropecuaria (Guatemala) 1(1): 46-66. [ Spanish, English summary]
153. Alegbejo, M.D. 2000. Whitefly transmitted plant viruses in Nigeria. J. Sustain. Agric. 17(2/3): 99-109.
154. Alegbejo, M.D. and Kashina, B.D. 2001. Status of legume viruses in Nigeria. J. Sustain. Agric. 18(1): 55-69.
155. Ali, A. and Ahmad, M. 1982. Biophysical resistance in different varieties of cotton against insect pests. Pakistan Entomol. 4(1/2): 27-32.
156. Ali, A.A. and El-Said, N.M. 1987. [Influence of seedling production methods on tomato transplant infestation with tobacco white fly and subsequent infection with tomato yellow leaf curl virus in plastic houses]. Arab J. Plant Prot. 5(1): 24-30. [ Cock (1993), Arabic, English summary]
157. Ali, A.A., Rajeb, A.S. and Al-Hussaini, H.H. 1986. Integrated means for the control of the tobacco whitefly *Bemisia tabaci* Gen and tomato yellow leaf virus in plastic houses. J. Agric. Water Resources Res. (Iraq) 5(2): 151-165. [ Arabic, English summary]
158. Ali, A.A., Razoki, I.A. and Hani, N.A. 1987. Influence of planting dates and plant age on the susceptibility of some cucurbit varieties to different pests during fall season. J. Agric. Water Resources Res. (Iraq) 6(2): 93-108.
159. Ali, A.A., Razoki, I.A.R. and Rajeb, A.S. 1986. Influence of planting date and crop variety on infestation with some spring cucurbit pests. J. Agric. Water Resources Res. (Iraq) 5(2): 37-52.
160. Ali, A.G. 1996. Survey of arthropods associated with sesame plants in Assiut Governorate, Upper Egypt. Assiut J. Agric. Sci. (Egypt) 27(2): 135-145.
161. Ali, A.G., Farghali, M.A. and Hussein, H.A. 1996. Susceptibility of some mungbean cultivars to whitefly (*Bemisia tabaci* (Genn.)) and mites (*Tetranychus urticae* Koch) with reference to pod setting and yield. Assiut J. Agric. Sci. (Egypt) 27(2): 147-156.
162. Ali, A.S.A. and Hani, N.A. 1993. Locally designed yellow sticky traps on the sweetpotato whitefly *Bemisia tabaci* and the incidence of the tomato yellow leaf curl virus in plastic houses. J. Agric. Res. (Iraq) 3(2): 217-228. [ Arabic, English summary]
163. Ali, M., Ahmad, Z., Hussain, T. and Tanveer, M. 1992. Cotton leaf curl virus situation in the Punjab 1991-92. Pakistan Central Cotton Committee, 33 pp.
164. Ali, M., Ahmad, Z., Tanveer, M. and Mahmood, T. 1995. Cotton leaf curl virus in the Punjab: Current situation and review of work 1994-1995. Central Cotton Research Institute, Multan. 117 pp.
165. Ali, M., Hossain, M.Z. and Sarker, N.C. 2000. Inheritance of yellow vein mosaic virus (YVMV) tolerance in a cultivar of okra (*Abelmoschus esculentus* (L.) Moench). Euphytica 111(3): 205-209.
166. Ali, M.I. and Ali, M.R. 1993. Okra leaf cotton: a potential source of resistance against the cotton jassid, *Amrasca devastans* (Dist.) in Bangladesh. Bangladesh J. Zool. 21(2): 45-49.
167. Ali, N.A. 1988. Entomology. Biochemistry. pp. 125-131. In Annu. Rep. Gezira Res. Stn. Substns. 1980/1981. Agric. Res. Corp., Sudan [Cock (1993)]
168. Ali, N.A. and Khalifa, H. 1980. Development of methods to measure cotton stickiness. Coton Fibres Trop. 35: 411-413.
169. Alicai, T., Adipala, E. and Gibson, R.W. 1999. Seasonal changes in whitefly numbers and their influence on incidence of sweetpotato chlorotic stunt virus and sweetpotato virus disease in sweetpotato in Uganda. Int. J. Pest Manage. 45(1): 51-55.
170. Alimukhamedov, S.N. 1991. The system of crop protection against whiteflies. Zashchita Rastenii (Moskva) 11: 52-53. [ Russian]
171. Alkaddoussi, A.R. and Abdel-Raheem, A.A. 1991. Gene action and genetics of resistance to aphid and white fly in *Gossypium* spp. Zagazig J. Agric. Res. (Egypt) 18(2): 349-356.
172. Allawi, T.R. 1994. Whitefly species in Jordan. Arab J. Plant Prot. 12(1): 30-32. [ Arabic, English Summary]
173. Allemann, D.V., Robinson, S.W. and Flückiger, C.R. 1995. Pymetrozine (CGA 215,944) - a novel control for sucking pests in cotton. pp. 519-521. In G.A. Constable and N.W. Forrester (Eds.), Challenging the Future: Proceedings of the World Cotton Research Conference-1. CSIRO, Melbourne, Australia.
174. Allen, J.C., Brewster, C.C., Paris, J.F., Riley, D.G. and Summers, C.G. 1996. Spatiotemporal modeling of whitefly dynamics in a regional cropping system using satellite data. pp. 111-124. In D. Gerling and R.T. Mayer (Eds.), *Bemisia*: 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
175. Allen, R.M., Tucker, H. and Nelson, R.A. 1960. Leaf crumple disease of cotton in Arizona. Plant Dis. Rep. 44: 246-250.
176. Alex, D., Laterrot, H., Kheyr-Pour, A., Marchoux, G. and Hostachy, B. 1994. La maladie des feuilles jaunes en cuillère de la tomate: une virose a venir. [Tomato Yellow Leaf Curl: a future virose for France]. PHM Rev. Hortic. (France) 350: 13-17. [ French]

177. Alonzo-Padilla, F. 1976. Uso de insecticidas granulados en frijol para el combate de *Empoasca* sp. y *Bemisia tabaci* (Genn), en el sur-oriente de Guatemala. [The use of granulated insecticides in bean for the control of *Empoasca* sp. and *Bemisia tabaci* (Genn) in Southeastern Guatemala.]. pp. 1-10. In Memoir. 22. Annual Meeting Central American Cooperative Program for the Improvement of Food Crops. Ministerio De Agricultura y Ganaderia, San Jose, Costa Rica; IICA, San Jose, Costa Rica; Rockefeller Foundation, New York, NY. [Spanish, English summary]
178. Alpizar, D. 1993. Aspectos basicos sobre las moscas blancas con enfasis en *Bemisia tabaci* y *Trialeurodes vaporariorum*. [Basic aspects of whiteflies with emphasis on *Bemisia tabaci* and *Trialeurodes vaporariorum*]. Boletin Divulgativo (Costa Rica) 112, 22 pp. [ Spanish]
179. Alvarez, P., Alfonseca, L., Abud, A., Villar, A., Rowland, R., Marcano, E., Borbón, J. and Garrido, L. 1992. Las moscas blancas en la Republica Dominicana. [The whiteflies in the Dominican Republic]. pp. 34-37. In L. Hilje and O. Arboleda (Eds.), Las moscas blancas (Homoptera:Aleyrodidae) en America Central y el Caribe. CATIE, Turrialba, Costa Rica. [Spanish]
180. Alves, S.B., Silveira, C.A., Lopes, R.B., Tamai, M.A., Ramos, E.Q. and De Salvo, S. 2001. Eficácia de *Beauveria bassiana*, imidacloprid e thiacloprid no controle de *Bemisia tabaci* e na incidência do BGMV. [Efficacy of *Beauveria bassiana*, imidacloprid and thiacloprid for the control of *Bemisia tabaci* and the incidence of BGMV]. Manejo Integrado de Plagas (Costa Rica) 61: 31-36. [ Spanish, English summary]
181. Aly, F.A., El-Adl, F.E., El-Sayed, A.M., Ibrahim, S.M., Nour El-Din, M.M., Samy, M.M., Taha, A.M. and Hamaky, M.A. 1993. Geographical distribution of the infestation levels of the cotton whitefly, *Bemisia tabaci* (Genn.) (Aleyrodidae: Homoptera) on cotton plants in Egypt. Mansoura Univ. J. Agric. Sci. (Egypt) 18(6): 1878-1887.
182. Aly, F.K. 1990. Effect of some agricultural practices on the melon aphid and the whitefly infesting three cucurbit vegetables. Minia J. Agric. Res. Devel. (Egypt) 12(2): 675-684.
183. Aly, F.K., Gharib, A.H. and Hussein, S.M. 1987. Efficacy of certain insect growth regulators (IGR's), insecticide-IGR mixtures and *Bacillus thuringiensis* on some sucking insects infesting cotton plants in Minia region. Minia J. Agric. Res. Devel. (Egypt) 9(1): 339-353.
184. Amador, R. and Hilje, L. 1993. Efecto de coberturas vivas e inertes sobre la atraccion de la mosca blanca, *Bemisia tabaci* (Gennadius), al tomate. [Effect of live and inert covers on the attraction of the whitefly, *Bemisia tabaci* (Gennadius) to tomato]. Manejo Integrado de Plagas (Costa Rica) 29: 14-21. [ Spanish, English summary]
185. Anciso, J.R. and Kern, J.L. 1992. Emergence of a new plant pest, *Bemisia tabaci* (Gennadius), in the Lower Rio Grande Valley, Texas. Subtrop. Plant Sci. 45: 54-57.
186. Anderson, M., Edmunds, P., Mellor, H.E. and Walbank, M.H. 1992. The role of the olfactory system of three crop pests: aphid, whitefly and thrips, in the detection of semiochemicals. pp. 1205-1210. In Brighton Crop Protection Conference: Pests and Diseases. British Crop Protection Council, Farnham, UK.
187. Anderson, P. 1994. La mosca blanca *Bemisia tabaci* (Gennadius) como vector del virus del mosaico dorado del frijol (BGMV). [*Bemisia tabaci*, the vector of bean golden mosaic virus]. pp. 125-143. In F. Morales (Ed.), Bean Golden Mosaic: Research Advances. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. [Spanish, English summary]
188. Anderson, P.K. 1992. Un modelo para la investigacion en mosca blanca, *Bemisia tabaci* (Gennadius). [A model for the investigation of the whitefly *Bemisia tabaci* (Gennadius)]. pp. 27-33. In L. Hilje and O. Arboleda (Eds.), Las moscas blancas (Homoptera: Aleyrodidae) en America Central y el Caribe. CATIE, Turrialba, Costa Rica. [Spanish]
189. Anderson, P.K. 2000. La mosca blanca vectora: *Bemisia tabaci* (Genn.). pp. 107-128. In Bean Golden Mosaic and Other Diseases of Common Bean Caused by Whitefly -Transmitted Geminiviruses in Latin America. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. [Spanish]
190. Andreeva, L., Jarvekulg, L., Rabenstein, F., Torrance, L., Harrison, B.D. and Saarma, M. 1994. Antigenic analysis of potato virus A particles and coat protein. Ann. Appl. Biol. 125(2): 337-348.
191. Andrews, F.W. 1936. The effect of leaf curl disease on the yield of cotton plant. Cotton Growing Rev. 13(4): 287-293. [ Ali et al. (1992)]
192. Angnen, M.C., Colting, L.M. and Tadawan, B.S. 1984. Insecticidal efficacy of microbial pathogens associated with whitefly, *Bemisia* sp. MSCA Res. J. (Philippines) 16: 1-8.
193. Anno-Nyako, F.O. 1986. Semipersistent transmission of an 'extra mild' isolate of cowpea mild mottle virus on soya bean by the whitefly *Bemisia tabaci* Genn. in Nigeria. [Spanish summary]. Trop. Agric. 63: 193-194. [ Cock (1993)]
194. Anno-Nyako, F.O., Vetten, H.J., Allen, D.J. and Thottappilly, G. 1983. The relation between cowpea golden mosaic and its vector, *Bemisia tabaci*. Ann. Appl. Biol. 102: 319-323. [ Cock (1986)]
195. Anon. 1963. Pests & diseases number. Plant Prot. Bull. (New Delhi) 11(1-4; 1959): 1-70. [ Cock (1986)]
196. Anon. 1982. New record of six species of whiteflies in Iraq with special reference to the population density of the cotton whitefly, *Bemisia tabaci* (Genn) on certain economic crops. Mesopotamia J. Agric. 17(1): 93-104.
197. Anon. 1986. Abstracts of papers presented at the 5th meeting on whiteflies in field crops and vegetables. Feb. 17, 1986. Phytoparasitica 14(2): 149-154. [ Cock (1993)]
198. Anon. 1986. Entomology. pp. 33-36. In Cyprus Agric. Res. Inst. Ann. Rpt.. 1985. Ministry of Agric. and Nat. Resources., Nicosia, Cyprus.
199. Anon. 1987. Abstracts of papers presented at the 6th meeting on whiteflies in field crops and vegetables. Gilat, Israel 26 February, 1987. Phytoparasitica 15(3): 259-265. [ Cock (1993)]
200. Anon. 1987. Save cotton and cash: kill whitefly. Pakistan Agric. 9(10): 7-10.
201. Anon. 1989. Abstracts of papers presented at the 7th meeting on whiteflies in field crops and vegetables held at Bet Dagan, Israel, 9 April, 1989. Phytoparasitica 17(3): 227-234. [ Cock (1993)]
202. Anon. 1989. [*Bemisia tabaci* - a new whitefly in greenhouse crops]. Gärtnermeister 13: 242-243. [ Cock (1993), German]
203. Anon. 1989. Cotton whitefly, *Bemisia tabaci*, identified. FAO Plant Prot. Bull. 37(2): 92. [ Cock (1993)]
204. Anon. 1989. EPPO data sheets on quarantine organisms List A2 No. 178. *Bemisia tabaci* Gennadius (Hemiptera: Aleyrodidae). Bull OEPP 19: 733-738.
205. Anon. 1989. Management of whitefly, *Bemisia tabaci* G. on cotton. Andhra Pradesh Agric. University, Rajendranagar, Hyderabad, 53 pp.
206. Anon. 1990. Outbreaks and new records. Trinidad and Tobago. New diseases and outbreak of whitefly reported. FAO Plant Prot. Bull. 35(1): 52. [ Cock (1993)]
207. Anon. 1991. Control of sweet potato whitefly, *Bemisia tabaci*, on ornamental plants. DSIR Plant Prot. Leaflet 91/1: 1-2. [ Cock (1993)]
208. Anon. 1993. Abstracts of papers presented at the 8th meeting on whiteflies in field crops, vegetables and ornamentals. Phytoparasitica 21(2): 169-180.
209. Anon. 1994. Abstracts of papers presented at the International Workshop of *Bemisia* spp. (An assessment of the biology and management strategies of *Bemisia* spp. from an international perspective). Phytoparasitica 22(4): 309-359.
210. Anon. 1994. Outbreaks and new records: Bermuda: Update on *Bemisia tabaci*: and new reports on citrus tristeza virus and *Aculops lycopersicae*. FAO Plant Prot. Bull. 42(3): 150-151.



211. Anon. 1994. Outbreaks and new records: Dominica: New pest and outbreak problems. *FAO Plant Prot. Bull.* 42(3): 151-152.
212. Anon. 1995. Outbreak of the silverleaf whitefly in the Cook Islands. *Agric. Alert No.* 13, 2pp.
213. Anon. 1996. A mosca blanca *Bemisia tabaci* e o virus frizado amarelo do tomateiro. [The whitefly *Bemisia tabaci* and tomato yellow leaf curl virus]. Instituto de Protecção da Produção Agro-Alimentar, Centro Nacional de Protecção da Produção Agrícola (CNPPA), Oeiras, Portugal [Portuguese]
214. Anon. 1998. Silverleaf whitefly extends range. *California Agric.* 52(2): 6-7.
215. Anon. 2001. Adversities of the industrial and table tomato. *Informatore Agrario Supplemento (Italy)* 57(16): 3-42. [Italian, English summary]
216. Ansolabehere, M.J. and Chernicky, J.P. 1996. Silverleaf whitefly control in cotton with KNACK insect growth regulator. pp. 819-820. *In P. Dugger and D. Richter (Eds.), Proceedings Belt wide Cotton Conferences. National Cotton Council, Memphis, TN.*
217. Ansolabehere, M.J. and Elliott, C. 1997. Silverleaf whitefly control in cotton with KNACK insect growth regulator. pp. 1246-1247. *In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
218. Anthony, K.R.M. and Jones, A.J. 1963. Cotton production in Thailand. *Empire Cotton Growing Rev.* 40: 170-178. [Cock (1986)]
219. Anthony, N.M., Brown, J.K., Feyereisen, R. and French-Constant, R.H. 1998. Diagnosis and characterization of insecticide-insensitive acetylcholinesterase in three populations of the sweetpotato whitefly *Bemisia tabaci*. *Pestic. Sci.* 52(1): 39-46.
220. Anthony, N.M., Brown, J.K., Markham, P.G. and French-Constant, R.H. 1995. Molecular analysis of cyclodiene resistance-associated mutations among populations of the sweetpotato whitefly, *Bemisia tabaci*. *Pestic. Biochem. Physiol.* 51(3): 220-228.
221. Anthony, W.S. 1999. Rapid estimation of cotton stickiness. pp. 690-695. *In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
222. Antignus, Y. 1995. The use of UV absorbing plastic film to protect crops from the white fly and spread of viral diseases and its vectors. *Hassadeh* 75(6): 39-42. [Hebrew]
223. Antignus, Y. 1999. Cultural control of insect-transmitted viruses. pp. 79-89. *In I.M.C. Gomez (Ed.), Current trends in epidemiology and virus control in horticultural crops. Fundacion para la investigacion Agraria en la Provincia de Almeria, Aguadulce (Almeria), Spain.* [Spanish and English]
224. Antignus, Y. 2000. Manipulation of wavelength-dependent behaviour of insects. An IPM tool to impede insects and restrict epidemics of insect-borne viruses. *Virus Res.* 71: 213-220.
225. Antignus, Y. and Cohen, S. 1992. Agroinfection of plants with a single genome component of the whitefly-borne tomato yellow leaf curl virus (TYLCV). pp. 59-60. *In I.S. Rumbos, P. Kyriakopoulou and F. Bem (Eds.), Recent Advances in Vegetable Virus Research. 7th Conference ISHS Vegetable Virus Working Group, Athens, Greece, July 12-16, 1992. Ores Publishing, Volos, Greece.*
226. Antignus, Y. and Cohen, S. 1994. Complete nucleotide sequence of an infectious clone of a mild isolate of tomato yellow leaf curl virus (TYLCV). *Phytopathology* 84(7): 707-712.
227. Antignus, Y., Cohen, S., Mor, N., Masika, Y. and Lapidot, M. 1996. The effects of UV-blocking greenhouse covers on insects and insect-borne virus diseases. *Plasticulture* 112: 15-20.
228. Antignus, Y., Lapidot, M. and Cohen, S. 2001. Interference with ultraviolet vision of insects to impede insect pests and insect-borne plant viruses. pp. 331-350. *In K.F. Harris, O.P. Smith and J.E. Duffus (Eds.), Virus-Insect-Plant Interactions. Academic Press, San Diego, CA.*
229. Antignus, Y., Lapidot, M., Hadar, D., Messika, Y. and Cohen, S. 1998. Ultraviolet-absorbing screens serve as optical barriers to protect crops from virus and insect pests. *J. Econ. Entomol.* 91(6): 1401-1405.
230. Antignus, Y., Mor, N., Joseph, B., Lapidot, M. and Cohen, S. 1996. Ultraviolet-absorbing plastic sheets protect crops from insect pests and from virus diseases vectored by insects. *Environ. Entomol.* 25(5): 919-924.
231. Antignus, Y., Nestel, D., Cohen, S. and Lapidot, M. 2001. Ultraviolet-deficient greenhouse environment affects whitefly attraction and flight-behavior. *Environ. Entomol.* 30(2): 394-399.
232. Antilla, L., El-Lissy, O., Staten, R.T., Walters, M.L. and Leggett, J.E. 1995. Silverleaf whitefly, *Bemisia argentifolii*, control on cotton in Paloma, AZ: a second year. pp. 844-845. *In D.A. Richter and J. Armour (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
233. Antilla, L., Leggett, J.E. and Walters, M.L. 1996. A three year study on the decline of fenpropathrin (Danitol) efficacy in Arizona cotton. pp. 151-153. *In P. Dugger and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
234. Anusonratchada, B., Phungcharoen, P. and Buntham, A. 1992. Survey soybean insect pest in Chiang Mai. pp. 728-736. *In Research Report in 1989: Soybean. Chiang Mai Field Crops Research Center, Thailand.* [Thai, English summary]
235. Anwar, M.P., Munshi, G.H., Hussain, T. and Shahwani, M.I. 1987. Insect pests associated with potato crop at Tandojam. pp. 149-151. *In Proceedings of the 5th Pakistan Congress of Zoology, University of Karachi, Karachi, January 8-11, 1986. Zoological Society of Pakistan, Karachi, Pakistan.* [Cock (1993)]
236. Anzola, D. and Lastra, R. 1978. Protección de semilleros de tomate y su relación con la incidencia del virus mosaico amarillo del tomate. *Agron. Trop.* 28: 473-482.
237. Anzola, D. and Lastra, R. 1985. Whiteflies population and its impact on the incidence of tomato yellow mosaic virus in Venezuela. *Phytopathol. Z.* 112: 363-366. [Cock (1993)]
238. Aoki, K., Yamada, H. and Simohata, T. 1995. Epidemiology and control of *Bemisia tabaci* Gennadius in Gifu Pref. *Bull. Gifu Agric. Res. Center (Japan)* 8: 23-26. [Japanese]
239. Appert, J. 1967. Notes techniques sur les insectes nuisibles aux cultures malagasy. Suite. *Agron. Trop. (Nogent-sur-Marne)* 22: 504-552. [Cock (1986)]
240. Aragao, F.J.L., Ribeiro, S.G., Barros, L.M.G., Brasileiro, A.C.M., Maxwell, D.P., Rech, E.L. and Faria, J.C. 1998. Transgenic beans (*Phaseolus vulgaris* L.) engineered to express viral antisense RNAs show delayed and attenuated symptoms to bean golden mosaic geminivirus. *Mol. Breeding* 4(6): 491-499.
241. Arakaki, N. and Okajima, S. 1998. Notes on the biology and morphology of a predatory thrips, *Franklinothrips vespiformis* (Crawford) (Thysanoptera: Aeolothripidae): first record from Japan. *Entomol. Sci.* 1(3): 359-363.
242. Araya, C.M. and Araya-Villalobos, R. 2000. Historia y situación de la producción de frijol en los países latinoamericanos afectados por geminivirus transmitidos por mosca blanca: América Central: Costa Rica. pp. 41-46. *In Bean Golden Mosaic and Other Diseases of Common Bean Caused by Whitefly-Transmitted Geminiviruses in Latin America. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia.* [Spanish]
243. Arcos-Cavazos, G. 1996. Dinámica poblacional de *Bemisia tabaci* Genn. en hospederos silvestres y cultivadas de Papantla, Veracruz. [Dynamic population of *Bemisia tabaci* Genn in wild and cropped hosts at Papantla, Veracruz]. pp. 289-296. *In Proceedings 9th Scientific and Technological Meeting of Forestry, Agriculture and Husbandry. INIFAP, Veracruz, Mexico.* [Spanish]

244. Arcos-Cavazos, G. 1996. Validación de un programa de manejo integrado de *Bemisia tabaci* Genn. en Chile jalapeño. [Integrated control program of *Bemisia tabaci* Genn. in Jalapeño pepper]. pp. 349-350. In Proceedings 9th Scientific and Technological Meeting of Forestry, Agriculture and Husbandry. INIFAP, Veracruz, Mexico. [Spanish]
245. Aref, N.M. and El-DougDoug, K.A. 1996. Biological and molecular diagnosis of three different symptoms of TYLC-disease in open field. Ain-Shams Univ. Ann. Agric. Sci. (Egypt) 41(1): 173-185.
246. Arenas, L.D.O. 1998. Resistencia de *Bemisia argentifolii* a insecticidas: implicaciones y estrategias de manejo en México. [Resistance of *Bemisia argentifolii* to insecticides: implications and strategies for management in Mexico]. Manejo Integrado de Plagas (Costa Rica) 49: 10-25. [Spanish, English summary]
247. Arevalo, C.E. and Diaz-Chávez, A.L. 1976. Determinación de los períodos mínimos requeridos por *Bemisia tabaci* Genn en la adquisición y transmisión del virus del mosaico dorado del frijol (VMDF). [Determination of the minimal periods required by *Bemisia tabaci* Genn. for the acquisition and transmission of the bean golden mosaic virus]. Reunion Anual del PCCMCA 22: L29-1-8. [CATIE (1992), Spanish]
248. Arguello -Astorga, G.R., Guevara-Gonzalez, G., Herrera-Estrella, L.R. and Rivera-Bustamante, R.F. 1994. Geminivirus replication origins have a group-specific organization of iterative elements: a model for replication. Virology 203: 90-100.
249. Arias T. R. and Hilje, L. 1993. Actividad diaria de los adultos de *Bemisia tabaci* (Gennadius) en el tomate y hospedantes alternos del insecto. [Daily activity of adult *Bemisia tabaci* (Gennadius) in tomatoes and the alternative host plants of the insect]. Manejo Integrado de Plagas (Costa Rica) 28: 20-25. [Spanish, English summary]
250. Arias T. R. and Hilje, L. 1993. Uso del frijol como cultivo trampa y de un aceite agrícola para disminuir la incidencia de virosis transmitida por *Bemisia tabaci* en el tomate. [The combined use of beans as a trap crop and a mineral oil to reduce the incidence of virus transmitted by *Bemisia tabaci* in tomatoes]. Manejo Integrado de Plagas (Costa Rica) 27: 27-34. [Spanish, English summary]
251. Arioglu, H.H. 1987. Research on growing possibilities of some determinate soybean varieties as a second crop in Çukurova, Turkey. Soy. Genet. Newsl. 14: 131-135. [Cock (1993)]
252. Arioglu, H.H. 1987. Screening of some soybean varieties for resistance to whitefly (*Bemisia tabaci* Genn.). Soy. Genet. Newsl. 14: 136-139. [Cock (1993)]
253. Arioglu, H.H., Ozgur, F. and Isler, N. 1989. The effect of whitefly (*Bemisia tabaci* Genn.) damage on yield components in double-cropped soybean production. Soy. Genet. Newsl. 16: 57-61. [Cock (1993)]
254. Aritua, V., Adipala, E., Carey, E.E. and Gibson, R.W. 1998. The incidence of sweet potato virus disease and virus resistance of sweet potato grown in Uganda. Ann. Appl. Biol. 132(3): 399-411.
255. Aritua, V., Alicai, T., Adipala, E., Carey, E.E. and Gibson, R.W. 1998. Aspects of resistance to sweet potato virus disease in sweet potato. Ann. Appl. Biol. 132(3): 387-398.
256. Aritua, V., Legg, J.P., Smit, N.E.J.M. and Gibson, R.W. 1999. Effect of local inoculum on the spread of sweet potato virus disease: limited infection of susceptible cultivars following widespread cultivation of a resistant sweet potato cultivar. Plant Pathol. 48(5): 655-661.
257. Arnal, E., Debrot, E., Marcano, R. and Montagne, A. 1993. Fluctuación poblacional de moscas blancas y su relación con el mosaico amarillo del tomate en una localidad de Venezuela. [Population fluctuation of whiteflies and its relation to tomato yellow mosaic in one location in Venezuela]. Fitopatol. Venezolana 6(1): 21-26. [Spanish, English summary]
258. Arnal, E. and Ramos, F. 2000. Additions of noteworthy records to the list of white flies of (Homoptera: Aleyrodidae) of Venezuela. Boletín Entomol. Venezolana 15(1): 97-107. [Spanish, English summary]
259. Arnal, E., Ramos, F. and Debrot, E. 1993. Plantas hospederas de la mosca blanca *Bemisia tabaci* (Gennadius) en Venezuela. [Host plants of the whitefly *Bemisia tabaci* (Gennadius) in Venezuela]. Agron. Trop. (Venezuela) 43(5/6): 267-285. [Spanish, English summary]
260. Arnal, E., Ramos, F., Soto, E. and Soto, E. 1996. Evaluación de insecticidas para controlar la mosca blanca en melón. [Evaluation of insecticides for control of whitefly on melon]. Agron. Trop. (Venezuela) 46(3): 237-249. [Spanish, English summary]
261. Arnal, E., Russell, L.M., Debrot, E., Ramos, F., Cermeli, M., Marcano, R. and Montagne, A. 1993. A listing of white flies (Homoptera, Aleyrodidae) and their host plants in Venezuela. Florida Entomol. 76: 365-381.
262. Arnó, J. and Gabarra, R. 1994. Whitefly species composition in winter tomato greenhouses. Bull. OILB/SROP (IOBC/WPRS) 17: 104-109.
263. Arnó, J. and Gabarra, R. 1996. Potential for biological control of mixed *Trialeurodes vaporariorum* and *Bemisia tabaci* populations in winter tomato crops grown in greenhouses. pp. 523-526. In D. Gerling and R.T. Mayer (Eds.), *Bemisia*: 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
264. Arutkani, K.E., Ayyanathan and Chandrasekaran, A. 1999. Evaluation of greengram genotypes for resistance to yellow mosaic disease. Madras Agric. J. 86(7-9): 527-528.
265. Arya, D.R., Yadav, P.R. and Singh, H.V. 1995. Insect pest complex of sunflower in relation to crop phenology. Indian J. Entomol. 57(2): 141-145.
266. Ascencio-Ibáñez, J.T., Diaz-Plaza, R., Méndez-Lozano, J., Monsalve-Fonnegra, Z.I., Argüello -Astorga, G.R. and Rivera-Bustamante, R.F. 1999. First report of tomato yellow leaf curl geminivirus in Yucatán, México. Plant Dis. 83(12): 1078.
267. Ascher, K.R.S. and Eliyahu, M. 1988. The ovidical properties of the juvenile hormone mimic Sumitomo S-31183 (SK-591) to insects. Phytoparasitica 16(1): 15-21. [Cock (1993)]
268. Ascher, K.R.S., Eliyahu, M., Nemny, N.E. and Ishaaya, I. 1986. The toxicity of synthetic pyrethroids and other insecticides and pesticides to *Spodoptera littoralis* (Boisd.), *Drosophila melanogaster* Meig. and *Bemisia tabaci* (Genn.). Int. Pest Control 28(3): 68-71, 74-78. [Cock (1993)]
269. Ascher, K.R.S., Ishaaya, I., Barros, J., Zur, M. and Ben-Moshe, E. 1985. The residual effect of three pyrethroids, PP 321 (cyhalothrin), Smash (fenprothrin) and Baythroid (cyfluthrin) on *Bemisia tabaci* adults under glasshouse and field conditions. Hassadeh 65: 888-891. [Cock (1993)]
270. Ashraf, M., Zafar, Z.U., McNeilly, T. and Veltkamp, C.J. 1999. Some morpho-anatomical characteristics of cotton (*Gossypium hirsutum* L.) in relation to resistance to cotton leaf curl virus (CLCuV). J. Appl. Bot. 73(3-4): 76-82.
271. Asiático, J. and Zoebisch, T.G. 1992. Control de mosca blanca *Bemisia tabaci* (Gennadius) en tomate con insecticidas de origen biológico y químico. [Control of the whitefly *Bemisia tabaci* (Gennadius) in tomatoes by biological and chemical insecticides]. Manejo Integrado de Plagas (Costa Rica) 24-25: 1-7. [Spanish, English summary]
272. Aslam, M. and Gebara, F. 1995. Host plant preference of vegetables by cotton whitefly, *Bemisia tabaci* (Genn.). Pakistan J. Zool. 27(3): 269-272.
273. Asly, O.J. 1987. Utilization of non woven fabrics against the TYLCV virus of tomatoes. Plasticulture (France) 74: 47-51.
274. Aspiazú, M.D., Cairo, V.G. and Hoz-Gonzalez, M.T. 1995. Ciclo de vida de *Bemisia tabaci* (Homoptera: Aleyrodidae) en tomate. [Life cycle of *Bemisia tabaci* (Homoptera: Aleyrodidae) in tomato. Rev. Biol. (Habana) 9: 113-115. [In Spanish]
275. Assad, Y.O.H., Bashir, N.H.H. and Eltoum, E.M.A. 1999. Development of resistance to endosulfan, deltamethrin, and chlorpyrifos in the cotton whitefly, *Bemisia tabaci* (Genn.), in the Sudan Gezira. Sudan J. Agric. Res. 2: 57-63.

276. Atta-Aly, M.A., Abdel-Megeed, M.I., Hegab, M.F. and Kamel, M.H. 1998. Increasing tomato plant growth and yield with improving fruit quality by controlling sap-sucking insects (whitefly and aphid) without insecticides. *Ain -Shams Univ. Ann. Agric. Sci. (Egypt)* 43: 845-863.
277. Attathom, D.T.S. and Sutabutra, T. 1986. Tomato yellow leaf curl virus in Thailand. [Chinese & Japanese summary]. pp. 60-63. *In* Plant Virus Diseases of Horticultural Crops in the Tropics and Subtropics. Food and Fertilizer Technology Centre for the Asian and Pacific Region [Cock (1993)]
278. Attia, M.B., Afifi, F.M.L. and El-Sherif, S.I. 1985. Effect of certain plant growth regulator and foliar fertilizer treatments on the population of the whitefly *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae), cotton aphid *Aphis gossypii* Glover (Homoptera: Aphididae) and the pink bollworm *Pectinophora gossypiella* (Saund.) (Lepidoptera, Gelechiidae) in cotton fields. *Bull. Faculty Agric. (Univ. Cairo)* 36(2): 1269-1277. [CATIE (1992)]
279. Attique, M.R., Arif, M.I., Ahmad, Z. and Mohyuddin, A.I. 2001. Effect of spray volume and group of insecticide on *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae) and its parasitoids. *Pakistan J. Zool.* 33(3): 201-204.
280. Attique, M.R. and Ghaffar, A. 1996. Control of early season sucking pests of cotton with seed protectant insecticides and their impact on natural enemies and yield of seed cotton. *Pakistan J. Zool.* 28(3): 253-255.
281. Attique, M.R., Ghaffar, A., Ahmad, Z. and Mohyuddin, A.I. 1999. Monitoring adult activity of *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae) with yellow sticky traps and its relationship with actual field population in Pakistan. *Pakistan J. Zool.* 31(4): 301-306.
282. Attique, M.R. and Shakeel, M.A. 1983. Comparison of ULV with conventional spraying on cotton in Pakistan. *Crop Prot.* 2: 231-234. [Cock (1986)]
283. Atwal, A.S. and Singh, K. 1969. Chemical control of cotton jassid (*Empoasca devastans* Distant) and whitefly (*Bemisia tabaci* Genn.). *J. Res. (Punjab Agric. Univ.)* 6 Suppl.(1): 237-240. [Cock (1986)]
284. Atwal, A.S. and Singh, K. 1969. Efficacy of various spraying schedules on cotton. *J. Res. (Punjab Agric. Univ.)* 6(1 suppl.): 661-667. [Cock (1986)]
285. Atzmon, G., van Oss, H. and Czosnek, H. 1998. PCR-amplification of tomato yellow leaf curl virus (TYLCV) DNA from squashes of plants and whitefly vectors: Application to the study of TYLCV acquisition and transmission. *Eur. J. Plant Pathol.* 104(2): 189-194.
286. Auad, A.M., Toscano, L.C., Boica, A.L. and Freitas, S. 2001. Aspectos biológicos dos estádios imaturos de *Chrysoperla externa* (Hagen) e *Ceraeochrysa cincta* (Schneider) (Neuroptera: Chrysopidae) alimentados com ovos e ninfas de *Bemisia tabaci* (Gennadius) Biotipo B (Homoptera: Aleyrodidae). *Neotrop. Entomol.* 30(3): 429-432. [Portuguese, English summary]
287. Ausher, R. 1996. Implementation of integrated pest management in Israel. pp. 659-665. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia* : 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
288. Ausher, R. 1997. Implementation of integrated pest management in Israel. *Phytoparasitica* 25(2): 119-141.
289. Avgelis, A.D., Roditakis, N., Dovas, C.I., Katis, N.I., Varveri, C., Vassilakos, N. and Bem, F. 2001. First report of Tomato yellow leaf curl virus on tomato crops in Greece. *Plant Dis.* 85(6): 678.
290. Avidov, Z. 1956. Bionomics of the tobacco whitefly (*Bemisia tabaci* Genn.) in Israel. *Khavim (English edition)* 7(1): 25-41. [Cock (1986)]
291. Avidov, Z. and Harpaz, I. 1969. Family Aleyrodidae. pp. 76-84. *In* Plant Pests of Israel. University Press, Jerusalem.
292. Avila-Valdez, J. 1998. Evaluación del impacto de las liberaciones masivas de *Chrysoperla carnea* (Neuroptera: Chrysopidae) sobre las poblaciones de *Bemisia tabaci* (Homoptera aleyrodidae) en tomate [Impact evaluation of mass liberation of *Chrysoperla carnea* (Neuroptera: Chrysopidae) on *Bemisia tabaci* (Homoptera aleyrodidae) in tomato]. pp. 186-188. *In* Memoria. 21. Congreso Nacional De Control Biológico, Rio Bravo, Tampaulipas, Mexico, 5-6 Nov 1998. INIFAP. [Spanish]
293. Aviles-Baeza, W.I. 1995. Plantas hospederas de mosquita blanca *Bemisia tabaci* Genn. asociadas al tomate *Lycopersicum esculentum* Mill. en el norte de Yucatan. [Host plants of the whitefly *Bemisia tabaci* Genn. associated with tomato *Lycopersicum esculentum* Mill. in the north of Yucatan]. *Agric. Tecnica Mexico* 21(1): 63-78. [Spanish]
294. Aviles-Gonzalez, M. and Martinez-Carrillo, J.L. 1998. Efecto de fecha de siembra sobre parasitoides de mosquita blanca *Bemisia argentifolii* (Homoptera: Aleyrodidae) Bellows and Perring, en variedades de soya. Valle de Culiacan. 1997. [Sowing date effect on white fly *Bemisia argentifolii* (Homoptera: Aleyrodidae), in soybean varieties in Culiacan Valley 1997. Sinaloa State]. pp. 326-328. *In* Memoria. 21. Congreso Nacional De Control Biológico, Rio Bravo, Tampaulipas, Mexico, 5-6 Nov 1998. INIFAP. [Spanish]
295. Aviles-Gonzalez, M. and Martinez-Carrillo, J.L. 1998. Parasitismo de la mosquita blanca *Bemisia argentifolii* Bellow & Perring (Homoptera: Aleyrodidae) en materiales de soya. Valle de Culiacan, 1997. [White fly *Bemisia argentifolii* Bellow and Perring parasitism (Homoptera: Aleyrodidae) in soybean. Culiacan Valley, 1997. Sinaloa State]. pp. 323-325. *In* Memoria. 21. Congreso Nacional De Control Biológico, Rio Bravo, Tampaulipas, Mexico, 5-6 Nov 1998. INIFAP. [Spanish]
296. Avilés, M. 1996. Reacción de materiales de tomate al ataque de la mosquita blanca (*Bemisia* spp.) en el Valle de Culiacán, Sin Mosquita blanca en el noroeste de Mexico. *Memoria Científica* 2: 72-23.
297. Awadallah, K.T., Tawfik, M.F.S. and Shalaby, F. 1976. Insect fauna of the bind-weed, *Convolvulus arvensis* L., in Giza, Egypt. *Bull. Entomol. Soc. Egypt* 60: 15-24. [Cock (1986)]
298. Azab, A.K., Megahed, M.M. and El-Mirsawi, H.D. 1969. Effect of degree of pubescence of host-plant on the number and distribution of dorsal spines in pupa of *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae). *Bull. Entomol. Soc. Egypt* 53: 353-357. [Cock (1986)]
299. Azab, A.K., Megahed, M.M. and El-Mirsawi, H.D. 1969. Parasitism of *Bemisia tabaci* (Genn.) in U.A.R. (Homoptera-Homoptera: Aleyrodidae). *Bull. Entomol. Soc. Egypt* 53: 439-441. [Cock (1986)]
300. Azab, A.K., Megahed, M.M. and El-Mirsawi, H.D. 1970. On the range of host-plants of *Bemisia tabaci* (Genn.). *Bull. Entomol. Soc. Egypt* 54: 319-326. [Cock (1986)]
301. Azab, A.K., Megahed, M.M. and El-Mirsawi, H.D. 1970. Studies on *Bemisia tabaci* (Genn.) (Homoptera-Homoptera: Aleyrodidae). *Bull. Entomol. Soc. Egypt* 53: 339-352. [Cock (1986)]
302. Azab, A.K., Megahed, M.M. and El-Mirsawi, H.D. 1971. On the biology of *Bemisia tabaci* (Genn.) (Homoptera-Homoptera: Aleyrodidae). *Bull. Entomol. Soc. Egypt* 55: 305-315. [Cock (1986)]
303. Azam, K.M., Razvi, S.A., Al-Muhthuri, M.H. and Al-Raeesi, A.A. 1997. Distribution pattern of sweet potato whitefly *Bemisia tabaci* (Gennadius) on tomato plants. *Sultan-Qaboos Univ. J. Sci. Res. Agric. Sci.* 2: 43-50.
304. Azam, K.M., Razvi, S.A., Zouba, A. and Al-Raeesi, A.A. 1997. Management of whitefly (*Bemisia tabaci* Gennadius) and tomato leaf curl virus in tomato crops. *Indian J. Plant Prot.* 25(1): 36-41.
305. Azam, K.M., Zouba, A., Razvi, S.A. and Al Raeesi, A.A. 1996. Evaluation of insecticides and Agril polyester cover against whitefly (*Bemisia tabaci* Gennadius) in tomato crops. *Sultan-Qaboos Univ. J. Sci. Res., Agric. Sci.* 1: 13-16.

306. Azevedo, A.C.S., Sosa, G.D.R., Faria, M. and Fungaro, M.H.P. 2000. Effects of double-stranded RNA on virulence of *Paecilomyces fumosoroseus* (Deuteromycotina: Hyphomycetes) against the silverleaf whitefly, *Bemisia tabaci* strain B (Homoptera: Aleyrodidae). *Genet. Mol. Biol.* 23(1): 61-63.
307. Azmi, O.R. and Rathi, Y.P.S. 1992. Efficacy of insecticides against whitefly - a vector of fresh bean crinkle stunt virus. *Plant. Dis. Res.* 7(2): 257-258.
308. Azzam, O., Frazer, J., Rosa, D., Beaver, J.S., Ahlquist, P. and Maxwell, D.P. 1994. Whitefly transmission and efficient ssDNA accumulation of bean golden mosaic geminivirus require functional coat protein. *Virology* 204(1): 289-296.
309. Azzam, O., Russell, D.R., Ahlquist, P. and Maxwell, D.P. 1994. Evaluation of different antiviral strategies to control bean golden mosaic geminivirus. pp. 275-282. In W.M. Roca, J.E. Mayer, M.A. Pastor-Corrales and M.J. Tohme (Eds.), *Phaseolus Beans Advanced Biotechnology Research Network: Proceedings of the Second International Scientific Meeting*, 7-10 September 1993, Cali, Colombia. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia.
310. Ba Angood, S.A., Ghaleb, A.M. and Ali, A.M. 2000. Effect of sowing dates on the occurrence of the whitefly *Bemisia tabaci* and the jassid *Jacobiasca lybica* on two different local cultivars of sesame in Yemen. *Univ. Aden J. Nat. Appl. Sci.* 4(1): 103-110.
311. Ba-Angood, S.A. and Mogahed, A.A. 1997. Republic of Yemen. pp. 85-91. In N. Ioannou (Ed.), *Management of the whitefly-virus complex*. FAO Plant Production and Protection Paper 143, Rome, Italy.
312. Babcock, C.S. and Heraty, J.M. 2000. Molecular markers distinguishing *Encarsia formosa* and *Encarsia luteola* (Hymenoptera: Aphelinidae). *Ann. Entomol. Soc. Am.* 93(4): 738-744.
313. Babcock, C.S., Heraty, J.M., De Barro, P.J., Driver, F. and Schmidt, S. 2001. Preliminary phylogeny of *Encarsia Forster* (Hymenoptera: Aphelinidae) based on morphology and 28S rDNA. *Mol. Phylogenet. Evol.* 18(2): 306-323.
314. Backus, E.A. 1988. Sensory systems and behaviors which mediate hemipteran plant-feeding: a taxonomic overview. *J. Insect Physiol.* 34: 151-165.
315. Badge, J., Brunt, A., Carson, R., Dagless, E., Karamagioli, M., Phillips, S., Seal, S., Turner, R. and Foster, G.D. 1996. A carlavirus-specific PCR primer and partial nucleotide sequence provides further evidence for the recognition of cowpea mild mottle virus as a whitefly-transmitted carlavirus. *Eur. J. Plant Pathol.* 102(3): 305-310.
316. Bailey, M.A. 1934. Leaf curl disease of cotton in the Sudan. *Empire Cotton Growing Rev.* 11: 280-288. [Cock (1986)]
317. Bailey, N.M., Bailey, C.A. and Reichard, S.M. 1982. Enzymatic evaluation of sugar content of cotton. *Textile Res. J.* 52: 321-327.
318. Baker, J.R., Crouse, M.B. and Shearin, E.A. 1993. Screening as part of insect and disease management in the greenhouse. *North Carolina Flower Growers' Bull.* 38(4): 12-17.
319. Baker, R.H.A. and Cheek, S. 1993. *Bemisia tabaci* in the United Kingdom. *Bull. OILB/SROP. (IOBC/WPRS)* 16(8): 6-11.
320. Bakheit, I.B. 1991. Entomology of soya bean, sesame, sunflower and sorghum. pp. 56-67. In *Annual Report of Kenana Agric. Research Station (Sudan)*. 1985-86.
321. Bakhetia, D.R.C., Kaur, S., Sandhu, I.S., Brar, D.S. and Kular, J.S. 1997. Monitoring of insect pests and quantification of yield losses in sunflower. *J. Insect Sci.* 10(2): 140-142.
322. Balaji, K. and Veeravel, R. 1994. Leaf morphological and biochemical studies on the four varieties of brinjal in relation to whitefly (*Bemisia tabaci* Genn.) incidence. *Prog. Hortic.* 26(1/2): 45-52.
323. Balaji, K. and Veeravel, R. 1995. Effect of constant temperature on the duration of different developmental stages of whitefly, *Bemisia tabaci* G. on brinjal. *Madras Agric. J.* 82(1): 62-63.
324. Balaji, K. and Veeravel, R. 1995. Effect of different levels of nitrogen on the incidence of *Bemisia tabaci* (Genn.) in four varieties of brinjal. *Indian J. Entomol.* 57(4): 356-361.
325. Balaji, K. and Veeravel, R. 1998. Incidence of whitefly, *Bemisia tabaci* G. (Aleyrodidae: Homoptera) on four varieties of brinjal at different periods of plant growth. *South Indian Hortic.* 46(3-6): 220-222.
326. Balakrishnan, S. and Nene, Y.L. 1980. A note on the mode of penetration of the fungus *Paecilomyces farinosus* (Dickson ex Fries) Brown & Smith into the whitefly *Bemisia tabaci* Gennadius. *Sci. Culture* 46: 231-232. [Cock (1993)]
327. Balan, J.S. 1967. Effect of different coloured lights on the development of *Bemisia tabaci* Genn. *Plant Prot. Bull. (Sci. in Practice)* 19(1): 30-33. [Cock (1986)]
328. Balasubramani, V. and Swamiappan, M. 1994. Development and feeding potential of the green lacewing *Chrysopa carnea* Steph. (Neur. Chrysopidae) on different insect pests of cotton. *Anz. Schadlingskunde, Pflanzenschutz, Umweltschutz* 67(8): 165-167.
329. Balasubramanian, G. and Chelliah, S. 1985. Chemical control of pests of sunflower. *Pesticides* 19(4): 21-22. [Cock (1993)]
330. Balasubramanya, R.H., Bhatawdekar, S.P. and Paralakar, K.M. 1985. A new method for reducing the stickiness of cotton. *Textile Res. J.* 55: 227-232.
331. Baldanza, F., Gaudio, L. and Viggiani, G. 1999. Cytotaxonomic studies of *Encarsia Forster* (Hymenoptera: Aphelinidae). *Bull. Entomol. Res.* 89(3): 209-215.
332. Baldin, E.L.L., Toscano, L.C., Lima, A.C.S., Lara, F.M. and Boica, A.L. 2000. Preferencia para oviposicao de *Bemisia tabaci* biotipo "B" por genotipos de *Cucurbita moschata* e *Cucurbita maxima*. [Oviposition preference of *Bemisia tabaci* "B" biotype for *Cucurbita moschata* and *Cucurbita maxima* genotypes]. *Boletim Sanidad Vegetal Plagas (Espana)* 26(3): 409-413. [Portuguese, English summary]
333. Baliadi, Y. and Saleh, N. 1993. The effect of three varieties and planting distance of soybean to cowpea mild mottle virus and its vector *Bemisia tabaci*. pp. 76-80. In *Proceeding of the Seminar on Food Crop Research Result in 1992*. Balai Penelitian Tanaman Pangan, Malang, Indonesia. [Indonesian, English summary]
334. Balla, A.N. 1978. Large-scale evaluation of some insecticides in the control of American bollworm (*Helicoverpa armigera armigera* Hubn), whitefly (*Bemisia tabaci*) and jassid (*Empoasca lybica*). pp. 275-280. In *Annu. Rep. Gezira Res. Stn. Substns. 1966/1967*. Agric. Res. Corp., Sudan.
335. Balla, A.N. 1978. Small-scale evaluation of some insecticides for the control of American bollworm (*Helicoverpa armigera armigera* Hubn), jassid (*Empoasca lybica*) and whitefly (*Bemisia tabaci*). pp. 264-274. In *Annu. Rep. Gezira Res. Stn. Substns. 1966/1967*. Agric. Res. Corp., Sudan.
336. Balla, A.N. 1981. Entomology. pp. 162-172. In *Annu. Rep. Gezira Res. Stn. Substns. 1974/1975*. Agric. Res. Corp., Sudan.
337. Balla, A.N. 1982. Entomology. pp. 166-181. In *Annu. Rep. Gezira Res. Stn. Substns. 1975/1976*. Agric. Res. Corp., Sudan.
338. Ballmer, G.R., Toscano, N.C. and Adams, C. 1991. The impact of sweetpotato whitefly, *Bemisia tabaci*, upon cotton quantity and quality in California. p. 714. In D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
339. Baloch, A.A. 1988. A review on the management of cotton whitefly. *Pakistan Cotton* 32(4): 214-233. [Cock (1993)]
340. Baloch, A.A., Kalroo, A.M. and Soomro, B.A. 1988. Studies on the effectiveness of granular pesticides against sucking pest complex of cotton. *Pakistan Cotton* 32(2): 79-86.
341. Baloch, A.A. and Soomro, B.A. 1980. Preliminary studies on plant profile and population dynamics of insect pests of cotton. *Türkiye Bitki Koruma Dergisi* 4(4): 203-217. [Cock (1986)]

342. Baloch, A.A., Soomro, B.A. and Korejo, A.K. 1990. Population dynamics studies on whitefly (*Bemisia tabaci*) in Sind: II. Population trend on alternate host plants. *Pakistan J. Agric. Res.* 6(1/2): 1-14.
343. Baloch, A.A., Soomro, B.A. and Mallah, G.H. 1982. Evaluation of some cotton varieties with known genetic markers for their resistance/tolerance against sucking and bollworm complex. *Türkiye Bitki Koruma Dergisi* 6(1): 3-14. [Cock (1986)]
344. Banerjee, M.K. and Kalloo, G. 1987. Inheritance of tomato leaf curl virus in *Lycopersicon hirsutum* f. *glabratum*. *Euphytica* 36(2): 581-584.
345. Banfield, M.J., Salvucci, M.E., Baker, E.N. and Smith, C.A. 2001. Crystal structure of the NADP(H)-dependent ketose reductase from *Bemisia argentifolii* at 2.3 Å resolution. *J. Mol. Biol.* 306(2): 239-250.
346. Banks, G.K., Bedford, I.D., Beitia, F.J., Rodriguez-Cerezo, E. and Markham, P.G. 1999. A novel geminivirus of *Ipomoea indica* (Convolvulaceae) from southern Spain. *Plant Dis.* 83(5): 486.
347. Banks, G.K., Colvin, J., Reddy, R.V.C., Maruthi, M.N. and Muniyappa, V. 2001. First report of the *Bemisia tabaci* B biotype in India and an associated tomato leaf curl disease epidemic. *Plant Dis.* 85(2): 231.
348. Bao, S.N., Kitajima, E.W., Callaini, G. and Dallai, R. 1996. Virus-like particles and rickettsia-like organisms in male germ and cyst cells of *Bemisia tabaci* (Homoptera, Aleyrodidae). *J. Invertebr. Pathol.* 67(3): 309-311.
349. Bao, S.N., Kitajima, E.W., Callaini, G., Lupetti, P. and Dallai, R. 1997. Spermatogenesis in three species of whitefly (Homoptera, Aleyrodidae). *Acta Zool. (Stockholm)* 78(2): 163-170.
350. Baraja, M.J., Gonzales, S. and Montalban, C. 1996. Manejo integrado en cultivo de melon entutorado bajo invernadero [Integrated management in a staked glasshouse melon crop]. *Horticultura, Revista de Hortalizas, Flores y Plantas Ornamentales* 113: 29-32. [Spanish]
351. Baranowski, T. and Dankowska, E. 1992. Quarantine pests of greenhouse crops. *Ochroza Roslin* 36(1): 13-15. [Polish]
352. Baranowski, T., Dankowska, E. and Gorski, R. 1992. New quarantine glasshouse pests in Poland and coloured sticky traps for their monitoring. *Bull. OEPP* 22: 347-349. [DeBarro (1995)]
353. Barbosa, F.R., Moreira, A.N., De Alencar, J.A., Haji, F.N.P. and Medina, V.D. 2000. Sampling methodology and activities against the main pests of mango in the Sao Francisco valley. *Circular Tecnica Da EMBRAPA Semi Arido*, No. 50, 23 pp. [Portuguese]
354. Barinaga, M. 1993. Is devastating whitefly invader really a new species? *Science* 259: 30.
355. Barnadas, I., Gabarra, R. and Albajes, R. 1998. Predatory capacity of two mirid bugs preying on *Bemisia tabaci*. *Entomol. Exp. Appl.* 86(2): 215-219.
356. Barnes, J.M., Trinidad-Correa, R., Orum, T.V., Felix -Gastelum, R. and Nelson, M.R. 1999. Landscape ecology as a new infrastructure for improved management of plant viruses and their insect vectors in agroecosystems. *Ecosyst. Health* 5(1): 26-35.
357. Barr, C.L. and Drees, B.M. 1992. The poinsettia strain of the sweetpotato whitefly. *Texas Nurseryman* 23(1): 8-12.
358. Barradas, M.M. and Chagas, C.M. 1982. Mosaico dourado de *Vigna luteola* (Jacq.) Betham, Leguminosa da vegetacao espontanea. *Arch. Inst. Biol.* 49: 85-88. [Cock (1986)]
359. Barreto, B.A., da Silva, T.L. and Teixeira, R.M. 1980. Ocorrencia de 'mosca branca' *Bemisia tabaci* (Gennadius 1889) (Homoptera: Aleyrodidae) em feijoeiro (*Phaseolus vulgaris* L.) no estado do Rio Grande do Sul. *Agron. Sulriograndense* 16(2): 363-365. [Cock (1986)]
360. Bartlett, A.C. and Gawel, N.J. 1993. Determining whitefly species. *Science* 261: 1333-1334.
361. Bartlett, P.W. 1992. Experience of polyphagous alien pests of protected crops in Great Britain. *Bull. OEPP* 22: 337-346. [DeBarro (1995)]
362. Bartsch, R. 1978. Economic problems of pest control. Examined for the case of the Gezira/Sudan. Munchen, German Federal Republic; Weltforum Verlag, 124 pp. [Cock (1986)]
363. Bashir, M. and Malik, B.A. 1988. Diseases of major pulse crops in Pakistan - a review. *Trop. Pest Manage.* 34: 309-314.
364. Bashir, N.H.H. 1999. *Bemisia tabaci* (Genn.) resistance in the Sudan: status quo. pp. 73-80. *In* Proceedings of the Fifth International Conference on Pests in Agriculture, Part I, Montpellier, France, 7-9 December, 1999. Association Nationale pour la Protection des Plantes (ANPP), Paris, France .
365. Bashir, N.H.H. and Abdalhadi, M.A. 1986. Screening of some insecticides against cucumber insect pests in the Sudan Gezira. [Arabic summary]. *Iraqi J. Agric. Sci., ZANCO* 4 Suppl.: 39-45. [Cock (1993)]
366. Basu, A.K. 1987. Resurgence of whitefly in cotton and strategies for its management. pp. 129-133. *In* S. Jayaraj (Ed.), Resurgence of Sucking Pests: Proceedings of National Symposium. Centre For Plant Protection Studies, Tamil Nadu Agric. Univ.
367. Basu, A.K. 1990. Cotton research in India with particular references to stickiness. pp. 8-15. *In* Cotton Production Research from a Farming Systems Perspective, with Special Emphasis on Stickiness, 49th Plenary Meeting. International Cotton Advisory Committee, Washington D. C.
368. Basu, A.N. 1995. *Bemisia tabaci* (Gennadius): Crop Pest and Principal Whitefly Vector of Plant Viruses. Westview Press, Boulder, Colorado, 183 pp.
369. Basyir, A. 1990. The effect of planting season on the pests population, growth, and yield of mungbean. pp. 100-104. *In* M. Dahlan, Sudaryono, A. Kasno, H. Suyamto, H.K. Hartojo, N. Saleh, Sunardi and A. Winarto (Eds.), Proceeding of Research Results of Food Crops in Malang Research Institute for Food Crops, 14-15 March 1990. Balai Penelitian Tanaman Pangan, Malang, Indonesia. [Indonesian, English summary]
370. Batarseh, S.F. and Jaddou, M.I. 1997. Jordan. pp. 33-40. *In* N. Ioannou (Ed.), Management of the whitefly-virus complex. FAO Plant Production and Protection Paper 143, Rome, Italy.
371. Bates, R.B., Byrne, D.N., Kane, V.V., Miller, W.B. and Taylor, S.R. 1990. N.M.R. Characterization of trehalulose from the excrement of the sweet potato whitefly, *Bemisia tabaci*. *Carbohydr. Res.* 201(2): 342-345.
372. Baufeld, P. and Unger, J.G. 1994. New aspects on the significance of *Bemisia tabaci* (Gennadius). *Nachrichtenblatt des Deutschen Pflanzenschutzdienstes* 46(11): 252-257. [German, English Summary]
373. Baumgärtner, J., Delucchi, V., Von Arx, R. and Rubli, D. 1986. Whitefly (*Bemisia tabaci* Genn., Stern.: Aleyrodidae) infestation patterns as influenced by cotton, weather and *Heliothis*: hypotheses testing by using simulation models. *Agric. Ecosystems Environ.* 17(1-2): 49-59. [Cock (1993)]
374. Baumgärtner, J. and Yano, E. 1990. Whitefly population dynamics and modelling. pp. 123-146. *In* D. Gerling (Ed.), Whiteflies: their Bionomics, Pest Status and Management. Intercept, Andover, UK.
375. Baur, E. 1906. Über die infektiöse Chlorose der Malvaceen. *Sitzungsberichte der Koniglich Preussischen Akademie der Wissenschaften* 1: 11-29. [Cock (1986)]
376. Bayart, J.D., Wilson, V., Salin, D. and Reynaud, P. 2001. *Bemisia tabaci* on imported plants: results of the French official control. *Phytoma (France)* 535: 45-46. [French, English summary]
377. Beaver, J. and Morales, F.J. 2000. Historia y situacion de la produccion de frijol en los paises latinoamericanos afectados por geminivirus transmitidos por mosca blanca: Region Caribe: Puerto Rico. pp. 65-68. *In* Bean Golden Mosaic and Other Diseases of Common Bean Caused by Whitefly -Transmitted Geminiviruses in Latin America. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. [Spanish]

378. Becker, H., Corliss, J., De Quattro, J., Gerriets, M., Senft, D., Stanley, D. and Wood, M. 1992. Get the whitefly swatters - fast! Agric. Res. (USDA) 40(11): 4-13.
379. Bedford, H.W. 1936. Entomological Section Agricultural Research Service. Report on work carried out by the staff of the section during the season 1934-35. Rep. Agric. Res. Ser. (Anglo-Egyptian Sudan) 1935: 63-96. [Cock (1986)]
380. Bedford, H.W. 1937. Entomological Section Agricultural Research Service. Report - season 1935-36. Rep. Agric. Res. Ser. (Anglo - Egyptian Sudan) 1936: 38-52. [Cock (1986)]
381. Bedford, H.W. 1938. Entomological Section Agricultural Research Service. Report ... 1936-37. Rep. Agric. Res. Ser. (Anglo-Egyptian Sudan) 1937: 50-65. [Cock (1986)]
382. Bedford, H.W. 1940. Entomological Section Agricultural Research Service. Report ... 1937-38. Rep. Agric. Res. Ser. (Anglo-Egyptian Sudan) 1938: 50-71. [Cock (1986)]
383. Bedford, I.D., Briddon, R.W., Brown, J.K., Rosell, R.C. and Markham, P.G. 1994. Geminivirus transmission and biological characterisation of *Bemisia tabaci* (Gennadius) biotypes from different geographic regions. Ann. Appl. Biol. 125(2): 311-325.
384. Bedford, I.D., Briddon, R.W., Jones, P., Alkaff, N. and Markham, P.G. 1994. Differentiation of three whitefly-transmitted geminiviruses from the Republic of Yemen. Eur. J. Plant Pathol. 100(3-4): 243-257.
385. Bedford, I.D., Briddon, R.W., Markham, P.G., Brown, J.K. and Rosell, R.C. 1992. *Bemisia tabaci* biotype characterisation and the threat of this whitefly species to agriculture. pp. 1235-1248. In Brighton Crop Protection Conference: Pests and Diseases. The British Crop Protection Council, Farnham, UK.
386. Bedford, I.D., Briddon, R.W., Markham, P.G., Brown, J.K. and Rosell, R.C. 1992. A new species of *Bemisia* or biotype of *Bemisia tabaci* (Genn.) as a future pest of European agriculture. Proc. Plant Health and the European Single Market. BCPC Monograph 54: 381-386.
387. Bedford, I.D., Kelly, A., Banks, G.K., Briddon, R.W., Cenis, J.L. and Markham, P.G. 1998. *Solanum nigrum*: An indigenous weed reservoir for a tomato yellow leaf curl geminivirus in southern Spain. Eur. J. Plant Pathol. 104(2): 221-222.
388. Bedford, I.D., Markham, P.G. and Strauss, P.A. 1994. A study of the effectiveness of crop covering within IPM, using an Amoco non-woven fleece as a barrier to aphids, whiteflies and their associated plant viruses. pp. 1163-1168. In Brighton Crop Protection Conference: Pests and Diseases. British Crop Protection Council, Farnham, UK.
389. Bedford, I.D., Pinner, M., Liu, S. and Markham, P.G. 1994. *Bemisia tabaci*: potential infestation, phytotoxicity and virus transmission within European agriculture. pp. 911-916. In Brighton Crop Protection Conference: Pests and Diseases. British Crop Protection Council, Farnham, UK.
390. Beevi, S.P. and Balasubramanian, M. 1991. Effect of buprofezin on adult life span, oviposition, egg hatch and progeny production of the cotton whitefly, *Bemisia tabaci*. Phytoparasitica 19(1): 33-47. [Cock (1993)]
391. Beevi, S.P. and Balasubramanian, M. 1995. Effect of buprofezin, a novel insect growth regulator, against cotton whitefly *Bemisia tabaci* Genn. Entomol. 20(1): 11-14.
392. Beevi, S.P., Natarajan, K. and Balasubramanian, M. 1988. Natural enemies of whitefly, *Bemisia tabaci*, on brinjal. pp. 175-181. In N. Mohandas and G. Koshy (Eds.), Proceedings National Symposium on Integrated Pest Control: Progress and Perspectives, 15-17 October. Association for Advancement of Entomology, Trivandrum, Kerala, India.
393. Beitia, F., Camero, A., Hernandez-Suarez, E., Onillon, J.C. and Guirao, P. 1996. Posibilidades de control biologico de *Bemisia tabaci*: situacion en Canarias. pp. 81-86. In J.L. Cenis (Ed.), El Virus del Rizado Amirillo (Hoja en Cuchara) del Tomate (TYLCV) y Su Vector *Bemisia tabaci*, Jornadas 8. Consejería de Medio Ambiente, Agricultura y Agua, Murcia, Spain.
394. Beitia, F., Mayo, I., Robles-Chillida, E.M., Guirao, P. and Cenis, J.L. 1997. Current status of *Bemisia tabaci* (Gennadius) in Spain: the presence of biotypes of this species. Bull. OILB/SROP (IOBC/WPRS) 20(4): 99-107.
395. Bell, M.L. 1997. Cultural, chemical and physical control for pest management in poinsettias. [North Carolina State University]. Dissertation Abst. Int. 58(03B): 1041.
396. Bell, M.L. and Baker, J.R. 2000. Comparison of greenhouse screening materials for excluding whitefly (Homoptera : Aleyrodidae) and thrips (Thysanoptera : Thripidae). J. Econ. Entomol. 93(3): 800-804.
397. Bellamy, D.E. and Byrne, D.N. 2001. Effects of gender and mating status on self-directed dispersal by the whitefly parasitoid *Eretmocerus eremicus* Ecol. Entomol. 26(6): 571-577.
398. Belli, G. 1974. Le virosi delle piante - come si manifestano, come si trasmettono. Italia Agric. 111(7/8): 71-86. [Cock (1986)]
399. Bello-Mendoza, L.E. 1998. Manejo integrado de plagas en tomate. Bucaramanga, Colombia, 8 pp. [Spanish]
400. Bellotti, A.C. and Arias, B. 2001. Host plant resistance to whiteflies with emphasis on cassava as a case study. In S. E. Naranjo and P. C. Ellsworth (Eds.), Special Issue: Challenges and Opportunities for Pest Management of *Bemisia tabaci* in the New Century. Crop Prot. 20(9): 813-823.
401. Bellows, T.S. and Headrick, D.H. 1995. Hot papers - entomology-description of a species of *Bemisia* (Homoptera: Aleyrodidae) by Bellows, T. S., Perring, T. M., Gill, R. J., Headrick, D. H. Scientist 9(23): 16.
402. Bellows, T.S., Jr. and Arakawa, K. 1986. Modeling the relationship between transient vector densities and plant disease incidence with special reference to *Bemisia tabaci* (Homoptera: Aleyrodidae) and lettuce infectious virus yellows. J. Econ. Entomol. 79(5): 1235-1239. [Cock (1993)]
403. Bellows, T.S., Jr. and Arakawa, K. 1988. Dynamics of preimaginal populations of *Bemisia tabaci* (Homoptera: Aleyrodidae) and *Eretmocerus* sp. (Hymenoptera: Aphelinidae) in southern California cotton. Environ. Entomol. 17(3): 483-487. [Cock (1993)]
404. Bellows, T.S., Jr., Perring, T.M., Arakawa, K. and Farrar, C.A. 1988. Patterns in diel flight activity of *Bemisia tabaci* (Homoptera: Aleyrodidae) in cropping systems in Southern California. Environ. Entomol. 17(2): 225-228. [Cock (1993)]
405. Bellows, T.S., Jr., Perring, T.M., Gill, R.J. and Headrick, D.H. 1994. Description of a species of *Bemisia* (Homoptera: Aleyrodidae). Ann. Entomol. Soc. Am. 87: 195-206.
406. Beltrao, N.E., Vieira, D.J., Azevedo, D.M.P., Nobrega, L.B. and Crisostomo, J.R. 1985. [Sticky cotton; causes, effects, prevention and control]. Documentos, Empresa Brasileira de Pesquisa Agropecuária - Centro Nacional de Pesquisa do Algodão 33: 1-19. [Cock (1993), Portuguese]
407. Bemis, F.E. [no date]. The aleyrodids, or mealy-winged flies, of California, with references to other American species. Proc. U.S. Nat. Museum 27: 471-537. [Cock (1986)]
408. Ben-Ze'ev, I.S. 1993. Check list of fungi pathogenic to insects and mites in Israel, update through 1992. Phytoparasitica 21: 213-237.
409. Ben-Ze'ev, I.S., Gindin, G., Barash, I. and Raccach, B. 1994. Entomopathogenic fungi attacking *Bemisia tabaci* in Israel. Bemisia Newsletter (special issue) 8: 36.
410. Ben-Ze'ev, I.S., Zelig, Y., Bitton, S. and Kenneth, R.G. 1989. The Entomophthorales of Israel and their arthropod hosts: additions 1980-1988. Phytoparasitica 16: 247-257. [Cock (1993)]
411. Bendahmane, M. and Gronenborn, B. 1997. Engineering resistance against tomato yellow leaf curl virus (TYLCV) using antisense RNA. Plant Mol. Biol. 33(2): 351-357.
412. Bene, G.D. 1990. [Biological control of *Trialeurodes vaporariorum* and *Bemisia tabaci*, with *Encarsia formosa* and indigenous parasitoids]. Rev. Agric. Subtropical Trop. 84(2): 237-248. [Cock (1993), Italian, English summary]

413. Bene, G.d., Gargani, E. and Landi, S. 1993. Lotta biologica e integrata contro insetti dannosi alle piante da fiore e ornamentali: risultati preliminari. [Biological and integrated pest control in protected ornamental crops: preliminary results]. *Culture Protette (Italy)* 22(1): 13-18. [ Italian, English summary]
414. Benigno, D.R.A. and Dolores, A.C. 1978. Virus diseases of mungbean in the Philippines. pp. 173-175. *In* R. Cowell (Ed.), 1st International Mungbean Symposium. Asian Vegetable Research and Development Center, Taiwan. [Cock (1986)]
415. Benmessaoud-Boukhalfa, H. 1991. Etude preliminaire de la bioecologie de *Bemisia tabaci* Gen. (Homoptera - Aleyrodidae) en Mitidja (Algerie). [Preliminary studies on bioecology of *Bemisia tabaci* Gen. (Homoptera - Aleyrodidae) in Mitidja (Algeria)]. *Bull. OILB/SROP (IOBC/WPRS)* 14: 98-104. [ French]
416. Benmessaoud-Boukhalfa, H. 1994. Etude preliminaire de la dynamique des populations de *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae) sur une culture protegee de tomate. [Preliminary study of population dynamics of *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae) on protected tomato crops]. *Bull. OILB/SROP (IOBC/WPRS)* 17: 116-119. [ French]
417. Benmessaoud-Boukhalfa, H. 1996. Fluctuations in populations of *Bemisia tabaci* Gen. (Homoptera: Aleyrodidae) on tomatoes under glass in Mitidja (Algeria). *Med. Fac. Landbouww. Univ. Gent* 61(3B): 1025-1029. [ French]
418. Benmessaoud-Boukhalfa, H. 1996. Study on the biological control of *Bemisia tabaci* Gen. (Homoptera: Aleyrodidae) by *Encarsia formosa* Gah. (Hymenoptera: Aphelinidae) on tomatoes of a greenhouse in Mitidja (Algeria). *Med. Fac. Landbouww. Univ. Gent* 61(3B): 1031-1034.
419. Benmessaoud-Boukhalfa, H., Benmessaoud, K. and Belkacem, N. 2000. A comparative study on the spatial and temporal distribution of two whiteflies of vegetable crops [*Trialeurodes vaporariorum* (Westwood) and *Bemisia tabaci* (Gennadius)] (Homoptera-Aleyrodidae) on tomato. *Bull. OILB/SROP (IOBC/WPRS)* 23: 125-131. [ French, English summary]
420. Benmessaoud-Boukhalfa, H., Nenon, J.P. and LeLannic, J. 2000. Wax secretions in *Bemisia tabaci* (Homoptera : Aleyrodidae). Evolution during the larval cycle. *Ann. Soc. Entomol. Fr.* 36(2): 165-170.
421. Benmessaoud-Boukhalfa, H., Sayoud, H. and Keroui, W. 1994. Essai de lutte biologique par l' utilisation d' *Encarsia formosa* Gah (Hymenoptera: Aphelinidae) sur *Bemisia tabaci* Gen. (Homoptera: Aleyrodidae). [Trials of biological control by use of *Encarsia formosa* Gah (Hymenoptera: Aphelinidae) on *Bemisia tabaci* Gen. (Homoptera: Aleyrodidae)]. *Bull. OILB/SROP (IOBC/WPRS)* 17: 110-115. [ French]
422. Benmessaoud-Boukhalfa, H.B. and Benhafid, A. 1998. Greenhouse study on the varietal susceptibility of vegetables to the aleurod *Bemisia tabaci* Gen. in the region of Biskra (south east Algeria). *Med. Fac. Landbouww. Univ. Gent* 63(2a): 385-388.
423. Bennett, F.D. and Alam, M.M. 1985. An annotated check-list of the insects and allied terrestrial arthropods of Barbados. *Caribbean Agric. Res. and Dev. Inst. (Barbados)*. 1-5,10-17,81. [ Cock (1986)]
424. Bennett, F.D. and Noyes, J.S. 1989. Three chalcidoid parasites of diaspines and whiteflies occurring in Florida USA and the Caribbean. *Florida Entomol.* 72: 370-373.
425. Bentz, J.A. 1993. The influence of nitrogen content of poinsettia, *Euphorbia pulcherrima*, on the sweetpotato whitefly, *Bemisia tabaci*, and its parasitoid *Encarsia formosa*. [University of Maryland]. *Dissertation Abst. Int.* 54(10B): 5009.
426. Bentz, J.A. and Neal, J.W., Jr. 1995. Effect of a natural insecticide from *Nicotiana glauca* on the whitefly parasitoid *Encarsia formosa* (Hymenoptera: Aphelinidae). *J. Econ. Entomol.* 88(6): 1611-1615.
427. Bentz, J.A., Reeves, J., Barbosa, P. and Francis, B. 1995. Effect of nitrogen fertilizer source and level on ovipositional choice of poinsettia by *Bemisia argentifolii* (Homoptera: Aleyrodidae). *J. Econ. Entomol.* 88(5): 1388-1392.
428. Bentz, J.A., Reeves, J., Barbosa, P. and Francis, B. 1995. Nitrogen fertilizer effect on selection, acceptance and suitability of *Euphorbia pulcherrima* (Euphorbiaceae) as a host plant to *Bemisia tabaci* (Homoptera, Aleyrodidae). *Environ. Entomol.* 24(1): 40-45.
429. Bentz, J.A., Reeves, J., Barbosa, P. and Francis, B. 1995. Within -plant variation in nitrogen and sugar content of poinsettia and its effects on the oviposition pattern, survival, and development of *Bemisia argentifolii* (Homoptera: Aleyrodidae). *Environ. Entomol.* 24(2): 271-277.
430. Bentz, J.A., Reeves, J., III, Barbosa, P. and Francis, B. 1996. The effect of nitrogen fertilizer applied to *Euphorbia pulcherrima* on the parasitization of *Bemisia argentifolii* by the parasitoid *Encarsia formosa*. *Entomol. Exp. Appl.* 78(1): 105-110.
431. Benuzzi, M. and Mosti, M. 1994. Mirid predators of aleyrodids. *Informatore Fitopatol.* 44(11): 25-30. [ Italian, English summary]
432. Benuzzi, M., Nicoli, G. and Manzaroli, G. 1990. Biological control of *Bemisia tabaci* (Genn.) and *Trialeurodes vaporariorum* (Westw.) by *Encarsia formosa* (Gahan) on poinsettia. *Bull. OILB/SROP (IOBC/WPRS)* 13: 27-31.
433. Benuzzi, M., Nicoli, G., Manzaroli, G. and Bravaccini, F. 1990. [Biological and integrated control in poinsettia]. *Informatore Agrario* 46(46): 77-80. [ Cock (1993), Italian]
434. Ber, R., Navot, N., Zamir, D., Antignus, Y., Cohen, S. and Czosnek, H. 1990. Infection of tomato by the tomato yellow leaf curl virus: susceptibility to infection, symptom development, and accumulation of viral DNA. *Arch. Virol.* 112(3-4): 169-180. [ Cock (1993)]
435. Berdiales, B., Bernal, J.J., Sáez, E., Woudt, B., Beitia, F. and Rodriguez-Cerezo, E. 1999. Occurrence of cucurbit yellow stunting disorder virus (CYSDV) and beet pseudo-yellows virus in cucurbit crops in Spain and transmission of CYSDV by two biotypes of *Bemisia tabaci*. *Eur. J. Plant Pathol.* 105(2): 211-215.
436. Berger, E.W. 1921. Natural enemies of scale insects and whiteflies in Florida. *Quart. Bull. Florida State Plant Board (Gainesville)* 5(3): 141-154. [ Cock (1986)]
437. Bergh, J.C., Perring, T.M. and Le Blanc, J.P.R. 1995. Identification of silverleaf whitefly, *Bemisia argentifolii* Bellows and Perring (Homoptera: Aleyrodidae), in Nova Scotia greenhouses. *Canadian Entomol.* 127(1): 141-142.
438. Berlinger, M.J. 1980. Resistance in tomato to the greenhouse whitefly in relation to integrated control in glasshouses. *Working Group Integrated Control in Glasshouses, Proc. 4th Meeting Bull. S.R.O.P./W.P.R.S.* III: 17-24.
439. Berlinger, M.J. 1980. A yellow sticky trap for whiteflies: *Trialeurodes vaporariorum* and *Bemisia tabaci* (Aleyrodidae). *Entomol. Exp. Appl.* 27: 98-102. [ Cock (1986)]
440. Berlinger, M.J. 1986. Host plant resistance to *Bemisia tabaci*. *Agric. Ecosystems Environ.* 17(1-2): 69-82.
441. Berlinger, M.J. 1986. Pests. pp. 391-441. *In* J.G. Atherton and J. Rudich (Eds.), *The Tomato Crop - A Scientific Basis For Improvement*. Chapman and Hall, London, UK.
442. Berlinger, M.J., Dahan, D. and Mordechi, S. 1988. Integrated pest management of organically grown greenhouse tomatoes in Israel. *Appl. Agric. Res.* 3(5): 233-238. [ Cock (1993)]
443. Berlinger, M.J. and Dahan, R. 1987. Breeding for resistance to virus transmission by whiteflies in tomatoes. [French summary]. *Insect. Sci. Appl.* 8(4-6): 783-784. [ Cock (1993)]
444. Berlinger, M.J. and Dahan, R. 1988. Progress in the study of tomato resistance to its pests, with special emphasis on the tobacco whitefly. *Bull. OILB/SROP (IOBC/WPRS)* 11: 17-19.
445. Berlinger, M.J. and Dahan, R. 1989. *Bemisia tabaci*, the vector of tomato yellow leaf curl virus: a challenge to southern European entomologists. pp. 67-71. *In* R. Cavalloro and C. Pelerents (Eds.), *Proceedings CEC/IBOC Group Meeting, Cabris, 27-29 May 1987*. A. A. Balkema, Rotterdam.

446. Berlinger, M.J. and Dahan, R. 1989. Importance of plant resistance in the control of whiteflies and whitefly-borne viruses in tomato and the development of screening methods. pp. 239-248. In S.K. Green, T.D. Griggs and B.T. McLean (Eds.), Tomato and Pepper Production in the Tropics. Proceedings of the International Symposium on Integrated Management Practices, Tainan, Taiwan, 21-26 March 1988. AVRDC, Shanhuah, Taiwan. [Cock (1993)]
447. Berlinger, M.J., Dahan, R., Berlinger, O.C. and Mordechi, S. 1990. Honeydew excretion as a possible tool to screen tomato resistance to virus transmission by *Bemisia tabaci*. Bull. OILB/SROP (IOBC/WPRS) 13(6): 121-131.
448. Berlinger, M.J., Dahan, R. and Cohen, S. 1983. Greenhouse tomato pests and their control in Israel. Bull. OILB/SROP (IOBC/WPRS) 6(3): 7-11. [Cock (1986)]
449. Berlinger, M.J., Dahan, R. and Mordechi, S. 1986. [Prevention of tomato yellow leaf curl virus by controlling its vector, *Bemisia tabaci*]. Hassadeh 66(4): 686-689. [Cock (1993), Hebrew, English summary]
450. Berlinger, M.J., Dahan, R. and Shevach-Urkin, E. 1983. Breeding for resistance to whiteflies in tomatoes - in relation to integrated pest control in greenhouses. Bull. OILB/SROP (IOBC/WPRS) 6(3): 172-176. [Cock (1986)]
451. Berlinger, M.J., Dahan, R. and Urkin-Shevach, E. 1984. Resistance to the tobacco whitefly, *Bemisia tabaci*, in tomato and related species: a quick screening method. Bull. OILB/SROP (IOBC/WPRS) 7: 39-40.
452. Berlinger, M.J. and De Ponti, O.M.B. 1981. Methods for testing resistance to whiteflies in tomato and related species. Bull. OILB/SROP (IOBC/WPRS) 6(1): 115-118.
453. Berlinger, M.J., Gol'berg, A.M., Dahan, R. and Cohen, S. 1983. The use of plastic covering to prevent the spread of tomato yellow leaf curl virus in greenhouses. Hassadeh 63: 1862-1865. [Hebrew, English summary]
454. Berlinger, M.J. and Lebiush-Mordechi, S. 1996. Physical methods for the control of *Bemisia*. pp. 617-634. In D. Gerling and R.T. Mayer (Eds.), *Bemisia* : 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
455. Berlinger, M.J., Lebiush-Mordechi, S., Dahan, R. and Taylor, R.A.J. 1996. A rapid method for screening insecticides in the laboratory. Pestic. Sci. 46(4): 345-353.
456. Berlinger, M.J., Lehmann-Sigura, N. and Taylor, R.A. 1996. Survival of *Bemisia tabaci* under different climatic conditions. Entomol. Exp. Appl. 80(3): 511-519.
457. Berlinger, M.J., Leibush-Mordechai, S., Fridja, D. and Pilowsky, M. 1993. Breeding tomatoes for whitefly-vector resistance. Bull. OILB/SROP (IOBC/WPRS) 16(5): 83-86.
458. Berlinger, M.J., Magal, Z. and Benzioni, A. 1983. The importance of pH in food selection by the tobacco whitefly, *Bemisia tabaci*. Phytoparasitica 11: 151-160. [Cock (1986)]
459. Berlinger, M.J., Mordechi, S. and Leeper, A. 1991. Application of screens to prevent whitefly penetration into greenhouses in the Mediterranean Basin. Bull. OILB/SROP (IOBC/WPRS) 14(5): 105-110.
460. Berlinger, M.J., Mordechi, S.L., Fridja, D., Chyzik, R., Klein, M., Dov, Y.B. and Aharon, Y. 1993. The development of an IPM programme for greenhouse crops in Israel. Bull. OILB/SROP (IOBC/WPRS) 16(8): 18-21.
461. Berlinger, M.J., Rylski, I., Dahan, R. and Lewisman, P. 1983. Plastic covering to prevent the spread of tomato yellow leaf curl virus by the tobacco whitefly (*Bemisia tabaci*) in the open field. Hassadeh 63: 2090-2094. [Hebrew, English summary]
462. Bernal-Vega, J.A., Neus, O., Eelen, H. and Bulcke, R. 2000. Parasitoids inventory of *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae) in tomato, chilli and four weeds in Panama. Med. Fac. Landbouww. Univ. Gent 65(2a): 293-301.
463. Bernays, E.A. 1999. Plasticity and the problem of choice in food selection. Ann. Entomol. Soc. Am. 92(6): 944-951.
464. Bernays, E.A. 1999. When host choice is a problem for a generalist herbivore: experiments with the whitefly, *Bemisia tabaci*. Ecol. Entomol. 24(3): 260-267.
465. Bernays, E.A. and Minkenberg, O.P.J.M. 1997. Insect herbivores: different reasons for being a generalist. Ecology 78(4): 1157-1169.
466. Berrie, L.C., Palmer, K.E., Rybicki, E.P. and Rey, M.E.C. 1998. Molecular characterisation of a distinct South African cassava infecting geminivirus. Arch. Virol. 143(11): 2253-2260.
467. Bethke, J.A., Nuessly, G.S., Paine, T.D. and Redak, R.A. 1991. Effect of host insect-host plant associations on selected fitness components of *Encarsia foenosa* (Gahan) (Hymenoptera: Aphelinidae). Biol. Control 1(2): 164-169.
468. Bethke, J.A. and Paine, T.D. 1991. Control of the sweetpotato whitefly on poinsettia under laboratory conditions Summer 1989. Insecticide Acaricide Tests 16: 256.
469. Bethke, J.A. and Paine, T.D. 1991. Screen hole size and barriers for exclusion of insect pests of glasshouse crops. J. Entomol. Sci. 26(1): 169-177. [Cock (1993)]
470. Bethke, J.A., Paine, T.D. and Nuessly, G.S. 1991. Comparative biology, morphometrics, and development of two populations of *Bemisia tabaci* (Homoptera: Aleyrodidae) on cotton and poinsettia. Ann. Entomol. Soc. Am. 84(4): 407-411. [Cock (1993)]
471. Bethke, J.A. and Redak, R.A. 1997. Effect of imidacloprid on the silverleaf whitefly, *Bemisia argentifolii* Bellows and Perring (Homoptera: Aleyrodidae) and whitefly parasitism. Ann. Appl. Biol. 130(3): 397-407.
472. Bethke, J.A. and Redak, R.A. 1998. Topical and residual assays of selected pesticides against the silverleaf whitefly, Winter 1997. Arthropod Management Tests 23: 370-371.
473. Bethke, J.A., Redak, R.A. and Paine, T.D. 1994. Screens deny specific pests entry to greenhouses. California Agric. 48: 37-40.
474. Bhagabati, K.N. and Goswami, B.K. 1992. Incidence of yellow vein mosaic virus disease of okra (*Abelmoschus esculentus* L. Moench) in relation to whitefly (*Bemisia tabaci* Genn.) population under different sowing dates. Indian J. Virol. 8(1): 37-39.
475. Bhagabati, K.N. and Mahato, K. 1999. Incidence of yellow mosaic virus disease on soybean under Assam conditions. Legume Res. 22(1): 59-61.
476. Bhagat, A.P., Yadav, B.I. and Prasad, Y. 1997. Management of bhindi yellow vein mosaic virus disease by insecticides. J. Mycol. Plant Pathol. 27(2): 215-216.
477. Bharathan, N., Graves, W.R., Narayanan, K.R., Schuster, D.J., Bryan, H.H. and McMillan, R.T., Jr. 1990. Association of double-stranded RNA with whitefly-mediated silvering in squash. Plant Pathol. 39(3): 530-538. [Cock (1993)]
478. Bharathan, N., Narayanan, K.R. and McMillan, R.T., Jr. 1992. Characteristics of sweetpotato whitefly-mediated silverleaf syndrome and associated double-stranded RNA in squash. Phytopathology 82(2): 136-141. [Cock (1993)]
479. Bhardwaj, S.C. 1992. A note on the transmission of tomato leaf curl virus (TLCV) with *Bemisia tabaci* (Gennadius). Indian J. Entomol. 54(1): 102-104.
480. Bhardwaj, S.C. and Kushwaha, K.S. 1984. Whitefly, *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae) infesting tomato in Rajasthan, I. Bionomics. Bull. Entomol. (New Delhi) 25(1): 76-97. [Cock (1993)]
481. Bhargava, K.S. and Joshi, R.D. 1962. Yellow mosaic, a virus disease of rose in Gorakhpur. Sci. Culture 28: 184-185. [Cock (1986)]
482. Bharpoda, T.M. and Chari, S.M. 1986. Bio-efficacy of some new insecticides against tobacco white-fly, *Bemisia tabaci* Gennadius. Pestology 10: 10-11.
483. Bhatnagar, P. and Sharma, P.D. 1991. Comparative incidence of sucking insects on different isogenic lines of cotton variety H777. J. Insect Sci. 4(2): 170-171.



484. Bhattacharyya, P.K., Ram, H. and Kole, P.C. 1999. Inheritance of resistance to yellow mosaic virus in interspecific crosses of soybean. *Euphytica* 108(3): 157-159.
485. Bhattacharjee, N.S. 1976. Control of the spread of the yellow mosaic virus of 'moong' in soybean. *Entomol. Newsletter* 6(11/12): 64-65. [Cock (1986)]
486. Bhattacharjee, N.S. 1990. *Bemisia tabaci* (Gennadius) on soybean and its control. *Indian J. Entomol.* 52(2): 265-273.
487. Bi, J.L., Ballmer, G.R., Hendrix, D.L., Henneberry, T.J. and Toscano, N.C. 2001. Effect of cotton nitrogen fertilization on *Bemisia argentifolii* populations and honeydew production. *Entomol. Exp. Appl.* 99(1): 25-36.
488. Bi, J.L., Ballmer, G.R., Toscano, N.C. and Madore, M.A. 2000. Effect of nitrogen fertility on cotton-whitefly interactions. pp. 1135-1142. *In* P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
489. Bi, J.L. and Toscano, N.C. 2001. Effect of cotton planting date and nitrogen fertilization on *Bemisia argentifolii* populations. pp. 918-924. *In* P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
490. Bianchini, A. 1999. Resistance to bean golden mosaic virus in bean genotypes. *Plant Dis.* 83(7): 615-620.
491. Bikash, M. and Anupam, V. 1996. Differentiation of natural variants of mungbean yellow mosaic geminivirus by host reactions and DNA-DNA hybridization. *Int. J. Trop. Plant Dis.* 14(2): 189-202.
492. Bikash, M., Anupam, V., Malathi, V.G., Mandal, B. and Varma, A. 1998. Some biological and genomic properties of pigeonpea isolate of mungbean yellow mosaic geminivirus. *Indian Phytopathol.* 51(2): 121-129.
493. Bindra, O.S. 1983. Insect resistance in cotton in Sudan. pp. 227-229. *In* F. Lamberti, J.M. Waller and N.A. van der Graaf (Eds.), *Durable Resistance in Crops*. Plenum Press, New York, USA. [Cock (1986)]
494. Bindra, O.S. 1985. Relation of cotton cultivars to the cotton-pest problem in the Sudan Gezira. *Euphytica* 34(3): 849-856. [Cock (1993)]
495. Bindra, O.S., Sidhu, A.S., Singh, G. and Brar, K.S. 1973. Control of sucking pests of cotton by soil application of granular systemic insecticides. *Indian J. Agric. Sci.* 43: 352-356. [Cock (1986)]
496. Bink, F.A. 1973. Nouvelle contribution a l'etude de la mosaïque du cotonnier au Tchad. I. Symptomes, transmission par *Bemisia tabaci* Genn.; II - Observation sur *B. tabaci*, III. - Autres maladies virales sur cotonniers et plantes voisines. [English edition available]. *Coton Fibres Trop.* 28: 365-378. [Cock (1986)]
497. Bink, F.A. 1975. Leafcurl and mosaic diseases of cotton in central Africa. *Cotton Growing Rev.* 52(3): 233-241. [Cock (1986)]
498. Bink-Moenen, R.M. 1983. Revision of the African whiteflies (Aleyrodidae), mainly based on a collection from Tchad. *Nederlandse Entomologische Vereniging (Amsterdam, Netherlands)*. 5-21,94-106,201-211. [Cock (1986)]
499. Bink-Moenen, R.M. and Mound, L.A. 1990. Whiteflies diversity biosystematics and evolutionary patterns. pp. 1-12. *In* D. Gerling (Ed.), *Whiteflies: their Bionomics, Pest Status, and Management*. Intercept, Andover, UK.
500. Biradar, A.P., Sunitha, N.D. and Jagginavar, S.B. 1999. Survey for major pests of cotton. *Insect Environ.* 5(2): 76.
501. Bird, J. 1957. A whitefly-transmitted mosaic of *Jatropha gossypifolia*. *Tech. Papers, Agric. Exp. Stn. (Puerto Rico)* 22: 5-35. [Cock (1986)]
502. Bird, J. 1958. Infectious chlorosis of *Sida carpinifolia* in Puerto Rico. *Tech. Papers, Agric. Exp. Stn. (Puerto Rico)* 26: 5-23. [Cock (1986)]
503. Bird, J., Idris, A.M., Rogan, D. and Brown, J.K. 2001. Introduction of the exotic tomato yellow leaf curl virus-Israel in tomato into Puerto Rico. *Plant Dis.* 85: 1028.
504. Bird, J. and Maramorosch, K. 1978. Viruses and virus diseases associated with whiteflies. pp. 55-110. *In* M.A. Lauffer, F.B. Bang, K. Maramorosch and K.M. Smith (Eds.), *Advances in virus research*. Academic Press, London, UK. [Cock (1986)]
505. Bird, J. and Maramorosch, K. (Eds.). 1975. *Tropical Diseases of Legumes*. Academic Press, New York, 171 pp. [Cock (1986)]
506. Bird, J., Perez, J.R., Alconero, R., Vakili, N.G. and Melendez, P.L. 1972. A whitefly-transmitted golden-yellow mosaic virus of *Phaseolus lunatus* in Puerto Rico. *J. Agric. (Univ. Puerto Rico)* 56(1): 64-74. [Cock (1986)]
507. Bird, J. and Sanchez, J. 1971. Whitefly-transmitted viruses in Puerto Rico. *J. Agric. (Univ. Puerto Rico)* 55: 461-467. [Cock (1986)]
508. Bird, J., Sanchez, J., Rodriguez, R.L. and Julia, F.J. 1975. Rugaceous (whitefly-transmitted) viruses in Puerto Rico. pp. 3-25. *In* J. Bird and K. Maramorosch (Eds.), *Tropical Diseases of Legumes*. Academic Press, New York, USA. [Cock (1986)]
509. Birnie, L.C. and Denholm, I. 1992. Field simulators: a novel approach to evaluating the impact of pesticides on beneficial arthropods in the laboratory. *Aspects Appl. Biol.* 31: 105-112.
510. Birnie, L.C. and Denholm, I. 1992. Use of field simulators to investigate integrated chemical and biological control tactics against the cotton whitefly, *Bemisia tabaci*. pp. 1003-1008. *In* Brighton Crop Protection Conference: Pests and Diseases. The British Crop Protection Council, Farnham, UK.
511. Bisaro, D.M., Hamilton, W.D.O., Coutts, R.H.A. and Buck, K.W. 1982. Molecular cloning and characterisation of the two DNA components of tomato golden mosaic virus. *Nucleic Acids Res.* 10(16): 4913-4922. [Cock (1986)]
512. Bisaro, D.M., Sunter, G., Revington, G.N., Brough, C.L., Homuzdi, S.G. and Haritiz, M. 1990. Molecular genetics of tomato golden mosaic virus replication: progress toward defining gene functions, transcription units and the origin of dna replication. pp. 89-105. *In* T.P. Pirone and J.G. Shaw (Eds.), *Viral Genes and Plant Pathogenesis*. Springer-Verlag, New York, USA.
513. Bisheya, F.A. 1997. Libya. pp. 41-42. *In* N. Ioannou (Ed.), *Management of the whitefly-virus complex*. FAO Plant Production and Protection Paper 143, Rome, Italy.
514. Bisht, N.S. and Banerjee, A.K. 1965. Occurrence of two new virus diseases in Uttar Pradesh. *Labdev J. Sci. Technol.* 3(4): 271-272. [Cock (1986)]
515. Bisht, N.S. and Mathur, R.S. 1964. Occurrence of two strains of jute mosaic virus in Uttar Pradesh. *Curr. Sci. (Bangalore)* 33: 434-435. [Cock (1986)]
516. Biswas, A.K. and Mandal, S.K. 1992. Occurrence of insect pests in different types of tobacco cultivated in West Bengal. *Crop Res. (Hisar)* 5(1): 169-171. [Cock (1993)]
517. Biswas, B. and Ghosh, L.K. 1994. Insecta: Hemiptera: Aleyrodidae. pp. 113-124. *In* A.K. Ghosh (Ed.), *Fauna of West Bengal. Part -5 (Insecta: Hemiptera)*. Zoological Survey of India, Calcutta, India
518. Blackman, R.L. and Cahill, M. 1998. The karyotype of *Bemisia tabaci* (Hemiptera: Aleyrodidae). *Bull. Entomol. Res.* 88(2): 213-215.
519. Blackmer, J.L. and Byrne, D.N. 1993. Environmental and physiological factors influencing phototactic flight of *Bemisia tabaci*. *Physiol. Entomol.* 18: 336-342.
520. Blackmer, J.L. and Byrne, D.N. 1993. Flight behaviour of *Bemisia tabaci* in a vertical flight chamber: effect of time of day, sex, age and host quality. *Physiol. Entomol.* 18(3): 223-232.
521. Blackmer, J.L. and Byrne, D.N. 1999. Changes in amino acids in *Cucumis melo* in relation to life-history traits and flight propensity of *Bemisia tabaci*. *Entomol. Exp. Appl.* 93(1): 29-40.
522. Blackmer, J.L. and Byrne, D.N. 1999. The effect of *Bemisia tabaci* on amino acid balance in *Cucumis melo*. *Entomol. Exp. Appl.* 93(3): 315-319.

523. Blackmer, J.L., Byrne, D.N. and Tu, Z. 1995. Behavioral, morphological, physiological traits associated with migratory *Bemisia tabaci* (Homoptera: Aleyrodidae). *J. Insect Behav.* 8(2): 251-267.
524. Blackmer, J.L. and Cross, D. 2001. Response of *Eretmocerus eremicus* to skylight and plant cues in a vertical flight chamber. *Entomol. Exp. Appl.* 100(3): 295-300.
525. Blackmer, J.L., Lindley, V.A. and Byrne, D.N. 1995. Histological examination of flight muscle development and breakdown in *Bemisia tabaci* (Homoptera: Aleyrodidae): Relationship to age and flight behavior. *J. Morphol.* 226(2): 213-221.
526. Blair, M.W., Bassett, M.J., Abouzid, A.M., Hiebert, E., Polston, J.E., McMillan, R.T., Jr., Graves, W. and Lamberts, M. 1995. Occurrence of bean golden mosaic virus in Florida. *Plant Dis.* 79(5): 529-533.
527. Blair, M.W. and Beaver, J.S. 1992. Resistance to the sweetpotato whitefly (*Bemisia tabaci*), the vector of bean golden mosaic virus in dry beans (*Phaseolus vulgaris*). *Annu. Rep. Bean Improvement Coop.* 35: 154-155.
528. Blair, M.W. and Beaver, J.S. 1993. Sweetpotato whitefly preference differs among Mesoamerican and Aedeon gene pools of common bean (*Phaseolus vulgaris* L.). *Annu. Rep. Bean Improvement Coop.* 36: 132-134.
529. Blanchard, E.E. 1937. Informaciones técnicas. Boletín Informativo Dirección Sanidad Vegetal (Buenos Aires) 1: 25-32. [Cock (1993)]
530. Blanco, J. and Hilje, L. 1995. Efecto de coberturas al suelo sobre la abundancia de *Bemisia tabaci* y la incidencia de virosis en tomate. [The effect of soil covers on the abundance of *Bemisia tabaci* and virus incidence in tomatoes]. *Manejo Integrado de Plagas (Costa Rica)* 35: 1-10. [Spanish, English summary]
531. Blanco, N. and Fature, B. 1994. Situación actual del mosaico dorado del frijol en el Caribe: Cuba. [Current situation of bean golden mosaic virus in Latin America: Cuba]. pp. 82-89. *In* F.J. Morales (Ed.), *Bean Golden Mosaic: Research Advances*. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. [Spanish]
532. Blanco-Sanchez, N. and Bencomo, I. 1978. Afluencia de la mosca blanca (*Bemisia tabaci*), vector del virus del mosaico dorado, en plantaciones de frijol. *Ciencias Agric.* 2: 39-46. [Cock (1986)]
533. Blanco-Sanchez, N. and Bencomo-Pérez, I. 1981. Presencia del virus del mosaico dorado del frijol (BGMV) en Cuba. *Ciencias Agric.* 9: 118. [Cock (1986)]
534. Blaney, W.M., Simmonds, M.S.J., Ley, S.V., Anderson, J.C. and Toogood, P.L. 1990. Antifeedant effects of azadirachtin and structurally related compounds on lepidopterous larvae. *Entomol. Exp. Appl.* 55(2): 149-160.
535. Bloch, G. and Wool, D. 1994. Methidathion resistance in the sweetpotato whitefly (Aleyrodidae: Homoptera) in Israel: Selection, heritability, and correlated changes of esterase activity. *J. Econ. Entomol.* 87(5): 1147-1156.
536. Bloch, G. and Wool, D. 1995. Esterase activity in populations of the whitefly, *Bemisia tabaci* (Homoptera: Aleyrodidae): Heritability and associated organophosphorus insecticide resistance. *Bull. Entomol. Res.* 85(1): 11-19.
537. Blua, M.J., Perring, T.M., Nuessly, G.S., Duffus, J.E. and Toscano, N.C. 1994. Seasonal cropping pattern effects on abundance of *Bemisia tabaci* (Homoptera: Aleyrodidae) and incidence of lettuce infectious yellows virus. *Environ. Entomol.* 23(6): 1422-1427.
538. Blua, M.J. and Toscano, N.C. 1994. *Bemisia argentifolii* (Homoptera, Aleyrodidae) development and honeydew production as a function of cotton nitrogen status. *Environ. Entomol.* 23(2): 316-321.
539. Blua, M.J., Toscano, N.C. and Henneberry, T.J. 1993. Effects of cotton nitrogen status on sweetpotato whitefly development and honeydew production. pp. 706-709. *In* D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
540. Blua, M. J., Yoshida, H.A. and Toscano, N.C. 1995. Oviposition preference of two *Bemisia* species (Homoptera: Aleyrodidae). *Environ. Entomol.* 24(1): 88-93.
541. Blumel, S. 1990. *Bemisia tabaci* (Genn.), the 'new' whitefly. now also in Austria. *Pflanzenschutz (Wien)* 2: 4-5.
542. Bob, M.A., Sithanatham, S. and Womdim, R.N. 1997. Whitefly problems in Africa: known importance and research needs. pp. 859-866. *In* Proceedings ANPP 4th International Conference on Pests in Agriculture, 6-9 Jan. 1997, Montpellier, France. ANPP, Paris.
543. Bock, K. 1983. Epidemiology of cassava mosaic disease in Kenya. pp. 337-347. *In* R.T. Plumb and J.M. Thresh (Eds.), *Plant Virus Epidemiology. The Spread and Control of Insect-borne Viruses*. Blackwell Scientific Publications, Oxford, UK. [Cock (1986)]
544. Bock, K.R. 1982. Geminivirus diseases in tropical crops. *Plant Dis.* 66: 266-270.
545. Bock, K.R. and Guthrie, E.J. 1978. African mosaic disease in Kenya. pp. 41-44. *In* T. Brekelbaum, A. Bellotti and J.C. Lozano (Eds.), *Proc. Cassava Protection Workshop CIAT, Cali, Columbia, November 7-12, 1977*. Centro Internacional de Agricultura Tropical, Cali, Colombia. [Cock (1986)]
546. Bock, K.R., Guthrie, E.J. and Meredith, G. 1978. Distribution, host range, properties and purification of cassava latent virus, a geminivirus. *Ann. Appl. Biol.* 90: 361-367. [Cock (1986)]
547. Bodnar, G. 1928. Alyrodideos do Brasil (2a Contribuicao). *Boletim Laboratoria Pathologia Vegetal, Estado de Bahia* 5: 1-37.
548. Bogran, C.E., Heinz, K.M. and Ciomperlik, M.A. 2002. Interspecific competition among insect parasitoids: Field experiments. *Ecology* 83(3): 653-668.
549. Bogran, C.E., Obrycki, J.J. and Cave, R. 1998. Assessment of biological control of *Bemisia tabaci* (Homoptera: Aleyrodidae) on common bean in Honduras. *Florida Entomol.* 81(3): 384-395.
550. Bohmer, B. 1989. [*Bemisia* and *Trialeurodes*. Two whitefly genera that damage euphorbias]. *Gärtnerbörse und Gartenwelt (Germany)* 89(28): 1362. [Cock (1993), German]
551. Boiça, A.L., Jr., Bolonhezi, A.C. and Paccini Neto, J. 1984. Levantamento de insetos-pragas e seus inimigos naturais em girassol (*Helianthus annuus* L.), cultivado em primeira e segunda época no Município de Selviria-MS. *Anais Soc. Entomol. Brasil* 13(2): 189-196. [Cock (1986)]
552. Boiça, A.L., Jr., Lopes, P.R., Silva, H.S., Fernandes, S.H.C., Costa, M.C. and Malimpence, R.A. 1991. Utilização de produtos químicos no controle de *Bemisia tabaci* (Gennadius, 1889), *Empoasca* spp. e *Thrips* spp., na cultura do feijão. [Efficiency of insecticides in the control of *Bemisia tabaci* (Gennadius, 1889), *Empoasca* spp. and *Thrips* spp. in bean crop (*Phaseolus vulgaris* L.)]. *Poliagro (Brazil)* 10(1): 1-7. [Portuguese, English summary]
553. Boiça, A.L., Jr. and Vendramim, J.D. 1986. Desenvolvimento de *Bemisia tabaci* (Gennadius, 1889) Homoptera, Aleyrodidae em genótipos de feijão (*Phaseolus vulgaris* L.). [Development of *Bemisia tabaci* (Gennadius, 1889) (Homoptera, Aleyrodidae) in genotypes of bean (*Phaseolus vulgaris* L.)]. *Anais Soc. Entomol. Brasil* 15(2): 231-238. [Cock (1993), Portuguese, English summary]
554. Boisclair, J., Brueren, G.J. and van Lenteren, J.C. 1990. Can *Bemisia tabaci* be controlled with *Encarsia formosa*? *Bull. OILB/SROP (IOBC/WPRS)* 13: 32-35. [DeBarro (1995)]
555. Boissot, N., Pavis, C., Guillaume, R., Lafortune, D. and Sauvion, N. 2000. Insect resistance in Cucumis melo accession 90625. *Acta Hort.* 510: 297-304.
556. Bolano, R.E. 1997. Determinación de niveles de dano económico de *Bemisia tabaci* de tomate en el norte del Cesar, Columbia. [*Bemisia tabaci* economic injury level in tomato crops in the northern area of Cesar, Columbia]. *Manejo Integrado de Plagas (Costa Rica)* 46: 26-33. [Spanish, English summary]
557. Bolaños, A. 1996. Germoplasma. pp. 42-50. *In* L. Hilje (Ed.), *Metodologías para el estudio y manejo de moscas blancas y geminivirus*. Centro Agronómico Tropical de Investigación y Enseñanza, Turrialba, Costa Rica.

558. Bolckmans, G., Sterk, J., Eyal, J., Sels, B. and Stepman, W. 1995. PreFeRal, (*Paecilomyces fumosoroseus* strain Apopka 97), a new microbial insecticide for the biological control of whiteflies in greenhouses. Med. Fac. Landbouww. Univ. Gent 60: 707-711.
559. Bond, W.E. 1945. Progress Reports from Experiment Stations season 1943-44. pp. 154-161. In Empire Cotton Growing Corp., London. [Cock (1986)]
560. Bondar, G. 1923. Aleyrodideos do Brasil. Bahia Secretaria da Agric., Industria e Obras Publicas: Seccao de Patologia Vegetal, 183 pp. [Cock (1986)]
561. Bondar, G. 1928. Aleyrodideos do Brasil (2a contribuicao). Bull. Lab. Pathol. Vegetal Estado Bahia 5: 37 Pp. [Cock (1986)]
562. Bonilla, S.F. 1995. Periodos de adquisicion, latencia y transmision de un geminivirus en tomate, por la mosca blanca *Bemisia tabaci* (Gennadius) en Costa Rica. [Acquisition, latency and transmission periods for a tomato geminivirus transmitted by the whitefly, *Bemisia tabaci* (Gennadius) in Costa Rica]. Manejo Integrado de Plagas (Costa Rica) 35: 11-13. [Spanish, English summary]
563. Borad, V.K. and Puri, S.N. 1991. Life table studies of *Bemisia tabaci* (Gennadius) on cotton and brinjal under field conditions by artificial infestation. Indian J. Ecol. 18: 186-189.
564. Borad, V.K. and Puri, S.N. 1993. Some field studies on behaviour of whitefly. J. Maharashtra Agric. Univ. (India) 18(1): 101-103.
565. Borad, V.K. and Puri, S.N. 1995. Dispersal flight activity of *Bemisia tabaci* (Gennadius) in relation to time of the day and weather parameters. Gujarat Agric. Univ. Res. J. 21(1): 64-68.
566. Borad, V.K., Puri, S.N., Brown, J.K. and Butler, G.D., Jr. 1992. Seasonal monitoring of sweetpotato whitefly, *Bemisia tabaci* Genn. and relationship to incidence of leaf curl disease in tomato. J. Appl. Zool. Res. 3(2): 124-127.
567. Borad, V.K., Puri, S.N., Brown, J.K. and Butler, G.D., Jr. 1993. Relationship of *Bemisia tabaci* population density and yellow vein mosaic disease incidence in okra. Pest Manag. Econ. Zool. 1(1): 14-19.
568. Borad, V.K., Puri, S.N., Butler, G.D., Jr. and Henneberry, T.J. 1992. Biology of sweetpotato whitefly, *Bemisia tabaci* (Genn.) on different plant hosts. J. Appl. Zool. Res. 3(2): 118-123.
569. Borah, R.K. 1994. Incidence of insect pests in cucumber, *Cucumis sativa* L., in the hill zone of Assam. Bull. Entomol. (New Delhi) 36(1/2): 120-122.
570. Borah, R.K. 1994. Influence of planting dates on the incidence of insect pests of brinjal (*Solanum melongena* L.) in a hilly area of Assam. J. Agric. Sci. Soc. North East India 7(2): 209-211.
571. Borah, R.K. 1995. Effect of insecticides on pest incidence in summer greengram (*Phaseolus radiatus*). Indian J. Agric. Sci. 65(12): 913-915.
572. Borah, R.K. 1995. Effect of synthetic pyrethroids and organophosphorus insecticides on the incidence of whitefly, *Bemisia tabaci* (Genn.) and yellow mosaic virus in green gram, *Vigna radiata* (L.) Wilczek. Indian J. Virol. 11(1): 75-76.
573. Borah, R.K. 1995. Incidence of insect pests in cotton (*Gossypium hirsutum* L.) in the hill zone of Assam. Ann. Agric. Res. 16(2): 218-219.
574. Borah, R.K. 1995. Insect pest complex in brinjal (*Solanum melongena* L.). Ann. Agric. Res. 16(1): 93-94.
575. Borah, R.K. 1995. Insect pests complex in summer greengram (*Vigna radiata* L.). Ann. Agric. Res. 16(1): 91-92.
576. Borah, R.K. and Bordoloi, D.K. 1998. Influence of planting time on the incidence of leaf curl virus disease and white fly population on tomato. Indian J. Virol. 14(1): 71-73.
577. Borah, R.K. and Nath, P.D. 1995. Evaluation of insecticide schedule on the incidence of whitefly, *Bemisia tabaci* (Genn.) and yellow vein mosaic in okra. Indian J. Virol. 11(2): 65-67.
578. Borah, R.K., Nath, P.D. and Deka, N. 1996. Effect of insecticides and crop trap on the incidence of white fly *Bemisia tabaci* (Genn.) and yellow mosaic virus in greengram *Vigna radiata* (L.) Wilczek. Indian J. Virol. 12(1): 75-77.
579. Bortoli, S.A. and Giacomini, P.L. 1981. Acao de alguns inseticidas granulados sistemicos sobre *Bemisia tabaci* (Gennadius, 1889) (Homoptera: Aleyrodidae) e *Empoasca kraemeri* Ross & Moore, 1957 (Homoptera-Cicadellidae) e seus efeitos na produtividade do feijoeiro (*Phaseolus vulgaris* L.). Anais Soc. Entomol. Brasil 10: 97-104. [Cock (1986), Portuguese, English summary]
580. Bos, L. 1986. Importance of ecological studies in plant virus research. [Arabic summary]. Arab J. Plant Prot. 4(1): 70-75. [Cock (1993)]
581. Bosco, D. and Caciagli, P. 1998. Bionomics and ecology of *Bemisia tabaci* (Sternorrhyncha: Aleyrodidae) in Italy. Eur. J. Entomol. 95(4): 519-527.
582. Bosco, D., Caciagli, P. and Noris, E. 1993. Indagini epidemiologiche sul virus dell' accartocciamento fogliare giallo del pomodoro (TYLC) in Italia. [Epidemiological studies on tomato yellow leaf curl geminivirus (TYLV) in Italy]. Informatore Fitopatol. 43(11): 33-36. [Italian, English summary]
583. Boulehya, S., Najjar, A., Sghairi, R. and Jarraya, A. 1997. Tunisia. pp. 71-75. In N. Ioannou (Ed.), Management of the whitefly-virus complex. FAO Plant Production and Protection Paper 143, Rome, Italy.
584. Boulton, M.I., King, D.I., Donson, J. and Davies, J.W. 1991. Point substitutions in a promoter-like region and the V1 gene affect the host range and symptoms of maize streak virus. Virology 183: 114-121.
585. Boulton, M.I., King, D.I., Markham, P.G., Pinner, M.S. and Davies, J.W. 1991. Host range and symptoms are determined by specific domains of the maize streak virus genome. Virology 181: 312-318.
586. Boulton, M.I., Steinkellner, H., Donson, J., Markham, P.G., King, D.I. and Davies, M.J.W. 1989. Mutational analysis of the viron-sense genes of maize streak virus. J. Gen. Virol. 70: 2309-2323.
587. Bourelly, J., Gutknecht, J. and Fournier, J. 1984. Chemical analysis of stickiness in cotton fiber. Part 1: Role of sugars and honeydews in the process of stickiness. Coton Fibres Trop. 39: 47-53. [French]
588. Bourgeois, H. 1998. Dossier ravageur. Cultures legumieres: coup d'oeil sur cinq ravageurs. [Pests file. Vegetables growing: a glance on five pests]. PHM Rev. Hort. (France) 390: 19-23. [French]
589. Bouriquet, G. 1938. Note concernant les maladies des plantes cultivees a la Reunion. Rev. Agric. l'Ile Reunion (N.S.) 43: 33-38. [Cock (1986)]
590. Bower, C.C. 1995. Poinsettia whitefly (*Bemisia tabaci* type B). Agriculture, Orange, New South Wales, Australia, 2 pp.
591. Braasch, D. and Nussbaum, P. 1992. Experiences in the recognition, control and eradication of the glasshouse whitefly (*Bemisia tabaci* Gennadius) in Eastern Germany. Nachrichtenblatt des Deutschen Pflanzenschutzdienstes 44: 9-11.
592. Brader, L. 1981. Resistance horizontal aux ravageurs et maladies des vegetaux. FAO, Rome, Italy, 10pp. [Cock (1986)]
593. Brar, D.S., Sohi, A.S., Singh, J. and Singh, J. 1999. Efficacy of insecticides against *Amrasca biguttula* (Ishida) and *Bemisia tabaci* (Gennadius) on Hirsutum cotton. Insect Environ. 5(2): 83.
594. Brar, J.S. and Rataul, H.S. 1987. Evidence against the transmission of urd bean leaf crinkle virus (ULCV) in mash bean, *Vignamungo* (L.) through insects - laboratory studies. Indian J. Entomol. 49(1): 69-72. [Cock (1993)]
595. Brar, J.S. and Ratul, H.S. 1987. Evidence against the transmission of urd bean leaf crinkle virus (ULCV) in mash, *Vignamungo* (L.) through insects - a field approach. Indian J. Entomol. 49(1): 57-63. [Cock (1993)]
596. Brasch, K., van Lenteren, J.C., Boisclair, J. and Henter, H. 1994. Biological control of *Bemisia tabaci* with *Encarsia formosa*: A realistic option? Med. Fac. Landbouww. Univ. Gent 59(2A): 325-332.

597. Brazzle, J. R., Fien, B., Goodell, P., Toscano, N. and Godfrey, L. 1998. Whitefly management in the San Joaquin Valley. pp. 73-76. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
598. Brazzle, J.R., Heinz, K.M. and Parrella, M.P. 1997. Multivariate approach to identifying patterns of *Bemisia argentifolii* (Homoptera: Aleyrodidae) infesting cotton. *Environ. Entomol.* 26(5): 995-1003.
599. Brazzle, J.R., Heinz, K.M., Parrella, M.P., Pickett, C. and Natwick, E.T. 1993. Biological control in the Imperial Valley : sweet potato whitefly vs. *Delphastus pusillus*. *Valley Grower (Winter)*: 5-7.
600. Breene, R.G., Dean, D.A. and Quarles, W. 1994. Predators of sweetpotato whitefly. *IPM Practitioner* 16(8): 1-9.
601. Breene, R.G., Meagher, R.L., Jr., Nordlund, D.A. and Wang, Y.T. 1992. Biological control of *Bemisia tabaci* (Homoptera: Aleyrodidae) in a greenhouse using *Chrysopa rufalibris* (Neuroptera: Chrysopidae). *Biol. Control* 2(1): 9-14.
602. Brettell, J.H. 1966. Eleven years work in Abyan (South Arabia) by entomologists of the Empire Cotton Growing Corp. *Empire Cotton Growing Rev.* 43: 286-295. [Cock (1986)]
603. Brettell, J.H. and Jowah, P. 1988. Entomology. pp. 133-151. *In* Annu. Rep. Cotton Res. Inst. (Zimbabwe) 1985/86. Zimbabwe Government Printer, Harare
604. Brettell, J.H., Jowah, P. and Brennan, M.J. 1990. Entomology. pp. 147, 162-175. *In* Annu. Rep. Cotton Res. Inst. (Zimbabwe) 1987/88. Zimbabwe Government Printer, Harare
605. Brettell, J.H., Musana, A.C.Z. and Jowah, P. 1982. Entomology. pp. 177-196. *In* Annu. Rep. Cotton Res. Inst. (Zimbabwe) 1980/81. Zimbabwe Government Printer, Harare
606. Brettell, J.H., Musana, A.C.Z. and Jowah, P. 1983. Entomology. pp. 113-118. *In* Annu. Rep. Cotton Res. Inst. (Zimbabwe) 1980/81. Zimbabwe Government Printer, Harare
607. Brettell, J.H., Musana, A.C.Z. and Jowah, P. 1984. Entomology. pp. 120-125. *In* Annu. Rep. Cotton Res. Inst. (Zimbabwe) 1982/83. Zimbabwe Government Printer, Harare
608. Brettell, J.H., Musana, A.C.Z. and Jowah, P. 1985. Entomology. pp. 121-186, 194,195. *In* Annu. Rep. Cotton Res. Inst. (Zimbabwe) 1983/84. Zimbabwe Government Printer, Harare
609. Brettell, J.H., Musana, A.C.Z. and Jowah, P. 1986. Entomology. pp. 101-166. *In* Annu. Rep. Cotton Res. Instit. (Zimbabwe) 1984/85. Zimbabwe Government Printer, Harare [Cock (1993)]
610. Brewster, C.C. 1996. Spatiotemporal dynamics of insects in large-scale agricultural systems: Methodology and analysis with the silverleaf whitefly, (Bellows and Perring). [University of Florida]. Dissertation Abst. Int. 58(07B): 3434.
611. Brewster, C.C. and Allen, J.C. 1997. Spatiotemporal model for studying insect dynamics in large-scale cropping systems. *Environ. Entomol* 26(3): 473-482.
612. Brewster, C.C., Allen, J.C. and Kopp, D.D. 1999. IPM from space: using satellite imagery to construct regional crop maps for studying crop-insect interaction. *Am. Entomol.* 45(2): 105-117.
613. Brewster, C.C., Allen, J.C., Schuster, D.J. and Stansly, P.A. 1997. Simulating the dynamics of *Bemisia argentifolii* (Homoptera: Aleyrodidae) in an organic cropping system with a spatiotemporal model. *Environ. Entomol.* 26(3): 603-616.
614. Briddon, R.W., Bedford, I.D., Tsai, J.H. and Markham, P.G. 1996. Analysis of the nucleotide sequence of the treehopper-transmitted geminivirus, tomato pseudo-curly top virus, suggests a recombinant origin. *Virology* 219(2): 387-394.
615. Briddon, R.W., Lunness, P., Chamberlin, L.C.L., Pinner, M.S., Brundish, H. and Markham, P.G. 1992. The nucleotide sequence of an infectious insect-transmissible clone of the geminivirus Panicum streak virus. *J. Gen. Virol.* 73: 1041-1047.
616. Briddon, R.W., Mansoor, S., Bedford, I.D., Pinner, M.S. and Markham, P.G. 2000. Clones of cotton leaf curl geminivirus induce symptoms atypical of cotton leaf curl disease. *Virus Genes* 20(1): 19-26.
617. Briddon, R.W. and Markham, P.G. 2000. Cotton leaf curl virus disease. *Virus Res.* 71(1/2): 151-159.
618. Briddon, R.W., Pinner, M.S., Standley, J. and Markham, P.G. 1990. Geminivirus coat protein gene replacement alters insect specificity. *Virology* 177: 85-94.
619. Briddon, R.W., Watts, J., Markham, P.G. and Stanley, J. 1989. The coat protein of beet curly top virus is essential for infectivity. *Virology* 172: 628-633.
620. Briggs, C.J. and Collier, T.R. 2001. Autoparasitism, interference, and parasitoid-pest population dynamics. *Theor. Popul. Biol.* 60(1): 33-57.
621. Broadbent, A.B., Footitt, R.G. and Murphy, G.D. 1989. Sweetpotato whitefly *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae), a potential insect pest in Canada. *Canadian Entomol.* 121(11): 1027-1028. [Cock (1993)]
622. Broadway, R.M., Gongora, C., Kain, W.C., Sanderson, J.P., Monroy, J.A., Bennett, K.C., Warner, J.B. and Hoffmann, M.P. 1998. Novel chitinolytic enzymes with biological activity against herbivorous insects. *J. Chem. Ecol.* 24(6): 985-998.
623. Broughton, R.M., Jr. and Wallace, R.W. 1998. A rapid test for honeydew contamination using the clintest reagent. pp. 1544-1547. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
624. Brown, J.K. 1984. Whitefly-transmitted viruses of the southwest [University of Arizona]. Dissertation Abst. Int. 46(02B): 375.
625. Brown, J.K. 1990. An update on the whitefly-transmitted geminiviruses in the Americas and the Caribbean Basin. *FAO Plant Prot. Bull.* 39(1): 5-23.
626. Brown, J.K. 1992. Biotypes of the sweetpotato whitefly: a current perspective. pp. 665-670. *In* D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
627. Brown, J.K. 1992. Virus diseases of cotton. pp. 275-330. *In* R.J. Hillocks (Ed.), Cotton Diseases. Commonwealth Agricultural Bureaux, Oxon, UK.
628. Brown, J.K. 1993. A critical assessment of biotypes of the sweetpotato whitefly in the Americas and adjacent locales from 1989-1992. pp. 1-9. *In* L. Hilje and O. Arboleda (Eds.), Proceedings Taller Centro Americano y Del Caribe Sobre Mosca Blancas. Turrialba, Costa Rica, August 3-5, 1992. CATIE, Turrialba, Costa Rica. [Spanish, English summary]
629. Brown, J.K. 1994. Current status of *Bemisia tabaci* as a plant pest and virus vector in agro-ecosystems worldwide. *FAO Plant Prot. Bull.* 42(1/2): 3-32. [English and French]
630. Brown, J.K. 1995. The biology and molecular epidemiology of the Geminiviridae Subgroup III. *Plant-Microbe Interactions Review Series*. Chapman and Hall, London.
631. Brown, J.K. 1997. Global diversity and distribution of cotton-infecting geminiviruses: an essential requisite to developing sustainable disease resistance. pp. 39-49. *In* International Cotton Advisory Committee, 56th Plenary Meeting, Paraguay.
632. Brown, J.K. 1997. Whitefly-transmitted geminiviruses of tomato and phylogenetic relationships predicted from virus coat protein gene sequences. pp. 140-144. *In* Proc. 1st International Symposium on Tropical Tomato Diseases. American Society Horticultural Science.
633. Brown, J.K. 1998. Diversity and global distribution of whitefly-transmitted geminiviruses of cotton. pp. 587-598. *In* Cotton, A College of Agriculture Report, Series P-112. University of Arizona, Tucson.
634. Brown, J.K. 1998. Global diversity and distribution of whitefly-transmitted geminiviruses of cotton. pp. 155-161. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
635. Brown, J.K. 2000. Molecular markers for the identification and global tracking of whitefly vector-Begomovirus complexes. *Virus Res.* 71(1/2): 233-260.

636. Brown, J.K. 2001. The molecular epidemiology of begomoviruses. pp. 279-315. In J.A. Khan and J. Dykstra (Eds.), Trends in Plant Virology: Plant Viruses as Molecular Pathogens. The Haworth Press, Inc., New York, NY.
637. Brown, J.K. and Bird, J. 1992. Whitefly-transmitted geminiviruses and associated disorders in the Americas and the Caribbean Basin. *Plant Dis.* 76(3): 220-225. [Cock (1993)]
638. Brown, J.K. and Bird, J. 1995. Variability within the *Bemisia tabaci* species complex and its relation to new epidemics caused by geminiviruses. *CEIBA* 36(1): 73-80.
639. Brown, J.K. and Bird, J. 1996. Introduction of an exotic whitefly (*Bemisia*) vector facilitates secondary spread of jatropha mosaic virus, a geminivirus previously vectored exclusively by the "jatropha" biotype. pp. 351-353. In D. Gerling and R.T. Mayer (Eds.), *Bemisia: 1995 Taxonomy, Biology, Damage, Control and Management*. Intercept Ltd., Andover, Hants, UK.
640. Brown, J.K., Bird, J., Frohlich, D.R., Rosell, R.C., Bedford, I.D. and Markham, P.G. 1996. The relevance of variability within the *Bemisia tabaci* species complex to epidemics caused by Subgroup III geminiviruses. pp. 77-89. In D. Gerling and R.T. Mayer (Eds.), *Bemisia: 1995 Taxonomy, Biology, Damage, Control and Management*. Intercept Ltd., Andover, Hants, UK.
641. Brown, J.K., Coats, S., Frohlich, D.R., Bedford, I.D. and Markham, P.G. 1995. Characterization and distribution of esterase electromorphs in the whitefly, *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae). *Biochem. Genet.* 33: 205-214.
642. Brown, J.K. and Czosnek, H. 2001. Whitefly transmitted viruses. pp. 65-100. In *Advances in Botanical Research*. Academic Press, NY.
643. Brown, J.K., Frohlich, D.R. and Rosell, R.C. 1995. The sweetpotato or silverleaf whiteflies: biotypes of *Bemisia tabaci* or a species complex? *Annu. Rev. Entomol.* 40: 511-534.
644. Brown, J.K., Idris, A.M., Olsen, M.W., Miller, M.E., Isakeit, T. and Anciso, J. 2000. Cucurbit leaf curl virus, a new whitefly transmitted geminivirus in Arizona, Texas, and Mexico. *Plant Dis.* 84(7): 809.
645. Brown, J.K., Idris, A.M., Rogan, D., Hussein, M.H. and Palmieri, M. 2001. Melon chlorotic leaf curl virus, a new begomovirus associated with *Bemisia tabaci* infestations in Guatemala. *Plant Dis.* 85: 1027.
646. Brown, J.K., Idris, A.M., Torres-Jerez, I., Banks, G.K. and Wyatt, S.D. 2001. The core region of the coat protein gene is highly useful for establishing the provisional identification and classification of begomoviruses. *Arch. Virol.* 146: 1581-1598.
647. Brown, J.K., Mihail, J.D. and Nelson, M.R. 1985. The effect of cotton leaf crumple on cotton inoculated at different growth stages. pp. 152-155. In *Cotton, A College of Agriculture Report, Series P-63*. University of Arizona, Tucson.
648. Brown, J.K., Mihail, J.D. and Nelson, M.R. 1987. Effects of cotton leaf crumple virus on cotton inoculated at different growth stages. *Plant Dis.* 71: 699-703.
649. Brown, J.K. and Nelson, M.R. 1984. Geminiate particles associated with cotton leaf crumple disease in Arizona. *Phytopathology* 74: 987-990.
650. Brown, J.K. and Nelson, M.R. 1985. Cotton leaf crumple virus, and whitefly-transmitted geminivirus of cotton in Arizona. pp. 156-157. In *Cotton, A College of Agriculture Report, Series P-63*. University of Arizona, Tucson.
651. Brown, J.K. and Nelson, M.R. 1986. Host range study of the cotton leaf crumple virus. pp. 171-176. In *Cotton, A College of Agriculture Report, Series P-63*. University of Arizona, Tucson.
652. Brown, J.K. and Nelson, M.R. 1986. Whitefly-borne viruses of melons and lettuce in Arizona. *Phytopathology* 76(2): 236-239. [Cock (1993)]
653. Brown, J.K. and Nelson, M.R. 1987. Host range and vector relationships of cotton leaf crumple virus. *Plant Dis.* 71(6): 522-524. [Cock (1993)]
654. Brown, J.K. and Nelson, M.R. 1988. Transmission, host range, and virus-vector relationships of chino del tomate virus (CdTV), a whitefly-transmitted geminivirus from Sinaloa. *Plant Dis.* 72(10): 866-869. [Cock (1993)]
655. Brown, J.K. and Nelson, M.R. 1989. Characterization of watermelon curly mottle virus a geminivirus distinct from squash leaf curl virus. *Ann. Appl. Biol.* 115(2): 243-252. [Cock (1993)]
656. Brown, J.K. and O'Leary, P.F. 1994. WFT geminiviruses in cotton: research imperative. *California-Arizona Cotton* 30(Sept/Oct): 23-25.
657. Brown, J.K. and O'Leary, P.F. 1994. Whitefly-transmitted geminiviruses. *California-Arizona Cotton* 30(Sept/Oct): 20-22.
658. Brown, J.K., Ostrow, K.M., Idris, A.M. and Stenger, D.C. 1999. Biotic, molecular, and phylogenetic characterization of bean calico mosaic virus, a distinct *Begomovirus* species with affiliation in the squash leaf curl virus cluster. *Phytopathology* 89(4): 273-280.
659. Brown, J.K., Ostrow, K.M., Idris, A.M. and Stenger, D.C. 2000. Chino del tomate virus: Relationships to other begomoviruses and identification of A-component variants that affect symptom expression. *Phytopathology* 90(5): 546-552.
660. Brown, J.K., Perring, T.M., Cooper, A.D., Bedford, I.D. and Markham, P.G. 2000. Genetic analysis of *Bemisia* (Hemiptera: Aleyrodidae) populations by isoelectric focusing electrophoresis. *Biochem. Genet.* 38(1/2): 13-25.
661. Brown, J.K. and Poulos, B.T. 1989. Detection of lettuce infectious yellows virus (LIYV) in greenhouse and field inoculated plants using an indirect enzyme-linked immunosorbent assay (indirect ELISA). *J. Rio Grande Valley Hortic. Soc.* 42: 13-18.
662. Brown, J.K. and Poulos, B.T. 1991. Isolation of geminivirus DNA and demonstration of infectivity by biolistic bombardment of host plants. *Annu. Rep. Bean Improvement Coop.* 34: 13-14.
663. Brown, J.K., Wendt, K.R. and Wyatt, S.D. 1995. Genetic variability among squash leaf curl virus isolates by component-specific polymerase chain reaction. pp. 5-11. In G.D. Lester and J.R. Dunlap (Eds.), *Proceedings of Cucurbitaceae 94: Education and enhancement of cucurbit germplasm*. South Padre Island, Texas Nov. 1-2, 1994.
664. Brown, L.G., Brown, J.K. and Tsai, J.H. 1990. Lettuce infectious yellows virus. *Plant Pathol. Cir. No. 335*, Florida Dept. Agric. Consumer Services, Div. Plant Industry
665. Brown, P.W., Watson, T.F. and Silvertooth, J.C. 1993. Weather conditions associated with outbreaks of severe whitefly infestations in Arizona. pp. 702-705. In D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
666. Brown, P.W., Watson, T.F. and Silvertooth, J.C. 1993. Weather conditions associated with outbreaks of severe whitefly infestations in Arizona. pp. 206-215. In *Cotton, A College of Agriculture Report, Series P-94*. University of Arizona, Tucson.
667. Brownbridge, M., Costa, S. and Jaronski, S.T. 2001. Effects of in vitro passage of *Beauveria bassiana* on virulence to *Bemisia argentifolii*. *J. Invertebr. Pathol.* 77(4): 280-283.
668. Broza, M., Butler, G.D., Jr. and Henneberry, T.J. 1988. Cottonseed oil for control of *Bemisia tabaci* on cotton. p. 301. In J.M. Brown and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Production Research Conferences*. National Cotton Council, Memphis, TN.
669. Brunetti, A., Tavazza, M., Noris, E., Tavazza, R., Caciagli, P., Ancora, G., Crespi, S. and Accotto, G.P. 1997. High expression of truncated viral rep protein confers resistance to tomato yellow leaf curl virus in transgenic tomato plants. *Mol. Plant-Microbe Interactions* 10(5): 571-579.
670. Brunt, A.A. 1986. Transmission of diseases. pp. 43-50. In M.J.W. Cock (Ed.), *Bemisia tabaci - A Literature Survey on the Cotton Whitefly with an Annotated Bibliography*. CAB International Institute of Biological Control, Ascot, UK.

671. Brunt, A.A., Atkey, P.T. and Woods, R.D. 1983. Intracellular occurrence of cowpea mild mottle virus in two unrelated plant species. *Intervirology* 20: 137-142. [ Cock (1986)]
672. Brunt, A.A., Crabtree, K. and Gibbs, A. 1990. Viruses of tropical plants - descriptions and lists from the VIDE database. C. A. B. International, Oxon, U. K., 707 pp. [Ioannou (1995)]
673. Brunt, A.A. and Kenten, R.H. 1973. Cowpea mild mottle, a newly recognised virus infecting cowpeas (*Vigna unguiculata*) in Ghana. *Ann. Appl. Sci.* 74: 67-74. [ Cock (1986)]
674. Brunt, A.A. and Phillips, S. 1981. 'Fuzzy vein', a disease of tomato (*Lycopersicon esculentum*) in Western Nigeria induced by cowpea mild mottle virus. *Trop. Agric. (Trinidad)* 58: 177-180. [ Cock (1986)]
675. Brushwood, D.E. 1998. The use of elevated temperatures to reduce the stickiness potential of honeydew contaminated cottons. pp. 1553-1557. *In* P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
676. Brushwood, D.E. and Perkins, H.H., Jr. 1995. Variations in cotton insect honeydew composition and the related effects on test methods and processing quality. pp. 1178-1181. *In* D.A. Richter and J. Armour (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
677. Bryan, H.H., Narayanan, K.R. and McMillan, R.T., Jr. 1992. Guar cultivar responses to nitrogen fertilizer rates on oolitic calcareous soils and disease incidence in South Florida. *Acta Hort.* 318: 271-280.
678. Buchner, E. 1995. Metodologia para cultivar tomates para proceso en el valle de comayagua (Honduras), bajo la limitante de la mosca blanca (*Bemisia tabaci*), transmisora del virus gemini. [A production method for limiting the transmission of geminivirus by the whitefly (*Bemisia tabaci*) in tomatoes in Comayagua Valley, Honduras]. *Proc. Interamerican Soc. Trop. Hortic.* 39: 46-61. [ Spanish, English summary]
679. Buckner, J.S., Freeman, T.P., Ruud, R.L., Chu, C.C. and Henneberry, T.J. 2002. Characterization and functions of the whitefly egg pedicel. *Arch. Insect Biochem. Physiol.* 49(1): 22-33.
680. Buckner, J.S., Hagen, M.M. and Nelson, D.R. 1999. The composition of the cuticular lipids from nymphs and exuviae of the silverleaf whitefly, *Bemisia argentifolii*. *Comp. Biochem. Physiol. B-Biochem. Mol. Biol.* 124(2): 201-207.
681. Buckner, J.S., Nelson, D.R. and Mardaus, M.C. 1994. The lipid composition of the wax particles from adult whiteflies, *Bemisia tabaci* and *Trialeurodes vaporariorum*. *Insect Biochem. Mol. Biol.* 24(10): 977-987.
682. Buckner, J.S., Poprawski, T.J., Jones, W.A. and Nelson, D.R. 2000. Effect of whitefly parasitoids on the cuticular lipid composition of *Bemisia argentifolii* (Homoptera : Aleyrodidae) nymphs. *Arch. Insect Biochem. Physiol.* 44(2): 82-89.
683. Bulut, E. and Gocmen, H. 2000. Pests and their natural enemies on greenhouse vegetables in Antalya. *Bull. OILB/SROP (IOBC/WPRS)* 23(1): 33-37.
684. Buntin, G.D., Gilbertz, D.A. and Oetting, R.D. 1993. Chlorophyll loss and gas exchange in tomato leaves after feeding injury by *Bemisia tabaci* (Homoptera, Aleyrodidae). *J. Econ. Entomol.* 86: 517-522.
685. Burban, C., Fishpool, L.D.C., Fauquet, C., Fargette, D. and Thouvenel, J.C. 1992. Host-associated biotypes within West African populations of the whitefly *Bemisia tabaci* (Genn.), (Hom., Aleyrodidae). *J. Appl. Entomol.* 113: 416-423.
686. Burghause, F. 1987. Neue Weisse Fliege fiel an Weihnachtssternen auf, *Bemisia tabaci*, ein Schaedling aus dem Mittelmeergebiet. [A new white fly species was observed in poinsettia. *Bemisia tabaci*, a Mediterranean pest]. *Taspo Gartenbaumagazin (Germany)* 121(50): 4. [ German]
687. Burghause, F. 1990. [White flies in greenhouses]. *Gärtnerbörse und Gartenwelt (Germany)* 90(14): 668-670, 672-673. [ German]
688. Bushara, A.G. and Salih, S.H. 1989. *Aphis* resistance. Control of the whitefly *Bemisia tabaci* on *Phaseolus vulgaris*. Screening insecticides for control of whiteflies on haricot bean. pp. 184-191. *In* *Annu. Rep. Hudeiba Res. Stn. (Sudan)*. 1985-1986.
689. Bustillo C, J. 1976. Informe de las labores del programa basico de mosca blanca: Seccion de Entomologia 1975-1976. [Basic program report on the white fly: entomology section 1975-1976]. Managua, Nicaragua, 75 pp. [ Spanish, English summary]
690. Butani, D.K. 1970. Les insectes ravageurs du cotonnier: XIII - Efficacite comparee de quelques poudres insecticides. [English version available]. *Coton Fibres Trop.* 25: 347-353. [ Cock (1986)]
691. Butani, D.K. 1980. Insect pests of vegetables and their control - cluster beans. *Pesticides* 14(1): 33-35. [ Cock (1986)]
692. Butani, D.K. and Jotwani, M.G. 1983. Insects as a limiting factor in vegetable production. *Pesticides* 17(9): 6-13. [ Cock (1986)]
693. Butani, D.K. and Sahni, V.M. 1970. Carbaryl - a versatile insecticide. *Pesticides* 4(3): 19-20. [ Cock (1986)]
694. Butani, D.K. and Singh, S. 1965. A note on the comparative efficacy of newer insecticides against pests of cotton. *Labdev J. Sci. Technol.* 3(1): 67-69. [ Cock (1986)]
695. Butler, G.D., Jr. 1982. Sweetpotato whitefly, *Bemisia tabaci*, a new pest of cotton in Arizona. pp. 49-50. *In* *Cotton, A College of Agriculture Report, Series P-56*. University of Arizona, Tucson.
696. Butler, G.D., Jr. 1984. Whitefly. p. 100. *In* *Cotton, A College of Agriculture Report, Series P-61*. University of Arizona, Tucson.
697. Butler, G.D., Jr. 1985. Populations of several insects on cotton in open-top carbon dioxide enrichment chambers. *Southwest. Entomol.* 10: 264-267.
698. Butler, G.D., Jr. 1986. Spring build-up of whiteflies in Central Arizona. pp. 226-229. *In* *Cotton, A College of Agriculture Report, Series P-63*. University of Arizona, Tucson.
699. Butler, G.D., Jr. 1986. Time for development of *Eretmocerus mundus*, a parasite of the sweet potato whitefly from Jordan. pp. 229-231. *In* *Cotton, A College of Agriculture Report, Series P-63*. University of Arizona, Tucson.
700. Butler, G.D., Jr. and Brown, J.K. 1985. Sweetpotato whitefly infection of cotton leaf crumple from weed hosts in 1984. pp. 149-151. *In* *Cotton, A College of Agriculture Report, Series P-63*. University of Arizona, Tucson.
701. Butler, G.D., Jr., Brown, J.K. and Henneberry, T.J. 1986. Effect of cotton seedling infection by cotton-leaf crumple virus on subsequent growth and yield. *J. Econ. Entomol.* 79: 208-211. [ Cock (1993)]
702. Butler, G.D., Jr., Coudriet, D. and Henneberry, T.J. 1988. Toxicity and repellency of soybean and cottonseed oils to the sweetpotato whitefly and the cotton aphid on cotton in greenhouse studies. *Southwest. Entomol.* 13: 81-86. [ Cock (1993)]
703. Butler, G.D., Jr., Coudriet, D. and Henneberry, T.J. 1989. Sweetpotato whitefly: Host plant preference and repellent effect of plant-derived oils on cotton, squash, lettuce and cantaloupe. *Southwest. Entomol.* 14: 9-16. [ Cock (1993)]
704. Butler, G.D., Jr., Coudriet, D.L. and Henneberry, T.J. 1991. Effect of plant-derived oils on sweetpotato whitefly on cotton. pp. 192-195. *In* *Cotton, A College of Agriculture Report, Series P-87*. University of Arizona, Tucson.
705. Butler, G.D., Jr., El-Lissy, O. and Antilla, L. 1993. Sweetpotato whitefly parasites abundant in some cotton fields during October. pp. 262-263. *In* *Cotton, A College of Agriculture Report, Series P-94*. University of Arizona, Tucson.
706. Butler, G.D., Jr. and Henneberry, T.J. 1983. Sweetpotato whitefly, *Bemisia tabaci*, 1982 Research Results. pp. 113-116. *In* *Cotton, A College of Agriculture Report, Series P-59*. University of Arizona, Tucson.

707. Butler, G.D., Jr. and Henneberry, T.J. 1984. *Bemisia tabaci* as a cotton pest in the desert cotton-growing areas of the southwestern United States. pp. 195-197. In J.M. Brown (Ed.), Proceedings Beltwide Cotton Production Research Conferences. National Cotton Council, Memphis, TN.
708. Butler, G.D., Jr. and Henneberry, T.J. 1984. *Bemisia tabaci*: Effect of cotton leaf pubescence on abundance. Southwest. Entomol. 9: 91-94. [Cock (1986)]
709. Butler, G.D., Jr. and Henneberry, T.J. 1984. Sweetpotato whitefly and parasite populations in sprayed cotton plots. pp. 97-99. In Cotton, A College of Agriculture Report, Series P-61. University of Arizona, Tucson.
710. Butler, G.D., Jr. and Henneberry, T.J. 1986. *Bemisia tabaci* (Gennadius), a pest of cotton in the Southwestern United States. USDA, Tech. Bull. 1701, 19 pp. [Cock (1993)]
711. Butler, G.D., Jr. and Henneberry, T.J. 1988. Laboratory studies on *Chrysopa carnea* predation on *Bemisia tabaci*. Southwest. Entomol. 13: 165-170. [Cock (1993)]
712. Butler, G.D., Jr. and Henneberry, T.J. 1989. Sweetpotato whitefly migration, population increase, and control on lettuce with cotton seed oil sprays. Southwest. Entomol. 14: 287-293. [Cock (1993)]
713. Butler, G.D., Jr. and Henneberry, T.J. 1990. Pest control on vegetables and cotton with household cooking oils and liquid detergents. Southwest. Entomol. 15: 123-131. [Cock (1993)]
714. Butler, G.D., Jr. and Henneberry, T.J. 1991. Effect of oil sprays on sweetpotato whitefly and phytotoxicity on watermelons, squash and cucumbers. Southwest. Entomol. 16: 63-72.
715. Butler, G.D., Jr. and Henneberry, T.J. 1991. Sweetpotato whitefly control: effect of tomato cultures and plant derived oils. Southwest. Entomol. 16: 37-43.
716. Butler, G.D., Jr. and Henneberry, T.J. 1992. Sweetpotato whitefly: flight activity, effects of wind velocity and precopulatory pairing activity patterns. pp. 140-145. In Cotton, A College of Agriculture Report, Series P-91. University of Arizona, Tucson.
717. Butler, G.D., Jr. and Henneberry, T.J. 1993. Sweetpotato whitefly control: enhancement of the repellency of oils. pp. 244-247. In Cotton, A College of Agriculture Report, Series P-94. University of Arizona, Tucson.
718. Butler, G.D., Jr. and Henneberry, T.J. 1993. Sweetpotato whitefly natural enemies: parasite surveys in urban areas and cotton fields and identification of a new predator. pp. 256-257. In Cotton, A College of Agriculture Report, Series P-94. University of Arizona, Tucson.
719. Butler, G.D., Jr. and Henneberry, T.J. 1994. *Bemisia* and *Trialeurodes* (Hemiptera: Aleyrodidae). pp. 325-352. In G.A. Matthews and J.P. Tunstall (Eds.), Insect Pests of Cotton. CAB International, Wallingford, UK.
720. Butler, G.D., Jr., Henneberry, T.J. and Brown, J.K. 1985. Cotton leaf crumple disease of Pima cotton. pp. 158-159. In Cotton, A College of Agriculture Report, Series P-63. University of Arizona, Tucson.
721. Butler, G.D., Jr., Henneberry, T.J. and Clayton, T.E. 1983. *Bemisia tabaci* (Homoptera: Aleyrodidae): development, oviposition, and longevity in relation to temperature. Ann. Entomol. Soc. Am. 76: 310-313. [Cock (1986)]
722. Butler, G.D., Jr., Henneberry, T.J. and Hutchison, W.D. 1986. Biology, sampling and population dynamics of *Bemisia tabaci*. pp. 167-195. In G.E. Russell (Ed.), Agricultural Zoology Reviews. Intercept [Cock (1993)]
723. Butler, G.D., Jr., Henneberry, T.J. and Hutchison, W.D. 1989. Biology, sampling and population dynamics of *Bemisia tabaci*. pp. 83-111. In G.E. Russell (Ed.), Biology and Population Dynamics of Invertebrate Crop Pests. Intercept, Andover, UK.
724. Butler, G.D., Jr., Henneberry, T.J. and Natwick, E.T. 1985. *Bemisia tabaci*: 1982 and 1983 populations in Arizona and California cotton fields. Southwest. Entomol. 10(1): 20-25. [Cock (1986)]
725. Butler, G.D., Jr., Henneberry, T.J. and Perkins, H.H., Jr. 1990. Sweetpotato whitefly populations in cotton genotypes at Poston, Arizona in 1988. pp. 144-145. In Cotton, A College of Agriculture Report, Series P-81. University of Arizona, Tucson.
726. Butler, G.D., Jr., Henneberry, T.J., Stansly, P.A. and Schuster, D.J. 1993. Insecticidal effects of selected soaps, oils and detergents on the sweetpotato whitefly: (Homoptera, Aleyrodidae). Florida Entomol. 76: 161-167.
727. Butler, G.D., Jr., Hutchison, W.D. and Broza, M. 1988. Effect of aldicarb treatments to cotton on *Bemisia tabaci* and *Aphis gossypii* populations in Israel. Southwest. Entomol. 13: 87-93. [Cock (1993)]
728. Butler, G.D., Jr., Kimball, B.A. and Mauney, J.R. 1985. Populations of the sweetpotato whitefly on cotton grown in open-top field carbon dioxide-enrichment chambers. pp. 175-176. In Cotton, A College of Agriculture Report, Series P-63. University of Arizona, Tucson.
729. Butler, G.D., Jr., Kimball, B.A. and Mauney, J.R. 1986. *Bemisia tabaci* (Homoptera: Aleyrodidae) on cotton: adult activity and cultivar oviposition preference. J. Econ. Entomol. 79: 350-354.
730. Butler, G.D., Jr., Kimball, B.A. and Mauney, J.R. 1986. Populations of *Bemisia tabaci* (Homoptera: Aleyrodidae) on cotton grown in open-top field chambers enriched with CO<sub>2</sub>. Environ. Entomol. 15: 61-63.
731. Butler, G.D., Jr., Naranjo, S.E., Henneberry, T.J. and Brown, J.K. 1995. Bibliography of *Bemisia tabaci* and *Bemisia argentifolii*. pp. 197-257. In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2.
732. Butler, G.D., Jr., Puri, S.N. and Henneberry, T.J. 1991. Plant-derived oil and detergent solutions as control agents for *Bemisia tabaci* and *Aphis gossypii* on cotton. Southwest. Entomol. 16: 331-337.
733. Butler, G.D., Jr. and Rao, S.B.P. 1990. Cottonseed oil to combat white-fly. Indian Textile J. February: 20-25.
734. Butler, G.D., Jr., Rimon, D. and Henneberry, T.J. 1988. *Bemisia tabaci* (Homoptera: Aleyrodidae): Populations on different cotton varieties and cotton stickiness in Israel. Crop Prot. 7: 43-47. [Cock (1993)]
735. Butler, G.D., Jr. and Wilson, F.D. 1984. Activity of adult whiteflies (Homoptera: Aleyrodidae) within plantings of different cotton strains and cultivars as determined by sticky trap catches. J. Econ. Entomol. 77: 1137-1140. [Cock (1986)]
736. Butler, G.D., Jr. and Wilson, F.D. 1986. Whitefly adults in okra-leaf and normal-leaf cottons. pp. 223-226. In Cotton, A College of Agriculture Report, Series P-63. University of Arizona, Tucson.
737. Butler, G.D., Jr., Wilson, F.D. and Fishler, G. 1991. Cotton leaf trichomes and populations of *Empoasca lybica* and *Bemisia tabaci*. Crop Prot. 10: 461-464.
738. Butler, G.D., Jr., Wilson, F.D. and Henneberry, T.J. 1985. Cotton leaf crumple in okra-leaf and normal-leaf cottons. J. Econ. Entomol. 78: 1500-1502. [Cock (1993)]
739. Butler, N.S. and Dhawan, A.K. 1996. Management of whitefly (*Bemisia tabaci*) on cotton in Punjab [India]. p. 60. In M.J. Iqbal and A. Kasana (Eds.), Second International Congress of Entomological Sciences, March 19-21, 1996, Islamabad, Pakistan.
740. Butler, N.S. and Kular, J.S. 1987. Effect of cotton whitefly damage on seed germination and fiber qualities of upland cotton. Indian J. Ecol. 14(1): 158-160.
741. Butler, N.S. and Kular, J.S. 1999. Resurgence of whitefly in cotton and its management. Indian J. Entomol. 61(1): 85-90.
742. Butler, N.S. and Rataul, H.S. 1973. Control of tomato leafcurl virus (TLCV) in tomatoes by controlling the vector whitefly *Bemisia tabaci* Gen. by mineral-oil sprays. Curr. Sci. (Bangalore) 42: 864-865. [Cock (1986)]
743. Butler, N.S. and Rataul, H.S. 1977. Effect of TLCV infection on *Bemisia tabaci*. Entomol. 2(2): 163-164. [Cock (1986)]

744. Butter, N.S. and Rataul, H.S. 1977. The virus-vector relationship of the tomato leafcurl virus (TLCV) and its vector, *Bemisia tabaci* (Genn.) (Hemiptera: Aleyrodidae). *Phytoparasitica* 5(3): 173-186. [Cock (1986)]
745. Butter, N.S. and Rataul, H.S. 1978. Influence of temperature on the transmission efficiency and acquisition threshold of whitefly, *Bemisia tabaci* Genn. in the transmission of tomato leafcurl virus. *Sci. Culture* 44(4): 168-170. [Cock (1986)]
746. Butter, N.S. and Rataul, H.S. 1981. Control strategies in whitefly - borne viruses - a review. *Pestology* 5(12): 7-14. [Cock (1986)]
747. Butter, N.S. and Rataul, H.S. 1981. Nature and extent of loss in tomatoes due to tomato leafcurl virus (TLCV) transmitted by whitefly, *Bemisia tabaci* Genn. (Hemiptera: Aleyrodidae). *Indian J. Ecol.* 8: 299-300.
748. Butter, N.S. and Sukhija, H.S. 1987. Efficacy of flucythrinate (Pay Off 10 EC) against bollworms (*Pectinophora gossypiella*, *Earias* spp. and *Heliothis armigera*) infesting cotton. *J. Res. (Punjab Agric. Univ.)* 24(4): 615-622. [Cock (1993)]
749. Butter, N.S. and Vir, B.K. 1989. Morphological basis of resistance in cotton to the whitefly *Bemisia tabaci*. *Phytoparasitica* 17(4): 251-261. [Cock (1993)]
750. Butter, N.S. and Vir, B.K. 1990. Sampling of whitefly *Bemisia tabaci* (Genn.) in cotton. *J. Res. (Punjab Agric. Univ.)* 27(4): 615-619.
751. Butter, N.S. and Vir, B.K. 1991. Response of whitefly, *Bemisia tabaci* Genn., to different cotton genotypes under glasshouse conditions. *Indian J. Entomol.* 53: 115-119.
752. Butter, N.S., Vir, B.K., Kaur, G., Singh, T.H. and Raheja, R.K. 1992. Biochemical basis of resistance to whitefly *Bemisia tabaci* (Genn.) (Aleyrodidae: Hemiptera) in cotton. *Trop. Agric.* 69(2): 119-122.
753. Butter, N.S., Vir, B.K., Kular, J.S., Brar, A.S. and Nagi, P.S. 1996. Relationship of plant nutrients and whitefly, *Bemisia tabaci* in upland cotton. *Indian J. Entomol.* 58(1): 1-6.
754. Buxton, J. and Clarke, A. 1994. Evaluation of insecticide DIPS to control *Bemisia tabaci* on poinsettia cuttings. *Pestic. Sci.* 42(2): 141-142.
755. Byabakama, B.A., Adipala, E., Ogenga-Latigo, M.W. and Otim-Nape, G.W. 1999. The effect of amount and disposition of inoculum on cassava mosaic virus disease development and tuberous root yield of cassava. *African Plant Prot.* 5(1): 21-29.
756. Byerly-Murphy, K.F., Martínez-Carrillo, J.L. and Nava-Camberos, U. 1998. Manejo integrado de plagas [Integrated pest management]. pp. 3-25. *In* J.J. Pacheco-Covarrubias and F. Pacheco-Mendivil (Eds.), *Temas Selectos Para El Manejo Integrado de la Mosquita Blanca*, Memoria Científica No. 6. Instituto Nacional De Investigaciones Forestales, Agrícolas y Pecuarias, Centro de Investigación Regional del Noroeste, CIRNO, Obregon, Sonora, Mexico.
757. Byrne, D.N. 1999. Migration and dispersal by the sweet potato whitefly, *Bemisia tabaci*. *Agric. Forest Meteorol.* 97(4): 309-316.
758. Byrne, D.N. and Bellows, T.S. 1991. Whitefly biology. *Annu. Rev. Entomol.* 36: 431-457.
759. Byrne, D.N., Bellows, T.S. and Parrella, M.P. 1990. Whiteflies in agricultural systems. pp. 227-261. *In* D. Gerling (Ed.), *Whiteflies: their Bionomics, Pest Status and Management*. Intercept, Andover, UK.
760. Byrne, D.N. and Blackmer, J.L. 1996. Examination of short-range migration by *Bemisia*. pp. 17-28. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia: 1995 Taxonomy, Biology, Damage, Control and Management*. Intercept Ltd., Andover, Hants, UK.
761. Byrne, D.N., Buchmann, S.L. and Spangler, H.G. 1988. Relationship between wing loading, wingbeat frequency and body mass in homopterous insects. *J. Exp. Biol.* 135: 9-23.
762. Byrne, D.N., Cohen, A.C. and Draeger, E.A. 1990. Water uptake from plant tissue by the egg pedicel of the greenhouse whitefly *Trialeurodes vaporariorum* Westwood (Homoptera: Aleyrodidae). *Canadian J. Zool.* 68(6): 1193-1195.
763. Byrne, D.N. and Draeger, E.A. 1989. Effect of plant maturity on oviposition and nymphal mortality of *Bemisia tabaci* (Homoptera: Aleyrodidae). *Environ. Entomol.* 18(3): 429-432. [Cock (1993)]
764. Byrne, D.N. and Hadley, N.F. 1988. Particulate surface waxes of whiteflies: morphology, composition and waxing behaviour. *Physiol. Entomol.* 13(3): 267-276. [Cock (1993)]
765. Byrne, D.N. and Houck, M.A. 1990. Morphometric identification of wing polymorphism in *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae). *Ann. Entomol. Soc. Am.* 83(3): 487-493. [Cock (1993)]
766. Byrne, D.N. and Miller, W.B. 1990. Carbohydrate and amino acid composition of phloem sap and honeydew produced by *Bemisia tabaci*. *J. Insect Physiol.* 36(6): 433-440. [Cock (1993)]
767. Byrne, D.N., Rathman, R.J., Orum, T.V. and Palumbo, J.C. 1996. Localized migration and dispersal by the sweet potato whitefly, *Bemisia tabaci*. *Oecologia* 105(3): 320-328.
768. Byrne, D.N. and von Bretzel, P.K. 1987. Similarity in flight activity rhythms in coexisting species of Aleyrodidae, *Bemisia tabaci* (Gennadius) and *Trialeurodes abutilonea* (Haldeman). *Entomol. Exp. Appl.* 43: 215-219.
769. Byrne, D.N., von Bretzel, P.K. and Hoffman, C.J. 1986. Impact of trap design and placement when monitoring for the bandedwinged whitefly and the sweetpotato whitefly (Homoptera: Aleyrodidae). *Environ. Entomol.* 15(2): 300-304. [Cock (1993)]
770. Byrne, F.J., Bedford, I.D., Devonshire, A.L. and Markham, P.G. 1995. Esterase variation and squash induction in B-type *Bemisia tabaci* (Homoptera: Aleyrodidae). *Bull. Entomol. Res.* 85(2): 175-179.
771. Byrne, F.J., Cahill, M., Denholm, I. and Devonshire, A.L. 1994. A biochemical and toxicological study of the role of insensitive acetylcholinesterase in organophosphorus resistant *Bemisia tabaci* (Homoptera: Aleyrodidae) from Israel. *Bull. Entomol. Res.* 84(2): 179-184.
772. Byrne, F.J., Cahill, M., Denholm, I. and Devonshire, A.L. 1995. Biochemical identification of interbreeding between B-type and non B-type strains of the tobacco whitefly *Bemisia tabaci*. *Biochem. Genet.* 33(1-2): 13-23.
773. Byrne, F.J., Denholm, I., Birnie, L.C., Devonshire, A.L. and Rowland, M.W. 1991. Analysis of insecticide resistance in the whitefly, *Bemisia tabaci*. pp. 165-178. *In* I. Denholm, A. Devonshire and D. Hollomon (Eds.), *Resistance 91: Achievements and Developments in Combating Pesticide Resistance*. Elsevier, London, UK.
774. Byrne, F.J. and Devonshire, A.L. 1991. *In vivo* inhibition of esterase and acetylcholin esterase activities by profenofos treatments in the tobacco whitefly *Bemisia tabaci* (Genn.): Implications for routine biochemical monitoring of the enzymes. *Pestic. Biochem. Physiol.* 40(3): 198-204. [Cock (1993)]
775. Byrne, F.J. and Devonshire, A.L. 1993. Insensitive acetylcholinesterase and esterase polymorphism in susceptible and resistant populations of the tobacco whitefly, *Bemisia tabaci*. *Pestic. Biochem. Physiol.* 45: 34-42.
776. Byrne, F.J. and Devonshire, A.L. 1996. Biochemical evidence of haplodiploidy in the whitefly *Bemisia tabaci*. *Biochem. Genet.* 34(3/4): 93-107.
777. Byrne, F.J. and Devonshire, A.L. 1997. Kinetics of insensitive acetylcholinesterases in organophosphate-resistant tobacco whitefly, *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae). *Pestic. Biochem. Physiol.* 58(2): 119-124.
778. Byrne, F.J., Gorman, K.J., Cahill, M., Denholm, I. and Devonshire, A.L. 2000. The role of B-type esterases in conferring insecticide resistance in the tobacco whitefly, *Bemisia tabaci* (Genn.). *Pest Manag. Sci.* 56(10): 867-874.
779. C.A.B. International Institute of Entomology. 1986. Distribution [throughout the tropics]. pp. 13-16. *In* M.J.W. Cock (Ed.), *Bemisia tabaci - A Literature Survey on the Cotton Whitefly with an Annotated Bibliography*. CAB International Institute of Biological Control, Ascot, UK.



780. Caballero, R. 1992. Moscas blancas neotropicales (Homoptera: Aleyrodidae): Hospedantes, distribución, enemigos naturales e importancia económica. [Neotropical whiteflies (Homoptera: Aleyrodidae): hosts, distribution, natural enemies and economic importance]. pp. 10-15. *In* L. Hilje and O. Arboleda (Eds.), Las moscas blancas (Homoptera: Aleyrodidae) en América Central y el Caribe. CATIE, Turrialba, Costa Rica. [Spanish]
781. Caballero, R. 1996. Identificación de mosca blanca. pp. 1-10. *In* L. Hilje (Ed.), Metodologías para el estudio y manejo de moscas blancas y geminivirus. Centro Agronómico Tropical de Investigación y Enseñanza, Turrialba, Costa Rica.
782. Caballero, R. and Rueda, A. 1992. Las moscas blancas en Honduras. [The whiteflies in Honduras]. pp. 50-53. *In* L. Hilje and O. Arboleda (Eds.), Las moscas blancas (Homoptera: Aleyrodidae) en América Central y el Caribe. CATIE, Turrialba, Costa Rica. [Spanish]
783. Cabello-García, T., Carricondo-Martínez, I., Justicia del Río, L. and Belda-Suárez, J.E. 1996. Biología y control de las especies de mosca blanca *Trialeurodes vaporariorum* (Gen.) y *Bemisia tabaci* (West.) (Hom.; Aleyrodidae) en cultivos hortícolas en invernadero. [Biology and control of white fly species *Trialeurodes vaporariorum* (Gen.) and *Bemisia tabaci* (West.) (Hom.; Aleyrodidae) in horticultural crops in greenhouses]. Dirección General de Investigación Agraria, Sevilla, Spain, 96 pp. [Spanish]
784. Cabello, T., Gómez, M., Barranco, P., Lucas, M. and Belda, J.E. 1997. Evaluation of oxamyl against homopteran pests in greenhouse-grown pepper, applied with drip irrigation [Spain]. *Tests Agrochemicals Cultivars* 18: 2-3.
785. CABI/EPP0. 1999. *Bemisia tabaci* (Gennadius) biotype B, Hemiptera: Aleyrodidae. Distribution Maps of Plant Pests 591. CAB International, Wallingford, Oxon, UK.
786. CABI/EPP0. 1999. *Bemisia tabaci* (Gennadius), Hemiptera: Aleyrodidae. Distribution Maps of Plant Pests 284, 1st Revision. CAB International, Wallingford, Oxon, UK.
787. Cabrera-Asencio, I. and Bastidas-López, H. 2000. Effect of *Wedelia trilobata* L. (Asterales: Asteraceae) on the whitefly *Bemisia tabaci* G. (Homoptera: Aleyrodidae). *J. Agric. (Univ. Puerto Rico)* 84(3-4): 181-184.
788. Cabrera-Asencio, I.S., Martínez, S. and Orenge, E. 1997. Insectos presentes en cultivos comerciales de *Brassica oleracea* G. *italica* en diferentes áreas de Puerto Rico. [Insects present on commercial cultivars of *Brassica oleracea* spp. *italica* (L.) in different areas of Puerto Rico]. *J. Agric. (Univ. Puerto Rico)* 81(1-2): 87-90. [Spanish, English summary]
789. Cacciola, S.O., Vacante, V., Garzia, G.T., San-Lio, G.M., Tropea-Garzia, G. and Magnano, G. 1995. Selection of fungicide-resistant strains of *Verticillium lecanii*. *Med. Fac. Landbouww. Univ. Gent* 60(3a): 725-733.
790. Caciagli, P. and Bosco, D. 1996. Quantitative determination of tomato yellow leaf curl geminivirus DNA by chemiluminescent assay using digoxigenin-labeled probes. *J. Virol. Methods* 57(1): 19-29.
791. Caciagli, P. and Bosco, D. 1997. Quantitation over time of tomato yellow leaf curl geminivirus DNA in its whitefly vector. *Phytopathology* 87(6): 610-613.
792. Caciagli, P., Bosco, D. and Al-Bitar, L. 1995. Relationships of the Sardinian isolate of tomato yellow leaf curl geminivirus with its whitefly vector *Bemisia tabaci* Gen. Eur. *J. Plant Pathol.* 101(2): 163-170.
793. Caciagli, P.C. 2001. Whitefly-borne viruses in Continental Europe. pp. 279-292. *In* K.F. Harris, O.P. Smith and J.E. Duffus (Eds.), Virus-Insect-Plant Interactions. Academic Press, San Diego, CA.
794. Cahill, M., Byrne, F.J., Denholm, I., Devonshire, A.L. and Gorman, K.J. 1994. Insecticide resistance in *Bemisia tabaci*. *Pestic. Sci.* 42(2): 137-139.
795. Cahill, M., Byrne, F.J., Gorman, K., Denholm, I. and Devonshire, A.L. 1995. Pyrethroid and organophosphate resistance in the tobacco whitefly *Bemisia tabaci* (Homoptera: Aleyrodidae). *Bull. Entomol. Res.* 85(2): 181-187.
796. Cahill, M. and Denholm, I. 1993. Detection of resistance to buprofezin in the whitefly *Bemisia tabaci*. *Resist. Pest Manage.* 5(1): 42.
797. Cahill, M. and Denholm, I. 1998. Pesticide resistance in protected agriculture - emphasis on *Bemisia*. pp. 45-53. *In* I. Cuadrado-Gómez and Viñuela-Sandoval, E. (Eds.), Resistencia a Los Pesticidas en los Cultivos Hortícolas, Almería.
798. Cahill, M. and Denholm, I. 1999. Managing resistance in the chloronicotinyl insecticides - rhetoric or reality. pp. 253-270. *In* I. Yamamoto and J.E. Casida (Eds.), Nicotinoid Insecticides and the Nicotinic Acetylcholine Receptor. Springer-Verlag, Tokyo.
799. Cahill, M., Denholm, I., Byrne, F.J. and Devonshire, A.L. 1996. Insecticide resistance in *Bemisia tabaci* - current status and implications for management. pp. 75-80. *In* Brighton Crop Protection Conference: Pests and Diseases. British Crop Protection Council, Brighton, UK.
800. Cahill, M., Denholm, I., Ross, G., Gorman, K. and Johnston, D. 1996. Relationship between bioassay data and the simulated field performance of insecticides against susceptible and resistant adult *Bemisia tabaci* (Homoptera: Aleyrodidae). *Bull. Entomol. Res.* 86(2): 109-116.
801. Cahill, M., Gorman, K., Day, S., Denholm, I., Elbert, A. and Nauen, R. 1996. Baseline determination and detection of resistance to imidacloprid in *Bemisia tabaci* (Homoptera: Aleyrodidae). *Bull. Entomol. Res.* 86(4): 343-349.
802. Cahill, M., Jarvis, W., Gorman, K. and Denholm, I. 1996. Resolution of baseline responses and documentation of resistance to buprofezin in *Bemisia tabaci* (Homoptera: Aleyrodidae). *Bull. Entomol. Res.* 86(2): 117-122.
803. Cahill, M., Johnston, D., Gorman, K. and Denholm, I. 1994. Insecticide resistance in *Bemisia tabaci* from Pakistan. pp. 431-436. *In* Brighton Crop Protection Conference: Pests and Diseases. British Crop Protection Council, Farnham, UK.
804. Cai, J.H., Wang, X.F. and Huang, F. 1993. Studies on tobacco leaf curl virus (TLCV) in Guangxi Province of China I. Biological properties and serological relationship with African cassava mosaic virus (ACMV). *Acta Microbiol. Sinica* 33(3): 166-169. [Chinese, English summary]
805. Cali, S. 1997. Turkey. pp. 77-80. *In* N. Ioannou (Ed.), Management of the whitefly-virus complex. FAO Plant Production and Protection Paper 143, Rome, Italy.
806. Calvert, L.A. 2000. Identificación molecular de los biotipos A y B de *Bemisia tabaci*. pp. 129-132. *In* Bean Golden Mosaic and Other Diseases of Common Bean Caused by Whitefly-Transmitted Geminiviruses in Latin America. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. [Spanish]
807. Calvert, L.A., Cuervo, M., Arroyave, J.A., Constantino, L.M., Bellotti, A. and Frohlich, D. 2001. Morphological and mitochondrial DNA marker analyses of whiteflies (Homoptera: Aleyrodidae) colonizing cassava and beans in Colombia. *Ann. Entomol. Soc. Am.* 94(4): 512-519.
808. Calvitti, M. and Remotti, P.C. 1998. Host preference and performance of *Bemisia argentifolii* (Homoptera: Aleyrodidae) on weeds in Central Italy. *Environ. Entomol.* 27(6): 1350-1356.
809. Calvo G. 1996. Análisis de datos económicos. pp. 104-111. *In* L. Hilje (Ed.), Metodologías para el estudio y manejo de moscas blancas y geminivirus. Centro Agronómico Tropical de Investigación y Enseñanza, Turrialba, Costa Rica.
810. Campbell, B.C. 1993. Congruent evolution between whiteflies (Homoptera: Aleyrodidae) and their bacterial endosymbionts based on respective 18S and 16S rDNAs. *Curr. Microbiol.* 26: 129-132.
811. Campbell, B.C., Duffus, J.E. and Baumann, P. 1993. Determining whitefly species. *Science* 261: 1333-1335.
812. Campbell, B.C., Steffen-Campbell, J.D. and Gill, R.J. 1994. Evolutionary origin of whiteflies (Hemiptera: Sternorrhyncha: Aleyrodidae) inferred from 18S rDNA sequences. *Insect Mol. Biol.* 3(2): 73-88.

813. Campbell, B.C., Steffen-Campbell, J.D. and Gill, R.J. 1996. Origin and radiation of whiteflies: an initial molecular phylogenetic assessment. pp. 29-51. In D. Gerling and R.T. Mayer (Eds.), *Bemisia*: 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
814. Cancino, M., Abouzid, A.M., Morales, F.J., Purcifull, D.E., Polston, J.E. and Hiebert, E. 1995. Generation and characterization of three monoclonal antibodies useful in detecting and distinguishing bean golden mosaic virus isolates. *Phytopathology* 85(4): 484-490.
815. Candido, G.F.O. 1999. Virulence of the entomopathogenic fungus *Paecilomyces amoeneroseus* (Hennings) Samson toward the whitefly *Bemisia argentifolii* Bellows and Perring. [University of California, Riverside]. Dissertation Abst. Int. 60(09B): 4332.
816. Caner, J., Fazio, G., Alexandre, M.A.V., Kudamatsu, M. and Vicente, M. 1985. [Action of antiviral chemicals in the control of bean golden mosaic virus on *Phaseolus lunatus* L.]. *Arch. Inst. Biol.* 52(1-4): 39-43. [Cock (1993), Portuguese, English summary]
817. Cano, M.C. 1986. Evaluación comparativa de cuatro insecticidas y una mezcla Lannate+Decis para el control de larvas de *Heliothis* spp. *Keiferia lycopericella* y adultos de *Bemisia tabaci* en el cultivo del tomate. [Comparative evaluation of 4 insecticides and one mixture (Lannate+Decis) for control of larval *Heliothis* spp. and *Keiferia lycopericella* and adult *Bemisia tabaci* in tomatoes]. Instituto Superior de Ciencias Agropecuarias, Escuela de Producción Vegetal, Managua, Nicaragua, 38 pp. [Spanish]
818. Cano-Rios, P., Avila-García, M.R., Nava-Camberos, U., Sanchez-Galvan, H., Lopez-Rios, E., Rangel-Santos, M., Blanco-Contreras, E. and Jimenez-Diaz, F. 2001. Plantas hospedantes de la "mosquita blanca de la hoja plateada", *Bemisia argentifolii* (Bellows & Perring) (Homoptera: Aleyrodidae) en la Comarca Lagunera, Mexico. *Folia Entomol. Mexicana* 40(1): 53-65. [Spanish, English summary]
819. Cano-Rios, P., Nava-Camberos, U. and Jimenez-Diaz, F. 2001. Efecto de la densidad de mosquita blanca, *Bemisia argentifolii* Bellows & Perring (Homoptera: Aleyrodidae) sobre el rendimiento y calidad del melón (*Cucumis melo* L.) en la Comarca Lagunera, Mexico. *Folia Entomol. Mexicana* 40(2): 145-154. [Spanish, English summary]
820. Cantliffe, D.J., Hochmuth, G.J., Locascio, S.J., Polston, J.E., Schuster, D.J., Chellemi, D.O., Olson, S.M., Stansly, P.A., Vavrina, C.S. and Seal, D.R. 1995. Production of solanacea for fresh market under field conditions: current problems and potential solutions. *Acta Hort.* 412: 229-244.
821. Capoor, S.P. 1939. Mosaic of *Jatropha curcas*. *Prog. Rep. Virus diseases scheme, Bombay (1939-1940)*, 18 pp. [Cock (1986)]
822. Capoor, S.P. and Ahmad, R.U. 1975. Yellow vein mosaic disease of field pumpkin and its relationship with the vector, *Bemisia tabaci*. *Indian Phytopathol.* 28: 241-246.
823. Capoor, S.P. and Varma, P.M. 1948. Yellow mosaic of *Phaseolus lunatus* L. *Curr. Sci. (Bangalore)* 17: 152-153. [Cock (1986)]
824. Capoor, S.P. and Varma, P.M. 1950. A new virus disease of *Dolichos lablab*. *Curr. Sci. (Bangalore)* 19: 248-249. [Cock (1986)]
825. Carazo, E., Martínez, J.L. and Bustamante, M. 1996. Insecticidas y resistencia. pp. 84-96. In L. Hilje (Ed.), *Metodologías para el estudio y manejo de moscas blancas y geminivirus*. Centro Agronómico Tropical de Investigación y Enseñanza, Turrialba, Costa Rica.
826. Cardenas-Alonso, M. 1982. Las enfermedades virales del frijol (*Phaseolus vulgaris* L.) transmitidas por la mosca blanca (*Bemisia tabaci* Genn.) con énfasis en Latinoamérica. [The viral diseases of bean (*Phaseolus vulgaris* L.) transmitted by the white fly, (*Bemisia tabaci* Genn.) with emphasis in Latin-America]. Universidad Autónoma Chapingo, Chapingo, Mexico, 25 pp. [Spanish]
827. Cardona, C. 1999. Las moscas blancas (Homoptera: Aleyrodidae) como plagas de cultivos semestrales en Colombia y Ecuador: visión general del problema. pp. 1-13. In *Proceedings, Moscas Blancas y Thrips: Un Agresivo Complejo de Plagas Agrícolas de Fin de Milenio*. Colombian Society of Entomology.
828. Cardoza, Y.J., McAuslane, H.J. and Webb, S.E. 1999. Mechanisms of resistance to whitefly-induced squash silverleaf disorder in zucchini. *J. Econ. Entomol.* 92(3): 700-707.
829. Cardoza, Y.J., McAuslane, H.J. and Webb, S.E. 2000. Effect of leaf age and silverleaf symptoms on oviposition site selection and development of *Bemisia argentifolii* (Homoptera: Aleyrodidae) on zucchini. *Environ. Entomol.* 29(2): 220-225.
830. Cardoza, Y.J., McAuslane, H.J., Webb, S.E. and Carle, R.B. 1999. Resistance to whitefly-induced squash silverleaf in *Cucurbita pepo* L., 1997. *Arthropod Management Tests* 24: 435.
831. Carlson, G.A. and Mohamed, A. 1986. Economic analysis of cotton-insect control in the Sudan Gezira. *Crop Prot.* 5(5): 348-354.
832. Carranza, L., Osorio, N. and Rios, D. 1998. Yield response of commercial cultivars of melon (*Cucumis melo* L.) in two ecosystems, La Villa de los Santos and Manaca in Baru, Chiriqui. *Ciencia Agropecuaria (Panama)* 9: 37-58. [Spanish, English summary]
833. Carruthers, R.I., Wraight, S.P. and Jones, W.A. 1993. An overview of biological control of the sweetpotato whitefly, *Bemisia tabaci*. pp. 680-685. In D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
834. Carter, F. 1990. The problem of sticky cotton in the USA and strategies for control. pp. 15-18. In *Cotton Production Research from a Farming Systems Perspective, with Special Emphasis on Stickiness: 49th Plenary Meeting*. International Cotton Advisory Committee
835. Carter, F.L. 1992. The sticky cotton issue. p. 645. In D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
836. Carter, F.L. 1993. Research and action plan for sweetpotato whitefly. pp. 120-121. In D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
837. Carter, F.L., Leigh, T.F., Natwick, E.T. and Perkins, H.H., Jr. 1985. Sticky cotton – a threat to our markets. *Proc. Western Cotton Production Conference*: 70-73.
838. Carter, F.L. and Perkins, H.H., Jr. 1986. Significance of insect honeydew contamination in textile processing. pp. 214-215. In J.M. Brown and T.C. Nelson (Eds.), *Proceedings Cotton Production Research Conferences. National Cotton Council, Memphis, TN.*
839. Carver, M. and Reid, I.A. 1996. Aleyrodidae (Hemiptera: Sternorrhyncha) of Australia. *CSIRO Div. Entomol. (Canberra)* 37: 1-55.
840. Castellani, E., Nur, A.M. and Mohamed, M.I. 1979. L'arricciamento fogliare del pomodoro in Somalia. [Tomato leaf curl disease in Somalia]. *Ann. Facolta Sci. Agrarie, Univ. Degli Studi Torino (Italy)* 12: 145-161. [Italian, English summary]
841. Castellani, E., Nur, A.M. and Mohamed, M.I. 1982. L'arricciamento fogliare del pomodoro in Somalia. *Ann. Facolta Sci. Agrarie Univ. Degli Studi Torino (Italy)* 12(1979-1982): 145-161. [Cock (1986)]
842. Castelo-Branco, M. and Pontes, L.A. 2001. Eficiência de tiacloprid para o controle de mosca-branca. [Efficiency of thiacloprid in controlling whiteflies]. *Hortic. Brasileira* 19(1): 97-101. [Portuguese, English summary]
843. Castillo-Torres, N., Montoya-Coronado, L., Garcia-Bernal, A., Navarro-Sandoval, F.J. and Rodríguez-Cota, F.G. 1998. Hector y esperanza: variedades de soya para el Noreste de México. INIFAP-CIRNO-CEVY, Folleto Técnico Núm. 32
844. Castineiras, A. 1995. Natural enemies of *Bemisia tabaci* (Homoptera: Aleyrodidae) in Cuba. *Florida Entomol.* 78(3): 538-540.
845. Castle, S., Henneberry, T., Toscano, N., Prabhaker, N., Birdsall, S. and Weddle, D. 1996. Silverleaf whiteflies show no increase in insecticide resistance. *California Agric.* 50(1): 18-23.
846. Castle, S., Henneberry, T.J., Prabhaker, N. and Toscaon, N.C. 1999. Perspectives on insecticide resistance and other causes of *Bemisia tabaci* outbreaks. pp. 32-42. In *Proceedings ICAC-CCRI Regional Consultation, Insecticide Resistance Management in Cotton*. Central Cotton Research Institute, Multan, Pakistan.

847. Castle, S.J. 1999. Agricultural intensification and pest outbreaks: a reappraisal of events in the Sudan Gezira. *Ann. Entomol. Soc. Am.* 92(6): 840-852.
848. Castle, S.J. 1999. Concentration and management of whiteflies in melons as a trap crop for cotton. pp. 412-417. *In Cotton, A College of Agriculture Report, Series P-116.* University of Arizona, Tucson.
849. Castle, S.J. 2000. Reduced whitefly infestations in cotton using a melon trap crop. pp. 315-319. *In Cotton, A College of Agriculture Report, Series P-121.* University of Arizona, Tucson.
850. Castle, S.J. 2001. Differences between cotton and melon in host acceptance by *Bemisia tabaci*. pp. 1056-1059. *In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences.* National Cotton Council, Memphis, TN.
851. Castle, S.J., Ellsworth, P.C., Prabhaker, N., Toscano, N.C. and Henneberry, T.J. 2001. Relative susceptibility of whiteflies to Danitol + Orthene over a 5-year period. pp. 269-277. *In Cotton, A College of Agriculture Report, Series P-125.* University of Arizona, Tucson.
852. Castle, S.J., Henneberry, T.J., Prabhaker, N. and Toscano, N.C. 1996. Trends in relative susceptibilities of whiteflies to insecticides through the cotton season in the Imperial Valley, CA. pp. 1032-1035. *In P. Dugger and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences.* National Cotton Council, Memphis, TN.
853. Castle, S.J., Henneberry, T.J. and Toscano, N.C. 1996. Suppression of *Bemisia tabaci* (Homoptera: Aleyrodidae) infestations in cantaloupe and cotton with sprinkler irrigation. *Crop Prot.* 15(7): 657-663.
854. Castle, S.J., Prabhaker, N. and Henneberry, T.J. 1999. Insecticide resistance and its management in cotton insects. *ICAC Review Article on Cotton Production Research, Int. Cotton Advisory Committee 5: 1-55.*
855. CATIE. 1992. Documentacion e informacion. Mosca blanca (*Bemisia tabaci* y *Trialeurodes vaporariorum*): Bibliografia. [Documentation and information. Whitefly (*Bemisia tabaci* and *Trialeurodes vaporariorum*): Bibliography]. Centro Agronomico Tropical de Investigacion y Ensenanza. CATIE, Turrialba, Costa Rica, 54 pp. [Spanish]
856. CATIE. 1992. Plan de accion regional para el manejo de las moscas blancas en America Central y el Caribe. CATIE, Turrialba, Costa Rica, 27 pp. [Spanish]
857. Cauquil, J. and Follin, J.E. 1983. Presumed virus and mycoplasma-like organism diseases in subsaharan africa and in the rest of the world. *Coton Fibres Trop.* 38(4): 309-317. [Ali et al. (1992)]
858. Cave, R.D. 1994. ¿Es viable el control biologico de un vector de geminivirus, como *Bemisia tabaci*? [Is it feasible to manage a vector of geminivirus like *Bemisia tabaci* through a biological control approach?]. *Manejo Integrado de Plagas (Costa Rica)* 34: 18-22. [Spanish, English summary]
859. Cave, R.D. 1996. Parasitodes y depredadores. pp. 69-76. *In L. Hilje (Ed.), Metodologias para el estudio y manejo de moscas blancas y geminivirus.* Centro Agronomico Tropical de Investigacion y Ensenanza, Turrialba, Costa Rica.
860. Cavero, E.S. and Menezes, A., Jr. 1984. Avaliacao dos efeitos de decis 2,5 no controle das pragas da batata. [Evaluation of the effects of decis 2,5 on the control of potato pests]. *Agros (Brazil)* 18(1/4): 50-56. [Portuguese, English summary]
861. Cebrian, R., Carnero, A. and Perez Padron, F. 1994. Pest status of *Bemisia tabaci* Gennadius (Homoptera: Aleyrodidae) on Canary Islands. *Bull. OILB/SROP (IOBC/WPRS)* 17: 47-51.
862. Ceglarska, E.B. 1999. *Dicyphus hyalinipennis* Burm. (Heteroptera: Miridae): a potential biological control agent for glasshouse pests in Hungary. *Bull. OILB/SROP (IOBC/WPRS)* 22: 33-36.
863. Celix, A., Lopez-Sese, A., Almarza, N., Gomez-Guillamon, L. and Rodriguez-Cerezo, R. 1996. Characterization of cucurbit yellow stunting disorder virus, a *Bemisia tabaci*-transmitted closterovirus. *Phytopathology* 86(12): 1370-1376.
864. Célix, A. and Rodríguez-Cerezo, E. 1996. Closterovirus transmitidos por mosca blanca. pp. 61-66. *In J.L. Cenis (Ed.), El Virus del Rizado Amirillo (Hoja en Cuchara) del Tomate (TYLCV) y Su Vector Bemisia tabaci, Jornadas 8. Consejería de Medio Ambiente, Agricultura y Agua, Murcia, Spain.*
865. Cenis, J.L. 1996. Introducción: Problemática planteada por *Bemisia tabaci*. pp. 9-12. *In J.L. Cenis (Ed.), El Virus del Rizado Amirillo (Hoja en Cuchara) del Tomate (TYLCV) y Su Vector Bemisia tabaci, Jornadas 8. Consejería de Medio Ambiente, Agricultura y Agua, Murcia, Spain.*
866. Cenis, J.L., Amorós, L., Hita, I., Reina, J. and Bejarano, E.R. 1996. Técnicas de detección del virus del rizado amarillo del tomate (TYLCV). pp. 23-34. *In J.L. Cenis (Ed.), El Virus del Rizado Amirillo (Hoja en Cuchara) del Tomate (TYLCV) y Su Vector Bemisia tabaci, Jornadas 8. Consejería de Medio Ambiente, Agricultura y Agua, Murcia, Spain.*
867. Centro de Investigaciones Agricolas de Sinaloa, M. 1974. Prueba de insecticida para combatir chupadores en algodonoero. Valle de Santo Domingo, Baja California Sur. 1975. [Insecticide test for the control of cotton plant sucking insects. Santo Domingo Valley. South Lower California 1975]. pp. 75-85. *In Informe Tecnico Labores Campo Agricola Experimental Valle De Santo Domingo, Mexico, 1974-1975.* [Spanish, English summary]
868. Cermeli, M. 1992. Control of *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae) on poinsettia with granular aldacar. *Boletin Entomol.* 7(1): 19-23. [Spanish, English summary]
869. Cervera, M.T., Cabezas, J.A., Simon, B., Martinez-Zapater, J.M., Beitia, F. and Cenis, J.L. 2000. Genetic relationships among biotypes of *Bemisia tabaci* (Hemiptera : Aleyrodidae) based on AFLP analysis. *Bull. Entomol. Res.* 90(5): 391-396.
870. Chagas, C.M., Barradas, M.M. and Vicente, M. 1981. Especies hospedeiras do virus do mosaico dourado do feijoeiro (VMDF). *Arch. Inst. Biol.* 48: 123-127. [Cock (1986)]
871. Chagas, C.M., Vicente, M. and Barradas, M.M. 1981. *Macropitium erythroloma* (Mart. ex Benth.) Urb. - Leguminosae - Possivel reservatorio do virus do mosaico dourado do feijoeiro (VMDF). *Arch. Inst. Biol.* 48: 113-116. [Cock (1986)]
872. Chakraborty, S., Pandey, P.K. and Singh, B. 1997. Okra enation leaf curl disease - a threat to cultivation of okra (*Abelmoschus esculentus* (L.) Moench). *Vegetable Sci.* 24(1): 52-54.
873. Chakravarthy, A.K., Prasad, R. and Mallikarjuna, G.D. 1997. Intercropping in cotton (*Gossypium hirsutum*) checks insect pest build-up. *Insect Environ.* 2(4): 131-132.
874. Chakravarthy, A.K. and Rao, P.K.A. 1985. Dispersion patterns, sample unit-sizes and techniques for sampling cotton jassid (*Amrasca biguttula biguttula* (Ishida)) and whitefly (*Bemisia tabaci*, Genn.) [French summary]. *Insect Sci. Appl.* 6(6): 661-665. [Cock (1993)]
875. Chakravarthy, A.K., Sidhu, A.S. and Singh, J. 1985. Effect of plant phenology and related factors on insect pest infestations in *arboresum* and *hirsutum* cotton varieties. *Insect Sci. Appl.* 6(4): 521-532. [Cock (1986)]
876. Chander, S. and Singh, Y. 1991. Effect of insecticides on white fly, *Bemisia tabaci* (Gennadius) and yellow mosaic virus in green gram, *Vigna radiata* (L) Wilczek. *Indian J. Entomol.* 53(2): 248-251.
877. Chandler, L.D. and Sumner, H.R. 1994. Evaluation of insecticides for control of sweetpotato whitefly (SPWF) on peanut, 1992. *Arthropod Management Tests* 19: 246.
878. Chang, Y.C. 1969. Host plant and morphological variation of *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae) in Taiwan. *Plant Prot. Bull. (Taiwan)* 11(1): 23-32. [Cock (1986), Chinese, English summary]
879. Channarayappa, G., Muniyappa, V., Schwegler-Berry, D. and Shivashankar, G. 1992. Ultrastructural changes in tomato infected with tomato leaf curl virus, a whitefly-transmitted geminivirus. *Canadian J. Bot.* 70(7-9): 1747-1753.

880. Channarayappa, S.G., Muniyappa, V. and Frist, R.H. 1992. Resistance of *Lycopersicum* species to *Bemisia tabaci*, a tomato leaf curl virus vector. *Canadian J. Bot.* 70(10-12): 2184-2192.
881. Chant, S.R. 1958. Studies on the transmission of cassava mosaic virus by *Bemisia* spp. (Aleyrodidae). *Ann. Appl. Biol.* 46: 210-215. [Cock (1986)]
882. Chao, S.E. 1997. Acetylcholine receptors as targets for imidacloprid and related insecticides. [University of California, Berkeley]. Dissertation Abst. Int. 58(07B): 3595.
883. Chao, S.L., Dennehy, T.J. and Casida, J.E. 1997. Whitefly (Hemiptera: Aleyrodidae) binding site for imidacloprid and related insecticides: A putative nicotinic acetylcholine receptor. *J. Econ. Entomol.* 90(4): 879-882.
884. Chari, M.S. and Rao, G.R. 1996. Influence of weather factors on the biology of whitefly, *Bemisia tabaci* Gennadius on brinjal. pp. 196-200. In S.C. Goel (Ed.), *Insect and Environment, VI: Integrated pest management and sustainable agriculture: An entomological approach*; Symposium, September 22-24, 1995. Uttar Pradesh Zool. Soc., Muzaffarnagar, India.
885. Chari, M.S., Rao, R.S.N., Prabhu, S.R. and Rao, S.G. 1990. Nicotine sulphate: An effective botanical pesticide. *Indian Farming* 34(11): 28-29.
886. Chatani, M. 1991. The nucleotide sequence and genome structure of the geminivirus miscanthus streak virus. *J. Gen. Virol.* 72: 2325-2331.
887. Chatchawankanphanich, O. 1997. Inhibition of geminiviral replication in a transient assay by expression of tomato leaf curl geminivirus REP gene antisense RNAs. [University of Wisconsin]. Dissertation Abst. Int. 58(06B): 2785.
888. Chatterjee, S.N. and Pandey, P.K. 1977. Susceptibility of sunflower cultivars to tobacco leafcurl virus transmitted by the whitefly *Bemisia tabaci*. *Indian J. Entomol.* 39: 259-261.
889. Chatterji, A. and Fauquet, C.M. 2000. Ecology of plant viruses, with special reference to whitefly-transmitted geminiviruses (WTCs). pp. 321-351. In C.J. Hurst (Ed.), *Viral Ecology*. Academic Press, San Diego, CA.
890. Chaudhary, H.R. and Dadheech, L.N. 1989. Incidence of insects attacking okra and avoidable losses caused by them. *Ann. Arid Zone* 28(3-4): 305-307. [Cock (1993)]
891. Chaudhary, J.P., Singh, A.K. and Yadav, L.S. 1981. Effect of disulfoton granules on pest complex, crop growth and yield of green gram. *Indian J. Entomol.* 43: 369-372. [Cock (1986)]
892. Chaudhary, R.R.P., Bhattacharya, A.K. and Rathore, R.R.S. 1977. Use of systemic granular insecticides for the control of stem miner, *Melanagromyza sojae* (Zehntner) and white fly, *Bemisia tabaci* Gennadius. *Indian J. Entomol.* 38(1976): 207-209. [Cock (1986)]
893. Chaudhary, R.R.P., Bhattacharya, A.K. and Rathore, R.R.S. 1981. Use of systemic insecticides for the control of stemfly *Melanagromyza sojae* (Zehnt.) and whitefly, *Bemisia tabaci* Genn. *Indian J. Entomol.* 43: 223-225. [Cock (1986)]
894. Chaudhuri, N., Deb, D.C. and Senapati, S.K. 2000. Evaluation of commonly grown tomato variety and hybrids in Terai region of West Bengal. *Environ. Ecol.* 18(4): 933-939.
895. Chaudhuri, N., Deb, D.C. and Senapati, S.K. 2001. Assessment of loss in yield caused by pest complex of tomato under terai region of West Bengal. *Res. Crops* 2(1): 71-79.
896. Chavan, V.M. 1983. Efficacy of systemic insecticides for the control of *Bemisia tabaci* Genn., a vector of the leaf-curl of cigar-wrapper tobacco. [tobacco leaf curl virus]. *Indian J. Agric. Sci.* 53: 585-589. [Cock (1986)]
897. Cheek, S. 1994. Assessing the potential use of biological control agents in the UK against plant pests of quarantine concern. pp. 175-182. In Brighton Crop Protection Conference: Pests and Diseases. British Crop Protection Council, Farnham, UK.
898. Cheek, S. 1996. *Bemisia tabaci* - the United Kingdom protected zone. pp. 397-398. In D. Gerling and R.T. Mayer (Eds.), *Bemisia* : 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
899. Cheek, S. 1999. Control options for eradication campaigns against quarantine pests: what now? *Bull. OEPP* 29(1-2): 55-61.
900. Cheek, S. and MacDonald, O. 1994. Statutory controls to prevent the establishment of *Bemisia tabaci* in the United Kingdom. *Pestic. Sci.* 42(2): 135-137.
901. Cheema, S.S., Sidhu, G.P.S. and Dhanju, K.C. 1997. Virus/virus-like diseases associated with tomato in Punjab: incidence, symptomatology and transmission tests. *J. Res. (Punjab Agric. Univ.)* 34(4): 398-401.
902. Chelliah, S., Murugesan, S., Sivakumar, C.V. and Ramakrishnan, L. 1976. Combination treatments for the control of insect pests, mite, virus vector, nematodes, fungal and viral diseases of bhendi, *Abelmoschus esculentus* (L.) Moench. *Madras Agric. J.* 63: 345-349. [Cock (1986)]
903. Chelliah, S., Murugesan, S., Sivakumar, C.V. and Ramakrishnan, L. 1976. Combination treatments for the control of virus vector, nematodes, fungal and viral diseases of tomato. *Madras Agric. J.* 63: 350-353. [Cock (1986)]
904. Chen, L. 1997. The damage and morphological variations of *Bemisia tabaci* (Gennadius) on ornamental plants. *J. Shanghai Agric. College* 15(3): 186-189.
905. Chen, W.W. and Feng, M.G. 1999. Current status in basic and applied research on the entomopathogenic fungus, *Paecilomyces fumosoroseus* Nat. *Enemies Insects* 21(3): 140-144. [Chinese, English summary]
906. Cheng, C.H. and Ho, S.C. 1997. Evaluation of the effects of ultraviolet-absorbing film on the population of insect pests and yield of muskmelon. *Plant Prot. Bull. (Taichung)* 39(4): 289-304. [Chinese]
907. Chenulu, V.V. and Phatak, H.C. 1965. Yellow mosaic of *Acalypha indica* L., a new white fly transmitted virus disease from India. *Curr. Sci. (Bangalore)* 34: 321-322. [Cock (1986)]
908. Cherkasov, V.A. 1986. [Effectiveness of the biological method in the greenhouse]. *Zashchita Rastenii (Moskva)* 2: 54-56. [Cock (1993), Russian]
909. Chermiti, B. 1991. Observations preliminaires sur la dynamique des populations de *Bemisia tabaci* (Genn.) (Homoptera, Aleyrodidae) sur une culture protegee d' aubergine. [Preliminary observations on population dynamics of *Bemisia tabaci* (Genn.) (Homoptera, Aleyrodidae) in eggplant protected crops]. *Bull. OILB/SROP (IOBC/WPRS)* 14: 89-97. [French]
910. Chermiti, B. 1994. Etude bio-ecologique de *Bemisia tabaci* (Genn.) sous tunnel plastique d' aubergine. [Bio-ecological study of *Bemisia tabaci* (Genn.) on plastic tunnels with eggplant crops]. *Bull. OILB/SROP (IOBC/WPRS)* 17: 89-95. [French]
911. Chermiti, B., Braham, M., Cenis, J.L., Alonso, C. and Beitia, F. 1997. Sur la presence en Tunisie des biotypes 'B' et 'non B' de *Bemisia tabaci* (Homoptera: Aleyrodidae) et de leurs parasitoides associes. [On the presence in Tunisia of the biotypes 'B' and 'non B' of *Bemisia tabaci* (Homoptera: Aleyrodidae) and of their associated parasitoids]. *Bull. OILB/SROP (IOBC/WPRS)* 20(4): 108-113. [French]
912. Chermiti, B., Onillon, J.C. and Aicha, B.B. 1993. Importance of tomato yellow leaf curl virus (TYLCV), transmitted by the whitefly *Bemisia tabaci* (Guen.) (Homoptera: Aleyrodidae) in late season cultivation in the irrigated area of Nabhana (Tunisia). *Med. Fac. Landbouww. Univ. Gent* 58(3a): 1057-1066. [French, English summary]
913. Cheung, P.S.R. and Roberts, C.W. 1980. Implications of disaccharides in sticky-cotton processing: Honeydew contamination. *Textile Res. J.* 50(1): 55-59.

914. Chew-Madinaveitia, Y.I., Zavaleta-Mejia, E., Delgadillo-Sanchez, F., Valdivia-Alcala, R., Pena-Martinez, M.R. and Cardenas-Soriano, E. 1995. Evaluacion de algunas estrategias de control de la virosis en el cultivo del chile (*Capsicum annuum* L.). [Evaluation of control strategies for virus diseases of pepper (*Capsicum annuum* L.)]. Fitopatologia 30(2): 74-84. [English summary]
915. Chhabra, K.S. and Kooner, B.S. 1980. Sources of whitefly, *Bemisia tabaci* G. and yellow mosaic virus resistance in *Vigna radiata* Willczek mung bean in India. Trop. Grain Legume Bull. 19: 26-29. [Cock (1986)]
916. Chhabra, K.S. and Kooner, B.S. 1981. Field resistance in black gram, *Vigna mungo* L. against insect-pests complex and yellow mosaic virus. Indian J. Entomol. 43: 288-293. [Cock (1986)]
917. Chhabra, K.S. and Kooner, B.S. 1993. Response of some promising mungbean genotypes towards whitefly, jassids and mungbean yellow mosaic virus. J. Insect Sci. 6(2): 215-218.
918. Chhabra, K.S. and Kooner, B.S. 1993. Screening of mungbean germplasm against insect pests and yellow mosaic virus. Indian J. Pulses Res. 6(1): 69-75.
919. Chhabra, K.S. and Kooner, B.S. 1994. Reaction of some promising mungbean genotypes towards whitefly, jassid and yellow mosaic virus. Pest Manag. Econ. Zool. 2(1): 11-14.
920. Chhabra, K.S., Kooner, B.S., Mahal, M.S. and Gill, A.S. 1983. The black aphid, *Aphis craccivora* Koch on pulses in Punjab. Pranikee 4: 251-258. [Cock (1986)]
921. Chhabra, K.S., Kooner, B.S., Saxena, A.K. and Sharma, A.K. 1981. Effect of biochemical components on the incidence of insect pest complex and yellow mosaic virus in mungbean. Crop Improvement 8: 56-59.
922. Chhabra, K.S., Kooner, B.S., Saxena, A.K. and Sharma, A.K. 1984. Influence of biochemical components on the incidence of insect pests and yellow mosaic virus in blackgram. Indian J. Entomol. 46(2): 148-156. [Cock (1993)]
923. Chhabra, K.S., Kooner, B.S., Sharma, A.K. and Saxena, A.K. 1987. Sources of resistance in mungbean (*Vigna radiata*) to insect pests and mungbean yellow mosaic virus. pp. 308-314. In S. Shanmugasundaram and B.T. McLean (Eds.), Mungbean, Proceedings Second International Symposium, Bangkok, Thailand 16-20 November 1987. Taipei, Taiwan.
924. Chhabra, K.S., Kooner, B.S., Sharma, A.K. and Saxena, A.K. 1993. Screening of blackgram genotypes against whitefly, jassids and yellow mosaic virus - role of phytochemicals in resistance. India J. Pulses Res. 6(1): 76-81.
925. Chhabra, K.S., Kooner, B.S. and Singh, G. 1981. Field resistance of certain cultivars of mungbean to whitefly, *Bemisia tabaci* G., and yellow-mosaic virus. J. Res. (Punjab Agric. Univ.) 16(4): 385-388. [Cock (1986)]
926. Chiaromonte, A. 1933. Considerazioni entomologiche sulla coltura delle piante ortensi nella Somalia Italiana. Agricoltura Coloniale 27: 523-529. [Cock (1986)]
927. Chiemsombat, P. 1992. Mungbean yellow mosaic disease in Thailand: a review. pp. 54-58. In S.K. Green and D. Kim (Eds.), Mungbean Yellow Mosaic Disease: Proceedings International Workshop, Bangkok, Thailand 2-3 July 1991. Asian Vegetable Research and Development Center, Taipei, Taiwan.
928. Chirinos, D.T. and Geraud-Pouey, F. 1996. Side effects of some insecticides on the entomofauna of tomato cultivars in the northeast region of the state of Zulia, Venezuela. Interciencia 21(1): 31-36, 54-56. [Spanish]
929. Cho, J.J., Mau, F.L., German, T.L., Hartmann, R.W., Yudin, L.S., Gonsalves, D. and Providenti, R. 1989. A multidisciplinary approach to management of tomato spotted wilt virus in Hawaii. Plant Dis. 73: 375-383.
930. Chorley, J.K. 1939. Report of the Division of Entomology for the year ending 31st December, 1938. Rhodesia Agric. J. 36: 598-622. [Cock (1986)]
931. Chortyk, O.T., Kays, S.J. and Teng, Q. 1997. Characterization of insecticidal sugar esters of petunia. J. Agric. Food Chem. 45(1): 270-275.
932. Chortyk, O.T., Pomonis, J.G. and Johnson, A.W. 1996. Syntheses and characterizations of insecticidal sucrose esters. J. Agric. Food Chem. 44(6): 1551-1557.
933. Chou, I. 1949. Listo de la konataj Aleurodoj "Homopteroj" en cinio. Entomol. Sinica 3(4): 18pp. [Cock (1986)]
934. Chowdhary, A.K., Biswas, B. and Saha, N.K. 1992. Inhibition of bhendi (okra) yellow vein mosaic virus (BYVMV) by different plant extracts. J. Mycopathol. Res. 30(2): 97-102.
935. Christie, R.G., Ko, N.J., Falk, B.W., Hiebert, E., Lastra, R., Bird, J. and Kim, K.S. 1986. Light microscopy of geminivirus-induced nuclear inclusion bodies. Phytopathology 76(1): 124-126. [Cock (1986)]
936. Chu, C.C., Cohen, A.C., Natwick, E.T., Simmons, G.S. and Henneberry, T.J. 1999. *Bemisia tabaci* (Homoptera: Aleyrodidae) biotype B colonization and leaf morphology relationships in upland cotton cultivars. Aust. J. Entomol. 38(2): 127-131.
937. Chu, C.C., Cohen, A.C., Natwick, E.T., Simmons, G.S. and Henneberry, T.J. 1999. Silverleaf whitefly colonization and leaf shape and vascular bundles relationships in cotton. pp. 909-910. In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
938. Chu, C.C., Freeman, T., Natwick, E.T., Buckner, J.S., Nelson, D.R. and Henneberry, T.J. 2000. *Bemisia argentifolii* adult, nymph and egg densities and egg distribution on selected upland cottons. J. Entomol. Sci. 35(1): 39-47.
939. Chu, C.C., Freeman, T.P., Buckner, J.S., Henneberry, T.J., Nelson, D.R. and Natwick, E.T. 2001. Susceptibility of upland cotton cultivars to *Bemisia tabaci* biotype B (Homoptera: Aleyrodidae) in relation to leaf age and trichome density. Ann. Entomol. Soc. Am. 94(5): 743-749.
940. Chu, C.C., Freeman, T.P., Buckner, J.S., Henneberry, T.J., Nelson, D.R., Walker, G.P. and Natwick, E.T. 2000. *Bemisia argentifolii* (Homoptera: Aleyrodidae) colonization on upland cottons and relationships to leaf morphology and leaf age. Ann. Entomol. Soc. Am. 93(4): 912-919.
941. Chu, C.C., Freeman, T.P., Buckner, J.S., Natwick, E.T., Henneberry, T.J. and Nelson, D.R. 2000. Silverleaf whitefly colonization and trichome density relationship on upland cotton cultivars. Southwest. Entomol. 25(4): 237-242.
942. Chu, C.C. and Henneberry, T.J. 1994. The effect of reflective plastic mulches and insecticides on silverleaf whitefly populations and broccoli production. pp. 137-146. In Proc. Amer. Soc. Plasticulture.
943. Chu, C.C. and Henneberry, T.J. 1996. Chemical control studies for silverleaf whitefly control. pp. 307-309. In Cotton, A College of Agriculture Report, Series P-103. University of Arizona, Tucson.
944. Chu, C.C. and Henneberry, T.J. 1997. Silverleaf whitefly on cotton. pp. 254-256. In Cotton, A College of Agriculture Report, Series P-108. University of Arizona, Tucson.
945. Chu, C.C. and Henneberry, T.J. 1998. Development of a new whitefly trap. J. Cotton Sci. 2: 104-109.
946. Chu, C.C. and Henneberry, T.J. 1998. Development of a silverleaf whitefly (Homoptera: Aleyrodidae) trap. Recent Res. Devel. Entomol. 2: 47-54.
947. Chu, C.C. and Henneberry, T.J. 1999. *Bemisia argentifolii*: action thresholds, upland cotton yields and cotton lint stickiness in the Imperial Valley, California. Southwest. Entomol. 24(2): 79-86.
948. Chu, C.C., Henneberry, T.J. and Akey, D.H. 1993. Insecticide control of sweetpotato whitefly on broccoli and lettuce 1991. Insecticide Acaricide Tests 18: 48-85.

949. Chu, C.C., Henneberry, T.J. and Akey, D.H. 1993. Results of field studies with insecticides for sweetpotato whitefly *Bemisia tabaci* control on cotton in the Imperial Valley, CA. p. 960. In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
950. Chu, C.C., Henneberry, T.J. and Akey, D.H. 1993. Sweetpotato whitefly (*Bemisia tabaci* Gennadius) control: field studies with insecticides on cotton in the Imperial Valley, CA. pp. 241-247. In Cotton, A College of Agriculture Report, Series P-94. University of Arizona, Tucson.
951. Chu, C.C., Henneberry, T.J. and Akey, D.H. 1993. Sweetpotato whitefly control on cantaloupe 1991. Insecticide Acaricide Tests 18: 116.
952. Chu, C.C., Henneberry, T.J. and Akey, D.H. 1993. Sweetpotato whitefly control on cotton. Insecticide Acaricide Tests 18: 220-221.
953. Chu, C.C., Henneberry, T.J. and Akey, D.H. 1994. Insecticide control of sweetpotato whitefly on broccoli and lettuce, 1992. Arthropod Management Tests 19: 57-59.
954. Chu, C.C., Henneberry, T.J. and Akey, D.H. 1994. Insecticide control of sweetpotato whitefly on cotton, 1992. Arthropod Management Tests 19: 221.
955. Chu, C.C., Henneberry, T.J. and Akey, D.H. 1994. Insecticide control of sweetpotato whitefly on spring cantaloupe, 1992. Arthropod Management Tests 19: 78-79.
956. Chu, C.C., Henneberry, T.J., Akey, D.H., Naranjo, S.E., Perkins, H.H., Prabhaker, N. and Mackey, B.E. 1995. Silverleaf whitefly: development of an action threshold for chemical control on cotton. pp. 873-874. In D.A. Richter and J. Armour (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
957. Chu, C.C., Henneberry, T.J., Akey, D.H., Prabhaker, N. and Perkins, H.H. 1994. A study on control action thresholds for *Bemisia tabaci* (Gennadius) on cotton. pp. 1239-1241. In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
958. Chu, C.C., Henneberry, T.J., Akey, D.H., Prabhaker, N. and Perkins, H.H. 1994. Sweetpotato whitefly (*Bemisia tabaci* Gennadius) population relationships to cotton yield and quality. pp. 304-307. In Cotton, A College of Agriculture Report, Series P-96. University of Arizona, Tucson.
959. Chu, C.C., Henneberry, T.J. and Boykin, M.A. 1998. Response of *Bemisia argentifolii* (Homoptera: Aleyrodidae) adults to white fluorescent and incandescent light in laboratory studies. Southwest. Entomol. 23(2): 169-181.
960. Chu, C.C., Henneberry, T.J., Boykin, M.A. and Cohen, A.C. 1997. A modified new whitefly trap (CC trap) to increase whitefly adult catches. pp. 1247-1248. In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
961. Chu, C.C., Henneberry, T.J. and Cohen, A.C. 1995. *Bemisia argentifolii* (Homoptera: Aleyrodidae): Host preference and factors affecting oviposition and feeding site preference. Environ. Entomol. 24(2): 354-360.
962. Chu, C.C., Henneberry, T.J. and Cohen, A.C. 1995. Host preference of silverleaf whitefly and factors associated with feeding site preference. pp. 280-286. In Cotton, A College of Agriculture Report, Series P-99. University of Arizona, Tucson.
963. Chu, C.C., Henneberry, T.J. and Cohen, A.C. 1996. Development and evaluation of a *Bemisia argentifolii* trap for survey and monitoring adult population density. pp. 1029-1031. In P. Dugger and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
964. Chu, C.C., Henneberry, T.J., Mackey, B.E. and Perkins, H.H. 1999. Effects of *Bemisia argentifolii* (Homoptera: Aleyrodidae) infestations on upland cotton yield and honeydew lint contamination. J. Agric. Assn. China 188: 94-105.
965. Chu, C.C., Henneberry, T.J. and Natwick, E.T. 1998. *Bemisia argentifolii* adults caught in CC whitefly traps at different trap heights and trap catch relationships to leaf-turn counts on cotton. Southwest. Entomol. 23(3): 259-268.
966. Chu, C.C., Henneberry, T.J., Natwick, E.T., Ritter, D. and Birdsall, S.L. 2001. Efficacy of CC traps and seasonal activity of adult *Bemisia argentifolii* (Homoptera: Aleyrodidae) in Imperial and Palo Verde Valleys, California. J. Econ. Entomol. 94(1): 47-54.
967. Chu, C.C., Henneberry, T.J., Perkins, H.H., Jr., Hendrix, D.L. and Steele, T. 1996. Sticky cotton and reduction of lint stickiness. pp. 1020-1022. In P. Dugger and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
968. Chu, C.C., Natwick, E.T., Brushwood, D.E., Henneberry, T.J. and Cohen, A.C. 1997. Susceptibility of upland cotton cultivars Deltapine 5461 and Louisiana 887 to silverleaf whitefly colonization. pp. 1064-1066. In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
969. Chu, C.C., Natwick, E.T., Cohen, A.C. and Henneberry, T.J. 1999. Relationship between upland cotton leaf shape, vascular bundles and *Bemisia argentifolii* (Homoptera: Aleyrodidae) colonization. pp. 418-422. In Cotton, A College of Agriculture Report, Series P-116. University of Arizona, Tucson.
970. Chu, C.C., Natwick, E.T., Cohen, A.C., Simmons, G.S., Brushwood, D.E. and Henneberry, T.J. 1998. *Bemisia argentifolii* colonization, vascular bundle depth relationships, lint yield and nymphal parasitism in selected deltapine cotton cultivars. Southwest. Entomol. 23(4): 293-299.
971. Chu, C.C., Natwick, E.T. and Henneberry, T.J. 1998. Effects of aldicarb on cotton insects and plant growth and yield. J. Agric. Assn. China 184: 98-108.
972. Chu, C.C., Natwick, E.T. and Henneberry, T.J. 2000. Silverleaf whitefly - trichome density relationships on selected upland cotton cultivars. pp. 290-295. In Cotton, A College of Agriculture Report, Series P-121. University of Arizona, Tucson.
973. Chu, C.C., Natwick, E.T. and Henneberry, T.J. 2000. Susceptibility of normal-leaf and okra-leaf shape cottons to silverleaf whiteflies and relationships to trichome densities. pp. 1157-1158. In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
974. Chu, C.C., Natwick, E.T., Henneberry, T.J., Cohen, A.C. and Castle, S.J. 1998. Cotton plant resistance to silverleaf whitefly as a management tool. pp. 1089-1091. In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
975. Chu, C.C., Natwick, E.T., Henneberry, T.J., Cohen, A.C. and Castle, S.J. 1998. Silverleaf whitefly cotton cultivar preference. pp. 362-366. In Cotton, A College of Agriculture Report, Series P-112. University of Arizona, Tucson.
976. Chu, C.C., Natwick, E.T., Henneberry, T.J. and Lee, R. 1998. Effects of pyrethroid insecticides alone and in mixtures on *Bemisia argentifolii* (Homoptera: Aleyrodidae) on cotton, cauliflower and broccoli yields. J. Agric. Assn. China 184: 57-66.
977. Chu, C.C., Natwick, E.T., Henneberry, T.J., Nelson, D.R., Buckner, J.S. and Freeman, T.P. 2001. Silverleaf whitefly populations and trichome densities on upland cottons. pp. 871-872. In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
978. Chu, C.C., Natwick, E.T., Henneberry, T.J., Nelson, D.R., Buckner, J.S. and Freeman, T.P. 2001. Silverleaf whitefly studies: effects of trichome density and leaf shape. pp. 296-298. In Cotton, A College of Agriculture Report, Series P-125. University of Arizona, Tucson.
979. Chu, C.C., Natwick, E.T., Perkins, H.H., Brushwood, D.E., Henneberry, T.J., Castle, S.J., Cohen, A.C. and Boykin, M.A. 1998. Upland cotton susceptibility of *Bemisia argentifolii* (Homoptera: Aleyrodidae) infestations. J. Cotton Sci. 2: 1-9.

980. Chu, C.C., Pinter, P.J., Henneberry, T.J., Umeda, K., Natwick, E.T., Wei, Y.A., Reddy, V.R. and Shrepatis, M. 2000. Use of CC traps with different trap base colors for silverleaf whiteflies (Homoptera: Aleyrodidae), thrips (Thysanoptera: Thripidae), and leafhoppers (Homoptera: Cicadellidae). *J. Econ. Entomol.* 93(4): 1329-1337.
981. Chudhry, H.S. and Gupta, P.C. 1970. Studies on the digestive system of *Bemisia gossypiperda*, M. & L. [*Bemisia tabaci* (Gennadius)] (Homoptera: Aleyrodidae). *Entomologist* 103((1282)): 49-52. [Cock (1986)]
982. Chun, D.T. and Brushwood, D.E. 1998. Heavy mechanical processing effects on cotton stickiness. pp. 1542-1544. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
983. Chung, M.L., Liao, C.H., Chen, M.J. and Chiu, R.J. 1985. [The isolation, transmission and host range of sweet potato leaf curl disease agent in Taiwan]. *Plant Prot. Bull. (Taiwan)* 27(4): 333-341. [Cock (1993), Chinese, English summary]
984. CIAT. 1987. Genetic improvement and related activities; Improvement of individual characters; Resistance to invertebrate pests. pp. 96-117. *In* Bean Program. Annual Report 1986. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia.
985. CIAT. 1988. Diseases and pests. 2. Viral pathogens. pp. 137-158. *In* Bean Program. Annual Report 1988. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia.
986. CIAT. 1991. Another victory over golden mosaic in Central America: new tolerant bean lines. [Otra victoria sobre el mosaico dorado en Centroamerica: nueva generacion de frijoles tolerantes]. Centro Internacional de Agricultura Tropical (CIAT) Report: Highlights for 1990 and Early 1991, Cali, Colombia [English and Spanish]
987. CIAT. 1992. Cassava entomology and acarology. pp. 119-171. *In* Cassava Program: Annual Report 1990. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia.
988. CIAT. 1996. Whiteflies in cassava and beans. Centro Internacional de Agricultura Tropical, Cali, Colombia, 61 pp.
989. CIAT. 1997. Output IV. Global whitefly research network reduce crop losses initiated. pp. 63-72. *In* Project PE-1: IPM for a Safer Environment: Integrated Pest Management in Major Agroecosystems in the Americas: Annual Report 1997. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia.
990. CIAT. 1998. Output 5. Analyses, scenarios, and options for sustainable land use in Latin America in general and in the CIAT priority agroecosystems. p. 5.1/95-5.5/111. *In* Project PE-4: Land Use Studies: Reconciling the Dynamics of Agriculture With the Environment. Annual Report 1998. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia.
991. CIAT. 1998. Sustainable integrated management of whiteflies as pests and vectors of plant viruses in the tropics: DANIDA projects: Phase 1: Network formation, diagnosis and analysis for integrated pest management of whiteflies in the tropics. Centro Internacional de Agricultura Tropical, Cali, Columbia, 131 pp.
992. CIAT. 1999. Output 6: Characterization of cassava germplasm for resistance/tolerance to major pests. pp. 83-117. *In* Project IP3: Improved Cassava for the Developing World: Annual Report 1999. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia.
993. CIAT. 1999. Output 7: Molecular markers for genes mapping and genetic diversity assessment in cassava. pp. 109-120. *In* Project IP3: Improved Cassava for the Developing World: Annual Report 1999. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia.
994. CIAT. 1999. Output IV. Global IPM networks and knowledge systems developed. pp. 119-130. *In* Project PE-1: Integrated Pest and Disease Management in Major Agroecosystems. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia.
995. CIBC Pakistan Station. 1983. Studies on potential biological control agents of whiteflies in Pakistan. March 1979-February 1982, Rawalpindi, Pakistan. CIBC Pakistan Station, 88 pp. [Cock (1986), Unpublished Report]
996. Cicero, J.M., Hiebert, E. and Webb, S.E. 1995. The alimentary canal of *Bemisia tabaci* and *Trialeurodes abutilonea* (Homoptera, Sternorrhynchi): histology, ultrastructure and correlations to function. *Zoomorphology* 115(1): 31-39.
997. Cicero, K. 1992. Lessons from the whitefly. *New Farm* 14(5): 29.
998. Cichocka, E. and Goszczynski, W. 1994. Nowe szkodniki upraw szklarniowych. [New species of pests in Polish glasshouses]. pp. 272-276. *In* S. Pruszyński and J.J. Lipa (Eds.), Materials 4th Research Session of Institute of Plant Protection. Instytut Ochrony Roslin, Poznan, Poland. [Polish, English summary]
999. Cisneros, F. and Mujica, N. 1999. Biological and selective control of the sweetpotato whitefly, *Bemisia tabaci* (Gennadius) (Hom.: Aleyrodidae). pp. 255-264. *In* Impact on a Changing World: International Potato Center Program Report 1997-1998. International Potato Center, Lima, Peru.
1000. Clark, M.A., Baumann, L., Munson, M.A., Baumann, P., Campbell, B.C., Duffus, J.E., Osborne, L.S. and Moran, N.A. 1992. The eubacterial endosymbionts of whiteflies (Homoptera: Aleyrodoidea) constitute a lineage distinct from the endosymbionts of aphids and mealybugs. *Curr. Microbiol.* 25: 119-123.
1001. Clausen, C.P. 1934. The natural enemies of Aleyrodidae in tropical Asia. *Philippine J. Sci.* 53: 253-265. [Cock (1986,1993)]
1002. Coates, W. 1996. Spraying technologies for cotton: deposition and efficacy. *Appl. Eng. Agric.* 12(3): 287-296.
1003. Coates, W. and Palumbo, J. 1997. Deposition off-target movement, and efficacy of Capture and Thiodan applied to cantaloupes using five sprayers. *Appl. Eng. Agric.* 13(2): 181-188.
1004. Coats, S.A., Brown, J.K. and Hendrix, D.L. 1994. Biochemical characterization of biotype-specific esterases in the whitefly, *Bemisia tabaci* Genn (Homoptera: Aleyrodidae). *Insect Biochem. Mol. Biol.* 24(7): 723-728.
1005. Cock, M.J.W. 1986. *Bemisia tabaci* - a literature survey on the cotton whitefly with an annotated bibliography. CAB International Institute of Biological Control, Ascot, UK, 121 pp.
1006. Cock, M.J.W. 1986. Other control methods. pp. 59-61. *In* M.J.W. Cock (Ed.), *Bemisia tabaci* - A Literature Survey on the Cotton Whitefly with an Annotated Bibliography. CAB International Institute of Biological Control, Ascot, UK.
1007. Cock, M.J.W. 1986. Population ecology. pp. 37-41. *In* Cock, M. J. W. (Ed.), *Bemisia tabaci* - A Literature Survey on the Cotton Whitefly with an Annotated Bibliography. CAB International Institute of Biological Control, Ascot, UK.
1008. Cock, M.J.W. 1986. Possibilities for classical biological control. pp. 63-72. *In* M.J.W. Cock (Ed.), *Bemisia tabaci* - A Literature Survey on the Cotton Whitefly with an Annotated Bibliography. CAB International Institute of Biological Control, Ascot, UK.
1009. Cock, M.J.W. 1986. Requirements for biological control: an ecological perspective. *Biocontrol News Information* 7: 7-16. [Cock (1986)]
1010. Cock, M.J.W. 1993. *Bemisia tabaci* an update 1986-1992 on the cotton whitefly with an annotated bibliography. CAB International Institute of Biological Control, Ascot, UK, 78 pp.
1011. Cock, M.J.W. 1994. Integrated management of whitefly pest problems in the Middle and Near East with special emphasis on biological control. *Arab J. Plant Prot.* 12(2): 127-136.
1012. Cohen, A. 1988. Cotton research in 1987: whitefly, irrigation and varieties (Summaries). *Hassadeh* 68(8): 1486-1491. [Hebrew]
1013. Cohen, A. and Hendrix, D. 1993. Demonstration and preliminary characterization of  $\alpha$ -amylase in sweetpotato whiteflies, *Bemisia tabaci* (Aleyrodidae: Homoptera). pp. 955-958. *In* D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
1014. Cohen, A. and Hendrix, D.L. 1994. Demonstration and preliminary characterization of  $\alpha$ -amylase in the sweetpotato whitefly, *Bemisia tabaci* (Aleyrodidae: Homoptera). *Comp. Biochem. Physiol.* 109(4): 593-601.

- 1015.Cohen, A., Newman, C.M. and Hendrix, D. 1993. Histochemical studies of feeding sites of *Bemisia tabaci* on cotton leaves. p. 959. In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 1016.Cohen, A.C. and Brummett, D.L. 1997. The non-abundant nutrient (NAN) concept as a determinant of predator-prey fitness. *Entomophaga* 42(1/2): 85-91.
- 1017.Cohen, A.C. and Byrne, D.N. 1992. *Geocoris punctipes* as a predator of *Bemisia tabaci* : a laboratory evaluation. *Entomol. Exp. Appl.* 64: 195-202.
- 1018.Cohen, A.C., Chu, C., Henneberry, T.J., Freeman, T., Buckner, J. and Nelson, D. 1996. Cotton leaf surface features serve as behavioral cues to the silverleaf whitefly. pp. 1017-1020. In P. Dugger and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 1019.Cohen, A.C., Chu, C.C., Henneberry, T.J., Freeman, T., Buckner, J. and Nelson, D. 1996. Cotton leaf surface features serve as behavioral cues to silverleaf whiteflies. *Southwest. Entomol.* 21(4): 377-385.
- 1020.Cohen, A.C., Chu, C.C., Henneberry, T.J., Freeman, T., Nelson, D., Buckner, J., Margosan, D., Vail, P. and Aung, L.H. 1998. Feeding biology of the silverleaf whitefly (Homoptera: Aleyrodidae). *Chinese J. Entomol.* 18(2): 65-82.
- 1021.Cohen, A.C., Hendrix, D.L. and Brown, J.K. 1992. Determination of trophic enzymes in *Bemisia tabaci* (Gennadius). pp. 951-952. In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 1022.Cohen, A.C., Henneberry, T.J. and Chu, C.C. 1996. Geometric relationships between whitefly feeding behavior and vascular bundle arrangements. *Entomol. Exp. Appl.* 78(2): 135-142.
- 1023.Cohen, A.C., Henneberry, T.J., Freeman, T., Margosan, D., Vail, P. and Chu, C. 1996. Microstructure of feeding in nymphal *Bemisia argentifolii* in cotton and cantaloupe. pp. 740-743. In P. Dugger and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 1024.Cohen, A.C., Jech, L.F., Newman, C. and Henneberry, T.J. 1992. Physiological ecology of putative strains A & B of *Bemisia tabaci* (Gennadius). pp. 948-950. In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 1025.Cohen, A.C. and Smith, L.K. 1998. A new concept in artificial diets for *Chrysoperla rufilabris*: the efficacy of solid diets. *Biol. Control* 13(1): 49-54.
- 1026.Cohen, A.C., Staten, R.T. and Henneberry, T.J. 1993. Evaluations of predators of sweetpotato whiteflies: laboratory and field cage studies. pp. 710-713. In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 1027.Cohen, J. 1993. Tomato yellow leaf curl virus is the causal agent of a new disease in *Lisianthus*. *Hassadeh* 74(2): 169-170, 197. [ Hebrew, English summary]
- 1028.Cohen, J., Gera, A., Ecker, R., Ben Joseph, R., Perlsman, M., Gokkes, M., Lachman, O. and Antignus, Y. 1995. *Lisianthus* leaf curl a new disease of *Lisianthus* caused by tomato leaf curl virus. *Plant Dis.* 79(4): 416-420.
- 1029.Cohen, J., Lapidot, M., Loebenstein, G. and Gera, A. 2001. First report of Sweet potato sunken vein virus occurring in *Lisianthus*. *Plant Dis.* 85(6): 679.
- 1030.Cohen, J. and Loebenstein, G. 1991. Role of a whitefly-transmitted agent in infection of sweet potato by cucumber mosaic virus. *Plant Dis.* 75(3): 291-292.
- 1031.Cohen, J., Milgram, M., Antignus, Y., Pearlsman, M., Lachman, O. and Loebenstein, G. 1997. Ipomoea crinkle leaf caused by a whitefly-transmitted gemini-like virus. *Ann. Appl. Biol.* 131(2): 273-282.
- 1032.Cohen, J., Vetten, A.F.H.J., Lesemann, D.E. and Lobenstein, G. 1992. Purification and properties of closterovirus-like particles associated with a whitefly-transmitted disease of sweet potato. *Ann. Appl. Biol.* 121: 257-268.
- 1033.Cohen, S. 1967. The occurrence in the body of *Bemisia tabaci* of a factor apparently related to the phenomenon of "periodic acquisition" of tomato yellow leaf curl virus. *Virology* 31: 180-183. [ Cock (1986)]
- 1034.Cohen, S. 1967. Transmission mechanism of tomato yellow leaf curl virus. *Summaries Lectures First Israel Congress Plant Pathology* (1967): 79. [ Cock (1986)]
- 1035.Cohen, S. 1969. In vivo effects in whiteflies of a possible antiviral factor. *Virology* 37: 448-454. [ Cock (1986)]
- 1036.Cohen S. 1982. Control of whitefly vectors of viruses by color mulches. pp. 45-56. In K.F. Harris and K. Maramorosch (Eds.), *Pathogens, Vectors and Plant Diseases: Approaches to Control*. Academic Press, New York.
- 1037.Cohen, S. 1990. Epidemiology of whitefly-transmitted viruses. pp. 211-225. In D. Gerling (Ed.), *Whiteflies: their Bionomics, Pest Status and Management*. Intercept, Andover, UK. [Cock (1993)]
- 1038.Cohen, S. and Antignus, Y. 1982. A noncirculative whitefly-borne virus affecting tomatoes in Israel. *Phytoparasitica* 10: 101-109. [ Cock (1986)]
- 1039.Cohen, S. and Antignus, Y. 1994. Tomato yellow leaf curl virus, a whitefly-borne geminivirus of tomatoes. *Adv. Disease Vector Res.* 10: 259-288.
- 1040.Cohen, S. and Berlinger, M.J. 1986. Transmission and cultural control of whitefly-borne viruses. *Agric. Ecosystems Environ.* 17(1-2): 89-97. [ Cock (1993)]
- 1041.Cohen, S., Duffus, J.E., Larsen, R.C., Liu, H.Y. and Flock, R.A. 1983. Purification, serology, and vector relationships of squash leaf curl virus, a whitefly-transmitted geminivirus. *Phytopathology* 73: 1669-1673. [ Cock (1986)]
- 1042.Cohen, S., Duffus, J.E. and Liu, H.Y. 1989. Acquisition, interference and retention of cucurbit leaf curl viruses in whiteflies. *Phytopathology* 79(1): 109-113. [ Cock (1993)]
- 1043.Cohen, S., Duffus, J.E. and Liu, H.Y. 1992. A new *Bemisia tabaci* biotype in the Southwestern United States and its role in silverleaf of squash and transmission of lettuce infectious yellows virus. *Phytopathology* 82: 86-90.
- 1044.Cohen, S., Duffus, J.E., Perry, R. and Dawson, R. 1989. A collection and marking system suitable for epidemiological studies on whitefly-borne viruses. *Plant Diseases.* 73(9): 765-768. [ Cock (1993)]
- 1045.Cohen, S., Eisenberg, M. and Sela, I. 1974. Antiviral effect of a factor from TMV-infected plants on the inoculativity of an insect vector. *Phytoparasitica* 2: 13-18. [ Cock (1986)]
- 1046.Cohen, S. and Harpaz, I. 1964. Periodic, rather than continual acquisition of a new tomato virus by its vector, the tobacco whitefly (*Bemisia tabaci* Gennadius). *Entomol. Exp. Appl.* 7: 155-166. [ Cock (1986)]
- 1047.Cohen, S., Kern, J., Harpaz, I. and Ben-Joseph, R. 1988. Epidemiological studies of the tomato yellow leaf curl virus (TYLCV) in the Jordan Valley, Israel. *Phytoparasitica* 16(3): 259-270. [ Cock (1993)]
- 1048.Cohen, S. and Marco, S. 1970. Periodic occurrence of an anti-TMV factor in the body of whiteflies carrying tomato yellow leaf curl virus. *Virology* 40: 363-368.
- 1049.Cohen, S. and Melamed-Madjar, V. 1978. Prevention by soil mulching of the spread of tomato yellow leaf curl virus transmitted by *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae) in Israel. *Bull. Entomol. Res.* 68: 465-470. [ Cock (1986)]
- 1050.Cohen, S., Melamed-Madjar, V. and Hameiri, J. 1974. Prevention of the spread of tomato yellow leaf curl virus transmitted by *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae) in Israel. *Bull. Entomol. Res.* 64: 193-197. [ Cock (1986)]



- 1051.Cohen, S. and Nitzany, F.E. 1960. A whitefly transmitted virus of cucurbits in Israel. *Phytopathol. Mediterr.* 1: 44-46.
- 1052.Cohen, S. and Nitzany, F.E. 1963. Identity of viruses affecting cucurbits in Israel. *Phytopathology* 53: 193-196. [Cock (1986)]
- 1053.Cohen, S. and Nitzany, F.E. 1966. Transmission and host range of the tomato yellow leaf curl virus. *Phytopathology* 56: 1127-1131. [Cock (1986)]
- 1054.Cohen, S., Nitzany, F.E. and Harpaz, I. 1963. Experiments in the control of yellow-top virus in tomatoes. *Hassadeh* 43: 576-578. [Cock (1986)]
- 1055.Cohen, S., Nitzany, F.E. and Vilda, T. 1961. The tomato yellow-top virus in Israel. *Hassadeh* 42: 139-140. [Cock (1986)]
- 1056.Cohic, F. 1966. Contribution a l'etude des aleurodes africains (1 Note). *Cahiers Office de la Res. Sci. et Tech. Outre-Mer (Serie Biologie)* 1: 3-59. [Cock (1986)]
- 1057.Cohic, F. 1966. Contribution a l'etude des aleurodes africains (2 Note). *Cahiers Office de la Res. Sci. et Tech. Outre-Mer (Serie Biologie)* 2: 13-72. [Cock (1986)]
- 1058.Cohic, F. 1968. Contribution a l'etude des aleurodes africains (4 Note). *Cahiers Office de la Res. Sci. et Tech. Outre-Mer (Serie Biologie)* 6: 63-143. [Cock (1986)]
- 1059.Cohic, F. 1969. Contribution a l'etude des aleurodes africains (5 Note). *Ann. Univ. Abidjan Serie E Ecol.* 2: 1-156. [Cock (1986)]
- 1060.Colinet, D., Kummert, J. and Lepoint, P. 1996. Molecular evidence that the whitefly-transmitted mild mottle virus belongs to a distinct genus of the Potyviridae. *Arch. Virol.* 141(1): 125-135.
- 1061.Colkesen, T. and Sekeroglu, E. 1987. [Development of *Bemisia tabaci* (Homoptera: Aleyrodidae) in relation to temperature]. *Türkiye Entomol. Dergisi* 11(3): 163-168. [Cock (1993), Turkish, English summary]
- 1062.Collier, T.R. and Hunter, M.S. 2001. Lethal interference competition in the whitefly parasitoids *Eretmocerus eremicus* and *Encarsia sophia*. *Oecologia* 129(1): 147-154.
- 1063.Colombo, M. 1993. Organismi ausiliari e possibilita tecniche nel contenimento di *Trialeurodes vaporariorum* (Westw.) e *Bemisia tabaci* (Genn.) in serre di *Euphorbia pulcherrima* Willd..[Control of *Trialeurodes vaporariorum* (Westw.) and *Bemisia tabaci* (Genn.) on *Euphorbia pulcherrima* Willd. by using natural enemies and other methods]. *Coltura Protette (Italy)* 22(Suppl. 1): 39-42. [Italian, English summary]
- 1064.Colombo, M. and Eoerdegh, R. 1992. Impiego di *Encarsia formosa* nel controllo di *Trialeurodes vaporariorum* e *Bemisia tabaci* in coltura di *Euphorbia pulcherrima*. [*Encarsia formosa* in controlling *Trialeurodes vaporariorum* and *Bemisia tabaci* in *Euphorbia pulcherrima* culture]. *Informatore Agrario (Italy)* 48(2): 51-53. [Italian]
- 1065.Colombo, M. and Eördegh, F.R. 1990. [Discovery of *Coenosia attenuata*, an active predator on aleyrodids, in protected crops in Liguria and Lombardia]. *Informatore Agrario* 47(10): 187-189. [Cock (1993), Italian]
- 1066.Colvin, J., Fishpool, L.D.C., Fargette, D., Sherington, J. and Fauquet, C. 1998. *Bemisia tabaci* (Homoptera: Aleyrodidae) trap catches in a cassava field in Cote d'Ivoire in relation to environmental factors and the distribution of African cassava mosaic disease. *Bull. Entomol. Res.* 88(4): 369-378.
- 1067.Comisión Nacional de Moscas Blancas. 1992. Las moscas blancas en Nicaragua. [The whiteflies in Nicaragua]. pp. 54-57. *In* L. Hilje and G. Arboleda (Eds.), *Las moscas blancas (Homoptera: Aleyrodidae) en America Central y el Caribe*. CATIE, Turrialba, Costa Rica. [Spanish]
- 1068.Comision Nacional del Algodon. 1977. Insecticidas para control de mosca blanca; *Bemisia tabaci* Ensayo no. 14. [Insecticides for the control of white fly; *Bemisia tabaci*: Test no. 14]. Report on Activities of the Entomology Section 1976-1977. Managua, Nicaragua, 10 pp. [Spanish]
- 1069.Commonwealth Institute of Entomology. 1971. Pest: *Bemisia tabaci* (Gennadius) Distribution Maps of Pests. Series A Map No. 284 [Cock (1986)]
- 1070.Conde, B., Young, G., Smith, S. and Gunning, R. 1995. Poinsettia whitefly. *Agnote (Darwin)* No. 649, 3pp.
- 1071.Conti, M.A. 1994. Whiteflies other than *Bemisia tabaci* as vectors of plant viruses. *Arab J. Plant Prot.* 12(2): 121-126.
- 1072.Cook, C.G. and Robinson, A.F. 1998. Reniform nematode and silverleaf whitefly influence on lint yield, fiber quality and seed quality of cotton. pp. 171-172. *In* P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
- 1073.Cook, C.G., Robinson, A.F., Namken, L.N. and Wolfenbarger, D.A. 1997. Effects of the reniform nematode and silverleaf whitefly on cotton. pp. 444-445. *In* P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
- 1074.Cook, C.G., Robinson, A.F. and Wolfenbarger, D.A. 1999. Silverleaf whitefly and reniform nematode effects on seed quality and stand establishment. pp. 489-490. *In* P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
- 1075.Cook, C.G., Wolfenbarger, D.A., Robinson, A.F. and Namken, L.N. 1996. Effect of reniform nematodes and silverleaf whiteflies on cottonseed quality. pp. 588-589. *In* P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
- 1076.Cook, M.T. 1931. New virus diseases of plants in Puerto Rico. *J. Dept. Agric. (Puerto Rico)* 15: 193-195. [Cock (1986)]
- 1077.Corbett, G.H. 1926. Contribution towards our knowledge of the Aleyrodidae of Ceylon. *Bull. Entomol. Res.* 16: 267-284. [Cock (1986)]
- 1078.Corbett, G.H. 1935. Malayan Aleurodidae. *J. Fed. Malay States Museums* 17: 722-852. [Cock (1986)]
- 1079.Corbett, G.H. 1935. On new Aleurodidae (Hem.). *Ann. Mag. Nat. Hist.* 16(10): 240-252. [Cock (1986)]
- 1080.Corbett, G.H. 1936. New Aleurodidae (Hem.). *Proc. Royal Entomol. Soc. London, (B)* 5: 18-22. [Cock (1986)]
- 1081.Cortes-Rodas, R.S. 1975. Evaluacion de insecticidas sistemicos para el control de mosca blanca (*Bemisia tabaci* Genn) vector del virus del mosaico dorado. [Evaluation of systemic insecticides to control the white fly (*Bemisia tabaci* Genn) vector of yellow mosaic virus]. pp. 327-335. *In* Memoir 21. Annual Meeting of the Central American Cooperative Program for Improving Food Crops, Santa Tecla, El Salvador, 7 April 1975. [Spanish]
- 1082.Costa, A.D. 1987. Fitovirose do feijoeiro no Brasil. pp. 173-257. *In* E.A. Bulisani (Ed.), *Feijão: Fatores de Produção e Qualidade*. Fundação Cargill, São Paulo, Brasil. [Portuguese]
- 1083.Costa, A.S. 1965. Three whitefly-transmitted virus diseases of beans in Sao Paulo, Brazil. *FAO Plant Prot. Bull.* 13(6): 121-130. [Cock (1986)]
- 1084.Costa, A.S. 1969. White flies as virus vectors. pp. 95-119. *In* K. Maramorosch (Ed.), *Viruses, Vectors, and Vegetation*. Interscience, New York, USA. [Cock (1986)]
- 1085.Costa, A.S. 1975. Increase in the populational density of *Bemisia tabaci*, a threat of widespread virus infection of legume crops in Brazil. pp. 27-29. *In* J. Bird and K. Maramorosch (Eds.), *Tropical Diseases of Legumes*. Academic Press, New York, USA. [Cock (1986)]
- 1086.Costa, A.S. 1976. Comparacao de machos e femeas de *Bemisia tabaci* na transmissao do mosaico dourado do feijoeiro. [Comparison of male and female *Bemisia tabaci* in the transmission of golden mosaic bean virus]. *Fitopatol. Brasileira* 1(2): 99-104. [CATIE (1992), Portuguese]
- 1087.Costa, A.S. 1976. Whitefly-transmitted plant diseases. *Annu. Rev. Phytopathol.* 14: 429-449. [Cock (1986)]

- 1088.Costa, A.S. 1980. Mosaico dourado do mussambe, uma invasora da familia Capparidaceae. Summa Phytopathol. 6(1-2): 3, 43. [ Cock (1986)]
- 1089.Costa, A.S. and Bennett, C.W. 1950. White-fly-transmitted mosaic of *Euphorbia prunifolia* . Phytopathology 40: 266-283. [ Cock (1986)]
- 1090.Costa, A.S. and Carvalho, A.M.B. 1960. Comparative studies between *Abutilon* and *Euphorbia* mosaic viruses. Phytopathol. Z. 38: 129-152. [ Cock (1986)]
- 1091.Costa, A.S., Costa, C.L. and Sauer, H.F.G. 1973. Surto de mosca-branca em culturas do Paraná e Sao Paulo [High populational increase of *Bemisia tabaci* on various crops in the States of Paraná and Sao Paulo]. Anais Soc. Entomol. Brasil 2: 20-30. [ Cock (1986), Portuguese, English summary]
- 1092.Costa, A.S., Gaspar, J.O. and Vega, J. 1983. Mosaico angular do feijoeiro jalo causado por um "carlavirus" transmitido pela mosca branca *Bemisia tabaci*. Fitopatol. Brasileira 8: 325-337. [ Cock (1986)]
- 1093.Costa, A.S. and Russell, L.M. 1975. Failure of *Bemisia tabaci* to breed on cassava plants in Brazil (Homoptera: Aleyrodidae). Ciencia Cultura (Sao Paulo) 27: 388-390. [ Cock (1986)]
- 1094.Costa, C.L. and Cupertino, F.P. 1977. Efelto repelente de background de casca di arroz sobre *Bemisia tabaci* vector del mosaico dourado do feijoeiro. [Repellent background effect of rice husks on *Bemisia tabaci* the vector of golden mosaic bean virus]. Fitopatol. Brasileira 2(1): 71-72. [ CATIE (1992), Portuguese]
- 1095.Costa, H.S. 1990. Effects of host plant on whitefly biology and behavior. [University of Arizona]. Dissertation Abst. Int. 51(06B): 2726.
- 1096.Costa, H.S. 1998. Interaction of *Beauveria bassiana* with various fungicides under exposed conditions, 1997. Arthropod Management Tests 23: 349.
- 1097.Costa, H.S., Bethke, J.A. and Redak, R.A. 1998. Efficacy of applications of acetamiprid and fipronil on the silverleaf whitefly under greenhouse conditions, 1997. Arthropod Management Tests 23: 349-350.
- 1098.Costa, H.S. and Brown, J.K. 1991. Biological characteristics and esterase patterns for *Bemisia tabaci* populations, and the association of silverleaf symptom development in squash with one population. pp. 79-86. In Cotton, A College of Agriculture Report, Series P-88. University of Arizona, Tucson.
- 1099.Costa, H.S. and Brown, J.K. 1991. Variation in biological characteristics and in esterase patterns among populations of *Bemisia tabaci* Genn. and the association of one population with silverleaf symptom development. Entomol. Exp. Appl. 61(3): 211-219. [ Cock (1993)]
- 1100.Costa, H.S., Brown, J.K. and Byrne, D.N. 1991. Host plant selection by the whitefly *Bemisia tabaci* (Gennadius)(Homoptera: Aleyrodidae) under greenhouse conditions. J. Appl. Entomol. 112(2): 146-152.
- 1101.Costa, H.S., Brown, J.K. and Byrne, D.N. 1991. Life history traits of the whitefly, *Bemisia tabaci* (Homoptera: Aleyrodidae) on six virus-infected or healthy plant species. Environ. Entomol. 20(4): 1102-1107.
- 1102.Costa, H.S., Brown, J.K., Sivasupramaniam, S. and Bird, J. 1993. Regional distribution, insecticide resistance, and reciprocal crosses between the 'A' and 'B' biotypes of *Bemisia tabaci*. Insect Sci. Appl. 14: 255-266.
- 1103.Costa, H.S. and Byrne, D.N. 1988. Neutron activation of a rare element to mark the sweetpotato whitefly *Bemisia tabaci* (Homoptera: Aleyrodidae). Ecol. Entomol. 13: 465-467.
- 1104.Costa, H.S., Henneberry, T.J. and Toscano, N.C. 1997. Effects of antibacterial materials on *Bemisia argentifolii* (Homoptera: Aleyrodidae) oviposition, growth, survival and sex ratio. J. Econ. Entomol. 90(2): 333-339.
- 1105.Costa, H.S., Johnson, M.W. and Ullman, D.E. 1994. Row covers effect on sweetpotato whitefly (Homoptera: Aleyrodidae) densities, incidence of silverleaf, and crop yield in zucchini. J. Econ. Entomol. 87(6): 1616-1621.
- 1106.Costa, H.S., Johnson, M.W., Ullman, D.E., Omer, A.D. and Tabashnik, B.E. 1993. Sweetpotato whitefly (Homoptera, Aleyrodidae) : Analysis of biotypes and distribution in Hawaii. Environ. Entomol. 22: 16-20.
- 1107.Costa, H.S. and Robb, K.L. 1999. Effects of ultraviolet -absorbing greenhouse plastic films on flight behavior of *Bemisia argentifolii* (Homoptera: Aleyrodidae) and *Frankliniella occidentalis* (Thysanoptera: Thripidae). J. Econ. Entomol. 92(3): 557-562.
- 1108.Costa, H.S., Toscano, N.C., Hendrix, D.L. and Henneberry, T.J. 1999. Patterns of honeydew droplet production by nymphal stages of *Bemisia argentifolii* (Homoptera: Aleyrodidae) and relative composition of honeydew sugars. J. Entomol. Sci. 34(3): 305-313.
- 1109.Costa, H.S., Toscano, N.C. and Henneberry, T.J. 1996. Mycetocyte inclusion in the oocytes of *Bemisia argentifolii* (Homoptera: Aleyrodidae). Ann. Entomol. Soc. Am. 89(5): 694-699.
- 1110.Costa, H.S., Ullman, D.E., Johnson, M.W. and Tabashnik, B.E. 1993. Antibiotic oxytetracycline interferes with *Bemisia tabaci* (Homoptera: Aleyrodidae) oviposition, development, and ability to induce squash silverleaf. Ann. Entomol. Soc. Am. 86(6): 740-748.
- 1111.Costa, H.S., Ullman, D.E., Johnson, M.W. and Tabashnik, B.E. 1993. Association between *Bemisia tabaci* density and reduced growth, yellowing, and stem blanching of lettuce and kai choy. Plant Dis. 77(10): 969-972.
- 1112.Costa, H.S., Ullman, D.E., Johnson, M.W. and Tabashnik, B.E. 1993. Squash silverleaf symptoms induced by immature, but not adult, *Bemisia tabaci*. Phytopathology 83: 763-766.
- 1113.Costa, H.S., Wescot, D.M., Ullman, D.E., Rosell, R.C., Brown, J.K. and Johnson, M.W. 1996. Virus-like particles in the mycetocytes of the sweetpotato whitefly, *Bemisia tabaci* (Homoptera, Aleyrodidae). J. Invertebr. Pathol. 67(2): 183-186.
- 1114.Costa, H.S., Westcot, D.M., Ullman, D.E. and Johnson, M.W. 1993. Ultrastructure of the endosymbionts of the whitefly, *Bemisia tabaci* and *Trialeurodes vaporariorum*. Protoplasma 176(3-4): 106-115.
- 1115.Costa, H.S., Westcot, D.M., Ullman, D.E., Rosell, R., Brown, J.K. and Johnson, M.W. 1995. Morphological variation in *Bemisia* endosymbionts. Protoplasma 189(3-4): 194-202.
- 1116.Costa, R.I.F., Almeida, S.A., Guerra, C.L. and Soares, J.J. 1999. Consumption of *Bemisia argentifolii* (Bellows & Perring) (Homoptera: Aleyrodidae) and *Aphis gossypii* (Glover, 1877) (Homoptera: Aphididae) by *Chrysoperla externa* (Hagen, 1861) (Neuroptera: Chrysopidae). pp. 256-258. In Anais Ii Congresso Brasileiro De Algodao: O Algodao No Seculo XX, Perspectivas Para a Seculo XXI, Ribeirao Preto, Sp, Brasil, 5-10 Setembro 1999. [Portuguese, English summary]
- 1117.Costa, R.I.F., Guerra, C.L., Almeida, S.A. and Soares, J.J. 1999. Food consumption of whitefly *Bemisia argentifolii* Bellows & Perring (Homoptera: Aleyrodidae) by *Chrysoperla externa* in different larval instars. pp. 244-246. In Anais Ii Congresso Brasileiro De Algodao: O Algodao No Seculo Xx, Perspectivar Para O Seculo Xxi, Ribeirao Preto, Sp, Brasil, 5-10 Setembro 1999. [Portuguese, English summary]
- 1118.Cota-Gomez, C., Rolsch, W. and Simmons, G. 1998. Introduccion de parasitoides exoticos de la especie *Eretmocerus emiratus* (Hymenoptera aphelinidae) contra mosquita blanca *Bemisia argentifolii* Bellows y Perring (Homoptera: aleyrodidae) en cultivo de algodonero y refugios en el valle de Mexicali, Baja California. [Introduction of exotic parasitoides of *Eretmocerus emiratus* (Hymenoptera aphelinidae) against white fly *Bemisia argentifolii* (Homoptera: aleyrodidae) in cotton and refuge area in Mexicali Valley, Baja California State]. pp. 182-185. In Proceedings 21st National Congress of Biological Control, Rio Bravo, Tampaulipas, Mexico, 5-6 Nov 1998. INIFAP [Spanish]

- 1119.Coudriet, D.L., Meyerdirk, D.E., Prabhaker, N. and Kishaba, A.N. 1986. Bionomics of sweetpotato whitefly (Homoptera:Aleyrodidae) on weed hosts in the Imperial Valley, California. *Environ. Entomol.* 15(6): 1179-1183. [ Cock (1993)]
- 1120.Coudriet, D.L., Prabhaker, N., Kishaba, A.N. and Meyerdirk, D.E. 1985. Variation in developmental rate on different hosts and overwintering of the sweetpotato whitefly, *Bemisia tabaci* (Homoptera: Aleyrodidae). *Environ. Entomol.* 14(4): 516-519. [ Cock (1986, 1993)]
- 1121.Coudriet, D.L., Prabhaker, N. and Meyerdirk, D.E. 1985. Sweetpotato whitefly (Homoptera: Aleyrodidae): Effects of neem-seed extract on oviposition and immature stages. *Environ. Entomol.* 14(6): 776-779. [ Cock (1993)]
- 1122.Couilloud, R. 1971. Elements de la biocenose du cotonnier en Iran. [English edition available]. *Coton Fibres Trop.* 26: 217-223. [ Cock (1986)]
- 1123.Couilloud, R. 1986. Bibliographical data on honeydew producing insects. *Coton Fibres Trop.* 41(3): 225-228. [ Cock (1993), English and French]
- 1124.Courshee, R.J. 1979. Whitefly survival rates in Sudanese cotton. pp. 1-12. *In Agricultural Aviation Research Unit. Progress Report 96/79.* Agricultural Aviation Research Unit, Cranfield, UK.
- 1125.Courshee, R.J. 1989. Numbers of whitefly in cotton in the Sudan and the use of DDT. *Int. Pest Control* 31: 38-40. [ Cock (1993)]
- 1126.Couteaux, L., Lefort, P.L. and Kuakuvi, E. 1968. Quelques observations sur le 'leaf-curl' du cotonnier chez *Gossypium barbadense* a la station d'Anie - Mono. *Coton Fibres Trop.* 23:506-507. [ Cock (1986)]
- 1127.Cowland, J.W. 1933. Gezira Entomological Section, G.A.R.S. Final report on experimental work, 1931-32. Rep. Gezira Agric. Res. Ser. (Anglo Egyptian Sudan) 1932: 93-112. [ Cock (1986)]
- 1128.Cowland, J.W. 1934. Gezira Entomological Section, G.A.R.S. Final report on experimental work, 1932-33. Rep. Gezira Agric. Res. Ser. (Anglo Egyptian Sudan) 1933: 107-125. [ Cock (1986)]
- 1129.Cowland, J.W. 1935. Gezira Entomological Section, G.A.R.S. Report on experimental work, 1933-34. Rep. Gezira Agric. Res. Ser. (Anglo Egyptian Sudan) 1934: 99-118. [ Cock (1986)]
- 1130.Crafts-Brandner, S.J. 2002. Plant nitrogen status rapidly alters amino acid metabolism and excretion in *Bemisia tabaci*. *J. Insect Physiol.* 48(1): 33-41.
- 1131.Crafts-Brandner, S.J. and Chu, C.C. 1999. Insect clip cages rapidly alter photosynthetic traits of leaves. *Crop Sci.* 39(6): 1896-1899.
- 1132.Crespi, S., Accotto, G.P., Caciagli, P. and Gronenborn, B. 1991. Use of digoxigenin-labeled probes for detection and host -range studies of tomato yellow leaf curl geminivirus. *Res. Virol.* 142(4): 283-288. [ Cock (1993)]
- 1133.Crowe, T.J. 1985. Field crop pests in Burma. An Annotated list. Rangoon, Burma; Office of the FAO Rep., 65 pp. [ Cock (1986)]
- 1134.Cruz Bernate, L. 1998. Sub pryecto de documentacion sobre las moscas blancas como plaga y vectores de virus de plantas en el tropico: informe final. Centro Internacional de Agricultura Tropical, Palmira, Valle Del Cauca Colombia, 48 pp. [ Spanish]
- 1135.Csizinszky, A.A., Schuster, D.J. and Kring, J.B. 1995. Color mulches influence yield and insect pest populations in tomatoes. *J. Am. Soc. Hortic. Sci.* 120(5): 778-784.
- 1136.Csizinszky, A.A., Schuster, D.J. and Kring, J.B. 1997. Evaluation of color mulches and oil sprays for yield and for the control of silverleaf whitefly, *Bemisia argentifolii* (Bellows and Perring) on tomatoes. *Crop Prot.* 16(5): 475-481.
- 1137.Csizinszky, A.A., Schuster, D.J. and Polston, J.E. 1999. Effect of ultraviolet-reflective mulches on tomato yields and on the silverleaf whitefly. *Hortscience* 34(5): 911-914.
- 1138.Csizinszky, A.A., Schuster, D.J., Polston, J.E. and Ben Yehoshua, S. 1998. Influence of color and reflective mulches on tomato (*Lycopersicon esculentum* Mill.) yields and on the silverleaf whitefly (*Bemisia argentifolii* (Bellows and Perring)). pp. 111-117. *In Proceedings of the 14th International Congress on Plastics in Agriculture.* Laser Pages Publishing, Jerusalem, Israel.
- 1139.Cuadrado, I.M., Janssen, D., Velasco, L., Ruiz, L. and Segundo, E. 2001. First report of Cucumber vein yellowing virus in Spain. *Plant Dis.* 85(3): 336.
- 1140.Cubillo, D., Chacón, A. and Hilje, L. 1994. Produccion de plantulas de tomate sin geminivirus transmitidos por la mosca blanca (*Bemisia tabaci*). [Production of tomato plants free from whitefly-borne geminivirus (*Bemisia tabaci*)]. *Manejo Integrado de Plagas (Costa Rica)* 34: 23-27. [ Spanish, English summary]
- 1141.Cubillo, D. and Hilje, L. 1996. Repelentes. pp. 77-83. *In* L. Hilje (Ed.), *Metodologias para el estudio y manejo de moscas blancas y geminivirus.* Centro Agronomico Tropical de Investigacion y Ensenanza, Turrialba, Costa Rica.
- 1142.Cubillo, D., Larriva, W., Quijije, R., Chacon, A. and Hilje, L. 1994. Evaluacion de la repelencia de varias sustancias sobre la mosca blanca, *Bemisia tabaci* (Homoptera: Aleyrodidae). [Evaluation of repellency of various substances to whitefly *Bemisia tabaci* (Homoptera: Aleyrodidae)]. *Manejo Integrado de Plagas (Costa Rica)* 33: 26-28. [ Spanish, English summary]
- 1143.Cubillo, D., Sanabria, G. and Hilje, L. 1997. Mortalidad de adultos de *Bemisia tabaci* con extractos de hombre grande (*Quassia amara*). [Mortality of *Bemisia tabaci* adults by bitterwood (*Quassia amara*) extracts]. *Manejo Integrado de Plagas (Costa Rica)* 45: 25-29. [ Spanish, English summary]
- 1144.Cubillo, D., Sanabria, G. and Hilje, L. 1999. Eficacia de coberturas vivas para el manejo de *Bemisia tabaci* como vector de geminivirus, en tomate. [Efficiency of living ground covers for the management of *Bemisia tabaci* as a vector of geminivirus in tomato. *Manejo Integrado de Plagas (Costa Rica)* 51: 10-20. [ Spanish, English summary]
- 1145.Cubillo, D., Sanabria, G. and Hilje, L. 1999. Evaluación de la repelencia y mortalidad causada por insecticidas comerciales y extractos vegetales sobre *Bemisia tabaci*. [Evaluation of repellency and mortality caused by commercial insecticides and plant extracts on *Bemisia tabaci*]. *Manejo Integrado de Plagas (Costa Rica)* 53: 65-71. [ Spanish, English summary]
- 1146.Cubillo, D., Sanabria, G. and Hilje, L. 1999. Evaluación de recipientes y mallas para el manejo de *Bemisia tabaci* mediante semilleros cubiertos, en tomate. [Evaluation of containers and nets for the management of *Bemisia tabaci* through covered seedbeds on tomato]. *Manejo Integrado de Plagas (Costa Rica)* 51: 29-35. [ Spanish, English summary]
- 1147.Culliney, T.W. and Nagamine, W.T. 2000. Introductions for biological control in Hawaii, 1987-1996. *Proc. Hawaiian Entomol. Soc.* 34: 121-133.
- 1148.Culotta, E. 1991. Biological immigrants under fire. *Science* 254: 1445-1447.
- 1149.Cyprus. 1977. Annual Report for 1976. Rep. Cyprus Agric. Res. Inst. 40-47. [ Cock (1986)]
- 1150.Cyprus. 1979. Annual Report for 1978. Rep. Cyprus Agric. Res. Inst. 48-50. [ Cock (1986)]
- 1151.Cyprus. 1980. Annual report for 1979. Rep. Cyprus Agric. Res. Inst. 33-39. [ Cock (1986)]
- 1152.Cyprus. 1981. Annual report for 1980. Rep. Cyprus Agric. Res. Inst. 34-37. [ Cock (1986)]
- 1153.Czosnek, H., Ber, R., Antignus, Y., Cohen, S., Navot, N. and Zamir, D. 1988. Isolation of tomato yellow leaf curl virus, a geminivirus. [German summary]. *Phytopathology* 78(5): 508-511. [ Cock (1993)]
- 1154.Czosnek, H., Ber, R., Navot, N., Antignus, Y., Cohen, S. and Zamir, D. 1989. Tomato leafcurl virus DNA forms in the viral capsid, in infected plants and in the insect vector. *J. Phytopathol.* 125(1): 47-54. [ Cock (1993)]

- 1155.Czosnek, H., Ber, R., Navot, N., Zamir, D., Antignus, Y. and Cohen, S. 1988. Detection of tomato yellow leaf curl virus in lysates of plants and insects by hybridization with a viral DNA probe. *Plant Dis.* 72(11): 949-951. [ Cock (1993)]
- 1156.Czosnek, H., Kheyr-pour, A., Gronenborn, B., Remetz, E., Zeidan, M., Rabinowitch, H.D., Vidavsky, S., Kedar, N., Gafni, Y. and Zamir, D. 1993. Replication of tomato yellow leaf curl virus (TYLCV) DNA in agroinoculated leaf discs from selected tomato genotypes. *Plant Mol. Biol.* 22(6): 995-1005.
- 1157.Czosnek, H. and Laterrot, H. 1997. A worldwide survey of tomato yellow leaf curl viruses. *Arch. Virol.* 142: 1391-1406.
- 1158.Czosnek, H., Navot, N. and Laterrot, H. 1990. Geographical distribution of tomato yellow leaf curl virus: a first survey using a specific DNA probe. *Phytopathol. Mediterr.* 29: 1-6. [ Ioannou (1995)]
- 1159.Czosnek, H., Navot, N., Sauri, D., Ovadia, S., Silberstein, A., Reinhartz, A. and Herzberg, M. 1989. [Identification of yellow top virus in tomato plants and in the tobacco white fly *Bemisia tabaci*]. *Hassadeh* 70(1): 60-61. [ Cock (1993), Hebrew]
- 1160.Czosnek, H., Zeidan, M., Ekstein, I., Zur-kunik, T., Gafni, Y., Gronenborn, B. and Zamir, D. 1994. Tomato yellow leaf curl virus, a geminivirus with a single genomic component: molecular analysis of infection and new ways for tomato protection. *Acta Hort.* 377: 251-257.
- 1161.Czosnek, H.K., Morin, S., Rubinstein, G., Fridman, V., Zeidan, M. and Ghanim, M. 2001. Tomato yellow leaf curl virus: a disease sexually transmitted by whiteflies. pp. 1-27. *In* K.F. Harris, O.P. Smith and J.E. Duffus (Eds.), *Virus-Insect-Plant Interactions*. Academic Press, San Diego, CA.
- 1162.d'Araujo-Silva, A.G., Goncalves, C.R., Galvao, D.M., Goncalves, A.J.L., Gomes, J., Silva, M. and Simoni, L. 1968. Quarto catalogo dos insetos que vivem nas plantas do Brasil seus Parasitos e predadores. Parte II 1 Tomo. pp. 117-118. *In* Rio de Janeiro, Brazil; Ministerio da Agric.
- 1163.Da Costa Lima, A. 1936. Terceiro catalogo dos insectos que vivem nas plantas do Brasil. Rio de Janeiro, Brazil; Ministerio da Agric., 460 pp. [ Cock (1986)]
- 1164.Da Silva, G.G., Praca, E.F., Menezes, J.B., Gomes, J., Jr. and Vieira, C.P.G. 2000. Qualidade de hibridos de melao apos a aplicacao de imidacloprid para controle de mosca-branca. [Fruit quality of hybrid melons after application of imidacloprid to control whitefly]. *Hortic. Brasileira* 18(3): 179-182. [ Portuguese, English summary]
- 1165.Dabi, R.K. and Gour, H.N. 1988. Field screening of mothbean (*Vigna aconitifolia*) for susceptibility to insect pests and diseases. *Indian J. Agric. Sci.* 58(11): 843-844. [ Cock (1993)]
- 1166.Dabire, R.A., Traore, S.N., Dabire, C.B. and Dicko, I.O. 1999. Effect of buprofezin, an insect growth regulator, on *Bemisia tabaci* (Homoptera: Aleyrodidae) on tomato. pp. 803-810. *In* Proceedings of the Fifth International Conference on Pests in Agriculture, Part 3, Montpellier, France, 7-9 December, 1999. Association Nationale pour la Protection des Plantes (ANPP), Paris, France. [French, English summary]
- 1167.Dafalla, G.A. and Siddig, S.A. 1997. Sudan. pp. 57-65. *In* N. Ioannou (Ed.), *Management of the whitefly -virus complex*. FAO Plant Production and Protection Paper 143, Rome. Italy.
- 1168.Dahal, G., Thapa, R.B. and Dangol, D.R. 1993. Epidemics of geminivirus diseases, monitoring and partial characterization of their suspected whitefly vector *Bemisia tabaci* in Nepal. *J. Inst. Agric. Anim. Sci.* 14: 55-68.
- 1169.Dahiya, A.S. and Singh, R. 1982. Bio-efficacy of some systemic insecticides against jassid, thrips and white fly attacking cotton. *Pesticides* 16: 13-14,22. [ Cock (1986)]
- 1170.Dalmon, A., Cailly, M. and David, C. 2000. Comparison of serological and molecular techniques for detection of Tomato yellow leaf curl begomovirus. *Bull. OEPP* 30(3-4): 457-462.
- 1171.Daly, P. 1999. Vegetable crops in New Caledonia: A report on diseases and pests. *Phytoma (France)* 519: 28-31. [ French, English summary]
- 1172.Dankowska, E. and Baranowski, T. 1997. Mozliwosci chemicznego zwalczania maczlika ostroskrzydlego (*Bemisia tabaci* Gennadius) na gwiezzdzie betlejmskiej (*Euphorbia pulcherrima* Willd.). [Possibilities of chemical control of the greenhouse whitefly (*Bemisia tabaci* Gennadius) on poinsettia (*Euphorbia pulcherrima* Willd.)]. *Roczniki Akademii Rolniczej w Poznaniu, Ogrodnictwo* 25: 23-27. [ Polish, English summary]
- 1173.Dantre, R.K., Keshwal, R.L. and Khare, M.N. 1996. Studies on a yellow mosaic disease of soybean. *Ann. Plant Prot. Sci.* 4(1): 67-70.
- 1174.Dantsig, E.M. 1964. Order Homoptera. Suborder Aleyrodinea. pp. 608-616. *In* G.Y. Bei-Bienko (Ed.), *Keys to the Insects of the European USSR*. [Cock (1986), Russian, English translation in 1967 by Israel Program for Scientific Translations Ltd., Jerusalem. Aleyrodinea]
- 1175.Dantsig, E.M. 1964. The whiteflies (Homoptera: Aleyrodidae) of the Caucasus. *Entomol. Obozrenie* 43: 633-646. [ Cock (1986), Russian, English translation in *Entomol. Rev.* 43:325-330]
- 1176.Dantsig, E.M. 1966. The whiteflies (Homoptera: Aleyrodoidea) of the Southern Primor'ye (Soviet Far East). *Entomol. Obozrenie* 45: 364-386. [ Cock (1986), Russian, English translation in *Entomol. Rev.* 45 (2):197-209]
- 1177.Dantsig, E.M. and Shenderovskaya, L.P. 1988. [Cotton whitefly]. *Zashchita Rastenii (Moskva)* 12: 40. [ Cock (1993), Russian]
- 1178.Daoud, M.A.R. 1996. Reduction in population density of the white fly, *Bemisia tabaci* after natural oil spray in tomato fields. *Al-Azhar J. Agric. Res. (Egypt)* 17: 21-28.
- 1179.Darby, A.C., Birkle, L.M., Turner, S.L. and Douglas, A.E. 2001. An aphid-borne bacterium allied to the secondary symbionts of whitefly. *Fems Microbiol. Ecol.* 36(1): 43-50.
- 1180.Dardón, A.D.E. 1992. Las moscas blancas en Guatemala. [The whiteflies in Guatemala]. pp. 38-41. *In* L. Hilje and O. Arboleda (Eds.), *Las moscas blancas (Homoptera:Aleyrodidae) en America Central y el Caribe*. CATIE, Turrialba, Costa Rica. [Spanish]
- 1181.Dardon, D., Calderon, F., Salguero, V., Lastra, R. and Brown, J. 1994. Identificacion de virus en el tomate (*Lycopersicon esculentum* L.) en cinco Departamentos de Guatemala. [Virus identification in tomatoes (*Lycopersicon esculentum* L.) in five Departments of Guatemala]. *Agron. Mesoamericana* 5: 109-117. [ Spanish, English summary]
- 1182.Darriet, F. 2000. Ebola and Marburg viruses; an entomological hypothesis to be confirmed. *Medecine Trop.* 60(3): 303-304. [ French]
- 1183.Darwish, Y.A., Abdel-Galil, F.A. and Younis, A.M. 1987. Population dynamics of the red scale insect *Aonidiella aurantii* Mask, the whitefly *Bemisia tabaci* (Gennadius) and the leaf hopper *Empoasca lybica* De-Berg on *Zizyphus* trees in upper Egypt (Homoptera: Diaspididae, Aleyrodidae, Cicadellidae). [Arabic summary]. *Assiut J. Agric. Sci. (Egypt)* 18(1): 267-278. [ Cock (1993)]
- 1184.Darwish, Y.A. and Farghal, A.I. 1990. Evaluation of certain pesticides activity against the cotton whitefly, *Bemisia tabaci* and associated natural enemies on cotton plants under field conditions in Assiut. *Assiut J. Agric. Sci. (Egypt)* 21(5): 331-339. [ Cock (1993)]
- 1185.Darwish, Y.A., Mannaa, S.H. and Abdel-Rahman, M.A.A. 2000. Effect of constant temperatures on the development of egg and nymphal stages of the cotton whitefly, *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae), and use of thermal requirements in determining its annual generation numbers. *Assiut J. Agric. Sci. (Egypt)* 31(1): 207-216.
- 1186.Dasgupta, B. and Chowdhury, A.K. 1985. Use of intervarietal intercropping to minimize the yellow mosaic virus of urid and mung beans. *Indian J. Plant Pathol.* 3(1): 100-101. [ Cock (1993)]
- 1187.Dash, P.C. 1989. Assessment of yield loss in groundnut due to whitefly in Orissa. *Madras Agric. J.* 76(5): 280-281.

- 1188.Dashad, S.S., Malipatil, M.V., Sharma, P.D. and Chaudhary, O.P. 2001. Relative performance of different sprayers, nozzles and spray volumes in controlling insect -pests of cotton in Haryana. *Crop Res. (Hisar)* 21(3): 324-331.
- 1189.Datar, V.V. 1980. Chemical control of chilli leaf curl complex in Maharashtra. *Pesticides* 14(9): 19-20.
- 1190.Datkar, D.S. and Dethé, M.D. 1994. Bioefficacy of new formulations of carbaryl, quinalphos and fluralinate against sucking pests of cotton. *Plant Prot. Bull. (Faridabad)* 46(2/3): 17-20.
- 1191.David, B.V. and Ananthkrishnan, T.N. 1976. Host correlated variation in *Trialeurodes rara* Singh and *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae: Insecta). *Curr. Sci. (Bangalore)* 45(6): 223-225. [Cock (1986)]
- 1192.David, B.V. and Jesudasan, R.W.A. 1986. Status of the cotton whitefly *Bemisia tabaci* (Gennadius) excluding its vector biology. *Pesticides* 20(7): 42-47. [Cock (1993)]
- 1193.David, B.V., Jesudasan, R.W.A. and Winstone, A.A. 1987. Effect of insecticides on the population build-up of *Bemisia tabaci* (Gennadius) on cotton. pp. 125-128. *In* S. Jayaraj (Ed.), Resurgence of Sucking Pests: Proceedings of National Symposium. Centre For Plant Protection Studies, Tamil Nadu Agric. Univ.
- 1194.David, B.V., Jesudasan, R.W.A. and Winstone, A.A. 1987. On the outbreak of *Bemisia tabaci* (Gennadius) on cotton and brinjal in South India. pp. 116-124. *In* S. Jayaraj (Ed.), Resurgence of Sucking Pests: Proceedings of National Symposium. Centre For Plant Protection Studies, Tamil Nadu Agric. Univ.
- 1195.David, B.V. and Subramanian, T.R. 1976. Studies on some Indian Aleyrodidae. *Record Zool. Survey of India* 70: 133-233. [Cock (1986)]
- 1196.David, B.V. and Winstone, A.A. 1988. A new whitefly *Bemisia graminis* sp. nov (Aleyrodidae: Homoptera) from India. *Entomol* 13(1): 33-35. [Cock (1993)]
- 1197.Davidson, E.W., Fay, M.L., Blackmer, J. and Lavine, M. 2000. Improved artificial feeding system for rearing the whitefly *Bemisia argentifolii* (Homoptera : Aleyrodidae). *Florida Entomol.* 83(4): 459-468.
- 1198.Davidson, E.W., Patron, R., Vey, A., Frutos, R., St. Leger, R., Lacey, L.A. and Hendrix, D. L. 1996. Effect of destruxins from *Metarhizium anisopliae*, ivermectin and *Bacillus thuringiensis* deltaendotoxins on adult silverleaf whitefly. pp. 739-740. *In* P. Dugger and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 1199.Davidson, E.W., Patron, R.B.R., Lacey, L.A., Frutos, R., Vey, A. and Hendrix, D.L. 1996. Activity of natural toxins against the silverleaf whitefly, *Bemisia argentifolii*, using a novel feeding bioassay system. *Entomol. Exp. Appl.* 79(1): 25-32.
- 1200.Davidson, E.W., Rosell, R.C. and Hendrix, D.L. 2000. Culturable bacteria associated with the whitefly, *Bemisia argentifolii* (Homoptera: Aleyrodidae). *Florida Entomol.* 83(2): 159-171.
- 1201.Davidson, E.W., Segura, B.J., Steele, T. and Hendrix, D.L. 1994. Microorganisms influence the composition of honeydew produced by the silverleaf whitefly, *Bemisia argentifolii*. *J. Insect Physiol.* 40(12): 1069-1076.
- 1202.Davies, J.W., Stanley, J., Donson, J., Mullineaux, P.M. and Boulton, M.I. 1987. Structure and replication of geminivirus genomes. *J. Cell Sci. Suppl.* 7: 95-107.
- 1203.Dávila, A.G.H. 1999. La mosca blanca (Homoptera: Aleyrodidae) en Guatemala. pp. 125-126. *In* 7th Taller Latinoamericano y del Caribe Sobre Moscas-Blancas y Geminivirus. IPA, Recife, PE, Brazil. [Spanish]
- 1204.Davino, M., Urso, F.D., Aredia, R., Carbone, M. and Mauromicale, G. 1994. Investigations on the epidemiology of tomato yellow leaf curl virus (TYLCV) in Sicily. [Italian Summary]. *Petria* 4(2): 151-160.
- 1205.Davis, E.F. 1929. Some chemical and physiological studies on the nature and transmission of 'infectious chlorosis' in variegated plants. *Ann. Missouri Bot. Gardens* 16: 145-227.
- 1206.Dawood, M.Z. 1999. Susceptibility of certain commonly cultivated squash and cucumber cultivars to *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae) in Beni-Suef Governorate. *Egyptian J. Agric. Res.* 77(3): 1075-1080.
- 1207.Dawood, M.Z., El-Rafie, K.K., Aly, S.A. and Hydar, M.F. 1999. Susceptibility of some tomato varieties and hybrids to whitefly *Bemisia tabaci* (Genn.) infestation in relation to rate of TYLCV infection and the yield. *Egyptian J. Agric. Res.* 77(3): 1059-1065.
- 1208.Dawson, W.O. and Hilf, M.E. 1992. Host-range determinants of plant viruses. *Annu. Rev. Plant Physiol.* 43: 527-555.
- 1209.De Arruda, G.P. 1998. Técnicas de preparacao de mosca branca para identificacao [Techniques for preparing whiteflies for identification]. *Manejo Integrado De Plagas (Costa Rica)* 47: 44-46. [Spanish, English summary]
- 1210.De Barro, P.J. 1995. *Bemisia tabaci* biotype B: a review of its biology, distribution and control. CSIRO Div. Entomol. Tech. Paper No. 33, 58 pp.
- 1211.De Barro, P.J. and Driver, F. 1997. Use of RAPD PCR to distinguish the B biotype from other biotypes of *Bemisia tabaci* (Gennadius) (Hemiptera: Aleyrodidae). *Aust. J. Entomol.* 36(2): 149-152.
- 1212.De Barro, P.J., Driver, F., Naumann, I.D., Schmidt, S., Clarke, G.M. and Curran, J. 2000. Descriptions of three species of *Eretmocerus* Haldeman (Hymenoptera : Aphelinidae) parasitising *Bemisia tabaci* (Gennadius) (Hemiptera : Aleyrodidae) and *Trialeurodes vaporariorum* (Westwood) (Hemiptera : Aleyrodidae) in Australia based on morphological and molecular data. *Aust. J. Entomol.* 39: 259-269.
- 1213.De Barro, P.J., Driver, F., Trueman, J.W.H. and Curran, J. 2000. Phylogenetic relationships of world populations of *Bemisia tabaci* (Gennadius) using ribosomal ITS1. *Mol. Phylogenet. Evol.* 16(1): 29-36.
- 1214.De Barro, P.J. and Hart, P.J. 2000. Mating interactions between two biotypes of the whitefly, *Bemisia tabaci* (Hemiptera : Aleyrodidae) in Australia. *Bull. Entomol. Res.* 90(2): 103-112.
- 1215.De Barro, P.J. and Hart, P.J. 2001. Antibiotic curing of parthenogenesis in *Eretmocerus mundus* (Australian parthenogenic form). *Entomol. Exp. Appl.* 99(2): 225-230.
- 1216.De Barro, P.J., Hart, P.J. and Morton, R. 2000. The biology of two *Eretmocerus* spp. (Haldeman) and three *Encarsia* spp. Forster and their potential as biological control agents of *Bemisia tabaci* biotype B in Australia. *Entomol. Exp. Appl.* 94(1): 93-102.
- 1217.De Barro, P.J., Liebrechts, W. and Carver, M. 1998. Distribution and identity of biotypes of *Bemisia tabaci* (Gennadius) (Hemiptera: Aleyrodidae) in member countries of the Secretariat of the Pacific Community. *Aust. J. Entomol.* 37(3): 214-218.
- 1218.De Bruijn, G.H. and Guthrie, E.J. 1982. Kenya . pp. 95-98. *In* Root Crops in Eastern Africa. Proceedings of a Workshop Held at Kigali, Rwanda, 23-27 November 1980. International Development Research Centre, Ottawa, Canada. [Cock (1986)]
- 1219.De Cock, A. 1993. Buprofezin: biologische activiteit en effect op de ultrastructuur van de cuticula van *Trialeurodes vaporariorum* (Westwood) en *Bemisia tabaci* (Gennadius). [Buprofezin: biological activity and effect on the ultrastructure of the cuticle of *Trialeurodes vaporariorum* (Westwood) and *Bemisia tabaci* (Gennadius)]. Ph.D. Dissertation, Faculteit van de Landbouwwetenschappen, Rijksuniversiteit, Gent, 139 pp. [Dutch]
- 1220.De Cock, A., Ishaaya, I., Degheele, D. and Veierov, D. 1990. Vapor toxicity and concentration-dependent persistence of buprofezin applied to cotton foliage for controlling the sweetpotato whitefly (Homoptera: Aleyrodidae). *J. Econ. Entomol.* 84: 1254-1260.
- 1221.De Courcy-Williams, M., Bedford, I.D., Kelly, A. and Markham, P.G. 1996. *Bemisia tabaci*: potential infestation and virus transmission within the ornamental plant industry. pp. 63-68. *In* Brighton Crop Protection Conference: Pests and Diseases. British Crop Protection Council, Farnham, UK.

1222. De Courcy-Williams, M. and Right, E. 1999. Parasitoid-induced mortality in the biological control of *Bemisia tabaci* on poinsettia. Bull. OILB/SROP (IOBC/WPRS) 22: 41-44.
1223. De La Torre, M. and Cardinas-Cota, H.M. 1996. Production of *Paecilomyces fumosoroseus* conidia in submerged culture. Entomophaga 41(2-4): 443-453.
1224. De Leon, E.D. 1976. Control químico de plagas del algodón en la región del Soconusco, Chiapas. Agric. Técnica Mexico 3: 447-451. [Cock (1986)]
1225. De León, F. and Sifuentes A.J.A. 1973. Control químico de la mosquita blanca en algodón en la región del Soconusco, Chis. [Chemical control of the whitefly on cotton in the region of Soconusco, Chis]. Agric. Técnica Mexico 3(7): 270-273. [Cock (1986)]
1226. De Nardo, E.A.B. and Costa, A.S. 1986. [Differentiation of isolates of the Brazilian complex of bean golden mosaic virus]. Fitopatol. Brasileira 11(3): 655-666. [Cock (1993), Portuguese, English summary]
1227. De Nardo, E.A.B., Costa, A.S. and Lourencao, A.L. 1997. *Melia azedarach* extract as an antifeedant to *Bemisia tabaci* (Homoptera: Aleyrodidae). Florida Entomol. 80(1): 92-94.
1228. De Oriani, M.A. and Lara, F.M. 2000. Antibiosis effects of wild bean lines containing arcelin on *Bemisia tabaci* (Genn.) biotype B (Homoptera: Aleyrodidae). Anais Soc. Entomol. Brasil 29(3): 573-582.
1229. De Oriani, M.A. and Lara, F.M. 2000. Oviposition preference of *Bemisia tabaci* (Genn.) biotype B (Homoptera: Aleyrodidae) for bean genotypes containing arcelin in the seeds. Anais Soc. Entomol. Brasil 29(3): 565-572.
1230. De Paula, S.V., Picanco, M., Vilela, E.F. and Fontes, P.C.R. 1998. Incidence of insect phyto-virus vectors on tomato (*Lycopersicon esculentum* Mill.) (Solanaceae) surrounded by crop strips. [Portuguese, English summary]. Rev. Brasil. Entomol. 41(2-4): 555-558.
1231. De Ponti, O. [Convenor]. 1984. Working group WPRS/EUCARPIA 'Breeding for Resistance to Insects and Mites', 3rd Meeting, Capbreton, France, 6-9 April 1983. Bull. OILB/SROP (IOBC/WPRS) 7(4): 1-82. [Cock (1986)]
1232. De Ponti, O.M.B., Romanow, L.R. and Berlinger, M.J. 1990. Whitefly-plant relationships: plant resistance. pp. 91-106. In D. Gerling (Ed.), Whiteflies: their Bionomics, Pest Status, and Management. Intercept, Andover, UK.
1233. De Quattro, J. 1994. Science update on whiteflies. Agric. Res. (USDA) 42(4): 23.
1234. De Quattro, J. 1995. Whitefly fungus on its way to growers. Agric. Res. (USDA) 43(5): 16-17.
1235. De Quattro, J., Senft, D. and Wood, M. 1997. The whitefly plan - 5-year update. Agric. Res. (USDA) 45(2): 4-12.
1236. De Santis, L. 1948. Estudio monográfico de los afelinidos de la República Argentina (Hymenoptera: Chalcidoidea). Rev. Museo Plata (Nueva Serie) 5: 149-159, 233-267. [Cock (1993)]
1237. De Santis, L. 1981. Sobre dos especies de *Encarsia* (Hymenoptera: Aphelinidae) del Brasil parasitoides de *Bemisia tabaci* (Homoptera: Aleyrodidae). Rev. Brasil. Entomol. 25: 37-39. [Cock (1993)]
1238. De Siqueira, K.M.M., De Farias, A.M.I. and Haji, F.N.P. 2001. Reconocimiento y oviposición del parasitoide *Encarsia lutea* en *Bemisia tabaci*. [Recognition and oviposition of the parasitoid *Encarsia lutea* on *Bemisia tabaci*]. Manejo Integrado de Plagas (Costa Rica) 61: 53-59. [Spanish, English summary]
1239. De Souza, A.P. and Vendramim, J.D. 2000. Efeito de extratos aquosos de meliáceas sobre *Bemisia tabaci* biotipo B em tomateiro. Bragantia 59(2): 173-179. [Portuguese, English summary]
1240. De Souza, A.P. and Vendramim, J.D. 2000. Ovicidal activity of aqueous extracts of Meliaceae on the silverleaf whitefly for tomato. Scientia Agric. 57(3): 403-406. [Portuguese, English summary]
1241. De Souza, A.P. and Vendramim, J.D. 2001. Atividade inseticida de extratos aquosos de Meliaceas sobre a mosca-branca *Bemisia tabaci* (Genn.) biotipo B (Homoptera: Aleyrodidae). [Insecticidal activity of aqueous extracts of Meliaceae plants on the silverleaf whitefly, *Bemisia tabaci* (Genn.) Biotipo B (Homoptera: Aleyrodidae)]. Neotrop. Entomol. 30(1): 133-137. [Portuguese, English summary]
1242. De Souza, V.B.V. and Kim, K.S. 1990. Cytopathology and transmission characteristics of a virus isolate from *Abutilon striatum*. Phytopathology 80(6): 548-552.
1243. Dean, D.E. 1994. Predaceous arthropods of the sweetpotato whitefly, *Bemisia tabaci* (Gennadius), on tomato in Florida. [University of Florida]. Dissertation Abst. Int. 56(11B): 5903.
1244. Dean, D.E. and Schuster, D.J. 1995. *Bemisia argentifolii* (Homoptera: Aleyrodidae) and *Macrosiphum euphorbiae* (Homoptera: Aphididae) as prey for two species of Chrysopidae. Environ. Entomol. 24(6): 1562-1568.
1245. Deang, R.T. 1969. An annotated list of insect pests of vegetables in the Philippines. Philippine Entomol. 1: 313-333.
1246. Debrot, E., Herold, F. and Dao, F. 1963. Nota preliminar sobre un "mosaico amarillento" del tomate en Venezuela. [with English summary]. Agron. Trop. 13: 33-41. [Cock (1986)]
1247. Debrot, E.A. and Centeno, F. 1985. Infección natural de la papa en Venezuela con el mosaico amarillo del tomate, un geminivirus transmitido por moscas blancas. Agron. Trop. 35: 125-138.
1248. Debrot, E.A. and Centeno, F. 1985. [Occurrence of *Euphorbia* mosaic virus infecting *Euphorbia heterophylla* L. in Venezuela]. Agron. Trop. 35(4-6): 5-12. [Cock (1993), Spanish, English summary]
1249. Debrot, E.A. and Ordosgoitti, F.A. 1975. Estudios sobre un mosaico amarillo de la soya en Venezuela. Agron. Trop. 25: 435-449. [Cock (1986)]
1250. Decanini, M.B., Smith, C.W. and Sparks, A.N., Jr. 1995. The use of no-choice fecundity study for the evaluation of cotton resistance to whitefly. pp. 846-847. In D.A. Richter and J. Armour (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
1251. Deguine, J.P. 1999. The strategic bases of cotton research at CIRAD. Agric. Devel. 22: 113-120. [French, English summary]
1252. Deguine, J.P. and Silvie, P. 1988. Un nouveau programme de protection insecticide en culture cotonnière au Tchad: augmentation des cadences de traitements et réduction des doses. [A new program for pest control in cotton fields in Tchad: increase of the treatment rates and reduction of the utilized amounts of pesticides]. pp. 771-787. In International Symposium Over Fytofarmacie En Fytiatrie, Belgium, Rijksuniversiteit Faculteit Landbouwwetenschappen, Gent, Volume 40. [French]
1253. Deguine, J.P., Vaissayre, M. and Hau, B. 1998. *Bemisia tabaci* on cotton in Senegal: Analysis of the situation and recommendations. Agric. Devel. 20: 19-23. [French, English summary]
1254. Dekker, E.L., Woolston, C.J., Xue, Y., Cox, B. and Mullineaux, P.M. 1991. Transcript mapping reveals different expression strategies for the bicistronic RNAs of the geminivirus wheat dwarf virus. Nucleic Acids Res. 19(15): 4075-4081.
1255. del Bene, G. and Landi, S. 1991. Biological pest control in glasshouse ornamental crops in Tuscany. Bull. OILB/SROP (IOBC/WPRS) 14: 13-21.
1256. Delattre, R. 1947. Insectes du cotonnier nouveaux ou peu connus en Cote d'Ivoire. Coton Fibres Trop. 2(1): 28-33. [Cock (1986)]
1257. Delattre, R. 1947. Insectes du cotonnier nouveaux ou peu connus en Cote d'Ivoire (II.). Coton Fibres Trop. 2(3): 97-100. [Cock (1986)]
1258. Delattre, R. 1961. Studies on the infestation and pesticide treatment of cotton in Africa. Outline of the work of the I.R.C.T. Phytiatric Phytopharmacie 10(1): 13-26. [Cock (1986), French]

1259. Delgado, A.M. 1996. El virus del rizado amarillo del tomate (TYLCV) en la region de murcia: aspectos generales para su control. pp. 67-70. *In* J.L. Cenis (Ed.), El Virus del Rizado Amarrillo (Hoja en Cuchara) del Tomate (TYLCV) y Su Vector *Bemisia tabaci*, Jornadas 8. Consejería de Medio Ambiente, Agricultura y Agua, Murcia, Spain.
1260. Demichelis, S., Bosco, D., Manino, A., Marian, D. and Caciagli, P. 2000. Distribution of *Bemisia tabaci* (Hemiptera: Aleyrodidae) biotypes in Italy. *Canadian Entomol.* 132(4): 519-527.
1261. Deng, D., McGrath, P.F., Robinson, D.J. and Harrison, B.D. 1994. Detection and differentiation of whitefly-transmitted geminiviruses in plants and vector insects by the polymerase chain reaction with degenerate primers. *Ann. Appl. Biol.* 125(2): 327-336.
1262. Dengel, H.J. 1981. Untersuchungen über das Auftreten der Imagines von *Bemisia tabaci* (Genn.) auf verschiedenen Manioksorten. *Zeitschrift für pflanzenkrankheiten und Pflanzenschutz* 88: 355-366. [Cock (1986)]
1263. Denholm, I. and Birnie, L.C. 1990. Prospects for managing resistance to insecticides in the whitefly. pp. 37-41. *In* Cotton Production Research from a Farming Systems Perspective, with Special Emphasis on Stickiness. 49th Plenary Meeting. International Cotton Advisory Committee
1264. Denholm, I., Cahill, M., Byrne, F.J. and Devonshire, A.L. 1996. Progress with documenting and combating insecticide resistance in *Bemisia*. pp. 577-603. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia*: 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
1265. Denholm, I., Cahill, M., Dennehy, T.J. and Horowitz, A.R. 1998. Challenges with managing insecticide resistance in agricultural pests, exemplified by the whitefly *Bemisia tabaci*. *Phil. Trans. R. Soc. (Lond. B)* 353(1376): 1757-1767.
1266. Denholm, I., Dennehy, T.J. and Horowitz, A.R. 1999. Dynamics of resistance to novel insecticides in whiteflies. pp. 44-47. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
1267. Denholm, I. and Jespersen, J.B. 1998. Insecticide resistance management in Europe: recent developments and prospects. *Pestic. Sci.* 52(2): 193-195.
1268. Denholm, I., Rollett, A.J., Cahill, M.R. and Ernst, G.H. 1995. Response of cotton aphids and whiteflies to diafenthiuron and pymetrozine in laboratory bioassays. pp. 991-994. *In* D.A. Richter and J. Armour (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
1269. Denholm, I., Rowland, M., Farnham, A.W. and Sawicki, R.M. 1990. Laboratory evaluation and empirical modeling of resistance-countering strategies. pp. 92-104. *In* M.B. Green, H.M. LeBaron and W.K. Moberg (Eds.), Symposium Series, American Chemical Society, No. 421.
1270. Dennehy, T.J. and Antilla, L. 1996. Whitefly resistance to insecticides in Arizona. pp. 144-145. *In* P. Dugger and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
1271. Dennehy, T.J. and Denholm, I. 1998. Goals, achievements and future challenges of the Arizona whitefly resistance management program. pp. 68-72. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
1272. Dennehy, T.J., Ellsworth, P.C. and Nichols, R.L. 1995. Whitefly management in Arizona cotton 1995. IPM Series No. 3, Univ. Arizona Coop. Ext., 4 pp.
1273. Dennehy, T.J., Ellsworth, P.C. and Nichols, R.L. 1996. Whitefly resistance management program for Arizona Cotton. IPM Series No. 8, Univ. Arizona, Coop. Ext. Publ. 196008, 16 pp.
1274. Dennehy, T.J., Ellsworth, P. C. and Watson, T.F. 1995. Whiteflies in Arizona: Pocket guide. Univ. Arizona Coop. Ext. Publ. 195009, 2 pp.
1275. Dennehy, T.J., Li, X. and Wigert, M. 1999. Successful management of whitefly resistance to pyrethroid insecticides in Arizona cotton: a four-year retrospective. pp. 392-401. *In* Cotton, A College of Agriculture Report, Series P-116. University of Arizona, Tucson.
1276. Dennehy, T.J., Wigert, M., Li, X. and Williams, L. III. 1999. Arizona whitefly susceptibility to insect growth regulators and chloronicotinyl insecticides: 1998 season summary. pp. 376-391. *In* Cotton, A College of Agriculture Report, Series P-116. University of Arizona, Tucson.
1277. Dennehy, T.J. and Williams, L. 1997. Management of resistance in *Bemisia* in Arizona cotton. *Pestic. Sci.* 51(3): 398-406.
1278. Dennehy, T.J., Williams, L. III, Li, X. and Wigert, M. 1998. 1997 season update on resistance of Arizona whiteflies to synergized pyrethroid and select non-pyrethroid insecticides. pp. 330-340. *In* Cotton, A College of Agriculture Report, Series P-112. University of Arizona, Tucson.
1279. Dennehy, T.J., Williams, L., III, Li, X., Wigert, M. and Birdwell, E. 1997. Status of whitefly resistance to insecticides in Arizona cotton. pp. 232-253. *In* Cotton, A College of Agriculture Report, Series P-108. University of Arizona, Tucson.
1280. Dennehy, T.J., Williams, L., III, Russell, J.S., Li, X. and Wigert, M. 1996. Monitoring and management of whitefly resistance to insecticides in Arizona. pp. 135-140. *In* P. Dugger and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
1281. Dennehy, T.J., Williams, L., III, Russell, J.S., Li, X. and Wigert, M. 1996. Monitoring and management of whitefly resistance to insecticides in Arizona. pp. 743-748. *In* P. Dugger and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
1282. Deotale, R.O., Nimbalkar, S.A. and Jumde, Y.S. 1992. Biology of white fly (*Bemisia tabaci* Genn.) on cotton. *J. Soils Crops* 2(2): 30-32.
1283. Department Oleagineux Annuels De l'Irho. 1982. Synthesis of studies presented on control of predators and groundnut diseases. *Oleagineux* 37(1): 25-28. [Cock (1986)]
1284. Desbiez, C., Lecoq, H., Aboulama, S. and Peterschmitt, M. 2000. First report of cucurbit yellow stunting disorder virus in Morocco. *Plant Dis.* 84(5): 596.
1285. Deshwal, H.L. and Sharma, J.K. 1996. Relative susceptibility of different varieties of mothbean against white fly, *Bemisia tabaci* Genn. *Ann. Biol.* 12(2): 252-254.
1286. Dessart, P. and Bournier, A. 1971. *Thrips tabaci* Lindman (Thysanoptera) hôte inattendu d' *Aphanogmus fumipennis* (Thompson) (Hym. Ceraphronidae). *Bull. Ann. Soc. Royale Entomol. Belgique* 107: 116-118. [Cock (1986)]
1287. Detzel, P. and Bathon, H. 1992. Nuetzlinge bei Poinsettien. Der Erfolg ist abhaengig vom Jungpflanzen-Lieferanten. [Useful insects in poinsettias. Success depends from the transplant supplier]. *Deutscher Gartenbau (Germany)* 46(4): 199-201. [German]
1288. Detzel, P. and Bathon, H. 1993. Poinsettien-Mutterpflanzen. Einsatz von *Encarsia formosa*. [Use of *Encarsia formosa* in poinsettia motherplants]. *Deutscher Gartenbau (Germany)* 47(11): 706-707. [German]
1289. Devine, G.J. and Denholm, I. 1998. An unconventional use of piperonyl butoxide for managing the cotton whitefly, *Bemisia tabaci* (Hemiptera: Aleyrodidae). *Bull. Entomol. Res.* 88(6): 601-610.
1290. Devine, G.J., Ishaaya, I., Horowitz, A.R. and Denholm, I. 1998. Effects of piperonyl butoxide on *Bemisia tabaci* Genn. (Homoptera: Aleyrodidae): mortality, development, parasitism and predation in Israeli cotton fields. *Crop Prot.* 17(9): 717-726.
1291. Devine, G.J., Ishaaya, I., Horowitz, A.R. and Denholm, I. 1999. The response of pyriproxyfen-resistant and susceptible *Bemisia tabaci* Genn (Homoptera: Aleyrodidae) to pyriproxyfen and fenoxycarb alone and in combination with piperonyl butoxide. *Pestic. Sci.* 55(4): 405-411.

- 1292.Devine, G.J., Wright, D.J. and Denholm, I. 2000. A parasitic wasp (*Eretmocerus mundus* Mercet) can exploit chemically induced delays in the development rates of its whitefly host (*Bemisia tabaci* Genn.). *Biol. Control* 19(1): 64-75.
- 1293.Devine, R.S. 1998. Alien invasion: America's battle with non-native animals and plants. National Geographic Society, Washington, D.C., 288 pp.
- 1294.Dhaliwal, J.S. 1998. Record of some new pests of forage crops. *J. Insect Sci.* 11(2): 180.
- 1295.Dhamdhare, S.V., Bahadur, J. and Misra, U.S. 1984. Studies on occurrence and succession of pests of okra at Gwalior. *Indian J. Plant Prot.* 12(1): 9-12. [ publ. 1985; Cock (1993)]
- 1296.Dhandapani, N., Kalyanasundaram, M., Swamiappan, M., Sundara-Babu, P.C. and Jayaraj, S. 1992. Experiments on management of major pests of cotton with biocontrol agents in India. *J. Appl. Entomol.* 114(1): 52-56.
- 1297.Dhanju, K.C., Cheema, S.S., Kapur, S.P., Arora, S.K. and Ghai, T.R. 1995. Occurrence of enation leaf curl disease of okra in Punjab. *Plant Dis. Res.* 10(2): 157-159.
- 1298.Dhanju, K.S. and Varma, J.P. 1986. Natural occurrence of mixed infection of tomato leaf curl virus and tobacco mosaic virus in tomato. *Indian J. Virol.* 2(1): 108-110. [ Cock (1993)]
- 1299.Dharmasena, C.M.D. 1998. Present status of managing chilli leaf curl complex in the North Central Province of Sri Lanka. *Trop. Agric. Res. Ext.* 1(2): 154-158.
- 1300.Dharmatti, P.R., Madalageri, B.B., Mannikeri, I.M. and Patil, R.V. 1999. Combining ability for tomato leaf curl virus resistance in summer tomatoes (*Lycopersicon esculentum*). *Adv. Agric. Res. (India)* 11: 67-72.
- 1301.Dhawan, A.K., Sidhu, A.S. and Simwat, G.S. 1989. Management of bollworm through chlorpyrifos in cotton system. *J. Res. (Punjab Agric. Univ.)* 26(4): 599-603. [ Cock (1993)]
- 1302.Dhawan, A.K. and Simwat, G.S. 1998. Population dynamics of whitefly, *Bemisia tabaci* on cotton: an ecobehavioural approach. pp. 435-448. *In* G.S. Dhaliwal, R. Arora, N.S. Randhawa and A.K. Dhawan (Eds.), *Ecological agriculture and sustainable development: Volume 1. Proceedings of an International Conference on Ecological Agriculture: Towards Sustainable Development, Chandigarh, India, 15-17 November, 1997. Centre for Research in Rural and Industrial Development, Chandigarh, India.*
- 1303.Dhawan, A.K., Simwat, G.S. and Sidhu, A.S. 1987. Effect of sowing dates on the incidence of sucking pests and bollworms in *arboreum* cotton. *J. Res. (Punjab Agric. Univ.)* 24(1): 75-85. [ Cock (1993)]
- 1304.Dhawan, A.K., Simwat, G.S. and Sidhu, A.S. 1988. Field evaluation of monocrotophos for the control of sucking pests on cotton. *Pesticides* 22(6): 25-28. [ Cock (1993)]
- 1305.Dhawan, A.K., Simwat, G.S. and Sidhu, A.S. 1990. Field reaction of different varieties of upland cotton to insect pests in Punjab. *J. Res. (Punjab Agric. Univ.)* 27(2): 263-266. [ Cock (1993)]
- 1306.Dhawan, A.K., Simwat, G.S. and Sidhu, A.S. 1990. Incidence of different insect pests on LH 900 upland cotton sown on different dates. *J. Res. (Punjab Agric. Univ.)* 27(2): 244-252. [ Cock (1993)]
- 1307.Dhawan, A.K., Simwat, G.S. and Sidhu, A.S. 1991. Field evaluation of Deltaphos for control of sucking pests and bollworms during reproductive phase of cotton crop. *Indian J. Plant. Prot.* 19(2): 172-176. [ Cock (1993)]
- 1308.Dhingra, K.L. and Chenulu, V.V. 1982. Control of yellow mosaic of soybean by systemic insecticides, 1976-78. *Fungicide Nematicide Tests, Am. Phytopathol. Soc.* 37: 103.
- 1309.Dhingra, K.L. and Ghosh, D.K. 1993. Efficiency of whitefly vector (*Bemisia tabaci* Gen.) in transmission of mungbean yellow mosaic virus in different source-test plant combinations. *Int. J. Trop. Agric.* 11(2): 149-152.
- 1310.Dhuri, A.V. and Singh, K.M. 1983. Pest complex and succession of insect pests in black gram *Vigna mungo* (L.) Hepper. *Indian J. Entomol.* 45: 396-401. [ Cock (1986)]
- 1311.Dhuri, A.V., Singh, K.M. and Singh, R.N. 1984. Incidence of insect pests in black gram *Vigna mungo* (L.) Hepper. *Indian J. Entomol.* 46(3): 270-276. [ Cock (1993)]
- 1312.Di Feo, L., Nome, S.F., Biderbost, E., Fuentes, S. and Salazar, L.F. 2000. Etiology of sweet potato chlorotic dwarf disease in Argentina. *Plant Dis.* 84(1): 35-39.
- 1313.Diana, G. and Sannino, L. 1995. Avversita della coltura del tabacco: insetti. [Adversities of tobacco culture: insects]. *Informatore Fitopatol.* 45(7-8): 24-31. [ Italian, English summary]
- 1314.Diaz-Franco, A. 1995. Produccion de cultivares de calabacita asociada con hoja plateada. [Production of squash cultivars associated with silverleaf]. *Rev. Mexicana Fitopatologia* 13(1): 26-28. [ Spanish, English summary]
- 1315.Diaz-Franco, A. and Ortegon, A.S. 1997. Influencia de la fecha de siembra y la poda sobre la produccion de cultivares de oca. [Influence of planting dates and pruning on the production of okra (*Abelmoschus esculentus*) cultivars]. *Agron. Mesoamericana* 8(1): 93-98. [ Spanish, English summary]
- 1316.Diaz, O.F. 1994. Evaluation of transgenic bean plants for their reaction to BGMV transmitted by *Bemisia tabaci*. pp. 181-182. *In* F.J. Morales (Ed.), *Bean Golden Mosaic: Research Advances. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia.* [English and Spanish]
- 1317.Diaz-Plaza, R. and Ramirez-Choza, J.L. 1993. Bioecologia y control integrado de la mosquita blanca *Bemisia tabaci* Genn (Homoptera: Aleyrodidae). [Bioecology and integrated control of *Bemisia tabaci* Genn (Homoptera: Aleyrodidae)]. *Campo Experimental Zona Henequenera, Cir Sureste, Mococho, Yucatan, Mexico*, 23 pp. [ Spanish]
- 1318.Díaz, R.O., Bellotti, A.C. and van Schoonhoven, A. 1978. Insectos y acaros que atacan al cultivo de la yuca en Colombia. *Turrialba* 28(1): 43-49. [ Cock (1986)]
- 1319.Dickson, R.C., Johnson, M.M. and Laird, E.F., Jr. 1954. Leaf crumple, a virus disease of cotton. *Phytopathology* 44: 479-480. [ Cock (1986)]
- 1320.Diehl, J.W. and Ellsworth, P.C. 1995. Community-wide implementation of sampling and action thresholds for whiteflies in cotton. pp. 255-258. *In* Cotton, A College of Agriculture Report, Series P-99. University of Arizona, Tucson.
- 1321.Diehl, J.W., Ellsworth, P.C. and Husman, S.H. 1994. A community-wide approach to whitefly management. pp. 299-303. *In* Cotton, A College of Agriculture Report, Series P-96. University of Arizona, Tucson.
- 1322.Diehl, J.W., Ellsworth, P.C. and Husman, S.H. 1994. Response to sweetpotato whitefly: organization of community IPM. pp. 1203-1204. *In* D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
- 1323.Diehl, J.W., Ellsworth, P.C. and Naranjo, S.E. 1997. *Bemisia* growth regulators: a field sampling-protocol for nymphs. pp. 929-931. *In* P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
- 1324.Diehl, J.W., Naranjo, S.E. and Ellsworth, P.C. 1997. Whitefly growth regulators: a field sampling protocol for nymphs. pp. 265-271. *In* Cotton, A College of Agriculture Report, Series P-108. University of Arizona, Tucson.
- 1325.Diethelm, V. 1990. Whiteflies on gerbera controlled with parasitic wasps. Research results in Switzerland. *Gärtnerbörse und Gartenwelt (Germany)* 90(11): 546-553.
- 1326.Díez, M.J., Picó, B. and Nuez, F. 1996. Mejora genética para la resistencia al TYLCV. pp. 91-98. *In* J.L. Cenis (Ed.), *El Virus del Rizado Amirillo (Hoja en Cuchara) del Tomate (TYLCV) y Su Vector Bemisia tabaci, Jornadas 8. Consejería de Medio Ambiente, Agricultura y Agua, Murcia, Spain.*
- 1327.Dimetry, N.Z., Gomaa, A.A., Salem, A.A. and Abd-El-Moniem, A.S.H. 1996. Bioactivity of some formulations of neem seed extracts against the whitefly *Bemisia tabaci* (Genn.). *Anz. Schadlingskunde, Pflanzenschutz, Umweltschutz* 69(6): 140-141.



- 1328.Dinçer, J. 1979. Progress made in integrated control of cotton pests with special emphasis on its practicability and economic interest. pp. 211-213. *In* K. Russ and H. Berger (Eds.), Proceedings International Symposium of IOBC/WPRS on Integrated Control in Agriculture and Forestry. Vienna Oct. 8-12, 1979. International Organization for Biological Control, Vienna, Austria. [Cock (1986)]
- 1329.Dinçer, J. 1984. Ege bölgesinde pamuk zararlılarına karşı entegre mücadele imkanlarının araştırılması. *Bitki Koruma Bulteni* 24(1): 15-32. [Cock (1986)]
- 1330.Diraviam, J. and Uthamasamy, S. 1992. Monitoring whitefly, *Bemisia tabaci* (Genn.) on sunflower with yellow sticky traps. *J. Entomol. Res. (New Delhi)* 16(2): 163-165.
- 1331.Direccao Geral de Protecçao das Culturas. 1998. Programa de erradicaçao dos virus TSWV e TYLCV na cultura do tomateiro: resultados da monitorizaçao dos vectores (*Frankliniella occidentalis*, *Thrips tabaci*, *Bemisia tabaci*). [Programme for the eradication of the TSWV and TYLCV viruses in tomato crops: results of monitoring of the different vectors (*Frankliniella occidentalis*, *Thrips tabaci*, *Bemisia tabaci*)]. Direccao Geral de Protecçao das Culturas, Oeiras, Portugal, 39 pp. [Portuguese]
- 1332.Dittrich V. 1987. Resistance and hormoligosis as driving forces behind pest outbreaks. pp. 169-181. *In* Rational Pesticide Use. Proceedings of the Ninth Long Ashton Symposium. Cambridge University Press, Cambridge, UK. [Cock (1993)]
- 1333.Dittrich, V. and Ernst, G.H. 1983. The resistance pattern in whiteflies of Sudanese cotton. *Mitteilungen der Deutschen Gesellschaft für Allgemeine und Angewandte Entomol.* 4(1-3): 96-97. [Cock (1986), German, English summary]
- 1334.Dittrich, V., Ernst, G.H., Ruesch, O. and Uk, S. 1990. Resistance mechanisms in sweetpotato whitefly (Homoptera: Aleyrodidae) populations from Sudan, Turkey, Guatemala, and Nicaragua. *J. Econ. Entomol.* 83(5): 1665-1670. [Cock (1993)]
- 1335.Dittrich, V., Hassan, S.O. and Ernst, G.H. 1985. Sudanese cotton and the whitefly: a case study of the emergence of a new primary pest. *Crop Prot.* 4: 161-176. [Cock (1986)]
- 1336.Dittrich, V., Hassan, S.O. and Ernst, G.H. 1986. Development of a new primary pest of cotton in the Sudan: *Bemisia tabaci*, the whitefly. *Agric. Ecosystems Environ.* 17(1-2): 137-142. [Cock (1993)]
- 1337.Dittrich, V., Uk, S. and Ernst, G.H. 1990. Chemical control and insecticide resistance of whiteflies. pp. 263-285. *In* D. Gerling (Ed.), Whiteflies: their Bionomics, Pest Status and Management. Intercept, Andover, UK. [Cock (1993)]
- 1338.Divakar, B.J., Sarma, P.V., Ragunathan, V., David, B.V., Reddy, G.R.S. and Swamy, S.V. 1994. Integrated pest management in cotton. *Indian J. Plant Prot.* 22(1): 98-104.
- 1339.Diwakar, M.P., Rajput, J.C. and D'Souza, T.F. 1986. Diseases of okra in Konkan region of Maharashtra. *Pesticides* 20(3): 34. [Cock (1993)]
- 1340.Djuwarso, T. and Suryawan, F. 1989. Major insect pests of legumes. pp. 356-367. *In* S. Hardjosumadi, M. Machmud, U.G. Kartasmita and A. Kurnia (Eds.), Proceedings of the Seminar of Food Crops Research, Balitran Bogor, Indonesia, 13-14 February 1989. [Indonesian, English summary]
- 1341.Do Queiros, M.R.P., Figueiredo, E. and Mexia, A. 2000. Monitoring for the whitefly *Bemisia tabaci* Genn. on Ribatejo and Oeste region of Portugal. *Bull. OILB/SROP (IOBC/WPRS)* 23(1): 97-99.
- 1342.Dodds, J.A., Lee, J.G., Nameth, S.T. and Laemmlen, F.F. 1984. Aphid- and whitefly-transmitted cucurbit viruses in Imperial County, California. *Phytopathology* 74: 221-225. [Cock (1986)]
- 1343.Dollet, M., Dubern, J., Fauquet, C., Thouvenel, J.C. and Bocklé-Morvan, A. 1987. [Groundnut viral diseases in West Africa]. *Oleagineux* 42(7): 291-297. [Cock (1993), French, English summary]
- 1344.Dolores, L.M., Valdez, R.B. and Roxas, V.P. 1988. Viral diseases of squash (*Cucurbita* spp.) in the Philippines and sources of resistance. *Philippine J. Crop Sci.* 13: 30. [CATIE (1992)]
- 1345.Domenichini, P. and Roffeni-Tirafferri, S. 1998. Risultati sperimentali di prove di campo condotte nel triennio 1995-97 con l'impiego di acetamiprid - nuovo insetticida sistemico. [Results of the field trials carried out with acetamiprid - a new systemic insecticide - during 1995-1997]. pp. 167-172. *In* Italian Phytopathological Society. Biennial meeting, Ragusa, Italy, 3-7 May 1998. [Italian, English summary]
- 1346.Donnely, J. 1966. Insect pests on kenaf. *Nigerian Entomol. Mag.* 1(1): 8-9. [Cock (1986)]
- 1347.Doreste S, E., Arias, C. and Bellotti, A. 1978. Field evaluations of cassava cultivars for resistance to tetranychid mites. pp. 161-164. *In* T. Brekelbaum, A. Bellotti and J.C. Lozano (Eds.), Proc. Cassava Protection Workshop CIAT, Cali, Columbia, November 7-12, 1977. Centro Internacional de Agricultura Tropical, Cali, Colombia. [Cock (1986)]
- 1348.Doss, S.A., Adam, K.M., Kerakly, F.A. and El-Hamaky, M.A. 1992. Population densities of the broad bean leafminer, *Liriomyza trifolii*, Berguss, and cotton whitefly, *Bemisia tabaci*, Genn, on protected cultivations. *Minia J. Agric. Res. Devel. (Egypt)* 14(3): 787-797.
- 1349.Doss, S.A., Ali, S.M. and Nour-El-Din, M.M. 1997. Relative susceptibility of some strawberry cultivars to *Tetranychus arabis* Atiah and whitefly, *Bemisia tabaci* (Gennadius) infestation. *Fayoum J. Agric. Res. Devel. (Egypt)* 11(2): 119-125.
- 1350.Doss, S.A. and Atallah, E.A. 1985. Effect of plant density on tomato infestation by different insect pests. *Agric. Res. Rev.* 63(1): 69-75. [CATIE (1992)]
- 1351.Dowell, R.V. 1990. Integrating biological control of whiteflies into crop management systems. pp. 315-335. *In* D. Gerling (Ed.), Whiteflies: their Bionomics, Pest Status and Management. Intercept, Andover, UK.
- 1352.Dozier, H.L. 1937. Descriptions of miscellaneous chalcidoid parasites from Puerto Rico (Hymenoptera). *J. Agric. Univ. Puerto Rico.* 21(2): 121-135. [Cock (1986)]
- 1353.Drees, B.M. 1988. A new pest for Texas: the sweetpotato whitefly. *Texas Nurseryman* 19(5): 8-12.
- 1354.Drees, B.M. 1989. Sweetpotato whitefly management on Texas ornamentals. *Ent Notes* 30(6): 1-5.
- 1355.Drees, B.M., Pianta, W. and Daniel, J. 1990. Sweetpotato whitefly treatment evaluations. *Texas Nurseryman* 21(9): 22-24.
- 1356.Dreyer, M. 1991. Neem: a promising natural insecticide for small scale vegetable producers in the Dominican Republic. pp. 491-500. *In* Caribbean Meetings on Biological Control. Guadeloupe (French Antilles), November 5-7, 1990. Institut National De La Recherche Agronomique, Paris.
- 1357.Dreyer, M. and Hellpap, C. 1991. Neem - a promising natural insecticide for small scale vegetable production in tropical and subtropical countries. *Zeitschrift für pflanzenkrankheiten und Pflanzenschutz* 89(4): 428-437. [German, English summary]
- 1358.Drost, Y.C. 1998. Searching behaviour of parasitoids of whiteflies. *Novenyvedelem (Hungary)* 34: 13, 79-80.
- 1359.Drost, Y.C., Elmula, A.F., Posthuma-Doodeman, C.J.A.M. and van Lenteren, J.C. 1996. Development of criteria for evaluation of natural enemies in biological control: bionomics of different parasitoids of *Bemisia argentifolii*. *Bull. OILB/SROP (IOBC/WPRS)* 19(1): 31-34.
- 1360.Drost, Y.C., Elmula, A.F., Posthuma-Doodeman, C.J.A.M. and van Lenteren, J.C. 1996. Development of selection criteria for natural enemies in biological control: parasitoids of *Bemisia argentifolii*. *Proc. Exp. Appl. Netherlands Entomol. Soc.* 7(1996): 165-170.
- 1361.Drost, Y.C., Qiu, Y.T., Posthuma-Doodeman, C.J.A.M. and van Lenteren, J.C. 1999. Life-history and oviposition behaviour of *Amitus bennetti*, a parasitoid of *Bemisia argentifolii*. *Entomol. Exp. Appl.* 90(2): 183-189.
- 1362.Drost, Y.C., Qiu, Y.T., Posthuma-Doodeman, C.J.A.M. and van Lenteren, J.C. 2000. Comparison of searching strategies of five parasitoid species of *Bemisia argentifolii* Bellows and Perring (Hom., Aleyrodidae). *J. Appl. Entomol.* 124(2): 105-112.

1363. Drost, Y.C., van Lenteren, J.C. and van Roermund, H.J.W. 1998. Life-history parameters of different biotypes of *Bemisia tabaci* (Hemiptera, Aleyrodidae) in relation to temperature and host plant: a selective review. *Bull. Entomol. Res.* 88(3): 219-229.
1364. Dry, I.B., Rigden, J.E., Krake, L.R., Mullineaux, P.M. and Rezaian, M.A. 1993. Nucleotide sequence and genome organization of tomato leaf curl geminivirus. *J. Gen. Virol.* 74: 147-151.
1365. Duan, Y.P., Powell, C.A., Webb, S.E., Purcifull, D.E. and Hiebert, E. 1997. Geminivirus resistance in transgenic tobacco expressing mutated BC1 protein. *Mol. Plant-Microbe Interactions* 10(5): 617-623.
1366. Dubern, J. 1994. Transmission of African cassava mosaic geminivirus by the whitefly (*Bemisia tabaci*). *Trop. Sci.* 34(1): 82-91.
1367. Dubern, J. and Dollet, M. 1981. Groundnut crinkle virus, a new member of the carlavirus group. *Phytopathol. Z.* 101: 337-347. [Cock (1986)]
1368. Dubitzki, E., Rosenberg, U. and Yogev, E. 1991. Inhibition of virus transmission and control of whitefly (*Bemisia tabaci*) by the detergent ZOHAR LQ-215' in melon. *Hassadeh* 72(2): 196-198.
1369. Duffus, J.E. 1965. Beet pseudo-yellow virus, transmitted by the greenhouse whitefly (*Trialeurodes vaporariorum*). *Phytopathology* 55: 450-453. [Cock (1986)]
1370. Duffus, J.E. 1987. Whitefly transmission of plant viruses. pp. 73-91. In K.F. Harris (Ed.), *Current Topics in Vector Research*, Vol. 4. Springer-Verlag, New York, USA.
1371. Duffus, J.E. 1994. Diseases vectored by whiteflies: etiology, ecology, geographical distribution and possible control measures. *Arab J. Plant Prot.* 12(2): 148-143 Arab journal reverses order.
1372. Duffus, J.E. 1996. Whitefly -borne viruses. pp. 255-263. In D. Gerling and R.T. Mayer (Eds.), *Bemisia : 1995 Taxonomy, Biology, Damage, Control and Management*. Intercept Ltd., Andover, Hants, UK.
1373. Duffus, J.E., Cohen, S. and Liu, H.Y. 1992. The sweetpotato whitefly in Western USA biotypes, plant interactions and virus epidemiology. pp. 76-77. In I.C. Rumbos, P. Kyriakopoulou and F. Bem (Eds.), *Recent Advances in Vegetable Virus Research*. 7th Conference ISHS Vegetable Virus Working Group, Athens, Greece, July 12-16, 1992. Ores Publishing, Volos, Greece.
1374. Duffus, J.E. and Flock, R.A. 1982. Whitefly-transmitted disease complex of the desert southwest. *California Agric.* 36(11-12): 4-6. [Cock (1986)]
1375. Duffus, J.E. and Johnstone, G.R. 1981. Beet pseudo-yellow virus in Tasmania. The first report of a whitefly transmitted virus in Australia. *Aust. Plant Pathol.* 10: 68-69. [Cock (1986)]
1376. Duffus, J.E., Larsen, R.C. and Liu, H.Y. 1986. Lettuce infectious yellows virus - a new type of whitefly transmitted virus. *Phytopathology* 76(1): 97-100. [Cock (1986, 1993)]
1377. Duffus, J.E., Liu, H.Y., Wisler, G.C. and Li, R.H. 1996. Lettuce chlorosis virus - a new whitefly-transmitted closterovirus. *Eur. J. Plant Pathol.* 102(6): 591-596.
1378. Duncombe, W.C. 1973. The acaricide spray rotation for cotton. *Rhodesia Agric. J.* 70: 115-118. [Cock (1986)]
1379. Dupuy, J.W. and Sencion, L.M. 1990. Whitefly (*Bemisia tabaci*) in tomato and proposed control in the Dominican Republic. *Proc. Interamerican Soc. Trop. Hortic.* 34: 97-100.
1380. Dussourd, D.E. 1995. Entrapment of aphids and whiteflies in lettuce latex. *Ann. Entomol. Soc. Am.* 88(2): 163-172.
1381. Duverger, C. 1986. [*Nephaspis maesi*, a new species of Scymnini from Nicaragua (Coleoptera, Coccinellidae)]. *Rev. Française Entomol.* 8(4): 167-169. [Cock (1993), French, English summary]
1382. Dysart, R.J. 1966. Natural enemies of the banded-wing whitefly, *Trialeurodes abutilonea* (Hemiptera: Aleyrodidae). *Ann. Entomol. Soc. Am.* 59: 28-33. [Cock (1986, 1993)]
1383. Echemendia, A.L., Ramos, P.L., Peral, R., Fuentes, A., Gonzalez, G., Sanpedro, J. and Morales, F. 2001. Cuban isolate of Bean golden yellow mosaic virus is a member of the mesoamerican BGYMV group. *Plant Dis.* 85(9): 1030.
1384. Edrisha, M.E. and Badr, T.S. 1994. Effect of some foliar fertilizers, pesticides and their mixtures on white fly *Bemisia tabaci* (Genn.) and their side effects on tomato fruits. *Moshtohor Ann. Agric. Sci. (Egypt)* 32(3): 1697-1706.
1385. Eichelkraut, K. and Cardona, C. 1989. [Biology, mass rearing and ecological aspects of the whitefly *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae), as a pest of beans]. *Turrialba* 1: 55-62. [Cock (1993), Spanish, English summary]
1386. Ekobom, B.S. and Rumei, X. 1990. Sampling and spatial patterns of whiteflies. pp. 107-121. In D. Gerling (Ed.), *Whiteflies: their Bionomics, Pest Status and Management*. Intercept, Andover, UK. [Cock (1993)]
1387. Ekukole, G. 1993. A check-list of cotton entomofauna in North Cameroon. I. Phytophagous species. *Coton Fibres Trop.* 48(3): 213-219.
1388. El-Amin, E.M., Hussein, I. and Talab, A.A. 1979. Large scale ultra low volume trial at Khashm El Girba scheme, season 1972-73. pp. 160-164. In *Annu. Rep. Gezira Res. Stn. Substns. 1972/1973*. Agric. Res. Corp., Sudan. [Cock (1993)]
1389. El-Amin, T.M. and El-Tayeb, Y.M. 1976. ULV [ultralow volume] application of insecticides gives Sudan better control [of cotton pests, *Empoasca lybica*, *Bemisia tabaci*, *Heliothis armigera*]. *Cotton Int.* Ed. 43: 206,208,210.
1390. El Badry, E.A. 1967. Three new species of phytoseiid mites preying on the cotton whitefly *Bemisia tabaci* in the Sudan (Acarina: Phytoseiidae). *Entomologist* 100: 106-111. [Cock (1986)]
1391. El Badry, E.A. 1968. Biological studies on *Amblyseius aleyrodii* a predator of the cotton whitefly (Acarina, pytoseiidae). *Entomophaga* 13: 323-329. [Cock (1986)]
1392. El-Bakheit, I.B. 1986. Entomology Section, 1981-1982. pp. 71-85. In Kenana, Sudan, Annual Report, Kenana Research Station. [Cock (1993)]
1393. El-Bakheit, I.B. 1991. Insect pests of vegetables. pp. 162-172. In *Annu. Rep. Gezira Res. Stn. Substns. 1986/1987*. Agric. Res. Corp., Sudan.
1394. El-Banhawy, E.M., Amer, S.A.A. and Saber, S.A. 2000. Development and reproduction of the predacious mite, *Amblyseius cydnodactylon* on different prey species; effect of plant leaf texture on the behaviour and reproduction of the predator. *Zeitschrift für pflanzenkrankheiten und Pflanzenschutz* 107(2): 218-224.
1395. El-Banhawy, E.M., Hafez, S.M. and Saber, S.A. 1999. Effect of the nymph prey density of the two spotted spider mite *Tetranychus urticae* Koch on the consumption and reproduction rates of the predacious mite *Cydnoseius negevi* (Swirski & Amital) in absence and presence of nymphs of the whitefly *Bemisia tabaci* (Genn.) (Acari: Phytoseiidae). *J. Pest Sci.* 72(2): 55-56.
1396. El-Banhawy, E.M., Hafez, S.M. and Saber, S.A. 2001. Response of *Amblyseius cydnodactylon* (Phytoseiidae) to increasing prey density of *Tetranychus urticae* (Tetranychidae) in absence or presence of nymphs of *Bemisia tabaci* (Homoptera) in Egypt. *Int. J. Acarol.* 27(3): 241-244.
1397. El-Bashir, S. 1974. Effect of some insecticides on immature stages of the cotton whitefly. *Cotton Growing Rev.* 51: 62-69. [Cock (1986)]
1398. El-Bessomy, M.A.E. 1998. Effect of the natural oil, Jojoba oil on reduction of different stages of whitefly, *Bemisia tabaci* (Genn.) and virus symptoms on tomato plants. *Mansoura Univ. J. Agric. Sci. (Egypt)* 23(7): 3375-3380.
1399. El-Bessomy, M.A.E., El-Khawalka, H.I.H.O. and El-Maghraby, H.M. 1997. Effect of the fungal insecticide (Biofly) compared with chemical insecticides in controlling different stages of whitefly *Bemisia tabaci* (Genn.) and its related virus. *Egyptian J. Agric. Res.* 75(4): 915-921.

- 1400.El-Borllosy, F.M. and Hemeida, E.A. 1986. Ecological studies on some insects attacking potato plant and tubers. I.- Effect of some potato varieties and planting date on the population density of some insects. *Zagazig J. Agric. Res. (Egypt)* 13(1): 475-489.
- 1401.El-Dahan, A.A., Hindy, M.A. and Moawad, G.M. 1997. Performance of different ground spraying equipment for controlling whitefly, *Bemisia tabaci* (Genn.) in cotton fields of Egypt. *Egyptian J. Agric. Res.* 75(2): 393-399.
- 1402.El-Gendi, S.S., Adam, K.M. and Bachatly, M. A. 1997. Effect of the planting date of tomato on the population density of *Bemisia tabaci* (Genn.) and *Heliothis armigera* (HB), viral infection and yield. *Arab J. Agric. Sci.* 5(1): 135-144.
- 1403.El-Ghany, A., El-Sayed, M., Afifi, F.M.L. and Haydar, M.F. 1990. Intereffects of temperature and type of food on adult longevity of *Eretmocerus mundus* Mercet, a primary parasitoid of *Bemisia tabaci* (Genn.) [Arabic summary]. *Bull. Faculty Agric. (Univ. Cairo)* 41(3 (Suppl. 1)): 913-922. [Cock (1993)]
- 1404.El-Ghany, A., El-Sayed, M. and El-Ghar, G.E.S.A. 1992. The influence of normal and low-rate application of insecticides on populations of the cotton whitefly and melon aphid and associated parasites and predators on cucumber. *Anz. Schadlingskunde, Pflanzenschutz, Umweltschutz* 65: 54-57.
- 1405.El-Ghar, G.E.S.A., El-Sayed, A.E.M., El-Shiekh, A.E. and Radwan, H.S.A. 1994. Field tests with insecticides and insect growth regulators to control insect pests of cowpea and its effects on certain beneficial insects. *Arch. Phytopathol. Plant Prot.* 28(6): 531-543.
- 1406.El-Hamaky, M.A. and Watson, W.M. 1993. Bioresidual activity of certain insecticides against adult and immature stage of whitefly *Bemisia tabaci* (Genn.) infesting cotton. *Mansoura Univ. J. Agric. Sci. (Egypt)* 18(7): 2150-2154.
- 1407.El-Hawary, I.S. 1988. Susceptibility of ten eggplant genotypes to the cotton and tomato whitefly, *Bemisia tabaci* (Genn.) in relation to yield in Kafr El Sheikh. *J. Agric. Res. (Egypt)* 14(3): 1288-1295. [CATIE (1992)]
- 1408.El-Hawary, I.S., Shenishen, Z., Tadros, M.S. and Ibrahim, M.M. 1995. Effect of the foliar fertilizers, and the plant growth regulator on *Aphis gossypii* Glover., *Bemisia tabaci* Genn. and consequences on *Pectinophora gossypiella* infestation in cotton fields. *Menofiya J. Agric. Res. (Egypt)* 20(4): 1595-1603.
- 1409.El-Helaly, M.S., El-Gayar, F.H. and El-Shazli, A.Y. 1975. Studies on the nutrition of the whitefly, *Bemisia tabaci* Genn. (Hom.: Aleyrodidae). I. A suitable device for artificial feeding. *Z. Angew. Entomol.* 78: 392-396. [Cock (1986)]
- 1410.El-Helaly, M.S., El-Gayar, F.H. and El-Shazli, A.Y. 1976. Development of a standard bioassay technique for the adult of *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae). *J. Agric. Sci.* 87: 543-548. [Cock (1986)]
- 1411.El-Helaly, M.S., El-Gayar, F.H. and El-Shazli, A.Y. 1977. Studies on the nutrition of the whitefly *Bemisia tabaci* Gennadius II. Soluble and non-soluble amino acids in adults. *Indian J. Entomol.* 38(1976): 263-265. [Cock (1986)]
- 1412.El-Helaly, M.S., El-Shazli, A.Y. and El-Gayar, F.H. 1971. Biological studies on *Bemisia tabaci* Genn. (Homopt.: Aleyrodidae) in Egypt. *Z. Angew. Entomol.* 69: 48-55.
- 1413.El-Helaly, M.S., El-Shazli, A.Y. and El-Gayar, F.H. 1971. Morphological studies on immature stages of *Bemisia tabaci* Gennadius (Homoptera: Aleyrodidae). *Z. Angew. Entomol.* 68: 403-408. [Cock (1986)]
- 1414.El-Helaly, M.S., Ibrahim, E.G. and Rawash, I.A. 1977. Photoperiodism of the whitefly *Bemisia tabaci* Gennadius (Aleyrodidae: Homoptera). *Z. Angew. Entomol.* 83: 393-397. [Cock (1986)]
- 1415.El-Helaly, M.S., Ibrahim, E.G. and Rawash, I.A. 1977. Sterilization of the whitefly *Bemisia tabaci* Genn. (Homop.: Aleyrodidae) by ultra-violet radiation. *Z. Angew. Entomol.* 83: 135-140. [Cock (1986)]
- 1416.El-Helaly, M.S., Rawash, I.A. and Ibrahim, E.G. 1981. Phototaxis of the adult whitefly, *Bemisia tabaci* Gennadius to the visible light. II. Effects of both light intensity and sex of the whitefly adults on the insect's response to different wavelengths of light spectrum. *Acta Phytopathologica Academiae Scientiarum Hungaricae* 16(3-4): 389-398. [Cock (1986)]
- 1417.El-Helaly, M.S., Rawash, I.A. and Ibrahim, E.G. 1981. Phototaxis of the adult whitefly, *Bemisia tabaci* Gennadius to visible light. I. Effect of the exposure period on the insect's response to different wavelengths of the visible light-spectrum using a devised simple technique. *Acta Phytopathologica Academiae Scientiarum Hungaricae* 16: 181-188. [Cock (1986)]
- 1418.El-Jadd, L. and Guirrou, Z. 1990. The cotton whitefly *Bemisia tabaci* Genn (Homoptera: Aleyrodidae) host plants and population dynamics at Tadla (Morocco). *Al Awamia* 71: 37-50. [French]
- 1419.El-Kabbany, S.M. 1994. Comparative studies on the efficacy of some insecticides on cotton whitefly, *Bemisia tabaci* (Genn.) and their side effects on non-target microorganisms. *Mansoura Univ. J. Agric. Sci. (Egypt)* 19(4): 1569-1577.
- 1420.El-Khawalka, M.H.M., El-Bessomy, M.A., El-Maghraby, H.M. and Omar, H.I.H. 1997. Effect of the insect growth regulator, Admiral on the different stages of the whitefly infesting tomato plants. *Egyptian J. Agric. Res.* 75(2): 377-382.
- 1421.El-Khayat, E.F., El-Sayed, A.M., Shalaby, F.F. and Hady, S.A. 1994. Infestation rates with *Bemisia tabaci* (Genn.) to different summer and winter vegetable crop plants. *Moshtohor Ann. Agric. Sci. (Egypt)* 32(1): 577-594.
- 1422.El-Khidir, E. 1965. Bionomics of cotton whitefly, (*Bemisia tabaci* Genn.) in the Sudan and the effects of irrigation on population density of whiteflies. *Sudan Agric. J.* 1(2): 8-22. [Cock (1986)]
- 1423.El-Khidir, E. and Khalifa, A. 1962. A new aleyrodid from the Sudan. *Proc. Royal Entomol. Soc. London (B)* (B) 31: 47-51. [Cock (1986)]
- 1424.El-Lissy, O., Antilla, L. and Butler, G.D., Jr. 1993. Sweetpotato whitefly control on cotton by treating only the field edges. pp. 248-252. *In Cotton, A College of Agriculture Report, Series P-94.* University of Arizona, Tucson.
- 1425.El-Lissy, O., Antilla, L. and Butler, G.D., Jr. 1994. Sweetpotato whitefly control of cotton by treating only the field edges. pp. 146-147. *In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences.* National Cotton Council, Memphis, TN.
- 1426.El-Lissy, O., Antilla, L., Staten, R.T., Leggett, J.E. and Walters, M. 1994. Control of sweetpotato (silverleaf) whitefly, *Bemisia tabaci*, on cotton in Paloma, Arizona. pp. 289-298. *In Cotton, A College of Agriculture Report, Series P-96.* University of Arizona, Tucson.
- 1427.El-Lissy, O., Antilla, L., Staten, R.T., Walters, M. and Leggett, J.E. 1994. Control of sweetpotato (silverleaf) whitefly, *Bemisia tabaci*, on cotton in Paloma, Arizona. pp. 148-151. *In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences.* National Cotton Council, Memphis, TN.
- 1428.El-Maghraby, M.M., El-Zoheri, M.M. and Hassanein, S.S. 1994. Relationship between insect predators and pests associated with different varieties of squash and cucumber cultivated in the newly reclaimed sandy areas of El-Khattara district, Egypt. *Zagazig J. Agric. Res. (Egypt)* 21(3b): 969-975.
- 1429.El-Maghraby, M.M.A., Shahein, A. and El-Deeb, M.A. 1990. Relationship between certain pests and their natural enemies on tomato and pepper plants in plastic tunnels in the newly reclaimed sandy areas of El-Kasasien district, Egypt. *Zagazig J. Agric. Res. (Egypt)* 17(5b): 1709-1713.
- 1430.El-Malky, K.G., Gergis, M.F. and Abdel-Alim, A.A. 2001. Population growth parameters of whitefly on *Bemisia tabaci* (Genn.). pp. 869-870. *In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences.* National Cotton Council, Memphis, TN.
- 1431.El-Meniawi, F.A. and Hashem, M. 1997. Insecticidal activity of detergents against the adult and immature stages of the cotton whitefly *Bemisia tabaci* Genn. on tomato. *Alexandria J. Agric. Res. (Egypt)* 42(3): 75-84.

- 1432.El-Mezayyen, G.A. and Abou-Attia, F.A. 1996. Population fluctuations of certain cotton sucking insect pests and associated predators as influenced by some weather factors at Kafer-El-Sheikh. *J. Agric. Res. (Egypt)* 22(4): 518-531.
- 1433.El-Nahal, A.K.M., Ammar, E.D., Dawood, M.Z. and Assem, M.A. 1980. Population studies on *Bemisia tabaci*, on some varieties of cucumber, snake cucumber and squash, at Damietta and Beni-Suef regions. pp. 291-303. *In Proceedings 1st Conference of Plant Protection Research Institute. Plant Protection Research Institute, Cairo.*
- 1434.El-Nawawy, A.S., Abd-El-Rahman, I., Ashry, M.A., Hosny, A. and Belal, A. 1983. Effect of mixtures of a foliar fertilizer and each of several insecticides on sucking pests and their predators in cotton fields. *Med. Fac. Landbouww. Univ. Gent* 48(1): 117-127. [Cock (1986)]
- 1435.El-Rafie, K.K. 1999. Effect of different rates of (N, P, K) fertilizers on *Bemisia tabaci* (Genn.) infestation on tomato and its effect on the yield. *Egyptian J. Agric. Res.* 77(3): 1067-1073.
- 1436.El-Rafie, K.K., Dawood, M.Z. and Hydar, M.F. 1999. The relation between daily temperature, and the population of *Bemisia tabaci* (Genn.) on tomato plantation. *Egyptian J. Agric. Res.* 77(4): 1501-1507.
- 1437.El-Said, A.M., Hegazi, E.M., Abo-El-Ghar, M.R. and Schalk, J.M. 1980. Ecological studies on the white fly, *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae) associated with the wild plants *Conyza dioscorides* in Egypt. *Natl. Agric. Library, Foreign Pub.*, 12 pp.
- 1438.El-Sayed, A.M. 1986. Further Studies on the white fly, *Bemisia tabaci* (Genn.) and its natural enemies in Egypt. Ph.D. Dissertation, Zagazig University, Moshtohor, Egypt, 246 pp.
- 1439.El-Sayed, A.M., Adl, F.E., Sami, M.A., Ali, S.M. and Hindy, M.A. 1997. Performance of different sprayers in relation to insecticidal activity against whitefly infesting eggplants in Egypt. *Moshtohor Ann. Agric. Sci. (Egypt)* 35(3): 1727-1740.
- 1440.El-Sayed, A.M. and El-Ghar, G.E.S.A. 1993. Effect of selected insecticides on population, adult longevity and reproduction of the whitefly, *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae). *Bull. Entomol. Soc. Egypt Econ. Ser.* 20: 161-171.
- 1441.El-Sayed, A.M., Moawad, G.M. and Samy, S.M. 1998. Natural enemies of *Bemisia tabaci* (Gennadius) on some vegetable crops in newly reclaimed lands in Egypt. *Zagazig J. Agric. Res. (Egypt)* 25(2): 337-350.
- 1442.El-Sayed, A.M., Shalaby, F.F., El-Khayat, E.F. and Hady, S.A. 1994. Relationship between certain weather factors and *Bemisia tabaci* populations on different host plants. *Moshtohor Ann. Agric. Sci. (Egypt)* 32(1): 617-631.
- 1443.El-Serwi, S., El-Haidari, H. and Saad, A. 1984. Population density of the whitefly *Bemisia tabaci* (Gennadius), (Homoptera: Aleyrodidae) on fall cucumber in Iraq. *J. Agric. Water Resources Res.* 3(2): 78-87 (Arabic), 135 (English). [Cock (1986)]
- 1444.El-Serwi, S.A., Ali, A.A. and Razoki, I.A. 1987. Effect of intercropping plantation of some host plants with tomato on population density of tobacco whitefly, *Bemisia tabaci* (Genn.), and the incidence of tomato yellow leaf curl virus (TYLCV) in plastic houses. *J. Agric. Water Resources Res. (Iraq)* 6(2): 81-97. [Arabic, English summary]
- 1445.El-Serwi, S.A., Ali, A.A. and Rozoki, I.A. 1987. [Effect of intercropping of some host plants with tomato on population density of tobacco whitefly, *Bemisia tabaci* (Genn.) and the incidence of tomato yellow leaf curl virus (TYLCV) in plastic houses]. *J. Agric. Water Resources Res.* 6: 81-79. [Cock (1993), Arabic, English summary]
- 1446.El-Serwi, S.A. and El-Haidari, H.S. 1983. Resistance of different varieties of tomato to *Bemisia tabaci* (Gennadius) in Iraq. *J. Agric. Water Resources Res. (Iraq)* 2(2): 81-84.
- 1447.El-Tayeb, Y.M. 1978. Spiny bollworm research [spiny bollworm; American bollworm; jassid; whitefly]. pp. 292-317. *In Annu. Rep. Gezira Res. Stn. Substns. 1970/1971. Agric. Res. Corp., Sudan.*
- 1448.El-Tayeb, Y.M. 1980. Large scale insecticide screening experiment. pp. 137-147. *In Annu. Rep. Gezira Res. Stn. Substns. 1973/1974. Agric. Res. Corp., Sudan [Cock (1993)]*
- 1449.El-Tayeb, Y.M. 1980. Small scale evaluation of insecticides for control of whitefly, american bollworm and spiny bollworm. pp. 130-137. *In Annu. Rep. Gezira Res. Stn. Substns. 1973/1974. Agric. Res. Corp., Sudan. [Cock (1993)]*
- 1450.El-Tayeb, Y.M. 1981. Entomology. pp. 91-108. *In Annu. Rep. Gezira Res. Stn. Substns. 1974/1975. Agric. Res. Corp., Sudan.*
- 1451.El-Tayeb, Y.M. and El-Amin, E.M. 1979. Large scale control of cotton pests in Gezira. pp. 152-159. *In Annu. Rep. Gezira Res. Stn. Substns. 1972/1973. Agric. Res. Corp., Sudan [Cock (1993)]*
- 1452.El-Tayeb, Y.M. and Mansi, M.G. 1980. Evaluation of the effect of some ultra low volume and conventional insecticide formulations on cotton pests. pp. 148-154. *In Annu. Rep. Gezira Res. Stn. Substns. 1973/1974. Agric. Res. Corp., Sudan [Cock (1993)]*
- 1453.El-Tigani, M.A. and Ahmed, M.A. 1991. Strategies for integrated cotton pest control in the Sudan. 1 - Cultural and legislative measures. *Insect Sci. Appl.* 12(5/6): 547-552.
- 1454.Elbert, A., Cahill, M., Nauen, R. and Steffens, R. 1997. From monitoring to implementation: a stepwise approach to resistance management with imidacloprid. *Resist. Pest Manage.* 9(1): 14-16.
- 1455.Elbert, A., Erdelen, C., Kuehnhold, J., Nauen, R. and Schmit, H.W. 2000. Thiacloprid, a novel neonicotinoid insecticides for foliar application. pp. 21-26. *In Brighton Crop Protection Conference: Pests and Diseases. British Crop Protection Council, Farnham, UK.*
- 1456.Elbert, A. and Nauen, R. 1996. Bioassays for imidacloprid for resistance monitoring against the whitefly *Bemisia tabaci*. pp. 731-738. *In Brighton Crop Protection Conference: Pests and Diseases. British Crop Protection Council, Farnham, UK.*
- 1457.Elbert, A. and Nauen, R. 2000. Resistance of *Bemisia tabaci* (Homoptera : Aleyrodidae) to insecticides in southern Spain with special reference to neonicotinoids. *Pest Manag. Sci.* 56(1): 60-64.
- 1458.Elbert, A., Nauen, R., Cahill, M., Devonshire, A.L., Scarr, A.W., Sone, S. and Steffens, R. 1996. Resistance management with chloronicotinyl insecticides using imidacloprid as an example. *Pflanzenschutz-Nachrichten Bayer (English Ed.)* 49(1): 5-54. [English, summaries in German, French, Spanish, Russian]
- 1459.Eldin, N.E.S. 1985. Investigation into the chemical control of whitefly on cotton. pp. 106-111. *In Annu. Rep. Gezira Res. Stn. Substns. 1977/1978. Agric. Res. Corp., Sudan [Cock (1993)]*
- 1460.Elewa, M.A., Saad, A.S.A. and Aly, N.M. 1979. Susceptibility of different gland and glandless cotton varieties to infestation with some cotton pests in relation to their chemical control. *Med. Fac. Landbouww. Univ. Gent* 44: 235-241. [Cock (1986)]
- 1461.Ellis, D., McAvoy, R., Abu-Ayyash, L., Flanagan, M. and Ciomperlik, M. 2001. Evaluation of *Serangium parcesetosum* (Coleoptera: Coccinellidae) for biological control of silverleaf whitefly, *Bemisia argentifolii* (Homoptera: Aleyrodidae), on poinsettia. *Florida Entomol.* 84(2): 215-221.
- 1462.Ellsworth, P., Diehl, J., Dennehy, T. and Naranjo, S. 1994. Sampling sweetpotato whiteflies in cotton. *IPM Series No. 2, Univ. Arizona Coop. Ext. Publ.* 194023, 2 pp.
- 1463.Ellsworth, P., Moore, L., Watson, T.F. and Dennehy, T. 1994. Insect pest management for cotton. *Univ. Arizona Coop. Ext. Publ.* #194022, Tucson, Arizona, 27 pp.
- 1464.Ellsworth, P.C. 1995. Whiteflies in Arizona: sampling, action thresholds & management. pp. 101-102. *In D.A. Richter and J. Armour (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
- 1465.Ellsworth, P.C. 1998. Whitefly management in Arizona: looking at the whole system. pp. 65-72. *In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*

1466. Ellsworth, P.C. 1999. Whitefly management in Arizona cotton - status and needs. pp. 41-43. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
1467. Ellsworth, P.C., Akey, D.H., Williams, L., Dennehy, T.J., Kirk, I.W., Carlton, J.B., Henneberry, T.J., Coppedge, J.R. and Diehl, J.W. 1996. Understanding whitefly control: threshold, insecticide rotation, & ground and air comparisons. pp. 133-135. *In* P. Dugger and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
1468. Ellsworth, P.C., Chernicky, J.P., Byrne, D.N., Gibson, R. and Meade, D. 1992. A native weed as a trap crop for whiteflies in cotton. pp. 911-913. *In* D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
1469. Ellsworth, P.C., Dennehy, T.J. and Nichols, R.L. 1996. Whitefly management in Arizona cotton 1996. IPM Series No. 3, Univ. Arizona Coop. Ext. Publ. 196004, 2 pp.
1470. Ellsworth, P.C., Diehl, J. and Naranjo, S.E. 1998. Impact of natural enemies and insecticides on whiteflies in cotton: a partial life table analysis. pp. 1087-1089. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
1471. Ellsworth, P.C., Diehl, J.W. and Husman, W.H. 1996. Establishment of integrated pest management infrastructure: a community-based action program for *Bemisia* management. pp. 681-693. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia* : 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
1472. Ellsworth, P.C., Diehl, J.W., Kirk, I.W. and Henneberry, T.J. 1997. *Bemisia* growth regulators: large-scale evaluation. pp. 922-929. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
1473. Ellsworth, P.C., Diehl, J.W., Kirk, I.W. and Henneberry, T.J. 1997. Whitefly growth regulators: large-scale evaluation. pp. 279-293. *In* Cotton, A College of Agriculture Report, Series P-108. University of Arizona, Tucson.
1474. Ellsworth, P.C., Diehl, J.W. and Meade, D.L. 1993. Cotton insect management in Arizona as impacted by whiteflies. pp. 966-968. *In* D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
1475. Ellsworth, P.C., Diehl, J.W. and Naranjo, S.E. 1995. Validation and implementation of a sampling model for whiteflies. pp. 978-979. *In* D.A. Richter and J. Armour (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
1476. Ellsworth, P.C., Diehl, J.W. and Naranjo, S.E. 1996. Sampling sweetpotato whitefly nymphs in cotton. IPM Series No. 6, Univ. Arizona, Coop. Ext. Publ. 196006, 2 pp.
1477. Ellsworth, P.C., Diehl, J.W., Silvertooth, J.C., Brown, P.W., Watson, T.F., Hood, L.R., Husman, S.H., Thacker, G.W., Clark, L.J., Cluff, R.E., Doerge, T.A., Howell, D.R., McCloskey, W.B., Stedman, S.W., Tronstad, R.E. and Wade, J.C. 1993. Sweetpotato whitefly in Arizona. pp. 283-293. *In* Cotton, A College of Agriculture Report, Series P-94. University of Arizona, Tucson.
1478. Ellsworth, P.C. and Jones, J.S. 2001. Cotton IPM in Arizona: a decade of research, implementation & education. pp. 1088-1096. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
1479. Ellsworth, P.C. and Martinez-Carrillo, J.L. 2001. IPM for *Bemisia tabaci*: a case study from North America. *In* S. E. Naranjo and P. C. Ellsworth (Eds.), Special Issue: Challenges and Opportunities for Pest Management of *Bemisia tabaci* in the New Century. Crop Prot. 20(9): 853-869.
1480. Ellsworth, P.C. and Meade, D.L. 1993. Performance of select ed insecticides against the sweetpotato whitefly and cotton aphid. pp. 280-282. *In* Cotton, A College of Agriculture Report, Series P-94. University of Arizona, Tucson.
1481. Ellsworth, P.C. and Meade, D.L. 1994. Action thresholds for whiteflies in Arizona. pp. 313-325. *In* Cotton, A College of Agriculture Report, Series P-96. University of Arizona, Tucson.
1482. Ellsworth, P.C. and Meade, D.L. 1994. Action thresholds for whiteflies in Arizona. pp. 878-881. *In* D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
1483. Ellsworth, P.C. and Meade, D.L. 1994. Novel pyrethroid combinations for control of sweetpotato whitefly and their impact on *Lygus*. pp. 346-351. *In* Cotton, A College of Agriculture Report, Series P-96. University of Arizona, Tucson.
1484. Ellsworth, P.C. and Meade, D.L. 1994. Validity of the pinhead square treatment program for pink bollworm suppression and impact of several insecticides on arthropod fauna in cotton. pp. 267-277. *In* Cotton, A College of Agriculture Report, Series P-96. University of Arizona, Tucson.
1485. Ellsworth, P.C., Meade, D.L., Byrne, D.N., Draeger, E.A. and Chernicky, J.P. 1993. Progress on use of trap crops for whitefly suppression. p. 274. *In* Cotton, A College of Agriculture Report, Series P-94. University of Arizona, Tucson.
1486. Ellsworth, P.C., Meade, D.L., Husman, S.H., Ramsey, C.S., Silvertooth, J.C. and Malcuit, J.E. 1993. Sweetpotato whitefly preference and performance on medium maturity cotton varieties in Arizona. pp. 275-279. *In* Cotton, A College of Agriculture Report, Series P-94. University of Arizona, Tucson.
1487. Ellsworth, P.C., Meade, D.L. and Odom, P. 1994. Preliminary field evaluation of an insect growth regulator, buprofezin, for control of the sweetpotato whitefly, *Bemisia tabaci*. pp. 363-367. *In* Cotton, A College of Agriculture Report, Series P-96. University of Arizona, Tucson.
1488. Ellsworth, P.C. and Naranjo, S.E. 1999. Whitefly management with insect growth regulators and the influence of *Lygus* controls. pp. 339-354. *In* Cotton, A College of Agriculture Report, Series P-116. University of Arizona, Tucson.
1489. Ellsworth, P.C., Naranjo, S.E., Castle, S.J., Hagler, J. and Henneberry, T.J. 1998. Whitefly management in Arizona: looking at whole systems. pp. 311-318. *In* Cotton, A College of Agriculture Report, Series P-112. University of Arizona, Tucson.
1490. Ellsworth, P.C., Sieglaff, D.H., Yasui, M. and Lublinkhof, J. 1999. Monitoring *Bemisia* susceptibility to Applaud (buprofezin) during the 1998 cotton season. pp. 361-375. *In* Cotton, A College of Agriculture Report, Series P-116. University of Arizona, Tucson.
1491. Ellsworth, P.C., Tronstad, R., Leser, J., Goodell, P.B., Godfrey, L.D., Henneberry, T.J., Hendrix, D., Brushwood, D., Naranjo, S.E., Castle, S. and Nichols, R.L. 1999. Sticky cotton sources and solutions. IPM Series No. 13, Univ. Arizona, Coop. Ext. Publ. AZ1156, 4 pp.
1492. Ellsworth, P.C. and Watson, T.F. 1996. Whiteflies in Arizona (No. 7): Pocket Guide '96. Univ. Arizona Coop. Ext. Publ. 196005, 2 pp.
1493. Elmer, J.S., Brand, L., Sunter, G., Gardiner, W., Bisaro, D.M. and Rogers, S.G. 1988. Genetic analysis of tomato golden mosaic virus. II. The conserved ALI ORF product is essential for replication. Nucleic Acids Res. 16(14): 7043-7061.
1494. Elsey, K.D. and Farnham, M.W. 1994. Response of *Brassica oleracea* L. to *Bemisia tabaci* (Gennadius). Hortscience 29(7): 814-817.
1495. Emam, A.K. 1996. Host plants suitability and control of the cotton whitefly *Bemisia tabaci*, Genn Homoptera: Aleyrodidae. Al-Azhar J. Agric. Res. (Egypt) 17: 45-58.
1496. Emam, A.K. 1999. The effect of squash as a plant trap and yellow sticky traps on the population density of the whiteflies, *Bemisia tabaci* in tomato fields. Ain-Shams Univ. Ann. Agric. Sci. (Egypt) 44(1): 395-402.
1497. Engle, C.E., Cohick, A.D. and Kroll, T.K. 1993. Combinations of Baythroid and Monitor for the control of whitefly in cotton. pp. 964-965. *In* D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.

- 1498.Engle, C.E., Scoggan, A.C. and Mullins, J.W. 1994. Imidacloprid for silverleaf whitefly control: a three year summary. pp. 1211-1213. *In* D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 1499.Enkegaard, A. 1989. The tobacco whitefly, *Bemisia tabaci*, and the possibilities for biological control by the parasitoid *Encarsia formosa*. pp. 83-89. *In* 6th Danish Plant Protection Conference: Pests and Diseases. Statens Planteavltsforsoeg, Lyngby, Denmark. [Danish, English summary]
- 1500.Enkegaard, A. 1990. Age-specific fecundity and adult longevity of the cotton whitefly, *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae) on poinsettia (*Euphorbia pulcherrima*) at different temperatures. Bull. OILB/SROP (IOBC/WPRS) 13(5): 55-60.
- 1501.Enkegaard, A. 1990. *Bemisia tabaci*: pests in greenhouses now and in future. Statens Planteavltsforsoeg, Lyngby, Denmark, 6 pp. [Danish]
- 1502.Enkegaard, A. 1990. Cotton whitefly - A pest in greenhouses - now and in the future. Gron Viden, Havebrug 56: 6.
- 1503.Enkegaard, A. 1993. The bionomics of the cotton whitefly, *Bemisia tabaci*, and its parasitoid, *Encarsia formosa*, on poinsettia. Bull. OILB/SROP (IOBC/WPRS) 16(8): 66-72.
- 1504.Enkegaard, A. 1993. *Encarsia formosa* parasitizing the poinsettia-strain of the cotton whitefly, *Bemisia tabaci*, on poinsettia - bionomics in relation to temperature. Entomol. Exp. Appl. 69(3): 251-261.
- 1505.Enkegaard, A. 1993. The poinsettia strain of the cotton whitefly, *Bemisia tabaci* (Homoptera: Aleyrodidae), biological and demographic parameters on poinsettia (*Euphorbia pulcherrima*) in relation to temperature. Bull. Entomol. Res. 83(4): 535-546.
- 1506.Enkegaard, A. 1994. Temperature dependent functional response of *Encarsia formosa* parasitizing the poinsettia-strain of the cotton whitefly, *Bemisia tabaci*, on poinsettia. Entomol. Exp. Appl. 73(1): 19-29.
- 1507.Ernst, G.H. 1994. Whiteflies (*Bemisia tabaci*) in cotton - possible origin of the problem and today's chemical control opportunities. Pestic. Sci. 42(2): 139-141.
- 1508.Esaki, T. 1940. A preliminary report on the entomological survey of the Micronesian Islands under the Japanese Mandate, with special reference to the insects of economic importance. Proc. 6th Pacific Sci. Congress 1939 4: 407-415. [Cock (1986)]
- 1509.Esau, K. 1977. Virus-like particles in the nuclei of phloem cells in spinach leaves infected with curly top virus. J. Ultrastruct. Res. 61: 78-88.
- 1510.Escoto, N.D. 2000. Historia y situacion de la produccion de frijol en los paises latinoamericanos afectados por geminivirus transmitidos por mosca blanca: America Central: Honduras. pp. 33-36. *In* Bean Golden Mosaic and Other Diseases of Common Bean Caused by Whitefly-Transmitted Geminiviruses in Latin America. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. [Spanish]
- 1511.Eshmatov, O.T. and Kadyrov, A.K. 1991. Whiteflies and their control. Zashchita Rastenii (Moskva) 2: 23-24. [Russian]
- 1512.Espanol, J.A. and Corredor, D. 1989. A method for interpreting yellow traps used in the evaluation of whitefly on commercial tomato crop. Rev. Colombiana Entomol. 15(2): 36-42. [Spanish, English summary]
- 1513.Etessami, P., Callis, R., Ellwood, S. and Stanley, J. 1988. Delimitation of essential genes of cassava latent virus DNA 2. Nucleic Acids Res. 16(11): 4811-4829.
- 1514.Etessami, P., Saunders, K., Watts, J. and Stanley, J. 1991. Mutational analysis of complementary-sense genes of African cassava mosaic virus DNA A. J. Gen. Virol. 72: 1005-1012.
- 1515.Etessami, P., Watts, J. and Stanley, J. 1989. Size reversion of African cassava mosaic virus coat protein gene deletion mutants during infection of *Nicotiana benthamiana*. J. Gen. Virol. 70: 277-289.
- 1516.Ethridge, M.D. and Hequet, E.F. 1999. Prospects for rapid measurement of stickiness in cotton. pp. 56-60. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 1517.Etienne, J., Quiot, J.B. and Russell, L.M. 1991. Les Aleyrodidae de Guadeloupe, cas de *Bemisia tabaci*. [Aleyrodidae from Guadeloupe: the case of *Bemisia tabaci*]. pp. 85-91. *In* Caribbean Meetings on biological control, Guadeloupe, French Antilles, November 5-7, 1990. Institut National De La Recherche Agronomique, Pairs. [French, English summary]
- 1518.Evans, D.D. 1965. Jassid populations on three hairy varieties of Sakel cotton. Empire Cotton Growing Rev. 42: 211-217. [Cock (1986)]
- 1519.Evans, G.A. 1993. Systematic studies of New World *Encarsia* species and a survey of the parasitoids of *Bemisia tabaci* in Florida, the Caribbean and Latin America. [University of Florida]. Dissertation Abst. Int. 55(09B): 3698.
- 1520.Evans, G.A. 1997. A new *Encarsia* (Hymenoptera: Aphelinidae) species reared from the *Bemisia tabaci* complex (Homoptera: Aleyrodidae). Florida Entomol. 80(1): 24-27.
- 1521.Evans, G.A. and Polaszek, A. 1997. Additions to the *Encarsia* parasitoids (Hymenoptera: Aphelinidae) of the *Bemisia tabaci*-complex (Hymenoptera: Aleyrodidae). Bull. Entomol. Res. 87(6): 563-571.
- 1522.Evans, G.A. and Polaszek, A. 1998. The *Encarsia cubensis* species-group (Hymenoptera: Aphelinidae). Proc. Entomol. Soc. Washington 100(2): 222-233.
- 1523.Eveleens, K.G. 1983. Cotton-insect control in the Sudan Gezira: analysis of a crisis. Crop Prot. 2: 273-287. [Cock (1986)]
- 1524.Everitt, J.H., Escobar, D.E., Summy, K.R., Alaniz, M.A. and Davis, M.R. 1996. Using spatial information technologies for detecting and mapping whitefly and harvester ant infestations in South Texas. Southwest. Entomol. 21(4): 421-432.
- 1525.Fabres, G., Boher, B., Bonato, O., Calatayud, P.A., Fargette, D., Le Gall, P., Le Ru, B., Savary, S. and Verdier, V. 1994. [Toward integrated management of the parasite biocenoses of cassava in Africa]. Comptes Rendus de L'Academie D'Agriculture de France 80(8): 37-54. [French]
- 1526.Fadl, G.M. and Burgstaller, H. 1986. Reduction of tomato leaf curl virus in Sudan through variety selection and insecticide application. Acta Hort. 190: 159-164. [Cock (1993)]
- 1527.Fadlalla, A.S. 1997. Improvement of the marketability of cotton produced in zones affected by stickiness. pp. 49-52. *In* International Cotton Advisory Committee, 56th Plenary Meeting, Paraguay.
- 1528.Falcon, L.A. 1971. Progreso del control integrado en el algodón de Nicaragua. Rev. Peruana Entomol. 14: 376-378. [Cock (1986)]
- 1529.Falk, B.W. and Klaassen, V. 1996. Lettuce infectious yellows virus: a bipartite closterovirus transmitted by *Bemisia*, and representative of a new genus of plant viruses. pp. 265-275. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia*: 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
- 1530.Fan, Y.Q. and Pettit, F.L. 1998. Dispersal of the broad mite, *Polyphagotarsonemus latus* (Acari: Tarsonemidae) on *Bemisia argentifolii* (Homoptera: Aleyrodidae). Exp. Appl. Acarol. 22(7): 411-415.
- 1531.Fancelli, M. 2001. Resistencia de genotipos de tomateiro a mosca-branca *Bemisia tabaci* (Gennadius, 1889) biotipo B. [Resistance of tomato genotypes to *Bemisia tabaci* (Gennadius, 1889) B-biotype]. Ph.D. Dissertation, Escola Superior de Agricultura Luiz de Queiroz, Piracicaba, SP, Brazil, 144 pp. [Portuguese]
- 1532.FAO. 1966. Plant Pest and Disease Situation Near East Regional. Report 20: 7. [Cock (1986)]
- 1533.FAO. 1995. Report. Workshop on Management of the Whitefly-Virus Complex in Vegetables and Cotton production in the Near East. Workshop on Management of the Whitefly Virus Complex in Vegetables and Cotton Production in the Near East, Lanarca, Cyprus, 2-6 Oct 1995. 5 pp.

1534. Farah, S.M. and Rahman, A.A.A. 1988. Effects of water stoppage on yield and quality of cotton variety Barac (67) B. pp. 38-41. *In* Annu. Rep. Gezira Res. Stn. Substns. 1980/1981. Agric. Res. Corp., Sudan [Cock (1993)]
1535. Fargette, D., Colon, L.T., Bouveau, R. and Fauquet, C. 1996. Components of resistance of cassava to African cassava mosaic virus. *Eur. J. Plant Pathol.* 102(7): 645-654.
1536. Fargette, D. and Fauquet, C. 1988. A preliminary study on the influence of intercropping maize and cassava on the spread of African cassava mosaic virus by whiteflies. *Aspects Appl. Biol.* 17: 195-202.
1537. Fargette, D., Fauquet, C., Grenier, E. and Thresh, J.M. 1990. The spread of African cassava mosaic virus into and within cassava fields. *J. Phytopathol.* 130(4): 289-302. [Cock (1993)]
1538. Fargette, D., Fauquet, C. and Thouvenel, J.C. 1985. Field studies on the spread of African cassava mosaic. *Ann. Appl. Biol.* 106(2): 285-294. [Cock (1986)]
1539. Fargette, D., Fauquet, C. and Thouvenel, J.C. 1988. Field losses induced by African cassava mosaic virus in relation to the mode and date of infection. *Trop. Pest Manage.* 34(1): 89-91. [Cock (1993)]
1540. Fargette, D., Jeger, M., Fauquet, C. and Fishpool, L.D.C. 1994. Analysis of temporal disease progress of African cassava mosaic virus. *Phytopathology* 84(1): 91-98.
1541. Fargette, D., Muniyappa, V., Fauquet, C., N'Guessan, P. and Thouvenel, J.C. 1993. Comparative epidemiology of three tropical whitefly-transmitted geminiviruses. *Biochimie (Paris)* 75(7): 547-554.
1542. Fargette, D., Thouvenel, J.C. and Fauquet, C. 1987. Virus content of leaves of cassava infected by African cassava mosaic virus. *Ann. Appl. Biol.* 110(1): 65-73. [Cock (1993)]
1543. Fargette, D., Thresh, J.M. and Otim-nape, G.W. 1994. The epidemiology of African cassava mosaic geminivirus: reversion and the concept of equilibrium. *Trop. Sci.* 34(1): 123-133.
1544. Fargette, D. and Vie, K. 1994. Modeling the temporal primary spread of African cassava mosaic virus into plantings. *Phytopathology* 84(4): 378-382.
1545. Faria, J.C. 2000. Historia y situacion de la produccion de frijol en los paises latinoamericanos afectados por geminivirus transmitidos por mosca blanca: America del Sur: Brasil. pp. 79-86. *In* Bean Golden Mosaic and Other Diseases of Common Bean Caused by Whitefly - Transmitted Geminiviruses in Latin America. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. [Spanish]
1546. Faria, J.C., Gilbertson, R.L., Hanson, S.F., Morales, F.J., Ahlquist, P., Loniello, A.O. and Maxwell, D.P. 1994. Bean golden mosaic geminivirus type ii isolates from the Dominican Republic and Guatemala: nucleotide sequences, infectious pseudorecombinants, and phylogenetic relationships. *Mol. Plant Pathol.* 84: 321-329.
1547. Faria, J.C., Oliveira, M.N. and Yokoyama, M. 1994. Resposta comparativa de genotipos de feijoeiro (*Phaseolus vulgaris*) a inoculacao com o virus do mosaico dourado no estagio de plantulas. [Comparative response of bean (*Phaseolus vulgaris*) genotypes to the inoculation with bean golden mosaic virus at the seedling stage]. *Fitopatol. Brasileira* 19(4): 566-572. [Portuguese, English summary]
1548. Faria, M. and Wraight, S.P. 2001. Biological control of *Bemisia tabaci* with fungi. *In* S. E. Naranjo and P. C. Ellsworth (Eds.), Special Issue: Challenges and Opportunities for Pest Management of *Bemisia tabaci* in the New Century. *Crop Prot.* 20(9): 767-778.
1549. Farias, A.R.N. 1990. Especies de "mosca branca": Situaçao atual e perspectivas de controle. pp. 1-9. *In* 7th Curso Intesivo Nacional de Mandioca, 12-24 Agosto, 1990, Cruz das Almas, Brasil. Empresa Brasileira de Pesquisa Agropecuaria - EMBRAPA, Centro Nacional de Pesquisa de Mandioca e Fruticultura. [Portuguese]
1550. Farnham, M.W. and Elsey, K.D. 1995. Recognition of *Brassica oleracea* L. resistance against the silverleaf whitefly. *Hortscience* 30(2): 343-347.
1551. Farrag, R.M., Kotob, F.K. and Nousier, N.I. 1994. Factors affecting the chemical control of the whitefly *Bemisia tabaci* on cabbage plants. *Alexandria J. Agric. Res. (Egypt)* 39(3): 307-316.
1552. Farrag, R.M., Nasr, F.N. and Noussier, N.I. 1996. Influence of diversity in plant species on insect populations in multiple cropping system. *Alexandria J. Agric. Res. (Egypt)* 41(2): 201-208.
1553. Fauquet, C. and Fargette, D. 1990. African cassava mosaic virus: etiology, epidemiology, and control. *Plant Dis.* 74(6): 404-411. [Cock (1993)]
1554. Fauquet, C., Fargette, D. and Thouvenel, J.C. 1988. Some aspects of the epidemiology of African cassava mosaic virus in Ivory Coast. *Trop. Pest Manage.* 34(1): 92-96. [Cock (1993)]
1555. Fauquet, C. and Thouvenel, J.C. 1980. Maladies virales des plantes en Cote D'Ivoire. Institut Francais de Recherche Scientifique Pour Le Developpement en Cooperatio n, Orstom, Paris, 243 pp.
1556. Faure-Alvarez, B., Chailloux-Laffita, M. and Vazquez-Moreno, L.L. 2000. Historia y situacion de la produccion de frijol en los paises latinoamericanos afectados por geminivirus transmitidos por mosca blanca: Region caribe: Cuba. pp. 49-54. *In* Bean Golden Mosaic and Other Diseases of Common Bean Caused by Whitefly-Transmitted Geminiviruses in Latin America. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. [Spanish]
1557. Faure, B., Rodriguez, O., Benitez, R. and Carballo, R.M. 1998. Lucha contra geminivirus transmitido por mosca blanca en frijol comun (*Phaseolus vulgaris* L.). [Geminivirus control in common bean (*Phaseolus vulgaris* L.) transmissible by white fly]. pp. 97-99. *In* Produccion De Cultivos En Condiciones Tropicales. Taller Internacional "Geminivirus en el Caribe", Habana, Cuba, Nov 1998. Instituto De Investigaciones Horticolas, La Habana, Cuba. [Spanish, English summary]
1558. Faust, R.M. (Ed.). 1992. Conference Report and 5-Year National Research and Action Plan for Development of Management and Control Methodology for the Sweetpotato Whitefly. U.S. Dept. Agric., Agric. Res. Serv., ARS-107, 165 pp.
1559. Felton, G.W. and Korth, K.L. 2000. Trade-offs between pathogen and herbivore resistance. *Curr. Opin. Plant Biol.* 3(4): 309-314.
1560. Fenigstein, A., Eliyahu, M., GanMor, S. and Veierov, D. 2001. Effects of five vegetable oils on the sweetpotato whitefly *Bemisia tabaci*. *Phytoparasitica* 29(3): 197-206.
1561. Fenoll, C., Schwarz, J.J., Black, D.M., Schneider, M. and Howell, S.H. 1990. The intergenic region of maize streak virus contains a GC-rich element that activates rightward transcription and binds maize nuclear factors. *Plant. Mol. Biol.* 15: 865-877.
1562. Fernandez, M. and Ramos, M. 1995. Incidencia de plagas y biorreguladores sobre variedades de papas adaptadas al calor. [Incidence of pests and bioregulators in varieties of potatoes adapted to heat]. *Rev. Proteccion Vegetal* 10(2): 133-142. [Spanish, English summary]
1563. Fernando, H.E. and Peiris, J.W.L. 1957. Investigations on the chilli leaf-curl complex and its control. *Trop. Agriculturist* 113: 305-323. [Cock (1986)]
1564. Fernando, M. and Udurawana, S.B. 1942. The nature of the mosaic disease of bandakka (*Hibiscus esculentus* L.). *Trop. Agriculturist* 98: 16-24. [Cock (1986)]
1565. Ferreira, P.T.O., Bezerra, I.C., Villas, B.G.L., Ribeiro, S.G. and Giordano, L.B. 1999. Evaluation of sources of resistance to a whitefly-transmitted geminivirus isolate with a bipartite genome in *Lycopersicon*. *Fitopatol. Brasileira* 24(2): 131-135.
1566. Ferrière, C. 1965. Hymenoptera Aphelinidae. pp. 132-133. *In* Faune de l'Europe et du Bassin Méditerranéen. Paris; Masson et Cie Editeurs. [Cock (1986)]
1567. French-Constant, R., Aronstein, K., Anthony, N. and Coustau, C. 1995. Polymerase chain reaction-based monitoring techniques for the detection of insecticide resistance-associated point mutations and their potential applications. *Pestic. Sci.* 43(3): 195-200.

1568. French-Constant, R.H., Anthony, N., Aronstein, K., Rocheleau, T. and Stilwell, G. 2000. Cyclodiene insecticide resistance: From molecular to population genetics. *Annu. Rev. Entomol.* 45: 449-466.
1569. French-Constant, R.H., Anthony, N.M., Andreev, D. and Aronstein, K. 1996. Single versus multiple origins of insecticide resistance: inferences from the cyclodiene resistance gene Rdl. pp. 106-116. *In* Molecular genetics and evolution of pesticide resistance. American Chem. Soc., Washington, D.C.
1570. Fishler, G., Butler, G.D., Jr. and Wilson, F.D. 1988. Cotton leaf pubescence and relationship to leafhopper and sweetpotato whitefly populations. pp. 301-302. *In* J.M. Brown and D.A. Richter (Eds.), Proceedings Beltwide Cotton Production Research Conferences. National Cotton Council, Memphis, TN.
1571. Fishpool, L.D.C. and Burban, C. 1994. *Bemisia tabaci*: The whitefly vector of African cassava mosaic geminivirus. *Trop. Sci.* 34(1): 55-72.
1572. Fishpool, L.D.C., Fargette, D., Colvin, J., Thouvenel, J.C., Burban, C. and Fauquet, C. 1996. Sexual dimorphism of fourth-instar whitefly nymphs on cassava in the Cote d'Ivoire. *Trop. Sci.* 36(3): 154-158.
1573. Fishpool, L.D.C., Fauquet, C., Fargette, D., Thouvenel, J.C., Burban, C. and Colvin, J. 1995. The phenology of *Bemisia tabaci* (Homoptera: Aleyrodidae) populations on cassava in Southern Cote D'Ivoire. *Bull. Entomol. Res.* 85(2): 197-207.
1574. Flanders, S.E. 1969. Herbert D. Smith's observations of citrus blackfly parasites in India and Mexico and the correlated circumstances. *Canadian Entomol.* 101: 467-480. [Cock (1986)]
1575. Flint, H.M., Henneberry, T.J., Wilson, F.D., Holguin, E., Parks, N. and Buehler, R.E. 1995. The effects of transgenic cotton, *Gossypium hirsutum* L., containing *Bacillus thuringiensis* toxin genes for the control of the pink bollworm, *Pectinophora gossypiella* (Saunders) and other arthropods. *Southwest. Entomol.* 20(3): 281-292.
1576. Flint, H.M., Leggett, J.E., Elhoff, L., Parks, N.J. and Davidson, E.W. 1996. Effects of the plant growth regulator mepiquat chloride on silverleaf whitefly (Homoptera: Aleyrodidae) infestations on cotton. *J. Entomol. Sci.* 31(1): 112-122.
1577. Flint, H.M., Naranjo, S.E., Henneberry, T.J., Leggett, J.E. and Hendrix, D.L. 1994. The effect of water stress on infestations of the sweetpotato whitefly. p. 867. *In* D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
1578. Flint, H.M., Naranjo, S.E., Leggett, J.E. and Henneberry, T.J. 1996. Cotton water stress, arthropod dynamics, and management of *Bemisia tabaci* (Homoptera: Aleyrodidae). *J. Econ. Entomol.* 89(5): 1288-1300.
1579. Flint, H.M. and Parks, N.J. 1989. Effect of azadirachtin from the neem tree on immature sweetpotato whitefly, *Bemisia tabaci*, (Homoptera: Aleyrodidae) and other selected pest species on cotton. *J. Agric. Entomol.* 6(4): 211-216. [Cock (1993)]
1580. Flint, H.M. and Parks, N.J. 1990. Infestation of germplasm lines and cultivars of cotton in Arizona USA by whitefly nymphs (Homoptera: Aleyrodidae). *J. Entomol. Sci.* 25(2): 223-229. [Cock (1993)]
1581. Flint, H.M., Parks, N.J., Hendrix, D.L., Wilson, F.D. and Radin, J. 1992. Whitefly population growth in cotton. A 3-year study in Maricopa, Arizona. U.S. Dept. Agric., Agric. Res. Serv., ARS-93, 15 pp.
1582. Flint, H.M., Parks, N.J., Holmes, J.E., Jones, J.A. and Higuera, C.M. 1995. Tests of garlic oil for control of the silverleaf whitefly, *Bemisia argentifolii* Bellows and Perring (Homoptera: Aleyrodidae) in cotton. *Southwest. Entomol.* 20(2): 137-150.
1583. Flint, H.M., Radin, J.W., Parks, N.J. and Reaves, L.L. 1995. The effects of drip or furrow irrigation of cotton on *Bemisia argentifolii* (Homoptera: Aleyrodidae). *J. Agric. Entomol.* 12(1): 25-32.
1584. Flint, H.M., Wilson, F.D., Hendrix, D., Leggett, J., Naranjo, S., Henneberry, T.J. and Radin, J.W. 1993. The effect of water stress on two short-season cultivars of cotton, *Gossypium hirsutum* L. and the sweetpotato whitefly, *Bemisia tabaci* Genn. pp. 253-255. *In* Cotton, A College of Agriculture Report, Series P-94. University of Arizona, Tucson.
1585. Flint, H.M., Wilson, F.D., Hendrix, D., Leggett, J., Naranjo, S., Henneberry, T.J. and Radin, J.W. 1994. The effect of plant water stress on beneficial and pest insects including the pink bollworm and the sweetpotato whitefly in two short-season cultivars of cotton. *Southwest. Entomol.* 19(1): 11-22.
1586. Flint, M.L. 1995. Whiteflies in California: A resource for cooperative extension. *Univ. California IPM Publ.* 19, 53 pp.
1587. Flock, R.A. and Mayhew, D.E. 1981. Squash leaf curl, a new disease of cucurbits in California. *Plant Dis.* 65: 75-76. [Cock (1986)]
1588. Flores, E. and Silberschmidt, K. 1958. Relations between insect and host plant in transmission experiments with infectious chlorosis of Malvaceae. *Ann. Acad. Brasileira Ciencias* 30: 535-560. [Cock (1986)]
1589. Flores, E. and Silberschmidt, K. 1966. Studies on a new virus disease of *Phaseolus longepedunculatus* Ann. Acad. Brasileira Ciencias 38: 327-334. [Cock (1986)]
1590. Flores, E. and Silberschmidt, K. 1967. Contribution to the problem of insect and mechanical transmission of infectious chlorosis of Malvaceae and the disease displayed by *Abutilon thompsonii*. *Phytopathol. Z.* 60: 181-195. [Cock (1986)]
1591. Flores, E., Silberschmidt, K. and Kramer, M. 1960. Observacoes de "clorose infecciosa" das malvaceas em tomateiros do campo. [English summary]. *Biologia* 26(4): 65-69. [Cock (1986)]
1592. Fluckiger, C.R., Kristinsson, H., Senn, R., Rindlisbacher, A., Buholzer, H. and Voss, G. 1992. A novel agent to control aphids and whiteflies. pp. 43-50. *In* Brighton Crop Protection Conference: Pests and Diseases. The British Crop Protection Council, Farnham, UK.
1593. Foda, M.E. 2000. Population dynamics, host preference and seasonal distribution patterns of whitefly, *Bemisia tabaci*, in middle Egypt. pp. 1380-1382. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
1594. Foda, M.E., Salem, M. and Attia, M.B. 1994. Population dynamics of white fly *Bemisia tabaci* (Genn.) in certain vegetable crops in Egypt. *Mansoura Univ. J. Agric. Sci. (Egypt)* 19(3): 1233-1243.
1595. Foltyn, S. and Gerling, D. 1985. The parasitoids of the aleyrodid *Bemisia tabaci* in Israel: development, host preference and discrimination of the aphelinid wasp *Eretmocerus mundus*. [French summary]. *Entomol. Exp. Appl.* 38(3): 255-260. [Cock (1986, 1993)]
1596. Fonseca, M.A., Machado, J., Perez, J., Bruqueta, D., Tornes, C., Verdecia, A., Guerrero, C. and Castella, M.I. 1999. Listado de plantas hospedantes de la mosca blanca (*Bemisia* spp) (Homoptera: Aleyrodidae) asociadas con el tomate (*Lycopersicon esculentum* Mill.) en agroecosistemas del Valle del Caucho. [Record of white fly (*Bemisia* spp)(Homoptera: Aleyrodidae) host plants related with tomato (*Lycopersicon esculentum* Mill.) in Valle del Caucho agroecosystems]. *Fitosanidad (Cuba)* 3(4): 13-15. [Spanish, English summary]
1597. Fontes, E.P.B., Gladfelter, H.J., Schaffer, R.L., Petty, I.T.D. and Hanley-Bowdoin, L. 1994. Geminivirus replication origins have a modular organization. *Plant Cell* 6: 405-416.
1598. Fontes, E.P.B., Luckow, V.A. and Hanley-Bowdoin, L. 1992. A geminivirus replication protein is a sequence-specific DNA binding protein. *Plant Cell* 4: 597-608.
1599. Fook, U.L. 1999. Evaluation of the agronomic characteristics of twelve tomato varieties under local ecological conditions. pp. 59-64. *In* G. Bala and M. Fortune (Eds.), Empowering Farmers Through Agricultural Research, Proceedings of Research Division, Research Seminar Series Held at Centeno, Trinidad and Tobago, October, 1998. Ministry of Agriculture, Land and Marine Resources, Trinidad and Tobago.



1600. Forer, G. 1990. Whitefly management in Israel to prevent honeydew contamination. pp. 33-37. *In* Cotton Production Research from a Farming Systems Perspective, with Special Emphasis on Stickiness. 49th Plenary Meeting. International Cotton Advisory Committee, .
1601. Forsyth, J. 1966. Agricultural insects of Ghana. Ghana University Press, Accra, Ghana, 163 pp. [Cock (1986), *Bemisia tabaci*; pp. 1-7, 54, 62, 111, 117-118]
1602. Fortmann, M. 1992. [Biological control organisms in tomato cultures]. Gärtnerbörse und Gartenwelt (Germany) 92(15): 723-727. [ German]
1603. Foster, R. 1987. The sweet potato whitefly: a potential "new" pest of vegetables in south Florida. pp. 3-4. *In* Lettuce Research Workshop, October 15, 1987, Belle Glade, Florida, No. 1987-3. Everglades Agricultural Research and Education Center, Belle Glade, Florida.
1604. Fouad, E.O. and Mohamed, A. 1997. Morocco. pp. 43-44. *In* N. Ioannou (Ed.), Management of the whitefly-virus complex. FAO Plant Production and Protection Paper 143, Rome, Italy.
1605. Fowler, H.D. 1956. Some physiological effects of attack by whitefly (*Bemisia gossypiperda*) and of spraying parathion on cotton in the Sudan Gezira. Empire Cotton Growing Rev. 33: 288-299. [Cock (1986)]
1606. Franca, F.H., Villas-Boas, G.L. and Branco, M.C. 1996. Ocorrência de *Bemisia argentifolii* Bellows & Perring (Homoptera: Aleyrodidae) em Distrito Federal. [Occurrence of *Bemisia argentifolii* Bellows & Perring (Homoptera: Aleyrodidae) in the Federal District, Brazil]. Anais Soc. Entomol. Brasil 25(2): 369-372. [Portuguese, English summary]
1607. Francki, R.I.B., Hatta, T., Boccardo, G. and Randles, J.W. 1980. The composition of chloris striate mosaic virus, a geminivirus. Virology 101: 233-241.
1608. Franco, M.I., Castillo-Garriga, A., Donoso, I., Reina, J. and Bejarano, E.R. 1999. Viruses transmitted by whitefly: tomato yellow leaf curl virus (TYLCV). Control strategies. pp. 9-39. *In* I.M.C. Gomez (Ed.), Current trends in epidemiology and virus control in horticultural crops. Fundacion para la investigacion Agraria en la Provincia de Almeria, Aguadulce, Spain. [Spanish and English]
1609. Franquinho-Aguiar, A.M. 1998. New records of whiteflies (Homoptera: Aleyrodidae) from Madeira Island with some taxonomical notes. Boletim Museu Municipal, Funchal Suplemento 5(a): 9-26.
1610. Fransen, J.J. 1990. Development of *Bemisia tabaci* Genn. (Homoptera: Aleyrodidae) on poinsettia and other pot plants grown under glass. Bull. OILB/SROP (IOBC/WPRS) 13: 61-63.
1611. Fransen, J.J. 1990. Natural enemies of whiteflies: fungi. pp. 187-210. *In* D. Gerling (Ed.), Whiteflies: their Bionomics, Pest Status and Management. Intercept, Andover, UK. [Cock (1993)]
1612. Fransen, J.J. 1994. *Bemisia tabaci* in the Netherlands - here to stay. Pestic. Sci. 42(2): 129-134.
1613. Franzmann, B., Lea, D. and De Barro, P. 1998. Silverleaf whitefly on the march. Aust. Cottongrower 19(4): 18-19.
1614. Franzmann, B.A., Lea, D.R. and De Barro, P.J. 1998. The distribution and parasitism of biotypes of the whitefly *Bemisia tabaci* in cotton areas of Queensland. pp. 461-464. *In* Proceeding 9th Australian Cotton Conference.
1615. Frappa, C. 1938. Description de *Bemisia manihotis* n. sp. (Hem. Hom. Aleyrodidae) nuisible au manioc a Madagascar. Bull. Entomol. Soc. France 43(1-2): 30-32. [Cock (1986)]
1616. Frappa, C. 1938. Les insectes nuisibles au manioc sur pied et aux tubercules de manioc en magasin a Madagascar. Rev. Bot. Appl. Agric. Trop. 18(197-198): 17-29, 104-109. [Cock (1986)]
1617. Frappa, C. 1939. Note sur une nouvelle espece d'aleurode nuisible aux plantations de tabac de la Tsiribihina. Bull. Economique Madagascar N. S. 16(1938 no. 4): 254-259. [Cock (1986)]
1618. Freeman, R. 1988. Sweet potato whitefly: then what? Long Island Hortic. News (January): 1.
1619. Freeman, T.P., Buckner, J.S., Nelson, D.R., Chu, C.C. and Henneberry, T.J. 2001. Stylet penetration by *Bemisia argentifolii* (Homoptera: Aleyrodidae) into host leaf tissue. Ann. Entomol. Soc. Am. 94(5): 761-768.
1620. Friedmann, M., Lapidot, M., Cohen, S. and Pilowsky, M. 1998. A novel source of resistance to tomato yellow leaf curl virus exhibiting a symptomless reaction to viral infection. J. Am. Soc. Hortic. Sci. 123(6): 1004-1007.
1621. Frischmuth, S., Frischmuth, T. and Jeske, H. 1991. Transcript mapping of abutilon mosaic virus, a geminivirus. Virology 185: 596-604.
1622. Frischmuth, S., Frischmuth, T., Latham, J.R. and Stanley, J. 1993. Transcriptional analysis of the virion-sense genes of the geminivirus beet curly top virus. Virology 197: 312-319.
1623. Frischmuth, T., Roberts, S., von Arnim, A. and Stanley, J. 1993. Specificity of bipartite geminivirus movement proteins. Virology 196: 666-673.
1624. Frischmuth, T. and Stanley, J. 1991. African cassava mosaic virus DNA interferes with the replication of both genomic components. Virology 183: 539-544.
1625. Frischmuth, T., Zimmat, G. and Jeske, H. 1990. The nucleotide sequence of abutilon mosaic virus reveals prokaryotic as well as eukaryotic features. Virology 178: 461-468.
1626. Frohlich, D.R., Brown, J.K., Bedford, I. and Markham, P. 1996. Mitochondrial 16S ribosomal subunit as a molecular marker in *Bemisia*, and implications for population variability. pp. 143-145. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia* : 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
1627. Frohlich, D.R., Torres-Jerez, I., Bedford, I.D., Markham, P.G. and Brown, J.K. 1999. A phylogeographical analysis of the *Bemisia tabaci* species complex based on mitochondrial DNA markers. Mol. Ecol. 8(10): 1683-1691.
1628. Fu-Castillo, A.A. and Silva-Sanchez, F.C. 1997. Manejo integrado de Mosquita blanca de la hoja plateada (*Bemisia argentifolii*). [Integral management of white fly of *Bemisia argentifolii* (regional experiences for handling and control)]. INIFAP, Hermosillo, Sonora, Mexico, 59 pp. [Spanish]
1629. Fuentes-Blandon, H., Cruz, G.E., Castillo, T., Calderon, O. and Cisnero, R. 1987. Reduccion del suelo excesivo de productos quimicos plagas (*Bemisia tabaci*) del frijol (*Phaseolus vulgaris*). [Reduction of excessive soil of chemical product against pests (*Bemisia tabaci*) of beans (*Phaseolus vulgaris*)]. Facultad de Agronomia, Universidad Nacional Agraria, Managua, Nicaragua, 15 pp. [Spanish]
1630. Fullerton, D. 1982. Effects of plant coverage in whitefly control. pp. 117-118. *In* Cotton, A College of Agriculture Report, Series P-56. University of Arizona, Tucson.
1631. Fullerton, D. and Morgan, J. 1982. Evaluation of whitefly control. p. 117. *In* Cotton, A College of Agriculture Report, Series P-56. University of Arizona, Tucson.
1632. Fulmek, L. 1943. Wirtsindex der Aleyrodiden-und Cocciden-Parasiten. Entomol. Beihefte aus Berlin-Dahlem 10: 1-100. [Cock (1986)]
1633. Funk, C.J. 2001. Alkaline phosphatase activity in whitefly salivary glands and saliva. Arch. Insect Biochem. Physiol. 46(4): 165-174.
1634. Funk, C.J., Hunter, W.B. and Achor, D.S. 2001. Replication of Insect iridescent virus 6 in a whitefly cell line. J. Invertebr. Pathol. 77(2): 144-146.
1635. Furuie, T. and Kiyota, H. 1996. Effect of a parasitoid *Encarsia formosa* Gahan on the density of the silverleaf whitefly, *Bemisia argentifolii* Bellows and Perring, on tomatoes in covered cultivation. Proc. Assoc. Plant Prot. Kyushu (Japan) 42: 114-118. [Japanese]
1636. Gabarra, R., Arnó, J., Alomar, O. and Albajes, R. 1999. Naturally occurring populations of *Encarsia pergandiella* (Hymenoptera: Aphelinidae) in tomato greenhouses. Bull. OILB/SROP (IOBC/WPRS) 22: 85-88.

1637. Gabarra, R. and Besri, M. 1999. Tomatoes. pp. 420-434. *In* R. Albajes, M.L. Gullino, van Lenteren J. C. and Y. Elad (Eds.), *Integrated Pest and Disease Management in Greenhouse Crops*. Kluwer, Dordrecht, The Netherlands.
1638. Gadd, C.H. and Loos, C.A. 1941. A virus disease of *Ageratum conyzoides* and tobacco. *Trop. Agriculturist* 96: 255-264. [Cock (1986)]
1639. Gallitelli, D., Luisoni, E., Martinelli, G.P., Caciagli, P., Milne, R.G., Accotto, G.P. and Antignus, Y. 1991. [Tomato yellow leaf curl disease in Sardinia]. *Informatore Fitopatol.* 41(7-8): 42-46. [Cock (1993), Italian, English summary]
1640. Galvez, G.E. 1989. Whitefly-transmitted viruses. pp. 379-406. *In* H.F. Schwartz and M.A. Pastor-Corrales (Eds.), *Bean Production Problems in the Tropics*. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia.
1641. Galvez, G.E. 1991. Informe Anual del Programa Regional de Frijol para la Zona Andina. [Annual report of the Regional Bean Program for the Andean Zone]. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia, 96 pp. [Spanish]
1642. Galvez, G.E. and Castaño, M.J. 1976. Purification of the whitefly-transmitted bean golden mosaic virus. *Turrialba* 26: 205-207. [Cock (1986)]
1643. Gameel, O.I. 1969. Studies on whitefly parasites *Encarsia lutea* Masi and *Eretmocerus mundus* Mercet. (Hymenoptera: Aphelinidae). *Rev. Zool. Bot. Afr.* 79: 65-77. [Cock (1986)]
1644. Gameel, O.I. 1970. The effects of whitefly on cotton. pp. 265-280. *In* M.A. Siddig and L.C. Hughes (Eds.), *Growth of Cotton in the Gezira Environment*. Agric. Res. Corp., Sudan, Cambridge: Heffer.
1645. Gameel, O.I. 1971. The whitefly eggs and first larval stages as prey for certain phytoseiid mites. *Rev. Zool. Bot. Afr.* 84: 79-82.
1646. Gameel, O.I. 1972. A new description, distribution and hosts of the cotton whitefly *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae). *Rev. Zool. Bot. Afr.* 86(1-2): 50-64. [Cock (1986)]
1647. Gameel, O.I. 1974. Field evaluation of insecticides for Jassid, *Empoasca lybica* De Berg, and whitefly, *Bemisia tabaci* (Gennadius), control on cotton (Hemiptera: Jassidae and Aleyrodidae). *Bull. Entomol. Soc. Egypt Econ. Ser.* 7(1973): 113-122. [Cock (1986)]
1648. Gameel, O.I. 1974. Some aspects of the mating and oviposition behaviour of the cotton whitefly *Bemisia tabaci* (Genn.). *Rev. Zool. Bot. Afr.* 88: 784-788. [Cock (1986)]
1649. Gameel, O.I. 1978. The cotton whitefly *Bemisia tabaci* (Genn.) in the Sudan Gezira. pp. 111-131. *In* 3rd Seminar on Strategy for Cotton Pest Control in the Sudan. 8-10 May 1978. Basle, Switzerland. [Cock (1993)]
1650. Gameel, O.I. 1978. Effectiveness of certain insecticides [against cotton pests] on populations of whitefly (*Bemisia tabaci* Genn.), jassid, and American bollworm (*Heliothis armigera* Hbn.). pp. 108-113. *In* Annu. Rep. Gezira Res. Stn. Substns. 1970/1971. Agric. Res. Corp., Sudan.
1651. Gameel, O.I. 1979. Effectiveness of insecticides against whitefly, jassids and American bollworm. pp. 121-125. *In* Annu. Rep. Gezira Res. Stn. Substns. 1972/1973. Agric. Res. Corp., Sudan [Cock (1993)]
1652. Gameel, O.I. 1985. Biology of the cotton whitefly. pp. 93-98. *In* Annu. Rep. Gezira Res. Stn. Substns. 1977/1978. Agric. Res. Corp., Sudan [Cock (1993)]
1653. Gameel, O.I. 1985. Effectiveness of insecticides against whitefly and jassid. pp. 91-92. *In* Annu. Rep. Gezira Res. Stn. Substns. 1977/1978. Agric. Res. Corp., Sudan [Cock (1993)]
1654. Gameel, O.I. and Osman, B.M. 1979. Insecticides and mixtures of insecticides to control insect pests attacking cotton. pp. 126-129. *In* Annu. Rep. Gezira Res. Stn. Substns. 1972/1973. Agric. Res. Corp., Sudan [Cock (1993)]
1655. Gamez, R. 1971. Los virus del frijol en Centroamerica. I. Transmision por moscas blancas (*Bemisia tabaci* Gen.) y plantas hospedantes del virus del mosaico dorado. *Turrialba* 21: 22-27. [Cock (1986)]
1656. Gan-Mor, S. and Grinstein, A. 1997. Recent developments in a sprayer for controlling *Bemisia* in adult cotton with pesticides of low toxicity. ASAE Annual International Meeting, Minneapolis, MN, 10-14 Aug. 1997. American Society of Agricultural Engineers, St Joseph, Michigan, 10 pp.
1657. Ganapathy, N. and Durairaj, C. 1995. Pest status of pulses in Pudukkottai District, Tamil Nadu. *Madras Agric. J.* 82(4): 322.
1658. Ganeshan, S. and Abeeluck, D. 2000. First record of the silverleaf whitefly, *Bemisia argentifolii* Bellows & Perring (Hemiptera: Aleyrodidae), from Mauritius. *African Entomol.* 8(2): 303-304.
1659. Gangwar, S.K. and Sachan, J.N. 1981. Seasonal incidence and control of insect pests of brinjal with special reference to shoot and fruit borer, *Leucinodes orbonalis* Guen. in Maghelaya. *J. Res. (Assam Agric. Univ.)* 2(2): 187-192. [Cock (1986)]
1660. Gara-Gonzalez, E. 1993. Enemigos naturales de accion microbial de mosquita blanca. pp. 54-56. *In* L.A. Rodriguez del Bosque (Ed.), *Proceedings 2nd Taller sobre Control Biologico de Mosquita Blanca*, SARH, Sinaloa, Mexico.
1661. Garcia-Carrasco, J.L., Sese, A.I.L. and Gomez-Guillamon, M.L. 1996. Host range of a melon yellowing virus transmitted by *Bemisia tabaci* (Gennadius) in Southern Spain. *Rep. Cucurbit Genetics Coop.* 19: 57-58.
1662. Garcia-Sandoval, J.A. and Gutierrez-Baeza, A.O. 1998. Impacto de *Paecilomyces fumosoroseus* contra la mosca blanca *Bemisia tabaci* en Quintana Roo. [Impact of *Paecilomyces fumosoroseus* against white fly *Bemisia tabaci* in Quintana Roo State]. pp. 189-192. *In* Proceedings 21st National Congress of Biological Control, Rio Bravo, Tampaulipas, Mexico, 5-6 Nov 1998. INIFAP. [Spanish]
1663. Gardiner, W.E., Sunter, G., Brand, L., Elmer, J.S., Rogers, S.G. and Bisaro, D.M. 1988. Genetic analysis of tomato golden mosaic virus: the coat protein is not required for systemic spread or symptom development. *EMBO J.* 7(4): 899-904.
1664. Garthwaite, D. 2000. Changes in biological control usage in Great Britain between 1968 and 1995 with particular reference to biological control on tomato crops. *Biocontrol Sci. Technol.* 10(4): 451-457.
1665. Garzon-Tiznado, J.A. 1998. Geminivirus transmitidos por mosquita blanca. [Geminivirus transmitted by whiteflies]. pp. 93-112. *In* J.J. Pacheco-Covarrubias and F. Pacheco-Mendivil (Eds.), *Temas Selectos Para El Manejo Integrado de la Mosquita Blanca*, Memoria Cientifica No. 6. Instituto Nacional De Investigaciones Forestales, Agricolas y Pecuarias, Centro de Investigacion Regional del Noroeste, CIRNO, Obregon, Sonora, Mexico.
1666. Garzon-Tiznado, J.A., Torres-Pacheco, I., Trinidad, J., Ibanez, A., Herrera-Estrella, L. and Rivera-Bustamante, R.F. 1993. Inoculation of peppers with infectious clones of a new geminivirus by a biolistic procedure. *Phytopathology* 83(5): 514-521.
1667. Gatehouse, A.G. 1997. Behavior and ecological genetics of wind-borne migration by insects. *Annu. Rev. Entomol.* 42: 475-502.
1668. Gaur, S.K. and Deshpande, R.R. 1998. Relative susceptibility of soybean *Glycine max* (E) varieties to insect pests complex. *Indian J. Plant Prot.* 26(2): 186-187.
1669. Gavriel, I., Patsias, A. and Ioannou, N. 1997. Cyprus. pp. 13-18. *In* N. Ioannou (Ed.), *Management of the whitefly-virus complex*. FAO Plant Production and Protection Paper 143, Rome, Italy.
1670. Gawel, N.J. and Bartlett, A.C. 1993. Characterization of differences between whiteflies using RAPD-PCR. *Insect Biochem. Mol. Biol.* 2(1): 33-38.
1671. Gawel, N.J. and Bartlett, A.C. 1993. Differentiation of SPWF biotypes using RAPD-PCR. pp. 953-954. *In* D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.

1672. Gawel, N.J. and Bartlett, A.C. 1993. Differentiation of sweet potato whitefly biotypes using RAPD-PCR. pp. 258-261. *In* Cotton, A College of Agriculture Report, Series P-94. University of Arizona, Tucson.
1673. Geneif, A.A. 1986. Attempts to control tomato leafcurl virus on tomato in the Sudan. *Acta Hort.* 190: 145-157. [CATIE (1992)]
1674. Gennadius, P. 1889. Disease of tobacco plantations in the Trikonía. The aleurodid of tobacco. *Ellenike Georgia* 5: 1-3. [Cock (1986), Greek]
1675. Gentry, J.W. 1965. Crop insects of Northeast Africa-southwest Asia. USDA Agric. Handbook 271 : 1-12, 12-14, 57, 62, 145, 147, 168-169, 184, 186. [Cock (1986)]
1676. Geraud-Pouey, F., Chirinos, D.T. and Vergara, J.A. 1996. Efectos colaterales de algunos tratamientos con insecticidas sobre entomofauna del tomate, *Lycopersicon esculentum* Miller, cv. Peto Seed 98, en la zona del río Limón, Estado Zulia, Venezuela. [Side effects of insecticide treatments on tomato, *Lycopersicon esculentum* Miller, cv. Peto Seed 98, entomofauna in the Limón River Zone, State of Zulia, Venezuela]. *Rev. Facultad Agron., Univ. Zulia (Venezuela)* 13(3): 313-325. [Spanish, English summary]
1677. Geraud-Pouey, F., Chirinos T. L., Chirinos, D.T., Miranda, M. and Tejera, A. 1997. Side effects of insecticide treatments on melon, *Cucumis melo* L. entomofauna. *Rev. Facultad Agron., Univ. Zulia (Venezuela)* 14(2): 225-232. [Spanish, English summary]
1678. Gergis, M.F. 1994. Comparative oviposition and feeding host preference of the silverleaf and sweetpotato whiteflies. pp. 870-872. *In* D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
1679. Gergis, M.F. 1994. Population growth and life table analysis in relation to temperature and host plants. pp. 1229-1230. *In* D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
1680. Gergis, M.F. 1994. Thermal requirements, developmental rates and prediction models of whiteflies in relation to temperature and host plants. pp. 1207-1210. *In* D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
1681. Gergis, M.F. and Adam, K.M. 1996. Variation in *Bemisia tabaci* populations based on host plant associations, developmental statistics and growth parameters in Egypt. pp. 1007-1011. *In* P. Dugger and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
1682. Gerling, D. 1966. Biological studies on *Encarsia formosa* (Hymenoptera: Aphelinidae). *Ann. Entomol. Soc. Am.* 59: 142-143.
1683. Gerling, D. 1966. Studies with whitefly parasites of Southern California. I. *Encarsia pergandiella* Howard (Hymenoptera: Aphelinidae). *Canadian Entomol.* 98: 707-724.
1684. Gerling, D. 1966. Studies with whitefly parasites of southern California. II. *Eretmocerus californicus* Howard (Hymenoptera: Aphelinidae). *Canadian Entomol.* 98: 1316-1329. [Cock (1986)]
1685. Gerling, D. 1967. Bionomics of the whitefly-parasite complex associated with cotton in southern California (Homoptera: Aleyrodidae; Hymenoptera: Aphelinidae). *Ann. Entomol. Soc. Am.* 60: 1306-1321.
1686. Gerling, D. 1972. Notes on three species of *Eretmocerus* Haldeman occurring in Israel with a description of a new species. *Entomol. Berichten* 32: 156-161. [Cock (1986)]
1687. Gerling, D. 1983. Factors affecting the establishment of *Bemisia tabaci* on cotton in Israel. p. 959. *In* Plant Protection for Human Welfare. British Crop Protection Council, Croydon, UK. [Cock (1986)]
1688. Gerling, D. 1984. The overwintering mode of *Bemisia tabaci* and its parasitoids in Israel. *Phytoparasitica* 12: 109-118. [Cock (1986)]
1689. Gerling, D. 1985. Parasitoids attacking *Bemisia tabaci* (Hom.: Aleyrodidae) in Eastern Africa. *Entomophaga* 30: 163-165. [Cock (1986)]
1690. Gerling, D. 1986. Natural enemies of *Bemisia tabaci*, biological characteristics and potential as biological control agents: A review. *Agric. Ecosystems Environ.* 17(1-2): 99-110. [Cock (1993)]
1691. Gerling, D. 1990. Natural enemies of whiteflies: predators and parasitoids. pp. 147-185. *In* D. Gerling (Ed.), *Whiteflies: their Bionomics, Pest Status and Management*. Intercept, Andover, UK. [Cock (1993)]
1692. Gerling, D. 1992. Approaches to the biological control of whiteflies. *Florida Entomol.* 75: 446-456.
1693. Gerling, D. 1996. Status of *Bemisia tabaci* in the Mediterranean countries: Opportunities for biological control. *Biol. Control* 6(1): 11-22.
1694. Gerling, D., Alomar, O. and Arnó, J. 2001. Biological control of *Bemisia tabaci* using predators and parasitoids. *In* S. E. Naranjo and P. C. Ellsworth (Eds.), *Special Issue: Challenges and Opportunities for Pest Management of Bemisia tabaci in the New Century*. *Crop Prot.* 20(9): 779-799.
1695. Gerling, D. and Foltyn, S. 1987. Development and host preference of *Encarsia lutea* (Masi) and interspecific host discrimination with *Eretmocerus mundus* (Mercet) (Hymenoptera, Aphelinidae) parasitoids of *Bemisia tabaci* (Gennadius) (Homoptera, Aleyrodidae). [German summary]. *J. Appl. Entomol.* 103(5): 425-433. [Cock (1993)]
1696. Gerling, D. and Fried, R. 1997. Density-related sterility in *Eretmocerus mundus*. *Entomol. Exp. Appl.* 84(1): 33-39.
1697. Gerling, D. and Fried, R. 2000. Biological studies with *Eretmocerus mundus* Mercet (Hymenoptera: Aphelinidae) in Israel. *Bull. OILB/SROP (IOBC/WPRS)* 23(1): 117-123.
1698. Gerling, D. and Horowitz, A.R. 1984. Yellow traps for evaluating the population levels and dispersal patterns of *Bemisia tabaci* (Homoptera: Aleyrodidae). *Ann. Entomol. Soc. Am.* 77: 753-759. [Cock (1986)]
1699. Gerling, D., Horowitz, A.R. and Baumgärtner, J. 1986. Autecology of *Bemisia tabaci*. *Agric. Ecosystems Environ.* 17: 5-19. [Cock (1993)]
1700. Gerling, D. and Kravchenko, V. 1996. Pest management of *Bemisia* out of doors. pp. 667-680. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia : 1995 Taxonomy, Biology, Damage, Control and Management*. Intercept Ltd., Andover, Hants, UK.
1701. Gerling, D., Kravchenko, V. and Lazare, M. 1997. Dynamics of common green lacewing (Neuroptera: Chrysopidae) in Israeli cotton fields in relation to whitefly (Homoptera: Aleyrodidae) populations. *Environ. Entomol.* 26(4): 815-827.
1702. Gerling, D. and Lindenbaum, M. 1991. Host plant related behavior of *Bemisia tabaci*. *Bull. OILB/SROP (IOBC/WPRS)* 14: 83-88.
1703. Gerling, D. and Mayer, R.T. (Eds.). 1996. *Bemisia 1995: taxonomy, biology, damage, control and management*. Intercept Limited, Andover, UK, 702 pp.
1704. Gerling, D., Motro, U. and Horowitz, A.R. 1980. Dynamics of *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae) attacking cotton in the Coastal Plain of Israel. *Bull. Entomol. Res.* 70: 213-219. [Cock (1986)]
1705. Gerling, D. and Naranjo, S.E. 1998. The effect of insecticide treatments in cotton fields on the levels of parasitism of *Bemisia tabaci* (Gennadius). *Biol. Control* 12(1): 33-41.
1706. Gerling, D., Orion, T. and Delarea, Y. 1990. *Eretmocerus* penetration and immature development: A novel approach to overcome host immunity. *Arch. Insect Biochem. Physiol.* 13(3-4): 247-254. [Cock (1993)]
1707. Gerling, D., Quicke, D.L.J. and Orion, T. 1998. Oviposition mechanisms in the whitefly parasitoids *Encarsia transvena* and *Eretmocerus mundus*. *Biocontrol* 43(3): 289-297.
1708. Gerling, D. and Rivnay, T. 1984. A new species of *Encarsia* [Hym.: Aphelinidae] parasitizing *Bemisia tabaci* [Hom.: Aleyrodidae]. *Entomophaga* 29: 439-444.

1709. Gerling, D. and Sinai, P. 1994. Buprofezin effects on two parasitoid species of whitefly (Homoptera: Aleyrodidae). *J. Econ. Entomol.* 87(4): 842-846.
1710. Gerling, D., Spivak, D. and Vinson, S.B. 1987. Life history and host discrimination of *Encarsia deserti* (Hymenoptera: Aphelinidae), a parasitoid of *Bemisia tabaci* (Homoptera: Aleyrodidae). *Ann. Entomol. Soc. Am.* 80(2): 224-229.
1711. Gerling, D. and Stern, T.U. 1993. Biology of *Delphastus pusillus*: the influence of density on fitness. *Bull. OILB/SROP (IOBC/WPRS)* 16: 39-42.
1712. Gerling, D., Tremblay, E. and Orion, T. 1991. Initial stages of the vital capsule formation in the *Eretmocerus-Bemisia tabaci* association. *Redia* 74(3): 411-415.
1713. Ghafoor, A., Ahmad, Z., Qureshi, A.S. and Iqbal, S.M. 2000. Significance of mungbean yellow mosaic virus on yield and its components in *Vignamungo* L. Hepper. *Pakistan J. Phytopathol.* 12(2): 74-78.
1714. Ghahari, H. and Hatami, B. 2000. Morphological and biological studies of greenhouse whitefly *Trialetrodes vaporariorum* Westwood (Homoptera: Aleyrodidae) in Isfahan. *J. Sci. Tech. Agric. Nat. Resources* 4(2): 141-154. [Persian, English summary]
1715. Ghanim, A.A., Nassar, O.A. and El-Adl, M.A. 1988. Biological studies on *Chrysopa carnea* Steph., preying on citrus brown mites (*Eutetranychus orientalis*) and white fly (*Bemisia tabaci* (Gennadius)). *Mansoura Univ. J. Agric. Sci. (Egypt)* 13(1): 300-304.
1716. Ghanim, A.A., Said, A.A.A. and El-Adl, M.A. 1988. Studies on soybean injurious insects and efficiency of certain insecticides and their combinations with two insect growth regulators on these insects. *Mansoura Univ. J. Agric. Sci. (Egypt)* 13(2): 934-941.
1717. Ghanim, M. and Czosnek, H. 2000. Tomato yellow leaf curl geminivirus (TYLCV-Is) is transmitted among whiteflies (*Bemisia tabaci*) in a sex-related manner. *J. Virol.* 74(10): 4738-4745.
1718. Ghanim, M., Morin, S. and Czosnek, H. 2001. Rate of tomato yellow leaf curl virus translocation in the circulative transmission pathway of its vector, the whitefly *Bemisia tabaci*. *Phytopathology* 91(2): 188-196.
1719. Ghanim, M., Morin, S., Zeidan, M. and Czosnek, H. 1998. Evidence for transovarial transmission of tomato yellow leaf curl virus by its vector, the whitefly *Bemisia tabaci*. *Virology* 240(2): 295-303.
1720. Ghanim, M., Rosell, R.C., Campbell, L.R., Czosnek, H., Brown, J.K. and Ullman, D.E. 2001. Digestive, salivary, and reproductive organs of *Bemisia tabaci* (Gennadius) (Hemiptera: Aleyrodidae) B type. *J. Morphol.* 248: 22-40.
1721. Gharib, A.H. and Aly, F.K. 1991. Population density and age structure of the whitefly, *Bemisia tabaci* on six field and vegetable crops in Minia region. *Minia J. Agric. Res. Devel. (Egypt)* 13(4): 1541-1556.
1722. Ghavami, M.D. 1994. The development period of the predator *Deraeocoris pallens* Reut. (Hemiptera: Miridae) at different prey and temperatures. pp. 387-394. *In* *Turkiye III. Biyolojik Mucadele Kongresi Bildirileeri*, Oct. 25-28, 1994, Ege Universitesi Ziraat Fakultesi, Bitki Koruma Bolumu, Izmir, Bornova, Turkey. [Turkish, English summary]
1723. Ghavami, M.D. and Ozgur, A.F. 1992. Population development of pests and their interaction with predatory insects in cotton fields. pp. 227-238. *In* *Proceeding 2nd Turkish National Congress Entomol.*, Ismir, Turkey; Ege Univ. [Turkish, English summary]
1724. Ghavami, M.D., Ozgur, A.F. and Kersting, U. 1998. Prey consumption by the predator *Deraeocoris pallens* Reuther (Hemiptera: Miridae) on six cotton pests. *Zeitschrift für pflanzenkrankheiten und Pflanzenschutz* 105(5): 526-531.
1725. Ghesquiére, J. 1934. Un Calliceratide (Hym. Proct.) nouveau du Congo Belge. *Ann. Soc. Zool. Belgique* 65: 59-62. [Cock (1986)]
1726. Ghosh, J., Ghosh, S.K., Chatterjee, H. and Senapati, S.K. 1999. Pest constraints of okra under Terai region of West Bengal. *Indian J. Entomol.* 61(4): 362-371.
1727. Ghosh, S.K. and Senapati, S.K. 2001. Evaluation of brinjal varieties commonly grown in terai region of West Bengal against pest complex. *Crop Res. (Hisar)* 21(2): 157-163.
1728. Ghosh, S.P., Pillai, K.S. and Thankappan, M. 1986. Cassava based multiple cropping system II. Incidence of pests and diseases. *J. Root Crops* 12(2): 83-89. [Cock (1993)]
1729. Gibson, R.W., Legg, J.P. and Otim-Nape, G.W. 1996. Unusually severe symptoms are a characteristic of the current epidemic of mosaic virus disease of cassava in Uganda. *Ann. Appl. Biol.* 128(3): 479-490.
1730. Gibson, R.W., Mpenbe, I., Alicai, T., Carey, E.E., Mwanga, R.O.M., Seal, S.E. and Vetten, H.J. 1998. Symptoms, aetiology and serological analysis of sweet potato virus disease in Uganda. *Plant Pathol.* 47(1): 95-102.
1731. Giha, O.H. and Nour, M.A. 1969. Epidemiology of cotton leafcurl virus in the Sudan. *Cotton Growing Rev.* 46(2): 105-118.
1732. Gilbertson, R.L., Faria, J.C., Hanson, S.F., Morales, F.J., Ahlquist, P., Maxwell, D.P. and Russell, D.R.M. 1991. Cloning of the complete DNA genomes of four bean-infecting geminiviruses and determining their infectivity by electric discharge particle acceleration. *Phytopathology* 81(9): 980-985.
1733. Gilbertson, R.L., Faria, J.C., Leong, S.A., Ahlquist, P.G. and Maxwell, D.P. 1989. Molecular characterization of a Brazilian isolate of bean golden mosaic virus. *Annu. Rep. Bean Improvement Coop.* 32: 92-93.
1734. Gilbertson, R.L., Hidayat, S.H., Martinez, R.T., Leong, S.A., Faria, J.C., Morales, F. and Maxwell, D.P. 1991. Differentiation of bean-infecting geminiviruses by nucleic acid hybridization probes and aspects of bean golden mosaic in Brazil. *Plant Dis.* 75: 336-342.
1735. Gilbertson, R.L., Hidayat, S.H., Paplomatas, E.J., Rojas, M.R., Hou, Y.M. and Maxwell, D.P. 1993. Pseudorecombination between infectious cloned DNA components of tomato mottle and bean dwarf mosaic geminiviruses. *J. Gen. Virol.* 74: 23-31.
1736. Gilbertson, R.L., Rojas, M.R., Russell, D.R. and Maxwell, D.P. 1991. Use of the asymmetric polymerase chain reaction and DNA sequencing to determine genetic variability of bean mosaic geminivirus in the Dominican Republic. *J. Gen. Virol.* 72: 2843-2848.
1737. Gilbertson, R.L., Ullman, D.E., Salati, R., Maxwell, D.P., Grafton-Cardwell, E.E. and Polek, M.L. 1998. Insect-transmitted viruses threaten agriculture. *California Agric.* 52(2): 23-28.
1738. Gill, C.K. and Rataul, H.S. 1986. A note on the incidence, estimation of losses and symptomology of yellow mosaic virus on soybean, *Glycine max* L. *Indian J. Entomol.* 48(4): 524-526.
1739. Gill, C.K. and Rataul, H.S. 1987. Virus vector relationship of soybean yellow mosaic virus and its vector whitefly, *Bemisia tabaci* Gen. in Punjab. *Indian J. Entomol.* 49(1): 46-57. [Cock (1993)]
1740. Gill, C.K. and Rataul, H.S. 1990. Studies on the length of proboscis of some insect vectors in relation to the depth of leaf tissues of some common crop plants. *Indian J. Entomol.* 52(4): 704-706. [Cock (1993)]
1741. Gill, C.K. and Rataul, H.S. 1991. Varietal response in soybean to yellow mosaic virus transmitted by *Bemisia tabaci* (Genn.). *J. Insect Sci.* 4(1): 85-86.
1742. Gill, C.K. and Rataul, H.S. 1994. Leaf cage for studies on cotton white fly *Bemisia tabaci* Guen. *Indian J. Entomol.* 56(2): 201-203.
1743. Gill, C.K. and Singh, L. 1999. Effect of mungbean yellow mosaic virus on yield components in mungbean cultivar. *Insect Environ.* 5(3): 112-113.
1744. Gill, C.K. and Singh, L. 2000. Biochemical changes in mungbean cultivar, ML-267 infected with yellow mosaic virus. *Insect Environ.* 6(2): 86-87.
1745. Gill, R.J. 1990. The morphology of whiteflies. pp. 13-46. *In* D. Gerling (Ed.), *Whiteflies: their Bionomics, Pest Status, and Management*. Intercept, Andover, UK.

- 1746.Gill, R.J. 1992. A review of the sweetpotato whitefly in Southern California. *Pan-Pacific Entomol.* 68(2): 144-152.
- 1747.Gill, R.J. [no date]. Color-photo and host keys to California whiteflies. Scale and whitefly key #2 . Dept. Food Agric., State of California : 1-6 .
- 1748.Gilrein, D. 1994. Let's get bio-rational. *Connecticut Greenhouse Newsl.* 181: 1-5.
- 1749.Gindin, G., Barash, I., Harari, N. and Raccach, B. 1994. Effect of endotoxic compounds isolated from *Verticillium lecanii* on the sweetpotato whitefly, *Bemisia tabaci*. *Phytoparasitica* 22(3): 189-196.
- 1750.Gindin, G. and Ben-Ze'ev, I.S. 1994. Virulence and persistence of *Conidiobolus coronatus* and *Conidiobolus* sp. in glasshouse populations of *Bemisia tabaci*. *Bull. OILB/SROP (IOBC/WPRS)* 17(3): 185-188.
- 1751.Gindin, G. and Benzeev, I.S. 1994. Natural occurrence of and inoculation experiments with *Conidiobolus coronatus* and *Conidiobolus* sp in glasshouse populations of *Bemisia tabaci*. *Phytoparasitica* 22(3): 197-208.
- 1752.Gindin, G., Geschtovt, N.U., Raccach, B. and Barash, I. 2000. Pathogenicity of *Verticillium lecanii* to different developmental stages of the silverleaf whitefly, *Bemisia argentifolii*. *Phytoparasitica* 28(3): 229-239.
- 1753.Giordano, L.B., Bezerra, I.C., Ferreira, P.T.O., Borges-Neto, C.R. and Bieche, B.J. 1999. Breeding tomatoes for resistance to whitefly - transmitted geminivirus with bipartite genome in Brazil. *Acta Hort.* 487: 357-360.
- 1754.Girardeau, J.H. 1958. The sweetpotato whitefly *Bemisia inconspicua* (Q.), as a vector of sweetpotato mosaic in South Georgia. *Plant Dis. Rep.* 42: 819. [ Cock (1986)]
- 1755.Girardeau, J.H. and Ratcliffe, T.J. 1960. The vector-virus relationship of the sweetpotato whitefly and a mosaic of sweetpotatoes in South Georgia. *Plant Dis. Rep.* 44: 48-50. [ Cock (1986)]
- 1756.Githunguri, C.M., Ndong'A, M.F.O. and Amadalo, B.A. 1984. Cassava production and constraints in Kenya. pp. 75-79. In A.H. Greethead, R.H. Markham, R.J. Murphy, S.T. Murphy and I.A.D. Robertson (Eds.), *Integrated Pest Management of Cassava Green Mite. Proceedings of a Regional Training Workshop in East Africa, 30 April - 4 May, 1984.* Commonwealth Institute of Biological Control, Ascot, UK. [Cock (1986)]
- 1757.Giustina, W.D., Martinez, M. and Bertaux, F. 1989. *Bemisia tabaci*: Le nouvel ennemi des cultures sous serres en Europe. *Phytoma (France)* 406: 48-52. [ French, English summary]
- 1758.Gocmen, H. 1995. Yeni bir gozlem: Pamukta sera beyazsinegi *Trialeurodes vaporariorum* (Westw.) (Homoptera, Aleyrodidae). [A new observation: greenhouse whitefly *Trialeurodes vaporariorum* (Westw.) (Homoptera, Aleyrodidae) on cotton. *Türkiye Entomol. Dergisi* 19(2): 111-115.
- 1759.Gocmen, H., Sekeroglu, E. and Ozgur, A.F. 1987. [Effectiveness of different installation heights and positions of yellow sticky traps for catching whiteflies (*Bemisia tabaci* (Genn.))(Homoptera: Aleyrodidae) on cotton]. *Türkiye I. Entomol. Kongresi. Bildirileri* : 367-376. [ Cock (1993), Turkish, English summary]
- 1760.Godfrey, L. and Rosenheim, J. 1996. Aphids and whiteflies in the San Joaquin Valley of California in 1995. pp. 128-132. In P. Dugger and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences.* National Cotton Council, Memphis, TN.
- 1761.Godfrey, L., Summers, C. and Goodell, P. 1996. Silverleaf whitefly in the San Joaquin Valley - year 4. pp. 1023-1026. In P. Dugger and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences.* National Cotton Council, Memphis, TN.
- 1762.Godfrey, L.D., Goodell, P.B., Bentley, W.J., Summers, C.G., Prather, T. and Coviello, R. 1994. Contribution of crops, weeds, and parasites to the abundance of silverleaf whitefly in San Joaquin Valley cotton. pp. 873-874. In D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences.* National Cotton Council, Memphis, TN.
- 1763.Godfrey, L.D., Goodell, P.B. and Summers, C.G. 1995. Silverleaf whitefly population dynamics and management in the San Joaquin Valley. pp. 95-97. In D.A. Richter and J. Armour (Eds.), *Proceedings Beltwide Cotton Conferences.* National Cotton Council, Memphis, TN.
- 1764.Godfrey, L.D., Goodell, P.B., Summers, C.G., Bentley, W.J. and Perring, T.M. 1995. Seasonal development of silverleaf whitefly populations in cotton and other crops in the San Joaquin Valley. pp. 831-834. In D.A. Richter and J. Armour (Eds.), *Proceedings Beltwide Cotton Conferences.* National Cotton Council, Memphis, TN.
- 1765.Godfrey, L.D., Keillor, K., Hutmacher, R.B. and Cisneros, J. 1999. Interaction of cotton aphid population dynamics and cotton fertilization regime in California cotton. pp. 1008-1011. In P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences.* National Cotton Council, Memphis, TN.
- 1766.Gold, C.S., Altieri, M.A. and Bellotti, A.C. 1989. Effects of cassava varietal mixtures on the whiteflies *Aleurotrachelus socialis* and *Trialeurodes variabilis* in Columbia. *Entomol. Exp. Appl.* 53(3): 195-202.
- 1767.Gold, C.S., Altieri, M.A. and Bellotti, A.C. 1989. The effects of intercropping and mixed varieties of predators and parasitoids of cassava whiteflies (Hemiptera: Aleyrodidae) in Columbia. *Bull. Entomol. Res.* 79: 115-122.
- 1768.Gold, C.S., Altieri, M.A. and Bellotti, A.C. 1990. Response of the cassava whitefly, *Trialeurodes variabilis* (Quaintance). (Homoptera: Aleyrodidae) to host plant size: Implications for cropping system management. *Acta Oecologica* 11(1): 35-41.
- 1769.Golding, F.D. 1930. A vector of leaf curl of cotton in Southern Nigeria. *Empire Cotton Growing Rev.* 7: 120-126. [ Cock (1986)]
- 1770.Golding, F.D. 1935. A probable vector of cassava mosaic in southern Nigeria. *Trop. Agric. (Trinidad)* 12: 215. [ Cock (1986)]
- 1771.Golding, F.D. 1936. *Bemisia nigeriensis*, Corb., a vector of cassava mosaic in southern Nigeria. *Trop. Agric. (Trinidad)* 13: 182-186. [ Cock (1986)]
- 1772.Golding, F.D. 1938. Notes on the insect pests of cotton in Nigeria. *Empire Cotton Growing Rev.* 15: 224-227. [ Cock (1986)]
- 1773.Gómez, D. 1996. Generación y transferencia de tecnología. pp. 97-103. In L. Hilje (Ed.), *Metodologías para el estudio y manejo de moscas blancas y geminivirus.* Centro Agronómico Tropical de Investigación y Enseñanza, Turrialba, Costa Rica.
- 1774.Gomez, J. and Garcia, O. 2000. A new species of *Encarsia* (Hymenoptera : Aphelinidae), a parasitoid of whitefly *Aleurodicus* sp (Homoptera : Aleyrodidae) in Mexico. *Pan-Pacific Entomol.* 76(1): 49-51.
- 1775.Gomez-Menor, J. 1953. Algunos insectos como pequenos enemigos: los aleurodidos. *Rev. Univ. Madrid* 2: 27-55. [ Cock (1986)]
- 1776.Gomez-Menor, J. 1954. Aleurodidos de Espana, islas Canarias y Africa occidental. *Eos, Madrid* 30: 363-377. [ Cock (1986)]
- 1777.Gomez-Menor, J. 1968. Estudio de la faunula de Homopteros Sternorrhyncha de la Provincia de Toledo. *Univ. Madrid*, 92 pp. [ Cock (1986)]
- 1778.Gomez, P., Cubillo, D., Mora, G.A. and Hilje, L. 1997. Evaluacion de posibles repelentes de *Bemisia tabaci*. I. Productos comerciales. [Evaluation of possible repellents for *Bemisia tabaci*. I. Commercial products]. *Manejo Integrado de Plagas (Costa Rica)* 46: 9-16. [ Spanish, English summary]
- 1779.Gomez, P., Cubillo, D., Mora, G.A. and Hilje, L. 1997. Evaluacion de posibles repelentes de *Bemisia tabaci*: II. Extractos vegetales. [Evaluation of possible repellent for *Bemisia tabaci*: II. Botanical substances]. *Manejo Integrado de Plagas (Costa Rica)* 46: 17-25. [ Spanish, English summary]
- 1780.Gonzalez, R.A., Goldman, G.E., Natwick, E.T., Rosenberg, H.R., Grieshop, J.I., Sutter, S.R., Funakoshi, T. and Davila-Garcia, S. 1992. Whitefly invasion in Imperial Valley costs growers, workers millions in losses. *California Agric.* 46(5): 7-8.

1781. Gonzalez-Zamora, J.E. and Gallardo, J.M. 1999. Development and reproduction of *Bemisia tabaci* (Gennadius) (Homoptera; Aleyrodidae) on sweet pepper at three temperatures. *Boletín de Sanidad Vegetal, Plagas* 25(1): 3-11. [ Spanish, English summary]
1782. Gonzalez-Zamora, J.E., Gallardo, J.M. and Garcia, M.M. 1997. Toxicity of different pesticides on pupae of *Eretmocerus mundus* Mercet (Hymenoptera: Aphelinidae) parasitizing *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae). *Bull. OILB/SROP (IOBC/WPRS)* 20(4): 114-120.
1783. Gonzalez-Zamora, J.E. and Moreno-Vazquez, R. 1996. Analisis de las tendencias poblacionales de *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae) en pimiento bajo plastico en Almeria. [Analysis of the population trends of *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae) in sweet peppers in plastic greenhouses in Almeria]. *Boletín Sanidad Vegetal Plagas (Argentina)* 22(2): 159-167. [ Spanish, English summary]
1784. Gonzalez-Zamora, J.E., Moreno-Vazquez, R., Rodriguez-Rodriguez, M.D., Rodriguez-Rodriguez, M.P., Mirasol-Carmona, E., Garcia-Teston, J. and Manzanera-Ruiz, C. 1996. Evolucion del parasitismo en *Bemisia tabaci* (Genn.) y *Trialeurodes vaporariorum* (West.) (Homoptera: Aleyrodidae) en invernaderos de Almeria. [Evolution of parasitism of *Bemisia tabaci* and *Trialeurodes vaporariorum* (West.) (Homoptera: Aleyrodidae) in winter pastures in Almeria. [Spain]]. *Boletín de Sanidad Vegetal, Plagas* 22(2): 373-389. [ Spanish, English summary]
1785. González-Zamora, J.G. 1996. Posibilidades de control biológico de *Bemisia tabaci*: situación en Almería. pp. 75-80. In J.L. Cenis (Ed.), *El Virus del Rizado Amirillo (Hoja en Cuchara) del Tomate (TYLCV) y Su Vector Bemisia tabaci*, Jornadas 8. Consejería de Medio Ambiente, Agricultura y Agua, Murcia, Spain.
1786. Goodell, P.B., Godfrey, L.D., Wood, J. and Eckert, J.W. 1995. Distribution of silverleaf whitefly in the San Joaquin Valley. pp. 834-836. In D.A. Richter and J. Armour (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
1787. Goodell, P.B. and Narbeth, S. 1996. Insect population dynamics in San Joaquin Valley cotton fields. pp. 1075-1078. In P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
1788. Goodman, R. 1981. Geminiviruses. *J. Gen. Virol.* 54: 9-21.
1789. Goodman, R.M. 1977. Infectious DNA from a whitefly-transmitted virus of *Phaseolus vulgaris*. *Nature (London)* 266: 54-55. [ Cock (1986)]
1790. Goodman, R.M. 1977. Single-stranded DNA genome in a whitefly-transmitted plant virus. *Virology* 83: 171-179. [ Cock (1986)]
1791. Goodman, R.M. 1981. Geminiviruses. pp. 879-910. In E. Kurstak (Ed.), *Handbook of Plant Virus Infections*. Elsevier/North Holland Biomedical Press, New York, USA. [Cock (1986)]
1792. Goodman, R.M., Bird, J. and Thongmeearkom, P. 1977. An unusual viruslike particle associated with golden yellow mosaic of beans. *Phytopathology* 67(1): 37-42. [ Cock (1986)]
1793. Goodman, R.M., Shock, T.L., Haber, S., Browning, K.S. and Bowers, G.R., Jr. 1980. The composition of bean golden mosaic virus and its single-stranded DNA genome. *Virology* 106: 168-172.
1794. Goolsby, J., Legaspi, J.C. and Legaspi, B.C. 1996. Quarantine evaluation of exotic parasitoids of the sweetpotato whitefly, *Bemisia tabaci* (Gennadius). *Southwest. Entomol.* 21(1): 13-21.
1795. Goolsby, J.A. and Ciomperlik, M.A. 1999. Development of parasitoid inoculated seedling transplants for augmentative biological control of silverleaf whitefly (Homoptera : Aleyrodidae). *Florida Entomol.* 82(4): 532-545.
1796. Goolsby, J.A., Ciomperlik, M.A., Kirk, A.A., Jones, W.A., Legaspi, B.C., Jr., Legaspi, J.C., Ruiz, R.A., Vacek, D.C. and Wendel, L.E. 2000. Predictive and empirical evaluation for parasitoids of *Bemisia tabaci* (biotype 'B'), based on morphological and molecular systematics. pp. 347-358. In A.D. Austin and M. Dowton (Eds.), *Hymenoptera: Evolution, Biodiversity and Biological Control*. CSIRO Publishing, Collingwood, Victoria, Australia.
1797. Goolsby, J.A., Ciomperlik, M.A., Legaspi, B.C., Legaspi, J.C. and Wendel, L.E. 1998. Laboratory and field evaluation of exotic parasitoids of *Bemisia tabaci* (Gennadius) (Biotype "B") (Homoptera: Aleyrodidae) in the lower Rio Grande Valley of Texas. *Biol. Control* 12(2): 127-135.
1798. Gopal, M., Mukherjee, I. and Srivastava, K.P. 1997. Efficacy of imidacloprid and its comparison with other insecticides for controlling whitefly in pulses. *Ann. Plant Prot. Sci.* 5(1): 29-33.
1799. Gorbalenya, A.E. and Koonin, E.V. 1989. Viral proteins containing the purine NTP-binding sequence pattern. *Nucleic Acids Res.* 17(21): 8413-8441.
1800. Gordon M. R., Franco, J., Gonzalez, A. and Gutierrez, O. 2000. Manejo quimico de las principales plagas del cultivo de melon bajo los sistemas de siembra, Los Santos, Panama, 1999. *Ciencia Agropecuaria (Panama)* 10: 101-112. [ Spanish, English summary]
1801. Gould, J.R. and Naranjo, S.E. 1999. Distribution and sampling of *Bemisia argentifolii* (Homoptera: Aleyrodidae) and *Eretmocerus eremicus* (Hymenoptera: Aphelinidae) on cantaloupe vines. *J. Econ. Entomol.* 92(2): 402-408.
1802. Gour, T.B. 1987. Factors influencing whitefly outbreaks--a review. pp. 95-102. In S. Jayaraj (Ed.), *Resurgence of Sucking Pests: Proceedings of National Symposium*. Centre for Plant Protection Studies, Tamil Nadu Agric. Univ., Coimbatore, India.
1803. Goze, E. 1990. Research on the causes of sticky cotton in farming systems in tropical Africa. pp. 19-24. In *Cotton Production Research from a Farming Systems Perspective, with Special Emphasis on Stickiness: 49th Plenary Meeting*. International Cotton Advisory Committee
1804. Grageda-Grageda, J., Loaiza-Villegas, J.M.I. and Cepeda-Vazquez, G. 1993. Evaluacion de practicas de cultivo para el combate de la mosquita blanca en calabaza italiana, en la region de Caborca, Son. [Evaluating practices to combat whitefly on zucchini: Caborca region, Sonora]. pp. 127-128. In J.M. Ramirez-Diaz (Ed.), *Research Advances Ciano. Fall-Winter 1989-1990*. Centro De Investigaciones Agricolas Del Noroeste, Obregon, Mexico. [Spanish, English summary]
1805. Graham, M.R.W. 1976. The British species of *Aphelinus* with notes and description of other European Aphelinidae (Hymenoptera). *Syst. Entomol.* 1: 123-146. [ Cock (1993)]
1806. Granges, A. and Leger, A. 1995. Sept annees d'experimentations de lutte biologique-integree en serres de tomates et de concombres au Centre des Fougères a Conthey. [Seven years of experiments on biological-integrated control on tomatoes and cucumbers under glass at the Fougères Centre at Conthey]. *Rev. Suisse de Viticulture, d'Arboriculture et d'Horticulture* 27(3): 189-191. [ French]
1807. Granillo, C., Diaz, A., Anaya, M. and Bermudez de Paz, L.A. 1975. Diseases transmitted by *Bemisia tabaci* in El Salvador. pp. 51-53. In J. Bird and K. Maramorosch (Eds.), *Tropical Diseases of Legumes*.
1808. Granillo, C.R. 1973. La mosca blanca *Bemisia tabaci* (Genn.) como vector del virus del algodonero (*Gossypium hirsutum*) en El Salvador. *Soc. Ingenieros Agron. (El Salvador)* 2: 31-34.
1809. Granillo, C.R., Diaz, A.D., Anaya, M.A. and Bermudez de Paz, L.A. 1975. Enfermedades transmitidas por *Bemisia tabaci* en el Salvados. *Soc. Ingenieros Agron. (El Salvador)* 4: 6-7.
1810. Gravena, S. 1984. [Integrated management of tomato pests]. pp. 129-149. In *Congresso Brasileiro Olericultura I Reuniao Latino-Americana Olericultura*. Brasilia, Brasil. [Cock (1993), Portuguese]
1811. Gravena, S., Churata-Masca, M.G.C., Arai, J. and Raga, A. 1984. Manejo integrado da mosca branca *Bemisia tabaci* (Gennadius, 1889) em cultivares de tomateiro de crescimento determinado visando reducao de virose do mosaico dourado. *Anais Soc. Entomol. Brasil* 13(1): 35-45. [ Cock (1986)]
1812. Gravena, S. and Nakano, O. 1975. Ensaio experimental com alguns inseticidas contra a "mosca branca" *Bemisia tabaci* Genn. (Homoptera: Aleyrodidae) em feijoeiro das secas no norte do Parana. *Cientifica* 3(1): 187-193. [ Cock (1986)]

- 1813.Gray, A., North, N.C. and Wright, A.N. 1985. The application of high performance liquid chromatography to the study of cotton stickiness. *Coton Fibres Trop.* 40(2): 105-111.
- 1814.Greathead, A.H. 1986. Host plants. pp. 17-25. In M.J.W. Cock (Ed.), *Bemisia tabaci* - A Literature Survey on the Cotton Whitefly with an Annotated Bibliography. CAB International Institute of Biological Control, Ascot, UK.
- 1815.Greathead, D.J. and Bennett, F.D. 1981. Possibilities for the use of biotic agents in the control of the white fly, *Bemisia tabaci*. *Biocontrol News Information* 2: 1-7.
- 1816.Green, S.K. and Kalloo, G. 1994. Leaf curl and yellowing viruses of pepper and tomato: an overview. *Tech. Bull., Asian Veg. Res. Dev. Center (Taiwan, ROC)*, 51 pp.
- 1817.Greenberg, S.M., Jones, W.A. and Legaspi, B.C., Jr. 2000. Interactions between *Encarsia pergandiella* (Hymenoptera: Aphelinidae) and its host *Bemisia argentifolii* (Homoptera: Aleyrodidae): effects of parasitoid densities and host-parasitoid ratios. *Subtrop. Plant Sci.* 52: 36-41.
- 1818.Greenberg, S.M., Jones, W.A. and Warfield, W.C. 1999. Comparative influence of cotton on whitefly species and parasitoids. pp. 911-914. In P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
- 1819.Greenberg, S.M., Legaspi, B.C. and Jones, W.A. 2001. Comparison of functional response and mutual interference between two aphelinid parasitoids of *Bemisia argentifolii* (Homoptera: Aleyrodidae). *J. Entomol. Sci.* 36(1): 1-8.
- 1820.Greenberg, S.M., Legaspi, B.C., Jones, W.A. and Enkegaard, A. 2000. Temperature-dependent life history of *Eretmocerus eremicus* (Hymenoptera: Aphelinidae) on two whitefly hosts (Homoptera: Aleyrodidae). *Environ. Entomol.* 29(4): 851-860.
- 1821.Greenberg, S.M., Liu, T.X., Sappington, T. and Elzen, G.W. 2000. Preliminary data on the effects of cotton defoliant chemicals on *Bemisia argentifolii* mortality and its parasitoid survival. pp. 1147-1151. In P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
- 1822.Grewal, J.S. 1978. Diseases of mungbean in India. pp. 165-168. In R. Cowell (Ed.), *The 1st International Mungbean Symposium*. Office of Information Services, Asian Vegetable Research and Development Center, Taiwan. [Cock (1986)]
- 1823.Grier, N. 1991. Lessons in sustainable agriculture from California's poinsettia whitefly. *J. Pestic. Reform* 1(4): 28.
- 1824.Griffiths, P.D. and Scott, J.W. 2001. Inheritance and linkage of tomato mottle virus resistance genes derived from *Lycopersicon chilense* accession LA 1932. *J. Am. Soc. Hortic. Sci.* 126(4): 462-467.
- 1825.Gronenborn, B., Bendahmane, M., David, D., Desbiez, C., Heyraud, F., Jupin, I., Kheyr-Pour A., Laufs, J., Schumacher, S. and Wartig, L. 1995. Geminiviruses: genome organization and protein functions. *Agronomie (Paris)* 15(7-8): 496-497.
- 1826.Groning, B.R., Abouzid, A. and Jeske, H. 1987. Single-stranded DNA from abutilon mosaic virus is present in the plastids of infected *Abutilon sellovianum*. *Proc. Natl. Acad. Sci.* 84(24): 8996-9000.
- 1827.Gruenhagen, N.M. 1996. Host plant effects on parasitism of silverleaf whitefly by *Eretmocerus* sp. nr. *californicus* [University of California, Riverside]. *Dissertation Abst. Int.* 57(11B): 6725.
- 1828.Gruenhagen, N.M. and Perring, T.M. 1999. Velvetleaf: A plant with adverse impacts on insect natural enemies. *Environ. Entomol.* 28(5): 884-889.
- 1829.Gruenhagen, N.M. and Perring, T.M. 2001. Plant influences on silverleaf whitefly oviposition and development and the potential for enemy-free space. *Entomol. Exp. Appl.* 99(3): 387-391.
- 1830.Gruenhagen, N.M., Perring, T.M., Bezark, L.G., Daoud, D.M. and Leigh, T.F. 1993. Silverleaf whitefly present in the San Joaquin Valley. *California Agric.* 47(1): 4-6.
- 1831.Grupo Paulista De Fitopatologia. 1983. Paulist Phytopathological Group. VI Paulist Phytopathology Congress, 24 to 26 January 1983, IAA/Planalsucar - Coordenadoria Regional Sul Araras - SP. *Summa Phytopathol.* 9(1-2): 3-97. [Cock (1986)]
- 1832.Guagliumi, P. 1967. Insetti e aracnidi delle piante comuni del Venezuela segnalati nel periodo 1938-1963. *Relazioni Monografiche Agrarie Subtropicali Tropicali Firenze (Nuova Serie)* 86: 28-29,68,122-123,137-138. [Cock (1986)]
- 1833.Guerra, J.A., Fernandez, O., Gutierrez, O., Murillo, A. and Villarreal, N. 1999. Identificación de especies y biotipos de moscas blancas en algunas áreas hortícolas de la Península de Azuero, Panamá. pp. 55-59. In 2nd Taller Regional sobre Manejo Integrado de Plagas: Combate del Picudo del Chile *Anthonomus Eugeni*. Cerro Punta, Panamá, 5-7 Oct 1999. [Spanish]
- 1834.Guershon, M. and Gerling, D. 1999. Predatory behavior of *Delphastus pusillus* in relation to the phenotypic plasticity of *Bemisia tabaci* nymphs. *Entomol. Exp. Appl.* 92(3): 239-248.
- 1835.Guershon, M. and Gerling, D. 2001. Effect of foliar tomentosity on phenotypic plasticity in *Bemisia tabaci* (Horn., Aleyrodidae). *J. Appl. Entomol.* 125(8): 449-453.
- 1836.Guest, E. 1931. Annual report on cotton (1929). *Memoirs Dep. Agric. (Iraq)* 15: 1-36. [Cock (1986)]
- 1837.Guevara, J.E. 1993. Diagnostico sobre la problematica de *Bemisia tabaci* (Gennadius) en el Valle Central de Costa Rica. [Diagnosis of the problem of *Bemisia tabaci* (Gennadius) in the Central Valley of Costa Rica]. *Manejo Integrado de Plagas (Costa Rica)* 30: 31-34. [Spanish, English summary]
- 1838.Guimaraes, J.M. 1996. The diagnostic value of the cement gland and other abdominal structures in aleyrodid taxonomy. *Bull. OEPP* 26(2): 413-419.
- 1839.Guimaraes, J.M., Louro, D. and Pereira, V.A. 1996. A mosca branca *Bemisia tabaci* e o virus do frisado amarelo do tomateiro. [The *Bemisia tabaci* white fly and the tomato yellow leaf curl virus]. IPPAA/CNPPA, Lisboa, Portugal, 7 pp. [Portuguese]
- 1840.Guirao, P., Beitia, F. and Cenis, J.L. 1994. Application of RAPD-PCR technique in whitefly taxonomy (Homoptera: Aleyrodidae). *Boletín de Sanidad Vegetal Plagas* 20 (3): 757-764.
- 1841.Guirao, P., Beitia, F. and Cenis, J.L. 1996. Biotipos de *Bemisia tabaci* presentes en España. pp. 47-54. In J.L. Cenis (Ed.), *El Virus del Rizado Amirillo (Hoja en Cuchara) del Tomate (TYLCV) y Su Vector Bemisia tabaci*. Jornadas 8. Consejería de Medio Ambiente, Agricultura y Agua, Murcia, Spain.
- 1842.Guirao, P., Beitia, F. and Cenis, J.L. 1997. Biotype determination of Spanish populations of *Bemisia tabaci* (Hemiptera: Aleyrodidae). *Bull. Entomol. Res.* 87(6): 587-593.
- 1843.Guirao, P., Cenis, J.L. and Beitia, F. 1996. Determinación de la presencia en España de biotipos de *Bemisia tabaci* (Gennadius). [Determining *Bemisia tabaci* (Gennadius) biotypes presence in Spain]. *Phytoma (España)* 81: 30-34. [Spanish]
- 1844.Guirao, P., Onillon, J.C., Beitia, F. and Cenis, J.L. 1997. Presence en France du biotype "B" de *Bemisia tabaci*. [The presence of the "B" biotype of *Bemisia tabaci* in France]. *Phytoma (France)* 498: 44, 47-48. [French, English summary]
- 1845.Guirguis, M.W., Khalil, F.A. and Watson, W.M. 1975. The effectiveness of certain insecticides against the jassid, *Empoasca lybica* (De Berg), and the whitefly *Bemisia tabaci* (Gennadius) attacking cotton. *Zagazig J. Agric. Res. (Egypt)* 2: 233-237.
- 1846.Gunning, R.V., Byrne, F.J., Conde, B.D., Connelly, M.L., Hergstrom, K. and Devonshire, A.L. 1995. First report of B-biotype *Bemisia tabaci* (Gennadius) (Hemiptera: Aleyrodidae) in Australia. *J. Aust. Entomol. Soc.* 34(2): 116.
- 1847.Gunning, R.V., Byrne, F.J. and Devonshire, A.L. 1997. Electrophoretic analysis of non-B and B-biotype *Bemisia tabaci* (Gennadius) (Hemiptera: Aleyrodidae) in Australia. *Aust. J. Entomol.* 36(3): 245-249.

- 1848.Gunning, R.V., Conde, B.D. and Byrne, F.J. 1995. Resistant B-biotype of *Bemisia tabaci* detected in Australia. *Resist. Pest Manage.* 7(1): 13.
- 1849.Gupta, G.P. and Katiyar, K.N. 1988. Effect of insecticidal application against bollworms and their response to whitefly in cotton. *Pesticides* 22(4): 33-35. [Cock (1993)]
- 1850.Gupta, G.P. and Katiyar, K.N. 1991. Bioefficacy of tank-mix insecticides for control of bollworm complex (*Earias* spp. and *Pectinophora gossypiella*) and impact on whitefly (*Bemisia tabaci*) in upland cotton (*Gossypium hirsutum*). *Indian J. Agric. Sci.* 61(7): 531-534.
- 1851.Gupta, G.P., Katiyar, K.N. and Sharma, K. 1999. Neem in the management strategies of insect pest of cotton. pp. 177-189. *In* Singh R. P. and R.C. Saxena (Eds.), *Azadirachta indica*. Science Publishers Inc., Enfield, New Hampshire.
- 1852.Gupta, G.P., Lal, R. and Roshan, L. 1998. Utilization of newer insecticides and neem in cotton pest management system. *Ann. Plant Prot. Sci.* 6(2): 155-160.
- 1853.Gupta, G.P., Mahapatro, G.K., Kundu, S.K. and Lal, R. 1998. Impact of abiotic factors on population dynamics of whitefly in cotton. *Indian J. Entomol.* 60(3): 293-296.
- 1854.Gupta, G.P. and Sharma, K. 1996. Utilization of biopesticides in managing the cotton pest complex in India. pp. 1135-1137. *In* P. Dugger and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
- 1855.Gupta, G.P. and Sharma, K. 1997. Neem based pest management strategy in cotton system. *Pestic. Res. J.* 9(2): 190-197.
- 1856.Gupta, P.C. 1972. External morphology of *Bemisia gossypiperda* (M. & L.) a vector of plant virus diseases (Homoptera: Aleurodidae). *Zool. Beitrage* 18: 1-23. [Cock (1986)]
- 1857.Gupta, P.C. and Chaudhry, H.S. 1972. New record of *Hemitarsonemus latus* Banks (Tarsonemidae) as a parasite of *Bemisia gossypiperda* M. & L. *Indian J. Entomol.* 33(1971): 476. [Cock (1986)]
- 1858.Gupta, P.K. and Singh, J. 1981. Important insect pests of cowpea (*Vigna unguiculata* L.) in agroecosystem of eastern Uttar Pradesh. *Indian J. Zootomy* 22(2): 91-95. [Cock (1986)]
- 1859.Gupta, P.K. and Singh, J. 1983. Effect of systemic granular insecticides on whitefly population and yellow-mosaic infection in greengram. *Indian J. Agric. Sci.* 53: 737-742. [Cock (1986)]
- 1860.Gupta, P.K. and Singh, J. 1993. Population studies on insect pests of greengram (*Vigna radiata* (L.) Wicczek). *Indian J. Entomol.* 55(1): 45-51.
- 1861.Gutierrez-Baeza, A.O. and Garcia-Sandoval, J.A. 1998. Efecto de *Paecilomyces fumosoroseus* contra mosca blanca (*Bemisia tabaci*) en chile Jalapeno. [Effect of *Paecilomyces fumosoroseus* against white fly (*Bemisia tabaci*) in Jalapeno chili]. pp. 193-195. *In* Proceedings 21st National Congress of Biological Control, Rio Bravo, Tampaulipas, Mexico, 5-6 Nov 1998. INIFAP [Spanish]
- 1862.Gutierrez, J.C. 1997. Registration of 'Victoria' cotton. *Crop Sci.* 37(4): 1389.
- 1863.Gutknecht, J. 1989. Assessing stickiness with thermodetection. pp. 123-125. *In* 56th Cotton Int., Willoughby, OH. Meister Publishing Co., Ohio.
- 1864.Haber, S., Ikegami, M., Bajet, N.B. and Goodman, R.M. 1981. Evidence for a divided genome in bean golden mosaic virus, a geminivirus. *Nature (London)* 289: 324-326. [Cock (1986)]
- 1865.Habib, A. and Farag, F.A. 1971. Studies on nine common aleurodids of Egypt. *Bull. Entomol. Soc. Egypt* 54: 1-41. [Cock (1986)]
- 1866.Habib, R. and Mohyuddin, A.I. 1981. Possibilities of biocontrol of some pests of cotton in Pakistan. *Biologia* 27(1): 107-113. [Cock (1986)]
- 1867.Habibi, J. 1975. The cotton whitefly *Bemisia tabaci* Genn. bioecology and methods of control. *Entomol. Phytopathol. Appl.* 38: 3-4. [Cock (1986)]
- 1868.Habu, N. 1991. [Effect of several insecticides on the sweetpotato whitefly, *Bemisia tabaci* (Gennadius)]. *Proc. Kanto-Tosan Plant Prot. Soc.* 38: 235-236. [Cock (1993), Japanese, English summary]
- 1869.Habu, N., Arai, S. and Iga, M. 1990. Occurrence of the sweetpotato whitefly, *Bemisia tabaci* (Genn.) in greenhouses in Tokyo (Honsu, Japan). *Proc. Kanto-tosan Plant Prot. Soc.* 37: 207-208. [Japanese]
- 1870.Hafeez, R., Khan, W.S., Khan, M.D. and Shah, M.K.N. 2001. Stability of cotton cultivars under leaf curl virus epidemic in Pakistan. *Field Crops Res.* 69(3): 251-257.
- 1871.Hafez, M., Tawfik, M.F.S., Awadallah, K.T. and Sarhan, A.A. 1983. Impact of the parasite, *Eretmocerus mundus* Mercet on population of the cotton whitefly, *Bemisia tabaci* (Genn.), in Egypt. *Bull. Entomol. Soc. Egypt* 62(1978-1979): 23-32. [Cock (1986)]
- 1872.Hafez, M., Tawfik, M.F.S., Awadallah, K.T. and Sarhan, A.A. 1983. Natural enemies of the cotton whitefly, *Bemisia tabaci* (Genn.), in the world and in Egypt. *Bull. Entomol. Soc. Egypt* 62(1978-1979): 9-13. [Cock (1986)]
- 1873.Hafez, M., Tawfik, M.F.S., Awadallah, K.T. and Sarhan, A.A. 1993. Studies on *Eretmocerus mundus* Mercet, a parasite of the cotton whitefly, *Bemisia tabaci* (Genn.) in Egypt. *Bull. Entomol. Soc. Egypt* 71: 15-22. [Cock (1986)]
- 1874.Hagler, J., Jackson, G. and Ciomperlik, M. 1998. A novel technique for labeling parasitoids of cotton pest. pp. 1310-1311. *In* P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
- 1875.Hagler, J.R., Brower, A.G., Tu, Z., Byrne, D.N., Bradley-Dunlop, D. and Enriquez, F.J. 1993. Development of a monoclonal antibody to detect predation of the sweetpotato whitefly, *Bemisia tabaci*. *Entomol. Exp. Appl.* 68(3): 231-236.
- 1876.Hagler, J.R. and Durand, C.M. 1994. A new method for immunologically marking prey and its use in predation studies. [French summary]. *Entomophaga* 29(3/4): 257-265.
- 1877.Hagler, J.R. and Naranjo, S.E. 1994. Determining the frequency of heteropteran predation on sweetpotato whitefly and pink bollworm using multiple ELISAs. *Entomol. Exp. Appl.* 72(1): 59-66.
- 1878.Hagler, J.R. and Naranjo, S.E. 1994. Qualitative survey of two coleopteran predators of *Bemisia tabaci* (Homoptera: Aleyrodidae) and *Pectinophora gossypiella* (Lepidoptera: Gelechiidae) using a multiple prey gut content ELISA. *Environ. Entomol.* 23(1): 193-197.
- 1879.Hagler, J.R. and Naranjo, S.E. 1996. Using gut content immunoassays to evaluate predaceous biological control agents: a case study. pp. 383-399. *In* W. Symondson and J. Liddell (Eds.), *The Ecology of Agricultural Pests: Biochemical Approaches*. Chapman Hall, London.
- 1880.Hagler, J.R. and Naranjo, S.E. 1997. A new approach to evaluate augmentative biological control agents. p. 1320. *In* D. Richter and J. Armour (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, Tenn.
- 1881.Hagler, J.R., Naranjo, S.E., Machtley, S., Durand, C., Figuli, P.J. and Henneberry, T.J. 1993. Identifying key predators of sweetpotato whitefly and pink bollworm using pest-specific monoclonal antibodies. pp. 283-285. *In* D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
- 1882.Hahn, S.K., Terry, E.R. and Leschner, K. 1980. Breeding cassava for resistance to cassava mosaic disease. *Euphytica* 29(3): 673-683.
- 1883.Haji, F.N.P., De Alencar, J.A. and Prezotti, L. 1998. Principal pests of tomato and alternatives for control [Spanish, English summary]. pp. 1-4, 20-31, 47-50. *In* Empresa Brasileira de Pesquisa Agropecuaria, Centro de Pesquisa Agropecuaria do Tropic Semi-Arido (CPATSA), Petrolina, Brazil.
- 1884.Haji, F.N.P., De Azevedo-Mattos, M.A., De Alencar, J.A., Barbosa, F.R. and Moreira, A.N. 2000. Biological aspects, damage and control strategies for white flies. *Circular Tecnica Da Embrapa Semi Arido* No.55, 38 pp. [Portuguese]



- 1885.Hajimorad, M.R., Kheyr-Pour, A., Alavi, V., Ahoonmanesh, A., Bahar, M., Rezaian, M.A. and Gronenborn, B. 1996. Identification of whitefly transmitted tomato yellow leaf curl geminivirus from Iran and a survey of its distribution with molecular probes. *Plant Pathol.* 45(3): 418-425.
- 1886.Haley, A., Zhan, Xiangcan, Richardson, K., Head, K. and Morris, B. 1992. Regulation of the activities of african cassava mosaic virus promoters by the AC1, AC2, and AC3 gene products. *Virology* 188: 905-909.
- 1887.Halimie, M.A., Ahmad, C.S., Khan, Q.A. and Khan, M.S. 1993. Control of cotton pest complex with some new organophosphates. *J. Agric. Res. (Pakistan)* 31(2): 215-219.
- 1888.Hallan, V., Saxena, S. and Singh, B.P. 1998. *Ageratum*, *Croton* and *Malvastrum* harbour geminiviruses: evidence through PCR amplification. *World J. Microbiol. Biotech.* 14(6): 931-932.
- 1889.Hamamura, T. 1999. Susceptibility of silver-leaf whitefly, *Bemisia argentifolii* to various spray-type insecticides. [Japanese, English summary]. *Bull. Nat. Res. Inst. Vegetables, Ornamental Plants and Tea* 14: 177-187.
- 1890.Hameed, S., Khalid, S., Ehsan-ul-Haq, E. and Hashmi, A.A. 1994. Cotton leaf curl disease in Pakistan caused by a whitefly-transmitted geminivirus. *Plant Dis.* 78(5): 529.
- 1891.Hameed, S., Khalid, S. and Naqvi, S.M.S. 1996. Occurrence of B-biotype of *Bemisia tabaci* in Pakistan. pp. 81-85. *In Brighton Crop Protection Conference: Pests and Diseases.* British Crop Protection Council, Farnham, UK.
- 1892.Hamid, A.M. and Korkor, A.A. 1998. New approaches for control of whitefly, *Bemisia tabaci* on cotton crop. *Alexandria Sci. Exchange (Egypt)* 19(4): 543-558.
- 1893.Hamilton, W.D.O., Bisaro, D.M. and Buck, K.W. 1982. Identification of novel DNA forms in tomato golden mosaic virus infected tissue. Evidence for a two component viral genome. *Nucleic Acids Res.* 10(16): 4901-4912. [Cock (1986)]
- 1894.Hamilton, W.D.O., Bisaro, D.M., Coutts, R.H.A. and Buck, K.W. 1983. Demonstration of the bipartite nature of the genome of a single-stranded DNA plant virus by infection with the cloned DNA components. *Nucleic Acids Res.* 11(21): 7387-7396. [Cock (1986)]
- 1895.Hamilton, W.D.O., Stein, V.E., Coutts, R.H.A. and Buck, K.W. 1984. Complete nucleotide sequence of the infectious cloned DNA components of tomato golden mosaic virus: potential coding regions and regulatory sequences. *EMBO J.* 3(9): 2197-2205.
- 1896.Hammad, S.M. 1978. Pests of grain legumes and their control in Egypt. pp. 135-137. *In S.R. Singh, H.F. van Emden and T.A. Taylor (Eds.), Pests of Grain Legumes: Ecology and Control.* Academic Press, London, UK. [Cock (1986)]
- 1897.Hammond, D.G., Rangel, S. and Kubo, I. 2000. Volatile aldehydes are promising broad-spectrum postharvest insecticides. *J. Agric. Food Chem.* 48(9): 4410-4417.
- 1898.Hamon, A.B. and Salguero, V. 1987. *Bemisia tabaci*, sweet potato whitefly, in Florida (Homoptera: Aleyrodidae: Aleyrodinae). *Entomol. Circular, Div. Plant Ind., Florida. Dep. Agric. Consumer Serv.* 292: 1-2. [Cock (1993)]
- 1899.Han, S.S. and Konieczny, J. 2000. Responses of whitefly and poinsettias to insecticidal controlled atmospheres. *J. Am. Soc. Hortic. Sci.* 125(4): 513-517.
- 1900.Hanafi, A. 2000. The threat of insect-transmitted viruses to vegetable production in Morocco. *Bull. OILB/SROP (IOBC/WPRS)* 23(1): 89-94.
- 1901.Handa, A. and Gupta, M.D. 1993. Management of bhindi yellow vein mosaic virus disease. *Indian Phytopathol.* 46(2): 123-130.
- 1902.Hanif-Khan, S., Brecht, J.K., Powell, C.A. and Stoffella, P.J. 2000. Ethylene levels and fruit quality of silverleaf whitefly-infested dwarf cherry tomato. *Proc. Florida State Hortic. Soc.* 112: 134-138.
- 1903.Hanif-Khan, S., Bullock, R.C., Stoffella, P.J., Powell, C.A., Brecht, J.K., McAuslane, H.J. and Yokomi, R.K. 1996. Tomato irregular ripening symptom development induced by silverleaf whitefly dwarf cherry tomatoes. *Proc. Florida State Hortic. Soc.* 108: 193-196.
- 1904.Hanif-Khan, S., Bullock, R.C., Stoffella, P.J., Powell, C.A., Brecht, J.K., McAuslane, H.J. and Yokomi, R.K. 1997. Possible involvement of altered gibberellin metabolism in the induction of tomato irregular ripening in dwarf cherry tomato by silverleaf whitefly. *J. Plant Grow. Reg.* 16(4): 245-251.
- 1905.Hanif-Khan, S., Bullock, R.C., Stoffella, P.J., Powell, C.A., Brecht, J.K., McAuslane, H.J. and Yokomi, R.K. 1998. Tomato irregular-ripening symptom development and ripening of silverleaf whitefly-infested dwarf cherry tomatoes. *J. Am. Soc. Hortic. Sci.* 123(1): 119-125.
- 1906.Hanley-Bowdoin, L., Elmer, J.S. and Rogers, S.G. 1988. Transient expression of heterologous RNAs using tomato golden mosaic virus. *Nucleic Acids Res.* 16(22): 10511-10528.
- 1907.Hanley-Bowdoin, L., Elmer, J.S. and Rogers, S.G. 1990. Expression of functional replication protein from tomato golden mosaic virus in transgenic tobacco plants. *Proc. Natl. Acad. Sci.* 87(4): 1446-1450.
- 1908.Hansford C. G. 1944. A probable virus disease of sweet potato. *East African Agric. Forestry J.* 10: 126-127. [Cock (1986)]
- 1909.Hanson, P.M., Bernacchi, D., Green, S., Tanksley, S.D., Muniyappa, V., Padmaja, S., Chen, H.M., Kuo, G., Fang, D. and Chen, J.T. 2000. Mapping a wild tomato introgression associated with tomato yellow leaf curl virus resistance in a cultivated tomato line. *J. Am. Soc. Hortic. Sci.* 125(1): 15-20.
- 1910.Harakly, F.A. 1974. Preliminary survey of pests infesting solanaceous truck crops in Egypt. *Bull. Entomol. Soc. Egypt* 58: 133-140. [Cock (1986)]
- 1911.Harakly, F.A. 1974. Variation in pupae of *Bemisia tabaci* (Gennadius) bred on different host (Homoptera: Aleyrodidae). *Bull. Entomol. Soc. Egypt.* 57(1973): 407-412. [Cock (1986)]
- 1912.Harakly, F.A. and Assem, M.A.H. 1978. Ecological studies on the truly pests of leguminous plants in Egypt. II. Piercing and sucking pests. pp. 237-242. *In Proceedings Fourth Conference of Pest Control, September 30 - October 3, 1978.* Academy of Scientific Research and Technology and National Research Center, Cairo, Egypt. [Cock (1986)]
- 1913.Hardee, D.D. 1993. Resistance in aphids and whiteflies: principles and keys to management. pp. 20-23. *In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences.* National Cotton Council, Memphis, TN.
- 1914.Hardee, D.D. and Herzog, G.A. 1992. 45th Annual conference report on cotton insect research and control. pp. 626-644. *In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences.* National Cotton Council, Memphis, TN.
- 1915.Hardee, D.D. and Herzog, G.A. 1993. 46th annual conference report on cotton insect research and control. pp. 635-660. *In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences.* National Cotton Council, Memphis, TN.
- 1916.Harpaz, I. 1982. Nonpesticidal control of vector-borne viruses. pp. 1-19. *In K.F. Harris and K. Maramorosch (Eds.), Pathogens, Vectors, and Plant Diseases: Approaches to Control.* Academic Press, New York.
- 1917.Harpaz, I. and Cohen, S. 1965. Semipersistent relationship between cucumber vein yellowing virus (CVYV) and its vector, the tobacco whitefly *Bemisia tabaci* (Gennadius). *Phytopathol. Z.* 54: 240-248. [Cock (1986)]
- 1918.Harris, K.F., Esbroeck, Z.P. and Duffus, J.E. 1996. Anatomy of a virus vector. pp. 289-318. *In D. Gerling and R.T. Mayer (Eds.), Bemisia : 1995 Taxonomy, Biology, Damage, Control and Management.* Intercept Ltd., Andover, Hants, UK.
- 1919.Harris, K.F. and Maramorosch, K. (Eds.). 1982. *Pathogens, vectors, and plant diseases: approaches to control.* Academic Press, New York, USA, 310 pp. [Cock (1986)]

- 1920.Harris, K.F., Pesic-Van Esbroeck, Z. and Duffus, J.E. 1996. Morphology of the sweet potato whitefly, *Bemisia tabaci* (Homoptera, Aleyrodidae), relative to virus transmission. *Zoomorphology* 116(3): 143-156.
- 1921.Harris, W.V. 1934. Report of the Acting Entomologist, 1933. Rep. Dept. Agric. (Tanganyika) 1933: 69-75. [ Cock (1986)]
- 1922.Harrison, B.D. 1985. Advances in geminivirus research. *Annu. Rev. Phytopathol.* 23: 55-82.
- 1923.Harrison, B.D. 1994. Detection, identification and assessment of variation of whitefly-transmitted geminiviruses. *Arab J. Plant Prot.* 12(2): 115-120.
- 1924.Harrison, B.D., Barker, H., Bock, K.R., Guthrie, E.J., Meredith G. and Atkinson, M. 1977. Plant viruses with circular single-stranded DNA. *Nature (London)* 270: 760-762. [ Cock (1986)]
- 1925.Harrison, B.D., Liu, Y.L., Khalid, S., Hameed, S., Otim-Nape, G.W. and Robinson, D.J. 1997. Detection and relationships of cotton leaf curl virus and allied whitefly-transmitted geminiviruses occurring in Pakistan. *Ann. Appl. Biol.* 130(1): 61-75.
- 1926.Harrison, B.D., Muniyappa, V., Swanson, M.M., Roberts, I.M. and Robinson, D.J. 1991. Recognition and differentiation of seven whitefly-transmitted geminiviruses from India, and their relationships to African cassava mosaic and Thailand mung bean yellow mosaic viruses. *Ann. Appl. Biol.* 118(2): 299-308. [ Cock (1993)]
- 1927.Harrison, B.D. and Robinson, D.J. 1988. Molecular variation in vector-borne plant viruses; epidemiological significance. *Phil. Trans. R. Soc. (Lond. B)* 321: 447-462.
- 1928.Harrison, B.D. and Robinson, D.J. 1999. Natural genomic and antigenic variation in whitefly-transmitted geminiviruses (Begomoviruses). *Annu. Rev. Phytopathol.* 37: 369-398.
- 1929.Harrison, B.D., Swanson, M.M. and Robinson, D.J. 1995. Cassava viruses in the old world, pp. 463-472. *In The Cassava Biotechnology Network: Proceedings 2nd International Scientific Meeting, Bogor, Indonesia, 22-26 August 1994. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia.*
- 1930.Harrison, B.D., Zhou, X., Otim-Nape, G.W., Liu, Y. and Robinson, D.J. 1997. Role of a novel type of double infection in the geminivirus-induced epidemic of severe cassava mosaic in Uganda. *Ann. Appl. Biol.* 131(3): 437-448.
- 1931.Hashmi, A.A., Ehsan-ul-Haq, E., Rana, M.A., Masih, R., Khalid, S., Hameed, S. and Aftab, M. 1993. A research compendium on cotton leaf curl viral disease and its vector - whitefly. *Pakistan Agric. Res. Council*, 62 pp.
- 1932.Hashmi, A.A. and Nizam-Ud-Din, M. 1978. Lethal exposure time as substitute for economic threshold. *Pakistan J. Zool.* 10: 229-234. [ Cock (1986)]
- 1933.Hassan, A.A. and Abdel-Ati, K.E.A. 1999. Genetics of tomato yellow leaf curl virus tolerance derived from *Lycopersicon pimpinellifolium* and *Lycopersicon pennellii*. *Egyptian J. Hortic.* 26(3): 323-338.
- 1934.Hassan, A.A. and Duffus, J.E. 1991. A review of a yellowing and stunting disorder of cucurbits in the United Arab Emirates. *Emirates J. Agric. Sci.* 2: 1-16. [ Ioannou (1995)]
- 1935.Hassan, A.A., Quronfilah, N.E., Obaji, U.A., Al-Rayis, M.A. and Wafi, M.S. 1991. Evaluation of domestic and wild citrullus germplasm for resistance to the yellow stunting disorder in the United Arab Emirates. *Egyptian J. Hortic.* 18(1): 11-21.
- 1936.Hassan, A.A. and Sayed, S.F. 1999. Chlorotic pod: A new physiological disorder of green-podded snap beans (*Phaseolus vulgaris* L.) associated with silverleaf whitefly infestation. *Egyptian J. Hortic.* 26(2): 213-228.
- 1937.Hassan, A.A., Wafi, M.S., Quronfilah, N.E., Obaji, U.A., Al-Rayis, M.A. and Al-Izabi, F. 1991. Evaluation of wild and domestic lycopersicon accessions for tomato yellow leaf curl virus. *Egyptian J. Hortic.* 18(1): 23-43.
- 1938.Hassan, S., Arif, M. and Defoer, T. 1993. Epidemiological studies of tomato viruses in Malakand Agency of North West Frontier Province of Pakistan. *Sarhad J. Agric. (Pakistan)* 9(1): 37-43.
- 1939.Hassan, S.M., Saad, A.S. and Mansour, M.H. 1975. Effect of certain insecticides on some cotton pests and on cotton plants. *Bull. Entomol. Soc. Egypt Econ. Ser.* 8(1974): 221-226. [ Cock (1986)]
- 1940.Hassan, S.M., Saad, A.S. and Mansour, M.H. 1975. Evaluation of certain insecticides against aphids, jassids, whiteflies and red spider mites attacking cotton. *Bull. Entomol. Soc. Egypt* 8(1974): 41-45. [ Cock (1986)]
- 1941.Hassanein, S.S.M. 1990. Seasonal abundance of certain leaf-insects infesting sunflower plants in Zagazig region, Sharkia Governorate, Egypt. *Zagazig J. Agric. Res. (Egypt)* 17(3b): 925-932.
- 1942.Hassanein, S.S.M. 1993. Effect of rape varieties, weather factors and sowing dates on the seasonal activity of *Bemisia tabaci*, Genn. and *Brevicoryne brassicae* L. *Zagazig J. Agric. Res. (Egypt)* 20(6): 1975-1986.
- 1943.Hassanein, S.S.M., El-Shakaa, S.M.A. and El-Maghraby, M.M.A. 1993. Seasonal occurrence of some leaf-insects attacking forage cowpea and guar plants at Zagazig region. *Mansoura Univ. J. Agric. Sci. (Egypt)* 18(4): 1221-1234.
- 1944.Hassanein, S.S.M., Metwally, E.M. and Hafsa, A.F.E. 1994. Varietal resistance of nine potato varieties to natural infestation by certain leaf pests at Gemmeza region, Egypt. *Egyptian J. Agric. Res.* 72(4): 965-976.
- 1945.Hassanein, S.S.M., Metwally, E.M. and Hafsa, A.F.E. 1995. Daily flight activity of certain sap sucking insects invading some vegetable crops and efficiency of two methods for trapping them at Gemmeza region, Egypt. *Egyptian J. Agric. Res.* 73(3): 639-651.
- 1946.Hassell, M.P. 1985. Insect natural enemies as regulating factors. *J. Anim. Ecol.* 54: 323-334.
- 1947.Hata, T.Y. and Hara, A.H. 1993. Control of whiteflies on *Anthurium*, Hawaii 1992. *Insecticide Acaricide Tests* 18: 303.
- 1948.Hatta, T. and Francki, R.I.B. 1979. The fine structure of chloris striate mosaic virus. *Virology* 92: 428-435.
- 1949.Hayat, M. 1972. The species of *Eretmocerus* Halderman, 1850 [Hymenoptera: Aphelinidae] from India. *Entomophaga* 17: 99-106. [ Cock (1986)]
- 1950.Hayat, M. 1989. Revision of the species of *Encarsia* Foerster (Hymenoptera: Aphelinidae) from India and the adjacent countries. *Oriental Insects* 23: 1-131. [ Cock (1993)]
- 1951.Hayati, J. and Varma, J.P. 1984. Host and environment response to white fly transmission of tomato leaf-curl virus in tomato. *Indian Phytopathol.* 37: 223-227.
- 1952.Haydar, M.F., Afifi, F.M.L. and Aly, F.A. 1990. A simple approach for the management of whitefly-borne virus diseases on tomatoes. [Arabic summary]. *Bull. Faculty Agric. (Univ. Cairo)* 41(3): 649-664. [ Cock (1993)]
- 1953.Haydar, M.F., Ahmed, M.A., El-Deeb, S.E. and Gabr, A.M. 1993. Effect of weed control on the whitefly, *Bemisia tabaci*, Genn. infesting tomato plants. *Moshtohor Ann. Agric. Sci. (Egypt)* 31(4): 2275-2279.
- 1954.Hayes, R.J. and Buck, K.W. 1989. Replication of tomato golden mosaic virus DNA B in transgenic plants expressing open reading frames (ORFs) of DNA A: requirement of ORF AL2 for production of single-stranded DNA. *Nucleic Acids Res.* 17(24): 10213-10222.
- 1955.Hayward, J.A. 1967. Cotton in Western Nigeria. 2. Entomological problems. *Cotton Growing Rev.* 44: 117-135. [ Cock (1986)]
- 1956.Head, J. and Morgan, D. 1996. Computer models in plant health campaign management: their use in *Bemisia tabaci* eradication. pp. 1047-1052. *In Brighton Crop Protection Conference: Pests and Diseases. British Crop Protection Council, Farnham, UK.*
- 1957.Headrick, D.H., Bellows, T.S. and Perring, T.M. 1995. Behaviors of female *Eretmocerus* sp. nr. *californicus* (Hymenoptera: Aphelinidae) attacking *Bemisia argentifolii* (Homoptera: Aleyrodidae) on sweet potato. *Environ. Entomol.* 24(2): 412-422.

1958. Headrick, D.H., Bellows, T.S. and Perring, T.M. 1996. Host-plant effects on the behavior of *Eretmocerus* sp. nr. *californicus* females raised from melon. *Entomophaga* 41(1): 15-26.
1959. Headrick, D.H., Bellows, T.S. and Perring, T.M. 1997. Behavior of female *Eretmocerus* sp. nr. *californicus* Howard (Aphelinidae: Hymenoptera) attacking *Bemisia argentifolii* Bellows and Perring (Aleyrodidae: Homoptera) on two native Californian weeds. *Canadian Entomol.* 129(2): 335-345.
1960. Headrick, D.H., Bellows, T.S. and Perring, T.M. 1999. Development and reproduction of a population of *Eretmocerus eremicus* (Hymenoptera: Aphelinidae) on *Bemisia argentifolii* (Homoptera: Aleyrodidae). *Environ. Entomol.* 28(2): 300-306.
1961. Headrick, D.H., Bellows, T.S., Jr. and Perring, T.M. 1996. Behaviors of female *Eretmocerus* sp. nr. *californicus* (Hymenoptera: Aphelinidae) attacking *Bemisia argentifolii* (Homoptera: Aleyrodidae) on cotton, *Gossypium hirsutum* (Malvaceae) and melon, *Cucumis melo* (Cucurbitaceae). *Biol. Control* 6(1): 64-75.
1962. Hector, D.J. and Hodgkinson, I.D. 1989. Stickiness in cotton. ICAC Review Article on Cotton Production Research, Int. Cotton Advisory Committee 2: 1-43. [Cock (1993)]
1963. Hegab, A.M. and Helaly, M.M. 1989. Occurrence and seasonal abundance of the white fly, *Bemisia tabaci* (Genn.) infesting certain cucurbitaceous and cruciferous vegetable plants in newly reclaimed sandy areas at Sahlia district, Egypt. *Zagazig J. Agric. Res. (Egypt)* 16(1): 130-136. [CATIE (1992)]
1964. Hegab, M.F.A.H., Sabek, A.M., Taha, A.M. and Mazyad, H.M. 1992. Evaluation of protective programmes to control the cotton whitefly, *Bemisia tabaci* (Genn.) and reduce the spread of tomato yellow leaf curl virus in tomato nurseries and in the open field. *Al-Azhar J. Agric. Res. (Egypt)* 16: 125-138.
1965. Hegab, M.F.A.H., Tayeb, E.H. and El-Sayed, N. 1992. Population density of the white fly *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae) associated with cucumber/cotton intercropping system in Menufiya Governorate, Egypt. *Mansoura Univ. J. Agric. Sci. (Egypt)* 17(12): 3941-3945.
1966. Heijne, C.G. and Peregrine, D.J. 1984. The effects of ULV spray characteristics on the activity of amitraz against the cotton whitefly, *Bemisia tabaci* (Gennadius). pp. 975-979. *In* British Crop Protection Conference. Pests and Diseases. British Crop Protection Council, Croydon, UK. [Cock (1986)]
1967. Hein, A. 1984. The leaf curl virus in tomatoes and possibilities of its control. A review. *Acta Hort.* 143: 439-450. [CATIE (1992)]
1968. Heinz, K.M. 1996. Predators and parasitoids as biological control agents of *Bemisia* in greenhouses. pp. 435-449. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia: 1995 Taxonomy, Biology, Damage, Control and Management*. Intercept Ltd., Andover, Hants, UK.
1969. Heinz, K.M., Brazzle, J.R., Parrella, M.P. and Pickett, C.H. 1999. Field evaluations of augmentative releases of *Delphastus catalinae* (Horn) (Coleoptera: Coccinellidae) for suppression of *Bemisia argentifolii* Bellows & Perring (Homoptera: Aleyrodidae) infesting cotton. *Biol. Control* 16(3): 241-251.
1970. Heinz, K.M., Brazzle, J.R., Pickett, C.H., Natwick, E.T., Nelson, J.M. and Parrella, M.P. 1994. Predatory beetle may suppress silverleaf whitefly. *California Agric.* 48(2): 35-40.
1971. Heinz, K.M., Lin, E. and Parrella, M.P. 1993. Behavioral response of *Bemisia tabaci* (Genn.) to olfactory cues emitted by poinsettia. *Bull. OILB/SROP (IOBC/WPRS)* 16: 59-61.
1972. Heinz, K.M. and Nelson, J.M. 1996. Interspecific interactions among natural enemies of *Bemisia* in an inundative biological control program. *Biol. Control* 6(3): 384-393.
1973. Heinz, K.M. and Parrella, M.P. 1994. Biological control of *Bemisia argentifolii* (Homoptera: Aleyrodidae) infesting *Euphorbia pulcherrima* - evaluations of releases of *Encarsia luteola* (Hymenoptera: Aphelinidae) and *Delphastus pusillus* (Coleoptera: Coccinellidae). *Environ. Entomol.* 23(5): 1346-1353.
1974. Heinz, K.M. and Parrella, M.P. 1994. Poinsettia (*Euphorbia pulcherrima* Willd ex Koltz) cultivar-mediated differences in performance of five natural enemies of *Bemisia argentifolii* Bellows and Perring, n sp (Homoptera: Aleyrodidae). *Biol. Control* 4(4): 305-318.
1975. Heinz, K.M. and Parrella, M.P. 1998. Host location and utilization by selected parasitoids of *Bemisia argentifolii* (Homoptera: Aleyrodidae): Implications for augmentative biological control. *Environ. Entomol.* 27(3): 773-784.
1976. Heinz, K.M., Parrella, M.P. and Newman, J.P. 1992. Time-efficient use of yellow sticky traps in monitoring insect populations. *J. Econ. Entomol.* 85: 2263-2269.
1977. Heinz, K.M. and Thompson, S.T. 1999. Development of biological control methods for use in Southwestern US greenhouses and nurseries. *Bull. OILB/SROP (IOBC/WPRS)* 22: 101-104.
1978. Heinz, K.M. and Zalom, F.G. 1995. Variation in trichome-based resistance to *Bemisia argentifolii* (Homoptera: Aleyrodidae) oviposition on tomato. *J. Econ. Entomol.* 88(5): 1494-1502.
1979. Heinz, K.M. and Zalom, F.G. 1996. Performance of the predator *Delphastus pusillus* on *Bemisia* resistant and susceptible tomato lines. *Entomol. Exp. Appl.* 81(3): 345-352.
1980. Held, D.W., Potter, D.A., Gates, R.S. and Anderson, R.G. 2001. Modified atmosphere treatments as a potential disinfestation technique for arthropod pests in greenhouses. *J. Econ. Entomol.* 94(2): 430-438.
1981. Helman, S., Peterlin, O. and Contreras, M. 1996. Parasitoids of *Bemisia* in Santiago del Estero cotton, Northwest Argentina. pp. 527-529. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia: 1995 Taxonomy, Biology, Damage, Control and Management*. Intercept Ltd., Andover, Hants, UK.
1982. Hemeida, E.A. 1981. Studies on some insects attacking certain solanaceous plants. Ph.D. Dissertation, Cairo University, Egypt, 214 pp.
1983. Hemmati, F. 1990. [Collecting and surveying of insect fauna on grapevine in Khuzestan province]. *Sci. J. Agric.* 13(13): 3-10. [Cock (1993), Arabic, English summary]
1984. Hempel, A. 1923. Hemipteros novos ou pouco conhecidos da familia Aleyrodidae. *Rev. Museu Paulista* 13: 1121-1191. [Cock (1986)]
1985. Hendi, A., Abdel-Fattah, M.I. and El-Sayed, A. 1987. Biological study on the white-fly, *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae). *Bull. Entomol. Soc. Egypt* 65: 101-108. [Cock (1993)]
1986. Hendrix, D.L. 1995. The relationship between whitefly populations, honeydew deposition, and stickiness in cotton lint. pp. 104-105. *In* D.A. Richter and J. Armour (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
1987. Hendrix, D.L. 1999. Sugar composition of cotton aphid and silverleaf whitefly honeydews. pp. 47-51. *In* P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
1988. Hendrix, D.L. and Henneberry, T.J. 2000. Differences in polyol accumulation and honeydew excretion in sweetpotato whitefly and cotton aphid. pp. 1296-1300. *In* P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
1989. Hendrix, D.L., Henneberry, T.J., Chu, C.C. and Coates, W.E. 2001. Enzyme treatment of honeydew-contaminated cotton fiber. *Appl. Eng. Agric.* 17(5): 571-576.
1990. Hendrix, D.L., Perkins, H.H., Jr. and Valaitis, A. 1995. Chemical characterization of cotton insect honeydew and elimination of stickiness of honeydew-contaminated lint. pp. 437-441. *In* G.A. Constable and N.W. Forrester (Eds.), *Challenging the Future: Proceedings of the World Cotton Research Conference-1*. CSIRO, Melbourne, Australia.

- 1991.Hendrix, D.L. and Salvucci, M.E. 1998. Polyol metabolism in homopterans at high temperatures: Accumulation of mannitol in aphids (Aphididae: Homoptera) and sorbitol in whiteflies (Aleyrodidae: Homoptera). *Comp. Biochem. Physiol.* 120(3): 487-494.
- 1992.Hendrix, D.L. and Salvucci, M.E. 2001. Isobemiose: an unusual trisaccharide abundant in the silverleaf whitefly, *Bemisia argentifolii*. *J. Insect Physiol.* 47(4-5): 423-432.
- 1993.Hendrix, D.L., Salvucci, M.E. and Wolfe, G.R. 1998. Effect of high temperature on polyol metabolism in the silverleaf whitefly and cotton aphid. pp. 1077-1080. *In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
- 1994.Hendrix, D.L., Steele, T.L. and Perkins, H.H., Jr. 1996. *Bemisia* honeydew and sticky cotton. pp. 189-199. *In D. Gerling and R.T. Mayer (Eds.), Bemisia : 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.*
- 1995.Hendrix, D.L., Taysom, D., Steele, T., Davidson, E.W. and Segura, B.J. 1994. Enzymes forming honeydew sugars from cotton phloem sap are found in sweetpotato whitefly eggs. pp. 1237-1238. *In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
- 1996.Hendrix, D.L. and Wei, Y. 1992. Detection and elimination of honeydew excreted by the sweetpotato whitefly feeding upon cotton. pp. 671-673. *In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
- 1997.Hendrix, D.L. and Wei, Y.A. 1994. Bemiose: an unusual trisaccharide in *Bemisia* honeydew. *Carbohydr. Res.* 253: 329-334.
- 1998.Hendrix, D.L., Wei, Y.A. and Leggett, J.E. 1992. Homopteran honeydew sugar composition is determined by both the insect and plant species. *Comp. Biochem. Physiol.* 101(1-2): 23-27.
- 1999.Henneberry, T.J. 1993. Sweetpotato whitefly - current status and national research and action plan. pp. 663-666. *In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
- 2000.Henneberry, T.J., Blackledge, B., Steele, T., Hendrix, D.L., Perkins, H.H. and Nichols, R.L. 1997. Preliminary evaluations of an enzyme approach to reduce cotton lint stickiness. pp. 430-436. *In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
- 2001.Henneberry, T.J. and Butler, G.D., Jr. 1992. Whiteflies as a factor in cotton production with specific reference to *Bemisia tabaci* (Gennadius). pp. 674-683. *In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
- 2002.Henneberry, T.J. and Castle, S.J. 2001. *Bemisia*: Pest status, economics, biology and population dynamics. pp. 247-278. *In K.F. Harris, O.P. Smith and J.E. Duffus (Eds.), Virus-Insect-Plant Interactions. Academic Press, San Diego, CA.*
- 2003.Henneberry, T.J., Faust, R.M., Jones, W.A. and Perring, T.M. (Eds.). 2000. Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000, 200 pp.
- 2004.Henneberry, T.J. and Faust, R.M. (Eds.). 1999. Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01, 185 pp.
- 2005.Henneberry, T.J., Forlow Jech, L. and De La Torre, T. 2001. Honeydew production by sweetpotato whitefly adults and nymphs. pp. 292-295. *In Cotton, A College of Agriculture Report, Series P-125. University of Arizona, Tucson.*
- 2006.Henneberry, T.J., Forlow Jech, L. and Hendrix, D.L. 2000. Sweetpotato whiteflies, cotton aphids, and sticky cotton. pp. 1160-1162. *In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
- 2007.Henneberry, T.J., Forlow Jech, L., Hendrix, D.L. and Steele, T. 2000. Effects of aqueous sprays of silverleaf whitefly honeydew sugars on cotton lint stickiness. pp. 285-289. *In Cotton, A College of Agriculture Report, Series P-121. University of Arizona, Tucson.*
- 2008.Henneberry, T.J., Hendrix, D.L. and Perkins, H.H. 1998. Effects of cotton ginning and lint cleaning on sticky cotton. pp. 400-402. *In Cotton, A College of Agriculture Report, Series P-112. University of Arizona, Tucson.*
- 2009.Henneberry, T.J., Hendrix, D.L., Perkins, H.H., Jech, L.F. and Burke, R.A. 1996. *Bemisia argentifolii* (Homoptera: Aleyrodidae) honeydew sugars and relationships to sticky cotton. *Environ. Entomol.* 25(3): 551-558.
- 2010.Henneberry, T.J., Hendrix, D.L., Perkins, H.H., Naranjo, S., Flint, H.M., Akey, D.H., Forlow Jech, L. and Burke, R.A. 1995. Silverleaf whitefly, sticky cotton, and cotton lint yields. pp. 836-838. *In D.A. Richter and J. Armour (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
- 2011.Henneberry, T.J., Hendrix, D.L., Perkins, H.H., Naranjo, S.E., Flint, H.M., Akey, D., Jech, L.F. and Burke, R.A. 1995. *Bemisia argentifolii* (Homoptera: Aleyrodidae) populations and relationships to sticky cotton and cotton yields. *Southwest. Entomol.* 20(3): 255-271.
- 2012.Henneberry, T.J., Jech, L., Hendrix, D.L. and Steele, T. 1999. *Bemisia argentifolii* (Homoptera: Aleyrodidae): Factors affecting adult and nymph honeydew production. *Southwest. Entomol.* 24(3): 207-231.
- 2013.Henneberry, T.J., Jech, L.F. and Hendrix, D.L. 1998. Seasonal distribution of *Bemisia argentifolii* (Homoptera: Aleyrodidae) honeydew sugars on Pima and upland cotton lint and lint stickiness at harvest. *Southwest Entomol.* 23(2): 105-121.
- 2014.Henneberry, T.J., Jech, L.F. and Hendrix, D.L. 1998. Seasonal distribution of *Bemisia* honeydew sugars on Pima and upland cotton lint. pp. 394-399. *In Cotton, A College of Agriculture Report, Series P-112. University of Arizona, Tucson.*
- 2015.Henneberry, T.J., Jech, L.F., Hendrix, D.L., Brushwood, D. and Steele, T. 1999. Open cotton boll exposure to whiteflies and development of sticky cotton. pp. 423-427. *In Cotton, A College of Agriculture Report, Series P-116. University of Arizona, Tucson.*
- 2016.Henneberry, T.J., Jech, L.F., Hendrix, D.L. and Brushwood, D.E. 1998. *Bemisia argentifolii* (Homoptera: Aleyrodidae) population relationships to cotton and lint stickiness in long and short staple cottons. *J. Econ. Entomol.* 91(5): 1196-1207.
- 2017.Henneberry, T.J., Jech, L.F., Hendrix, D.L., Perkins, H.H. and Brushwood, D.E. 1997. Silverleaf whitefly and cotton lint stickiness. pp. 332-338. *In Cotton, A College of Agriculture Report, Series P-108. University of Arizona, Tucson.*
- 2018.Henneberry, T.J., Jech, L.F., Hendrix, D.L., Perkins, H.H. and Brushwood, D.E. 1997. Silverleaf whitefly lint stickiness, and cotton yields in low and high plant populations of untreated and insecticide-treated long and short staple cottons. pp. 899-907. *In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
- 2019.Henneberry, T.J., Jech, L.F., Hendrix, D.L. and Steele, T. 2000. *Bemisia argentifolii* (Homoptera : Aleyrodidae) honeydew and honeydew sugar relationships to sticky cotton. *Southwest. Entomol.* 25(1): 1-14.
- 2020.Henneberry, T.J., Jech, L.F., Hendrix, D.L., Steele, T. and Brushwood, D. 1999. Relationship between days of open cotton boll exposure to whitefly populations and development of sticky cotton. pp. 1189-1193. *In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
- 2021.Henneberry, T.J. and Naranjo, S.E. 1999. *Bemisia* whiteflies, sticky cotton and sticky cotton sampling. pp. 51-56. *In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*

- 2022.Henneberry, T.J., Prabhaker, N., Castle, S.J. and Toscano, N.C. 1999. A brief review of *Bemisia tabaci*, its occurrence and resistance to insecticides in southern California and Arizona. pp. 21-31. *In* Proceedings ICAC-CCRI Regional Consultation, Insecticide Resistance Management in Cotton. Central Cotton Research Institute, Multan, Pakistan.
- 2023.Henneberry, T.J. and Toscano, N.C. 1997. Current status of silverleaf and sweetpotato whiteflies in the United States. pp. 95-101. *In* N. Ioannou (Ed.), Management of the whitefly-virus complex. FAO Plant Production and Protection Paper 143, Rome, Italy.
- 2024.Henneberry, T.J. and Toscano, N.C. 1997. Whitefly sampling methods and action thresholds. pp. 115-123. *In* N. Ioannou (Ed.), Management of the whitefly-virus complex. FAO Plant Production and Protection Paper 143, Rome, Italy.
- 2025.Henneberry, T.J., Toscano, N.C. and Castle, S.J. 1998. *Bemisia* spp. (Homoptera: Aleyrodidae) in the United States history, pest status, and management. *Recent Res. Devel. Entomol.* 2: 151-161.
- 2026.Henneberry, T.J., Toscano, N.C., Faust, R.M. and Coppedge, J.R. 1995. Silverleaf whitefly: the 5-year national research and action plan - with particular reference to current status and research progress. pp. 157-160. *In* D.A. Richter and J. Armour (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 2027.Henneberry, T.J., Toscano, N.C., Faust, R.M. and Coppedge, J.R. (Eds.). 1993. Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112, 175 pp.
- 2028.Henneberry, T.J., Toscano, N.C., Faust, R.M. and Coppedge, J.R. (Eds.). 1994. Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125, 228 pp.
- 2029.Henneberry, T.J., Toscano, N.C., Faust, R.M. and Coppedge, J.R. (Eds.). 1995. Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2, 299 pp.
- 2030.Henneberry, T.J., Toscano, N.C., Faust, R.M. and Coppedge, J.R. (Eds.). 1996. Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01, 232 pp.
- 2031.Henneberry, T.J., Toscano, N.C., Perring, T.M. and Faust, R.M. (Eds.). 1997. Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02, 268 pp.
- 2032.Henneberry, T.J., Toscano, N.C., Perring, T.M. and Faust, R.M. (Eds.). 1998. Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01, 169 pp.
- 2033.Hennessey, R.D., Arredondo-Bernal, H.C. and Rodriguez del Bosque, L.A. 1995. Distribucion geografica y huéspedes alternos de parasitoides afelinidos de *Bemisia tabaci* (Homoptera: Aleyrodidae). [Distribucion geografica mundial y huéspedes alternos de 30 especies de afelinidos que atacan a *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae)]. *Vedalia Revista Internacional De Control Biologico (Mexico)* 2(1): 61-75. [Spanish, English summary]
- 2034.Henrard, P. 1937. Les insectes parasites du cotonnier dans la region de Lisala. *Bull. Agric. Congo Belge* 28: 609-624. [Cock (1986)]
- 2035.Henry, S., Gururajan, K.N., Natarajan, K. and Krisnamurthy, R. 1990. "Kanachana", a whitefly-tolerant medium staple cotton. *Indian Farming* 40(5): 25-26.
- 2036.Henter, H.J., Brasch, K. and van Lenteren, J.C. 1996. Variation between laboratory populations of *Encarsia formosa* in their parasitization behavior on the host *Bemisia tabaci*. *Entomol. Exp. Appl.* 80(2): 435-441.
- 2037.Henter, H.J., Luttkhuizen, P.C., Visser, J.W. and van Lenteren, J.C. 1993. Variation in host use in *Encarsia formosa*. *Bull. OILB/SROP (IOBC/WPRS)* 16: 67-70.
- 2038.Henter, H.J. and van Lenteren, J.C. 1996. Variation between laboratory populations in the performance of the parasitoid *Encarsia formosa* on two host species, *Bemisia tabaci* and *Trialeurodes vaporariorum*. *Entomol. Exp. Appl.* 80(2): 427-434.
- 2039.Hequet, E. and Wyatt, B.G. 1999. Evidence on the origins of sugars causing stickiness in cotton. pp. 698-701. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 2040.Herakly, F.A. and El-Ezz, A.A. 1970. The cotton white fly, *Bemisia tabaci* Genn., infesting cucurbits, in U.A.R. *Agric. Res. Rev.* 48: 110-118. [Cock (1986)]
- 2041.Heraty, J.M. and Polaszek, A. 2000. Morphometric analysis and descriptions of selected species in the *Encarsia strenua* group (Hymenoptera: Aphelinidae). *J. Hymenoptera Res.* 9(1): 142-169.
- 2042.Hernandez, A. and Pacheco, J.J. 1998. Chancing a planting date: a silverleaf whitefly case. pp. 574-577. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 2043.Hernandez, D., Mansanet, V. and Puiggroc-Jove, J.M. 1999. Use of Confidor 200SL in vegetable cultivation in Spain. *Planzenschutz-Nachrichten Bayer* 52: 364-375.
- 2044.Hernandez-Jasso, A. and Gutierrez-Zamoran, F.J. 1996. Response of cotton to early defoliation in the Yaqui Valley, Mexico. pp. 1219-1221. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 2045.Hernandez-Jasso, A. and Gutierrez-Zamoran, F.J. 1998. Response of NuCotn 33B to early defoliation in the Yaqui Valley, Sonora, Mexico. pp. 1487-1489. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 2046.Hernandez-Jasso, A. and Pacheco-Covarrubias, J.J. 1998. Control of the cotton whitefly with planting dates in the Yaqui Valley, Sonora, Mexico. [Spanish, English summary]. *Rev. Fitotecnia Mexicana* 21(1): 83-92.
- 2047.Hernández-Jasso, A. and Pacheco-Covarrubias, J.J. 1999. Response of cotton cultivars to silverleaf whitefly and its relation to yield, in Yaqui Valley, Sonora, México. pp. 490-491. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 2048.Hernández, P.T.A., Arenas, L.D.O., Aguilera, G.M. and Bravo, J.M.C. 1999. Cubiertas flotantes como barreras contra insectos vectores de virus en sandía en Veracruz, México. [Floating covers as barriers against insect virus vectors in watermelon in Veracruz, Mexico]. *Manejo Integrado de Plagas (Costa Rica)* 51: 1-9. [Spanish, English summary]
- 2049.Hernandez-Torres, I. and Pereyra-Alferez, B. 1998. Obtencion de una cepa mutante de *Paecilomyces fumosoroseus* (Deuteromycotina: Hyphomycetes) sobreproductora de quitinasas para el control de mosquita blanca *Bemisia tabaci* (Homoptera: Aleyrodidae) [Mutant strain of chintinasas overproductive *Paecilomyces fumosoroseus* (Deuteromycotina: Hyphomycetes) for the control of white fly *Bemisia tabaci* (Homoptera: Aleyrodidae)]. pp. 361-363. *In* Proceedings 21st National Congress of Biological Control, Rio Bravo, Tampaulipas, Mexico, 5-6 Nov 1998. INIFAP. [Spanish]
- 2050.Hernandez-Velazquez, V.M., Garza-Gonzalez, E. and Berlanga-Padilla, A.M. 1995. Control microbial de mosquitas blancas con *Paecilomyces* spp en Mexico. pp. 29-36. *In* Memoria, Simposio sobre Control Biologico de Mosquita Blanca. Sociedad Mexicana de Control Biologico, Tapachula, Chiapas, Mexico.
- 2051.Hernandez, Y. 2000. *Justicia pectoralis* Jacq. new host of B biotype of *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae) in Cuba. *Rev. Proteccion Vegetal* 15(2): 132-133. [Spanish]
- 2052.Herren, H.R. and Bennett, F.D. 1984. Cassava pests, their spread and control. pp. 110-114. *In* D.L. Hawksworth (Ed.), Advancing Agricultural Production in Africa. Proceedings of CAB's First Scientific Conference, Arusha, Tanzania, Feb. 12-18, 1984. Commonwealth Agricultural Bureaux, Farnham Royal, UK. [Cock (1986)]

- 2053.Herrera, F. 1995. Uso de hongos entomopatógenos para el control microbiano de *Bemisia tabaci* (Homoptera: Aleyrodidae). [Use of entomopathogenic fungi for controlling *Bemisia tabaci* (Homoptera: Aleyrodidae)]. M. Sc. Thesis. CATIE, Turrialba, Costa Rica, 69 pp. [ Spanish]
- 2054.Herrera, F., Carballo, M. and Shannon, P. 1999. Eficacia de cepas nativas de hongos entomopatogenos sobre *Bemisia tabaci*, en el laboratorio [Efficacy of native entomopathogenic fungi strains against *Bemisia tabaci* in the laboratory]. Manejo Integrado de Plagas (Costa Rica) 54: 37-43. [ Spanish, English summary]
- 2055.Herting, B. 1972. A catalogue of parasites and predators of terrestrial arthropods. Section A Host or Prey/enemy. Volume II Homoptera. Commonwealth Agricultural Bureaux, Farnham Royal, UK, 210 pp. [Cock (1986), *Bemisia tabaci*; pp. 103-107]
- 2056.Heydari, H., Attaran, M.R. and Daniali, M. 1997. Use of *Chrysoperla carnea* Steph. against cotton pests in the northern part of Iran. Arab and Near East Plant Protection Newsletter. 25: 28.
- 2057.Heyer, W., Aviles, R., Chiang-lok, M.L. and Cruz, B. 1994. [Towards more effective chemical control measures against pests of bean (*Phaseolus vulgaris* L.) in Cuba]. Arch. Phytopathol. Plant Prot. 29(1): 89-100. [ German, English summary]
- 2058.Heyer, W., Caballero, R. and Lok, M.L.C. 1988. [The present status and possibilities for integrated control of insect pests in bean crops in the Republic of Cuba]. Beiträge zur Tropischen Landwirtschaft und Veterinärmedizin 26(3): 291-301. [ Cock (1993), German, English, Spanish, French & Russian summaries]
- 2059.Heyer, W., Lok, M.L.C. and Cruz, B. 1989. The population dynamics of the cotton whitefly, *Bemisia tabaci* Genn., in bean fields in the Republic of Cuba. Arch. Phytopathol. Plant Prot. 25(5): 473-479. [ German, English summary]
- 2060.Hidalgo-Salvatierra, O., Leon-Quant, G., Lindo -Espinoza, O. and Vaughan-Rodriguez, M. 1975. Informe de la Mision de Estudio de la Mosca Blanca. Managua, Nicaragua; Banco Nacional de Nicaragua, Comision Nacional Del Algodon & Ministerio de Agric. y Ganaderia, 120 pp. [ Cock (1986)]
- 2061.Hiebert, E., Abouzid, A.M. and Polston, J.E. 1996. Whitefly-transmitted geminiviruses. pp. 277-288. In D. Gerling and R.T. Mayer (Eds.), *Bemisia* : 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
- 2062.Hildebrand, E.M. 1960. The feathery mottle virus complex of sweetpotato. Phytopathology 50: 751-757. [ Cock (1986)]
- 2063.Hilje, L. 1993. Un esquema conceptual para el manejo integrado de la mosca blanca (*Bemisia tabaci*) en el cultivo de tomate. [A conceptual scheme for the integrated management of the whitefly (*Bemisia tabaci*) in tomato]. Manejo Integrado de Plagas (Costa Rica) 29: 51-57. [ Spanish, English summary]
- 2064.Hilje, L. 1995. Aspectos bioecologicos de *Bemisia tabaci* in Mesoamerica. [Bio-ecological aspects of *Bemisia tabaci* in Middle America]. Manejo Integrado de Plagas (Costa Rica) 35: 46-54. [ Spanish, English summary]
- 2065.Hilje, L. 1996. Escritura de artículos científicos. pp. 127-133. In L. Hilje (Ed.), Metodologias para el estudio y manejo de moscas blancas y geminivirus. Centro Agronomico Tropical de Investigacion y Ensenanza, Turrialba, Costa Rica.
- 2066.Hilje, L. 1997. Posibilidades para el manejo integrado del complejo *Bemisia tabaci*-geminivirus en Costa Rica. [Possibilities of integrated management of the *Bemisia tabaci*-geminivirus complex in Costa Rica]. Agron. Costarricense 21(1): 139-142. [ Spanish, English summary]
- 2067.Hilje, L. 1998. Un modelo de colaboracion agricola internacional para el manejo de moscas blancas y geminivirus en America Latina y el Caribe. [A model of international agricultural collaboration for whitefly and geminivirus management in Latin America and the Caribbean]. Manejo Integrado de Plagas (Costa Rica) 49: 1-9. [ Spanish, English summary]
- 2068.Hilje, L. 2000. Prácticas agrícolas para el manejo de *Bemisia tabaci*. [Cultural practices for managing *Bemisia tabaci*]. Manejo Integrado de Plagas (Costa Rica) 56: 22-30. [ Spanish, English summary]
- 2069.Hilje, L. 2000. Use of living ground covers for managing the whitefly *Bemisia tabaci* as a geminivirus vector in tomatoes. pp. 167-170. In Brighton Crop Protection Conference: Pests and Diseases. British Crop Protection Council, Farnham, UK.
- 2070.Hilje, L. 2001. Avances hacia el manejo sostenible del complejo mosca blanca-geminivirus en tomate, en Costa Rica. [Advances towards the sustainable management of the whitefly-geminivirus complex in tomato in Costa Rica]. Manejo Integrado de Plagas (Costa Rica) 61: 69-80. [ Spanish, English summary]
- 2071.Hilje, L., Costa, H.S. and Stansly, P.A. 2001. Cultural practices for managing *Bemisia tabaci* and associated viral diseases. In S. E. Naranjo and P. C. Ellsworth (Eds.), Special Issue: Challenges and Opportunities for Pest Management of *Bemisia tabaci* in the New Century. Crop Prot. 20(9): 801-812.
- 2072.Hilje, L. and Cubillo, D. 1996. Prácticas agrícolas. pp. 51-59. In L. Hilje (Ed.), Metodologias para el estudio y manejo de moscas blancas y geminivirus. Centro Agronomico Tropical de Investigacion y Ensenanza, Turrialba, Costa Rica.
- 2073.Hilje, L., Cubillo, D. and Segura, L. 1993. Observaciones ecologicas sobre la mosca blanca *Bemisia tabaci* (Gennadius) en Costa Rica. [Ecological observations on the whitefly *Bemisia tabaci* (Gennadius) in Costa Rica]. Manejo Integrado de Plagas (Costa Rica) 30: 24-30. [ Spanish, English summary]
- 2074.Hilje, L., Lastra, R., Zoebisch, T., Calvo, G., Segura, L., Barrantes, L., Alpizar, D. and Amador, R. 1992. Las moscas blancas en Costa Rica. [The whiteflies in Costa Rica]. pp. 58-63. In L. Hilje and O. Arboleda (Eds.), Las moscas blancas (Homoptera: Aleyrodidae) en America Central y el Caribe. CATIE, Turrialba, Costa Rica. [Spanish]
- 2075.Hilje, L. and Stansly, P.A. 2000. Coberturas vivas para el manejo de la mosca blanca en tomate (Hoja Tecnica No. 33) [Living covers for the management of whitefly in tomato]. Manejo Integrado de Plagas (Costa Rica) 56: i-iv. [ Spanish, English summary]
- 2076.Hill, B.G. 1968. Occurrence of *Bemisia tabaci* (Genn.) in the field and its relation to the leaf curl disease of tobacco. South African J. Agric. Sci. 11: 583-594. [ Cock (1986)]
- 2077.Hill, B.G. 1969. A morphological comparison between two species of whitefly, *Trialeurodes vaporariorum* (West). and *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae) which occur on tobacco in the Transvall. Phytophylactica 1(34): 127-145. [ Cock (1986)]
- 2078.Hinduja, C.P., Duhon, S.S. and Bunerjee, S.K. 1984. Effect of different combinations of contact and systemic insecticides on yield and profit in *desi* cotton *Gossypium arboreum* Linn. Indian J. Entomol. 46(1): 105-106. [ Cock (1993)]
- 2079.Hindy, M.A., El-Sayed, A.M., El-Salam, S.M.A. and Samy, M.A. 1997. Qualitative assessment of certain insecticides applied by different ground sprayers against whitefly, *Bemisia tabaci* (Genn.) on eggplant. Egyptian J. Agric. Res. 75(3): 565-577.
- 2080.Hinz, S.E. and Wright, J.E. 1997. Naturalis-L: a biological product (*Beauveria bassiana* JW-1) for the control of cotton pests. pp. 1300-1302. In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 2081.Hirano, K., Budiyanto, E., Swastika, N. and Fuji, K. 1995. Population dynamics of the whitefly *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae) in Java, Indonesia, with special reference to spatio-temporal changes in the quantity of food resources. Ecol. Res. 10(1): 75-85.
- 2082.Hirano, K., Budiyanto, E. and Winarni, S. 1993. Biological characteristics and forecasting outbreaks of the whitefly, *Bemisia tabaci*, a vector of virus diseases in soybean fields. Tech. Bull. No. 135, Inst. Biol. Sci. (Univ. Tsukuba, Ibaraki, Japan), 14 pp.
- 2083.Hnizdil, M. 1996. *Bemisia tabaci* Gennadius. Ochrona Roslin 32(4): 1-2. [ Czech]
- 2084.Hobolth, L.A. 1988. [Particularly noticeable pathogens in 1987]. pp. 119-123. In 5th Danish Plant Protection Conference: Pests and Diseases. Statens Planteavltsforsoeg, Lyngby, Denmark. [Danish, English summary]

- 2085.Hodde, M., van Driesche, R. and Sanderson, J. 1997. Biological control of *Bemisia argentifolii* (Homoptera: Aleyrodidae) on poinsettia with inundative releases of *Encarsia formosa* (Hymenoptera: Aphelinidae): Are higher release rates necessarily better? *Biol. Control* 10(3): 166-179.
- 2086.Hodde, M.S. 1997. Evaluating parasitoids (Hymenoptera: Aphelinidae) for biological control of *Bemisia argentifolii* (Homoptera: Aleyrodidae) on poinsettia. [University of Massachusetts]. Dissertation Abst. Int. 58(02B): 513.
- 2087.Hodde, M.S., Sanderson, J.P. and van Driesche, R.G. 1999. Biological control of *Bemisia argentifolii* (Hemiptera: Aleyrodidae) on poinsettia with inundative releases of *Eretmocerus eremicus* (Hymenoptera: Aphelinidae): does varying the weekly release rate affect control? *Bull. Entomol. Res.* 89(1): 41-51.
- 2088.Hodde, M.S. and van Driesche, R. 1996. Evaluation of *Encarsia formosa* (Hymenoptera: Aphelinidae) to control *Bemisia argentifolii* (Homoptera: Aleyrodidae) on poinsettia (*Euphorbia pulcherrima*): a life table analysis. *Florida Entomol.* 79(1): 1-12.
- 2089.Hodde, M.S. and van Driesche, R. 1999. Evaluation of *Eretmocerus eremicus* and *Encarsia formosa* (Hymenoptera: Aphelinidae) Beltsville strain in commercial greenhouses for biological control of *Bemisia argentifolii* (Homoptera: Aleyrodidae) on colored poinsettia plants. *Florida Entomol.* 82(4): 556-569.
- 2090.Hodde, M.S., van Driesche, R. and Sanderson, J. 1996. Greenhouse trials of *Eretmocerus californicus* Howard (Hymenoptera: Aphelinidae) for control of *Bemisia argentifolii* Bellows and Perring (Homoptera: Aleyrodidae) on poinsettia in Northeastern U. S. A. *Bull. OILB/SROP (IOBC/WPRS)* 19(1): 55-62.
- 2091.Hodde, M.S., van Driesche, R., Sanderson, J. and Rose, M. 1996. A photographic technique for constructing life tables for *Bemisia argentifolii* (Homoptera: Aleyrodidae) on poinsettia. *Florida Entomol.* 79(3): 464-468.
- 2092.Hodde, M.S. and van Driesche, R.G. 1999. Evaluation of inundative releases of *Eretmocerus eremicus* and *Encarsia formosa* Beltsville strain in commercial greenhouses for control of *Bemisia argentifolii* (Hemiptera: Aleyrodidae) on poinsettia stock plants. *J. Econ. Entomol.* 92(4): 811-824.
- 2093.Hodde, M.S., van Driesche, R.G., Elkinton, J.S. and Sanderson, J.P. 1998. Discovery and utilization of *Bemisia argentifolii* patches by *Eretmocerus eremicus* and *Encarsia formosa* (Beltsville strain) in greenhouses. *Entomol. Exp. Appl.* 87(1): 15-28.
- 2094.Hodde, M.S., van Driesche, R.G., Lyon, S.M. and Sanderson, J.P. 2001. Compatibility of insect growth regulators with *Eretmocerus eremicus* (Hymenoptera: Aphelinidae) for whitefly (Homoptera: Aleyrodidae) control on poinsettias. *Biol. Control* 20(2): 122-131.
- 2095.Hodde, M.S., van Driesche, R.G. and Sanderson, J.P. 1997. Biological control of *Bemisia argentifolii* (Homoptera: Aleyrodidae) on poinsettia with inundative releases of *Encarsia formosa* Beltsville strain (Hymenoptera: Aphelinidae): Can parasitoid reproduction augment inundative releases? *J. Econ. Entomol.* 90(4): 910-924.
- 2096.Hodde, M.S., van Driesche, R.G. and Sanderson, J.P. 1998. Biology and use of the whitefly parasitoid *Encarsia formosa*. *Annu. Rev. Entomol.* 43: 645-669.
- 2097.Hodde, M.S., van Driesche, R.G., Sanderson, J.P. and Minkenberg, O.P.J.M. 1998. Biological control of *Bemisia argentifolii* (Hemiptera: Aleyrodidae) on poinsettia with inundative releases of *Eretmocerus eremicus* (Hymenoptera: Aphelinidae): do release rates affect parasitism? *Bull. Entomol. Res.* 88(1): 47-58.
- 2098.Hoefert, L.L. 1987. Association of squash leaf curl virus with nuclei of squash vascular cells. *Phytopathology* 77(11): 1596-1600. [Cock (1993)]
- 2099.Hoelmer, K., Kirk, A. and Simmons, G. 1999. An overview of natural enemy explorations and evaluations for *Bemisia* in the U.S. pp. 689-696. *In Proceedings of the Fifth International Conference on Pests in Agriculture, Part 3, Montpellier, France, 7-9 December, 1999.* Association Nationale pour la Protection des Plantes (ANPP), Paris, France.
- 2100.Hoelmer, K.A. 1996. Whitefly parasitoids: can they control field populations of *Bemisia*? pp. 451-476. *In D. Gerling and R.T. Mayer (Eds.), Bemisia: 1995 Taxonomy, Biology, Damage, Control and Management.* Intercept Ltd., Andover, Hants, UK.
- 2101.Hoelmer, K.A. and Kirk, A.A. 1999. An overview of natural enemy explorations and evaluations for *Bemisia* in the U. S. *Bull. OILB/SROP (IOBC/WPRS)* 22: 109-112.
- 2102.Hoelmer, K.A., Osborne, L.S., Bennett, F.D. and Yokomi, R.K. 1994. Biological control of sweetpotato whitefly in Florida. pp. 101-113. *In D. Rosen, F.D. Bennett and J.L. Capinera (Eds.), Pest Management in the Subtropics: Biological Control: A Florida Perspective.* Intercept Ltd., Andover, UK.
- 2103.Hoelmer, K.A., Osborne, L.S. and Yokomi, R.K. 1991. Foliage disorders in Florida associated with feeding by sweetpotato whitefly, *Bemisia tabaci*. *Florida Entomol.* 74(1): 162-166.
- 2104.Hoelmer, K.A., Osborne, L.S. and Yokomi, R.K. 1993. Reproduction and feeding behavior of *Delphastus pusillus* (Coleoptera, Coccinellidae), a predator of *Bemisia tabaci* (Homoptera: Aleyrodidae). *J. Econ. Entomol.* 86: 322-329.
- 2105.Hoelmer, K.A., Osborne, L.S. and Yokomi, R.K. 1994. Interactions of the whitefly predator *Delphastus pusillus* (Coleoptera: Coccinellidae) with parasitized sweetpotato whitefly (Homoptera: Aleyrodidae). *Environ. Entomol.* 23(1): 136-139.
- 2106.Hoelmer, K.A., Roltsch, W.J., Chu, C.C. and Henneberry, T.J. 1998. Selectivity of whitefly traps in cotton for *Eretmocerus eremicus* (Hymenoptera: Aphelinidae), a native parasitoid of *Bemisia argentifolii* (Homoptera: Aleyrodidae). *Environ. Entomol.* 27(4): 1039-1044.
- 2107.Hofer, P., Bedford, I.D., Markham, P.G., Jeske, H. and Frischmuth, T. 1997. Coat protein gene replacement results in whitefly transmission of an insect nontransmissible geminivirus isolate. *Virology* 236(2): 288-295.
- 2108.Hoffman C. J. and Byrne, D.N. 1986. Effects of temperature and photoperiod upon adult eclosion of sweetpotato whiteflies, *Bemisia tabaci* (Gennadius). [German summary]. *Entomol. Exp. Appl.* 42(2): 139-143. [Cock (1993)]
- 2109.Hohmann, C.L., van Schoonhoven, A. and Cardona, C. 1980. Manejo de las plagas de frijol (*Phaseolus vulgaris* Linnaeus, 1753) a través de la utilización de sistemas de diversificación del cultivo con malezas asociado a resistencia varietal [English summary]. *Anais Soc. Entomol. Brasil* 9: 143-153. [Cock (1986)]
- 2110.Hohnle, M., Hofer, P., Bedford, I.D., Briddon, R.W., Markham, P.G. and Frischmuth, T. 2001. Exchange of three amino acids in the coat protein results in efficient whitefly transmission of a nontransmissible Abutilon mosaic virus isolate. *Virology* 290(1): 164-171.
- 2111.Hokama, N., Matsui, M., Kawano, S. and Tokashiki, I. 1993. Disorder of various vegetables caused by releasing a new type of the sweetpotato whitefly, *Bemisia tabaci* Gennadius. *Proc. Kanto-Tosan Plant Prot. Soc.* 40: 217-219. [Japanese, English summary]
- 2112.Hollands, J. 1998. Mosca blanca: detectamos nuevas especies en Canete. [White flies: new species detected in Canete]. *RAAA Boletín (Peru)* 28: 15. [Spanish]
- 2113.Hollings, M., Stone, O.M. and Bock, K.R. 1976. Purification and properties of sweetpotato mild mottle, a whitefly *Bemisia tabaci* borne virus from sweet potato *Ipomoea batatas* in east Africa. *Ann. Appl. Biol.* 82: 511-528.
- 2114.Hollings, M., Stone, O.M. and Bock, K.R. 1976. Sweet potato mild mottle virus. *CMI/AAB Descriptions of Plant Viruses* 162: 1-4. [Cock (1986)]
- 2115.Holt, J., Colvin, J. and Muniyappa, V. 1999. Identifying control strategies for tomato leaf curl virus disease using an epidemiological model. *J. Appl. Ecol.* 36(5): 625-633.
- 2116.Holt, J., Jeger, M.J., Thresh, J.M. and Otim-Nape, G.W. 1997. An epidemiological model incorporating vector population dynamics applied to African cassava mosaic virus disease. *J. Appl. Ecol.* 34(3): 793-806.

- 2117.Honda, Y., Iwaki, M., Saito, Y., Thongmeearkom, P., Kittisak, K. and Deema, N. 1983. Mechanical transmission, purification, and some properties of whitefly -borne mungbean yellow mosaic virus in Thailand. *Plant Dis.* 67: 801-804. [Cock (1986)]
- 2118.Honda, Y., Kiratiya-Angul, K., Srithongchai, W. and Kiratiya-Angul, S. 1986. Virus diseases of solanaceous plants transmitted by whitefly. pp. 51-59. *In Plant Virus Diseases of Horticultural Crops in the Tropics and Subtropics.* Food and Fertilizer Technology Centre for the Asian and Pacific Region, Taipei, Taiwan. [Cock (1993)]
- 2119.Hong, Y., Wang, X., Tian, B. and Cai, J. 1995. Chinese squash leaf curl virus: a new whitefly-transmitted geminivirus. *Science in China Series B Chemistry Life Sciences & Earth Sciences* 38(2): 179-186.
- 2120.Hong, Y.G. and Harrison, B.D. 1995. Nucleotide sequences from tomato leaf curl viruses from different countries: evidence for three geographically separate branches in evolution of the coat protein of whitefly-transmitted geminiviruses. *J. Gen. Virol.* 76(8): 2043-2049.
- 2121.Hong, Y.G., Robinson, D.J. and Harrison, B.D. 1993. Nucleotide sequence evidence for the occurrence of three distinct whitefly-transmitted geminiviruses in cassava. *J. Gen. Virol.* 74(11): 2437-2443.
- 2122.Hooks, C.R.R., Valenzuela, H.R. and Defrank, J. 1998. Incidence of pests and arthropod natural enemies in zucchini grown with living mulches. *Agric. Ecosystems Environ.* 69(3): 217-231.
- 2123.Hopkins, J.C.F. 1956. Tobacco diseases: with special reference to Africa. Commonwealth Mycological Institute, Kew, UK, 178 pp. [Cock (1986), *Bemisia tabaci*; pp. 119-136]
- 2124.Horns, T. and Jeske, H. 1991. Localization of abutilon mosaic virus (abmv) DNA within leaf tissue by in situ hybridization. *Virology* 181: 580-588.
- 2125.Horowitz, A.R. 1986. Population dynamics of *Bemisia tabaci* (Gennadius): with special emphasis on cotton fields. *Agric. Ecosystems Environ.* 17: 37-47.
- 2126.Horowitz, A.R. 1993. Control strategy for the sweetpotato whitefly, *Bemisia tabaci*, late in the cotton-growing season. *Phytoparasitica* 21(4): 281-291.
- 2127.Horowitz, A.R., Denholm, I., Groman, K. and Ishaaya, I. 1999. Insecticide resistance in whiteflies: Current status and implication for management. pp. 96-98. *In I. Denholm and P. Ioannidis (Eds.), Proceedings ENMARIA Symposium: Combating Insecticide Resistance, Thessalonikis, Greece, May 1999.*
- 2128.Horowitz, A.R., Forer, G. and Ishaaya, I. 1994. Managing resistance in *Bemisia tabaci* in Israel with emphasis on cotton. *Pestic. Sci.* 42(2): 113-122.
- 2129.Horowitz, A.R., Forer, G. and Ishaaya, I. 1995. Insecticide resistance management as a part of an IPM strategy in Israeli cotton fields. pp. 537-544. *In G.A. Constable and N.W. Forrester (Eds.), Challenging the Future: Proceedings of the World Cotton Research Conference-1.* CSIRO, Melbourne, Australia.
- 2130.Horowitz, A.R. and Gerling, D. 1992. Seasonal variation of sex ratio in *Bemisia tabaci* on cotton in Israel. *Environ. Entomol.* 21: 556-559.
- 2131.Horowitz, A.R. and Ishaaya, I. 1992. Susceptibility of the sweetpotato whitefly (Homoptera: Aleyrodidae) to buprofezin during the cotton season. *J. Econ. Entomol.* 85(2): 318-324.
- 2132.Horowitz, A.R. and Ishaaya, I. 1994. Managing resistance to insect growth regulators in the sweetpotato whitefly (Homoptera: Aleyrodidae). *J. Econ. Entomol.* 87(4): 866-871.
- 2133.Horowitz, A.R. and Ishaaya, I. 1996. Chemical control of *Bemisia* - management and application. pp. 537-556. *In D. Gerling and R.T. Mayer (Eds.), Bemisia : 1995 Taxonomy, Biology, Damage, Control and Management.* Intercept Ltd., Andover, Hants, UK .
- 2134.Horowitz, A.R., Mendelson, Z., Cahill, M., Denholm, I. and Ishaaya, I. 1999. Managing resistance to the insect growth regulator, pyriproxyfen, in *Bemisia tabaci*. *Pestic. Sci.* 55(3): 272-276.
- 2135.Horowitz, A.R., Mendelson, Z. and Ishaaya, I. 1997. Effect of abamectin mixed with mineral oil on the sweetpotato whitefly (Homoptera: Aleyrodidae). *J. Econ. Entomol.* 90(2): 349-353.
- 2136.Horowitz, A.R., Mendelson, Z., Weintraub, P.G. and Ishaaya, I. 1998. Comparative toxicity of foliar and systemic applications of acetamiprid and imidacloprid against the cotton whitefly, *Bemisia tabaci* (Hemiptera: Aleyrodidae). *Bull. Entomol. Res.* 88(4): 437-442.
- 2137.Horowitz, A.R., Podoler, H. and Gerling, D. 1984. Life table analysis of the tobacco whitefly *Bemisia tabaci* (Gennadius) in cotton fields in Israel. *Acta Oecologica* 5: 221-233. [Cock (1986)]
- 2138.Horowitz, A.R., Toscano, N.C., Youngman, R.R. and Georghiou, G.P. 1988. Synergism of insecticides with DEF in sweetpotato whitefly. (Homoptera: Aleyrodidae). *J. Econ. Entomol.* 81(1): 110-114.
- 2139.Horowitz, A.R., Toscano, N.C., Youngman, R.R., Kido, K., Knabke, J.J. and Georghiou, G.P. 1988. Synergism: potential new approach to whitefly control. *California Agric.* 43(1): 21-22, 29.
- 2140.Horowitz, A.R., Weintraub, P.G. and Ishaaya, I. 1998. Status of pesticide resistance in arthropod pests in Israel. *Phytoparasitica* 26(3): 231-240.
- 2141.Horowitz, R. 1992. Insecticide resistance monitoring for the whitefly, *Bemisia tabaci*, in flower greenhouses. *Hassadeh* 72(12): 1515-1517. [Hebrew, English summary]
- 2142.Horvat, F. and Verhoyen, M. 1981. Cytological modifications and presence of virus-like particles in cells of *Nicotiana benthamiana* (Domin) and *Manihot utilissima* (Pohl) infected with the geminivirus isolated from cassava infected with the cassava African mosaic disease. *Parasitica* 37: 119-130. [Cock (1986)]
- 2143.Hostachy, B. and Alex, D. 1993. Un nouveau defi pour les maraichers des Antilles francaises: un geminivirus de la tomate transmis par *Bemisia tabaci*. [A geminivirus on tomato crops transmitted by the white fly *Bemisia tabaci*]. *Phytoma (France)* 456: 24-27. [French]
- 2144.Hoto, F.V. and Pascholati, S.F. 2000. Powdery mildew (*Sphaerotheca fuliginea*) on zucchini squash (*Cucurbita pepo*) silvered leaves: Conidium germination, appressorium formation and biochemical aspects of the silvering. *Summa Phytopathol.* 26(1): 84-88. [Portuguese, English summary]
- 2145.Howarth, A.J. 1986. Geminiviruses. The plant viruses with single-stranded DNA genomes. *Genet. Engineering* 8: 85-99.
- 2146.Howarth, A.J., Caton, J., Bossert, M. and Goodman, R.M. 1985. Nucleotide sequence of bean golden mosaic virus and a model for gene regulation in geminiviruses. *Proc. Natl. Acad. Sci.* 82(11): 3572-3576.
- 2147.Howarth, A.J. and Goodman, R.M. 1986. Divergence and evolution of geminivirus genomes. *J. Mol. Evol.* 23: 313-319.
- 2148.Howarth, A.J. and Vandemark, G.J. 1989. Phylogeny of geminiviruses. *J. Gen. Virol.* 70: 2717-2727.
- 2149.Howell, D.R., Palumbo, J., Nelson, J., Hernandez, H. and Gayler, R. 1997. Upland and Pima cotton demonstration using IGR's Knack & Applaud to control silverleaf whitefly at the Yuma Valley Ag Center in 1996. pp. 324-331. *In Cotton, A College of Agriculture Report, Series P-108.* University of Arizona, Tucson.
- 2150.Howell, H.N., Jr. 1978. Notas sobre el complejo de las plagas del algodon en Honduras, C. A. su ecologia y su control. *Ceiba* 22(1): 29-33. [Cock (1986)]
- 2151.Hoyer, U., Maiss, E., Jelkmann, W., Lesemann, D.E. and Vetten, H.J. 1996. Identification of the coat protein gene of a sweet potato sunken vein closterovirus isolated from Keyna and evidence for a serological relationship among geographically diverse closterovirus isolates from sweet potato. *Phytopathology* 86(7): 744-750.
- 2152.Huang, J., Huang, B. and Fu, J. 1996. The revision of scientific name on rice-aleyrodid from Fujian China. *Entomol. J. East China* 5(1): 97-98. [Chinese]
- 2153.Huang, J. and Polaszek, A. 1998. A revision of the Chinese species of *Encarsia* Forster (Hymenoptera: Aphelinidae): parasitoids of whiteflies, scale insects and aphids (Hemiptera: Aleyrodidae, Diaspididae, Aphidoidea). *J. Nat. Hist.* 32(12): 1825-1966.
- 2154.Hull, R. 1989. The movement of viruses in plants. *Annu. Rev. Phytopathol.* 27: 213-240.



2155. Hulspar-Jordan, P.M. and van Lenteren, J.C. 1989. The parasite-host relationship between *Encarsia formosa* and *Trialeurodes vaporariorum* (Homoptera: Aleyrodidae). Modelling population growth of greenhouse whitefly on tomato. Agric. Univ. Wageningen Papers 89(2): 1-54.
2156. Hummer, P. 1977. Geographic, economic and social consequences of catastrophic insect pest infestation in the cotton growing areas of Cukurova, Turkey. Z. Ausl. Lanwirtsch. 16(4): 372-381.
2157. Hunter, M.S., Antolin, M.F. and Rose, M. 1996. Courtship behavior, reproductive relationships, and allozyme patterns of three North American populations of *Eretmocerus* nr. *californicus* (Hymenoptera: Aphelinidae) parasitizing the whitefly *Bemisia* sp., *tabaci* complex (Homoptera: Aleyrodidae). Proc. Entomol. Soc. Washington 98(1): 126-137.
2158. Hunter, M.S. and Kelly, S.E. 1998. Hyperparasitism by an exotic autoparasitoid: secondary host selection and the window of vulnerability of conspecific and native heterospecific hosts. Entomol. Exp. Appl. 89(3): 249-259.
2159. Hunter, M.S., Rose, M. and Polaszek, A. 1996. Divergent host relationships of males and females in the parasitoid *Encarsia porteri* (Hymenoptera: Aphelinidae). Ann. Entomol. Soc. Am. 89(5): 667-675.
2160. Hunter, M.S. and Woolley, J.B. 2001. Evolution and behavioral ecology of heteronomous aphelinid parasitoids. Annu. Rev. Entomol. 46: 251-290.
2161. Hunter, W.B., Hiebert, E., Webb, S.E., Polston, J.E. and Tsai, J.H. 1996. Precibarial and cibarial chemosensilla in the whitefly, *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae). Int. J. Insect Morph. Embryol. 25(3): 295-304.
2162. Hunter, W.B., Hiebert, E., Webb, S.E., Tsai, J.H. and Polston, J.E. 1998. Location of geminiviruses in the whitefly *Bemisia tabaci* (Homoptera: Aleyrodidae). Plant Dis. 82(10): 1147-1151.
2163. Hunter, W.B. and Polston, J.E. 2001. Development of a continuous whitefly cell line [Homoptera: Aleyrodidae: *Bemisia tabaci* (Gennadius)] for the study of begomovirus. J. Invertebr. Pathol. 77(1): 33-36.
2164. Huque, H. and Baloch, A.A. 1986. Population dynamics and bionomics studies of white flies on cotton crop and biological means to develop optimum pest management practices in Sindh. Annu. Res. Rep., Cotton Res. Institute, (Sakrand, Pakistan), 38 pp.
2165. Hurvitz, R. 1993. The policy of *Bemisia tabaci* control towards the end of the cotton season. Hassadeh 73(10): 1101-1103. [Hebrew]
2166. Husain, M.A. 1931. Annual Report to the entomologist for the year ending 30th June 1930. Punjab Dep. Agric., Lahore, 46 pp. [Cock (1986)]
2167. Husain, M.A. 1931. A preliminary note on the white-fly of cottons in the Punjab. Agric. J. India 25: 508-526. [Cock (1986)]
2168. Husain, M.A., Puri, A.N. and Trehan, K.N. 1936. Cell sap acidity and the incidence of white-fly (*Bemisia gossypiperda*) on cottons. Curr. Sci. (Bangalore) 4: 486-487. [Cock (1986)]
2169. Husain, M.A. and Trehan, K.N. 1940. Final report on the scheme of investigation on the white-fly of cotton in the Punjab. Indian J. Agric. Sci. 10(2): 101-109.
2170. Husain, M.A. and Trehan, K.N. 1942. The nature and extent of damage caused by *Bemisia gossypiperda* M. and L., the white-fly of cotton in the Punjab. Indian J. Agric. Sci. 12: 793-821. [Cock (1986)]
2171. Husain, M.A., Trehan, K.N. and Verma, P.A. 1936. Studies on *Bemisia gossypiperda* M. & L. No. 3: Seasonal activities of *Bemisia gossypiperda* M. & L. (The white-fly of cotton) in the Punjab. Indian J. Agric. Sci. 6: 893-903. [Cock (1986)]
2172. Husain, M.A., Trehan, K.N. and Verma, P.M. 1939. Economics of field-scale spraying against the white-fly of cotton (*Bemisia gossypiperda*, M. & L.). Indian J. Agric. Sci. 9: 109-126. [Cock (1986)]
2173. Husain, M.F. and Trehan, K.K. 1933. Observations on the life-history, bionomics and control of the white-fly of cotton (*Bemisia gossypiperda* M. & L.). Indian J. Agric. Sci. 3: 701-753. [Cock (1986)]
2174. Husman, S.H. and Jech, L.E. 1998. Voluntary area-wide whitefly monitoring project implementation 1995-1997 Gila Bend, Arizona. pp. 352-361. In Cotton, A College of Agriculture Report, Series P-112. University of Arizona, Tucson.
2175. Hussain, T. and Ali, M. 1975. A review of cotton diseases of Pakistan. Pakistan Cotton 19(2): 71-86. [Ali et al. (1992)]
2176. Hussain, T., Khan, M.M., Akbar, M.F., Naqvi, S.M.S.H. and Rajput, M.A. 1992. Relative toxicity and persistence of different insecticides against white fly, *Bemisia tabaci* Genn. on soybean. pp. 295-297. In M. Ahmad and A.R. Shakoori (Eds.), Proceedings of Pakistan Congress of Zoology. University of Karachi, Karachi, Pakistan.
2177. Hussain, T. and Mahmood, T. 1988. A note on leaf curl disease of cotton. Pakistan Cotton 32(4): 248-251. [Ali et al. 1992]
2178. Hussein, M.H. and Kinawy, M.M. 1987. Chemical control of white fly (*Bemisia tabaci* genn) and its effect on tomato yields. Bull. Faculty Agric. (Univ. Cairo) 38(2): 513-520.
2179. Hussein, S.M., Makadey, M.A. and Abdel-Alim, A. 1993. Susceptibility of cotton whitefly, *Bemisia tabaci* (Aleyrodidae, Homoptera) to selected insecticides, their mixtures and their synergism with DEF. Minia J. Agric. Res. Devel. (Egypt) 15(3): 681-693.
2180. Hussey, N.W. and Gurney, B. 1959. Some host plant factors affecting fecundity of white flies. Annu. Rep. Glasshouse Crops Res. Inst. (Littlehamton, UK), 5 pp.
2181. Hutchinson, J.B., Knight, R.I. and Pearson, E.O. 1949. Response of cotton to leaf-curl disease. J. Genet. 50: 100-111.
2182. Iaccarino, F.M. 1981. Aleirodidi nuovi o poco noti per l'Italia. Bull. Lab. Entomol. Agraria 'Filippo Silvestri' Portici 38: 143-157. [Cock (1986)]
2183. Idris, A.M. 1990. Cotton leaf curl virus disease in the Sudan. Med. Fac. Landbouww. Univ. Gent 55(2a): 263-267. [Cock (1993)]
2184. Idris, A.M., Bird, J. and Brown, J.K. 1999. First report of a bean-infecting begomovirus from *Macrotillium lathyroides* in Puerto Rico. Plant Dis. 83(11): 1071.
2185. Idris, A.M. and Brown, J.K. 1998. Sinaloa tomato leaf curl geminivirus: Biological and molecular evidence for a new subgroup III virus. Phytopathology 88 (7): 648-657.
2186. Idris, A.M. and Brown, J.K. 2000. Identification of a new, monopartite begomovirus associated with leaf curl disease of cotton in Gezira, Sudan. Plant Dis. 84(7): 809.
2187. Idris, A.M. and Brown, J.K. 2001. Three previously unidentified begomoviral genomes from tomato exhibiting leaf curl disease symptoms from Central Sudan. Plant Dis. 85: 1209.
2188. Idris, A.M., Smith, S.E. and Brown, J.K. 2001. Ingestion, transmission, and persistence of Chino del tomate virus (CdTV), a New World begomovirus, by Old and New World biotypes of the whitefly vector *Bemisia tabaci*. Ann. Appl. Biol. 139(1): 145-154.
2189. IITA. 1986. Infection rate of selected cassava varieties by cassava mosaic virus. p. 111. In IITA Annu. Rep. Res. Highlights 1985. (Ibadan, Nigeria). [Cock (1993)]
2190. Iizuka, N., Rajeshwari, R., Reddy, D.V.R., Goto, T., Muniyappa, V., Bharathan, N. and Ghanekar, A.M. 1984. Natural occurrence of a strain of cowpea mild mottle virus on groundnut (*Arachis hypogaea*) in India. Phytopathol. Z. 109: 245-253. [Cock (1986)]
2191. Ilyas, M., Puri, S.N. and Rote, N.B. 1991. Effects of some morphophysiological characters of leaf on incidence of cotton whitefly. J. Maharashtra Agric. Univ. (India) 16(3): 386-388.

2192. Ilyas, M.D., Puri, S.N., Lavekar, R.C., Butler, G.D., Jr. and Henneberry, T.J. 1994. Synthetic pyrethroid-induced cotton plant leaf tissue morphological changes and incidence of *Bemisia tabaci*. pp. 442-445. In G.A. Constable and N.W. Forrester (Eds.), Challenging the Future: Proceedings of the World Cotton Research Conference-1. CSIRO, Melbourne, Australia.
2193. Immaraju, J.A. 1989. The whitefly problem in cotton diagnosis and needs. *Pesticides* 23(8): 19-21.
2194. Inayatullah, C. and Ghani, M.A. 1987. Incidence of cotton whitefly and its natural enemies in Pakistan. *Pakistan J. Agric. Res.* 8(2): 179-183. [CATIE (1992)]
2195. Inayatullah, C., Ghani, M.A. and Ghaffar, A. 1985. Cotton Whitefly, *Bemisia tabaci* and its Control. Agric. Dept., Govt. Punjab, Lahore, Pakistan, 90 pp.
2196. Inbar, M., Doostdar, H., Gerling, D. and Mayer, R.T. 2001. Induction of systemic acquired resistance in cotton by BTH has a negligible effect on phytophagous insects. *Entomol. Exp. Appl.* 99(1): 65-70.
2197. Inbar, M., Doostdar, H., Leibee, G.L. and Mayer, R.T. 1999. The role of plant rapidly induced responses in asymmetric interspecific interactions among insect herbivores. *J. Chem. Ecol.* 25(8): 1961-1979.
2198. Inbar, M., Doostdar, H. and Mayer, R.T. 1999. Effects of sessile whitefly nymphs (Homoptera: Aleyrodidae) on leaf-chewing larvae (Lepidoptera: Noctuidae). *Environ. Entomol.* 28(3): 353-357.
2199. Inbar, M., Doostdar, H. and Mayer, R.T. 2001. Suitability of stressed and vigorous plants to various insect herbivores. *Oikos* 94(2): 228-235.
2200. Inbar, M., Doostdar, H., Sonoda, R.M., Leibee, G.L. and Mayer, R.T. 1998. Elicitors of plant defensive systems reduce insect densities and disease incidence. *J. Chem. Ecol.* 24(1): 135-149.
2201. Ingham, D.J. and Lazarowitz, S.G. 1993. Single missense mutation in the BR1 movement protein alters the host range of the squash leaf curl geminivirus. *Virology* 196: 694-702.
2202. Ingham, D.J., Pascal, E. and Lazarowitz, S.G. 1995. Both bipartite geminivirus movement proteins define viral host range, but only B1 determines viral pathogenicity. *Virology* 207: 191-204.
2203. Inouye, T. and Osaki, T. 1980. The first record in the literature of the possible plant virus disease that appeared in "Manyoshu", a Japanese classic anthology, as far back as the time of the 8th century. *Ann. Phytopathol. Soc. Japan* 46(1): 49-50. [Cock (1986)]
2204. Ioannou, N. 1985. Yellow leaf curl and other virus diseases of tomato in Cyprus. *Plant Pathol.* 34: 428-434. [Cock (1993)]
2205. Ioannou, N. 1987. Cultural management of tomato yellow leaf curl disease in Cyprus. *Plant Pathol.* 36(3): 367-373. [Cock (1993)]
2206. Ioannou, N. 1992. Diseases of tomato caused by whitefly-transmitted geminiviruses. pp. 53-55. In I.S. Rumbos, P. Kyriakopoulou and F. Bem (Eds.), Recent Advances in Vegetable Virus Research. 7th Conference ISHS Vegetable Virus Working Group, Athens, Greece, July 12-16, 1992. Ores Publishing, Volos, Greece.
2207. Ioannou, N. 1995. Virus transmission by whiteflies. FAO Workshop TCP/RAH/4451. Management of Whiteflies-Viruses Complex in Vegetable and Cotton Production in the Near East. Larnaca, [Cyprus] October 2-6, 1995. FAO, 32 pp.
2208. Ioannou, N. 1997. Current status of the whitefly-virus complex in the Near East. pp. 137-157. In N. Ioannou (Ed.), Management of the whitefly-virus complex. FAO Plant Production and Protection Paper 143, Rome, Italy.
2209. Ioannou, N. 1997. Integrated pest management strategies for the whitefly-virus complex in the Near East. pp. 159-180. In N. Ioannou (Ed.), Management of the whitefly-virus complex. FAO Plant Production and Protection Paper 143, Rome, Italy.
2210. Ioannou, N. 1997. Virus transmission by whiteflies. pp. 103-113. In N. Ioannou (Ed.), Management of the whitefly-virus complex. FAO Plant Production and Protection Paper 143, Rome, Italy.
2211. Ioannou, N. and Iordanou, N. 1985. Chemical control of the whitefly *Bemisia tabaci* (Gennadius), and its effect on tomato yellow leaf curl virus. *Tech. Bull., Agric. Res. Inst., Nicosia, Cyprus*, 8 pp.
2212. Ioannou, N. and Iordanou, N. 1985. Epidemiology of tomato yellow leaf curl virus in relation to the population density of its whitefly vector, *Bemisia tabaci* (Gennadius). *Tech. Bull., Agric. Res. Inst. (Cyprus)* 71: 2-7. [Cock (1993)]
2213. Ioannou, N., Kyriakou, A. and Hadjinicolis, A. 1987. Host range and natural reservoirs of tomato yellow leaf curl virus. [Greek summary]. *Tech. Bull., Agric. Res. Inst. (Nicosia)* 85: 1-8. [Cock (1993)]
2214. Ioannou, N. (Ed.). 1997. Management of the Whitefly-Virus Complex. FAO Plant Production and Protection Paper 143, Rome, Italy.
2215. Isaac, P.V. 1934. Report of the Imperial Entomologist. *Sci. Rep., Inst. Agric. Res. (Pusa)* 1932-33: 161-166. [Cock (1986)]
2216. Isaac, P.V. 1946. Report of the Imperial Entomologist. *Sci. Rep., Agric. Res. Inst. (New Delhi)* 1944-45: 73-79. [Cock (1986)]
2217. Isaacs, R. and Byrne, D.N. 1998. Aerial distribution, flight behaviour and eggload: their inter-relationship during dispersal by the sweetpotato whitefly. *J. Anim. Ecol.* 67(5): 741-750.
2218. Isaacs, R., Byrne, D.N. and Hendrix, D.L. 1998. Feeding rates and carbohydrate metabolism by *Bemisia tabaci* (Homoptera: Aleyrodidae) on different quality phloem saps. *Physiol. Entomol.* 23(3): 241-248.
2219. Isaacs, R., Cahill, M. and Byrne, D.N. 1999. Host plant evaluation behavior of *Bemisia tabaci* and its modification by external or internal uptake of imidacloprid. *Physiol. Entomol.* 24(2): 101-108.
2220. Isaacs, R., Willis, M.A. and Byrne, D.N. 1999. Modulation of whitefly take-off and flight orientation by wind speed and visual clues. *Physiol. Entomol.* 24(4): 311-318.
2221. Ishaaya, I. 1990. Benzoylphenyl ureas and other selective control agents - mechanism and application. pp. 365-376. In J.E. Casida (Ed.), *Pesticides and Alternatives*. Elsevier, Amsterdam.
2222. Ishaaya, I. 1990. Buprofezin and other insect growth regulators for controlling cotton pests. *Pestic. Outlook* 1(2): 30-33. [Cock (1993)]
2223. Ishaaya, I. 1993. Insect detoxifying enzymes: their importance in pesticide synergism and resistance. *Arch. Insect Biochem. Physiol.* 22(1/2): 263-276.
2224. Ishaaya, I., Austerweil, M. and Frankel, H. 1986. Effect of the petroleum oil virol on toxicity and chemical residue of fenprothrin applied against adults of *Bemisia tabaci* (Homoptera: Aleyrodidae) as high- and low-volume sprays. *J. Econ. Entomol.* 79: 596-599.
2225. Ishaaya, I., Blumberg, D. and Yarom, I. 1989. Buprofezin - A novel IGR for controlling whiteflies and scale insects. *Med. Fac. Landbouww. Univ. Gent* 54(3b): 1003-1008. [Cock (1993)]
2226. Ishaaya, I. and Horowitz, A.R. 1992. Novel phenoxy juvenile hormone analog (pyriproxyfen) suppresses embryogenesis and adult emergence of sweetpotato whitefly (Homoptera, Aleyrodidae). *J. Econ. Entomol.* 85: 2113-2117.
2227. Ishaaya, I. and Horowitz, A.R. 1995. Pyriproxyfen, a novel insect growth regulator for controlling whiteflies: mechanisms and resistance management. *Pestic. Sci.* 43(3): 227-232.
2228. Ishaaya, I. and Horowitz, A.R. 1998. Insecticides with novel modes of action: an overview. pp. 1-24. In I. Ishaaya and D. Degheele (Eds.), *Insecticides with Novel Modes of Action, Mechanisms and Application*. Springer-Verlag, New York.
2229. Ishaaya, I., Kontsedalov, S., Mazirov, D. and Horowitz, A.R. 2001. Biorational agents - mechanisms and importance in IPM and IRM program for controlling agricultural pests. Proceedings 53rd International Symposium on Crop Protection. *Med. Fac. Landbouww. Univ. Gent* 66(2a): 363-374.
2230. Ishaaya, I., Mendelson, Z., Ascher, K.R.S. and Casida, J.E. 1987. Cypermethrin synergism by pyrethroid esterase inhibitors in adults of the whitefly *Bemisia tabaci*. *Pestic. Biochem. Physiol.* 28(2): 155-162.

2231. Ishaaya, I., Mendelson, Z. and Horowitz, A.R. 1993. Toxicity and growth-suppression exerted by diafenthiuron in the sweetpotato whitefly, *Bemisia tabaci*. *Phytoparasitica* 21(3): 199-204.
2232. Ishaaya, I., Mendelson, Z. and Melamed-Madjar, V. 1988. Effect of buprofezin on embryogenesis and progeny formation of sweetpotato whitefly (Homoptera: Aleyrodidae). *J. Econ. Entomol.* 81(3): 781-784.
2233. Ishaaya, I., Yablonski, S., Mendelson, Z., Mansour, Y. and Horowitz, A.R. 1996. Novaluron (MCW -275), a novel benzoylphenyl urea, suppressing developing stages of lepidopteran, whitefly and leafminer pests. pp. 1013-1020. *In* Brighton Crop Protection Conference: Pests and Diseases. British Crop Protection Council, Farnham, UK.
2234. Ishii, T. 1938. Descriptions of six new species belonging to the Aphelinae from Japan. *Kontyu* 12(1): 27-32. [Cock (1986)]
2235. Iskandar, N.N. 1992. Studies on the potato tuber moth *Phthorimaea operculella* (Zell.) and the tomato whitefly *Bemisia tabaci* (Genn.). Ph.D. Dissertation, Cairo University, Egypt, 184 pp.
2236. Ismail, I.I. and Sharaf-El-Din, A.A.A. 1994. Susceptibility of lentil varieties and strains to infestation with *Bemisia tabaci* Genn. *Aphis craccivora* (Koch.) and *Liriomyza congesta* Becker in Egypt. *Zagazig J. Agric. Res. (Egypt)* 21(1): 269-277.
2237. Iwaki, M. 1986. Soybean crinkle leaf and cowpea mild mottle viruses. pp. 92-100. *In* International Symposium on Virus Diseases of Rice and Leguminous Crops in the Tropics. Tropical Agriculture Research Centre, Yatabe, Tsukuba, Ibaraki, Japan. [Cock (1993)]
2238. Iwaki, M., Thongmeearkom, P., Honda, Y. and Deema, N. 1983. Soybean crinkle leaf: a new whitefly-borne disease of soybean. *Plant Dis.* 67: 546-548. [Cock (1986)]
2239. Iwaki, M., Thongmeearkom, P., Honda, Y., Sarindu, N., Deema, N. and Surin, P. 1986. Soybean crinkle leaf disease occurring on soybean in Thailand. *Tech. Bull., Trop. Agric. Res. Center* 21: 132-143. [Cock (1993)]
2240. Iwaki, M., Thongmeearkom, P., Prommin, M., Honda, Y. and Hibi, T. 1982. Whitefly transmission and some properties of cowpea mild mottle virus on soybean in Thailand. *Plant Dis.* 66: 365-368. [Cock (1986)]
2241. Jack, R.W. 1936. Annual Report of the Division of Entomology, for the year ended 31st December 1935. *Rhodesia Agric. J.* 33: 329-335. [Cock (1986)]
2242. Jack, R.W. 1938. Annual Report of the Division of Entomology, for the year ended 31st December, 1937. *Rhodesia Agric. J.* 35: 652-659. [Cock (1986)]
2243. Jackson, D.M., Farnham, M.W., Simmons, A.M., van Giessen, W.A. and Elsey, K.D. 2000. Effects of planting pattern of collards on resistance to whiteflies (Homoptera: Aleyrodidae) and on parasitoid abundance. *J. Econ. Entomol.* 93(4): 1227-1236.
2244. Jackson, G.V.H. 1980. Disease and pests of taro. Noumea, New Caledonia; South Pacific Commission: 9-20, 39-41. [Cock (1986)]
2245. Jackson, J.E. 1970. The influence of environment on cotton growth in the Gezira. pp. 78-104. *In* M.A. Siddig and L.C. Hughes (Eds.), *Growth of Cotton in the Gezira Environment*. Agric. Res. Corp. Sudan, Cambridge: Heffer.
2246. Jackson, J.E., Burhan, H.O. and Hassan, H.M. 1973. Effects of season, sowing date, nitrogenous fertilizer and insecticide spraying on the incidence of insect pests on cotton in the Sudan Gezira. *J. Agric. Sci. (Cambridge)* 81: 491-505.
2247. Jackson, M.A., McGuire, M.R., Lacey, L.A. and Wraight, S.P. 1997. Liquid culture production of desiccation tolerant blastospores of the bioinsecticidal fungus *Paecilomyces fumosoroseus*. *Mycol. Res.* 101(1): 35-41.
2248. Jadhav, D.R., Kranthi, K.R., Tawar, K.B. and Russell, D.A. 1999. Insecticide resistance scenario on cotton pests in India. pp. 103-117. *In* Proceedings ICAC-CCRI Regional Consultation, Insecticide Resistance Management in Cotton. Central Cotton Research Institute, Multan, Pakistan.
2249. Jain, H.K., Agnihotri, N.P. and Srivastava, K.P. 1979. Toxicity of fenvalerate and estimation of its residues on some vegetables. *J. Entomol. Res. (New Delhi)* 3(2): no pages.
2250. Jambhrunkar, S.R., Nachane, M.N., Sonalkar, V.U. and Sadawarte, A.K. 1998. Management of sucking pests in cotton through cropping systems. *J. Soils Crops* 8(1): 50-52.
2251. James, R.R. 2001. Effects of exogenous nutrients on conidial germination and virulence against the silverleaf whitefly for two hyphomycetes. *J. Invertebr. Pathol.* 77(2): 99-107.
2252. James, R.R. and Elzen, G.W. 2001. Antagonism between *Beauveria bassiana* and imidacloprid when combined for *Bemisia argentifolii* (Homoptera: Aleyrodidae) control. *J. Econ. Entomol.* 94(2): 357-361.
2253. James, R.R. and Jaronski, S.T. 2000. Effect of low viability on infectivity of *Beauveria bassiana* conidia toward the silverleaf whitefly. *J. Invertebr. Pathol.* 76(3): 227-228.
2254. Jamil, F.F., Qureshi, M.J., Haq, A., Bashir, N. and Naqvi, S.H.M. 1988. Efficacy of the controlled release of 14C carbofuran formulation for pest control in cotton. pp. 169-175. *In* Pesticides: Food and Environmental Implications. International Atomic Energy Agency, Vienna, Austria. [Cock (1993)]
2255. Jancovich, J.K., Davidson, E.W. and Hendrix, D.L. 1997. Feeding chamber and diet for culture of nymphal *Bemisia argentifolii* (Homoptera: Aleyrodidae). *J. Econ. Entomol.* 90(2): 628-633.
2256. Jani, S.M. and Bharodia, R.K. 1978. Varietal susceptibility of sesamum (*Sesamum indicum* L.) to leaf curl disease. *Gujarat Agric. Univ. Res. J.* 4(1): 28-29. [CATIE (1992)]
2257. Jansen, M.G.M. and Oudman, L. 1994. Identification of whiteflies. *Netherlands Entomol. Soc.* 5: 187-188.
2258. Jaronski, S.T., Lord, J., Rosinska, J., Bradley, C., Hoelmer, K., Simmons, G., Osterlind, R., Brown, C., Staten, R. and Antilla, L. 1998. Effect of a *Beauveria bassiana*-based mycoinsecticide on beneficial insects under field conditions. pp. 651-656. *In* Brighton Crop Protection Conference: Pests and Diseases. British Crop Protection Council, Farnham, UK.
2259. Jaronski, S.T. and Lord, J.C. 1996. Evaluation of *Beauveria bassiana* (Mycotrol WF) for control of whitefly in spring cantaloupes, 1995. *Arthropod Management Tests* 21: 103.
2260. Jaronski, S.T., Lord, J.C. and Padem, R. 1996. Evaluation of *Beauveria bassiana* (Mycotrol WP) with pyrethroids for control of whiteflies on spring cantaloupes. *Arthropod Management Tests* 21: 102-103.
2261. Javid, I. 1976. Efficacy of some insecticides against cotton insects. *East African Agric. Forestry J.* 42(2): 125-126. [Cock (1986)]
2262. Jayanthi, M., Singh, K.M. and Singh, R.N. 1993. Differential influence of drip and surface irrigation on pest complex of groundnut. *Indian J. Entomol.* 55(2): 124-131.
2263. Jayanthi, M., Singh, K.M. and Singh, R.N. 1993. Pest complex of a high yielding groundnut variety MH-4 under Delhi conditions. *Indian J. Entomol.* 55(1): 30-33.
2264. Jayanthi, M., Singh, K.M. and Singh, R.N. 1993. Population build up of insect pests on MH-4 variety of groundnut influenced by abiotic factors. *Indian J. Entomol.* 55(2): 109-123.
2265. Jayanthi, M., Singh, K.M. and Singh, R.N. 1993. Succession of insect pests on high-yielding variety MH-4 groundnut under Delhi conditions. *Indian J. Entomol.* 55(1): 24-29.
2266. Jayanthi, M., Singh, K.M. and Singh, R.N. 2000. Insecticidal schedule against groundnut pests. *Indian J. Entomol.* 62(2): 109-126.
2267. Jayaraj, S. 1990. Basic research for the management of vectors of plant virus diseases. pp. 151-164. *In* P. Vidhyasekaran (Ed.), *Basic Research for Crop Disease Management*. [Cock (1993)]
2268. Jayaraj, S., Rangarajan, A.V., Murugesan, S., Santharam, G., Vijayaraghavan, S. and Thangaraj, D. 1987. Studies on the outbreak of whitefly, *Bemisia tabaci* (Gennadius) on cotton in Tamil Nadu. pp. 103-115. *In* S. Jayaraj (Ed.), *Resurgence of Sucking Pests: Proceedings of National Symposium*. Centre For Plant Protection Studies, Tamil Nadu Agric. Univ.

2269. Jayaraj, S., Rangarajan, A.V., Santharam, G. and Vijayaraghavan, S. 1986. Whitefly – a threat to cotton cultivation. pp. 185-188. *In* S. Jayaraj (Ed.), Pest and Disease Management: Oilseeds, Pulses, Millets and Cotton. Centre for Plant Protection Studies, Tamil Nadu Agric. Univ.
2270. Jayashree, K., Pun, K.B. and Doraiswamy, S. 1998. Host range of pumpkin yellow vein mosaic virus. *Madras Agric. J.* 85(1): 63-65.
2271. Jayashree, K., Pun, K.B. and Doraiswamy, S. 1998. Management of yellow vein mosaic virus disease of pumpkin. *Madras Agric. J.* 85(2): 127-129.
2272. Jayashree, K., Pun, K.B. and Doraiswamy, S. 1999. Effect of plant extracts and derivatives, butter milk and virus inhibitory chemicals on pumpkin yellow vein mosaic virus transmission. *Indian Phytopathol.* 52(4): 357-361.
2273. Jayashree, K., Pun, K.B. and Doraiswamy, S. 1999. Virus - vector relationships of yellow vein mosaic virus and whitefly (*Bemisia tabaci*) in pumpkin. *Indian Phytopathol.* 52(1): 10-13.
2274. Jayaswal, A.P. 1989. Whitefly on cotton and its management - A review. *J. Cotton Res. Dev.* 3(1): 10-22.
2275. Jayaswal, A.P. and Pundarikakshudu, R. 1989. Evaluation of cotton germplasm for resistance to whitefly *Bemisia tabaci* Gennadius. *Curr. Sci. (Bangalore)* 58: 197-198.
2276. Jayaswal, A.P. and Singh, D.P. 1987. Resurgence of whitefly on cotton and its management. pp. 78-84. *In* P. Jayaraj (Ed.), Resurgence of Sucking Pests: Proceedings of National Symposium. Centre for Plant Protection Studies, Tamil Nadu Agric. Univ., Coimbatore, India.
2277. Jech, L.E. and Husman, S.H. 1997. Gila Basin voluntary pest management project, 1995-1996. pp. 225-231. *In* Cotton, A College of Agriculture Report, Series P-108. University of Arizona, Tucson.
2278. Jech, L.E. and Husman, S.H. 1998. Correlation between early season insecticide control of pink bollworm and other pests and subsequent whitefly applications near Gila Bend, AZ, 1997. pp. 300-304. *In* Cotton, A College of Agriculture Report, Series P-112. University of Arizona, Tucson.
2279. Jech, L.E. and Husman, S.H. 1998. Improved areawide whitefly management through industry and extension partnership. pp. 1081-1083. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
2280. Jech, L.E. and Husman, S.H. 1998. Voluntary area-wide whitefly monitoring project implementation 1995-1997 Gila Bend, Arizona. pp. 1084-1087. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
2281. Jech, L.E., Husman, S.H., Ellsworth, P.C. and Diehl, J.W. 1997. Whitefly control using insect growth regulators. pp. 272-278. *In* Cotton, A College of Agriculture Report, Series P-108. University of Arizona, Tucson.
2282. Jeger, M.J., van den Bosch, F., Madden, L.V. and Holt, J. 1998. A model of analysing plant-virus transmission characteristics and epidemic development. *J. Math. Appl. Med. Biol.* 15(1): 1-18.
2283. Jennings, D.L. 1994. Breeding for resistance to African cassava mosaic geminivirus in East-Africa. *Trop. Sci.* 344: 110-122.
2284. Jensen, M.H., Valenzuela, M. and Fangmeler, D.D. 1999. Using non-woven floating covers on summer squash for exclusion of whitefly-transmitted gemini viruses. *Plasticulture* 118: 14-19.
2285. Jeske, H., Menzel, D. and Wertz, G. 1977. Electron microscopic studies on intranuclear virus-like inclusions in mosaic-diseased *Abutilon sellowianum* Reg. *Phytopathol. Z.* 89(4): 289-295.
2286. Jeske, H. and Schuchalter-Eicke, G. 1984. The abutilon mosaic virus (AbMV) in different leaf tissues of several host plant species in the Malvaceae. *Phytopathol. Z.* 109: 353-362.
2287. Jeske, H. and Wertz, G. 1980. Cytochemical characterization of plastidial inclusions in Abutilon mosaic infected *Malva parviflora* mesophyll cells. *Virology* 106: 155-158.
2288. Jeske, H. and Wertz, G. 1980. Ultrastructural and biochemical investigations on the whitefly transmitted Abutilon mosaic virus (AbMV). *J. Phytopathol.* 97(1): 43-55. [CATIE (1992)]
2289. Jesudasan, R.W.A. 1997. Biodiversity and systematics of whiteflies (Aleyrodidae: Homoptera: Insecta). pp. 113-118. *In* H. Ulrich (Ed.), Tropical Biodiversity and Systematics; International Symposium. Bonn, Germany, May 2-7, 1994. Zoologisches Museum, Zurich, Switzerland.
2290. Jesudasan, R.W.A. and David, B.V. 1993. Record on natural enemies of some economically important whiteflies. *J. Appl. Zool. Res.* 4(2): 161.
2291. Jesudasan, R.W.A., David, B.V. and Masilamani, S.R. 1997. Numerical analyses of intraspecific diversity in *Bemisia tabaci* (Gennadius) and *Trialeurodes ricini* (Misra) (Homoptera: Aleyrodidae). *Indian J. Environ. Toxic.* 7(1): 52-55.
2292. Jeyanandarajah, P. and Brunt, A.A. 1993. The natural occurrence, transmission, properties and possible affinities of cowpea mild mottle virus. *J. Phytopathol.* 137(2): 148-156.
2293. Jeyarajan, R., Doraiswamy, S., Sivaprakasam, K., Venkata-Rao, V.A. and Ramakrishnan, L. 1988. Incidence of whitefly transmitted viruses in Tamil Nadu. *Madras, Agric. J.* 75(5-6): 212-213. [Cock (1993)]
2294. Jha, A. and Misra, J.N. 1955. Yellow-vein mosaic of bhindi (*Hibiscus esculentus* L.) in Bihar. *Proc. Bihar Acad. Agric. Sci.* 4: 129-130. [Cock (1986)]
2295. Jiang, Y.X., DeBlas, C., Barrios, L. and Fereres, A. 2000. Correlation between whitefly (Homoptera: Aleyrodidae) feeding behavior and transmission of tomato yellow leaf curl virus. *Ann. Entomol. Soc. Am.* 93(3): 573-579.
2296. Jiang, Y.X., Lei, H., Collar, J.L., Martin, B., Muniz, M. and Fereres, A. 1999. Probing and feeding behavior of two distinct biotypes of *Bemisia tabaci* (Homoptera: Aleyrodidae) on tomato plants. *J. Econ. Entomol.* 92(2): 357-366.
2297. Jiang, Y.X., Nombela, G. and Muniz, M. 2001. Analysis by DC-EPG of the resistance to *Bemisia tabaci* on an Mi-tomato line. *Entomol. Exp. Appl.* 99(3): 295-302.
2298. Jianhe, C., Suyan, W., Xiaofeng, W. and Bo, T. 1995. Transmission, serology and PCR analysis of tomato leaf curl virus. *Weishengwu Xuebao* 35(5): 394-396. [Chinese, English summary]
2299. Jimenez, D.R., Shapiro, J.P. and Yokomi, R.K. 1994. Biotypic-specific expression of DSRNA in the sweetpotato whitefly. *Entomol. Exp. Appl.* 70(2): 143-152.
2300. Jimenez, D.R., Yokomi, R.K., Mayer, R.T. and Shapiro, J.P. 1995. Cytology and physiology of silverleaf whitefly-induced squash silverleaf. *Physiol. Mol. Plant Pathol.* 46(3): 227-242.
2301. Jimenez-Diaz, F. 1996. Weed hosts of viruses, population fluctuations of vectors and their relationship with virus diseases of melon (*Cucumis melo* L.) in the Comarca Lagunera, Mexico. [Spanish, English summary]. *Rev. Mexicana Fitopatología* 14(1): 31-37.
2302. Jiménez, J.I. 1996. Evaluación de inducidos de resistencia a geminivirus y promotores del crecimiento en el cultivo del tomate. [Evaluation of several substances for inducing resistance to geminivirus in tomatoes]. M. Sc. Thesis. CATIE. Turrialba, Costa Rica, 74 pp. [Spanish]
2303. Johansen, N.S. 1997. [New outbreak of cotton whitefly]. *Gartneryrket (Norway)* 87(18): 9-11. [Norwegian]
2304. Johnson, A.W. 1996. Sub-group collaborative study on insect host plant resistance. 1996 report. *Bull. D'information Coresta (France)* 3/4: 52-55.
2305. Johnson, A.W. 1997. Sub-group collaborative study on tobacco resistance to field insect pests. *Bull. D'information Coresta (France)* 3: 86-92.
2306. Johnson, D.D. and Walker, G. P. 1999. Intracellular punctures by the adult whitefly *Bemisia argentifolii* on DC and AC electronic feeding monitors. *Entomol. Exp. Appl.* 92(3): 257-270.

- 2307 Johnson, E.A. and Nuessly, G.S. 1994. Whiteflies. pp. 97-99. In L.G. Higley and D.J. Boethel (Eds.), Handbook of soybean insect pests. Entomological Society of America, Latham, MD.
- 2308 Johnson, M.W., Toscano, N.C., Reynolds, H.T., Sylvester, E.S., Kido, K.R. and Natwick, E.T. 1982. Whiteflies cause problems for southern California growers. California Agric. 9-10: 24-26. [Cock (1986)]
- 2309 Jones, G.H. and Mason, T.G. 1926. On two obscure diseases of cotton. Ann. Bot. 40: 759-772. [Cock (1986)]
- 2310 Jones, W.A., Ciomperlik, M.A. and Wolfenbarger, D.A. 1998. Lethal and sublethal effects of insecticides on two parasitoids attacking *Bemisia argentifolii* (Homoptera: Aleyrodidae). Biol. Control 11(1): 70-76.
- 2311 Jones, W.A. and Greenberg, S.M. 1998. Suitability of *Bemisia argentifolii* (Homoptera: Aleyrodidae) instars for the parasitoid *Eretmocerus mundus* (Hymenoptera: Aphelinidae). Environ. Entomol. 27(6): 1569-1573.
- 2312 Jones, W.A. and Greenberg, S.M. 1999. Host instar suitability of *Bemisia argentifolii* (Homoptera: Aleyrodidae) for the parasitoid *Encarsia pergandiella* (Hymenoptera: Aphelinidae). J. Agric. Urban Entomol. 16 (1): 49-57.
- 2313 Jones, W.A., Greenberg, S.M. and Legaspi, B. 1999. The effect of varying *Bemisia argentifolii* and *Eretmocerus mundus* ratios on parasitism. Biocontrol 44(1): 13-28.
- 2314 Jones, W.A. and Snodgrass, G.L. 1998. Development and fecundity of *Deraeocoris nebulosus* (Heteroptera: Miridae) on *Bemisia argentifolii* (Homoptera: Aleyrodidae). Florida Entomol. 81(3): 345-350.
- 2315 Jones, W.A., Wolfenbarger, D.A. and Kirk, A.A. 1994. Comparative tolerance of native and exotic parasitoids of the sweetpotato whitefly to cotton insecticides. pp. 1234-1236. In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 2316 Jones, W.A., Wolfenbarger, D.A. and Kirk, A.A. 1995. Response of adult parasitoids of *Bemisia tabaci* (Hom.: Aleyrodidae) to leaf residues of selected cotton insecticides. Entomophaga 40(2): 153-162.
- 2317 Jordá, C. 1996. Aspectos generales del tomate yellow leaf curl virus (TYLCV) y los geminivirus. pp. 13-18. In J.L. Cenis (Ed.), El Virus del Rizado Amirillo (Hoja en Cuchara) del Tomate (TYLCV) y Su Vector *Bemisia tabaci*, Jornadas 8. Consejería de Medio Ambiente, Agricultura y Agua, Murcia, Spain.
- 2318 Jorgensen, K. 1995. Western Australia prepares restrictions for white fly. Queensland Fruit and Vegetable News (Australia) 66(2): 10-11.
- 2319 Joshi, B.C. and Yadav, D.N. 1990. Biology and feeding of potential of *Mallada boninensis* (Okamoto), a chrysopid predator of white fly *Bemisia tabaci* Gennadius. J. Biol. Control 4(1): 18-21. [Cock (1993)]
- 2320 Joshi, R.D. and Dubey, L.N. 1976. Efficiency of certain insecticides in controlling leaf curl disease in chillies. Sci. Culture 42(5): 273-275. [Cock (1986)]
- 2321 Jovel, J. 1997. Daily movement of *Bemisia tabaci* in tomato plots, local dissemination of yellow mosaic disease and sources of inoculum of the ToYMoV-CR in Guayabo, Costa Rica. M. Sc. Thesis. CATIE. Turrialba, Costa Rica, 93 pp.
- 2322 Jovel, J., Hilje, L., Kleinn, C., Cartin, V. and Valverde, B. 2000. Movimientos diarios de *Bemisia tabaci* en parcelas de tomate, en Turrialba, Costa Rica. [Daily movements of *Bemisia tabaci* in tomato plants in Turrialba, Costa Rica]. Manejo Integrado de Plagas (Costa Rica) 55: 49-55. [Spanish, English summary]
- 2323 Jovel, J., Ramirez, P., Valverde, B.E. and Hilje, L. 1999. Determinación de las fuentes de inóculo del moteado amarillo del tomate (ToYMoV), en Guayabo, Costa Rica [Determination of the sources of tomato yellow mottle (ToYMoV), in Guayabo, Costa Rica]. Manejo Integrado de Plagas (Costa Rica) 54: 20-26. [Spanish, English summary]
- 2324 Joyce, A.L. and Bellows, T.S. 2000. Field evaluation of *Amitus bennetti* (Hymenoptera: Platygasteridae), a parasitoid of *Bemisia argentifolii* (Homoptera: Aleyrodidae), in cotton and bean. Biol. Control 17(3): 258-266.
- 2325 Joyce, A.L., Bellows, T.S. and Headrick, D.H. 1999. Reproductive biology and search behavior of *Amitus bennetti* (Hymenoptera: Platygasteridae), a parasitoid of *Bemisia argentifolii* (Homoptera: Aleyrodidae). Environ. Entomol. 28(2): 282-289.
- 2326 Joyce, R.J.V. 1955. Cotton spraying in the Sudan Gezira. I. Yield increase from spraying and spraying methods. II. Entomological problems arising from spraying. Bull. FAO Plant Prot. 3: 86-96, 97-103. [Cock (1986)]
- 2327 Joyce, R.J.V. 1958. Effect of the cotton plant in the Sudan Gezira on certain leaf-feeding insect pests. Nature (London) 182: 1463-1464.
- 2328 Joyce, R.J.V. 1973. Insect mobility and the philosophy of crop protection with reference to the Sudan Gezira. Pest Abstracts and News Summaries 19: 62-70. [Cock (1986)]
- 2329 Joyce, R.J.V. 1981. The control of migrant pests. pp. 209-229. In Aidley (Ed.), Animal Migration. Cambridge University Press, Cambridge, UK.
- 2330 Joyce, R.J.V. 1983. Aerial transport of pests and pest outbreaks. Bull. OEPP 13(2): 111-119. [Cock (1986)]
- 2331 Joyce, R.J.V. and Roberts, P. 1959. The determination of the size of plot suitable for cotton spraying experiments in the Sudan Gezira. Ann. Appl. Biol. 47: 287-305. [Cock (1986)]
- 2332 Jupin, I., Kouchkovsky, F., Jouanneau, F. and Gronenborn, B. 1994. Movement of tomato yellow leaf curl geminivirus (TYLCV): involvement of the protein encoded by ORF C4. Virology 204(1): 82-90.
- 2333 Jyani, D.B., Patel, N.C., Patel, J.R. and Ratanpara, H.C. 1997. Field evaluation of some brinjal varieties for resistance to insect pests and little leaf disease. Gujarat Agric. Univ. Res. J. 22(2): 75-77.
- 2334 Jyani, D.B., Patel, N.C., Ratanpara, H.C., Patel, J.R. and Borad, P.K. 1995. Varietal resistance in brinjal to insect pests and disease. Gujarat Agric. Univ. Res. J. 21(1): 59-63.
- 2335 Kadkao, S., Anusonratchada, B. and Phungcharoen, P. 1992. Distribution and factors influence the outbreak of sweetpotato whitefly. pp. 698-710. In Research Report in 1989: Soybean. Chiang Mai Field Crops Research Center, Chiang Mai, Thailand. [Thai, English summary]
- 2336 Kadkao, S., Anusonratchada, B. and Phungcharoen, P. 1992. Evaluation the yield losses of promising line soybeans which infested by sweetpotato whitefly (*Bemisia tabaci* (Gennadius)). pp. 711-720. In Research Report in 1989: Soybean. Chiang Mai Field Crops Research Center, Chiang Mai, Thailand. [Thai, English summary]
- 2337 Kaiser, W.J. and Louie, R. 1982. Heat therapy of cassava infected with African cassava mosaic disease. Plant Dis. 66(6): 475-477.
- 2338 Kajita, H. 1993. Influence of developmental stage and density of the tobacco whitefly *Bemisia tabaci* (Gennadius) on host feeding and parasitism by *Encarsia formosa* Gahan and *E. transvena* (Timberlake). Proc. Assoc. Plant Prot. Kyushu (Japan) 39: 108-110. [Japanese, English summary]
- 2339 Kajita, H. 2000. Geographical distribution and species composition of parasitoids (Hymenoptera: Chalcidoidea) of *Trialeurodes vaporariorum* and *Bemisia tabaci*-complex (Homoptera: Aleyrodidae) in Japan. Appl. Entomol. Zool. 35(1): 155-162.
- 2340 Kajita, H. and Alam, M.Z. 1996. Whiteflies on guava and vegetables in Bangladesh and their aphelinid parasitoids. Appl. Entomol. Zool. 31(1): 159-162.
- 2341 Kajita, H., Samudra, I.M. and Naito, A. 1992. Parasitism of the tobacco whitefly, *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae), by *Encarsia transvena* (Timberlake) (Hymenoptera: Aphelinidae) in Indonesia. Appl. Entomol. Zool. 27(3): 468-470.

2342. Kakahel, S.A., Amjad, M. and Islam, N. 1997. Chemical control of whitefly (*Bemisia tabaci* Gennad) and jassid (*Empoasca devastans* Dist.) in sunflower (*Helianthus annuus* L.). Sarhad J. Agric. (Pakistan) 13(1): 87-93.
2343. Kallender, H., Petty, I.T.D., Stein, V.E., Panico, M., Blench, I.P., Etienne, A.T., Morris, H.R., Coutts, R.H.A. and Buck, K.W. 1988. Identification of the coat protein gene of tomato golden mosaic virus. J. Gen. Virol. 69: 1351-1357.
2344. Kalshoven, L.G.E. and van der Vecht, J. 1950. De plagen van de cultuurgewassen in Indonesie. Deel 1. 's-Gravenhage/Bandoeng; N.V. Uitgevenj W. van Hoeve, 512 pp. [Cock (1986)]
2345. Kamel, E.H., Ibrahiem, S.A. and Tantawy, M.M. 1990. Synergism of permethrin toxicity by formamidines and formanilides in whitefly *Bemisia tabaci* (Gennadius). Minia J. Agric. Res. Devel. (Egypt) 12(3): 1771-1782.
2346. Kamel, M.H.M. 1997. Field evaluation of some new methods for controlling some tomato insect pests to minimize the insecticidal pollution. Ph.D. Dissertation, Ain-Shams University, Cairo, Egypt, 208 pp.
2347. Kanmiya, K. 1998. Mating behavior and vibratory signals on whiteflies. [Japanese, English summary]. Shokubutsu Boeki 52(1)(1): 17-22.
2348. Kannan, H.O. 2000. Side effects of common insecticides on the population of the predator *Chrysoperla carnea* in cotton. Trop. Sci. 40(4): 188-191.
2349. Kao, J., Jia, L., Tian, T., Rubio, L. and Falk, B.W. 2000. First report of cucurbit yellow stunting disorder virus (genus *Crinivirus*) in North America. Plant Dis. 84(1): 101.
2350. Kapadia, M.N. and Mittal, V.P. 1994. A technique for mass collection of cotton whitefly parasitoids. Gujarat Agric. Univ. Res. J. 19(2): 123-124.
2351. Kapadia, M.N. and Puri, S.N. 1989. Seasonal incidence of natural enemies of *Bemisia tabaci* (Gennadius). Indian J. Ecol. 16: 164-168.
2352. Kapadia, M.N. and Puri, S.N. 1990. Development relative proportions and emergence of *Encarsia transvena* (Timberlake) and *Eretmocerus mundus* Mercet important parasitoids of *Bemisia tabaci* (Gennadius). Entomon 15(3-4): 235-239. [Cock (1993)]
2353. Kapadia, M.N. and Puri, S.N. 1990. Feeding behavior of *Chrysoperla carnea* (Stephens) on the parasitized pupae of *Bemisia tabaci* (Gennadius). Entomon 15(3-4): 283-284. [Cock (1993)]
2354. Kapadia, M.N. and Puri, S.N. 1991. Biology and comparative predation efficacy of three heteropteran species recorded as predators of *Bemisia tabaci* in Maharashtra. Entomophaga 36(4): 555-559.
2355. Kapadia, M.N. and Puri, S.N. 1991. Effect of host plants of *Bemisia tabaci* Genn. on the development of its parasitoids. J. Biol. Control 5(1): 45-46.
2356. Kapadia, M.N. and Puri, S.N. 1991. Parasitization of *Chrysoperla carnea* (Stephens) in cotton field under Parbhani conditions. J. Maharashtra Agric. Univ. (India) 16(3): 453-454.
2357. Kapadia, M.N. and Puri, S.N. 1991. Persistence of different insecticides on cotton leaves against the larvae of *Chrysoperla carnea* (Stephens). Int. J. Trop. Agric. 9(2): 85-87.
2358. Kapadia, M.N. and Puri, S.N. 1991. Toxicity of different insecticides against two parasitoids of *Bemisia tabaci* (Gennadius) and their persistence against *Encarsia transvena* (Timberlake). Int. J. Trop. Agric. 9(2): 81-84.
2359. Kapadia, M.N. and Puri, S.N. 1992. Biology of *Serangium parcesetosum* Sicard as a predator of cotton whitefly. J. Maharashtra Agric. Univ. (India) 17(1): 162-163.
2360. Kapadia, M.N. and Puri, S.N. 1992. Development of *Chrysoperla carnea* reared on aphids and whitefly. J. Maharashtra Agric. Univ. (India) 17(1): 163-164.
2361. Kapadia, M.N. and Puri, S.N. 1993. Parasitism of whitefly in relation to variety, plant protection and sowing date of cotton. J. Maharashtra Agric. Univ. (India) 18(2): 313.
2362. Kapadia, M.N. and Puri, S.N. 1993. Parasitism of whitefly on different host plants under greenhouse conditions. J. Maharashtra Agric. Univ. (India) 18(2): 312-313.
2363. Kapadia, M.N. and Puri, S.N. 1994. Varietal preference of parasitoids of cotton whitefly. J. Maharashtra Agric. Univ. [India] 19(2): 278.
2364. Kapadia, M.N., Puri, S.N., Butler, G.D., Jr. and Henneberry, T.J. 1992. Whitefly, *Bemisia tabaci* Genn., and parasitoid populations in insecticide treated cotton. J. Appl. Zool. Res. 3(1): 7-10.
2365. Kapoor, S.P., Singh, J., Chopra, B.L., Sohi, A.S., Rewal, H.S. and Narang, D.D. 1994. Cotton leaf curl in Punjab. Plant Dis. Res. 9(1): 86-91.
2366. Karaman, G.A., Mostafa, N.R. and Abdel Alim, A.A. 1983. The simultaneous effect of certain weather factors on the population activity of the whitefly *Bemisia tabaci* (Gennadius) in Minia Region. Minia J. Agric. Res. Devel. (Egypt) 5(2): 153-167. [CATIE (1992)]
2367. Karaman, G.A., Mostafa, N.R., Ali, M.A. and Abdel-Alim, A.A. 1984. Studies on the age structure and specific population trend of the white fly, *Bemisia tabaci* (Gennadius) infesting soy-beans in middle Egypt. Minia J. Agric. Res. Devel. (Egypt) 6(1): 1-14.
2368. Karel, A.K. and Autrique, A. 1994. Insectos y otras plagas en Africa. [Insects and other bean pests in Africa]. pp. 519-575. In M.A. Pastor and H.F. Schwartz (Eds.), Bean Production Problems in the Tropics. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. [Spanish]
2369. Karim, S., Nasreen, Z., Malik, K. and Riazuddin, S. 1998. A simple and fast method for mass rearing of sweet potato whitefly (*Bemisia tabaci* and cotton aphid (*Aphis gossypii*) in contained conditions. Asia Life Sci. 7(1): 103-108.
2370. Karnkowski, W. 1999. Quarantine insects, mites and nematodes intercepted in consignments of ornamental plants imported to Poland in 1993-1998. Progress Plant Prot. 39(1): 312-320. [Polish, English summary]
2371. Karnkowski, W. 1999. Quarantine pests intercepted in consignments of ornamental plants imported into Poland in 1993/1998. Bull. OEPP 29(1-2): 45-49.
2372. Kashima, T. and Mat sui, M. 1998. Effects of the elimination of ultraviolet rays on major pests of tomato and their parasitoids. [Japanese, English summary]. Proc. Kanto-Tosan Plant Prot. Soc. 45: 185-189.
2373. Kasrawi, M.A. 1989. Inheritance of resistance to tomato yellow leaf curl virus (TYLCV) in *Lycopersicon pimpinellifolium*. Plant Dis. 73: 435-437.
2374. Kasrawi, M.A. and Mansour, A. 1994. Genetics of resistance to tomato yellow leaf curl virus in tomato. J. Hortic. Sci. 69: 1095-1100.
2375. Katiyar, K.N. 1987. Resurgence of cotton whitefly – a review. pp. 85-89. In P. Jayaraj (Ed.), Resurgence of Sucking Pests: Proceedings of National Symposium. Centre for Plant Protection Studies, Tamil Nadu Agric. Univ., Coimbatore, India.
2376. Kato, K., Onuki, M., Fuji, S. and Hanada, K. 1998. The first occurrence of tomato yellow leaf curl virus in tomato (*Lycopersicon esculentum* Mill.) in Japan. Ann. Phytopathol. Soc. Japan 64(6): 552-559.
2377. Kawana, T. and Fukuda, H. 1992. Chemical control of the sweetpotato whitefly, *Bemisia tabaci* Gennadius. Proc. Kanto-Tosan Plant Prot. Soc. 39: 215-218. [Japanese]
2378. Kawana, T., Fukuda, H. and Shimizu, K. 1990. Notes on control methods of the sweetpotato whitefly, *Bemisia tabaci*. Proc. Kanto-tosan Plant Prot. Soc. 37: 209-211. [Japanese]
2379. Kaygisiz, H. 1976. Mediterranean region cotton damage caused by the white fly (*Bemisia tabaci*). (Turkish). Bolge Zirai Mucadele Arastirma Enstitusu Mudurlugu arastirma eserleri serisi 45: 1-58.
2380. Kays, S.J., Severson, R.F., Nottingham, S.F., Chalfant, R.B. and Chortyk, O. 1995. Possible biopesticide from *Petunia* for the control of the sweetpotato whitefly (*Bemisia tabaci*) on vegetable crops. Proc. Florida State Hortic. Soc. 107: 163-167.

2381. Kayser, H., Kaufmann, L., Schurmann, F. and Harrewijn, P. 1994. Pymetrozine (GGA 215'944): a novel compound for aphid and whitefly control: an overview of its mode of action. pp. 737-742. *In* Brighton Crop Protection Conference: Pests and Diseases. British Crop Protection Council, Farnham, UK.
2382. Kegler, H. 1994. Incidence, properties and control of tomato yellow leaf curl virus - a review. *Arch. Phytopathol. Plant Prot.* 29(2): 119-132.
2383. Kelly, J.W., Adler, P.H., Decoteau, D.R. and Lawrence, S. 1989. Colored reflective surfaces to control whitefly on poinsettia. *HortScience* 24(6): 1045.
2384. Kelly-Johnson, S., Sivasupramaniam, S. and Watson, T.F. 1995. Seasonal population dynamics of sweetpotato whitefly in Maricopa County, Arizona. pp. 979-984. *In* D.A. Richter and J. Armour (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
2385. Kenchaiah, R.P.M. and Porte, B.S. 1989. Some observations on population fluctuation of insect pests of groundnut in Karnataka. *Plant Prot. Bull. (Faridabad)* 41(3-4): 7-10. [Cock (1993)]
2386. Kennedy, F.G. 1994. International results with Naturalis L for control of sweetpotato whitefly (silverleaf whitefly). pp. 1111-1112. *In* D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
2387. Kennedy, G.G. and Storer, N.P. 2000. Life systems of polyphagous arthropod pests in temporally unstable cropping systems. *Annu. Rev. Entomol.* 45: 467-493.
2388. Kerns, D.L. and Palumbo, J.C. 1995. Using Admire on desert vegetable crops. *IPM Series No. 4*, Univ. Arizona Coop. Ext. Publ. 195017, 4 pp.
2389. Kerns, D.L. and Tellez, T. 1998. Efficacy of experimental insecticides for insect control of cotton grown in the Low Desert Region of Arizona, 1997. pp. 422-434. *In* Cotton, A College of Agriculture Report, Series P-112. University of Arizona, Tucson.
2390. Kerns, D.L. and Tellez, T. 1998. Efficacy of experimental insecticides for whitefly control in cotton, 1996. pp. 367-377. *In* Cotton, A College of Agriculture Report, Series P-112. University of Arizona, Tucson.
2391. Kerns, D.L. and Tellez, T. 1998. Efficacy of experimental insecticides for whitefly control in cotton, 1997. pp. 378-393. *In* Cotton, A College of Agriculture Report, Series P-112. University of Arizona, Tucson.
2392. Kerns, D.L. and Tellez, T. 1999. Evaluation of insecticides for whitefly control in cotton, 1998. pp. 355-360. *In* Cotton, A College of Agriculture Report, Series P-116. University of Arizona, Tucson.
2393. Keshwal, R.L., Gupta, K.N. and Khatri, R.K. 1999. Appearance and sources of soybean yellow mosaic virus in Jabalpur (M.P.). *Indian J. Virol.* 15(2): 97-99.
2394. Kfoury, L., Kfoury, R. and Traboulsi, A. 1997. En cultures de concombres sous serres au Liban. Effet de trois insecticides sur les aleurodes. [Greenhouse cucumbers in Lebanon. The effect of three insecticides against whiteflies]. *Phytoma (France)* 495: 38-39. [French, English summary]
2395. Khalid, S., Shah, H. and Masood, A.A. 1999. Cotton leaf curl, virus induced symptoms and their relation with virus titer and epitope profile. pp. 245-249. *In* Proceedings ICAC-CCRI Regional Consultation, Insecticide Resistance Management in Cotton. Central Cotton Research Institute, Multan, Pakistan.
2396. Khalid, S., Soomro, M.H. and Ahmad, I. 1997. Occurrence of cotton leaf curl virus (CLCuV) in Sindh. *Pakistan J. Bot.* 29(1): 173-174.
2397. Khalifa, A. and El Khidir, E. 1964. Biological study on *Trialeurodes lubia* and *Bemisia tabaci* (Aleyrodidae). *Bull. Entomol. Soc. Egypt* 48: 115-129. [Cock (1986)]
2398. Khalifa, H. and Gameel, O.I. 1982. Control of cotton stickiness through breeding cultivars resistant to whitefly (*Bemisia tabaci* (Genn)) infestation. pp. 181-186. *In* Meeting on Use of Induced Mutations on Improvement of Oil Seed and Other Industrial Crops. International Atomic Energy Agency, Vienna, Austria.
2399. Khalifa, H. and Gameel, O.I. 1983. Breeding cotton cultivars resistant to whitefly (*Bemisia tabaci* (Genn)). pp. 231-236. *In* F. Lamberti, J.M. Waller and N.A. van der Graaf (Eds.), *Durable Resistance in Crops*. Plenum Press, New York, USA. [Cock (1986)]
2400. Khalil, F.A., Watson, W.M. and Guirguis, M.W. 1983. Evaluation of dimilin and its combinations with different insecticides against some cotton pests in Egypt. *Bull. Entomol. Soc. Egypt Econ. Ser.* 1978-79(11): 71-76.
2401. Khalil, O. and Sabet, K.A. 1971. A study of certain aetiological aspects of bacterial leaf-blight of castor bean. *Sudan Agric. J.* 6: 26-33. [Cock (1986)]
2402. Khambay, B.P.S., Batty, D., Beddie, D.G., Denholm, I. and Cahill, M.R. 1997. A new group of plant-derived [*Calceolaria andina*] naphthoquinone pesticides. *Pestic. Sci.* 50(4): 291-296.
2403. Khambay, B.P.S., Batty, D., Cahill, M., Denholm, I., Mead-Briggs, M., Vinall, S., Niemeyer, H.M. and Simmonds, M.S.J. 1999. Isolation, characterization, and biological activity of naphthoquinones from *Calceolaria andina* L. *J. Agric. Food Chem.* 47(2): 770-775.
2404. Khan, J.A. 2000. Detection of tomato leaf curl geminivirus in its vector *Bemisia tabaci*. *Indian J. Exp. Biol.* 38(5): 512-515.
2405. Khan, M., Khalid, S. and Hassan, S. 2000. Effect of some neem products on transmission of cotton leaf curl virus through *Bemisia tabaci*. *Sarhad J. Agric. (Pakistan)* 16(6): 593-600.
2406. Khan, M.A. 1987. Occurrence of whiteflies - an important virus vector in Kashmir Valley. *Res. Dev. Rep.* 4(1): 102.
2407. Khan, M.A. and Mukhopadhyay, S. 1985. Effect of different pesticide combinations on the incidence of yellow vein mosaic virus disease of okra (*Abelmoschus esculentus*) and its whitefly vector *Bemisia tabaci* Genn. *Indian J. Virol.* 1(2): 147-151. [Cock (1993)]
2408. Khan, S.M. and Ullah, Z. 1994. Population dynamics of insect pests of cotton in Dera Ismail Khan. *Sarhad J. Agric. (Pakistan)* 10(3): 285-290.
2409. Khattab, M.M., Fayad, A.N. and Mikhael, R.H.G. 1995. Relative susceptibility of different potato varieties to infestation with insect pests. *Moshtohor Ann. Agric. Sci. (Egypt)* 33(4): 1539-1552.
2410. Khattat, A.R., Zwain, Q.K., Kasim, A.K. and Khefaji, A.A. 1997. Iraq. pp. 27-32. *In* N. Ioannou (Ed.), *Management of the whitefly - virus complex*. FAO Plant Production and Protection Paper 143, Rome, Italy.
2411. Khey-Pour, A., Bananej, K., Dafalla, G.A., Caciagli, P., Noris, E., Ahoonmanesh, A., Lecoq, H. and Gronenborn, B. 2000. Watermelon chlorotic stunt virus from the Sudan and Iran: Sequence comparisons and identification of a whitefly-transmission determinant. *Phytopathology* 90(6): 629-635.
2412. Khey-Pour, A., Bendahmane, M., Matzeit, V., Accotto, G.P., Crespi, S. and Gronenborn, B. 1991. Tomato yellow leaf curl virus from Sardinia is a whitefly-transmitted monopartite geminivirus. *Nucleic Acids Res.* 19(24): 6763-6770.
2413. Kim, K.S., Bird, J., Rodriguez, R.L., Martin, E.M. and Escudero, J. 1986. Ultrastructural studies of *Jatropha gossypifolia* infested with *Jatropha* mosaic virus, a whitefly-transmitted Geminivirus. *Phytopathology* 76: 80-85. [Cock (1986)]
2414. Kim, K.S., Chung, B.J. and Kim, H.K. 2000. DBI-3204: a new benzoylphenyl urea insecticide with a particular activity against whitefly. pp. 41-46. *In* Brighton Crop Protection Conference: Pests and Diseases. British Crop Protection Council, Farnham, UK.
2415. Kim, K.S. and Flores, E.M. 1979. Nuclear changes associated with *Euphorbia* mosaic virus transmitted by the whitefly. *Phytopathology* 69: 980-984. [Cock (1986)]
2416. Kim, K.S. and Fulton, R.W. 1984. Ultrastructure of *Datura stramonium* infected with *Euphorbia* virus suggestive of a whitefly-transmitted geminivirus. *Phytopathology* 74: 236-241. [Cock (1986)]

- 2417.Kim, K.S. and Lee, K.W. 1992. Geminivirus-induced macrotubules and their suggested role in cell-to-cell movement. *Phytopathology* 82(6): 664-669.
- 2418.Kim, K.S., Shock, T.L. and Goodman, R.M. 1978. Infection of *Phaseolus vulgaris* by bean golden mosaic virus: ultrastructural aspects. *Virology* 89: 22-33. [Cock (1986)]
- 2419.King, H.H. 1932. Report of the Government Entomologist for the year 1931. *Bull. Wellcome Trop. Res. Lab., Entomol. Sect.* 35: 1-12. [Cock (1986)]
- 2420.King, W.J. 1976. Ultra-low-volume application of insecticides to cotton in The Gambia. *Misc. Rep., Centre for Overseas Pest Res.* 27: 1-13. [Cock (1986)]
- 2421.King, W.J. 1978. Very-low-volume application of insecticides to cotton in The Gambia. *Misc. Rep., Ministry of Overseas Dev.* 44: 1-15. [Cock (1986)]
- 2422.King, W.J. 1980. Cotton in the Gambia (report on the Cotton Development Project 1975-1978). *Misc. Rep., Overseas Dev. Admin.* 51: 1-18. [Cock (1986)]
- 2423.Kingdon, L. 1985. Cotton leaf crumple causes yield loss. *Arizona Farmer Stockman* 64(4): 41.
- 2424.Kinjo, K. and Matsui, M. 1994. The possible use of nets treated with insecticides in the control of the sweetpotato whitefly, *Bemisia tabaci* (Gennadius), in greenhouses or nurseries. *Proc. Kanto-tosan Plant Prot. Soc.* 41: 217-221. [Japanese, English summary]
- 2425.Kirby, R., Clarke, B.A. and Rybicki, E.P. 1989. Evolutionary relationship of three Southern African maize streak virus isolates. *Intervirology* 30: 96-101.
- 2426.Kiriukhin, G. 1947. Quelques Aleurododea de l'Iran. *Entomol. Phytopathol. Appl.* 5: 8-10,22-28. [Cock (1986), Persian, French summary]
- 2427.Kirk, A.A. and Lacey, L.A. 1996. A systematic approach to foreign exploration for natural enemies of *Bemisia* and some current results. pp. 531-533. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia* : 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
- 2428.Kirk, A.A., Lacey, L.A., Brown, J.K., Ciomperlik, M.A., Goolsby, J.A., Vacek, D.C., Wendel, L.E. and Napompeh, B. 2000. Variation in the *Bemisia tabaci* s. l. species complex (Hemiptera : Aleyrodidae) and its natural enemies leading to successful biological control of *Bemisia* biotype B in the USA. *Bull. Entomol. Res.* 90(4): 317-327.
- 2429.Kirk, A.A., Lacey, L.A. and Goolsby, J.A. 2001. Classical biological control of *Bemisia* and successful integration of management strategies in the United States. pp. 309-331. *In* K.F. Harris, O. Smith and J.E. Duffus (Eds.), *Virus-Insect-Plant Interactions*. Academic Press, San Diego, CA.
- 2430.Kirk, A.A., Lacey, L.A., Roidakis, N. and Brown, J.K. 1993. The status of *Bemisia tabaci* (Hom.: Aleyrodidae), *Trialeurodes vaporariorum* (Hom.: Aleyrodidae) and their natural enemies in Crete. *Entomophaga* 38(3): 405-410.
- 2431.Kirk, A.D. and Thistlewood, H. 1999. Development of host specificity tests for predators as biological control agents: an example for *Clitostethus arcuatus* (Rossi) (Coleoptera: Coccinellidae) on *Bemisia tabaci* Gennadius (Homoptera: Aleyrodidae) B-biotype species complex. *Bull. OILB/SROP (IOBC/WPRS)* 22: 33.
- 2432.Kirk, I.W., Bouse, L.F., Carlton, J.B., Franz, E., Latheef, M.A., Weight, J.E. and Wolfenbarger, D.A. 1994. Within-canopy spray distribution from fixed-wing aircraft. *Trans. Am. Soc. Agric. Eng.* 37(3): 745-752.
- 2433.Kirk, I.W., Bouse, L.F., Carlton, J.B., Franz, E., Latheef, M.A. and Wright, J.E. 1993. Aerial spray deposition for sweetpotato whitefly control. pp. 1623-1625. *In* D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
- 2434.Kirk, I.W., Bouse, L.F., Latheef, M.A. and Carlton, J.B. 1994. Spray deposition by different aerial delivery systems for control of sweetpotato whitefly in cotton. pp. 546-548. *In* D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
- 2435.Kirkpatrick, T.W. 1931. Further studies on leaf-curl of cotton in the Sudan. *Bull. Entomol. Res.* 22: 323-363. [Cock (1986)]
- 2436.Kisha, J. 1979. Insect pests of vegetables. pp. 130-141. *In* Annu. Rep. Gezira Res. Stn. Substns. 1972/1973. *Agric. Res. Corp., Sudan* [Cock (1993)]
- 2437.Kisha, J. 1980. Insect pests of vegetables. pp. 154-168. *In* Annu. Rep. Gezira Res. Stn. Substns. 1973/1974. *Agric. Res. Corp., Sudan* [Cock (1993)]
- 2438.Kisha, J. 1985. Entomology. pp. 120-127. *In* Annu. Rep. Gezira Res. Stn. Substns. 1977/1978. *Agric. Res. Corp., Sudan* [Cock (1993)]
- 2439.Kisha, J.S.A. 1981. The effect of insecticides on *Bemisia tabaci*, tomato leaf curl virus disease incidence and yield of tomatoes in the Sudan. *Ann. Appl. Biol.* 99: 231-239. [Cock (1986)]
- 2440.Kisha, J.S.A. 1981. Observations on the trapping of the whitefly *Bemisia tabaci* by glandular hairs on tomato leaves. *Ann. Appl. Biol.* 97: 123-127. [Cock (1986)]
- 2441.Kisha, J.S.A. 1984. Whitefly, *Bemisia tabaci*, infestations on tomato varieties and a wild *Lycopersicon* species. *Ann. Appl. Biol. (Suppl., Tests of Agrochemicals and Cultivars)* 104(5): 124-125. [Cock (1986)]
- 2442.Kisha, J.S.A. 1986. Comparison of electrodynamic spraying with use of a knapsack sprayer for control of whitefly on tomato in the Sudan Gezira. *Ann. Appl. Biol. (Suppl., Tests of Agrochemicals and Cultivars)* 108(7): 36-37. [Cock (1993)]
- 2443.Kishaba, A.N., Castle, S., McCreight, J.D. and Desjardins, P.R. 1992. Resistance of white-flowered gourd to sweetpotato whitefly. *HortScience* 27: 1217-1221.
- 2444.Kiss, F. and Budai, C. 1995. Termeszetes alapu keszitmenyek alkalmazasa a hajtatott paprika karositoi ellen. [Preparations based on natural materials used against pests of forced paprika in greenhouses]. *Novenyvedelem (Hungary)* 31(8): 387-389. [Hungarian]
- 2445.Klaassen, V.A., Boeshore, M.L., Koonin, E.V., Tian, T. and Falk, B.W. 1995. Genome structure and phylogenetic analysis of lettuce infectious yellows virus, a whitefly-transmitted, bipartite closterovirus. *Virology* 208(1): 99-110.
- 2446.Kleinn, C., Jovel, J. and Hilje, L. 1999. A model for assessing the effect of distance on disease spread in crop fields. *Crop Prot.* 18: 609-617.
- 2447.Klinkenberg, F.A., Ellwood, S. and Stanley, J. 1989. Fate of African cassava mosaic virus coat protein deletion mutants after agroinoculation. *J. Gen. Virol.* 70: 1837-1844.
- 2448.Knauf, T.A. 1992. Naturalis L: a biorational insecticide for boll weevil and whitefly control. pp. 31-32. *In* D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
- 2449.Knauf, T.A. and Wright, J.E. 1994. *Beauveria bassiana* (ATCC 74040): control of insect pests in field crops and ornamentals. pp. 1103-1108. *In* Brighton Crop Protection Conference: Pests and Diseases. British Crop Protection Council, Farnham, UK.
- 2450.Knorr, L.C., Paterson, E.C. and Proctor, J.H. 1961. World citrus problems. I. Aden Protectorate. *FAO Plant Prot. Bull.* 9(6): 91-98. [Cock (1986)]
- 2451.Knowles, T.C. and Jaronski, S.T. 1997. Control of silverleaf whitefly in fall cantaloupe with *Beauveria bassiana*, 1995. *Arthropod Management Tests* 22: 115.
- 2452.Kobatake, H., Osaki, T. and Inouye, T. 1981. Ecology and control of yellow dwarf disease of tomato caused by tobacco leaf curl virus. *Proc. Kansai Plant Prot. Soc.* 23: 8-14. [Cock (1986)]



- 2453.Koch, W. 1989. [A 'new' whitefly in the greenhouse. Description of and prospects for control of *Bemisia tabaci*]. Deutscher Gartenbau 43(14): 892-894. [ Cock (1993), German]
- 2454.Kocheili, F. and Komeili, A.A. 1993. A study on population dynamics of whitefly *Bemisia tabaci* Genn. (Homop. Aleyroidae) and its attraction rate to soybean varieties in Ahvaz Iran. Shahid Chamran Univ. Sci. J. Agric.(Iran) 16(1/2): 19. [ Persian]
- 2455.Koellner, V. and Moll, M. 1993. Studies on the biological control of the cotton whitefly, *Bemisia tabaci* (Gennadius), with the parasitic wasp *Eretmocerus mundus* Mercet. pp. 133-143. In S.E. Smolka, P. Mattusch and M. Hommes (Eds.), Components of Integrated Plant Protection in Horticulture -Current Research Work at the Institute for Plant Protection in Horticultural Crops. Biologische Bundesanstalt Fuer Land Und Forstwirtschaft, Berlin. [German]
- 2456.Koenig, J.P., Lawson, D.S. and White, S.M. 1998. Utility of Fulfill 50 WG for aphid and whitefly management in cotton. pp. 997-999. In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 2457.Kogan, M. 1996. Areawide management of major pests: is the concept applicable to the *Bemisia* complex ? pp. 643-657. In D. Gerling and R.T. Mayer (Eds.), *Bemisia* : 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
- 2458.Kok, F.N., Arica, M.Y., Gencer, O., Abak, K. and Hasirci, V. 1999. Controlled release of aldicarb from carboxymethyl cellulose microspheres: *In vitro* and field applications. Pestic. Sci. 55(12): 1194-1202.
- 2459.Konate, G., Barro, N., Fargette, D., Swanson, M.M. and Harrison, B.D. 1995. Occurrence of whitefly -transmitted geminiviruses in crops in Burkina Faso, and their serological detection and differentiation. Ann. Appl. Biol. 126(1): 121-129.
- 2460.Kooner, B.S. and Singh, H. 1981. Control of whitefly and the yellow-mosaic virus in green gram with granular insecticides. J. Res. (Punjab Agric. Univ. ) 17(1980): 268-271. [ Cock (1986)]
- 2461.Kooner, B.S., Singh, H. and Sandhu, G.S. 1986. Control of yellow mosaic virus in soybean by the soil application of granular systemic insecticides. Indian J. Entomol. 48(2): 125-129.
- 2462.Kooner, B.S., Singh, K., Singh, H. and Singh, K.B. 1977. Field-screening of mungbean germplasm against whitefly (*Bemisia tabaci* Genn.) and yellow mosaic. J. Res. (Punjab Agric. Univ.) 14(1): 75-76.
- 2463.Koonin, E.V. and Illyina, T.V. 1992. Geminivirus replication proteins are related to prokaryotic plasmid rolling circle DNA replications initiator proteins. J. Gen. Virol. 73: 2763-2766.
- 2464.Korkor, A.A., Hamid, A.M. and El-Beltagi, A.M. 1998. Effect of different local inorganic products on the population density of some cotton pests. Alexandria Sci. Exchange (Egypt) 19(4): 559-570.
- 2465.Korolev, N. and Gindin, G. 1999. Vegetative compatibility in the entomopathogen *Verticillium lecanii*. Mycol. Res. 103(7): 833-840.
- 2466.Koya, K.M.A., Gautam, S.S.S. and Banerjee, S.K. 1983. *Bemisia tabaci* Guen - a new pest of cinnamon. Indian J. Entomol. 45: 198. [ Cock (1986)]
- 2467.Koyama, K. and Matsui, M. 1994. Nutritional physiology of the sweetpotato whitefly, *Bemisia tabaci* (Gennadius): survival period of the adults reared on distilled water alone under various temperatures. Proc. Kanto-Tosan Plant Prot. Soc. 41: 211-212. [ Japanese]
- 2468.Koyama, K. and Matsui, M. 1997. Survival of silverleaf whitefly, *Bemisia argentifolii* Bellows & Perring on various sugar solutions. Proc. Kanto-Tosan Plant Prot. Soc. 44: 231-234. [ Japanese]
- 2469.Kraemer, P. 1966. Serious increase of cotton whitefly and virus transmission in Central America. J. Econ. Entomol. 59: 1531. [ Cock (1986)]
- 2470.Kranthi, K.R., Jadhav, D.R., Wanjari, R.R., Ali, S.S. and Russell, D. 2001. Carbamate and organophosphate resistance in cotton pests in India, 1995 to 1999. Bull. Entomol. Res. 91(1): 37-46.
- 2471.Krebs, E.K. 1989. [The control of whiteflies. a subject worthy for discussion also in this poinsettia season]. Gärtnerbörse und Gartenwelt (Germany) 89(28): 1358-1362. [ Cock (1993), German]
- 2472.Krebs, E.K. 1992. [Plant protection in elatior begonias]. Gärtnerbörse und Gartenwelt (Germany) 92(23): 1123-1126. [ German]
- 2473.Krebs, E.K. 1993. [*Bemisia tabaci* in poinsettias can be controlled biologically]. Gartenbau Magazin (Germany) 2(3): 58-60. [ German]
- 2474.Kring, J.B. and Schuster, D.J. 1992. Management of insects on pepper and tomato with UV-reflective mulches. Florida Entomol. 75(1): 119-129.
- 2475.Krishna, A., Nema, A.G., Nadkarni, P.G. and Bhalla, P.L. 1982. Control of leaf curl of chillies by controlling the white fly vector. Indian J. Mycol. Plant Pathol. 12: 121.
- 2476.Kubuta, A. 1991. [Effect of insecticides on sweet potato whitefly *Bemisia tabaci*]. Bulletin of the Saitama Horticultural Experimental Station. 18: 29-36. [ Cock (1993), Japanese, English summary]
- 2477.Kuehne, S. 1998. Open rearing of generalist predators: a strategy for improvement of biological pest control in greenhouses. Phytoparasitica 26(4): 277-281.
- 2478.Kular, J.S. and Butter, N.S. 1995. Screening cotton germplasm against the whitefly, *Bemisia tabaci* Genn. J. Entomol. Res. (New Delhi) 19(4): 341-344.
- 2479.Kular, J.S. and Butter, N.S. 1996. An improved technique for screening cotton germplasm against whitefly, *Bemisia tabaci*. Indian J. Entomol. 58(3): 210-214.
- 2480.Kular, J.S. and Butter, N.S. 1999. Influence of some morphological traits of cotton genotypes on resistance to whitefly, *Bemisia tabaci* Genn. J. Insect Sci. 12(1): 81-83.
- 2481.Kumar, N.K.K. and Moorthy, P.N.K. 2000. Transmission of yellow vein mosaic Geminivirus to imidacloprid treated okra by the whitefly, *Bemisia tabaci* Gennadius (Homoptera: Aleyrodidae). Insect Environ. 6(1): 46-47.
- 2482.Kumar, S. and Bhattacharya, A.K. 1989. Effect of plant density on the incidence of insects associated with soybean. Indian J. Entomol. 50(2): 185-192. [ Cock (1993)]
- 2483.Kumar, S. and Bhattacharya, A.K. 1989. Influence of different row spacings on the incidence of some insects of soybean. Indian J. Entomol. 51(1): 1-7. [ Cock (1993)]
- 2484.Kumar, S., Naresh, J.S. and Dahiya, B. 1998. Pest complex of a high yielding pigeonpea variety 'Manak' under Hisar conditions. Haryana Agric. Univ. J. Res. 28(1): 27-30.
- 2485.Kumar, S., Padam and Jain, C. 1992. Comparative bio-efficacy of various insecticides against white fly *Bemisia tabaci* Genn. on brinjal. Indian J. Entomol. 54(2): 124-131.
- 2486.Kumawat, K.C. and Kumawat, G.L. 1995. Effect of some insecticides on sucking pests and yellow mosaic incidence of moth bean, *Vigna aconitifolia* (Jacq.) Marechel. Indian J. Virol. 11(2): 69-72.
- 2487.Kumawat, R.L., Pareek, B.L. and Meena, B.L. 2000. Seasonal incidence of jassid and whitefly on okra and their correlation with abiotic factors. Ann. Biol. 16(2): 167-169.
- 2488.Kundu, G.G., Sekhar, J.C., Trimohan and Srivastava, K.P. 1995. A few soybean varieties highly resistant to yellow mosaic virus disease. Ann. Agric. Res. 16(4): 502-504.
- 2489.Kunik, T., Gafni, Y., Czosnek, H. and Citovsky, V. 1997. Transgenic tomato plants expressing TYLCV capsid protein are resistant to the virus: the role of the nuclear localization signal (NLS) in the resistance. Acta Hort. 447: 387-391.
- 2490.Kunik, T., Palanichelvam, K., Czosnek, H., Citovsky, V. and Gafni, Y. 1998. Nuclear import of the capsid protein of tomato yellow leaf curl virus (TYLCV) in plant and insect cells. Plant J. 13(3): 393-399.
- 2491.Kuno, E. 1977. Sequential estimation of population density by quadrat sampling. pp. 13-21. In M. Morisita (Ed.), Studies on Methods of Estimating Population Density, Biomass and Productivity in Terrestrial Animals. University of Tokyo Press, Tokyo. [Cock (1986)]

- 2492.Kurogi, S., Kuroki, F., Kawasaki, Y. and Nonaka, K. 1993. Studies on a fungus, *Beauveria bassiana*, isolated from *Thripsparmi* Karny. I. Pathogenicity to *Thripsparmi* and *Bemisia tabaci* and effect of pesticides on hyphal growth. Proc. Assoc. Plant Prot. Kyushu 39: 111-113. [ Japanese]
- 2493.Labanowski, G. 1994. Możliwości ograniczenia zabiegów chemicznych w ochronie roślin ozdobnych przed szkodnikami. [Possibilities of reduction of chemical treatments on ornamental crops against pests]. Materiały Sesji Instytutu Ochrony Roslin 34(1): 164-169. [ Polish, English summary]
- 2494.Labanowski, G.S. 1999. Occurrence and chemical control of introduced ornamental glasshouse pests in Poland. Bull. OEPP 29(1-2): 73-76.
- 2495.Labanowski, G.S. and Soika, G. 1999. Effectiveness of microbial and botanical insecticides in the control of *Bemisia tabaci* and *Frankliniella occidentalis* on ornamental plants. Bull. OEPP 29(1-2): 77-80.
- 2496.Laboucheix, J. 1973. Recherche et production cotonnières au Nicaragua. Bilan de cinq années de lutte phytosanitaire. Coton Fibres Trop. 28: 323-335. [ Cock (1986)]
- 2497.Lacasa, A., Sanchez, J.A., Gutierrez, L., Contreras, J., Guirao, P., Ros, C., Molina, J., Lorca, M. and Hita, I. 1996. Dinámica poblacional de *Bemisia tabaci* (Gennadius) y evolución de la incidencia del TYLCV en cultivos de tomate de Murcia. pp. 35-46. In J.L. Cenis (Ed.), El Virus del Rizado Amirillo (Hoja en Cuchara) del Tomate (TYLCV) y Su Vector *Bemisia tabaci*, Jornadas 8. Consejería de Medio Ambiente, Agricultura y Agua, Murcia, Spain.
- 2498.Lacey, L.A., Fransen, J.J. and Carruthers, R. 1996. Global distribution of naturally occurring fungi of *Bemisia*, their biologies and use as biological control agents. pp. 401-433. In D. Gerling and R.T. Mayer (Eds.), *Bemisia* : 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
- 2499.Lacey, L.A. and Kirk, A.A. 1993. Foreign exploration for natural enemies of *Bemisia tabaci* and implementation in integrated control programs in the United States. pp. 351-360. In Third International Conference on Pests in Agriculture, Montpellier 7-9 December 1993. Association Nationale de Protection des Plantes
- 2500.Lacey, L.A., Kirk, A.A., Millar, L., Mercadier, G. and Vidal, C. 1999. Ovicidal and larvicidal activity of conidia and blastospores of *Paecilomyces fumosoroseus* (Deuteromycotina: Hyphomycetes) against *Bemisia argentifolii* (Homoptera: Aleyrodidae) with a description of a bioassay system allowing prolonged survival of control insects. Biocontrol Sci. Technol. 9(1): 9-18.
- 2501.Lacey, L.A., Millar, L., Kirk, A.A. and Perring, T.M. 1999. Effect of storage temperature and duration on survival of eggs and nymphs of *Bemisia argentifolii* (Homoptera : Aleyrodidae) and pupae of the whitefly parasitoid *Encarsia formosa* (Hymenoptera : Aphelinidae). Ann. Entomol. Soc. Am. 92(3): 430-434.
- 2502.Laemmlen, F.F. 1982. The whitefly -virus-host complex: a new dilemma for the desert valleys of California. California Plant Pathol. Coop. Ext., Univ. California No. 56, 4 pp.
- 2503.Laird, E.F., Jr. and Dickson, R.C. 1959. Insect transmission of the leaf-crumple virus of cotton. Phytopathology 49: 324-327. [ Cock (1986)]
- 2504.Lal, O.P. 1997. Okra variety, Parbhani Kranti, highly prone to leafhopper, *Amrasca biguttula biguttula* Ishida, creating epidemic conditions in the hills of Uttarkashi in Bhagirathi Valley in central Himalaya (India). J. Entomol. Res. (New Delhi) 21(1): 81-83.
- 2505.Lal, R. and Gupta, G.P. 1998. Impact of spraying schedules on sucking pests in cotton system. Indian J. Entomol. 60(3): 305-310.
- 2506.Lal, S., Kumar, R. and Ali, S. 2000. Studies on the insect-pest succession of sunflower in eastern Uttar Pradesh. Shashpa 7(1): 95-97.
- 2507.Lal, S.S. 1980. A note on *Prospaltella flava* Shafee, and aphelinid parasitoid of *Bemisia tabaci* (Gennadius). Madras Agric. J. 67: 557-558. [ Cock (1986)]
- 2508.Lal, S.S. 1981. An ecological study of the whitefly, *Bemisia tabaci* (Genn.) population on cassava *Manihot esculenta* Crantz. Pestology 5(1): 11-17. [ Cock (1986)]
- 2509.Lal, S.S. 1985. A review of insect pests of mungbean and their control in India. Trop. Pest Manage. 31(2): 105-114. [ Cock (1993)]
- 2510.Lal, S.S. and Hrish, N. 1981. Note on the relative resistance of high-yielding cassava cultivars to infestations by tetranychid spider mites and whitefly. Indian J. Agric. Sci. 51: 536-538. [ Cock (1986)]
- 2511.Lal, S.S. and Pillai, K.S. 1981. Cassava pests and their control in southern India. Trop. Pest Manage. 27: 480-491. [ Cock (1986)]
- 2512.Lal, S.S. and Pillai, K.S. 1982. Ecological studies on whitefly, *Bemisia tabaci* (Genn.) infesting cassava in Kerala. Entomol. 7: 101-102. [ Cock (1986)]
- 2513.Lambert, A.L., McPherson, R.M. and Espelie, K.E. 1995. Soybean host plant resistance mechanisms that alter abundance of whiteflies (Homoptera: Aleyrodidae). Environ. Entomol. 24(6): 1381-1386.
- 2514.Lambert, A.L., McPherson, R.M. and Herzog, G.A. 1997. Field evaluation of fourteen soybean genotypes for resistance to whitefly (Homoptera: Aleyrodidae) infestations. J. Econ. Entomol. 90(2): 658-662.
- 2515.Lambert, A.L., McPherson, R.M. and Sparks, B. 1995. Evaluation of selected soybean genotypes for resistance to two whitefly species (Homoptera: Aleyrodidae) in the greenhouse. J. Entomol. Sci. 30(4): 519-526.
- 2516.Lamberts, M., Swanson, S. and Nesheim, O.N. 1995. Use of endosulfan on vegetable crops in Dade and Collier counties [Florida] from 1989-90 to 1994-95. Proc. Florida State Hort. Soc. 108: 248-251.
- 2517.Lana, A.F. and Wilson, G.F. 1976. A new viruslike disease of tomato in Nigeria. Plant Dis. Rep. 60: 296-298. [ Cock (1986)]
- 2518.Lana, A.O. and Taylor, T.A. 1976. The insect transmission of an isolate of okra mosaic virus occurring in Nigeria. Ann. Appl. Biol. 82: 361-364. [ Cock (1986)]
- 2519.Landa, Z., Osborne, L., Lopez, F. and Eyal, J. 1994. A bioassay for determining pathogenicity of entomogenous fungi on whiteflies. Biol. Control 4(4): 341-350.
- 2520.Landa Z., Osborne, L.S. and Eyal, J. 1994. Standard in vivo bioassay to assess entomogenous fungi on whiteflies. Sbornik Jihoceska Univerzita Zemedelska Fakulta Ceske Budejovice Fytotechnicka Rada 11(2): 3-14.
- 2521.Lapidot, M., Friedmann, M., Lachman, O., Yehezkel, A., Nahon, S., Cohen, S. and Pilowsky, M. 1997. Comparison of resistance level to tomato yellow leaf curl virus among commercial cultivars and breeding lines. Plant Dis. 81(12): 1425-1428.
- 2522.Lapidot, M., Friedmann, M., Pilowsky, M., Ben-Joseph, R. and Cohen, S. 2001. Effect of host plant resistance to Tomato yellow leaf curl virus (TYLCV) on virus acquisition and transmission by its whitefly vector. Phytopathology 91(12): 1209-1213.
- 2523.Larew, H.G. 1990. Activity of neem seed oil against greenhouse pests. pp. 128-131. In Paper presented at the USDA Neem Workshop, April 16-17, 1990, Beltsville, Maryland. U.S. Dept. Agric., Agric. Res. Serv. No. 86.
- 2524.Larew, H.G. and Lock, J.C. 1990. Repellency and toxicity of a horticultural oil against whiteflies on chrysanthemum. HortScience 25(11): 1406-1407.
- 2525.Larios, J.F. 1979. Niveles críticos de insectos que transmiten fitopatogenos: el caso de mosca blanca (*Bemisia tabaci* Genn.). Turrialba 29: 237-241. [ Cock (1993)]
- 2526.Lastra, J.R. and Uzcategui, R.C. 1975. Viruses affecting tomatoes in Venezuela. Phytopathol. Z. 84: 253-258. [ Cock (1986)]
- 2527.Lastra, R. 1992. Los geminivirus: un grupo de fitovirus con características especiales. [Geminivirus: A special type of plant viruses]. pp. 16-19. In L. Hilje and O. Arboleda (Eds.), Las moscas blancas (Homoptera: Aleyrodidae) en América Central y el Caribe. CATIE, Turrialba, Costa Rica. [Spanish]

- 2528.Lastra, R. and Gil, F. 1981. Ultrastructural host cell changes associated with tomato yellow mosaic. *Phytopathology* 71: 524-528. [Cock (1986)]
- 2529.Laterrot, H. 1990. An EEC programme to improve the resistance of the tomato to tomato yellow leaf curl virus. pp. 31-36. *In* EUCARPIA, Synop. XI Meet. Torremolinos, Malaga.
- 2530.Laterrot, H. 1992. Resistance genetics to tomato yellow leaf curl virus (TYLCV). *Tomato Yellow Leaf Curl Newsletter* 1: 2-4.
- 2531.Laterrot, H. 1993. Present state of the genetic control of tomato yellow leaf curl virus and of the EEC-supported breeding programme. pp. 19-24. *In* L. Stamova (Ed.), *Maritsa Vegetable Crops Research Institute, Plovdiv, Bulgaria*.
- 2532.Laterrot, H. 1995. Breeding network to create tomato varieties resistant to tomato yellow leaf curl virus (TLCV). *Fruits (Paris)* 50(6): 439-444. [English and French]
- 2533.Laterrot, H. 1996. Cultures de tomates en region mediterraneene pour le marche de frais. Lutte genetique: situation pratique et espoirs. [Tomato crops in the Mediterranean region for the fresh food market. Genetic control: practical situation and aspirations]. *Phytoma (France)* 484: 48-51. [French, English summary]
- 2534.Laterrot, H. and Cuartero, J. 1997. Control de patogenos en el Mediterraneo. 2a parte. [Control of pathogens in the Mediterranean region. 2nd part]. *Horticultura, Revista de Hortalizas, Flores y Plantas Ornamentales* 122: 95-100. [Spanish]
- 2535.Latha, C.R.P., Sriramulu, M. and Babu, T.R. 1993. Evaluation of insecticides against insect pest complex of soybean *Glycine max* (L.) Merrill. *Pestic. Res. J.* 5(2): 212-214.
- 2536.Latha, P. and Gopalakrishnan, T.R. 1995. Seasonal variation in white fly population and its influence on the yellow vein mosaic and productivity in pumpkin (*Cucurbita moschata* Poir.). *J. Trop. Agric.* 33(2): 144-146.
- 2537.Latheef, M.A., Bouse, L.F. and Kirk, I.W. 1994. Efficacy of aerially-applied sprays for control of sweetpotato whitefly in cotton. pp. 891-893. *In* D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN*.
- 2538.Latheef, M.A., Carlton, J.B. and Kirk, I.W. 1996. Seasonal control of sweet potato whiteflies in cotton using aerial electrostatic charged sprays. pp. 1035-1036. *In* P. Dugger and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN*.
- 2539.Latheef, M.A. and Wolfenbarger, D. 1995. Response of a greenhouse strain of silverleaf whitefly to danitol plus orthene and capture plus orthene. pp. 860-863. *In* D.A. Richter and J. Armour (Eds.), *Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN*.
- 2540.Lauritsen, K. and Paulson, G.S. 1998. A microscopic examination of whitefly (Homoptera: Aleyrodidae) egg pedicel insertion into host plant tissues. *J. Pennsylvania Acad. Sci.* 71(3): 99-103.
- 2541.Lawson, D.S., Ngo, N. and Koenig, J.P. 2000. Comparison of aerial and ground applied thiamethoxam (Actara™ & Centric™) for control of cotton pests. pp. 1330-1333. *In* P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN*.
- 2542.Lazarevic, B.M. 1970. Effects of the number of sprayings and different chemicals in controlling cotton pests in the Sudan. *J. Econ. Entomol.* 63: 629-633. [Cock (1986)]
- 2543.Lazarowitz, S.G. 1987. The molecular characterization of geminiviruses. *Plant. Mol. Biol. Rep.* 4: 177-192.
- 2544.Lazarowitz, S.G. 1991. Molecular characterization of two bipartite geminiviruses causing squash leaf curl disease: role of viral replication and movement functions in determining host range. *Virology* 180: 70-80.
- 2545.Lazarowitz, S.G. 1992. Geminiviruses: genome structure and gene function. *Crit. Rev. Plant Sci.* 11: 327-349.
- 2546.Lazarowitz, S.G. and Lazdins, I.B. 1991. Infectivity and complete nucleotide sequence of the cloned genomic components of a bipartite squash leaf curl geminivirus with a broad host range phenotype. *Virology* 180: 58-69.
- 2547.Lazarowitz, S.G., Wu, L.C., Rogers, S.G. and Elmer, J.S. 1992. Sequence-specific interaction with the viral AL1 protein identifies a geminivirus DNA replication origin. *Plant Cell* 4: 799-809.
- 2548.Lecoq, H., Desbiez, C., Delecalle, B., Cohen, S. and Mansour, A. 2000. Cytological and molecular evidence that the whitefly-transmitted cucumber vein yellowing virus is a tentative member of the family Potyviridae. *J. Gen. Virol.* 81(9): 2289-2293.
- 2549.Ledson, T.M. and Thornback, N. 1994. Control of *Bemisia tabaci*, Gennadius (Sternorrhyncha: Aleyrodidae) in Ethiopian cotton by the insect growth regulator buprofezin. *Int. Pest Control* 36(2): 42-43.
- 2550.Lee, M.L., Ahn, S.B. and Cho, W.S. 2000. Morphological characteristics of *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae) and discrimination of their biotypes in Korea by DNA markers. *Korean J. Appl. Entomol.* 39(1): 5-12. [Korean, English summary]
- 2551.Lee, M.L. and De Barro, P.J. 2000. Characterization of different biotypes of *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae) in South Korea based on 16S ribosomal RNA sequences. *Korean J. Entomol.* 30(2): 125-130.
- 2552.Legal, J. 1961. Phytosanitary note on the cultivation of cotton in Morocco. Present state of pest infestation and the possibilities of control. *Phytopathologie* 10(1): 27-37. [Cock (1986), French]
- 2553.Legaspi, B.C., Jr., Legaspi, J.C., Carruthers, R.I., Goolsby, J., Hadman, J., Jones, W., Murden, D. and Wendel, L. 1997. Areawide population dynamics of silverleaf whitefly (Homoptera: Aleyrodidae) and its parasitoids in the lower Rio Grande Valley of Texas. *J. Entomol. Sci.* 32(4): 445-459.
- 2554.Legaspi-Diaz, F., Machain-Lillingston, M., Castro-Silva, E., Gamboa-Bernal, G. and Rodriguez-Martinez, J. 1998. Efectividad biologica de buprofezin contra "Mosquita blanca de la hoja plateada" *Bemisia argentifolii* (Homoptera: Aleyrodidae) en algodonero en el Valle de Mexicali, B.C. 1997. [Biological effectivity of buprofezin against white fly of silver leave *Bemisia argentifolii* (Homoptera: Aleyrodidae) in cotton at the Mexicali Valley, Baja California Norte. 1997]. pp. 282-286. *In* *Proceedings 33rd National Entomological Congress, Acapulco, Mexico, 24-27 May 1998. Sociedad Mexicana De Entomologia.* [Spanish]
- 2555.Legaspi, J.C. and Carruthers, R. 1995. Population dynamics of silverleaf whitefly *Bemisia argentifolii* on cotton in mixed crop fields. pp. 828-831. *In* D.A. Richter and J. Armour (Eds.), *Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN*.
- 2556.Legaspi, J.C., Carruthers, R.I. and Nordlund, D.A. 1994. Life history of *Chrysoperla rufilabris* (Neuroptera: Chrysopidae) provided sweetpotato whitefly *Bemisia tabaci* (Homoptera: Aleyrodidae) and other food. *Biol. Control* 4(2): 178-184.
- 2557.Legaspi, J.C., Ciomperlik, M.A. and Legaspi, B.C., Jr. 2001. Field cage evaluation of *Serangium parcesetosum* as a predator of citrus blackfly eggs. *Southwest. Entomol.* 26(2): 171-172.
- 2558.Legaspi, J.C., Correa, J.A., Carruthers, R.I., Legaspi, B.C., Jr. and Nordlund, D.A. 1996. Effect of short-term releases of *Chrysoperla rufilabris* (Neuroptera: Chrysopidae) against silverleaf whitefly (Homoptera: Aleyrodidae) in field cages. *J. Entomol. Sci.* 31(1): 102-111.
- 2559.Legaspi, J.C., Legaspi, B.C., Jr., Carruthers, R.I., Goolsby, J., Jones, W.A., Kirk, A.A., Moomaw, C., Poprawski, T.J., Ruiz, R.A., Talekar, N.S. and Vacek, D. 1996. Foreign exploration for natural enemies of *Bemisia tabaci* from southeast Asia. *Subtrop. Plant Sci.* 48: 43-48.
- 2560.Legaspi, J.C., Legaspi, B.C., Jr., Meagher, R.L., Jr. and Ciomperlik, M.A. 1996. Evaluation of *Serangium parcesetosum* (Coleoptera: Coccinellidae) as a biological control agent of the silverleaf whitefly (Homoptera: Aleyrodidae). *Environ. Entomol.* 25(6): 1421-1427.

2561. Legaspi, J.C., Nordlund, D.A. and Legaspi, B.C., Jr. 1996. Tri-trophic interactions and predation rates in *Chrysoperla* spp. attacking the silverleaf whitefly. *Southwest. Entomol.* 21(1): 33-42.
2562. Legg, J.P. 1994. *Bemisia tabaci*: the whitefly vector of cassava mosaic geminiviruses in Africa: an ecological perspective. *African Crop Sci. J.* 2(4): 437-448.
2563. Legg, J.P. 1996. Host-associated strains within Ugandan populations of the whitefly, *Bemisia tabaci* (Genn), (Hom., Aleyrodidae). *J. Appl. Entomol.* 120(9): 523-527.
2564. Legg, J.P. 1999. Emergence, spread and strategies for controlling the pandemic of cassava mosaic virus disease in east and central Africa. *Crop Prot.* 18(10): 627-637.
2565. Legg, J.P., Gibson, R.W. and Otim-nape, G.W. 1994. Genetic polymorphism amongst Ugandan populations of *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae), vector of African cassava mosaic geminivirus. *Trop. Sci.* 34(1): 73-81.
2566. Legg, J.P. and Ogwal, S. 1998. Changes in the incidence of African cassava mosaic virus disease and the abundance of its whitefly vector along south-north transects in Uganda. *J. Appl. Entomol.* 122(4): 169-178.
2567. Legg, J.P. and Raya, M.D. 1998. Survey of cassava virus diseases of Tanzania. *Int. J. Pest Manage.* 44(1): 17-23.
2568. Legg, J.P., Sseruwgi, P., Kamau, J., Ajanga, S., Seremiah, S.C., Aritua, V., Otim-Nape, G.W., Muimba-Kankolongo, A., Gibson, R.W. and Thresh, J.M. 1999. The pandemic of severe cassava mosaic disease in East Africa: current status and future threats. pp. 236-251. *In* M.O. Akoroda and J.M. Teri (Eds.), *Proceedings Scientific Workshop of the Southern African Root Crops Research Network (SARRNET)*, Lusaka, Zambia, 17-19 August 1998.
2569. Legg, J.P. and Thresh, J.M. 2000. Cassava mosaic virus disease in East Africa: a dynamic disease in a changing environment. *Virus Res.* 71(1/2): 135-149.
2570. Leggett, J.E. 1993. Comparison of arthropods sampled from cultivars of upland and Pima cotton with drip and furrow irrigation. *Southwest. Entomol.* 18(1): 37-43.
2571. Leggett, J.E., Antilla, L. and Elhoff, L.K. 1995. Effectiveness of edge and late season full field treatments in reducing silverleaf whiteflies in central Arizona cotton fields. *Southwest. Entomol.* 20(4): 453-460.
2572. Lei, H., van Lenteren, J.C. and Tjallingii, W.F. 1999. Analysis of resistance in tomato and sweet pepper against the greenhouse whitefly using electrically monitored and visually observed probing and feeding behaviour. *Entomol. Exp. Appl.* 92(3): 299-309.
2573. Leigh, T.F., Wynholds, P.F. and Godfrey, L.D. 1994. Silverleaf whitefly: infestation development in relation to cotton genotype. pp. 865-866. *In* D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
2574. Leite, G.L.D., Picanco, M., Zanuncio, J.C. and Gonring, A.H.R. 1998. Effect of fertilization levels, age and canopy height of *Lycopersicon* spp. on attack rate of *Bemisia tabaci* (Homoptera: Aleyrodidae). *Agron. Lusitana* 46(2/4): 53-60.
2575. Lenfant, C., Ridray, G. and Schoen, L. 1998. Protection intégrée de la tomate de serre en région méditerranéenne. *PHM Rev. Hortic. (France)* 388: 34-38. [French]
2576. Lenfant, C., Ridray, G. and Schoen, L. 2000. Biopropagation of *Macrolophus caliginosus* Wagner for a quicker establishment in southern tomato greenhouses. *Bull. OILB/SROP (IOBC/WPRS)* 23(1): 247-251.
2577. Leon, A., Rodriguez, H., Terry, E. and Pino, M.A. 1997. Evaluacion de poblaciones de mosca blanca (*Bemisia tabaci* Guennadius) en variantes de policultivo tomate-maiz. [The evaluation of white fly (*Bemisia tabaci* Gennadius) populations in some tomato-maize polycultural treatments]. *Cultivos Tropicales* 18(2): 28-30. [Spanish, English summary]
2578. Leon-Lopez, R., Cervantes-Ramirez, M. and Sanchez-Damian, J.B. 1995. Estrategia para plagas agricolas de contingencia nacional: caso de mosca blanca *Bemisia argentifolii* en el Valle de Mexicali y la region de San Luis Rio Colorado. [Agricultural plagues of national contingence. Strategie: The case of *Bemisia argentifolii* en Mexicali Valley and San Luis Rio Colorado]. pp. 153-164. *In* M.M. Aguilera-Pena, P.R. Perez-Ayala and A. Lagunes-Tejeda (Eds.), *Tercera Asamblea Anual Del Conacofi.*, Chapingo, Mexico. Consejo Nacional Consultivo Fitosanitario. [Spanish]
2579. León-López, R.L. 1993. Mosquita blanca: daños e impacto en la agricultura regional. pp. 23-25. *In* Memoria 1st Reunión Regional sobre Problemas Fitosanitarios del Noroeste de México. IAP, A. C. [Spanish]
2580. Leppla, N.C., Delfosse, E.S. and Soper, R.S. 1996. Technical and regulatory constraints to international cooperation in biological control. pp. 385-395. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia : 1995 Taxonomy, Biology, Damage, Control and Management.* Intercept Ltd., Andover, Hants, UK.
2581. Leskovar, D.I. and Boales, A.K. 1996. Azadirachtin: potential use for controlling lepidopterous insects and increasing marketability of cabbage. *HortScience* 31(3): 405-409.
2582. Leuschner, K. 1978. Whiteflies: biology and transmission of African mosaic disease. pp. 51-58. *In* T. Brekelbaum, A. Bellotti and J.C. Lozano (Eds.), *Proc. Cassava Protection Workshop CIAT, Cali, Columbia, November 7-12, 1977.* Centro Internacional de Agricultura Tropical, Cali, Colombia. [Cock (1986)]
2583. Ley-Flores, J.H. 1975. Estudio de transmisión del virus del enchinamiento del tomate por mosquita blanca (*Bemisia tabaci*) (Genn.) en el Valle del Cullacán. [A study of transmission of the tomato leaf-curl virus by the whitefly *Bemisia tabaci* in the Culican Valley. Univ. Autónoma de Sinaloa, 36 pp. [CATIE (1992)]
2584. Lezhneva, I.P. and Anisimov, A.I. 1995. Predatory aphidophages for protection of greenhouse crops. *Zashchita Rastenii (Moskva)* 11: 39-40. [Russian]
2585. Li, A.Y.S., Dennehy, T.J., Li, S.X.H., Wigert, M.E., Zaborae, M. and Nichols, R.L. 2001. Sustaining Arizona's fragile success in whitefly resistance management. pp. 1108-1114. *In* P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
2586. Li, T., Vinson, S.B. and Gerling, D. 1989. Courtship and mating behavior of *Bemisia tabaci* (Homoptera: Aleyrodidae). *Environ. Entomol.* 18(5): 800-806.
2587. Li, Y., Dennehy, T.J., Li, X. and Wigert, M.E. 2000. Susceptibility of Arizona whiteflies to chloronicotynyl insecticides and IGR's: New developments in the 1999 season. pp. 296-306. *In* Cotton, A College of Agriculture Report, Series P-121. University of Arizona, Tucson.
2588. Li, Y., Dennehy, T.J., Li, X. and Wigert, M.E. 2000. Susceptibility of Arizona whiteflies to chloronicotynyl insecticides and IGR's: New developments in the 1999 season. pp. 1325-1330. *In* P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
2589. Liao, C.H., Chien, I.C., Chung, M.L., Chiu, R.J. and Han, Y.H. 1979. A study of sweet potato virus disease in Taiwan. I. Sweet potato yellow spot virus disease. *J. Agric. Res. (China)* 28(3): 127-137. [Cock (1986), Chinese, English summary]
2590. Lican, R. and Seljak, G. 1999. Tobacco whitefly [*Bemisia tabaci* (Gennadius)], a serious pest in glasshouses. pp. 221-229. *In* Zbornik Predavanj in Referatov 4. Slovenskega Posvetovanja O Varstvu Rastlin V Portorožu Od 3. Do 4. Marca 1999. Društvo Za Varstvo Rastlin Slovenije, Ljubljana, Slovenia. [Slovenian, English summary]
2591. Liedl, B.E., Lawson, D.M., White, K.K., Shapiro, J.A., Cohen, D.E., Carson, W.G., Trumble, J.T. and Matschler, M.A. 1995. Acylsugars of wild tomato *Lycopersicon pennelli* alters settling and reduces oviposition of *Bemisia argentifolii* (Homoptera: Aleyrodidae). *J. Econ. Entomol.* 88: 742-748.

- 2592.Lima, L.H.C., Campos, L., Moretzsohn, M.C., Návia, D., Silva, O.L.R. and Oliveira, M.R.V. 1999. II. Populações do complexo *Bemisia tabaci* (Gennadius) através de RAPD-PCR análise da diversidade genética. Pesquisa em Andamento, Empresa Brasileira de Pesquisa Agropecuária, EMBRAPA No. 22, 5pp. 1-5. [Portuguese]
- 2593.Lima, L.H.C., Navia, D., Inglis, P.W. and Oliveira, M.R.V. 2000. Survey of *Bemisia tabaci* (Gennadius) (Hemiptera: Aleyrodidae) biotypes in Brazil using RAPD markers. Genet. Mol. Biol. 23(4): 781-785.
- 2594.Lin, F.C., Su, T.H. and Wang, C.L. 1997. Effect of temperature on the development and reproduction of silverleaf whitefly (*Bemisia argentifolii* Bellows and Perring) and its population fluctuation on poinsettia. Chinese J. Entomol. 17(2): 66-79. [Chinese]
- 2595.Lin, T.B., Schwartz, A. and Saranga, Y. 1999. Effect of silverleaf whitefly on cotton photosynthesis. pp. 586-587. In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 2596.Lin, T.B., Schwartz, A. and Saranga, Y. 1999. Photosynthesis and productivity of cotton under silverleaf whitefly stress. Crop Sci. 39(1): 174-184.
- 2597.Lin, T.B., Schwartz, L. and Saranga, Y. 1999. Non-stomatal factors limit cotton photosynthesis under silverleaf whitefly stress. Physiol. Plantarum 107(3): 303-311.
- 2598.Lin, T.B., Wolf, S., Schwartz, A. and Saranga, Y. 2000. Silverleaf whitefly stress impairs sugar export from cotton source leaves. Physiol. Plantarum 109(3): 291-297.
- 2599.Lindley, C.D. 1970. Control of some pests of *Citrus*, date palm, wheat and cotton with ULV sprays. pp. 213-220. In British Crop Protection Council, London, UK. [Cock (1986)]
- 2600.Lindley, C.D. 1971. Control of pests of cotton, rice and maize with EI 47470. pp. 492-501. In Proceedings of the Sixth British Insecticide and Fungicide Conference, 15th to 18th November, 1971, Hotel Metropole, Brighton, England. [Cock (1986)]
- 2601.Lindquist, R.K. and Casey, M.L. 1991. Evaluation of conventional and biorational pesticides for sweetpotato and greenhouse whitefly control on poinsettia. Ohio Florists' Assoc. Bull. 74(1): 11-14. [Cock (1993)]
- 2602.Lindquist, R.K., Casey, M.L., Moore, T. and Taylor, R.A.J. 1996. Control of silverleaf whitefly on poinsettia, 1995. Arthropod Management Tests 21: 358-359.
- 2603.Link, D., Alvarez F. A. and Concatto, L.C. 1979. Plantas hospedeiras da mosca branca *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae), em Santa Maria, RS. Rev. Centro Ciencias Rurais 9(1): 55-59. [Cock (1986)]
- 2604.Link, D. and Costa, E.C. 1980. Ocorrência de inimigos naturais da mosca branca, *Bemisia tabaci* (Gennadius, 1889), na cultura da soja. Rev. Centro Ciencias Rurais 10(2): 111-113. [Cock (1986), Portuguese, English summary]
- 2605.Link, D. and Costa, E.C. 1982. Preferencia varietal da mosca-branca, *Bemisia tabaci* (Gennadius 1889) em soja. [Varietal preference of the White-fly, *Bemisia tabaci* (Gennadius 1889) in soybean]. pp. 158-163. In Annals 2nd National Seminar of Soybean Research, Brasília, DF, Brazil, 16 Feb 1981. [Portuguese, English summary]
- 2606.Link, D., Costa, E.C., Carvalho, S., Tarrago, M.F.S. and Veiga, P. 1979. Infestação da mosca branca, *Bemisia tabaci* em soja. [Infestation of the white fly, *Bemisia tabaci*, on soybean]. pp. 23-27. In Annals of the 1. National Seminar of Soybean Research, Londrina, PR, Brazil, 24 Sep 1978. [Portuguese, English summary]
- 2607.Linss, H. and Richter, M. 1994. White flies on poinsettias. Testing susceptibility of cultivars with red bracts. Gärtnerbörse und Gartenwelt (Germany) 94(46): 2273-2275. [German]
- 2608.Lipa, J.J. 1999. Problems of plant quarantine discussed at the 14th International Plant Protection Congress. Ochrona Roslin 43: 43. [Polish]
- 2609.Lipes, J.E. 1966. Outbreaks and new records. FAO Plant Prot. Bull. 14(1): 24. [Cock (1986)]
- 2610.Lipes, J.E. 1968. Outbreaks and new records. FAO Plant Prot. Bull. 16(2): 32. [Cock (1986)]
- 2611.Litsinger, J.A. and Apostol, R.F. 1994. Control of foliar insect pests on eggplant with systemic granular insecticides. Philippine Entomol. 9(3): 268-301.
- 2612.Liu, H.Y. 1993. Whitefly -borne viruses and virus epidemiology in Western USA. [Chinese summary]. Plant Pathol. Bull. 2(4): 187-194.
- 2613.Liu, H.Y., Cohen, S. and Duffus, J.E. 1992. The use of isozyme patterns to distinguish sweetpotato whitefly *Bemisia tabaci* biotypes. Phytoparasitica 20(3): 187-194.
- 2614.Liu, H.Y., Duffus, J.E. and Cohen, S. 1992. Isozyme patterns as a tool to monitor changes in *Bemisia [tabaci]* populations. pp. 78-79. In I.C. Rumbos, P. Kyriakopoulou and F. Bem (Eds.), Recent Advances in Vegetable Virus Research. 7th Conference ISHS Vegetable Virus Working Group, Athens, Greece, July 12-16, 1992. Ores Publishing, Volos, Greece.
- 2615.Liu, H.Y., Wisler, G.C. and Duffus, J.E. 2000. Particle lengths of whitefly-transmitted criniviruses. Plant Dis. 84(7): 803-805.
- 2616.Liu, S., Bedford, I.D., Briddon, R.W. and Markham, P.G. 1997. Efficient whitefly transmission of African cassava mosaic geminivirus requires sequences from both genomic components. J. Gen. Virol. 78(7): 1791-1794.
- 2617.Liu, S.J., Briddon, R.W., Bedford, I.D., Pinner, M.S. and Markham, P.G. 1999. Identification of genes directly and indirectly involved in the insect transmission of African cassava mosaic geminivirus by *Bemisia tabaci*. Virus Genes 18(1): 5-11.
- 2618.Liu, T.X. 1992. Ecological and biological studies of *Trialeurodes vaporariorum* and *Bemisia tabaci* on some greenhouse-grown ornamental plants. [University of Georgia]. Dissertation Abst. Int. 53(07B): 3276.
- 2619.Liu, T.X. 1999. Control of *Bemisia argentifolii* nymphs with the entomopathogen *Beauveria bassiana* under greenhouse and laboratory conditions, 1997-1998. Arthropod Management Tests 24: 403-404.
- 2620.Liu, T.X. 2000. Effects of thiamethoxam and imidacloprid on silverleaf whitefly on cantaloupe in spring 1999. Arthropod Management Tests 25: 20.
- 2621.Liu, T.X. 2000. Population dynamics of *Bemisia argentifolii* (Homoptera : Aleyrodidae) on spring collard and relationship to yield in the Lower Rio Grande Valley of Texas. J. Econ. Entomol. 93(3): 750-756.
- 2622.Liu, T.X. and Chen, T.Y. 2000. Effects of the chitin synthesis inhibitor buprofezin on survival and development of immatures of *Chrysoperla rufilabris* (Neuroptera : Chrysopidae). J. Econ. Entomol. 93(2): 234-239.
- 2623.Liu, T.X., Conner, J.M. and Stansly, P.A. 1997. Use of *Beauveria bassiana*, an entomopathogen, to control nymphs of *Bemisia argentifolii* under field conditions, 1995. Arthropod Management Tests 22: 133.
- 2624.Liu, T.X. and Oetting, R.D. 1993. Morphological comparisons of three species of whiteflies (Homoptera: Aleyrodidae) found on greenhouse-grown plants. Georgia Agric. Exp. Stn. Res. Bull. 412: 1-11.
- 2625.Liu, T.X. and Oetting, R.D. 1994. Oviposition preference of *Bemisia tabaci* (Gennadius) on eight species of greenhouse-grown plants. J. Agric. Entomol. 11(2): 177-179.
- 2626.Liu, T.X., Oetting, R.D. and Buntin, G.D. 1993. Distribution of *Trialeurodes vaporariorum* and *Bemisia tabaci* (Homoptera, Aleyrodidae) on some greenhouse-grown ornamental plants. J. Entomol. Sci. 28: 102-112.
- 2627.Liu, T.X., Oetting, R.D. and Buntin, G.D. 1993. Population dynamics and distribution of *Trialeurodes vaporariorum* and *Bemisia tabaci* on poinsettia following applications of three chemical insecticides. J. Entomol. Sci. 28: 126-135.

- 2628.Liu, T.X., Oetting, R.D. and Buntin, G.D. 1994. Evidence of interspecific competition between *Trialeurodes vaporariorum* and *Bemisia tabaci* (Gennadius) (Homoptera, Aleyrodidae) on some greenhouse-grown plants. *J. Entomol. Sci.* 29(1): 55-65.
- 2629.Liu, T.X., Oetting, R.D. and Buntin, G.D. 1994. Temperature and diel catches of *Trialeurodes vaporariorum* and *Bemisia tabaci* (Homoptera, Aleyrodidae) adults on sticky traps in the greenhouse. *J. Entomol. Sci.* 29(2): 222-230.
- 2630.Liu, T.X., Sparks, A.N., Jr., Liang, G.M. and Greenberg, S.M. 2001. Effects of defoliant alone and in combination with insecticides on boll weevil and whiteflies in cotton: C. Effects on silverleaf whitefly. pp. 980-984. *In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
- 2631.Liu, T.X. and Stansly, P.A. 1994. Leaf-dip contact toxicity bioassays of biorational insecticides to sweetpotato whitefly immature stages, 1993. *Arthropod Management Tests* 19: 374.
- 2632.Liu, T.X. and Stansly, P.A. 1994. Leaf-dip contact toxicity bioassays of surfactant materials to sweetpotato whitefly nymphs, 1993. *Arthropod Management Tests* 19: 373-374.
- 2633.Liu, T.X. and Stansly, P.A. 1994. Leaf-dip dry residual toxicity bioassays of biorational insecticides to sweetpotato whitefly adults, 1993. *Arthropod Management Tests* 19: 375.
- 2634.Liu, T.X. and Stansly, P.A. 1995. Deposition and bioassay of insecticides applied by leaf dip and spray tower against *Bemisia argentifolii* nymphs (Homoptera: Aleyrodidae). *Pestic. Sci.* 44(4): 317-322.
- 2635.Liu, T.X. and Stansly, P.A. 1995. Oviposition by *Bemisia argentifolii* (Homoptera: Aleyrodidae) on tomato: Effects of leaf factors and insecticide residues. *J. Econ. Entomol.* 88(4): 992-997.
- 2636.Liu, T.X. and Stansly, P.A. 1995. Toxicity and repellency of some biorational insecticides to *Bemisia argentifolii* on tomato plants. *Entomol. Exp. Appl.* 74(2): 137-143.
- 2637.Liu, T.X. and Stansly, P.A. 1995. Toxicity of Admire to second and third instar silverleaf whitefly nymphs on tomato leaves, 1994. *Arthropod Management Tests* 20: 360-361.
- 2638.Liu, T.X. and Stansly, P.A. 1995. Toxicity of biorational insecticides to *Bemisia argentifolii* (Homoptera: Aleyrodidae) on tomato leaves. *J. Econ. Entomol.* 88(3): 564-568.
- 2639.Liu, T.X. and Stansly, P.A. 1996. Morphology of *Nephaspis oculatus* and *Delphastus pusillus* (Coleoptera: Coccinellidae), predators of *Bemisia argentifolii* (Homoptera: Aleyrodidae). *Proc. Entomol. Soc. Washington* 98(2): 292-300.
- 2640.Liu, T.X. and Stansly, P.A. 1996. Oviposition, development, and survivorship of *Encarsia pergandiella* (Hymenoptera: Aphelinidae) in four instars of *Bemisia argentifolii* (Homoptera: Aleyrodidae). *Ann. Entomol. Soc. Am.* 89(1): 96-102.
- 2641.Liu, T.X. and Stansly, P.A. 1996. Pupal orientation and emergence of some aphelinid parasitoids (Hymenoptera: Aphelinidae) of *Bemisia argentifolii* (Homoptera: Aleyrodidae). *Ann. Entomol. Soc. Am.* 89(3): 385-390.
- 2642.Liu, T.X. and Stansly, P.A. 1996. Toxicological effects of selected insecticides on *Nephaspis oculatus* (Col. Coccinellidae), a predator of *Bemisia argentifolii* Hem, Aleyrodidae). *J. Appl. Entomol.* 120(6): 369-373.
- 2643.Liu, T.X. and Stansly, P.A. 1997. Effects of pyriproxyfen on three species of *Encarsia* (Hymenoptera: Aphelinidae), endoparasitoids of *Bemisia argentifolii* (Homoptera: Aleyrodidae). *J. Econ. Entomol.* 90(2): 404-411.
- 2644.Liu, T.X. and Stansly, P.A. 1997. Insecticidal effects of vegetable oil on silverleaf whitefly, 1996. *Arthropod Management Tests* 22: 414.
- 2645.Liu, T.X. and Stansly, P.A. 1998. Life history of *Bemisia argentifolii* (Homoptera: Aleyrodidae) on *Hibiscus rosa-sinensis* (Malvaceae). *Florida Entomol.* 3: 437-445.
- 2646.Liu, T.X. and Stansly, P. A. 1999. Searching and feeding behavior of *Nephaspis oculatus* and *Delphastus catalinae* (Coleoptera: Coccinellidae), predators of *Bemisia argentifolii* (Homoptera: Aleyrodidae). *Environ. Entomol.* 28(5): 901-906.
- 2647.Liu, T.X. and Stansly, P.A. 2000. Insecticidal activity of surfactants and oils against silverleaf whitefly (*Bemisia argentifolii*) nymphs (Homoptera : Aleyrodidae) on collards and tomato. *Pest Manag. Sci.* 56(10): 861-866.
- 2648.Liu, T.X., Stansly, P.A. and Chortyk, O.T. 1996. Insecticidal activity of natural and synthetic sugar esters against *Bemisia argentifolii* (Homoptera: Aleyrodidae). *J. Econ. Entomol.* 89(5): 1233-1239.
- 2649.Liu, T.X., Stansly, P.A., Hoelmer, K.A. and Osborne, L.S. 1997. Life history of *Nephaspis oculatus* (Coleoptera: Coccinellidae), a predator of *Bemisia argentifolii* (Homoptera: Aleyrodidae). *Ann. Entomol. Soc. Am.* 90(6): 776-782.
- 2650.Liu, T.X., Stansly, P.A., Sparks, A.N., Jr., Knowles, T.C. and Chu, C.C. 1999. Application of Mycotrol and Naturalis-L (*Beauveria bassiana*) for management of *Bemisia argentifolii* (Homoptera: Aleyrodidae) on vegetables, cotton and ornamentals in Southern United States. *Subtrop. Plant Sci.* 51: 41-48.
- 2651.Liu, Y., Robinson, D.J. and Harrison, B.D. 1998. Defective forms of cotton leaf curl virus DNA -A that have different combinations of sequence deletion, duplication, inversion and rearrangement. *J. Gen. Virol.* 79(6): 1501-1508.
- 2652.Livieratos, I.C., Avgelis, A.D. and Coutts, R.H.A. 1999. Molecular characterization of the cucurbit yellow stunting disorder virus coat protein gene. *Phytopathology* 89(11): 1050-1055.
- 2653.Livieratos, I.C., Katis, N. and Coutts, R.H.A. 1998. Differentiation between cucurbit yellow stunting disorder virus and beet pseudo-yellows virus by a reverse transcription-polymerase chain reaction assay. *Plant. Pathol.* 47(3): 362-369.
- 2654.Livingston, S.D., Cowan, B.F. and Norman, J.W. 1992. An evaluation of sixteen commercial picker cotton varieties for performance factors and feeding damage sustained under heavy whitefly pressure in the Lower Rio Grande Valley. pp. 557-559. *In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
- 2655.Llano, A. and Vanegas, J.A. 2000. Historia y situacion de la produccion de frijol en los paises latinoamericanos afectados por geminivirus transmitidos por mosca blanca: America Central: Nicaragua. pp. 37-40. *In Bean Golden Mosaic and Other Diseases of Common Bean Caused by Whitefly -Transmitted Geminiviruses in Latin America. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. [Spanish]*
- 2656.Lobo-Lima, M.L. and Klein-Koch, C. 1981. Schwerpunkte in der integrierten Bekämpfung von Schadarthropoden auf den Kapverdischen Inseln (W-Afrika). *Mitteilungen der Deutschen Gesellschaft für Allgemeine und Angewandte Entomol.* 3: 60-65. [ Cock (1986), German, English summary]
- 2657.Loebenstein, G. and Harpaz, I. 1960. Virus diseases of sweet potatoes in Israel. *Pathology* 50: 100-104. [ Cock (1986)]
- 2658.Loera-Gallardo, J. and Kokubu, H. 1998. Cria, liberacion y capacidad depredadora de *Hippodamia convergens* Guerin-Meneville (Coleoptera: Coccinellidae). [Rearing, release and depredation capacity of *Hippodamia convergens* Guerin-Meneville (Coleoptera: Coccinellidae)]. pp. 332-335. *In Proceedings 21st National Congress of Biological Control, Rio Bravo, Tampaulipas, Mexico, 5-6 Nov 1998. INIFAP. [Spanish]*
- 2659.Loera-Gallardo, J., Wolfenbarger, D.A. and Riley, D.G. 1998. Insecticidal mixture interactions against B-strain sweetpotato whitefly (Homoptera: Aleyrodidae). *J. Entomol. Sci.* 33(4): 407-411.
- 2660.Loginova, E. 1993. Sweetpotato white fly threaten to attack Bulgaria. *Rastitelna Zashchita* 34(8): 22-23. [ Bulgarian]
- 2661.Lohar, M.K. 1987. Field evaluation of insecticides against jassid (*Amrasca devastans* D.) and white fly (*Bemisia tabaci* G.) on sunflower crop. *Sarhad J. Agric. (Pakistan)* 3(2): 215-200.

- 2662.Lohrer, T. 1994. Plant protection in poinsettias. Gärtnerbörse und Gartenwelt (Germany) 94(43): 2134-2138. [ German]
- 2663.Loomans, A.J.M. and van Lenteren, J.C. 1999. Evaluating environmental effects of *Encarsia* species (Hymenoptera: Aphelinidae) introduced for whitefly control into Europe. Bull. OILB/SROP (IOBC/WPRS) 22: 153-156.
- 2664.Lopez-Avila, A. 1986. Natural enemies. pp. 27-35. In M.J.W. Cock (Ed.), *Bemisia tabaci* - A Literature Survey on the Cotton Whitefly with an Annotated Bibliography. CAB International Institute of Biological Control, Ascot, UK. [Cock (1993)]
- 2665.Lopez-Avila, A. 1986. Taxonomy and biology. pp. 3-11. In M.J.W. Cock (Ed.), *Bemisia tabaci* - A Literature Survey on the Cotton Whitefly with an Annotated Bibliography. CAB International Institute of Biological Control
- 2666.Lopez-Avila, A. 1987. Two new species of *Encarsia* Foerster (Hymenoptera: Aphelinidae) from Pakistan, associated with the cotton whitefly, *Bemisia tabaci* (Hemiptera: Aleyrodidae). Bull. Entomol. Res. 77: 425-438.
- 2667.Lopez, B. 1996. Susceptibilidad de variedades comerciales de frijol, al ataque de la osputita blanca (*Bemisia* spp.) en el Valle de Fuate, Sinaloa. Memoria Científica 2: 69-70
- 2668.Lopez, M. 1998. The whitefly (*Bemisia* spp.) and its natural enemies. [Spanish, English summary]. Rev. Proteccion Vegetal 13(2): 75-79.
- 2669.Lopez, M.V. 1999. Growth, yield and leaf NPK concentrations in crop-covered squash. J. Sustain. Agric. 12(4): 25-38.
- 2670.Lopez, R.L. and Rivera, I.R.T. 1997. Rescate 200 (acetamiprid) a new alternative for silverleaf whitefly control in the Mexicali Valley, Mexico, 1996. p. 1063. In P. Dugger and D. Richter (Eds.), National Cotton Council, Memphis, TN.
- 2671.Lopez-Salinas, E. and Becerra-Leor, E.N. 2000. Historia y situacion de la produccion de frijol en los paises latinoamericanos afect ados por geminivirus transmitidos por mosca blanca: Mexico: Mexico-Sur. pp. 19-23. In Bean Golden Mosaic and Other Diseases of Common Bean Caused by Whitefly-Transmitted Geminiviruses in Latin America. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. [Spanish]
- 2672.Lopez-Sese, A.I. and Gomez-Guillamon, M.L. 2000. Resistance to cucurbit yellowing stunting disorder virus (CYSDV) in *Cucumis melo* L. Hortscience 35(1): 110-113.
- 2673.Lot, H., Delecolle, B. and Lecoq, H. 1983. A whitefly transmitted virus causing muskmelon yellows in France. Acta Hortic. 127: 175-182. [ Cock (1986)]
- 2674.Lotrakul, P., Valverde, R.A., Clark, C.A., Sim, J. and De la Torre, R. 1998. Detection of a geminivirus infecting sweet potato in the United States. Plant Dis. 82(11): 1253-1257.
- 2675.Lotrakul, P., Valverde, R.A., De La Torre, R., Sim, J.G. and Gomez, A. 2000. Occurrence of a strain of Texas pepper virus in Tabasco and Habanero pepper in Costa Rica. Plant Dis. 84(2): 168-172.
- 2676.Lotrakul, P., Valverde, R.A. and Landry, A.D. 2000. Biolo gical and molecular properties of a begomovirus from *Dicliptera sexangularis*. Phytopathology 90(7): 723-729.
- 2677.Lourençao, A.L. and Miranda, M.A.C. 1987. [Soyabean resistance to insects: VIII. IAC 78-2318, a line with multiple resistance]. Bragantia 46(1): 65-72. [ Cock (1993), Portuguese]
- 2678.Lourençao, A.L., Miranda, M.A.C. and Alves, S.B. 2001. Epizootic occurrence of *Verticillium lecanii* on *Bemisia tabaci* B biotype (Hemiptera: Aleyrodidae) in the State of Maranhao, Brazil. Neotrop. Entomol. 30(1): 183-185. [ Portuguese, English summary]
- 2679.Lourençao, A.L. and Nagai, H. 1994. Outbreaks of *Bemisia tabaci* in the Sao Paulo State, Brazil. Bragantia 53(1): 53-59.
- 2680.Lourençao, A.L. and Yuki, V.A. 1982. Oviposicao de *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae) em tres variedades de soja sem chance de escolha. [With English summary]. Bragantia 41: 199-202. [ Cock (1986)]
- 2681.Lourençao, A.L., Yuki, V.A. and Alves, S.B. 1999. Epizootics of *Aschersonia* cf. *goldiana* on *Bemisia tabaci* (Homoptera: Aleyrodidae) biotype B in the State of Sao Paulo, Brazil. Anais Soc. Entomol. Brasil 28(2): 343-345. [ Portuguese, English summary]
- 2682.Lourens, J.H., van der Laan, P.A. and Brader, L. 1972. Contribution à l'étude d'une 'mosaïque' du cotonnier au Tchad; distribution dans un champ; Aleurodidae communs; essais de transmission de cotonnier à cotonnier par les Aleurodidae. Coton Fibres Trop. 27: 225-230. [ Cock (1986)]
- 2683.Louro, D., Accotto, G.P. and Vaira, A.M. 2000. Occurrence and diagnosis of Tomato chlorosis virus in Portugal. Eur. J. Plant Pathol. 106(6): 589-592.
- 2684.Louro, D. and Fernandes, J.M.E. 1999. Recent fitovirological problems associated with the whitefly *Bemisia tabaci* Genn. p. 102. In New Directions in Plant Protection, 2nd Biennial Meeting of the Portuguese Phytopathology Society, Oeiras, Portugal, 24-25 Sep 1998. Estacao Agronomica Nacional, Oeiras, Portugal.
- 2685.Louro, D., Vicente, M., Vaira, A.M., Accotto, G.P. and Nolasco, G. 2000. Cucurbit yellow stunting disorder virus (Genus *Crimivirus*) associated with the yellowing disease of cucurbit crops in Portugal. Plant Dis. 84(10): 1156.
- 2686.Loxdale, H.D. and Lushai, G. 1999. Slaves of the environment: the movement of herbivorous insects in relation to their ecology and genotype. Phil. Trans. R. Soc. (Lond. B.) 354: 1479-1495.
- 2687.Lublinkhof, J., Comer, D. and Moore, L. 1997. Performance of Applaud R (Buprofezin) against silverleaf whitefly in first year's commercial use in Arizona. p. 934. In P. Dugger and D. Richter (Eds.), National Cotton Council, Memphis, TN.
- 2688.Lublinkhof, J. and Odom, P.N. 1993. Control of whitefly with OVASYN. p. 951. In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 2689.Luft, P.A. and Paine, T.D. 1997. Evaluation of environmental and plant-associated cues for nymphal settling preference by *Trioza eugeniae*. Entomol. Exp. Appl. 85(2): 105-111.
- 2690.Luo, Z.Y., Zhang, W.N. and Gan, G.P. 1989. Population dynamics of tobacco whitefly in cotton fields and the influence of insecticide application. Acta Entomol. Sinica 32(3): 293-299. [ Chinese, English summary]
- 2691.Luo, Z.Y. and Zhou, C.M. 2001. Record of the citrus whiteflies in China. South China Fruits 30(1): 14-16. [ Chinese]
- 2692.Lutzinsky, U., Hama, M. and Roso, R. 1996. L'emploi de paillages plastiques colores pour reduire l' incidence du virus TYLCV en culture de tomate. [The use of coloured plastics to reduce the incidence of Tomato Yellow Leaf Curl Virus in market tomatoes]. Plasticulture (France) 112: 21-22. [ French, English summary]
- 2693.Lynch, R.E. and Simmons, A.M. 1993. Distribution of immatures and monitoring of adult sweetpotato whitefly, *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae), in peanut, *Arachis hypogaea*. Environ. Entomol. 22: 375-380.
- 2694.Mabbett, T. 1978. A revision of the economic insect pests of cotton in Thailand. Description, infestation and control. I. Cotton whitefly (*Bemisia tabaci* Gennadius). Rep. Cotton Pest Mngt. Project Dep. Agric., (Bangkok) 13: 1-12. [ Cock (1993)]
- 2695.Mabbett, T. 1983. Pin-point your cotton pests. Part 2: distribution on the plant and in the field. World Crops 35(4): 133-134. [ Cock (1986)]
- 2696.Mabbett, T., Nachapong, M. and Mekdaeng, J. 1980. The within-canopy distribution of adult cotton whitefly (*Bemisia tabaci* Gennadius) incorporating economic thresholds and meteorological conditions. Thai J. Agric. Sci. 13: 97-108. [ Cock (1993)]
- 2697.Mabbett, T.H. 1979. Guidelines for the management of cotton pests, Pt. 3: the cotton whitefly (*Bemisia tabaci* Gennadius). Ministry Agric. Cooperatives, Bangkok, Thailand, 6 pp.
- 2698.Mabbett, T.H. 1979. A review of the economic insect pests of cotton in Thailand. Description, infestation and control. Cotton Pests Res. Branch, Dept. Agric. (Bangkok, Thailand), 80 pp. [ Cock (1986)]

- 2699.Mabbett, T.H. 1980. Management of four major cotton pests in Thailand. *World Crops* 32(4): 101-104. [Cock (1986)]
- 2700.MacDonald, O. and Mitchell, R. 1998. Efficacy of propoxur smoke for the control of juvenile *Bemisia tabaci* (Gennadius) (Homoptera:Aleyrodidae). *Ann. Appl. Biol.* 132(Suppl. S): 4-5.
- 2701.MacDonald, O.C. and Cheek, S. 1994. Efficacy of methyl bromide fumigation as a quarantine treatment for *Bemisia tabaci* on poinsettia. *Med. Fac. Landbouww. Univ. Gent* 59(2B): 619-623.
- 2702.Machain-Lillingston, M., Legaspi-Diaz, F., Machado-Ibarra, S., Valdes-Castro, J.R. and Martinez-Vargas, A. 1998. Manejo integrado de insecticidas contra "Mosquita blanca" de la hoja plateada *Bemisia argentifolii* en algodonoero, con base en tipo de producto y umbrales de accion. Valle de Mexicali, B.C. 1997. [Insecticide integrated management against white fly of silver leave *Bemisia argentifolii* in cotton with base in the type of product and action threshold. Mexicali Valley, Baja California Norte State 1997]. pp. 293-296. *In Proceedings 33rd National Entomological Congress, Acapulco, Mexico, 24-27 May 1998. Sociedad Mexicana De Entomologia. [Spanish]*
- 2703.MacIntosh, S., Robinson, D.J. and Harrison, B.D. 1992. Detection of three whitefly-transmitted geminiviruses occurring in Europe by tests with heterologous monoclonal antibodies. *Ann. Appl. Biol.* 121: 297-303.
- 2704.Mahal, M.S. and Singh, B. 1981. Resistance to white fly -transmitted yellow vein mosaic virus in okra *Bemisia tabaci*. *Indian J. Ecol.* 8(2): 301-303. [CATIE (1992)]
- 2705.Mahanta, I.C., Dhal, A. and Mohanty, A.K. 1998. Field performance of tomato cultivars in relation to wilt and leaf curl diseases in North Central Plateau zone of Orissa. *Orissa J. Hortic.* 26(1): 34-39.
- 2706.Mahmood, T. 1999. Cotton leaf curl virus disease and its status in Pakistan. pp. 234-244. *In Proceedings ICAC-CCRI Regional Consultation, Insecticide Resistance Management in Cotton. Central Cotton Research Institute, Multan, Pakistan.*
- 2707.Mahto, D.N. and Sinha, D.C. 1978. Evaluation of insecticides for the control of white fly, *Bemisia tabaci* Genn. in relation to the incidence of mosaic of cassava. *Indian J. Entomol.* 40: 316-319. [Cock (1986)]
- 2708.Mahto, D.N. and Sinha, D.C. 1978. Mosaic disease of cassava and its relationship with the vector, *Bemisia tabaci* Genn. *Indian J. Entomol.* 40: 117-120. [Cock (1986)]
- 2709.Maienfisch, P., Gsell, L. and Rindlisbacher, A. 1999. Synthesis and insecticidal activity of CGA 293'343 - a novel broad-spectrum insecticide. *Pestic. Sci.* 55(3): 351-355.
- 2710.Maignet, P. and Onillon, J.C. 1997. Premieres donnees sur le potentiel biotique d'*Encarsia hispida* De Santis (Hymenop. : Aphelinidae, endoparasitoide du biotype 'B' de *Bemisia tabaci* (Gennadius) et de *Trialetrodes vaporariorum* West. (Homoptera: Aleyrodidae) . [Initial data on the biotic potential of *Encarsia hispida* De Santis (Hymenopt: Aphelinidae), endoparasitoid of biotype 'B' of *Bemisia tabaci* (Gennadius) and of *Trialetrodes vaporariorum* West. (Homoptera: Aleyrodidae)]. *Bull. OILB/SROP (IOBC/WPRS)* 20(4): 121-129. [French]
- 2711.Maisano, J.J., Jr. 1988. Poinsettia whitefly control. *Connecticut Greenhouse Newsl.* 146: 4-7.
- 2712.Maisonneuve, J.C. 1992. New glasshouse pests in Europe. *Bull. OEPP* 22(3): 331-335. [French, English summary]
- 2713.Makadey, M.A., Ibrahim, S.A. and Gergis, M.F. 1994. Whitefly, *Bemisia tabaci* (Gen.): developmental biology, sampling and management on cotton in Middle Egypt. pp. 1226-1228. *In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
- 2714.Makkouk, K.M. 1978. A study on tomato viruses in the Jordan Valley with special emphasis on tomato yellow leaf curl. *Plant Dis. Rep.* 62(3): 259-262. [Cock (1986)]
- 2715.Makkouk, K.M. and Laterrot, H. 1983. Epidemiology and control of tomato yellow leaf curl virus. pp. 315-321. *In R.T. Plumb and J.M. Thresh (Eds.), Plant Virus Epidemiology. The Spread and Control of Insect-borne Viruses. Blackwell Scientific Publications, Oxford, UK. [Cock (1986)]*
- 2716.Makkouk, K.M., Shebab, S. and Maadjalani, S.E. 1979. Tomato yellow leaf curl: incidence, yield losses, and transmission in Lebanon. *Phytopathology* : 263-267.
- 2717.Mali, V.R. 1977. Cotton leaf crumple virus disease - a new record for India. *Indian Phytopathol.* 30: 326-329.
- 2718.Malik, D.M., Saleem, A. and Khan, A.G. 1992. Cotton leaf curl virus a status report. *Ayub Agric. Res. Instit., Faisalabad*, 16 pp.
- 2719.Malipatil, M.V., Dashad, S.S., Sharma, P.D. and Chaudhary, O.P. 2001. Evaluation of different spray nozzles in controlling insect-pests of cotton in Haryana. *Crop Res. (Hisar)* 21(2): 219-224.
- 2720.Mallick, S.C. and Chowdhury, A.K. 1996. Virus diseases of some vegetable crops in Nadia. [West Bengal]. *J. Mycopathol. Res.* 34(1): 29-33.
- 2721.Mancia, J.E. 1974. Utilizacion de insecticidas sistemicos granulados en el control de mosca blanca *Bemisia tabaci* Genn., e infeccion virosa en frijol comun. [Utilization of granulated systemic insecticides in the control of whitefly *Bemisia tabaci* Genn., and virus infection in beans]. *Soc. Ingenieros Agron. (El Salvador)* 3(3): 77-81. [Spanish]
- 2722.Mandal, B., Langston, D.B., Jr., Kucharek, T.A., Pappu, H.R., Beard, G.H., Flanders, J.T., Whiddon, J.P., Smith, J.E. and Kelly, W.T. 2001. First report of cabbage leaf curl virus (Family Geminiviridae) in Georgia. *Plant Dis.* 85(5): 561.
- 2723.Mandal, B., Varma, A. and Malathi, V.G. 1997. Systemic infection of *Vigna mungo* using the cloned DNAs of the blackgram isolate of mungbean yellow mosaic geminivirus through agroinoculation and transmission of the progeny virus by whiteflies. *J. Phytopathol. (Berlin)* 145(11-12): 505-510.
- 2724.Manjunatha, B., Muniyappa, V. and Nateshan, H.M. 1995. Purification and transmission of dolichos yellow mosaic geminivirus. *Indian J. Mycol. Plant Pathol.* 25(1-2): 91-92.
- 2725.Mann, G.S., Dhaliwal, G.S. and Dhawan, A.K. 2001. Effect of alternate application of neem products and insecticides on population of *Bemisia tabaci* Gennadius and its impact on bollworm damage in upland cotton. *Ann. Plant Prot. Sci.* 9(1): 22-25.
- 2726.Mannaa, S.H. 2000. Monitoring of insecticide sensitivity change in different developmental stages of the cotton whitefly, *Bemisia tabaci* (Genn.) to certain insecticides. *Assiut J. Agric. Sci. (Egypt)* 31(2): 131-144.
- 2727.Manor, G., Hofner, A., Or, R., Phishler, G., Epstein, Y., Nakash, J. and Jacobi, M. 1987. Air sleeves field sprayer for cotton. pp. Paper No. 87-1609. *In Am. Soc. Agr. Eng. Microfische Collect. American Society of Agricultural Engineers, St. Joseph, MI.*
- 2728.Manor, G., Hofner, A., Or, R., Phishler, G., Epstein, Y., Nakash, J. and Jacobi, M. 1989. Air stream facilitated application of cotton foliage treatments. *Trans. Am. Soc. Agric. Eng.* 32(1): 37-40. [Cock (1993)]
- 2729.Mansoor, S., Amin, I., Hussain, M. and Zafar, Y. 2001. Association of a disease complex involving a begomovirus, DNA 1 and a. *Plant Dis.* 85(8): 922.
- 2730.Mansoor, S., Bashir, A., Khan, S.H., Hussain, M., Saeed, M., Zafar, Y., Markham, P.G. and Malik, K.A. 1999. Rapid multiplex PCR for the specific detection of two whitefly-transmitted geminivirus species associated with cotton leaf curl disease in Pakistan. *Pakistan J. Bot.* 31(1): 115-123.
- 2731.Mansoor, S., Bedford, I., Pinner, M.S., Stanley, J. and Markham, P.G. 1993. A whitefly-transmitted geminivirus associated with cotton leaf curl disease in Pakistan. *Pakistan J. Bot.* 25: 105-107.
- 2732.Mansoor, S., Khan, S.H., Bashir, A., Saeed, M., Zafar, Y., Malik, K.A., Briddon, R., Stanley, J. and Markham, P.G. 1999. Identification of a novel circular single-stranded DNA associated with cotton leaf curl disease in Pakistan. *Virology* 259(1): 190-199.



- 2733.Mansoor, S., Khan, S.H., Hussain, M., Bashir, A., Saeed, M., Zafar, Y., Stanley, J., Briddon, R., Markham, P. and Malik, K.A. 1999. DNA variants among Pakistan isolates of cotton leaf curl virus. pp. 261-276. *In* Proceedings ICAC-CCRI Regional Consultation, Insecticide Resistance Management in Cotton. Central Cotton Research Institute, Multan, Pakistan.
- 2734.Mansoor, S., Mukhta S., Hussain, M., Amin, I., Zafar, Y. and Malik, K.A. 2000. Widespread occurrence of cotton leaf curl virus on radish in Pakistan. *Plant Dis.* 84(7): 809.
- 2735.Mansour, A. and Al-Musa, A. 1992. Tomato yellow leaf curl virus: host range and virus-vector relationships. *Plant Pathol.* 41(2): 122-125. [Cock (1993)]
- 2736.Mansour, A. and Al-Musa, A. 1993. Cucumber vein yellowing virus: Host range and virus vector relationships. *J. Phytopathol.* 137: 73-78.
- 2737.Mansour, A., Al-Musa, A., Vetten, H.J. and Lesemann, D.E. 1998. Properties of a cowpea mild mottle virus (CPMMV) isolate from eggplant in Jordan and evidence for biological and serological differences between CPMMV isolates from leguminous and solanaceous hosts. *J. Phytopathol.* 146(11-12): 539-547.
- 2738.Mansour, M.M., Eissa, I.S. and Metwally, H.E. 1977. Abundance and seasonal fluctuation of *Bemisia tabaci* on different vegetable plants in three localities at Sharkia Governorate. *Egyptian Ann. Agric. Sci. (Moshtohor)* 7: 227-235.
- 2739.Mansour, M.M., Gouhar, K.A. and Guirguis, M.W. 1974. The susceptibility of five soya bean varieties to infestation with different pests at Zagazig Region, Egypt. *Bull. Entomol. Soc. Egypt* 58: 285-290. [Cock (1986)]
- 2740.Mantilla-Gonzalez, C.E. 1991. Evaluation of two bioassay methods for determining toxicity of selected insecticides to sweetpotato whitefly, *Bemisia tabaci*. [University of Florida]. Dissertation Abst. Int. 52(10B): 5076.
- 2741.Manzaroli, G. and Benuzzi, M. 1995. Pomodoro in serra, lotta biologica e integrata. [Protected tomato: biological and integrated control]. *Colture Protette (Italy)* 24(1): 41-47. [Italian]
- 2742.Manzaroli, G., Tommasini, M.G., Mosti, M. and Dradi, D. 1997. Biological control of whitefly on poinsettia in Italy. *Bull. OILB/SROP (IOBC/WPRS)* 20(4): 130-142.
- 2743.Marcano, R. and Gonzalez, E. 1993. Evaluacion de insecticidas para el control de la mosca blanca *Bemisia tabaci* (Gennadius) en tomate. [Evaluation of insecticides for the control of the whitefly *Bemisia tabaci* (Gennadius), in tomato]. *Bull. Entomol. Venezuela* 8(2): 123-132. [Spanish, English summary]
- 2744.Marchoux, G., Leclant, F. and Mathai, P.J. 1970. Maladies de type jaunisse et maladies voisines affectant principalement les solanacees et transmises par des insectes. *Ann. Phytopathol.* 2: 735-773. [Cock (1986)]
- 2745.Marco, S., Cohen, S., Harpaz, I. and Birk, Y. 1972. In vivo suppression of plant virus transmissibility by an anti-TMV factor occurring in an inoculative vector's body. *Virology* 47: 761-766. [Cock (1986)]
- 2746.Marco, S., Cohen, S., Harpaz, I. and Birk, Y. 1975. An anti-TMV factor in the tobacco whitefly after acquisition of tomato yellow leaf curl virus. *J. Insect Physiol.* 21: 1821-1826. [Cock (1986)]
- 2747.Mardiningasih, T.L., Iskandar, M. and Balfas, R. 1996. [Pest on several drug plants]. pp. 149-160. *In* D. Sitepu, S.M.D. Rosita, Soediarto, Hernani, H. Moko and Supriadi (Eds.), Proceedings on Consultative Forum of Strategy and Coordination of Agroindustrial Development of Drug Plants, Bogor, Indonesia, 28-29 Nov 1995. Balai Penelitian Tanaman Rempah Dan Obat, Bogor, Indonesia. [Indonesian, English summary]
- 2748.Mariappan, F. and Narayanasamy, P. 1972. *Acanthospermum hispidum* D.C., a new host of tomato leaf curl virus. *Madras Agric. J.* 59: 355-357. [Cock (1986)]
- 2749.Mariappan, V. and Ramanujam, K. 1975. Yellow ring mosaic of *Jasminum* spp. with particular reference to its transmission by an aleurodid. *South Indian Hortic.* 23(1-2): 77-78. [Cock (1986)]
- 2750.Markham, P.G. and Bedford, I.D. 1993. Comportamiento y modos de accion de *Bemisia tabaci* (Gennadius): capacidad de transmitir virosis. [Behaviour and action ways of *Bemisia tabaci* (Gennadius): capacity to transmit viroses]. *Phytoma (España)* 50: 47-52. [Spanish]
- 2751.Markham, P.G., Bedford, I.D., Liu, S., Frohlich, D.R., Rosell, R. and Brown, J.K. 1996. The transmission of geminiviruses by biotypes of *Bemisia tabaci* (Gennadius). pp. 69-75. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia* : 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
- 2752.Markham, P.G., Bedford, I.D., Liu, S.J. and Pinner, M.S. 1994. The transmission of geminiviruses by *Bemisia tabaci*. *Pestic. Sci.* 42(2): 123-128.
- 2753.Markkula, M. 1988. Pests of cultivated plants in Finland during 1987. *Ann. Agric. Fenniae* 27(4): 323-327. [Cock (1993)]
- 2754.Mart, C., Kismir, A., Belli, A., Tunc, A., Turhan, N., Pala, Y., Kisakurek, N., Karaat, S. and Aktura, T. 1995. Researches on the chemical control of whitefly (*Bemisia tabaci* Genn.) on cotton fields in Mediterranean Region. *Plant Protection Research Institute, Adana, Turkey*, 51 pp. [Turkish, English summary]
- 2755.Mart, C., Kismir, A., Belli, A., Tunc, A., Turhan, N., Pala, Y., Kisakurek, N., Karaat, S. and Laktura, T. 1995. Studies on the chemical control of whitefly (*Bemisia tabaci* Genn.) on cotton in Mediterranean region. *Zirai Mucadele Arastirma Yilligi* 30: 13-14. [Turkish]
- 2756.Martin, E.C., Dittmar, S.H., Ellsworth, P.C., Silvertooth, J.C., McCloskey, W.B., Olsen, M.W., Roth, R.L. and Tronstad, R.E. 2000. 1999 Integrated cotton management demonstration. pp. 35-43. *In* Cotton, A College of Agriculture Report, SeriesP -121 . University of Arizona, Tucson.
- 2757.Martin, J.H. 1985. The whitefly of New Guinea (Homoptera: Aleyrodidae). *Bull. British Museum (Natural History) Entomol. Series.* 50(3): 303-351. [Cock (1986)]
- 2758.Martin, J.H. 1987. An identification guide to common whitefly species of the world (Homoptera: Aleyrodidae). *Trop. Pest Manage.* 33: 298-322. [Cock (1993)]
- 2759.Martin, J.H. 1999. The whitefly fauna of Australia (Sternorrhyncha: Aleyrodidae). A taxonomic account and identification guide. Division of Entomology Tech. Paper 38. Australia Commonwealth Scientific and Industrial Research Organization, 197 pp.
- 2760.Martin, J.H., Franquinho-Aguiar, A.M. and Pita, M.T. 1996. Aleyrodidae of Madeira: descriptions of three new species, with notes on a pan-Mediterranean species of *Aleurotrachelus*. *J. Nat. Hist.* 30(1): 113-125.
- 2761.Martin, J.H., Mifsud, D. and Rapisarda, C. 2000. The whiteflies (Hemiptera : Aleyrodidae) of Europe and the Mediterranean Basin. *Bull. Entomol. Res.* 90: 407-448.
- 2762.Martin, N.A. 1989. Greenhouse tomatoes. A survey of pest and disease control. *DSIR Plant Prot. Rep.* 1: 1-42. [Cock (1993)]
- 2763.Martinez-Carrillo, J.L. 1998. Control químico de la mosquita blanca. [Chemical control of the whitefly]. pp. 113-118. *In* J.J. Pacheco-Covarrubias and F. Pacheco-Mendivil (Eds.), *Temas Selectos Para El Manejo Integrado de la Mosquita Blanca, Memoria Cientifica No. 6.* Instituto Nacional De Investigaciones Forestales, Agrícolas y Pecuarias, Centro de Investigacion Regional del Noroeste, CIRNO, Obregon, Sonora, Mexico.
- 2764.Martinez-Carrillo, J.L. 1998. Estrategia para el manejo de la resistencia en mosquita blanca. [Strategies for resistance management in whiteflies]. pp. 119-126. *In* J.J. Pacheco-Covarrubias and F. Pacheco-Mendivil (Eds.), *Temas Selectos Para El Manejo Integrado de la Mosquita Blanca, Memoria Cientifica No. 6.* Instituto Nacional De Investigaciones Forestales, Agrícolas y Pecuarias, Centro de Investigacion Regional del Noroeste, CIRNO, Obregon, Sonora, Mexico. [Spanish]

2765. Martínez-Carrillo, J.L. 1998. Generalidades de las mosquitas blancas [Generalities of whiteflies]. pp. 27-30. In J.J. Pacheco-Covarrubias and F. Pacheco-Mendivil (Eds.), *Temas Selectos Para El Manejo Integrado de la Mosquita Blanca*, Memoria Científica No. 6. Instituto Nacional De Investigaciones Forestales, Agrícolas y Pecuarias, Centro de Investigación Regional del Noroeste, CIRNO, Obregon, Sonora, Mexico.
2766. Martínez-Carrillo, J.L. 1998. Resistencia a insecticidas en poblaciones de mosquita blanca de la hoja plateada (*Bemisia argentifolii* Bellows and Perring) del Valle de Yaqui, Sonora. *Memoria Científica* 5: 86-95.
2767. Martínez-Carrillo, J.L., Camberos, U.N., González, M.A., Díaz-Ortiz, B.E. and Villegas, R.S. 2001. Monitoring resistance to insecticides on silverleaf whitefly (*Bemisia argentifolii*) from northwestern Mexico. pp. 872-874. In P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
2768. Martínez-Carrillo, J.L., Rodríguez-Cota, F.G., Navarro-Sandoval, F.J. and López-Ahumada, B. 1998. Establecimiento de un manejo integrado para mosquita blanca en soya en el Norte de Sinaloa. INIFAP-CIRNO-CEVAF, Folleto Técnico No. 14
2769. Martínez, R.T., Beaver, J.S., Saladin, F. and Maxwell, D.P. 1991. Lack of seed transmission by bean golden mosaic geminivirus from the Dominican Republic and Puerto Rico. *Annu. Rep. Bean Improvement Coop.* 34: 17.
2770. Martínez, S.S., De Carvalho, A.O.R., Vieira, L.G., Nunes, L.M. and Bianchini, A. 2000. Identification, geographical distribution and host plants of *Bemisia tabaci* (Genn.) biotypes (Homoptera: Aleyrodidae) in the state of Parana, Brazil. *Anais Soc. Entomol. Brasil* 29(3): 597-603.
2771. Martins-Santana, F., Ribeiro, S.D., Moita, A.W., Moreira, D.J. and Giordano, L.B. 2001. Sources of resistance in *Lycopersicon* spp. to a bipartite whitefly-transmitted geminivirus from Brazil. *Euphytica* 122(1): 45-51.
2772. Marutani, M. and Rivera, E. 1996. Control of silverleaf whitefly on tomato, 1995. *Arthropod Management Tests* 21: 182.
2773. Maruthi, M.N., Colvin, J. and Seal, S. 2001. Mating compatibility, life-history traits, and RAPD-PCR variation in *Bemisia tabaci* associated with the cassava mosaic disease pandemic in East Africa. *Entomol. Exp. Appl.* 99(1): 13-23.
2774. Masi, L. 1909. Contribuzioni alla conoscenza dei calcididi italiani. *Bull. Lab. Zool. General Agraria Della Facolta Agraria Portici* 4: 25-26. [Cock (1986)]
2775. Mason, G., Rancati, M. and Bosco, D. 2000. The effect of thiamethoxam, a second generation neonicotinoid insecticide, in preventing transmission of tomato yellow leaf curl geminivirus (TYLCV) by the whitefly *Bemisia tabaci* (Gennadius). *Crop Prot.* 19(7): 473-479.
2776. Masuda, T. 1993. Pathogenicity of *Verticillium lecanii* to the larvae of *Bemisia tabaci*. *Annu. Rep. Soc. Plant Prot. N. Japan* 44: 189-190. [Japanese, English summary]
2777. Masuda, T. and Kikuchi, O. 1992. Pathogenicity of *Verticillium lecanii* isolates to whitefly and aphids. *Japan J. Appl. Entomol. Zool.* 36: 239-245.
2778. Mathew, A.V. and Balakrishnan, S. 1982. Yellow mosaic of *Micrococca mercurialis* Benth. - a sap transmissible whitefly-borne virus. pp. 99-105. In Alexander, K. M. and R.S. Prasad (Eds.), *Vectors and Vector-borne Diseases*. Proceedings of the All India Symposium. Trivandrum, Kerala State, India, February 26-28, 1982. University of Kerala, Trivandrum, India. [Cock (1993)]
2779. Mathew, A.V., Mathew, J. and Mathai, G. 1991. A whitefly transmitted mosaic disease of bittergourd. *Indian Phytopathol.* 44: 497-499.
2780. Mathew, A.V. and Muniyappa, V. 1991. Transmission of Indian cassava mosaic virus by *Bemisia tabaci*. *Fitopatol. Brasileira* 16(1): 46-49.
2781. Mathew, A.V. and Muniyappa, V. 1992. Purification and characterization of Indian cassava mosaic virus. *J. Phytopathol.* 135(4): 299-308.
2782. Mathew, A.V. and Muniyappa, V. 1993. Host range of Indian cassava mosaic virus. *Indian Phytopathol.* 46(1): 16-23.
2783. Mathur, R.N. 1932. Leaf curl of cotton in garden zinnia in North India. *Nature (London)* 129: 797. [Ali et al. (1992)]
2784. Mathur, R.N. 1933. Leaf-curl in *Zinnia elegans* at Dehra Dun. *Indian J. Agric. Sci.* 129: 89-96. [Cock (1986)]
2785. Matile-Ferrero, D. 1978. Cassava mealybug in the People's Republic of Congo. pp. 29-46. In K.F. Nwanze and K. Leuschner (Eds.), *Proceedings International Workshop on the Cassava Mealybug *Phenacoccus manihoti* Mat.-Ferr. (Pseudococcidae) held at INERA - M'vuazi, Bas-Zaire, Zaire, June 26-29, 1977*. International Institute for Tropical Agriculture, Ibadan, Nigeria. [Cock (1986)]
2786. Matsui, M. 1992. Irregular ripening of tomato fruit caused by the sweetpotato whitefly *Bemisia tabaci* (Gennadius) in Japan. *Japan. J. Appl. Entomol. Zool.* 36(1): 47-49.
2787. Matsui, M. 1995. Efficiency of *Encarsia formosa* Gahan in suppressing population density of *Bemisia argentifolii* Bellows and Perring on tomatoes in plastic greenhouses. *Japan J. Appl. Entomol. Zool.* 39(1): 25-31.
2788. Matsui, M. and Nakashima, T. 1992. Damage to vegetables and ornamental plants by the sweetpotato whitefly and its control in Japan. *Japan Pestic. Information* 60: 15-18.
2789. Matthews, G.A. 1986. Overview of chemical control with special references to cotton crops. pp. 55-58. In M.J.W. Cock (Ed.), *Bemisia tabaci - A Literature Survey on the Cotton Whitefly with an Annotated Bibliography*. CAB International Institute of Biological Control, Ascot, UK.
2790. Matthews, R.E.F. 1979. The classification and nomenclature of viruses. *Intervirology* 12: 133-135. [Cock (1986)]
2791. Matyis, J.C., Silva, D.M., Oliveira, A.R. and Costa, A.S. 1975. Purificao e morfologia do virus do mosaico dourado do tomateiro. *Summa Phytopathol.* 1: 267-274. [Cock (1986)]
2792. Mauromicale, G., Davino, M., Nucifora, S., Sortino, O., Raccuia, S. and D'Urso, F. 1996. Pacciamatura del pomodoro: difesa e comportamento agronomico. [Effects of mulching with polyethylene sheets of different characteristics on infestation by *Bemisia tabaci*, TYLCV infections and yield of tomato grown in greenhouses]. *Culture Protette (Italy)* 25(6): 73-79. [Italian, English summary]
2793. Maxwell, D.P., Hanson, S.F., Hoogstraten, R.A., Ahlquist, P., Beaver, J.S., Azzam, O. and Karkashian, J. 1994. La organizacion y funcion del genoma del virus del mosaico dorado del frijol. [Genome organization and functions of bean golden mosaic geminivirus]. pp. 119-124. In F.J. Morales (Ed.), *Bean Golden Mosaic: Research Advances*. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. [Spanish, English summary]
2794. Maya-Hernandez, V. and Garza-Urbina, E. 1998. Identificacion de hospederos de mosquita blanca (Homoptera: aleyrodidae) en el estado de San Luis Potosi. [White fly host identification (Homoptera: aleyrodidae) in San Luis Potosi State]. pp. 206-207. In *Proceedings 21st National Congress of Biological Control*, Rio Bravo, Tampaulipas, Mexico, 5-6 Nov 1998. INIFAP [Spanish]
2795. Mayer, R.T., McCollum, T.G., McDonald, R.E., Polston, J.E. and Doodstar, H. 1996. *Bemisia* feeding induces pathogenesis-related proteins in tomato. pp. 179-188. In D. Gerling and R.T. Mayer (Eds.), *Bemisia: 1995 Taxonomy, Biology, Damage, Control and Management*. Intercept Ltd., Andover, Hants, UK.
2796. Maynard, D.N. and Cantliffe, D.J. [no date]. Squash silverleaf and tomato irregular ripening: New vegetable disorders in Florida. *Univ. Florida Coop. Ext., Vegetable Crops Fact Sheet*, VC-37, 4 pp.
2797. Mayné, R. and Ghesquière, J. 1934. Hémiptères nuisibles aux végétaux du Congo Belge. *Ann. Gembloux* 40: 1-41. [Cock (1986)]

2798. Mazumder, N., Borthakur, U. and Choudhury, D. 1996. Incidence of yellow vein mosaic virus of bhindi (*Abelmoschus esculentus* L., Moench) in relation to cultivar and vector population under different sowing dates. *Indian J. Virol.* 12(2): 137-141.
2799. Mazyad, H.M., Hassan, A.A., Nakhla, M.K. and Moustagfa, S.E. 1982. Evaluation of some wild *Lycopersicon* species as sources of resistance to tomato yellow leaf curl. *J. Hortic.* 9: 241-246.
2800. Mazyad, H.M. and Moawad, G.M. 1997. Egypt. pp. 19-25. *In* N. Ioannou (Ed.), Management of the whitefly-virus complex. FAO Plant Production and Protection Paper 143, Rome, Italy.
2801. Mazyad, H.M., Omar, F., Al-Taher, K. and Salha, M. 1979. Observations on the epidemiology of tomato yellow leaf curl virus disease on tomato plants. *Plant Dis. Rep.* 63: 695-698.
2802. McAuslane, H.J. 1996. Influence of leaf pubescence on ovipositional preference of *Bemisia argentifolii* (Homoptera: Aleyrodidae) on soybean. *Environ. Entomol.* 25(4): 834-841.
2803. McAuslane, H.J., Golub, R. and Schuster, D.J. 1996. Evaluation of tomato cultivars, isolines, and near-isolines for resistance to silverleaf whitefly, Spring 1995. *Arthropod Management Tests* 21: 426-427.
2804. McAuslane, H.J., Johnson, F.A., Colvin, D.L. and Sojack, B. 1995. Influence of foliar pubescence on abundance and parasitism of *Bemisia argentifolii* (Homoptera: Aleyrodidae) on soybean and peanut. *Environ. Entomol.* 24: 1135-1143.
2805. McAuslane, H.J., Johnson, F.A. and Knauff, D.A. 1994. Population levels and parasitism of *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae) on peanut cultivars. *Environ. Entomol.* 23(5): 1203-1210.
2806. McAuslane, H.J., Johnson, F.A., Knauff, D.A. and Colvin, D.L. 1993. Seasonal abundance and within-plant distribution of parasitoids of *Bemisia tabaci* (Homoptera, Aleyrodidae) in peanuts. *Environ. Entomol.* 22: 1043-1050.
2807. McAuslane, H.J., Knauff, D.A. and Johnson, F.A. 1995. Evaluation of peanut breeding lines for resistance to silverleaf whitefly (Homoptera: Aleyrodidae) [Spanish summary]. *Florida Entomol.* 78(1): 75-81.
2808. McAuslane, H.J. and Nguyen, R. 1996. Reproductive biology and behavior of thelytokous species of *Eretmocerus* (Hymenoptera: Aphelinidae) parasitizing *Bemisia argentifolii* (Homoptera: Aleyrodidae). *Am. Entomol. Soc. Am.* 89(5): 686-693.
2809. McAuslane, H.J., Simmons, A.M. and Jackson, D.M. 2000. Parasitism of *Bemisia argentifolii* on collard with reduced or normal leaf wax. *Florida Entomol.* 83(4): 428-437.
2810. McAuslane, H.J., Webb, S.E. and Elmstrom, G.W. 1996. Resistance in germplasm of *Cucurbita pepo* to silverleaf, a disorder associated with *Bemisia argentifolii* (Homoptera: Aleyrodidae). *Florida Entomol.* 79(2): 206-221.
2811. McCreight, J.D. 1992. Preliminary screening of melons for sweetpotato whitefly resistance. *Cucurbit Genet. Coop.* 15: 59-61.
2812. McCreight, J.D. 1993. Screening of melons for sweetpotato whitefly resistance: 1992. [Salinas, CA]. *Cucurbit Genet. Coop.* 16: 49-52.
2813. McCreight, J.D. 1994. Screening of melons for sweetpotato whitefly (*Bemisia tabaci*) resistance: 1993. *Cucurbit Genet. Coop.* 17: 83-85.
2814. McCreight, J.D. 2000. Inheritance of resistance to lettuce infectious yellows virus in melon. *Hortscience* 35(6): 1118-1120.
2815. McCreight, J.D. and Kishaba, A.N. 1991. Reaction of cucurbit species to squash leaf curl virus and sweetpotato whitefly. *J. Am. Soc. Hortic. Sci.* 116(1): 137-141. [Cock (1986,1993)]
2816. McCreight, J.D., Kishaba, A.N. and Mayberry, K.S. 1986. Lettuce infectious yellows tolerance in lettuce. *J. Am. Soc. Hortic. Sci.* 3(5): 788-792. [CATIE (1992)]
2817. McCutcheon, G.S. and Simmons, A.M. 2001. Relationship between temperature and rate of parasitism by *Eretmocerus* sp (Hymenoptera: Aphelinidae), a parasitoid of *Bemisia tabaci* (Homoptera: Aleyrodidae). *J. Agric. Urban Entomol.* 18(2): 97-104.
2818. McGovern, R.J., Polston, J.E., Danyluk, G.M., Hiebert, E., Abouzid, A.M. and Stansly, P.A. 1994. Identification of a natural weed host of tomato mottle geminivirus in Florida. *Plant Dis.* 78(11): 1102-1106.
2819. McGrath, P.F. and Harrison, B.D. 1995. Transmission of tomato leaf curl geminiviruses by *Bemisia tabaci*: effects of virus isolate and vector biotype. *Ann. Appl. Biol.* 126(2): 307-316.
2820. McKenzie, C.L., Toscano, N.C. and Beehler, L. 1996. Field & greenhouse performance of Mustang 1.5 EW and Capture 2EC on immature and adult whitefly. pp. 810-816. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
2821. McLain, J., Castle, S., Holmes, G. and Creamer, R. 1998. Physiochemical characterization and field assessment of lettuce chlorosis virus. *Plant Dis.* 82(11): 1248-1252.
2822. McMahan, R.W., Lindquist, R.K., Baith, B.D., Makin, T.L. and Casey, M.L. 1994. Comparison of commercially and locally produced *Encarsia formosa* Gahan on the control of sweetpotato whitefly on poinsettias. *HortTechnology* 4(3): 295-298.
2823. McPherson, R., Hall, E.K., Lambert, A.L. and Herzog, G.A. 1995. Effects of a natural pesticide from *Nicotiana glauca* on controlling whiteflies (Homoptera: Aleyrodidae) on a susceptible and resistant soybean line. *J. Entomol. Sci.* 30(3): 312-317.
2824. McPherson, R.M. 1996. Rating soybean germplasm for whitefly (Homoptera: Aleyrodidae) infestations. *J. Agric. Entomol.* 13(1): 65-71.
2825. McPherson, R.M. and Lambert, A.L. 1995. Abundance of two whitefly species (Homoptera: Aleyrodidae) on Georgia soybean. *J. Entomol. Sci.* 30(4): 527-533.
2826. McWhorter, F.P. 1957. Virus diseases of geranium in the Pacific Northwest. *Plant Dis. Rep.* 41: 83-88. [Cock (1986)]
2827. Meade, D.L. and Byrne, D.N. 1991. The use of *Verticillium lecanii* against subimaginal instars of *Bemisia tabaci*. *J. Invertebr. Pathol.* 57: 296-298.
2828. Meagher, R.L., Smith, C.W. and Smith, W.J. 1997. Preference of *Gossypium* genotypes to *Bemisia argentifolii* (Homoptera: Aleyrodidae). *J. Econ. Entomol.* 90(4): 1046-1052.
2829. Meagher, R.L., Jr. 1994. Resistance of bedding plants to twospotted spider mite and sweetpotato whitefly. *Subtrop. Plant Sci.* 46: 62-65.
2830. Meagher, R.L., Jr. and Estrada, J.A. 1994. Hibiscus resistance to sweetpotato whitefly. *Subtrop. Plant Sci.* 46: 69-71.
2831. Medeiros, F.A.S.B., Bleicher, E. and Menezes, J.B. 2001. Efeito do oleo mineral e do detergente neutro na eficiencia de controle da mosca-branca por betacyfluthrin, dimethoato e methomyl no meloeiro. [Effect of mineral oil and neutral detergent on the efficiency of control of whitefly in melon plants by betacyfluthrin, dimethoate and methomyl]. *Hortic. Brasileira* 19(1): 74-76. [Portuguese, English summary]
2832. Meekes, E.T.M. 2001. Entomopathogenic fungi against whiteflies: tritrophic interactions between *Aschersonia* species, *Trialeurodes vaporariorum* and *Bemisia argentifolii*, and glasshouse crops. Ph.D. Dissertation, Wageningen University, 181 pp.
2833. Meekes, E.T.M., Franssen, J.J. and van Lenteren, J.C. 1994. The use of entomopathogenic fungi for the control of whiteflies. *Med. Fac. Landbouww. Univ. Gent* 59(2A): 371-377.
2834. Meekes, E.T.M., Franssen, J.J. and van Lenteren, J.C. 1996. Pathogenicity of entomopathogenic fungi of the genus *Aschersonia* against whitefly. *Bull. OILB/SROP (IOBC/WPRS)* 19(1): 103-106.
2835. Meena, R.S., Rathore, G.S., Shekhawat, B.S., Yadav, L.D. and Agnathotri, J.P. 1984. Efficacy of sowing dates and trap crops in management of yellow mosaic of moth (*Vigna aconitifolia* (Jacq.) Marechal). *Indian J. Mycol. Plant Pathol.* 14(3): 304-309. [Cock (1993)]
2836. Mehrotra, K.N., Singh, A.K., Srivastava, M., Singh, S.P. and Phokela, A. 1995. Response of white fly, *Bemisia tabaci* (Genn.) to cypermethrin. *Pestic. Res. J.* 7(2): 155-156.

- 2837.Mehta, P. 1995. Transmission and molecular detection of tomato-infecting geminiviruses vectored by the sweetpotato whitefly, *Bemisia tabaci* (Homoptera: Aleyrodidae). [University of Wisconsin]. Dissertation Abst. Int. 56(12B): 6525.
- 2838.Mehta, P., Wyman, J.A., Nakhla, M.K. and Maxwell, D.P. 1994. Polymerase chain reaction detection of viruliferous *Bemisia tabaci* (Homoptera: Aleyrodidae) with two tomato-infecting geminiviruses. *J. Econ. Entomol.* 87(5): 1285-1290.
- 2839.Mehta, P., Wyman, J.A., Nakhla, M.K. and Maxwell, D.P. 1994. Transmission of tomato yellow leaf curl geminivirus by *Bemisia tabaci* (Homoptera: Aleyrodidae). *J. Econ. Entomol.* 87(5): 1291-1297.
- 2840.Melamed-Madjar, V., Chen, M. and Rosilio, D. 1975. Screening insecticides against the tobacco whitefly (*Bemisia tabaci*) on cotton, using a leaf cage laboratory method. *Phytoparasitica* 12: 119-125. [Cock (1986)]
- 2841.Melamed-Madjar, V., Cohen, S., Chen, M., Tam, S. and Rosilio, D. 1979. Observations on populations of *Bemisia tabaci* Gennadius (Homoptera: Aleyrodidae) on cotton adjacent to sunflower and potato in Israel. *Israel J. Entomol.* 13: 71-78. [Cock (1986)]
- 2842.Melamed-Madjar, V., Cohen, S., Chen, M., Tam, S. and Rosilio, D. 1982. A method for monitoring *Bemisia tabaci* and timing spray applications against the pest in cotton fields in Israel. *Phytoparasitica* 10: 85-91. [Cock (1986)]
- 2843.Melamed-Madjar, V., Cohen, S., Tomer, S., Yoles, K., Yossef, R., Rozolio, D. and Tam, S. 1977. Screening of insecticides for the control of tobacco whitefly [*Bemisia tabaci*] in cotton. *Hassadeh* 57(9): 1716-1720.
- 2844.Melamed-Madjar, V., Gerson, U. and Tal, S. 1983. A laboratory method for estimating survival of tobacco whitefly nymphs after insecticide treatment, based on honeydew excretion. *Phytoparasitica* 11: 25-32. [Cock (1986)]
- 2845.Melamed-Madjar, V., Navon, A. and Tal, S. 1984. Honeydew staining to evaluate survival of tobacco whitefly nymphs after insecticide application. *Phytoparasitica* 12: 157-161. [Cock (1986)]
- 2846.Melamed-Madjar, V. and Yatom, S. 1992. Gamma radiation, a quarantine measure against *Bemisia tabaci* in ornamentals. *Hassadeh* 72(9): 1167-1168. [Hebrew]
- 2847.Mellor, H.E. and Anderson, M. 1995. Antennal sensilla of whiteflies: *Trialeurodes vaporariorum* (Westwood), the glasshouse whitefly, *Aleyrodes proletella* (Linnaeus), the cabbage whitefly, and *Bemisia tabaci* (Gennadius), the tobacco whitefly (Homoptera, Aleyrodidae). Part 1: external morphology. *Int. J. Insect Morph. Embryol.* 24(2): 133-143.
- 2848.Memane, S.A., Joi, M.B. and Kale, P.N. 1987. Screening of chilli cultivars against leaf curl complex. *Curr. Res. Rep., Mahatma Phule Agric. Univ.* 3(1): 98-99. [Cock (1993)]
- 2849.Men, U.B. and Kandalkar, H.G. 1997. Pest complex of sunflower, *Helianthus annuus* Linn, in Maharashtra. *Punjabrao Krishi Vidyapeeth Res. J. (India)* 21(1): 61-63.
- 2850.Men, U.B., Kandalkar, H.G. and Pawar, N.P. 1997. Whiteflies on sunflower - weather parameters influence. *Punjabrao Krishi Vidyapeeth Res. J. (India)* 21(2): 195-197.
- 2851.Men, U.B. and Sarode, S.B. 1999. Vertical distribution of whiteflies, *Bemisia tabaci* Genn. on sunflower. *Insect Environ.* 5(3): 111.
- 2852.Mendoza-Zamora, C. and Ponce-Gonzalez, F. 1998. Efecto del piriproxyfen (Knack) en el control de huevos y ninfas de mosquita blanca en gerbera. [Effect of pyriproxyfen (Knack) on the control of eggs and nymphs of white fly in gerbera]. pp. 287-292. *In Proceedings 33rd National Entomological Congress, Acapulco, Mexico, 24-27 May 1998. Sociedad Mexicana De Entomologia.* [Spanish]
- 2853.Menn, J.J. 1996. The *Bemisia* complex, an international crop protection problem waiting for a solution. pp. 381-383. *In D. Gerling and R.T. Mayer (Eds.), Bemisia : 1995 Taxonomy, Biology, Damage, Control and Management.* Intercept Ltd., Andover, Hants, UK.
- 2854.Menozzi, P. 1997. Characterization of strains of *Bemisia tabaci* (Gennadius) using molecular biology techniques. *Med. Fac. Landbouww. Univ. Gent* 62(2A): 281-288. [French]
- 2855.Menvielle, J.P. 1995. Managing the whitefly. *Valley Grower* (May-June): 22,23,28.
- 2856.Mercet, R. 1931. Notas sobre Afelinidos (4) (*Eretmocerus, Coccophagus*). *Eos, Madrid* 7: 395-410. [Cock (1986)]
- 2857.Merriam, T.L., Burkart, S.E. and von Maltzahn, C. 1990. Field evaluation of AC 801,757 (MK-239) acaricide. pp. 63-69. *In Brighton Crop Protection Conference: Pests and Diseases.* [Cock (1993)]
- 2858.Mesquita, A.L.M., Lacey, L.A., Mercadier, G. and Leclant, F. 1996. Entomopathogenic activity of a whitefly-derived isolate of *Paecilomyces fumosoroseus* (Deuteromycotina: Hyphomycetes) against the Russian wheat aphid, *Diuraphis noxia* (Hemiptera: Sternorrhyncha: Aphididae) with the description of an effective bioassay method. *Eur. J. Entomol.* 93(1): 69-75.
- 2859.Metwally, E.M., Hassanein, S.S.M. and Afsah, A.F.E. 1994. Effect of planting date on population abundance of certain leaf pests infesting some vegetable crops at Gemmeza region, Egypt. *Egyptian J. Agric. Res.* 72(4): 977-990.
- 2860.Metwally, E.M., Hassanein, S.S.M. and Hafsa, A.F.E. 1995. Seasonal population activity of sucking pests infesting common bean and squash plants at Gemmeza region, Gharbia Governorate, Egypt. *Egyptian J. Agric. Res.* 73(3): 653-667.
- 2861.Metwally, S.A.G. 1999. Effect of planting date and certain weather factors on the population fluctuations of three insect pests infesting kidney beans in Qalyobia governorate. *Egyptian J. Agric. Res.* 77(1): 139-149.
- 2862.Metwally, S.A.G. and Gabr, A.M. 1997. The effect of macro- and micro- elements on the population of Aphids and whiteflies. *Fayoum J. Agric. Res. Devel. (Egypt)* 11(2): 206-212.
- 2863.Metwally, S.A.G., Hegab, M.F.A.H. and Abdel-Mageed, H.A. 1993. Evaluation for some treatment in controlling *Bemisia tabaci* Genn on squash and its relation with crop measurements in Egypt. *Al-Azhar J. Agric. Res. (Egypt)* 18: 219-224.
- 2864.Metwally, S.A.G., Ibrahim, S.M. and Gabr, A.M. 1999. Studies on the effect of certain substances on the population density of whitefly, *Bemisia tabaci* (Genn), and cotton aphid, *Aphis gossypii* glover attacking squash crop. *Mansoura Univ. J. Agric. Sci. (Egypt)* 24(5): 2629-2634.
- 2865.Meyerdirk, D.E. and Coudriet, D.L. 1985. Evaluation of two biotypes of *Euseius scutalis* (Acari: Phytoseiidae) as predators of *Bemisia tabaci* (Homoptera: Aleyrodidae). *J. Econ. Entomol.* 79(3): 659-663. [Cock (1993)]
- 2866.Meyerdirk, D.E. and Coudriet, D.L. 1985. Predation and developmental studies of *Euseius hibisci* (Chant) (Acarina: Phytoseiidae) feeding on *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae). *Environ. Entomol.* 14: 24-27. [Cock (1986)]
- 2867.Meyerdirk, D.E., Coudriet, D.L. and Prabhaker, N. 1986. Population dynamics and control strategy for *Bemisia tabaci* in the Imperial Valley, California. *Agric. Ecosystems Environ.* 17(1-2): 62-67. [Cock (1993)]
- 2868.Michelson, I., Zamir, D. and Czosnek, H. 1994. Accumulation and translocation of tomato yellow leaf curl virus (TYLCV) in a *Lycopersicon esculentum* breeding line containing the *L. chilense* TYLCV tolerance gene Ty-1. *Phytopathology* 84: 928-933.
- 2869.Michelson, I., Zeidan, M., Zamir, D., Czosnek, H. and Zamski, E. 1997. Localization of tomato yellow leaf curl virus (TYLCV) in susceptible and tolerant nearly isogenic tomato lines. *Acta Hort.* 447: 407-414.
- 2870.Middelburg, H.A. 1939. Jaarverslag 1937-1938. Mededelingen, Proefstation voor vorstenlandsche Tabak 87: 1-73. [Cock (1986), German]

2871. Mier, T., Rivera, F., Bermudez, J.C., Dominguez, Y., Benavides, C. and Ulloa, M. 1991. Primer reporte en México del aislamiento de *Verticillium lecanii* a partir de la mosquito blanca e pruebas de patogenicidad in vitro sobre este insecto. [First report in Mexico on the isolation of *Verticillium lecanii* from whitefly and in vitro pathogenicity tests on this insect]. Rev. Mexicana Micol. 7: 149-156. [Spanish, English summary]
2872. Mifsud, D. 1997. Biological control in the Maltese Islands - past initiatives and future programmes. Bull. OEPP 27(1): 77-84.
2873. Miklas, P.N. and Santiago, J. 1996. Reaction of selected tepary bean to bean golden mosaic virus. Hortscience 31: 430-432.
2874. Milgram, M., Cohen, J. and Loebenstein, G. 1996. Effects of sweet potato feathery mottle virus and sweet potato sunken vein virus on sweet potato yields and rates of reinfection of virus-free planting material in Israel. Phytoparasitica 24(3): 189-193.
2875. Miliev, K.D., Alexiev, N.P. and Lazic, B. 1997. Intensive greenhouse tomato production through interplanting. Acta Hort. 462: 649-658.
2876. Miller, R. 1987. Sweet potato whitefly: This pest intends to stay. Grower Talks (Nov): 56, 58-60.
2877. Miller, T.A. 1986. Status of resistance in the cotton insect complex. pp. 162-165. In J.M. Brown and T.C. Nelson (Eds.), Proceedings Beltwide Cotton Production Research Conferences. National Cotton Council, Memphis, TN.
2878. Mills, N.J. and Gutierrez, A.P. 1996. Prospective modeling in biological control: an analysis of the dynamics of heteronomous hyperparasitism in a cotton-whitefly-parasitoid system. J. Appl. Ecol. 33(6): 1379-1394.
2879. Mimeur, J.M. 1946. Aleurodidae du Maroc (Ire note). Bull. Soc. Sci. Naturelles Physiques Maroc. 24(1944): 87-89. [Cock (1986)]
2880. Mineo, G., Sciortino, A. and Fazzari, A. 1994. Fitofagi del melone d'inverno in Sicilia e danno economico dell'*Aphis gossypii* Glov. (Hom. Aphididae). [Phytophages of winter melon in Sicily and economic damage by *Aphis gossypii* Glov. (Hom. Aphididae)]. Informatore Fitopatol. 44(3): 58-63. [Italian, English summary]
2881. Ministerio de Desarrollo Agropecuario. 2000. Memoria. 8th Congreso Latinoamericano y del Caribe de Manejo Integrado de Plagas y 9th Taller Latinoamericano y del Caribe sobre Mosca Blanca y Geminivirus, 22-24 Nov 2000, Panama, Direccion Nacional de Sanidad Vegetal Instituto de Investigacion Agropecuaria de Panama, 205 pp. [Spanish]
2882. Minkenberg, O., Simmons, G.S., Malloy, R., Kaltenbach, J. and Leonard, C. 1994. Biological control of whiteflies on cotton: a reality check. pp. 887-890. In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
2883. Minton, B.W., Bachman, W.W., Ngo, N.D. and Allemann, D.V. 1994. Control of aphids and whiteflies with CGA-215944. pp. 1016-1017. In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
2884. Miranda, M.M.M., Picanco, M., Leite, G.L.D., Zannuncio, J.C. and De Clercq, P. 1998. Sampling and non-action levels for predators and parasitoids of virus vectors and leaf miners of tomato in Brazil. Med. Fac. Landbouww. Univ. Gent 63(2B): 519-526.
2885. Mishra, M.D., Raychaudhuri, S.P. and Jha, A. 1963. Virus causing leaf curl of chilli (*Capsicum annum* L.). Indian J. Microbiol. 3: 73-76. [Cock (1986)]
2886. Mishra, P.M. 1984. Studies on bio-efficacy of some insecticides against the pest complex of tomato, *Lycopersicon esculentum* Mill., var Pusaruby. Madras Agric. J. 71(10): 673-676. [Cock (1993)]
2887. Misra, C.S. and Lamba, K.S. 1929. The cotton white-fly (*Bemisia gossypiperda*, n. sp.). Bull. Agric. Res. Inst., Pusa 196: 1-7. [Cock (1986)]
2888. Misra, H.P. 1994. Golden yellow mosaic virus on yard bean (*Vigna unguiculata* subsp. *sesquipedalis* and its management. Indian J. Agric. Sci 64(11): 804-806.
2889. Misra, R.M. and Mishra, R.K. 1995. White-fly epidemic in *Holoptelea integrifolia* forest in Kanpur Zoological Park, Uttar Pradesh. [Hindi summary]. Indian Forester 121(1): 55-58.
2890. Mitchell, H.R. 2001. Cotton pest control with Capture 2EC insecticide/miticide. pp. 930-934. In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
2891. Mizuno, A.C.R. and Boas, G.L.V. 1997. Biologia da mosca-branca (*Bemisia argentifolii*) em tomate e repolho. [White fly (*Bemisia argentifolii*) biology in tomato and cabbage]. Centro Nacional de Pesquisa de Hortaliças, Brasília, DF, Brazil, 5 pp. [Portuguese]
2892. Moawad, G.M. 1997. Integrated management of cotton sucking insect pests in Egypt. pp. 6-16. In International Cotton Advisory Committee, 56th Plenary Meeting, Paraguay.
2893. Moawad, G.M., Gergis, M.F. and Mohamed, M.A. 1995. Population dynamics, host preference and seasonal distribution patterns of whitefly *Bemisia tabaci* (Genn.) in Middle Egypt. pp. 826-828. In D.A. Richter and J. Armour (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
2894. Moghaddam, H.J., Bashar, G., Nemati, N. and Noori, P. 1997. Comparative estimation of damage caused by *Bemisia tabaci* (Genn.) in different varieties of cotton in Varamin. [Iran]. Appl. Entomol. Phytopathol. 65(1): 13-14 (English), 54-61 (Persian).
2895. Mohamed, M.A. 1986. Ecological studies on some sucking insects infesting cotton in Middle Egypt. Ph.D. Dissertation, Minia University, Egypt, 149 pp.
2896. Mohamed, O.S.A. and Adam, S.E.I. 1990. Toxicity of sumicidin (fenvalerate) to Nubian goats. J. Comp. Pathol. 102(1): 1-6. [Cock (1993)]
2897. Mohammad, Z.K. and Ghabbour, M.W. 1997. Overbridging certain gaps in the classification of the nymphal stages of whiteflies in Egypt and a recent method for controlling *Dialeurodes citri* (Ashmead) (Homoptera: Aleyrodidae). J. Egypt. German Soc. Zool. 24 (E): 233-261.
2898. Mohan, M. and Katiyar, K.N. 2000. Impact of different insecticides used for bollworm control on the population of jassid and whitefly in cotton. Pestic. Res. J. 12(1): 99-102.
2899. Mohanty, A.K. and Basu, A.N. 1986. Effect of host plant and seasonal factors on intraspecific variation in pupal morphology of the whitefly vector, *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae). J. Entomol. Res. (New Delhi) 10(1): 19-26. [Cock (1993)]
2900. Mohanty, A.K. and Basu, A.N. 1987. Biology of the whitefly vector, *Bemisia tabaci* Genn. on four host plants throughout the year. J. Entomol. Res. (New Delhi) 11(1): 15-18. [Cock (1993)]
2901. Mohanty, A.K. and Basu, A.N. 1990. Disease transmitting ability of *Bemisia tabaci* Genn. reared on different host plants during different periods. Indian J. Virol. 6(1-2): 108-109. [Cock (1993)]
2902. Mohanty, A.K. and Basu, A.N. 1991. Relative preference of *Bemisia tabaci* Genn. for some of its host plants during different seasons. J. Entomol. Res. (New Delhi) 15(1): 70-71.
2903. Mohanty, A.K. and Basu, A.N. 1991. Seasonal variations in the aerial populations of the whitefly vector *Bemisia tabaci* under Delhi conditions. Indian Phytopathol. 44: 494-496.
2904. Mohanty, A.K., Kar, A.K., Sethi, P.N. and Dhal, A. 1996. Brinjal varieties as sources of rearing host plants for *Bemisia tabaci* Genn. Crop Res. (Hisar) 11(3): 386-387.
2905. Mohite, P.B. and Uthamasamy, S. 1997. Influence of varied spacings and fertilizer levels on the incidence of key pests of cotton in Tamil Nadu. Indian J. Agric. Res. 31(4): 222-226.
2906. Mohyuddin, A.I., Jilani, G., Khan, A.G., Hamza, A., Ahmed, I. and Mahmood, Z. 1997. Integrated pest management of major cotton pests by conservation, redistribution and augmentation of natural enemies. Pakistan J. Zool. 29(3): 293-298.

2907. Mohyuddin, A.I., Khan, A.G. and Goraya, A.A. 1989. Population dynamics of cotton whitefly, *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae) and its natural enemies in Pakistan. *Pakistan J. Zool.* 21(3): 273-288.
2908. Mojenji, T.D., Bayatasadi, H. and Poorghaz, A. 1999. Evaluation of insecticides on cotton aphid and *Bemisia* in the Golestan Province of Iran. pp. 43-47. *In Proceedings ICAC-CCRI Regional Consultation, Insecticide Resistance Management in Cotton.* Central Cotton Research Institute, Multan, Pakistan.
2909. Monci, F., Navas-Castillo, J., Cenis, J.L., Lacasa, A., Benazoun, A. and Moriones, E. 2000. Spread of tomato yellow leaf curl virus Sar from the Mediterranean Basin: Presence in the Canary Islands and Morocco. *Plant Dis.* 84(4): 490.
2910. Monga, D., Kumar, S. and Raj, S. 2000. Preliminary investigations on the potential loss caused due to cotton leaf curl virus disease. *Adv. Plant Sci.* 13(2): 615-617.
2911. Monkman, K.D. 1989. A new insect pest in Bermuda. *Monthly Bull. Bermuda Dept. Agric. and Fisheries* 60(1): 2. [Cock (1993)]
2912. Monsef, A.A. and Kashkooli, A. 1978. Die Baumwollweissfliege *Bemisia tabaci* Gen. in der Provinz Fars und die Kontrolle. *Entomol. Phytopathol. Appl.* 46(1-2): 66-77. [Cock (1986), Persian, German summary]
2913. Montasser, M.S., Al-Sharidah, A., Ali, N.Y., Nakhla, M.K., Farag, B.L. and Maxwell, D.P. 1999. A single DNA of tomato yellow leaf curl geminivirus causing epidemics in the State of Kuwait. *Kuwait J. Sci. Eng.* 26(1): 127-141.
2914. Monteiro-Guimaraes, J. 1995. *Bemisia tabaci*. *Proteccao da Producao Agricola* 1: 49-52. [Portuguese]
2915. Monteiro-Neves, A. and Viereck, A. 1987. [Cultivation problems of the Cape Verde Islands with special reference to the island of Santo Antao]. *Courier Forschungsinstituts Senckenberg* 95: 41-49. [Cock (1993), German, English summary]
2916. Montesdeoca-Montesdeoca, M., Camero-Hernandez, A., Perez-Padron, F., Siverio-Nunez, A. and Rodriguez-Lopez, P. 1992. Presencia de *Bemisia tabaci* (Genn.) en cultivos comerciales hortícolas y ornamentales en las Islas Canarias. [Presence of *Bemisia tabaci* (Genn.) in horticultural and ornamental commercial crops in Canary Islands]. *Agric. Vergel (Spain)* 123: 152-157. [Spanish]
2917. Montilla, D. and Teran, H. 1996. UCLA-1, una nueva variedad de ajonjolí (*Sesamun indicum* L.). [UCLA-1, a new (*Sesamun indicum* L.) variety]. *Rev. Decanato Agron., Bioagro (Venezuela)* 8(1): 26-29. [Spanish, English summary]
2918. Moore, E.S. and Smith, A.J. 1933. Pest and diseases in tobacco seedbeds. *Farming in South Africa* 1933. 89(8): 314-316. [Cock (1986)]
2919. Moores, G.D., Denholm, I., Byrne, F.J., Kennedy, A.L. and Devonshire, A.L. 1988. Characterising acetylcholinesterase genotypes in resistant insect populations. pp. 451-456. *In Brighton Crop Protection Conference: Pests and Diseases.* British Crop Protection Council, Surrey, UK. [Cock (1993)]
2920. Moores, G.D., Devonshire, A.L., Stumpf, N. and Nauen, R. 2000. A fluorometric method to detect insensitive acetylcholinesterase in resistant pests. pp. 447-452. *In Brighton Crop Protection Conference: Pests and Diseases.* British Crop Protection Council, Farnham, UK.
2921. Mor, H., Gindin, G., Benzeev, I.S., Raccach, B., Geschtovt, N.U. and Ajtkhozina, N. 1996. Diversity among isolates of *Verticillium lecanii* as expressed by DNA polymorphism and virulence towards *Bemisia tabaci*. *Phytoparasitica* 24(2): 111-118.
2922. Morales, F., Niessen, A., Ramirez, B. and Castaño, M. 1990. Isolation and partial characterization of a geminivirus causing bean dwarf mosaic. *Phytopathology* 80(1): 96-101. [Cock (1993)]
2923. Morales, F.J. 1994. Situacion actual del mosaico dorado del frijol en la America del Sur: Brasil. [Current situation of bean golden mosaic virus in Latin America: Brazil]. pp. 90-95. *In F.J. Morales (Ed.), Bean Golden Mosaic: Research Advances.* Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. [Spanish]
2924. Morales, F.J. 2000. Current situation of common bean production and whitefly transmitted begomoviruses affecting *Phaseolus vulgaris* in Latin America. pp. 159-168. *In Bean Golden Mosaic and Other Diseases of Common Bean Caused by Whitefly-Transmitted Geminiviruses in Latin America.* Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia.
2925. Morales, F.J. 2000. El mosaico dorado y otras enfermedades del frijol comun causadas por geminivirus transmitidos por mosca blanca en la America Latina: 1960-2000, 40 anos de investigacion colaborativa. Centro Internacional de Agricultura Tropical, Palmira, Valle Del Cauca, Colombia, 169 pp. [Spanish]
2926. Morales, F.J. 2000. Importancia socioeconomica del frijol en la America Latina. pp. 1-11. *In Bean Golden Mosaic and Other Diseases of Common Bean Caused by Whitefly-Transmitted Geminiviruses in Latin America.* Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. [Spanish]
2927. Morales, F.J. 2000. Metodos de control de begomovirus del frijol. pp. 133-154. *In Bean Golden Mosaic and Other Diseases of Common Bean Caused by Whitefly-Transmitted Geminiviruses in Latin America.* Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. [Spanish]
2928. Morales, F.J. 2000. Necesidades de investigacion futura. pp. 155-158. *In Bean Golden Mosaic and Other Diseases of Common Bean Caused by Whitefly-Transmitted Geminiviruses in Latin America.* Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. [Spanish]
2929. Morales, F.J. 2001. Conventional breeding for resistance to *Bemisia tabaci*-transmitted geminiviruses. *In S. E. Naranjo and P. C. Ellsworth (Eds.), Special Issue: Challenges and Opportunities for Pest Management of Bemisia tabaci in the New Century.* *Crop Prot.* 20(9): 825-834.
2930. Morales, F.J. and Anderson, P.K. 2001. The emergence and dissemination of whitefly-transmitted geminiviruses. *Arch. Virol.* 146(3): 415-441.
2931. Morales, F.J., Munoz, C., Castano, M. and Velasco, A.C. 2000. Geminivirus transmitidos por mosca blanca en Colombia. *Fitopatol. Colombiana* 24(1-2): 95-98. [Spanish, English summary]
2932. Morales, F.J. and Niessen, A.I. 1988. Comparative responses of selected *Phaseolus vulgaris* germ plasm inoculated artificially and naturally with bean golden mosaic virus. *Plant Dis.* 72(12): 1020-1023. [Cock (1993)]
2933. Morales, F.J. and Rivera, E.E. 1994. Situacion actual del mosaico dorado del frijol en la America Central: El Salvador. [Current situation of bean golden mosaic virus in Latin America: El Salvador]. pp. 40-44. *In F.J. Morales (Ed.), Bean Golden Mosaic: Research Advances.* Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. [Spanish]
2934. Morales, F.J., Saladin, F., Figueroa, A. and Sanchez, A. 1994. Situacion actual del mosaico dorado del frijol en el Caribe: Republica Dominicana. [Current situation of bean golden mosaic virus in Latin America: Dominican Republic]. pp. 58-71. *In F.J. Morales (Ed.), Bean Golden Mosaic: Research Advances.* Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. [Spanish]
2935. Morales, F.J. and Singh, S.P. 1991. Genetics of resistance to bean golden mosaic virus in *Phaseolus vulgaris* L. *Euphytica* 52: 113-117.
2936. Morales, F.J. and Singh, S.P. 1993. Breeding for resistance to bean golden mosaic virus in an interracial population of *Phaseolus vulgaris* L. *Euphytica* 67: 59-63.
2937. Morales, F.J. (Ed.). 1994. *Bean Golden Mosaic: Research Advances.* Centro Internacional de Agricultura Tropical, Cali, Colombia, 193 pp. [English and Spanish]

2938. Morales, P.A. and Bastidas, Y.R. 1997. Evaluacion de la resistencia de ocho cultivares de melon (*Cucumis melo* L.) al ataque de la mosca blanca *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae) en el sector los Perozos, Estado Falcon, Venezuela. [Evaluation of the resistance of eight cultivars of melon (*Cucumis melo* L.) to attack by the whitefly *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae) in the Los Perozos area, Estado Falcon, Venezuela]. Bull. Entomol. Venezuela 12(2): 141-149. [ Spanish, English summary]
2939. Morales-Valles, P.A. 1997. Resistencia al ataque de *Bemisia tabaci* Gennadius (Homoptera: Aleyrodidae) en ocho cultivares de melon en Paraguana, estado Falcon, Venezuela. [Resistance to attack of *Bemisia tabaci* Gennadius (Homoptera: Aleyrodidae) in eight melon cultivars in Paraguana, Falcon State, Venezuela]. Boletin Entomol. Venezolana 12(1): 63-72. [ Spanish, English summary]
2940. Moreno, R., Garijo, C., Rodriguez, M.D., Garcia, S., Téllez, M.M., Garcia, E. and Rodriguez, M.P. 1994. IPM development on tomato under plastic in the south of Spain. Bull. OILB/SROP (IOBC/WPRS) 17(5): 22-26.
2941. Moreno, V., Gomez-Aguilera, J.L., Guerau de Arellano, C. and Roig, L.A. 1993. Preliminary screening of cucurbits species for *Bemisia tabaci* Genn. whitefly resistance. [Valencia, Spain]. Cucurbit Genet. Coop. 16: 87-89.
2942. Morgan, D. and MacLeod, A. 1996. Assessing the economic threat of *Bemisia tabaci* and tomato yellow leaf curl virus to the tomato industry in England and Wales. pp. 1077-1082. In Brighton Crop Protection Conference: Pests and Diseases. British Crop Protection Council, Farnham, UK.
2943. Morillo, F.E. and Marcano, R.V.B. 1997. Estudio del desarrollo de la mosca blanca en diferentes genotipos de tomate. [Development of the whitefly on different genotypes of tomato]. Agron. Trop. (Maracay) 47(3): 271-286. [ Spanish, English summary]
2944. Morin, S., Ghanim, M., Sobol, I. and Czosnek, H. 2000. The GroEL protein of the whitefly *Bemisia tabaci* interacts with the coat protein of transmissible and nontransmissible begomoviruses in the yeast two-hybrid system. Virology 276(2): 404-416.
2945. Morin, S., Ghanim, M., Zeidan, M., Czosnek, H., Verbeek, M. and van den Heuvel, J.F.J.M. 1999. A GroEL homologue from endosymbiotic bacteria of the whitefly *Bemisia tabaci* is implicated in the circulative transmission of tomato yellow leaf curl virus. Virology 256(1): 75-84.
2946. Morinaga, T., Ikegami, M. and Miura, K. 1993. The nucleotide sequence and genome structure of mung bean yellow mosaic geminivirus. Microbiol. Immunol. 37: 471-476.
2947. Moriones, E. 1996. El virus del rizado amarillo del tomate (tomato yellow leaf curl virus, TYLCV): una nueva virosis en España. pp. 19-22. In J.L. Cenis (Ed.), El Virus del Rizado Amirillo (Hoja en Cuchara) del Tomate (TYLCV) y Su Vector *Bemisia tabaci*, Jornadas 8. Consejería de Medio Ambiente, Agricultura y Agua, Murcia, Spain.
2948. Moriones, E. and Luis-Arteaga, M. 1999. Viral Diseases. pp. 16-33. In R. Albajes, M.L. Gullino, J.C. van Lenteren and Y. Elad (Eds.), Integrated Pest and Disease Management in Greenhouse Crops. Kluwer, Dordrecht, The Netherlands.
2949. Moriones, E. and Navas-Castillo, J. 2000. Tomato yellow leaf curl virus, an emerging virus complex causing epidemics worldwide. Virus Res. 71(1/2): 123-134.
2950. Morrill, A.W. and Back, E.A. 1991. Natural control of white flies in Florida. USDA Bull. 102, 78 pp.
2951. Morris, B., Coates, L., Lowe, S., Richardson, K. and Eddy, P. 1990. Nucleotide sequence of the infectious cloned DNA components of African cassava mosaic virus (Nigerian strain). Nucleic Acids Res. 18(1): 197-198.
2952. Morris, B., Richardson, K., Eddy, P., Zhan, X., Haley, A. and Gardner, R. 1991. Mutagenesis of the AC3 open reading frame of African cassava mosaic virus DNA A reduces DNA B replication and ameliorates disease symptoms. J. Gen. Virol. 72: 1205-1213.
2953. Morris, B.A.M., Richardson, K.A., Haley, A., Zhan, X. and Thomas, J.E. 1992. The nucleotide sequence of the infectious cloned dna component of tobacco yellow dwarf virus reveals features of geminiviruses infecting monocotyledonous plants. Virology 187: 633-642.
2954. Morton, N., Byrne, J.E., Vigil, O., Rodrigues, R., Rodrigues, A., Cabezas, G. and Allen, G.W. 1986. PP321: a new insecticide for boll weevil control. pp. 137-144. In British Crop Protection Conference: Pests and Diseases. British Crop Protection Council, Thornton Heath, UK. [Cock (1993)]
2955. Moskovetz, S.N. 1941. Virus disease of cotton and its control. pp. 173-190. In Transactions of the Conference on Plant Virus Diseases, Moscow Feb. 4-7, 1940. Institut Mikrobiologie Akademik Nauk SSSR, Moscow. [Cock (1986), Russian]
2956. Mossop, M.C. 1932. Cultural methods and tobacco whitefly in Southern Rhodesia. Rhodesia Agric. J. 29: 869-872. [Cock (1986)]
2957. Mostafa, N.R., Karaman, G.A., Ali, M.A. and Abdel Alim, A.A. 1983. Effect of different levels of nitrogen fertilization on the population density of the whitefly *Bemisia tabaci* (Gennadius) infesting soybean plants. Minia J. Agric. Res. Devel. (Egypt) 5(2): 143-151. [CATIE (1992)]
2958. Mote, U.N. 1978. Effect of a few insecticides alone and in combination with agricultural spray oil on the control of whitefly (*Bemisia tabaci* Gennadius) population and incidence of leaf curl virus. Indian J. Plant Prot. 6(1): 19-22. [Cock (1986)]
2959. Mound, L.A. 1962. Studies on the olfaction and colour sensitivity of *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae). Entomol. Exp. Appl. 5: 99-104. [Cock (1986)]
2960. Mound, L.A. 1963. Host -correlated variation in *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae). Proc. Royal Entomol. Soc. London, (A) 38: 171-180. [Cock (1986)]
2961. Mound, L.A. 1965. Effect of leaf hair on cotton whitefly populations in the Sudan Gezira. Empire Cotton Growing Rev. 42: 33-40. [Cock (1986)]
2962. Mound, L.A. 1965. Effect of whitefly (*Bemisia tabaci*) on cotton in the Sudan Gezira. Empire Cotton Growing Rev. 42: 290-294.
2963. Mound, L.A. 1983. Biology and identity of whitefly vectors of plant pathogens. pp. 305-313. In R.T. Plumb and J.M. Thresh (Eds.), Plant Virus Epidemiology. The Spread and Control of Insect -borne Viruses. Blackwell Scientific Publications, Oxford, UK. [Cock (1986)]
2964. Mound, L.A. and Halsey, S.H. 1978. Whitefly of the world. A systematic catalogue of the Aleyrodidae (Homoptera) with host plant and natural enemy data. British Museum (Natural History) and Chichester; John Wiley & Sons, London, UK, 340 pp. [Cock (1986)]
2965. Mourad, E.I. 1996. Population dynamics of *Aphis gossypii* and *Bemisia tabaci* under the cotton chemical control regime. Egyptian J. Agric. Res. 70(2): 451-459.
2966. Mourad, E.I., Keddis, M.E. and Khouneim, Y.F. 1991. Potency of certain insecticides against the cotton whitefly, *Bemisia tabaci* (Genn.). Mansoura Univ. J. Agric. Sci. (Egypt) 16(1): 172-175.
2967. Moustafa, S.S., Nakhla, M.K., Fadl, F.A. and El-Safty, N. 1991. Effect of nursery treatments on the control of tomato yellow leaf curl virus disease. Egyptian J. Agric. Res. 69(3): 807-820.
2968. Moya, A., Guirao, P., Cifuentes, D., Beitia, F. and Cenis, J.L. 2001. Genetic diversity of Iberian populations of *Bemisia tabaci* (Homoptera: Aleyrodidae) based on random amplified polymorphic DNA-polymerase chain reaction. Mol. Ecol. 10(4): 891-897.
2969. Mrig, K.K. and Singh, R. 1985. Incidence of insect-pests on garden bean, *Dolichos lablab* Linn. Bull. Entomol. (New Delhi) 26(1): 5-7. [Cock (1993)]
2970. Muhlberger, E. and Maignet, P. 1999. Aleurodes sur tomate: *Trialetrodes vaporariorum* et *Bemisia argentifolii*. PHM Rev. Hortic. (France) 407: 21-25. [French]

2971. Muigai, S.G. 1997. Enhancement of wild *Lycopersicon* germplasm for resistance to *Bemisia argentifolii* (Homoptera: Aleyrodidae). [University of Florida]. Dissertation Abst. Int. 58(07B): 3393.
2972. Mukherjee, A.K. and Raychaudhuri, S.P. 1964. A note on leafcurl of *Malvaviscus arboreus* Cav. Indian J. Hortic. 21: 176-177. [Cock (1986)]
2973. Mulindangabo, J. and Birandano, B. 1984. Some problems of cassava production in Rwanda. pp. 95-100. In A.H. Greathead, R.H. Markham, R.J. Murphy, S.T. Murphy and I.A.D. Robertson (Eds.), Integrated Pest Management of Cassava Green Mite. Proceedings of a Regional Training Workshop in East Africa, 30 April - 4 May 1984. Commonwealth Institute of Biological Control, Ascot, UK. [Cock (1986)]
2974. Mullineaux, P.M., Rigden, J.E., Dry, I.B., Krake, L.R. and Rezaian, M.A. 1993. Mapping of the polycistronic RNAs of tomato leaf curl geminivirus. Virology 193: 414-423.
2975. Mullins, J.W. and Engle, C.E. 1993. Imidacloprid (BAY NTN 33893): A novel chemistry for sweetpotato whitefly control in cotton. pp. 719-720. In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
2976. Mullins, W. and Christie, D. 1995. Management of aphids, whiteflies and plant bugs with foliarly applied imidacloprid. pp. 868-870. In D.A. Richter and J. Armour (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
2977. Mumtaz, M.K., Parvez, A., Ahmad, A., Irshad, A. and Ali, A. 1997. Abiotic factors affecting population fluctuation of whitefly, *Bemisia tabaci* (Genn.) and their impact on leaf curl virus incidence in different genotypes of cotton. J. Anim. Plant Sci. (Pakistan) 7(3/4): 119-121.
2978. Muniyappa, V. 1980. Whiteflies. pp. 39-85. In K.F. Harris and K. Maramorosch (Eds.), Vectors of Plant Pathogens. Academic Press, New York, USA. [Cock (1986)]
2979. Muniyappa, V. 1983. Epidemiology of yellow mosaic disease of horsegram (*Macrotyloma uniflorum*) in southern India. pp. 331-335. In R.T. Plumb and J.M. Thresh (Eds.), Plant Virus Epidemiology. The Spread and Control of Insect-borne Viruses. Blackwell Scientific Publications, Oxford, UK. [Cock (1986)]
2980. Muniyappa, V., Jalikop, S.H., Saikia, A.K., Chennarayappa, Shivashankar, G., Bhat, A.I. and Ramappa, H.K. 1991. Reaction of *Lycopersicon* cultivars and wild accessions to tomato leaf curl virus. Euphytica 56(1): 37-41. [Cock (1993)]
2981. Muniyappa, V., Rajeshwari, R., Bharathan, N., Reddy, D.V.R. and Nolt, B.L. 1987. Isolation and characterization of a geminivirus causing yellow mosaic disease of horsegram (*Macrotyloma uniflorum* (Lam.) Verdc.) in India. [German summary]. J. Phytopathol. 119(1): 81-87. [Cock (1993)]
2982. Muniyappa, V., Rao, M.R.G., Ravi, K.S. and Shivashankar, G. 1983. The natural occurrence of a yellow mosaic disease of *Voandzeia subterranea* transmitted by white-flies. Int. J. Trop. Plant Dis. 1: 193-194. [Cock (1986)]
2983. Muniyappa, V. and Reddy, D.V.R. 1983. Transmission of cowpea mild mottle virus by *Bemisia tabaci* in a nonpersistent manner. Plant Dis. 67: 391-393. [Cock (1986)]
2984. Muniyappa, V. and Reddy, H.R. 1976. Studies on the yellow mosaic disease of horse gram yellow (*Dolichos biflorus* Linn.). I. Virus vector relationships. Mysore J. Agric. Sci. 10: 605-610. [Cock (1986)]
2985. Muniyappa, V. and Reddy, H.R. 1979. *Indigofera hirsuta* a natural reservoir of horsegram yellow mosaic virus. Madras Agric. J. 66: 350. [Cock (1986)]
2986. Muniyappa, V., Reddy, H.R. and Mustak Ali, T.M. 1978. Studies on the yellow mosaic disease of horsegram (*Dolichos biflorus*), IV. Epidemiology of the disease. Mysore J. Agric. Sci. 12(2): 277-279. [Cock (1986)]
2987. Muniyappa, V., Reddy, H.R. and Shivashankar, G. 1975. Yellow mosaic disease *Dolichos biflorus* Linn. (horsegram). Curr. Res., Univ. Agric. Sci. (Bangalore) 4(10): 176. [Cock (1986)]
2988. Muniyappa, V., Reddy, H.R. and Shivashankar, G. 1976. Studies on the yellow mosaic disease of horsegram (*Dolichos biflorus* Linn.). II. Host range studies. Mysore J. Agric. Sci. 10: 611-614. [Cock (1986)]
2989. Muniyappa, V. and Veeresh, G.K. 1984. Plant virus diseases transmitted by whiteflies in Karnataka. Proc. Indian Acad. Sci., Animal Sci. 93: 397-406. [Cock (1986)]
2990. Muniyappa, V., Venkatesh, H.M., Ramappa, H.K., Kulkarni, R.S., Zeidan, M., Tarba, C.Y., Ghanim, M. and Czosnek, H. 2000. Tomato leaf curl virus from Bangalore (ToLCV-Ban4): Sequence comparison with Indian ToLCV isolates, detection in plants and insects, and vector relationships. Arch. Virol. 145(8): 1583-1598.
2991. Muniz, M. 2000. Host suitability of two biotypes of *Bemisia tabaci* on some common weeds. Entomol. Exp. Appl. 95(1): 63-70.
2992. Muñoz, M. 1996. Preferencia y reproducción de *Bemisia tabaci* en malas hierbas de invierno y en cultivos hortícolas de interés. pp. 55-60. In J.L. Cenis (Ed.), El Virus del Rizado Amirillo (Hoja en Cuchara) del Tomate (TYLCV) y Su Vector *Bemisia tabaci*, Jornadas 8. Consejería de Medio Ambiente, Agricultura y Agua, Murcia, Spain.
2993. Muniz, M. and Nombela, G. 1997. Development, oviposition and female longevity of two biotypes of *Bemisia tabaci* (Homoptera: Aleyrodidae) on three varieties of *Capsicum annum* L. Bull. OILB/SROP (IOBC/WPRS) 20(4): 143-146.
2994. Muniz, M. and Nombela, G. 2001. Differential variation in development of the B- and Q-biotypes of *Bemisia tabaci* (Homoptera: Aleyrodidae) on sweet pepper at constant temperatures. Environ. Entomol. 30(4): 720-727.
2995. Munoz, R., Santamaria, E. and Pitty, A. 1993. Efecto de tres manejos de malezas sobre las plagas enemigos naturales, rendimiento y rentabilidad del frijol. [Effect of three weed management systems on the pests, natural enemies, yield and profitability of beans]. Manejo Integrado De Plagas (Costa Rica) 27: 46-53. [Spanish, English summary]
2996. Munoz, S.C. 1994. Non-preference induced effect of the sweetpotato whitefly, *Bemisia tabaci* (Genn.) type B on carboxy treated cotton plants. pp. 1231-1233. In D.A. Richter and J. Armour (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
2997. Murguido, C. 1983. Efecto comparado de algunos insecticidas organofosforados en el control de la mosca blanca (*Bemisia tabaci*) y del saltahoja (*Empoasca* sp.) en el cultivo del frijol. Ciencia Tecnica Agric., Proteccion de Plantas 6(4): 59-65. [Cock (1986)]
2998. Murguido, C., Gonzalez, G. and Nieves, C. 1991. Preguntas y respuestas sobre la mosca blanca *Bemisia tabaci* (Gennadius) transmisora del Virus del Encrespamiento Amarillo de las Hojas del Tomate (ToYLCV). [Questions and answers about the white fly *Bemisia tabaci* (Gennadius) transmitter of the Tomato Yellow Leaf Curl Virus (ToLCV)]. INISAV, Ciudad de la Habana, Cuba, 5 pp. [Spanish]
2999. Murphy, B.C., Morisawa, T. and Parrella, M.P. 1997. Insect-killing fungi: floriculture's IPM future? Grower Talks 61(10): 60,62,64,66,68.
3000. Murtaza, M.A., Bhatti, M.A. and Qayyum, H.A. 1983. Susceptibility of mung bean varieties to whitefly (*Bemisia tabaci* Genn.) and yellow mosaic. Pakistan Entomol. 5(1-2): 51-56. [CATIE (1992)]
3001. Murthy, K.V.V.S. and Reddy, D.R.R. 1992. Chemical control of yellow mosaic disease of bhendi. Indian J. Plant Prot. 20(2): 198-201.
3002. Murugesan, S. and Chelliah, S. 1977. Transmission of greengram yellow mosaic virus by the white fly, *Bemisia tabaci* (Genn.). Madras Agric. J. 64: 437-441. [Cock (1986)]



3003. Murugesan, S. and Chelliah, S. 1978. Effect of yellow mosaic infection of the host green gram on the biology of *Bemisia tabaci* (Genn.). *Entomol.* 3(1): 41-43. [Cock (1986)]
3004. Murugesan, S. and Chelliah, S. 1981. Efficacy of insecticides in the control of *Bemisia tabaci* (Genn.), a vector of the yellow-mosaic virus disease on greengram. *Indian J. Agric. Sci.* 51: 583-584. [Cock (1986)]
3005. Musana, A.C.Z. 1983. A potentially serious pest. Whitefly. *Zimbabwe Agric. J.* 80(4): 143-146. [Cock (1986)]
3006. Musana, A.C.Z. 1986. A method for monitoring whitefly, *Bemisia tabaci* (Genn.), in cotton in Zimbabwe. *Agric. Ecosystems Environ.* 17(1-2): 29-35. [Cock (1993)]
3007. Myartseva, S.N. 1997. New pests in Turkmenistan. *Zashchita Karantin Rastenii* 10: 29-30. [Russian]
3008. Nachapong, M. and Mabbett, T. 1979. A survey of some wild hosts of *Bemisia tabaci* Genn. around cotton fields in Thailand. *Thai J. Agric. Sci.* 12: 217-222. [Cock (1986)]
3009. Naguib, S.M. 1992. Effect of host plants on the life cycle of the whitefly, *Bemisia tabaci* and the potential of two parasitoids, *Encarsia deserti* and *Eretmocerus californicus*. *Moshtohor Ann. Agric. Sci. (Egypt)* 30(4): 1981-1988.
3010. Nahia, D.K., Kumar, S., Sharma, S.P., Saini, M.L., Prasad, H. and Dikshit, A.K. 1992. Field studies on effectiveness and residues of some organophosphorous insecticides against pest complex of cotton. *Plant Prot. Bull. (Faridabad)* 44(1/2): 18-20.
3011. Naidu, R.A., Gowda, S., Satyanarayana, T., Boyko, V., Reddy, A.S., Dawson, W.O. and Reddy, D.V.R. 1998. Evidence that whitefly-transmitted cowpea mild mottle virus belongs to the genus *Carlavirus*. *Arch. Virol.* 143(4): 769-780.
3012. Naik, B.G., Verma, S. and Phadke, K.G. 1993. Occurrence of pests in relation to degradation of insecticides in brinjal crop during summer and kharif seasons. *Pestic. Res. J.* 5(1): 94-103.
3013. Naik, L.K. and Lingappa, S. 1992. Distribution pattern of *Bemisia tabaci* (Gennadius) in cotton plant. *Insect Sci. Appl.* 13: 377-379.
3014. Nair, N.G. 1981. Relationship between cassava mosaic disease spread and whitefly (*Bemisia tabaci* Gen.) population under different insecticide treatments. *J. Root Crops* 7(1-2): 15-19. [Cock (1986, 1993)]
3015. Nair, N.G. and Daniel, R.S. 1983. Preference of *Bemisia tabaci* Gen. to cassava varieties and their reaction to cassava mosaic disease. *J. Root Crops* 9(1-2): 45-49. [Cock (1993)]
3016. Nair, R.G. and Nambiar, T.A. 1984. Annual progress report 1983. (For the period January-December 1983). Central Tuber Crops Research Institute, Trivandrum, India, 140 pp. [Cock (1986)]
3017. Nair, R.R. and Menon, M.R. 1978. Yellow mosaic of *Micrococca mercurialis* Benth. *Agric. Res. J. (Kerala)* 16: 256. [Cock (1986)]
3018. Nakahara, S. and Hilburn, D.J. 1989. Annotated checklist of the whiteflies (Homoptera: Aleyrodidae) of Bermuda. *J. New York Entomol. Soc.* 97(3): 261-264. [Cock (1993)]
3019. Nakamura, S., Inoue, M., Fujimoto, H. and Kasamatsu, K. 1994. A new application method of pyriproxyfen for controlling sweet potato whitefly, *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae). *Appl. Entomol. Zool.* 29(3): 454-456.
3020. Nakano, A. and Sadano, M. 1994. Outbreak of sweet potato whitefly in the sweet potato culture area in Tokushima prefecture. *Tokushima Agric. Exp. Stn. Rep. (Japan)* 30: 32-40. [Japanese]
3021. Nakhla, M.K. and Maxwell, D.P. 1998. Epidemiology and management of tomato yellow leaf curl disease. pp. 565-583. *In* A. Hadidi, R.K. Khetarpal and A. Koganezawa (Eds.), *Plant virus disease control*. American Phytopathological Society Press, St. Paul, MN.
3022. Nameth, S.T., Laemmlein, F.F. and Dodds, J.A. 1985. Viruses cause heavy melon losses in desert valleys. *California Agric.* 39 (7-8): 28-29. [Cock (1993)]
3023. Nandihalli, B.S., Hugar, P. and Patil, B.V. 1990. Evaluation of neem and neem products against cotton whitefly, *Bemisia tabaci* (Gennadius). *Karnataka J. Agric. Sci. [India]* 3: 58-61.
3024. Nandihalli, B.S., Patil, B.V. and Hugar, P. 1990. The plant that kills notorious insect pests. *My Forest (India)* 26(2): 118-120.
3025. Nandihalli, B.S., Patil, B.V. and Lingappa, S. 1993. Population dynamics of cotton whitefly *Bemisia tabaci* (Genn.). *Karnataka J. Agric. Sci. [India]* 6(1): 25-29.
3026. Nandihalli, B.S. and Thontadarya, T.S. 1986. Symptoms of damage caused by different pests on chilli. *Curr. Res., Univ. Agric. Sci. (Bangalore)* 15(10): 99-101. [Cock (1993)]
3027. Naqvi, K.M., Khanzada, A.G. and Abassi, F.D. 1984. Field evaluation of systemic insecticides; their comparative efficacy against potato sucking insect pests. *Sind J. Agric. Res. (Pakistan)* 4(1): 1-7.
3028. Naqvi, K.M. and Qureshi, A.R. 1974. Control of *Empoasca devastans* Dist., *Thrips tabaci* L., and *Bemisia tabaci* Gen., on cotton with systemic insecticides. *Pakistan Cotton* 18(4): 141-146.
3029. Naqvi, S.H., Talpur, M.A., Rustamani, M.A., Khan, M.M. and Hussain, T. 1995. Relative resistance of mung bean, *Vigna radiata* (L.) Wilczek, varieties to whitefly and yellow mosaic virus. *Proc. Pakistan Congress Zool.* 15: 247-251.
3030. Naranjo, S.E. 1996. Overview of biological control programs for *Bemisia* in the USA with emphasis on Arizona. pp. 4-10. *In* Memoria, Simposium de Control Biologico de Mosquita Blanca. XIX Congreso Nacional de Control Biologico, Culiacan, Sinaloa, Mexico.
3031. Naranjo, S.E. 1996. Sampling *Bemisia* for research and pest management applications. pp. 209-224. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia: 1995 Taxonomy, Biology, Damage, Control and Management*. Intercept Ltd., Andover, Hants, UK.
3032. Naranjo, S.E. 2001. Conservation and evaluation of natural enemies in IPM systems for *Bemisia tabaci*. *In* S. E. Naranjo and P. C. Ellsworth (Eds.), *Special Issue: Challenges and Opportunities for Pest Management of Bemisia tabaci in the New Century*. *Crop Prot.* 20(9): 835-852.
3033. Naranjo, S.E., Butler, G.D., Jr. and Henneberry, T.J. 1996. Bibliography of *Bemisia tabaci* and *Bemisia argentifolii* - 1996 addendum. pp. 188-200. *In* Silverleaf Whitefly: 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01.
3034. Naranjo, S.E., Butler, G.D., Jr. and Henneberry, T.J. 1997. Bibliography of *Bemisia tabaci* and *Bemisia argentifolii* - 1997 addendum. pp. 220-238. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02.
3035. Naranjo, S.E., Butler, G.D., Jr. and Henneberry, T.J. 1998. Bibliography of *Bemisia tabaci* and *Bemisia argentifolii* - 1998 Addendum. pp. 119-137. *In* Silverleaf Whitefly, National Research, Action and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01.
3036. Naranjo, S.E., Butler, G.D., Jr. and Henneberry, T.J. 1999. Bibliography of *Bemisia tabaci* and *Bemisia argentifolii* - 1999 Addendum. pp. 133-154. *In* Silverleaf Whitefly, National Research, Action and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01.
3037. Naranjo, S.E., Butler, G.D., Jr. and Henneberry, T.J. 2000. Bibliography of *Bemisia tabaci* and *Bemisia argentifolii* - 2000 Addendum. pp. 150-170. *In* Silverleaf Whitefly, National Research, Action and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000.
3038. Naranjo, S.E., Butler, G.D., Jr. and Henneberry, T.J. 2001. Bibliography of *Bemisia tabaci* and *Bemisia argentifolii* - 2001 Addendum. U.S. Dept. Agric., Agric. Res. Serv., Western Cotton Research Laboratory, 15 pp.

- 3039.Naranjo, S.E., Chu, C.C., Ellsworth, P.C., Henneberry, T.J., Nichols, R.L., Riley, D.G., Toscano, N.C. and Watson, T.F. 1996. Results of a two year regional project to determine an action threshold to manage whiteflies in cotton. pp. 132-133. *In* P. Dugger and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3040.Naranjo, S.E., Chu, C.C. and Henneberry, T.J. 1996. Economic injury levels for *Bemisia tabaci* (Homoptera: Aleyrodidae) in cotton: impact of crop price, control costs, and efficacy of control. *Crop Prot.* 15(8): 779-788.
- 3041.Naranjo, S.E., Diehl, J.W. and Ellsworth, P.C. 1997. Sampling whiteflies in cotton: validation and analysis of enumerative and binomial plans. *Environ. Entomol.* 26(4): 777-788.
- 3042.Naranjo, S.E. and Ellsworth, P.C. 1999. Mortality factors affecting whitefly populations in Arizona cotton management systems: life table analysis. pp. 402-411. *In* Cotton, A College of Agriculture Report, Series P-116. University of Arizona, Tucson.
- 3043.Naranjo, S.E. and Ellsworth, P.C. 2001. Introduction. *In* S. E. Naranjo and P. C. Ellsworth (Eds.), Special Issue: Challenges and Opportunities for Pest Management of *Bemisia tabaci* in the New Century. *Crop Prot.* 20(9): 707.
- 3044.Naranjo, S.E., Ellsworth, P.C., Chu, C.C., Henneberry, T.J., Riley, D.G., Watson, T.F. and Nichols, R.L. 1998. Action thresholds for the management of *Bemisia tabaci* (Homoptera: Aleyrodidae) in cotton. *J. Econ. Entomol.* 91(6): 1415-1426.
- 3045.Naranjo, S.E., Ellsworth, P.C. and Diehl, J.W. 1998. Whitefly management in Arizona: Contribution of natural enemies to whitefly mortality. pp. 324-329. *In* Cotton, A College of Agriculture Report, Series P-112. University of Arizona, Tucson.
- 3046.Naranjo, S.E. and Ellsworth, P.C. (Eds.). 2001. Special Issue: Challenges and Opportunities for Pest Management of *Bemisia tabaci* in the New Century. *Crop Prot.* 20(9): 707-869.
- 3047.Naranjo, S.E. and Flint, H.M. 1993. Progress in the development of sampling plans for *Bemisia tabaci* in cotton. pp. 673-674. *In* D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3048.Naranjo, S.E. and Flint, H.M. 1993. Sequential sampling plans for *Bemisia tabaci* eggs and nymphs in cotton. pp. 216-220. *In* Cotton, A College of Agriculture Report, Series P-94. University of Arizona, Tucson.
- 3049.Naranjo, S.E. and Flint, H.M. 1994. Spatial distribution of preimaginal *Bemisia tabaci* (Homoptera: Aleyrodidae) in cotton and development of fixed-precision sequential sampling plans. *Environ. Entomol.* 23(2): 254-266.
- 3050.Naranjo, S.E. and Flint, H.M. 1995. Spatial distribution of adult *Bemisia tabaci* (Homoptera: Aleyrodidae) in cotton and development and validation of fixed-precision sampling plans for estimating population density. *Environ. Entomol.* 24: 261-270.
- 3051.Naranjo, S.E., Flint, H.M. and Henneberry, T.J. 1994. Numerical and binomial sequential sampling plans for adult *Bemisia tabaci* in cotton. pp. 308-312. *In* Cotton, A College of Agriculture Report, Series P-96. University of Arizona, Tucson.
- 3052.Naranjo, S.E., Flint, H.M. and Henneberry, T.J. 1994. Progress in the development of sampling plans for *Bemisia tabaci*: evaluation of binomial sampling methods. pp. 875-877. *In* D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3053.Naranjo, S.E., Flint, H.M. and Henneberry, T.J. 1995. Comparative analysis of selected sampling methods for adult *Bemisia tabaci* (Homoptera: Aleyrodidae) in cotton. *J. Econ. Entomol.* 88(6): 1666-1678.
- 3054.Naranjo, S.E., Flint, H.M. and Henneberry, T.J. 1995. Comparative analysis of two sampling methods for estimating abundance of adult *Bemisia tabaci* in cotton. pp. 247-254. *In* Cotton, A College of Agriculture Report, Series P-99. University of Arizona, Tucson.
- 3055.Naranjo, S.E., Flint, H.M. and Henneberry, T.J. 1996. Binomial sampling plans for estimating and classifying population density of adult *Bemisia tabaci* on cotton. *Entomol. Exp. Appl.* 80: 343-353.
- 3056.Naranjo, S.E. and Hagler, J.R. 1997. *Bemisia* growth regulators: conservation of natural enemies? pp. 932-933. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3057.Naranjo, S.E. and Hagler, J.R. 1997. Conservation of natural enemies relative to use of insect growth regulators for control of sweetpotato whitefly. pp. 318-323. *In* Cotton, A College of Agriculture Report, Series P-108. University of Arizona, Tucson.
- 3058.Naranjo, S.E. and Hagler, J.R. 1998. Characterizing and estimating the effect of heteropteran predation. pp. 171-197. *In* M. Coll and J. Ruberson (Eds.), Predatory Heteroptera: Their ecology and use in biological control. Thomas Say Symposium Proc. Entomological Society of America, Lanham, Maryland
- 3059.Naranjo, S.E., Hagler, J.R. and Ellsworth, P.C. 1998. Whitefly management in Arizona: conservation of natural enemies relative to insecticide regime. pp. 319-323. *In* Cotton, A College of Agriculture Report, Series P-112. University of Arizona, Tucson.
- 3060.Naranjo, S.E. and Henneberry, T.J. 1998. Sampling methods to estimate lint stickiness. *California-Arizona Cotton* 34(3): 11.
- 3061.Naranjo, S.E., Henneberry, T.J. and Chu, C.C. 1998. Progress in the development of sampling methods to estimate cotton lint stickiness due to sweetpotato whitefly infestation. p. 1087. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3062.Naranjo, S.E. and Hutchison, W.D. 1997. Validation of arthropod sampling plans using a resampling approach: Software and analysis. *Am. Entomol.* 43(1): 48-57.
- 3063.Narayanan, P. and Jaganathan, T. 1973. Vector transmission of black gram leaf crinkle virus. *Madras Agric. J.* 60: 651-652. [Cock (1986)]
- 3064.Naresh, J.S. and Nene, Y.L. 1980. Host range, host preference for oviposition and development and the dispersal of *Bemisia tabaci* Gennadius, a vector of several plant viruses. *Indian J. Agric. Sci.* 50: 620-623. [Cock (1986)]
- 3065.Naresh, J.S. and Thakur, R.P. 1972. Efficacy of systemic granular and spray insecticides for the control of insect pests of black-gram (*Phaseolus mungo* Roxb.). *Indian J. Agric. Sci.* 42: 732-735. [Cock (1986)]
- 3066.Nariani, T.K. 1960. Yellow mosaic of mung (*Phaseolus aureus* L.). *Indian Phytopathol.* 8: 24-29. [Cock (1986)]
- 3067.Nariko, S. 1991. Squash silver leaf by sweetpotato whitefly. *Ann. Phytopathol. Soc. Japan* 57: 216.
- 3068.Nasir, M.M. 1947. Biology of *Chrysopa selestes* Banks. *Indian J. Entomol.* 9: 177-192. [Cock (1986)]
- 3069.Nasir, M.M. 1947. *Chrysopa cybele* Banks and its two new varieties. *Indian J. Entomol.* 8: 119-120. [Cock (1986)]
- 3070.Nassef, M.A. 1999. Juvenile hormone mimic and plant-derived oils as control agents against whitefly, *Bemisia tabaci* (Genn.), on cotton. *Egyptian J. Agric. Res.* 77(2): 691-699.
- 3071.Nassef, M.A. and Watson, W.M. 1999. Sequential spray schedules of insecticides to control bollworms as target pests in addition to certain sap suckers as non-target pests in cotton fields. *Egyptian J. Agric. Res.* 77(3): 1155-1162.
- 3072.Natarajan, K. 1986. Influence of NPK fertilizer on the population density of cotton whitefly. pp. 134-136. *In* S. Jayaraj (Ed.), Resurgence of Sucking Pests: Proceedings of National Symposium. Centre For Plant Protection Studies, Tamil Nadu Agric. Univ.
- 3073.Natarajan, K. 1988. Transport of yellow mite *Polyphagotarsonemus latus* by cotton whitefly. *Curr. Sci. (Bangalore)* 57(20): 1142-1143. [Cock (1993)]
- 3074.Natarajan, K. 1990. Effect of leaf pubescence in cotton *Gossypium hirsutum* on the parasitism of whitefly *Bemisia tabaci* (Gennadius). *J. Biol. Control* 4(1): 57-58. [Cock (1993)]

- 3075.Natarajan, K. 1990. Natural enemies of *Bemisia tabaci* Gennadius and effect of insecticides on their activity. *J. Biol. Control* 4(2): 86-88. [Cock (1993)]
- 3076.Natarajan, K. and Sundaramurthy, V.T. 1988. Management of *Bemisia tabaci* in cotton system through host plant resistance. pp. 112-115. *In* N. Mohandas and G. Koshy (Eds.), Proceedings National Symposium on Integrated Pest Control: Progress and Perspectives, 15-17 October. Association for Advancement of Entomology, Trivandrum, Kerala, India.
- 3077.Natarajan, K. and Sundaramurthy, V.T. 1990. Effect of neem oil on cotton whitefly (*Bemisia tabaci*). *Indian J. Agric. Sci.* 60(4): 290-291.
- 3078.Natarajan, K., Sundaramurthy, V.T. and Basu, A.K. 1986. Meet the menace of whitefly to cotton. *Indian Farming* 36(4): 37,39,44. [Cock (1993)]
- 3079.Natarajan, K., Sundaramurthy, V.T. and Chidambaram, P. 1987. Whitefly and aphid resurgence in cotton as induced by certain insecticides. pp. 137-143. *In* S. Jayaraj (Ed.), Resurgence of Sucking Pests: Proceedings of National Symposium. Centre for Plant Protection Studies, Tamil Nadu Agric. Univ., Coimbatore, India.
- 3080.Natarajan, K., Sundaramurthy, V.T. and Chidambaram, P. 1991. Usefulness of fish oil rosin soap in the management of whitefly and other sap feeding insects of cotton. *Entomon* 16: 229-232.
- 3081.Natarajan, N., Rao, P.V.S. and Gopal, S. 1991. Effect of intercropping of pulses in cereals on the incidence of major pests. *Madras Agric. J.* 78(1/4): 59-67.
- 3082.Nateshan, H.M., Muniyappa, V., Jalikop, S.H. and Ramappa, H.K. 1996. Resistance of *Lycopersicon* species and hybrids to tomato leaf curl geminivirus. pp. 369-377. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia* : 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
- 3083.Nateshan, H.M., Muniyappa, V., Swanson, M.M. and Harrison, B.D. 1996. Host range, vector relations and serological relationships of cotton leaf curl virus from southern India. *Ann. Appl. Biol.* 128(2): 233-244.
- 3084.Nath, P. 1993. Monitoring of whitefly (*Bemisia tabaci*) population in mungbean yellow mosaic infected greengram (*Vigna radiata*) field. *Ann. Agric. Res.* 14(3): 360-362.
- 3085.Nath, P., Chaudhary, O.P., Sharma, P.D. and Kaushik, H.D. 2000. Studies on incidence of important insect-pests of cotton with special reference to *Gossypium arboreum* (desi) cotton. *Indian J. Entomol.* 62(4): 391-395.
- 3086.Nath, P. and Saikia, A.K. 1993. Assessment of yield loss due to yellow vein mosaic of bhendi (*Abelmoschus esculentus* (L.) Moench in Assam. *J. Agric. Sci. Soc. North East India* 6: 87-88.
- 3087.Nath, P. and Saikia, A.K. 1995. Influence of sowing time on yellow vein mosaic virus of okra. *Indian J. Mycol. Plant Pathol.* 25(3): 277-279.
- 3088.Nath, P.D. 1994. Effect of sowing time on the incidence of yellow mosaic virus disease and whitefly population on greengram. *Ann. Agric. Res.* 15(2): 174-177.
- 3089.Nath, P.D. 1994. Split cage for monitoring whitefly (*Bemisia tabaci*) in yellow mosaic infested greengram field. *Ann. Agric. Res.* 15(3): 371-373.
- 3090.Nath, P.D., Gupta, M.K. and Bora, P. 1992. Influence of sowing time on the incidence of yellow vein mosaic and whitefly population on okra. *Indian J. Virol.* 8: 45-48.
- 3091.Nath, P.D. and Saikia, A.K. 1995. Effect of time of sowing on the incidence of mungbean yellow mosaic virus disease and white fly (*Bemisia tabaci* Genn.) population in greengram. *Ann. Agric. Res.* 16(4): 483-484.
- 3092.Natwick, E., Durazo, A. III and Laemmlen, F. 1987. Row covers for insects and virus diseases protection in desert agriculture. *Proc. National Agric. Plastics Congress* 20: 159-169.
- 3093.Natwick, E., Turini, T., Cook, C., Gilbertson, R. and Seo, Y.S. 2001. Molecular techniques corroborate resistance to whitefly transmitted cotton leaf crumple disease. pp. 793-796. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3094.Natwick, E.T. 1984. Effects of Temick 15G on the sweetpotato whitefly *Bemisia tabaci* (Gennadius). *California Cotton Prog. Rep.* 59-60.
- 3095.Natwick, E.T. 1985. Whitefly and virus diseases continue to plague vegetable growers! *Valley Grower* (Fall): 9,11,12,14,16.
- 3096.Natwick, E.T. 1993. Silverleaf whitefly control in cotton using various insecticides in the Imperial Valley of California. pp. 722-727. *In* D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3097.Natwick, E.T. 1994. Silverleaf whitefly control in cotton using insecticides and an insect growth regulator. pp. 896-900. *In* D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3098.Natwick, E.T. 1995. Efficacy of selected insecticides for control of silverleaf whitefly in Imperial Valley cotton. pp. 863-866. *In* D.A. Richter and J. Armour (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3099.Natwick, E.T. 1999. New insecticides for control of silverleaf whitefly: an efficacy evaluation. pp. 919-921. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3100.Natwick, E.T., Chu, C.C., Cohen, A.C., Simmons, G.S. and Brushwood, D.E. 1997. Silverleaf whitefly infestation levels and percent parasitism in relation to cotton variety and insecticide treatments. pp. 915-918. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3101.Natwick, E.T., Chu, C.C. and Leimgruber, W. 1991. Cotton varietal susceptibility to whitefly infestation under low desert conditions. pp. 715-717. *In* D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3102.Natwick, E.T., Chu, C.C., Perkins, H.H., Henneberry, T.J. and Cohen, A.C. 1995. Pima and upland cotton susceptibility to *Bemisia argentifolii* under desert conditions. *Southwest. Entomol.* 20(4): 429-438.
- 3103.Natwick, E.T., Cook, C., Gilbertson, R., Seo, Y.S. and Turini, T. 2000. Resistance in upland cotton to the silverleaf whitefly transmitted cotton leaf crumple disease. pp. 164-167. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3104.Natwick, E.T., Cook, C.G. and Gilbertson, R.L. 1998. Silverleaf whitefly and cotton leaf crumple virus resistance screening in upland cotton. pp. 1091-1093. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3105.Natwick, E.T. and Deeter, B.D. 2001. Comparison of neonicotinoid with pyrethroid insecticides for control whitefly in cotton. pp. 906-908. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3106.Natwick, E.T. and Durazo, A., III. 1985. Polyester covers protect vegetables from whiteflies and virus diseases. *California Agric.* 39(7/8): 21-22. [Cock (1993)]
- 3107.Natwick, E.T., Durazo, A., III and Laemmlen, F. 1988. Direct row covers for insects and virus diseases protection in desert agriculture. *Plasticulture* 78: 35-46. [Cock (1993), English and French]
- 3108.Natwick, E.T. and Laemmlen, F.F. 1993. Protection from phytophagous insects and virus vectors in honeydew melons using row covers. *Florida Entomol.* 76: 120-126.
- 3109.Natwick, E.T. and Leigh, T.F. 1985. Whitefly problems affecting cotton production in the San Joaquin Valley and in southern California. *California Cotton Prog. Rep.* 63-67.

3110. Natwick, E.T., Leimgruber, W., Toscano, N.C. and Yates, L. 1992. Sampling adult sweetpotato whitefly in cotton. pp. 693-697. *In* D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
3111. Natwick, E.T., Mayberry, K. and Laemmlen, F. 1993. Silverleaf whitefly (formerly sweetpotato whitefly Strain B) Fact Sheet. Imperial County Coop. Extn., California, 8pp.
3112. Natwick, E.T., Palumbo, J.C. and Engle, C.E. 1996. Effects of imidacloprid on colonization of aphids and silverleaf whitefly and growth, yield and phytotoxicity in cauliflower. *Southwest. Entomol.* 21(3): 283-292.
3113. Natwick, E.T., Summers, C.G., Chu, C.C., Henneberry, T.J., Bell, C.E. and Godfrey, L.D. 2000. *Bemisia argentifolii* hosts in Imperial and southern San Joaquin Valleys, California. *Southwest. Entomol.* 25(4): 243-254.
3114. Natwick, E.T., Toscano, N.C. and Yates, L. 1995. Comparisons of adult whitefly sampling techniques in cotton with whitefly adult populations from whole plant samples. *Southwest. Entomol.* 20(1): 33-41.
3115. Natwick, E.T., Toscano, N.C. and Yates, L. 1996. Correlations of adult *Bemisia* sampling techniques in cotton to whole plants samples. pp. 247-252. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia*: 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
3116. Natwick, E.T. and Zalom, F.G. 1984. Surveying sweetpotato whitefly in the Imperial Valley. *California Agric.* 38 (3-4): 11. [Cock (1986)]
3117. Natwick, E.T. and Zalom, F.G. 1985. Verification of the cotton whitefly population model, *Bemisia tabaci* Gennadius (Homoptera, Aleyrodidae). pp. 174-177. *In* J.M. Brown and T.C. Nelson (Eds.), Proceedings Beltwide Cotton Production Research Conferences. National Cotton Council, Memphis, TN. [Cock (1993)]
3118. Natwick, E.T. and Zalom, F.G. 1987. Development of sweetpotato whitefly in clip cages on cotton. pp. 250-252. *In* J.M. Brown and T.C. Nelson (Eds.), Proceedings Beltwide Cotton Production Research Conferences. National Cotton Council, Memphis, TN.
3119. Natwick, E.T., Zalom, F.G., Toscano, N.C. and Kido, K. 1984. Monitoring of the cotton whitefly, *Bemisia tabaci* (Gennadius): Studies in the insect's development and control in cotton. pp. 197-202. *In* J.M. Brown (Ed.), Proceedings Beltwide Cotton Production Research Conferences. National Cotton Council, Memphis, TN.
3120. Nauen, R., Koob, B. and Elbert, A. 1998. Antifeedant effects of sublethal dosages of imidacloprid on *Bemisia tabaci*. *Entomol. Exp. Appl.* 88(3): 287-293.
3121. Nauen, R., Koob, B., Kluver, T. and Elbert, A. 1997. Biochemical characterization of insecticide resistant strains of the tobacco whitefly *Bemisia tabaci* (Homoptera: Aleyrodidae). *Proc. German Soc. Gen. Appl. Entomol.* 11(1-6): 217-221. [German, English summary]
3122. Nauen, R., Reckmann, U., Armbrorst, S., Stupp, H.P. and Elbert, A. 1999. Whitefly-active metabolites of imidacloprid: biological efficacy and translocation in cotton plants. *Pestic. Sci.* 55(3): 265-271.
3123. Nava-Camberos, U. 1996. Bionomics of *Bemisia argentifolii* Bellow and Perring on cotton, cantaloupe and pepper. [Texas A&M University]. Dissertation Abst. Int. 57(08B): 4854.
3124. Nava-Camberos, U. 1998. Disposición especial y muestreo de mosquitas blancas. [Spatial distribution and sampling of whitefly]. pp. 47-71. *In* J.J. Pacheco-Covarrubias and F. Pacheco-Mendivil (Eds.), *Temas Selectos Para El Manejo Integrado de la Mosquita Blanca*, Memoria Científica No. 6. Instituto Nacional De Investigaciones Forestales, Agrícolas y Pecuaria, Centro de Investigación Regional del Noroeste, CIRNO, Obregon, Sonora, Mexico.
3125. Nava-Camberos, U. 1998. Relaciones densidad-rendimiento y estimación de umbrales económicos para la mosquita blanca de la hoja plateada (*Bemisia argentifolii* Bellows & Perring). [Density-yield relationships and estimation of economic thresholds for silverleaf whitefly (*Bemisia argentifolii* Bellows & Perring)]. pp. 73-91. *In* J.J. Pacheco-Covarrubias and F. Pacheco-Mendivil (Eds.), *Temas Selectos Para El Manejo Integrado de la Mosquita Blanca*, Memoria Científica No. 6. Instituto Nacional De Investigaciones Forestales, Agrícolas y Pecuaria, Centro de Investigación Regional del Noroeste, CIRNO, Obregon, Sonora, Mexico. [Spanish]
3126. Nava-Camberos, U. and Cano Rios, P. 1998. Umbrales económicos para la "Mosquita blanca de la hoja plateada", *Bemisia argentifolii* Bellows and Perring, en melón. [Economic threshold for white fly of silver leaf, *Bemisia argentifolii* Bellows and Perring, in melon]. pp. 277-281. *In* Proceedings 33rd National Entomological Congress, Acapulco, Mexico, 24-27 May 1998. Sociedad Mexicana De Entomología. [Spanish]
3127. Nava-Camberos, U. and Cano-Rios, P. 2000. Economic threshold for the silverleaf whitefly in cantaloupe at the Comarca Lagunera, Mexico. *Agrociencia* 34(2): 227-234. [Spanish and English]
3128. Nava-Camberos, U., Riley, D.G. and Harris, M.K. 2001. Density-yield relationships and economic injury levels for *Bemisia argentifolii* (Homoptera: Aleyrodidae) in cantaloupe in Texas. *J. Econ. Entomol.* 94(1): 180-189.
3129. Nava-Camberos, U., Riley, D.G. and Harris, M.K. 2001. Temperature and host plant effects on development, survival, and fecundity of *Bemisia argentifolii* (Homoptera: Aleyrodidae). *Environ. Entomol.* 30(1): 55-63.
3130. Navas-Castillo, J., Camero, R., Bueno, M. and Moriones, E. 2000. Severe yellowing outbreaks in tomato in Spain associated with infections of Tomato chlorosis virus. *Plant Dis.* 84(8): 835-837.
3131. Navas-Castillo, J., Diaz, J.A., Sanchez-Campos, S. and Moriones, E. 1998. Short communication. Improvement of the print-capture polymerase chain reaction procedure for efficient amplification of DNA virus genomes from plants and insect vectors. *J. Virol. Methods* 75(2): 195-198.
3132. Navas-Castillo, J., Diaz, J.A., Sanchez-Campos, S., Moriones, E. and Barba, M. 1999. Detection of geminivirus genomes by print PCR. *Petria* 9(1-2): 175-178.
3133. Navas-Castillo, J., Sánchez-Campos, J.A., Díaz, E. and Sáez-Alonso, E.M. 1999. Tomato yellow leaf curl virus-Is causes a novel disease of common bean and severe epidemics in tomato in Spain. *Plant Dis.* 83: 29-32.
3134. Navon, A. and Melamed-Madjar, V. 1984. Honeydew staining techniques for estimating the live tobacco whitefly nymph population on cotton leaves. *Phytoparasitica* 12: 199-202. [Cock (1986)]
3135. Navon, A., Melamed-Madjar, V. and Zur, M. 1991. Effects of a primitive cotton accession on feeding of *Spodoptera littoralis* and *Helicoverpa armigera* and on oviposition of *Bemisia tabaci*. *Phytoparasitica* 19(2): 143-147. [Cock (1993)]
3136. Navon, A., Melamed-Madjar, V., Zur, M. and Benmoshe, E. 1991. Effects of cotton cultivars on feeding of *Heliothis armigera* and *Spodoptera littoralis* larvae and on oviposition of *Bemisia tabaci*. *Agric. Ecosystems Environ.* 35: 73-80.
3137. Navot, N., Ber, R. and Czosnek, H. 1989. Rapid detection of tomato yellow leaf curl virus in squashes of plants and insect vectors. *Phytopathology* 79(5): 562-568. [Cock (1993)]
3138. Navot, N., Picherski, E., Zeidan, M., Zamir, D. and Czosnek, H. 1991. Tomato yellow leaf curl virus: a whitefly-transmitted geminivirus with a single genomic component. *Virology* 185: 151-161.
3139. Navot, N., Zeidan, M., Picherski, E., Zamir, D. and Czosnek, H. 1992. Use of the polymerase chain reaction to amplify tomato yellow leaf curl virus DNA from infected plants and viruliferous whiteflies. *Phytopathology* 82(10): 1199-1202.
3140. Nawar, M.S. and El-Sjerif, A.A. 1993. *Neoseiulus cucumeris* (Oudemans), a predator of whitefly *Bemisia tabaci* (Gennadius). *Bull. Entomol. Soc. Egypt* 71: 9-17.

- 3141.Nazer, I. and Sharaf, N. 1982. Control of the immature stages of the tobacco whitefly (*Bemisia tabaci* gen.) by certain insecticides on tomatoes in the Jordan Valley. *Dirasat* 9(1): 183-190.
- 3142.Nazer, I.K. and Sharaf, N.S. 1985. Susceptibility of the sweetpotato whitefly (*Bemisia tabaci*, Genn) to certain insecticides. *Dirasat* 12(4): 151-159.
- 3143.Ndayiragije, P. and Autrique, A. 1992. [To a more integrated control of the principal pest insects of the cultures in Burundi]. pp. 439-445. *In* C. Verstraeten, P. Grootaert and G. Coulon (Eds.), Proceedings 3rd International Conference of the French-Speaking Entomologists. Methods and Responsibilities of the Entomologists Today, Gembloux, Belgium, 9-14 Jul 1990. Societe Royale Belge D'Entomologie, Bruxelles, Belgium. [French]
- 3144.Nderitu, J.H., Anyango, J.J. and Ampofo, J.K.O. 1997. A survey on insect pests and farmers' control measures on snap beans in Kirinyaga district, Kenya. CIAT African Occasional Publications Series. No. 23, 16 pp.
- 3145.Neal, J.W. and Bentz, J.A. 1999. Evidence for the stage inducing phenotypic plasticity in pupae of the polyphagous whiteflies *Trialeurodes vaporariorum* and *Bemisia argentifolii* (Homoptera: Aleyrodidae) and the *raison d'etre*. *Ann. Entomol. Soc. Am.* 92(6): 774-787.
- 3146.Neal, J.W., Davis, J.C., Bentz, J.A., Warthen, J.D., Griesbach, R.J. and Santamour, F.S. 1998. Allelochemical activity in *Ardisia* species (Myrsinaceae) against selected arthropods. *J. Econ. Entomol.* 91(3): 608-617.
- 3147.Neal, J.W., Jr., Buta, J.G., Pittarelli, G.W., Lusby, W.R. and Bentz, J.A. 1994. Novel sucrose esters from *Nicotiana glauca*: effective biorationals against selected horticultural insect pests. *J. Econ. Entomol.* 87(6): 1600-1607.
- 3148.Neal, J.W., Jr., Leonhardt, B.A., Brown, J.K., Bentz, J. and Devilbiss, E.D. 1994. Cuticular lipids of greenhouse whitefly and sweetpotato whitefly type A and B (Homoptera: Aleyrodidae) pupal exuviae on the same hosts. *Ann. Entomol. Soc. Am.* 87(5): 609-618.
- 3149.Nedstam, B. 1988. A new whitefly species, *Bemisia tabaci* (Homoptera: Aleyrodidae), in Swedish greenhouses. *Vaxtskyddsnotiser* 52: 71-72.
- 3150.Nedstam, B. 1989. Summary from one year of biocontrol in a small-scale mixed production of ornamentals. *Bull. OILB/SROP (IOBC/WPRS)* 12: 12-14.
- 3151.Negasi, A., Parker, B.L. and Brownbridge, M. 1998. Screening and bioassay of entomopathogenic fungi for the control of silverleaf whitefly, *Bemisia argentifolii*. *Insect Sci. Appl.* 18(1): 37-44.
- 3152.Nel, C. 1997. Whiteflies - farmers can help. *Farmer's Weekly (South Africa)* (17 October): 14-15.
- 3153.Nelson, D.R. and Buckner, J.S. 2000. Waxy particles - the 'overcoat' on adult whiteflies. pp. 1276-1281. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3154.Nelson, D.R., Buckner, J.S. and Fatland, C.L. 1994. The composition of external lipids from adult whiteflies, *Bemisia tabaci* and *Trialeurodes vaporariorum*. *Comp. Biochem. Physiol.* 109(2-3): 293-303.
- 3155.Nelson, D.R., Buckner, J.S., Freeman, T.P., Chu, C.C. and Henneberry, T.J. 2001. How adult whiteflies feed on cotton leaves. pp. 1182-1184. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3156.Nelson, J. and Parrella, M.P. 1993. Potential interference among natural enemies of *Bemisia tabaci*. *Bull. OILB/SROP (IOBC/WPRS)* 16: 121-124.
- 3157.Nelson, J.M., Nakayama, F.S., Flint, H.M., Garcia, R.L. and Hart, G.L. 1994. Methanol treatments on pima and upland cotton. pp. 1341-1342. *In* D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3158.Nelson, M., Orum, T., Byrne, D., El-Lissy, O., Antilla, L. and Staten, R. 1993. Preliminary investigation of sweet potato whitefly population dynamics across Arizona. pp. 197-205. *In* Cotton, A College of Agriculture Report, Series P-94. University of Arizona, Tucson.
- 3159.Nelson, M.R., Crum, T.V. and Nadeem, A. 1994. Regional analysis of epidemics of whitefly transmitted gemini viruses in cotton. pp. 270-271. *In* D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3160.Nelson, M.R., Nadeem, A., Ahmed, W. and Orum, T.V. 1998. Cotton virus diseases. pp. 584-586. *In* Cotton, A College of Agriculture Report, Series P-112. University of Arizona, Tucson.
- 3161.Nelson, M.R., Nadeem, A., Ahmed, W. and Orum, T.V. 1998. Global assessment of cotton virus diseases. pp. 161-162. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3162.Nene, S.R. 1987. An appraisal of cotton whitefly problem and for further research. pp. 90-94. *In* P. Jayaraj (Ed.), Resurgence of Sucking Pests: Proceedings of National Symposium. Centre for Plant Protection Studies, Tamil Nadu Agric. Univ., Coimbatore, India. [Cock (1986)]
- 3163.Nene, Y.L. 1972. A survey of viral diseases of pulse crops in Uttar Pradesh: final technical report. *Res. Bull., G. B. Pant Univ. Agric. Tech.* 4: 1-109,178-187.
- 3164.Nene, Y.L. 1973. Control of *Bemisia tabaci* Genn., a vector of several plant viruses. *Indian J. Agric. Sci.* 43: 433-436. [Cock (1986)]
- 3165.Nene, Y.L. 1973. Note on a fungus parasite of *Bemisia tabaci* Genn., a vector of several plant viruses. *Indian J. Agric. Sci.* 43: 514-516. [Cock (1986,1993)]
- 3166.Nene, Y.L. 1973. Viral diseases of some warm weather pulse crops in India. *Plant Dis. Rep.* 57: 463-467. [Cock (1986)]
- 3167.Nennmann, H. and Klatt, J. 1996. [White flies firmly in the grip. Useful insects protect enemies of *Euphorbia pulcherrima*]. *Gärtnerbörse und Gartenwelt (Germany)* 96(24): 1126-1127. [German]
- 3168.Nerkar, Y.S. 1991. The use of related species in transferring disease and pest resistance genes to okra. pp. 110-113. *In* International Workshop on Okra Genetic Resources, New Delhi, India, 8-12 October 1990. International Crop Network Series 5, Rome, Italy.
- 3169.Nguyen, R. and Bennett, F.D. 1995. Importation and field release of parasites against silverleaf whitefly, *Bemisia argentifolii* (Bellows and Perring) in Florida from 1990-1994. *Proc. Florida State Hort. Soc.* 108: 43-47.
- 3170.Nibouche, S. 1992. Mites, diplopods and phytophagous insects associated with cotton cultivation in Burkina Faso. [French summary]. *Coton Fibres Trop.* 47(4): 305-311.
- 3171.Nibouche, S., Chazeaux, R., Deguine, J.P., Martin, J. and Vaissayre, M. 1998. Damage caused by *Bemisia tabaci* (Gennadius) in cotton crops: Recent developments in West Africa. *Agric. Devel.* 20: 13-18. [French, English summary]
- 3172.Nichols, R., Ellsworth, P. and Dennehy, T. 1996. Insect growth regulators for whitefly management. p. 153. *In* P. Dugger and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3173.Nichols, R.L. 1994. Getting a handle on whiteflies. pp. 9-12. *In* D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3174.Nichols, R.L., Miller, W.B., Mysore, K. and Perkins, H.H., Jr. 1998. Honeydew sugar estimates differ among reducing-sugar test methods. pp. 1547-1549. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3175.Nicholson, W.F., Senn, R., Fluckiger, C.R. and Fuog, D. 1996. Pymetrozine - a novel compound for control of whiteflies. pp. 635-639. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia* : 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.

3176. Nicoli, G. and Burgio, G. 1997. Mediterranean biodiversity as source of new entomophagous species for biological control in protected crops. *Bull. OILB/SROP (IOBC/WPRS)* 20(4): 27-38.
3177. Nimbalkar, S.A., Khodke, S.M., Taley, Y.M. and Patil, K.J. 1993. Bioefficacy of some new insecticides including neem seed extract and neem oil for control of whitefly, *Bemisia tabaci* Genn. on cotton. pp. 256-260. *In* Botanical Pesticides in Integrated Pest Management. Indian Society of Tobacco Sciences.
3178. Nitzany, F.E. 1975. Tomato yellow leaf curl virus. *Phytopathol. Mediterr.* 14: 127-129.
3179. Nitzany, F.E., Geisenberg, H. and Koch, B. 1964. Tests for the protection of cucumbers from a white fly-borne virus. *Phytopathology* 54: 1059-1061. [Cock (1986)]
3180. Nkouka, N., Onore, G. and Fabres, G. 1981. Elements d'un inventaire de l'entomofaune phytophage du manioc en vue de l'identification des insectes vecteurs de la bacteriose vasculaire. *Cahiers ORSTOM, Serie Biologie* 44: 9-10. [Cock (1986)]
3181. Nombela, G., Beitia, F. and Muniz, M. 2000. Variation in tomato host response to *Bemisia tabaci* (Hemiptera : Aleyrodidae) in relation to acyl sugar content and presence of the nematode and potato aphid resistance gene Mi. *Bull. Entomol. Res.* 90(2): 161-167.
3182. Nombela, G., Beitia, F. and Muniz, M. 2001. A differential interaction study of *Bemisia tabaci* Q-biotype on commercial tomato varieties with or without the Mi resistance gene, and comparative host responses with the B-biotype. *Entomol. Exp. Appl.* 98(3): 339-344.
3183. Nomikou, M., Janssen, A., Schraag, R. and Sabelis, M.W. 2001. Phytoseiid predators as potential biological control agents for *Bemisia tabaci*. *Exp. Appl. Acarol.* 25(4): 271-291.
3184. Nordlund, D.A. and Legaspi, J.C. 1996. Whitefly predators and their potential for use in biological control. pp. 499-513. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia : 1995 Taxonomy, Biology, Damage, Control and Management*. Intercept Ltd., Andover, Hants, UK.
3185. Noris, E., Accotto, G.P., Tavazza, R., Brunetti, A., Crespi, S. and Tavazza, M. 1996. Resistance to tomato yellow leaf curl geminivirus in *Nicotiana benthamiana* plants transformed with a truncated viral C1 gene. *Virology* 224: 130-138.
3186. Noris, E., Hidalgo, E., Accotto, G.P. and Moriones, E. 1994. High similarity among the tomato yellow leaf curl virus isolates from the West Mediterranean Basin: the nucleotide sequence of an infectious clone from Spain. *Arch. Virol.* 135: 165-170.
3187. Noris, E., Vaira, A.M., Caciagli, P., Masenga, V., Gronenborn, B. and Accotto, G.P. 1998. Amino acids in the capsid protein of tomato yellow leaf curl virus that are crucial for systemic infection, particle formation, and insect transmission. *J. Virol.* 72(12): 10050-10057.
3188. Norman, J.W., Jr., Riley, D.G., Sparks, A.N., Jr., and Lester, J.F. 1993. Texas suggestions for managing sweetpotato whitefly and aphids in cotton 1993. pp. 36-37. *In* D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
3189. Norman, J.W., Jr., Riley, D.G., Stansly, P.A., Ellsworth, P.C. and Toscano, N.C. 1995. Management of silverleaf whitefly: a comprehensive manual on the biology, economic impact and control tactics. U.S. Dept. Agric./Coop. State Res. Ext. Education Serv., Grant Publ. 93-EPIX-1-0102, 13 pp.
3190. Norman, J.W., Jr. and Sparks, A.N., Jr. 1996. Silverleaf whiteflies and cotton leaf hairs, lower Rio Grande Valley, 1994-1995. pp. 1026-1027. *In* P. Dugger and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
3191. Norman, J.W., Jr. and Sparks, A.N., Jr. 1997. Cotton leafhairs and silverleaf whiteflies in the lower Rio Grande Valley of Texas three year research summary. pp. 1063-1064. *In* P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
3192. Norman, J.W., Jr., Sparks, A.N., Jr. and Riley, D. 1992. Sweetpotato whiteflies in Lower Rio Grande Valley cotton. pp. 687-690. *In* D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
3193. Norman, J.W., Jr., Sparks, A.N., Jr. and Riley, D.G. 1994. Keys to management of SPWF. pp. 144-145. *In* D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
3194. Norman, J.W., Jr., Sparks, A.N., Jr. and Riley, D.G. 1995. Impact of cotton leaf-hairs and whitefly populations on yields in the Lower Rio Grande Valley. pp. 102-104. *In* D.A. Richter and J. Armour (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
3195. Norman, J.W., Jr., Sparks, A.N., Jr. and Riley, D.G. 1996. Whiteflies in the Lower Rio Grande Valley of Texas in 1995. pp. 127-128. *In* P. Dugger and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
3196. Nottingham, S.F., Chortyk, O.T. and Stephenson, M.G. 1996. Sugar esters from *Nicotiana* species as potential insecticides against the sweetpotato whitefly (Homoptera: Aleyrodidae). *J. Entomol. Sci.* 31(3): 331-339.
3197. Noueiry, A.O., Lucas, W.J. and Gilbertson, R.L. 1994. Two proteins of a plant DNA virus coordinate nuclear and plasmodesmal transport. *Cell* 76: 925-932.
3198. Nour, M.A. 1960. On "leaf curl" of cotton in the Philippines. *FAO Plant Prot. Bull.* 8(5): 55-56. [Cock (1986)]
3199. Nour, M.A. and Nour, J.J. 1962. A mosaic disease of *Dolichos lablab* and diseases of other crops caused by alfalfa mosaic virus in the Sudan. *Phytopathology* 52: 427-432. [Cock (1986)]
3200. Nour, M.A. and Nour, J.J. 1964. Identification, transmission and host range of leaf curl viruses infecting cotton in the Sudan. *Empire Cotton Growing Rev.* 41: 27-37. [Cock (1986)]
3201. Nousier, N.I.M. 1988. Survey of some vegetable insect pests with biological and ecological studies on certain species. Ph.D. Dissertation, Alexandria University, Egypt, 139 pp.
3202. Nucifora, A., Mauromicale, G., Sortino, O. and Davino, M. 1992. Effetti dell'impiego di bioreti sulla dinamica di infestazione di *Bemisia tabaci* - sulle infezioni da TYLCV e sulla produzione di pomodoro. [Effects of the use of the network protection system on *Bemisia tabaci* activity - infection by TYLCV and tomato production]. *Culture Protette (Italy)* 21(12): 59-63. [Italian]
3203. Nucifora, M.T. 1992. Infezione da TYLCV, trasmessa da *Bemisia tabaci*, su alcuni ibridi di pomodoro. [TYLCV infection transmitted from *Bemisia tabaci* on some hybrids of tomato]. *Culture Protette (Italy)* 21(12): 69-73. [Italian]
3204. Nucifora, S. 1994. Mallas antiviral para la defensa del cultivo de tomate protegido contra *Bemisia tabaci*, vector del TYLCV (virus del rizado amarillo del tomate). [Antiviral mesh for defense of tomatoes protected cultivation against *Bemisia tabaci*, vector of TYLCV (tomato yellow leaf curl virus)]. *Phytoma (España)* 57: 40-48. [Spanish]
3205. Nuessly, G.S., Henneberry, T.J. and Perkins, H.H., Jr. 1989. Effect of sweetpotato whitefly population density on cotton fiber stickiness and reducing sugars. pp. 281-284. *In* J.M. Brown and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Production Research Conferences*. National Cotton Council, Memphis, TN.
3206. Nuessly, G.S., Meyerdirk, D.E., Coudriet, D.L. and Henneberry, T.J. 1994. The effect of short season cotton production schedules on *Bemisia tabaci* (Gennadius). *Southwest. Entomol.* 19(3): 209-217.
3207. Nuessly, G.S., Meyerdirk, D.E., Hart, W.G. and Davis, M.R. 1987. Evaluation of color infrared aerial photography as a tool for the identification of sweetpotato whitefly induced fungal and viral infestations of cotton and lettuce. pp. 141-148. *In* J.H. Everett and P.R. Nixon (Eds.), *Eleventh Annual Workshop on Color Aerial Photography and Videography in the Plant Sciences and Related Fields*, April 27 - May 1, 1987, Weslaco, TX. Am. Soc. Photogrammetry and Remote Sensing, Falls Church, Virginia.
3208. Nuessly, G.S. and Perring, T.M. 1995. Influence of endosulfan on *Bemisia tabaci* (Homoptera: Aleyrodidae) populations, parasitism, and lettuce infectious yellows virus in late-summer planted cantaloupe. *J. Entomol. Sci.* 30(1): 49-61.

- 3209.Nuez, F. and Esteva, J. 1992. Virus de las cucurbitáceas transmitidos por aleurodíidos. [Cucurbitaceae viruses transmitted by Aleyrodidae]. *Phytoma (España)* 42: 31-36. [Spanish]
- 3210.Nyirenda, G.K.C. 1982. Daily application of ultra-low-volume (UVL) insecticides at low dosages to control insect pests of cotton in Malawi. *Crop Prot.* 1: 213-220. [Cock (1986)]
- 3211.O'Neil, R.J., Giles, K.L., Obyrcki, J.J., Mahr, D.L., Legaspi, J.C. and Katovich, K. 1998. Evaluation of the quality of four commercially available natural enemies. *Biol. Control* 11(1): 1-8.
- 3212.Oatman, E.R. 1970. Parasitization of whiteflies on strawberry plants in southern California. *J. Econ. Entomol.* 63(4): 1377-1378. [Cock (1986)]
- 3213.Obando, A., Darby, N. and Navarro, L. 1996. Chemical control of silverleaf whitefly in cotton, with karate in mixture with conventional insecticides in Mexico. pp. 1027-1029. *In* P. Dugger and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
- 3214.Ochou, G.O., Matthews, G.A. and Mumford, J.D. 1998. Farmers' knowledge and perception of cotton insect pest problems in Cote d'Ivoire. *Int. J. Pest. Manage.* 44(1): 5-9.
- 3215.Oetting, R.D. and Anderson, A.L. 1990. Imidacloprid for control of whiteflies, *Trialeurodes vaporariorum* and *Bemisia tabaci*, on greenhouse grown poinsettias. pp. 367-372. *In* British Crop Protection Conference: Pests and Diseases. Brighton Crop Protection Council, Thornton Heath, UK. [Cock (1993)]
- 3216.Oetting, R.D. and Buntin, G.D. 1996. *Bemisia* damage expression in commercial greenhouse production. pp. 201-208. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia: 1995 Taxonomy, Biology, Damage, Control and Management*. Intercept Ltd., Andover, Hants, UK.
- 3217.Ogata, Y. 1999. Integration of biological and chemical control in case of Japan. *Bull. OILB/SROP (IOBC/WPRS)* 22: 189-192.
- 3218.Ohnesorge, B. 1981. Untersuchungen zur Populationsdynamik der Weissen Fliege *Bemisia tabaci* Genn. in den Wintermonaten. *Mitteilungen der Deutschen Gesellschaft für Allgemeine und Angewandte Entomol.* 3: 324-327. [Cock (1986), German, English summary]
- 3219.Ohnesorge, B. and Rapp, G. 1986. Methods for estimating the density of whitefly nymphs (*Bemisia tabaci* Genn.) in cotton. [Spanish & French summaries]. *Trop. Pest Manage.* 32(3): 207-211. [Cock (1993)]
- 3220.Ohnesorge, B. and Rapp, G. 1986. Monitoring *Bemisia tabaci*: a review. *Agric. Ecosystems Environ.* 17(1-2): 21-27. [Cock (1993)]
- 3221.Ohnesorge, B., Sharaf, N. and Allawi, T. 1980. Population studies on the tobacco whitefly *Bemisia tabaci* Genn. (Homoptera, Aleyrodidae) during the winter season. I. The spatial distribution on some host plants. *Z. Angew. Entomol.* 90: 226-232. [Cock (1986)]
- 3222.Ohnesorge, B., Sharaf, N. and Allawi, T. 1981. Population studies on the tobacco whitefly *Bemisia tabaci* Genn. (Homoptera, Aleyrodidae) during the winter season. II. Some mortality factors of the immature stages. *Z. Angew. Entomol.* 92: 127-136. [Cock (1986)]
- 3223.Ohno, I. 1992. Whiteflies problem in the United States of America. *Japan Pestic. Information* 60: 19-20.
- 3224.Ohta, M., Ozawa, A. and Kobayashi, H. 1998. Efficacy of *Paecilomyces fumosoroseus* preparation against whitefly on tomato in the greenhouse [Japanese, English summary]. *Proc. Kanto-Tosan Plant Prot. Soc.* 45: 181-184.
- 3225.Ohto, K. 1990. Occurrence of the sweetpotato whitefly, *Bemisia tabaci* (Gennadius), on the poinsettia. *Plant Prot. (Japan)* 44(6): 264-266. [Japanese]
- 3226.Okada, T., Kawamura, M. and Kobayashi, H. 1998. A broad mite, *Polyphagotersonemus latus*, adhering to legs of whiteflies, *Bemisia tabaci* and *B. argentifolii*. [Japanese, English summary]. *Rostria* 47: 48.
- 3227.Okoth, V.A.G. 1991. Whitefly as a vector of African cassava mosaic virus (ACMV). *Ministry Agric., Animal Industry and Fisheries, Namulonge Res. Stn.*, 1 p.
- 3228.Olivares, F.M., Jr. and San Juan, M.O. 1966. The transmission, virus-vector relationship and host range of tobacco leaf curl virus. pp. 283-299. *In* Eleventh Pacific Science Congress. Japan Plant Protection Association, Tokyo. [Cock (1986)]
- 3229.Oliveira, M.A.S., Icuma, I.M., Alves, R.T., Oliveira, J.N.S., Oliveira, M.R.V., Lima, L.H.C. and Lira, G.S. 2000. Avaliação de surtos de mosca-branca em áreas do sistema produtivo de melão, soja e feijão. *EMBRAPA Recursos Genéticos e Biotecnologia, Comunicado Técnico No. 29*, 10 pp. [Portuguese]
- 3230.Oliveira, M.R.V. 1995. O emprego de casas de vegetação no Brasil: vantagens e desvantagens [The use of greenhouses in Brazil: advantages and disadvantages]. *Pesquisa Agropecuária Brasileira* 30(8): 1049-1060. [Portuguese, English summary]
- 3231.Oliveira, M.R.V. and DeLima, L.H.C. 1997. Padrões isoenzimáticos de *Trialeurodes vaporariorum* e *Bemisia tabaci* (Homoptera, Aleyrodidae) e de *Encarsia formosa* e *E. lycopersici* (Hymenoptera, Aphelinidae). [Isoenzyme patterns of *Trialeurodes vaporariorum* and *Bemisia tabaci* (Homoptera, Aleyrodidae) and of *Encarsia formosa* and *E. lycopersici* (Hymenoptera, Aphelinidae)]. *Pesquisa Agropecuária Brasileira* 32(7): 683-687. [Portuguese, English summary]
- 3232.Oliveira, M.R.V. and Faria, M.R. 2000. Mosca branca do complexo *Bemisia tabaci* (Gennadius) (Homoptera, Aleyrodidae): bioecologia e medidas de controle. *EMBRAPA Recursos Genéticos e Biotecnologia Documentos No. 48*, 111 pp. [Portuguese]
- 3233.Oliveira, M.R.V., Fernandes, E.R. and Rocha, H.G.C. 1999. I. Alternativas ao controle da mosca branca, *Bemisia tabaci* raça B, em plantas de melão. *Pesquisa em Andamento, Empresa Brasileira de Pesquisa Agropecuária, EMBRAPA No. 23*, 14 pp. [Portuguese]
- 3234.Oliveira, M.R.V., Henneberry, T.J. and Anderson, P. 2001. History, current status, and collaborative research projects for *Bemisia tabaci*. *In* S. E. Naranjo and P. C. Ellsworth (Eds.), *Special Issue: Challenges and Opportunities for Pest Management of Bemisia tabaci in the New Century*. *Crop Prot.* 20(9): 709-723.
- 3235.Oliveira, M.R.V., Lima, L.H.C. and Ferreira, L.T. 1997. Análise eletroforética de populações da mosca-branca, *Bemisia tabaci* raça B (= *B. argentifolii*) (Homoptera, Aleyrodidae). *Pesquisa em Andamento, Empresa Brasileira de Pesquisa Agropecuária, EMBRAPA No. 11*, 6 pp. [Portuguese]
- 3236.Oliveira, M.R.V., Lima, L.H.C., Návía, D. and Vieira, P.R.G. 1998. Avaliação das populações de *Bemisia tabaci* (Gennadius) a través de RAPD-PCR, no Brasil. *Pesquisa em Andamento, Empresa Brasileira de Pesquisa Agropecuária, EMBRAPA No. 18*, 6 pp. [Portuguese]
- 3237.Oliveira, M.R.V. and Silva, O.L.R. 1997. Alerta fitossanitário. prevenção e controle da mosca branca, *Bemisia argentifolii* (Homoptera, Aleyrodidae). *Brasília Ministério da Agricultura e do Abastecimento, Departamento de Defesa e Inspeção Vegetal*, 16 pp.
- 3238.Oliveira, M.R.V., Tigano, M.S. and Aljanabi, S. 2000. Molecular characterization of whitefly (*Bemisia* spp.) in Brazil. *Pesquisa Agropecuária Brasileira* 35(6): 1261-1268.
- 3239.Olivera, H., Lastres, N., Chiang, M.L., Gonzalez, M. and Zayas, M.A. 1998. Manejo del cultivo del tomate (*Lycopersicon esculentum* Mill) en primavera-verano frente al TYLCV y su insecto vector. [Management of tomatoes crop (*Lycopersicon esculentum* Mill) a spring-summer in front of TYLCV and its vector insect]. pp. 102-104. *In* Produccion De Cultivos En Condiciones Tropicales. Taller Internacional "Geminivirus en el Caribe", Habana, Cuba, Nov 1998. Instituto De Investigaciones Hortícolas, La Habana, Cuba. [Spanish, English summary]
- 3240.Olson, D.L. and Oetting, R.D. 1999. The efficacy of mycoinsecticides of *Beauveria bassiana* against silverleaf whitefly (Homoptera: Aleyrodidae) on poinsettia. *J. Agric. Urban Entomol.* 16(3): 179-185.
- 3241.Oltman, D. 1991. In the wake of the whitefly. *California Farmer (Dec. 7)*: 6-7,26,35.

- 3242.Omar, H.I., Hegab, M.F.A. and El-Hamaky, M.A. 1988. Combined action of some insecticides and fertilizers on pests infesting sweet melon. [Arabic summary]. *Agric. Res. Rev.* 66(1): 85-90. [Cock (1993)]
- 3243.Omar, H.I.H., Haydar, M.F. and El-Sorady, A.E.M. 1994. Effect of sowing date of intercropping cowpea with cotton on infestation with some major pests. *Egyptian J. Agric. Res.* 72(3): 691-698.
- 3244.Omer, A.D., Johnson, M.W., Tabashnik, B.E., Costa, H.S. and Ullman, D.E. 1993. Sweetpotato whitefly resistance to insecticides in Hawaii: Intra-island variation is related to insecticide use. *Entomol. Exp. Appl.* 67(2): 173-182.
- 3245.Omer, A.D., Tabashnik, B.E., Johnson, M.W., Costa, H.S. and Ullman, D.E. 1993. Genetic and environmental influences on susceptibility to acephate in sweetpotato whitefly (Homoptera, Aleyrodidae). *J. Econ. Entomol.* 86(3): 652-659.
- 3246.Omran, H.H. and El-Khidir, E. 1978. Über die Bevorzugung von Blattlaaren zur Eiablage bei *Bemisia tabaci* (Genn.) (Hom., Aleyrodidae). [On the preference of leaf hair sites for egg laying by the cotton whitefly]. *Anz. Schadlingskunde, Pflanzenschutz, Umweltschutz* 51(11): 175. [Cock (1986)]
- 3247.Ondieki, J.J. 1975. Diseases and pests of passion fruit in Kenya. *Acta Hort.* 49: 291-293. [Cock (1986)]
- 3248.Onillon, J.C. 1990. The use of natural enemies for the biological control of whiteflies. pp. 287-313. *In* Gerling, D. (Ed.), *Whiteflies: their Bionomics, Pest Status, and Management*. Intercept, Andover, UK.
- 3249.Onillon, J.C. and Maignet, P. 2000. Native parasitoids of the B-biotype of *Bemisia tabaci* Gennadius (Homoptera: Aleyrodidae). What can be expected from them for the biological control of the pest? *Bull. OILB/SROP (IOBC/WPRS)* 23(1): 101-107. [French, English summary]
- 3250.Onillon, J.C., Maignet, P. and Cocquempot, C. 1994. Premiers resultats sur l'efficacite d'*Encarsia pergandiella* (Hymenopt., Aphelinidae) dans le controle de *Bemisia tabaci* (Homopt., Aleyrodidae) en serres de tomate. [First results on *Encarsia pergandiella* (Hymenopt., Aphelinidae) efficiency in the control of *Bemisia tabaci* (Homopt., Aleyrodidae) in greenhouse tomato crops]. *Bull. OILB/SROP (IOBC/WPRS)* 17: 71-80. [French, English summary]
- 3251.Onillon, J.C., Polaszek, A., Cocquempot, C. and Maignet, P. 1994. Premieres observations sur l'entomofaune parasitaire infeodee a *Bemisia tabaci* (Homopt., Aleyrodidae) dans le sud-est de la France. [First observations on *Bemisia tabaci* (Homopt., Aleyrodidae) parasitic entomofauna in Southeastern France]. *Bull. OILB/SROP (IOBC/WPRS)* 17: 97-69. [French, English summary]
- 3252.Oñoro, P.R. 1996. Diseño y análisis estadístico. pp. 112-126. *In* L. Hilje (Ed.), *Metodologias para el estudio y manejo de moscas blancas y geminivirus*. Centro Agronomico Tropical de Investigacion y Enseñanza, Turrialba, Costa Rica.
- 3253.Onuki, M. and Hanada, K. 1998. PCR amplification and partial nucleotide sequences of three dicot-infesting geminiviruses occurring in Japan. *Ann. Phytopathol. Soc. Japan* 64(2): 116-120.
- 3254.Onuki, M. and Hanada, K. 2000. A rapid and simple procedure for distinguishing geminivirus-infected viruliferous silverleaf whiteflies by print-capture PCR (P-PCR). *Proc. Assoc. Plant Prot. Kyushu (Japan)* 46: 54-57. [Japanese]
- 3255.Opoku-Asiama, Y. and Oduro-Owusu, E. 1996. A survey of the incidence and severity of *Nesidiocoris tenuis*, *Bemisia tabaci* and tomato leaf curl virus in tomato growing areas in Ghana. 4th National Workshop on Improving Farming Systems in the Savannah Zone, Nyankpala, Tamale, Ghana, 28 pp.
- 3256.Organisation Europeenne et Mediterraneenne pour la Protection des Plantes. 1989. EPPO data sheets on quarantine organisms, no. 178: *Bemisia tabaci* (Gennadius) (Hemiptera: Aleyrodidae). *Bull. OEPP* 19(4): 733-756.
- 3257.Organisation Europeenne et Mediterraneenne pour la Protection des Plantes. 1990. Insecticide acaricide susceptibility tests. IRAC method No. 8 *Bemisia tabaci* (adults); amitraz. *Bull. OEPP* 20(2): 400-401.
- 3258.Orlando, A. and Silberschmidt, K. 1946. Estudos sobre a disseminacao natural do virus da "clorose infecciosa" das malvaceas (Abutilon virus 1. Baur) e a sua relacao com o inseto-vetor *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae). [English summary]. *Arch. Inst. Biol.* 17: 1-36. [Cock (1986)]
- 3259.Orozco, M., Lopez, O., Perez, O. and Delgado-Sanchez, F. 1994. Effect of transparent mulch, floating row covers and oil sprays on insect populations, virus diseases and yield of cantaloup. *Biol. Agric. Hortic.* 10(4): 229-234.
- 3260.Orozco-Santos, M., Farias-Larios, J., López-Pérez, J. and Ramírez-Vázquez, N.R. 2000. Uso de *Beauveria bassiana* para el control de *Bemisia argentifolii* en melón. [Use of *Beauveria bassiana* for the control of *Bemisia argentifolii* in melon]. *Manejo Integrado de Plagas (Costa Rica)* 56: 45-51. [Spanish, English summary]
- 3261.Orozco-Santos, M., Perez-Zamora, O. and Lopez-Arriaga, O. 1995. Effect of transparent mulch on insect populations, virus diseases, soil temperature, and yield of cantaloup in a tropical region. *New Zealand J. Crop Hortic. Sci.* 23(2): 199-204.
- 3262.Orozco-Santos, M., Perez-Zamora, O. and Lopez-Arriaga, O. 1995. Floating row cover and transparent mulch to reduce insect populations, virus diseases and increase yield in cantaloupe. *Florida Entomol.* 78(3): 493-501.
- 3263.Orozco-Santos, M., Robles-Gonzalez, M. and Farias-Larios, J. 1995. Some host plants of *Bemisia* (Homoptera, Aleyrodidae) under mesh-shade house conditions in Colima, Mexico. *Southwest. Entomol.* 20(1): 111-112.
- 3264.Osaki, T. and Inouye, T. 1978. Resemblance in morphology and intranuclear appearance of viruses isolated from yellow dwarf diseased tomato and leaf curl diseased tobacco. *Ann. Phytopathol. Soc. Japan* 44: 167-178. [Cock (1986)]
- 3265.Osaki, T. and Inouye, T. 1991. Transmission characteristics and cytopathology of a whitefly-transmitted virus isolated from sweet potato leaf curl disease. *Bull. Univ. Osaka Prefecture. Series B, Agric. Bio.* 43: 11-19. [Cock (1993)]
- 3266.Osaki, T., Kobatake, H. and Inouye, T. 1979. Yellow vein mosaic of honeysuckle (*Lonicera japonica* Thunb.), a disease caused by tobacco leaf curl virus in Japan. *Ann. Phytopathol. Soc. Japan* 45: 62-69. [Cock (1986)]
- 3267.Osborne, L.S. 1988. The not so sweet sweetpotato whitefly. *Florida Foliage (May)*: 8-10, 12,14,15.
- 3268.Osborne, L.S., Hoelmer, K. and Gerling, D. 1990. Prospects for biological control of *Bemisia tabaci*. *Bull. OILB/SROP (IOBC/WPRS)* 13: 153-160.
- 3269.Osborne, L.S. and Landa, Z. 1992. Biological control of whiteflies with entomopathogenic fungi. *Florida Entomol.* 75: 456-471.
- 3270.Osborne, L.S. and Landa, Z. 1994. Utilization of entomogenous fungus *Paecilomyces fimosorus* against sweetpotato whitefly, *Bemisia tabaci*. *Bull. OILB/SROP (IOBC/WPRS)* 17(3): 201-206.
- 3271.Oschek, W. 1989. Possibilities for controlling the whitefly on poinsettias. *Zierpflanzenbau* 29(18): 744-745.
- 3272.Osman, A.A., Attia, M.B., Darwish, E.T.E. and Anderyas, M.S. 1990. Biological control studies on *Aphis gossypii*, Glover and *Bemisia tabaci*, Genn. *Minufiya J. Agric. Res. (Egypt)* 15(1): 1023-1034.
- 3273.Osman, A.A., Attia, M.B., Darwish, E.T.E. and Anderyas, M.S. 1990. Potency of some predacious mites as biological control of certain cotton piercing sucking insects. *Minufiya J. Agric. Res. (Egypt)* 15(1): 1015-1022.
- 3274.Ota, M. and Ozawa, A. 1997. Vertical dispersion of the silverleaf whitefly, *Bemisia argentifolii* (Bellows & Perring) in field. *Proc. Kanto-Tosan Plant Prot. Soc.* 44: 229-230. [Japanese]
- 3275.Ota, M., Ozawa, A. and Kobayashi, H. 1999. Efficacy of *Beauveria bassiana* preparation against whitefly on tomato. *Annu. Rep. Kanto Tosan Plant Prot. Soc.* 46: 109-112. [Japanese, English summary]
- 3276.Otanes, F.Q. and Butac, F.L. 1935. A preliminary study of the insect pests of cotton in the Philippines with suggestions for their control. *Philippine J. Agric.* 6: 147-174. [Cock (1986)]



- 3277.Otanes, F.Q. and Butac, F.L. 1939. Cotton pests in the Philippines. *Philippine J. Agric.* 10: 341-371. [Cock (1986)]
- 3278.Otim-Nape, G.W., Alicai, T. and Thresh, J.M. 2001. Changes in the incidence and severity of Cassava mosaic virus disease, varietal diversity and cassava production in Uganda. *Ann. Appl. Biol.* 138(3): 313-327.
- 3279.Otim-Nape, G.W. and Ingoot, D. 1987. Effect of cultural practices on the African cassava mosaic disease and its vector, *Bemisia tabaci*. *Int. Dev. Res. Center IDRC-258e*: 105-108. [Cock (1993)]
- 3280.Otim-Nape, G.W., Shaw, M.W. and Thresh, J.M. 1994. The effects of African cassava mosaic geminivirus on the growth and yield of cassava in Uganda. *Trop. Sci.* 34(1): 43-54.
- 3281.Otim-Nape, G.W., Thresh, J.M., Bua, A., Baguma, Y. and Shaw, M.W. 1998. Temporal spread of cassava mosaic virus disease in a range of cassava cultivars in different agro-ecological regions of Uganda. *Ann. Appl. Biol.* 133(3): 415-430.
- 3282.Otim-nape, G.W., Thresh, J.M. and Fargette, D. 1996. *Bemisia tabaci* and cassava mosaic virus disease in Africa. pp. 319-350. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia* : 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
- 3283.Otim-Nape, G.W., Thresh, J.M. and Shaw, M.W. 1998. The incidence and severity of cassava mosaic virus disease in Uganda: 1990-1992. *Trop. Sci.* 38(1): 25-37.
- 3284.Ounchaichon, K., Rumakom, M. and Wangboonkong, S. 1990. Optimization of foliar-applied formulations for the control of tobacco whitefly *Bemisia tabaci*, under field conditions. pp. 75-84. *In* Agricultural Biotechnology Conference, Bangkok, Thailand, 16-18 Aug 1990.
- 3285.Ozawa, A., Satou, M. and Masuda, T. 1992. Parasitism of an indigenous parasitoid, *Encarsia transvena* (Timberlake) on the sweetpotato whitefly, *Bemisia tabaci* Gennadius, in sweetpotato fields. *Proc. Kanto-Tosan Plant Prot. Soc.* 39: 199-200. [Japanese, English summary]
- 3286.Ozgür, A.F. and Sekeroglu, E. 1986. Population development of *Bemisia tabaci* (Homoptera: Aleyrodidae) on various cotton cultivars in Cukurova, Turkey. *Agric. Ecosystems Environ.* 17(1-2): 83-88. [Cock (1993)]
- 3287.Ozgür, A.F., Sekeroglu, E., Gençer, O., Göçmen, H., Yelin, D. and Isler, N. 1988. [Study of population development of important cotton pests in relation to various cotton varieties and plant phenology]. *Doga, Türk Tarm ve Ormanlık Dergisi* 12(1): 48-74. [Cock (1993), Turkish, English summary]
- 3288.Ozgür, A.F., Sekeroglu, E., Ohnesorge, B. and Gocmen, H. 1990. Studies on host plant changes, migration and population dynamics of the cotton whitefly, *Bemisia tabaci*, in Cukurova (Turkey). *Mitteilungen der Deutschen Gesellschaft für Allgemeine und Angewandte Entomol.* 7(4-6): 653-656. [German, English summary]
- 3289.Ozgür, A.F., Sekeroglu, E., Ohnesorge, B. and Göçmen, H. 1989. Studies on the population dynamics of *Bemisia tabaci* Genn. (Homopt., Aleyrodidae) in Cukurova, Turkey. [German summary]. *J. Appl. Entomol.* 107(3): 217-227. [Cock (1993)]
- 3290.Pacheco-Covarrubias, J.J. 1998. Conceptualización y organigrama de la campaña contra la mosquita blanca de la hoja plateada (*Bemisia argentifolii* Bellows & Perring). [Conceptualization and chartflow of the campaign against the silverleaf whitefly (*Bemisia argentifolii* Bellows & Perring)]. pp. 149-155. *In* J.J. Pacheco-Covarrubias and F. Pacheco-Mendivil (Eds.), *Temas Selectos Para El Manejo Integrado de la Mosquita Blanca*, Memoria Científica No. 6. Instituto Nacional De Investigaciones Forestales, Agrícolas y Pecuarias, Centro de Investigación Regional del Noroeste, CIRNO, Obregon, Sonora, Mexico.
- 3291.Pacheco-Covarrubias, J.J. 1998. Crecimiento poblacional de la mosquita blanca de la hoja plateada (*Bemisia argentifolii* Bellows & Perring) como base para la implementación de medidas de combate. [Population growth of the silverleaf whitefly (*Bemisia argentifolii* Bellows & Perring) as a base for implementation of control measures]. pp. 39-45. *In* J.J. Pacheco-Covarrubias and F. Pacheco-Mendivil (Eds.), *Temas Selectos Para El Manejo Integrado de la Mosquita Blanca*, Memoria Científica No. 6. Instituto Nacional De Investigaciones Forestales, Agrícolas y Pecuarias, Centro de Investigación Regional del Noroeste, CIRNO, Obregon, Sonora, Mexico. [Spanish]
- 3292.Pacheco-Covarrubias, J.J. 1998. Estrategia para el manejo regional de insecticidas para la mosquita blanca de la hoja plateada (*Bemisia argentifolii* Bellows & Perring). [Strategy for regional insecticide management for the silverleaf whitefly (*Bemisia argentifolii* Bellows & Perring)]. pp. 127-147. *In* J.J. Pacheco-Covarrubias and F. Pacheco-Mendivil (Eds.), *Temas Selectos Para El Manejo Integrado de la Mosquita Blanca*, Memoria Científica No. 6. Instituto Nacional De Investigaciones Forestales, Agrícolas y Pecuarias, Centro de Investigación Regional del Noroeste, CIRNO, Obregon, Sonora, Mexico. [Spanish]
- 3293.Pacheco-Covarrubias, J.J. and Hernández, A. 1998. Differential susceptibility of cotton cultivars to silverleaf whitefly in the Yaqui Valley, Sonora, Mexico. pp. 1281-1282. *In* P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
- 3294.Pacheco-Covarrubias, J.J. and Hernandez-Jasso, A. 1999. Median life of silverleaf whitefly in cotton cultivars. pp. 491-492. *In* P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
- 3295.Pacheco, J.L. 1998. A five year review of *Lygus* efficacy and cotton yield studies in Central Arizona. pp. 984-992. *In* P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
- 3296.Pacheco-Mendivil, F. 1995. Identificación de la "Mosquita blanca" *B. tabaci* (Gennadius) y de la "Mosquita blanca de la hoja plateada" *B. argentifolii* (Bellows & Perring). [Identification of *Bemisia tabaci* (Genn.) and *B. argentifolii* (Bellows & Perrings)]. *Bemisia tabaci* in Northwest Mexico, Research Report 1993, Ciudad Obregon, Sonora, Mexico [Spanish]
- 3297.Pacheco-Mendivil, F. 1998. Generalidades del género *Bemisia* [Generalities of the genus *Bemisia*]. pp. 31-38. *In* J.J. Pacheco-Covarrubias and F. Pacheco-Mendivil (Eds.), *Temas Selectos Para El Manejo Integrado de la Mosquita Blanca*, Memoria Científica No. 6. Instituto Nacional De Investigaciones Forestales, Agrícolas y Pecuarias, Centro de Investigación Regional del Noroeste, CIRNO, Obregon, Sonora, Mexico.
- 3298.Padidam, M., Beachy, R.N. and Fauquet, C.M. 1995. Classification and identification of geminiviruses using sequence comparisons. *J. Gen. Virol.* 76(2): 249-263.
- 3299.Padidam, M., Beachy, R.N. and Fauquet, C.M. 1995. Tomato leaf curl geminivirus from India has a bipartite genome and coat protein is not essential for infectivity. *J. Gen. Virol.* 76(1): 25-35.
- 3300.Padidam, M., Sawyer, S. and Fauquet, C.M. 1999. Possible emergence of new geminiviruses by frequent recombination. *Virology* 265: 218-225.
- 3301.Padilla, M.R. 1995. Reducing tomato yellow mosaic severity through soil fertilization. M. Sc. Thesis. CATIE. Turrialba, Costa Rica, 90 pp.
- 3302.Pai, K.F. and Chen, C.C. 1998. Biology of silverleaf whitefly, *Bemisia argentifolii* (Homoptera: Aleyrodidae) on three host plants. [Chinese, English summary]. *Bull. Tichung District Agric. Improvement Stn.* 58: 33-41.
- 3303.Paiva, F.D.A. and Goulart, A.C.P. 1995. Variation of the whitefly population and incidence of bean golden mosaic at Dourados, MS. *Fitopatol. Brasileira* 20(2): 199-202. [Portuguese]
- 3304.Pakistan Central Cotton Committee. 1986. Summary Annual Progress Report 1985-1986. pp. 45-92. *In* Central Cotton Research Institute, Sakrand, Pakistan.

- 3305.Pal, B.P. and Tandon, R.K. 1937. Types of tobacco leaf-curl in northern India. *Indian J. Agric. Sci.* 7: 363-393. [Cock (1986)]
- 3306.Palaniswami, M.S. 1999. Whitefly *Bemisia tabaci* Genn. and its natural enemies. CTCRI (Central Tuber Crops Research Institute (Kerala, India) 15(1): 4-6.
- 3307.Palaniswami, M.S., Antony, B. and Lisha Vijayan, S., (Eds.). 2001. Indian bibliography of sweetpotato whitefly, *Bemisia tabaci* (Genn.) 1979-1998. Bibliography Series 5. Central Tuber Research Institute, Thiruvananthapuram, India.
- 3308.Palaniswami, M.S., Nair, R.R., Pillai, K.S. and Thankappan, M. 1996. Whiteflies on cassava and its role as vector of cassava mosaic disease in India. *J. Root Crops* 22(1): 1-8.
- 3309.Palaniswami, M.S. and Pillai, K.S. 1990. Bioecology of *Bemisia tabaci* G. on cassava. ISRC National Symposium. *J. Root Crops* (Special): 169-173.
- 3310.Palaniswami, M.S., Rajamma, P. and Pillai, K.S. 1988. Biological agents of the pests of tropical tuber crops in India - A review. *J. Root Crops* 14(2): 41-46.
- 3311.Palevsky, E., Soroker, V., Weintraub, P., Mansour, F. and Abo-Moch, F. 2001. How species-specific is the phoretic relationship between the broad mite, *Polypagotarsonemus latus* (Acari: Tarsonemidae), and its insect hosts? *Exp. Appl. Acarol.* 25(3): 217-224.
- 3312.Palumbo, J.C. 1998. Evaluation of insect growth regulators for management of whiteflies in melons. pp. 35-40. *In* N. Oebker (Ed.), *Vegetable Report, Series P-115, AZ 1101*. University of Arizona, College of Agriculture, Tucson.
- 3313.Palumbo, J.C. 1998. Management of whiteflies and leafminers in melons in the southwestern United States. pp. 97-106. *In* J.H. Nunez-Moreno, G. Martinez-Diaz, E. Valenzuela-Cornejo and L.A. Maldonado-Navarro (Eds.), *Memories International Symposium on Phytosanitary Protection, Hermosillo, Sonora, Mexico, 27-30 Oct 1998*. INIFAP. [English and Spanish]
- 3314.Palumbo, J.C. 1999. Field performance of admire against silverleaf whitefly on commercial iceberg lettuce, 1993-1998. pp. 121-129. *In* D.N. Byrne (Ed.), *Vegetable Report Series P-117, AZ 1143*. University of Arizona, College of Agriculture, Tucson.
- 3315.Palumbo, J.C. 2001. Residual efficacy of foliar chloronicotinyls against silverleaf whiteflies on melons. *Arthropod Management Tests* 26: E21.
- 3316.Palumbo, J.C. and Coates, W.E. 1996. Air-assisted electrostatic application of pyrethroid and endosulfan mixtures for sweetpotato whitefly (Homoptera: Aleyrodidae) control and spray deposition in cauliflower. *J. Econ. Entomol.* 89(4): 970-980.
- 3317.Palumbo, J.C., Ellsworth, P.C., Dennehy, T.J. and Umeda, K. 1999. Cross commodity management of whiteflies and chemical efficacy in Arizona. pp. 108-120. *In* D.N. Byrne (Ed.), *Vegetable Report Series P-117, AZ 1143*. University of Arizona, College of Agriculture, Tucson.
- 3318.Palumbo, J.C. and Hannan, T.A. 1997. Commercial field performance of KNACK on cotton in the Yuma Valley. pp. 294-305. *In* Cotton, A College of Agriculture Report, Series P-108. University of Arizona, Tucson.
- 3319.Palumbo, J.C., Horowitz, A.R. and Prabhaker, N. 2001. Insecticidal control and resistance management for *Bemisia tabaci*. *In* S. E. Naranjo and P. C. Ellsworth (Eds.), *Special Issue: Challenges and Opportunities for Pest Management of Bemisia tabaci in the New Century*. *Crop Prot.* 20(9): 739-765.
- 3320.Palumbo, J.C., Kerns, D.L., Engle, C.E., Sanchez, C.A. and Wilcox, M. 1996. Imidacloprid formation and soil placement effects on colonization by sweetpotato whitefly (Homoptera: Aleyrodidae): Head size and incidence of chlorosis in lettuce. *J. Econ. Entomol.* 89(3): 735-742.
- 3321.Palumbo, J.C., Kerns, D.L. and Umeda, K. 2000. Whitefly management on desert melons. *IPM Series No. 14, Univ. Arizona Coop. Ext. Publ.* AZ1190, 7 pp.
- 3322.Palumbo, J.C., Mullins, C. and Reyes, F.J. 1995. Control of sweetpotato whitefly on fall broccoli, 1993. *Arthropod Management Tests* 20: 68.
- 3323.Palumbo, J.C., Mullins, C.H. and Reyes, F.J. 1993. Control of sweetpotato whitefly in cantaloupe with various pesticides 1992. *Insecticide Acaricide Tests* 18: 117.
- 3324.Palumbo, J.C., Mullins, C.H. and Reyes, F.J. 1994. Control of sweetpotato whitefly in cantaloupe with various pesticides, 1992. *Arthropod Management Tests* 19: 80-81.
- 3325.Palumbo, J.C., Mullins, C.H. and Reyes, F.J. 1995. Evaluation of imidacloprid for control of sweetpotato whitefly in lettuce, 1994. *Arthropod Management Tests* 20: 93.
- 3326.Palumbo, J.C., Mullins, C.H., Jr. and Reyes, F.J. 1995. Efficacy of imidacloprid on whitefly in fall cantaloupes, 1994. *Arthropod Management Tests* 20: 82.
- 3327.Palumbo, J.C. and Mullins, C.H., Jr. 1996. Efficacy of pyrethroid & endosulfan combinations against whitefly adults on spring cantaloupes, 1995. *Arthropod Management Tests* 21: 103-104.
- 3328.Palumbo, J.C., Mullins, C.H., Jr. and Reyes, F.J. 1996. Control of whiteflies with imidacloprid on fall cantaloupes, 1995. *Arthropod Management Tests* 21: 104.
- 3329.Palumbo, J.C., Mullins, C.H., Jr. and Reyes, F.J. 1997. Evaluation of imidacloprid seed treatments for control of whiteflies on fall cantaloupes, 1996. *Arthropod Management Tests* 22: 116.
- 3330.Palumbo, J.C., Mullins, C.H., Jr. and Reyes, F.J. 2001. Residual efficacy and field performance of thiacloprid (Calypto) against silverleaf whitefly in melons. pp. 43-52. *In* D.N. Byrne and P. Baciewicz (Eds.), *Vegetable Report*. University of Arizona, College of Agriculture and Life Sciences, Tucson.
- 3331.Palumbo, J.C., Mullins, C.H., Jr., Reyes, F.J., Amaya, A., Ledesma, L. and Carey, L. 2000. Comparison of neonicotinoid use patterns for silverleaf whitefly management in melons and broccoli. pp. 22-34. *In* D.N. Byrne and P. Baciewicz (Eds.), *Vegetable Report, AZ1143*. University of Arizona, College of Agriculture, Tucson.
- 3332.Palumbo, J.C. and Sanchez, C.A. 1995. Imidacloprid does not enhance growth and yield of muskmelon in the absence of whitefly. *Hortscience* 30(5): 997-999.
- 3333.Palumbo, J.C., Tonhasca, A. and Byrne, D.N. 1995. Evaluation of three sampling methods for estimating adult sweetpotato whitefly (Homoptera: Aleyrodidae) abundance on cantaloupes. *J. Econ. Entomol.* 88(5): 1393-1400.
- 3334.Palumbo, J.C., Tonhasca, A., Jr. and Byrne, D.N. 1994. Sampling plans and action thresholds for whiteflies on spring melons. *IPM Series No. 1, Univ. Arizona, Coop. Ext. Publ.*, 2 pp.
- 3335.Palumbo, J.C., Toscano, N.C., Blua, M.J. and Yoshida, H.A. 2000. Impact of *Bemisia* whiteflies (Homoptera: Aleyrodidae) on alfalfa growth, forage yield, and quality. *J. Econ. Entomol.* 93(6): 1688-1694.
- 3336.Pane, A., Garzia, G.T., Cacciola, S.O., Lio, G.M., Grasso, S. and Perrotta, G. 1999. Biological control of insect pests of agricultural crops with entomopathogenic fungi. *Phytophaga Palermo* 9(Suppl.): 105-115. [Italian, English summary]
- 3337.Paoli, G. 1930. Notizie sull' arriccamento del cotone nella somalia italiana. *Atti primo Congresso Studi coloniale, Firenze, 8-12 April 1931*, 324-336. [Cock (1986)]
- 3338.Paplomatas, E.J., Patel, V.P., Hou, Y.M., Noueir, A.O. and Gilbertson, R.L. 1994. Molecular characterization of a new sap-transmissible bipartite genome geminivirus infecting tomatoes in Mexico. *Phytopathology* 84(10): 1215-1224.
- 3339.Pappu, S.S., Pappu, H.R., Langston, D.B., Jr., Flanders, J.T., Riley, D.G. and Diaz-Perez, J.C. 2000. Outbreak of tomato yellow leaf curl virus (family Geminiviridae) in Georgia. *Plant Dis.* 84(3): 370.
- 3340.Pareek, B.L., Sharma, R.C. and Yadav, C.P.S. 1983. Record of insect faunal complex on mothbean, *Vigna aconitifolia* (Jacq.) Marechal in semi arid zone of Rajasthan. *Bull. Entomol. (New Delhi)* 24: 44-45. [Cock (1986)]

- 3341.Parth, I.A., Mgbemena, M.O., Anozie, O.C. and Tanyimboh, E.N. 1997-1998. Incidence and control of insect pests of vegetable crops on Jos Plateau, Northern Nigeria. *Nigerian J. Entomol.* 14/15: 7-22.
- 3342.Paris, H.S. 1993. Leaf silvering of squash: a brief review. [Haifa, Israel]. *Cucurbit Genet. Coop.* 16: 75-76.
- 3343.Paris, H.S., Stoffella, P.J. and Powell, C.A. 1993. Differential susceptibility to leaf silvering in *Cucurbita pepo*. *Hortscience* 28(6): 657-658.
- 3344.Paris, H.S., Stoffella, P.J. and Powell, C.A. 1993. Susceptibility to leaf silvering in the cultivar groups of summer squash. *Euphytica* 69(1/2): 69-72.
- 3345.Paris, H.S., Stoffella, P.J. and Powell, C.A. 1993. Sweetpotato whitefly, drought stress, and leaf silvering of squash. *HortScience* 28(2): 157-158.
- 3346.Park, M. and Fernando, M. 1938. The nature of chilli leaf curl. *Trop. Agriculturist* 91: 263-265. [Cock (1986)]
- 3347.Parkash, O., Pandita, M.L. and Malik, Y.S. 1979. Effect of fertilizer and irrigation applications on the efficacy of granular insecticides against insect pests of brinjal (*Solanum melongena* L.). *Haryana J. Hortic. Sci.* 8(3-4): 143-146. [Cock (1986)]
- 3348.Parkash, O. and Verma, A.N. 1985. Effect of different granular insecticides applied by different methods, against jassid, *Amrasca devastans* (Dist.) and white fly, *Bemisia tabaci* (Genn.) on brinjal during pre-fruited crop stage. *Indian J. Entomol.* 47(1): 66-70. [Cock (1993)]
- 3349.Parker, B.L., Booth, R.H. and Bellotti, A.C. 1978. *Bemisia tabaci* identified on *Manihot esculenta* in Malaysia. *Malaysian Appl. Biol.* 7(1): 85-86. [Cock (1986)]
- 3350.Parker, B.L. and Brownbridge, M. 1992. Fungal pathogens for biological control of greenhouse insect pests. pp. 1-14. *In* Sustain. Agric. Res. and Education (SARE) or Agric. in Concert with the Environment (ACE) Research Projects 1992.
- 3351.Parker, R. 1995. Identification of poinsettia whitefly (*Bemisia tabaci* Type B). *Queensland Fruit and Vegetable News* (Australia) 66(2): 12.
- 3352.Parrella, M.P., Bellows, T.S., Gill, R.J., Brown, J.K. and Heinz, K.M. 1992. Sweetpotato whitefly: prospects for biological control. *California Agric.* 46(1): 25-26.
- 3353.Parrella, M.P., Jones, V.P., Malais, M.S. and Heinz, K.M. 1989. Advances in sampling in ornamentals. [Spanish summary]. *Florida Entomol.* 72(3): 394-403. [Cock (1993)]
- 3354.Parrella, M.P. and Murphy, B.C. 1998. Insect growth regulators. *Grower Talks* 62(2): 86-89.
- 3355.Parrella, M.P., Paine, T.D., Bethke, J.A., Robb, K.L. and Hall, J. 1991. Evaluation of *Encarsia formosa* (Hymenoptera: Aphelinidae) for biological control of sweetpotato whitefly (Homoptera: Aleyrodidae) on poinsettia. *Environ. Entomol.* 20(2): 713-719. [Cock (1993)]
- 3356.Parrish, M.D. 2001. Assail TM: a new tool for insect pest management in cotton. pp. 46-47. *In* P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
- 3357.Pascal, E., Goodlove, P.E., Wu, L.C. and Lazarowitz, S.G. 1993. Transgenic tobacco plants expressing the geminivirus BL1 protein exhibit symptoms of viral disease. *Plant Cell* 5: 795-807.
- 3358.Pascal, E., Sanderfoot, A.A., Ward, B.M., Medville, R., Turgeon, R. and Lazarowitz, S.G. 1994. The geminivirus BR1 movement protein binds single-stranded dna and localizes to the cell nucleus. *Plant Cell* 6: 995-1006.
- 3359.Pascoe, D.R. 1997. ICAC/CFC whitefly project activities in Zimbabwe. p. 17. *In* International Cotton Advisory Committee, 56th Plenary Meeting, Paraguay.
- 3360.Pascual, S., Aviles, M., Nombela, G., Muniz, M. and Beitia, F. 2000. Development of *Bemisia tabaci* (biotype Q) on tomato cultivars with/without the Mi gene. *Med. Fac. Landbouww. Univ. Gent* 65(2a): 291-292.
- 3361.Pasian, C., Taylor, R.A.J., McMahon, R.W. and Lindquist, R.K. 2000. New method of acephate application to potted plants for control of *Aphis gossypii*, *Frankliniella occidentalis* and *Bemisia tabaci*. *Crop Prot.* 19(4): 263-271.
- 3362.Pasian, C.C., Lindquist, R.K. and Struve, D.K. 1997. A new method of applying imidacloprid to potted plants for controlling aphids and whiteflies. *HortTechnology* 7(3): 265-269.
- 3363.Pasini, C., D'Aquila, F. and Gandolfo, M. 1997. *Gerbera*: sali di acidi grassi nella lotta agli aleirodidi. [ *Gerbera*: salts of fatty acids in controlling aleyrodids]. *Colture Protette* (Italy) 26(4): 73-76. [Italian, English summary]
- 3364.Pasini, C., D'Aquila, F., Gandolfo, M., Costanzi, M. and Mirto, L. 1998. *Macrolophus caliginosus* nella lotta biologica. [Experience in biological control of whiteflies on Gerbera in greenhouse by the mirid bug *Macrolophus caliginosus*]. *Colture Protette* (Italy) 27(Suppl. 12): 43-46. [Italian, English summary]
- 3365.Paszkowski, U., Zhang, S.B., Potrykus, I. and Paszkowski, J. 1993. Replication of the DNA A component of African cassava mosaic virus in a heterologous system. *J. Gen. Virol.* 74: 2725-2729.
- 3366.Patel, B.H., Patel, D.M., Patel, M.J. and Patel, J.R. 1998. Bio-efficacy of various ready mix synthetic insecticides against pest complex of 'H 6' cotton (*Gossypium hirsutum*). *Indian J. Agric. Sci.* 68(12): 780-781.
- 3367.Patel, B. N. and Patel, J.B. 1997. Failure of insecticides to control whitefly vector and thereby leaf curl on bidi tobacco. *Indian J. Virol.* 13(1): 29-31.
- 3368.Patel, G.J., Chari, M.S. and Jaisani, B.G. 1976. Differential response of whiteflies (*Bemisia tabaci* Gen.) to *Nicotiana* species. *Gujarat Agric. Univ. Res. J.* 1(2): 89-92. [Cock (1986)]
- 3369.Patel, H.M. and Jhala, R.C. 1992. Studies on host range, host preference and population dynamics of whitefly, *Bemisia tabaci* (Gennadius) in south Gujarat, India. *Gujarat Agric. Univ. Res. J.* 17: 76-81.
- 3370.Patel, H.M., Jhala, R.C., Pandya, A.V. and Patel, C.B. 1992. Biology of whitefly (*Bemisia tabaci*) on okra (*Hibiscus esculentus*). *Indian J. Agric. Sci.* 62(7): 497-499.
- 3371.Patel, J.R. and Thanki, K.V. 1988. Effect of planting time on the incidence of the pests of brinjal in Gujarat. pp. 512-515. *In* N. Mohandas and G. Koshy (Eds.), *Proceedings National Symposium on Integrated Pest Control: Progress and Perspectives*, 15-17 October. Association for Advancement of Entomology, Trivandrum, Kerala, India.
- 3372.Patel, M.B. and Srivastava, K.P. 1989. Biology of insect vector *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae) on cowpea, *Vigna unguiculata* (L.). *Bull. Entomol. (New Delhi)* 30(1): 99-109.
- 3373.Patel, M.B. and Srivastava, K.P. 1989. Host preference of whitefly, *Bemisia tabaci* Gennadius for oviposition and development on different grain legumes. *Bull. Entomol. (New Delhi)* 30(1): 118-120.
- 3374.Patel, M.B. and Srivastava, K.P. 1990. Field screening of some high yielding genotypes of mungbean, *Vigna radiata* (Linnaeus) Wilczrk to whitefly *Bemisia tabaci* (Gennadius) and yellow mosaic virus (YMV). *Indian J. Entomol.* 52(4): 547-551. [Cock (1993)]
- 3375.Patel, M.B. and Srivastava, K.P. 1990. Phytotoxic effect of insecticides on cowpea, *Vigna unguiculata* (Linn.) and green gram, *Vigna radiata* (Linn.). *Indian J. Entomol.* 52(4): 583-588. [Cock (1993)]
- 3376.Patel, M.B. and Srivastava, K.P. 1996. Efficacy of various insecticides against *Bemisia tabaci* (Genn.) and CGMV disease transmission in cowpea. *Gujarat Agric. Univ. Res. J.* 22(1): 43-50.
- 3377.Patel, M.B., Srivastava, K.P. and Patel, G.M. 1992. Incidence of whitefly and virus diseases in cowpeas and green gram intercropped with cereals and oilseeds. *Gujarat Agric. Univ. Res. J.* 18(1): 56-62.
- 3378.Patel, M.M., Patel, C.B. and Patel, M.B. 1995. Screening of brinjal varieties against insect pests. *Gujarat Agric. Univ. Res. J.* 20(2): 98-102.

3379. Patel, V.C. and Patel, H.K. 1966. Inter-relationship between whitefly (*Bemisia tabaci* Genn.) population and the incidence of leaf curl in bidi tobacco (*Nicotiana tabacum* L.) in relation to different planting dates. *Indian J. Entomol.* 28: 339-344. [Cock (1986)]
3380. Patil, B.V. 1996. Competitive displacement of *Bemisia* with leafhoppers and aphids in a cotton ecosystem. pp. 243-245. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia* : 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
3381. Patil, B.V., Nandihalli, B.S. and Hugar, P. 1990. Effect of synthetic pyrethroids on population buildup of cotton whitefly, *Bemisia tabaci* (G.). *Cotton Development* 18: 25-27.
3382. Patil, B.V., Nandihalli, B.S. and Hugar, P. 1991. Management of whitefly, *Bemisia tabaci* (G.) on cotton through certain plant and animal products. *J. Indian Soc. Cotton Improvement* 16(1): 21-26.
3383. Patil, B.V., Nandihalli, B.S., Hugar, P., Thimmanagoud, B.R. and Lingappa, S. 1993. Bioefficacy of newer insecticides against cotton whitefly *Bemisia tabaci* (Gennadius). *Karnataka J. Agric. Sci. [India]* 6(1): 20-24.
3384. Patil, B.V., Nandihalli, B.S., Hugar, P., Thimmanagouda, B.R. and Lingappa, S. 1993. Efficacy of some plant and animal products against cotton pest complex. pp. 183-189. *In* Botanical Pesticides and Integrated Pest Management. Indian Society of Tobacco Science, Rajahmundry, India.
3385. Patil, S.P. and Pokharkar, R.N. 1979. Some new records of insect pests infesting cruciferous vegetable crops in Maharashtra State. *J. Maharashtra Agric. Univ. (India)* 4(2): 222-223. [Cock (1986)]
3386. Patti, I. and Rapisarda, C. 1981. Reperti morfo-biologici sugli aleirodidi nocivi alle piante coltivate in Italia. *Bull. Zool. Agraria Bachicoltura* 16: 135-190. [Cock (1986)]
3387. Paulson, G.S. and Beardsley, J.W. 1985. Whitefly (Hemiptera: Aleyrodidae) egg pedicel insertion into host plant stomata. *Ann. Entomol. Soc. Am.* 78(4): 506-508. [Cock (1986, 1993)]
3388. Paulson, G.S. and Kumashiro, B.R. 1985. Hawaiian Aleyrodidae. *Proc. Hawaiian Entomol. Soc.* 25: 103-124. [Cock (1986)]
3389. Paximadis, M., Idris, A.M., Torres-Jerez, I., Villarreal, A., Rey, M.E.C. and Brown, J.K. 1999. Characterization of tobacco geminiviruses in the old and new world. *Arch. Virol.* 144: 703-717.
3390. Paximadis, M., Muniyappa, V. and Rey, M.E.C. 2001. A mixture of begomoviruses in leaf curl-affected tobacco in Karnataka, South India. *Ann. Appl. Biol.* 139(1): 101-109.
3391. Paximadis, M. and Rey, M.E.C. 1997. Aetiology of tobacco leaf curl in southern Africa. *Ann. Appl. Biol.* 131(3): 449-457.
3392. Peacock, B. and Summers, C. 1997. New threat to valley grapes. *California Grower* 21(10): 20-21.
3393. Pearson, E.O. and Maxwell-Darling, R.C. 1958. *Bemisia tabaci* (Gennadius) (Aleyrodidae). pp. 232-236. *In* E.O. Pearson and R.C. Maxwell Darling (Eds.), *The Insect Pests of Cotton in Tropical Africa*. Empire Cotton Growing Corp. and Commonwealth Inst. Entomol., London, UK. [Cock (1986)]
3394. Pedata, P.A. and Viggiani, G. 1991. Notes on *Encarsia transvena* (Timberlake) (Hymenoptera: Aphelinidae), a new parasitoid of Aleyrodidae in Italy. *Bull. Lab. Entomol. Agraria 'Filippo Silvestri' Portici* 48: 241-244. [Italian, English summary]
3395. Pedersen, T.J. and Hanley-Bowdoin, L. 1994. Molecular characterization of the a3 protein encoded by a bipartite geminivirus. *Virology* 202: 1070-1075.
3396. Pedgley, D.E. 1984. Weather and insect plagues in Africa. pp. 312-314. *In* D.L. Hawksworth (Ed.), *Advancing Agricultural Production in Africa*. Proceedings CAB's First Scientific Conference, Arusha, Tanzania 12-18 February 1984. Commonwealth Agricultural Bureaux, Carnham Royal, Slough, UK. [Cock (1986)]
3397. Peña, E.A., Pantoja, A. and Beaver, J. 1993. Development of *Bemisia tabaci* Gennadius in 4 Genotypes of *Phaseolus vulgaris* with different pubescence levels. *J. Agric. (Univ. Puerto Rico)* 77(1-2): 61-67.
3398. Peña, E.A., Pantoja, A., Beaver, J. and Armstrong, A. 1993. Oviposition of *Bemisia tabaci* Genn. (Homoptera: Aleyrodidae) in four genotypes of *Phaseolus vulgaris* L. (Leguminosae) with different degrees of pubescence. *Folia Entomol. Mexicana* 0(87): 1-12. [Spanish]
3399. Pena-Rojas, E.A., Pantoja, A. and Beaver, J.S. 1992. Determination of the pubescence of four genotypes of beans (*Phaseolus vulgaris* L.) and its effect on the insect *Bemisia tabaci* (Gennadius). *Rev. Colombiana Entomol.* 18(2): 41-48. [Spanish, English summary]
3400. Penny, D.D. 1922. A catalog of the California Aleyrodidae and the descriptions of four new species. *J. Entomol. Zool.* 14: 21-35. [Cock (1986)]
3401. Peralta, L. and Hilje, L. 1993. Un intento de control de *Bemisia tabaci* con insecticidas sistemicos incorporados a la vainica como cultivo trampa, mas aplicaciones de aceite en el tomate. [AN attempt to control *Bemisia tabaci* in tomatoes with systemic insecticides incorporated in beans as a trap crop, plus oil applications]. *Manejo Integrado de Plagas (Costa Rica)* 30: 21-23. [Spanish, English summary]
3402. Percy, R.G. 2001. Registration of PS-6ne, PS-6L, PS-6neL, P62ne, P62L, and P62neL extra-long staple cotton germplasm. *Crop Sci.* 41(2): 602-603.
3403. Peregrine, D.J. and Lemon, R.W. 1986. The value of amitraz for control of *Bemisia tabaci* on cotton. *Agric. Ecosystems Environ.* 17(1-2): 129-135. [Cock (1993)]
3404. Pereira, R.A.S., De Menezes, A. Jr. and Sugawara, L.M. 1996. Rearing whitefly *Bemisia tabaci* (Gennadius) (Hemiptera: Aleyrodidae) on rooted leaves of soybean (*Glycine max* (L.) Merr.). [Portuguese, English summary]. *Semina Londrina* 17(1): 45-48.
3405. Perez-Alvarez-Castellanos, M.P. 1999. Actividad plaguicida de aceites esenciales, extractos y material vegetal de *Chrysanthemum coronarium* L. Ph.D. Dissertation, Universidad Politecnica de Madrid, 254 pp.
3406. Perez, C.J., Alvarado, P., Narvaez, C., Miranda, F., Hernandez, L., Vanegas, H., Hruska, A. and Shelton, A.M. 2000. Assessment of insecticide resistance in five insect pests attacking field and vegetable crops in Nicaragua. *J. Econ. Entomol.* 93(6): 1779-1787.
3407. Perez-Cabrera, C.A. and Mendoza-Puquire, E. 2000. Historia y situacion de la produccion de frijol en los paises latinoamericanos afectados por geminivirus transmitidos por mosca blanca: America Central: El Salvador. pp. 29-31. *In* Bean Golden Mosaic and Other Diseases of Common Bean Caused by Whitefly-Transmitted Geminiviruses in Latin America. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. [Spanish]
3408. Perez, J.J., Obando, A. and Darby, N. 1993. Efficacy of buprofezin and conventional insecticides under different levels of SPWF populations. p. 721. *In* D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
3409. Perez, J.J., Obando, A. and Darby, N. 1994. Evaluation of the growth regulator buprofezin mixed with conventional insecticides, for whitefly control. pp. 904-905. *In* D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
3410. Perez, M.P. and Pascual-Villalobos, M.J. 1999. Effects of the essential oil of flower heads of *Chrysanthemum coronarium* L. on whitefly and stored product pests. *Investigacion Agraria, Produccion Proteccion Vegetales* 14(1-2): 249-258. [Spanish, English summary]
3411. Perez, O., Ramirez, O., Hilje, L. and Karremans, J. 1997. Potencial de adopcion de dos opciones tecnologicas de manejo integrado de plagas (MIP), aplicando tres tecnicas de extension con productores de tomate en el valle Central Occidental, Costa Rica. [Adoption potential of two integrated pest management technological options by tomato producers of Western Central Valley of Costa Rica, applying three extension techniques]. *Manejo Integrado de Plagas (Costa Rica)* 43: 19-30. [Spanish, English summary]

- 3412.Perez, O.E. 1996. Evaluating the adoption potential of two technologies of integrated management of pests (IPM) using three extension techniques with tomato producers in Grecia and Valverde Vega, Alajuela, Costa Rica. M. Sc. Thesis. CATIE. Turrialba, Costa Rica, 144 pp.
- 3413.Perez, P., Avila, E. and Martinez, M. 1998. La mosca blanca (*Bemisia* sp) como vector del plateado de la hoja de la calabaza. [White fly (*Bemisia* sp.) as vector of squash silvered leaf]. pp. 95-97. In Produccion De Cultivos En Condiciones Tropicales. Taller Internacional "Geminivirus en el Caribe", Habana, Cuba, Nov 1998. Instituto De Investigaciones Horticolas, La Habana, Cuba. [Spanish, English summary]
- 3414.Perkins, H.H., Jr. 1971. Rapid screening test for sugar content of cotton. Textile Bull. 97: 21, 25, 34.
- 3415.Perkins, H.H., Jr. 1971. Some observations on sticky cottons. Textile Manag. Eng. J. 135(3): 49-64.
- 3416.Perkins, H.H., Jr. 1977. Effects of whitefly contamination on cotton lint quality. pp. 93-94. In J.M. Brown (Ed.), Proceedings Beltwide Cotton Production Research Conferences. National Cotton Council, Memphis, TN.
- 3417.Perkins, H.H., Jr. 1983. Effects of whitefly contamination on lint quality of U. S. cottons. pp. 102-103. In J.M. Brown (Ed.), Proceedings Beltwide Cotton Production Research Conferences. National Cotton Council, Memphis, TN.
- 3418.Perkins, H.H., Jr. 1983. Identification and processing of honeydew-contaminated cottons. Textile Res. J. 53(8): 508-512.
- 3419.Perkins, H.H., Jr. 1984. Causes of a sticky problem for the cotton spinner. Textile Month : 42-43.
- 3420.Perkins, H.H., Jr. 1984. Use of additives to improve the processing quality of whitefly contaminated cotton. pp. 379-381. In J.M. Brown (Ed.), Proceedings Beltwide Cotton Production Research Conferences. National Cotton Council, Memphis, TN.
- 3421.Perkins, H.H., Jr. 1986. Whitefly honeydew in U. S. cottons: update on methods for detecting and processing contaminated cotton. pp. 106-107. In J.M. Brown and T.C. Nelson (Eds.), Proceedings Beltwide Cotton Production Research Conferences. National Cotton Council, Memphis, TN.
- 3422.Perkins, H.H., Jr. 1987. Sticky cotton. pp. 53-55. In Proc. 1987 Western Cotton Production Conf.Proc. National Cotton Council, Memphis, TN.
- 3423.Perkins, H.H., Jr. and Bassett, D.M. 1988. Variations in stickiness of variety test cottons - San Joaquin Valley, California, 1986. pp. 135-136. In J.M. Brown and D.A. Richter (Eds.), Proceedings Beltwide Cotton Production Research Conferences. National Cotton Council, Memphis, TN.
- 3424.Perrin, R.M. 1999. The role of industry in the management of insecticide resistance. pp. 197-205. In Proceedings ICAC-CCRI Regional Consultation, Insecticide Resistance Management in Cotton. Central Cotton Research Institute, Multan, Pakistan.
- 3425.Perring, T.M. 1996. Biological differences of two species of *Bemisia* that contribute to adaptive advantage. pp. 3-16. In D. Gerling and R.T. Mayer (Eds.), *Bemisia* : 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
- 3426.Perring, T.M. 2001. The *Bemisia tabaci* species complex. In S. E. Naranjo and P. C. Ellsworth (Eds.), Special Issue: Challenges and Opportunities for Pest Management of *Bemisia tabaci* in the New Century. Crop Prot. 20(9): 725-737.
- 3427.Perring, T.M., Cooper, A. and Kazmer, D.J. 1992. Identification of the poinsettia strain of *Bemisia tabaci* (Homoptera, Aleyrodidae) on broccoli by electrophoresis. J. Econ. Entomol. 85: 1278-1284.
- 3428.Perring, T.M., Cooper, A., Kazmer, D.J., Shields, C. and Shields, J. 1991. New strain of sweetpotato whitefly invades California vegetables. California Agric. 45(6): 10-12.
- 3429.Perring, T.M., Cooper, A.D., Rodriguez, R.J., Farrar, C.A. and Bellows, T.S. 1993. Identification of a whitefly species by genomic and behavioral studies. Science 259: 74-77.
- 3430.Perring, T.M., Farrar, C.A., Bellows, T.S., Cooper, A.D. and Rodriguez, R.J. 1993. Evidence for a new species of whitefly: UCR findings and implications. California Agric. 47(1): 7-8.
- 3431.Perring, T.M., Farrar, C.A., Cooper, A.D., Bellows, T.S. and Rodriguez, R.J. 1993. Determining whitefly species - response. Science 261: 1334-1335.
- 3432.Perring, T.M., Royalty, R.N. and Farrar, C.A. 1989. Floating row covers for the exclusion of virus vectors and the effect on disease incidence and yield of cantaloupe. J. Econ. Entomol. 82(6): 1709-1715. [ Cock (1993)]
- 3433.Peshney, N.L. and Moghe, P.G. 1984. Role of mites, thrips and whiteflies in transmission of viruses and development of churda murda of chillies in Vidharba. Punjabrao Krishi Vidyapeeth Res. J. (India) 8(1): 33-35.
- 3434.Pessoni, L.A., Zimmermann, M.J.D.O. and Faria, J.C.D. 1997. Genetic control of characters associated with bean golden mosaic geminivirus resistance in *Phaseolus vulgaris* L. Brazilian J. Genetics 20(1): 51-58.
- 3435.Peterlin, O. and Helman, S. 1996. Some aspects of the population dynamics of *Bemisia tabaci* as a cotton plague in Santiago del Estero, northwestern Argentina. pp. 133-141. In D. Gerling and R.T. Mayer (Eds.), *Bemisia* : 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
- 3436.Peterschmitt, M., Granier, M. and Aboulama, S. 1999. First report of tomato yellow leaf curl geminivirus in Morocco. Plant Dis. 83(11): 1074.
- 3437.Pettit, F.L. 1993. Biological control in the integrated pest management program at the Land, Epcot Center. Bull. OILB/SROP (IOBC/WPRS) 16: 129-132.
- 3438.Pettersson, M.L. and Rämert, B. 1988. [Plant protection year 1987 - horticulture]. Växtskyddsnotiser 52(3): 66-70. [ Cock (1993), Swedish, English summary]
- 3439.Petty, I.T.D., Coutts, R.H.A. and Buck, K.W. 1988. Transcriptional mapping of the coat protein gene of tomato golden mosaic virus. J. Gen. Virol. 69: 1359-1365.
- 3440.Phadke, A.D., Khandal, V.S. and Rahalkar, S.R. 1988. Use of neem product in insecticide resistance management IPM in cotton. Pesticides 22(4): 36-37. [ Cock (1993)]
- 3441.Philemon, E.C. 1987. Plant pathology note no 32. A virus disease of peanut.(publ. 1990). Harvest 12(4): 15-16. [ Cock (1993)]
- 3442.Pickett, C.H., Casanave, K.A., Schoenig, S.E. and Heinz, K.M. 1999. Rearing *Delphastus catalinae* (Coleoptera: Coccinellidae): practical experience and a modeling analysis. Canadian Entomol. 131(1): 115-129.
- 3443.Pico, B., Diez, M. and Nuez, F. 1998. Evaluation of whitefly-mediated inoculation techniques to screen *Lycopersicon esculentum* and wild relatives for resistance to Tomato yellow leaf curl virus. Euphytica 101(3): 259-271.
- 3444.Pico, B., Diez, M.J. and Nuez, F. 1996. Viral diseases causing the greatest economic losses to the tomato crop. II: The tomato yellow leaf curl virus - a review. Sci. Hortic. 67(3-4): 151-196.
- 3445.Pico, B., Diez, M.J. and Nuez, F. 1999. Improved diagnostic techniques for tomato yellow leaf curl virus in tomato breeding programs. Plant Dis. 83(11): 1006-1012.
- 3446.Pico, B., Ferriol, M., Diez, M. and Nuez, F. 1999. Developing tomato breeding lines resistant to tomato yellow leaf curl virus. Plant Breeding 118(6): 537-542.
- 3447.Pierre, R.E. 1979. Studies on golden mosaic of bean (*Phaseolus vulgaris*) in Jamaica. pp. 113-121. In Hillside Farming in Jamaica Training Seminar, Kingston, Jamaica, 11-13 Dec 1978. Ministry of Agriculture, Kingston, Jamaica.
- 3448.Pietersen, G., Idris, A.M., Kruger, K. and Brown, J.K. 2000. Tomato curly stunt virus, a new begomovirus of tomato within the tomato yellow leaf curl virus-IS cluster in South Africa. Plant Dis. 84(7): 810.

- 3449.Pillai, K.S. and Daniel, R.S. 1979. Monthly variation on population of whitefly (*Bemisia tabaci*) the vector of cassava mosaic disease. *J. Root Crops* 5(1/2): 8-10.
- 3450.Pilowsky, M. and Cohen, S. 1974. Inheritance of resistance of tomato yellow leaf curl virus in tomatoes. *Phytopathology* 64: 632-635.
- 3451.Pilowsky, M. and Cohen, S. 1990. Tolerance to tomato yellow leaf curl virus derived from *Lycopersicon peruvianum*. *Plant Dis.* 74: 248-250.
- 3452.Pilowsky, M. and Cohen, S. 2000. Screening additional wild tomatoes for resistance to the whitefly -borne tomato yellow leaf curl virus. *Acta Physiol. Plantarum* 22(3): 351-353.
- 3453.Pimpale, T.D. and Summanwar, A.S. 1984. Studies on different stages in the life cycle and influence of season on the duration of different generations of the whitefly *Bemisia tabaci* Genn. *Pestology* 8: 15-19.
- 3454.Pinto, R.L., Hoefert, L.L. and Fail, G.L. 1988. Plasmalemma deposits in tissues infected with lettuce infectious yellows virus. *J. Ultrastructure Mol. Structure Res.* 100(3): 245-254.
- 3455.Piven, N.M., De Uzcatagui, R.C. and Infante, H.D. 1995. Resistance to tomato yellow mosaic virus in species of *Lycopersicon*. *Plant Dis.* 79(6): 590-594.
- 3456.Plana, L., Suris, M., Pino, M.A., Quintana, E., Hernandez, Y. and Martinez, M.A. 1995. Incidencia de *Bemisia tabaci* Genn. en tomate (*Lycopersicon esculentum* Mill), asociado a maiz como cultivo protector, en epoca no optima. [Incidence of *Bemisia tabaci* Genn. on tomato (*Lycopersicon esculentum* Mill), associated with maize as a protective crop, out of the optimal growing season]. *Rev. Proteccion Vegetal* 10(2): 129-132. [ Spanish, English summary]
- 3457.Ploper, L.D., Rodriguez-Pardina, P.E., Laguna, I.G., Truol, G.A., Hanada, K., Rivas-Platero, G.G., Ramirez, P. and Herrera, P.S. 1998. Presence of a geminivirus in soyabeans in Northwest Argentina. [Spanish, English summary]. *Avance Agroindustrial* 19(74): 38-41.
- 3458.Pluschke, U., Horowitz, A.R. and Ishaaya, I. 1999. Effect of milbemectin on the sweetpotato whitefly, *Bemisia tabaci*. *Phytoparasitica* 27(3): 183-191.
- 3459.Polaszek, A. 1991. Egg parasitism in Aphelinidae (Hymenoptera: Chalcidoidea) with special reference to *Centrodora* and *Encarsia* species. *Bull. Entomol. Res.* 81: 97-106. [ Cock (1993)]
- 3460.Polaszek, A., Abd-Rabou, S. and Huang, J. 1999. The Egyptian species of *Encarsia* (Hymenoptera: Aphelinidae): a preliminary review. *Zool. Mediterr. Leiden* 73: 131-163.
- 3461.Polaszek, A., Evans, G. and Bennett, F.D. 1992. *Encarsia* parasitoids of *Bemisia tabaci* (Hymenoptera: Aphelinidae, Homoptera: Aleyrodidae) - a preliminary guide to identification. *Bull. Entomol. Res.* 82: 375-391. [ Cock (1993)]
- 3462.Polizzi, G. and Asero, C. 1994. Epidemiology and incidence of tomato yellow leaf curl virus (TYLCV) in greenhouse protected by screens in Italy. *Acta Hort.* 366: 345-352.
- 3463.Pollard, D.G. 1955. Feeding habits of the cotton whitefly, *Bemisia tabaci* Genn. (Homoptera, Aleyrodidae). *Ann. Appl. Biol.* 43: 664-671. [ Cock (1986)]
- 3464.Pollard, D.G. and Saunders, J.H. 1956. Relations of some cotton pests to jassid resistant sakel. *Empire Cotton Growing Rev.* 33: 197-202. [ Cock (1986)]
- 3465.Polston, J.E. 1988. Interactions between divided genome viruses under field conditions: A study of cucurbit geminiviruses. [University of California, Riverside]. *Dissertation Abst. Int.* 50(03B): 814.
- 3466.Polston, J.E., Al-Musa, A., Perring, T.M. and Dodds, J.A. 1990. Association of the nucleic acid of squash leaf curl geminivirus with the whitefly *Bemisia tabaci*. *Phytopathology* 80(9): 850-856. [ Cock (1993)]
- 3467.Polston, J.E. and Anderson, P.K. 1997. The emergence of whitefly-transmitted geminiviruses in tomato in the western hemisphere. *Plant Dis.* 81(12): 1358-1369.
- 3468.Polston, J.E. and Anderson P. K. 1999. Surgimiento y distribución de geminivirus transmitidos por mosca blanca en tomate en el Hemisfero Occidental. [The emergence of whitefly -transmitted geminiviruses in tomato in the Western Hemisphere]. *Manejo Integrado de Plagas (Costa Rica)* 53: 24-42. [ Spanish, English summary]
- 3469.Polston, J.E., Chellemi, D.O., Schuster, D.J., McGovern, R.J. and Stansly, P.A. 1996. Spatial and temporal dynamics of tomato mottle geminivirus and *Bemisia tabaci* (Genn.) in Florida tomato fields. *Plant. Dis.* 80(9): 1022-1028.
- 3470.Polston, J.E., Dodds, J.A. and Perring, T.M. 1989. Nucleic acid probes for detection and strain discrimination of cucurbit geminiviruses. *Phytopathology* 79: 1123-1127.
- 3471.Polston, J.E., Hiebert, E., McGovern, R.J., Stansly, P.A. and Schuster, D.J. 1993. Host range of tomato mottle virus, a new geminivirus infecting tomato in Florida. *Plant Dis.* 77(12): 1181-1184.
- 3472.Polston, J.E., McGovern, R.J. and Brown, L.G. 1999. Introduction of tomato yellow leaf curl virus in Florida and implications for the spread of this and other geminiviruses of tomato. *Plant Dis.* 83(11): 984-988.
- 3473.Pompeu, A.S. and Krantz, W.M. 1977. Linhagens de feijoeiro (*Phaseolus vulgaris* L.) resistance ao virus do mosaico dourado. *Summa Phytopathol.* 3: 162-163. [ Portuguese]
- 3474.Poprawski, T.J. 1999. Control of *Bemisia argentifolii* on collards using different formulations and rates of *Beauveria bassiana*, 1997. *Arthropod Management Tests* 24: 122-123.
- 3475.Poprawski, T.J. and Jackson, M.A. 1999. Laboratory activity of blastospores of *Paecilomyces fumosoroseus* on *Bemisia argentifolii* nymphs, 1997. *Arthropod Management Tests* 24: 399-400.
- 3476.Poprawski, T.J. and Jones, W.J. 2001. Host plant effects on activity of the mitosporic fungi *Beauveria bassiana* and *Paecilomyces fumosoroseus* against two populations of *Bemisia* whiteflies (Homoptera: Aleyrodidae). *Mycopathologia* 151(1): 11-20.
- 3477.Poprawski, T.J., Legaspi, J.C. and Parker, P.E. 1998. Influence of entomopathogenic fungi on *Serangium parcesetosum* (Coleoptera: Coccinellidae), an important predator of whiteflies (Homoptera: Aleyrodidae). *Environ. Entomol.* 27(3): 785-795.
- 3478.Poswal, A. and Williamson, S. 1998. Off the 'treadmill': Cotton IPM in Pakistan. *Pestic. News* 40: 12-13.
- 3479.Powell, C.A., De Paulo, J.J. and Borovsky, D. 1993. Induction of trypsin-like enzymes in the sweetpotato whitefly, pp. 77-83. *In* D. Borovsky and A. Spielman (Eds.), *Proceedings Third Symposium*, Vero Beach, Florida, February 8-11, 1993. Florida Medical Entomology Laboratory, Vero Beach, Florida, USA.
- 3480.Powell, C.A. and Stoffella, P.J. 1991. Endosulfan and silver reflective mulch effects on sweet potato whitefly populations and yields of zucchini squash and tomatoes. *Proc. Florida State Hort. Soc.* 103: 117-119.
- 3481.Powell, C.A. and Stoffella, P.J. 1993. Influence of endosulfan sprays and aluminum mulch on sweetpotato whitefly disorders of zucchini squash and tomatoes. *J. Prod. Agric.* 6: 118-121.
- 3482.Powell, C.A. and Stoffella, P.J. 1995. Culling tomatoes with external symptoms of irregular ripening is of limited benefit. *Hortscience* 30(2): 316-317.
- 3483.Powell, C.A. and Stoffella, P.J. 1995. Susceptibility of tomato cultivars to internal and external tomato irregular ripening. *Hortscience* 30(6): 1307.
- 3484.Powell, C.A. and Stoffella, P.J. 1998. Control of tomato irregular ripening with imidacloprid. *Hortscience* 33(2): 283-284.
- 3485.Powell, C.A., Stoffella, P.J. and Paris, H.S. 1993. Plant population influence on squash yield, sweetpotato whitefly, squash silverleaf and zucchini yellow mosaic. *HortScience* 28(8): 796-798.

- 3486.Powell, D.A. 1990. Life tables and demography of sweetpotato whitefly, *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae), and two *Eretmocerus* sp. parasitoids (Hymenoptera: Aphelinidae). [University of California, Riverside]. Dissertation Abst. Int. 52(01B): 42.
- 3487.Powell, D.A. and Bellows, T.S. 1992. Adult longevity, fertility and population growth rates for *Bemisia tabaci* (Genn) (Homoptera: Aleyrodidae) on two host plant species. J. Appl. Entomol. 113: 68-78.
- 3488.Powell, D.A. and Bellows, T.S. 1992. Development and reproduction of two populations of *Eretmocerus* species (Hymenoptera, Aphelinidae) on *Bemisia tabaci* (Homoptera, Aleyrodidae). Environ. Entomol. 21: 651-658.
- 3489.Powell, D.A. and Bellows, T.S., Jr. 1992. Preimaginal development and survival of *Bemisia tabaci* on cotton and cucumber. Environ. Entomol. 21: 359-363.
- 3490.Powell, R.K., Stoffella, P.J. and Powell, C.A. 1998. Internal tomato irregular ripening symptoms do not diminish upon storage. Hortscience 33(1): 157.
- 3491.Power, A.G. 2000. Insect transmission of plant viruses: A constraint on virus variability. Curr. Opin. Plant Biol. 3(4): 336-340.
- 3492.Prabhaker, N., Coudriet, D.L. and Meyerdirk, D.E. 1985. Insecticide resistance in the sweetpotato whitefly, *Bemisia tabaci* (Homoptera: Aleyrodidae). J. Econ. Entomol. 78(4): 748-752. [Cock (1993)]
- 3493.Prabhaker, N., Coudriet, D.L. and Meyerdirk, D.E. 1987. Discrimination of three whitefly species (Homoptera: Aleyrodidae) by electrophoresis of non-specific esterases. [German summary]. J. Appl. Entomol. 103(5): 447-451. [Cock (1993)]
- 3494.Prabhaker, N., Coudriet, D.L. and Toscano, N.C. 1988. Effect of synergists on organophosphate and permethrin resistance in sweetpotato whitefly (Homoptera: Aleyrodidae). J. Econ. Entomol. 81(1): 34-39. [Cock (1993)]
- 3495.Prabhaker, N., Toscano, N.C., Castle, S.J. and Henneberry, T.J. 1996. Evaluation of insecticide rotation and mixtures for delaying insecticide resistance in whiteflies in Imperial Valley, CA. pp. 1031-1032. In P. Dugger and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3496.Prabhaker, N., Toscano, N.C., Castle, S.J. and Henneberry, T.J. 1997. Selection for imidacloprid resistance in silverleaf whiteflies from the Imperial Valley and development of a hydroponic bioassay for resistance monitoring. Pestic. Sci. 51: 419-428.
- 3497.Prabhaker, N., Toscano, N.C. and Coudriet, D.L. 1989. Susceptibility of the immature and adult stages of the sweetpotato whitefly (Homoptera: Aleyrodidae) to selected insecticides. J. Econ. Entomol. 82(4): 983-988. [Cock (1993)]
- 3498.Prabhaker, N., Toscano, N.C. and Henneberry, T.J. 1994. Management strategies to extend the effectiveness of chemicals for whitefly control. pp. 894-895. In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3499.Prabhaker, N., Toscano, N.C. and Henneberry, T.J. 1998. Evaluation of insecticide rotations and mixtures as resistance management strategies for *Bemisia argentifolii* (Homoptera: Aleyrodidae). J. Econ. Entomol. 91(4): 820-826.
- 3500.Prabhaker, N., Toscano, N.C. and Henneberry, T.J. 1999. Comparison of neem, urea, and amitraz as oviposition suppressants and larvicides against *Bemisia argentifolii* (Homoptera: Aleyrodidae). J. Econ. Entomol. 92(1): 40-46.
- 3501.Prabhaker, N., Toscano, N.C., Henneberry, T.J., Castle, S.J. and Weddle, D. 1996. Assessment of two bioassay techniques for resistance monitoring of silverleaf whitefly (Homoptera: Aleyrodidae) in California. J. Econ. Entomol. 89(4): 805-815.
- 3502.Prabhaker, N., Toscano, N.C., Perring, T.M., Nuessly, G., Kido, K. and Youngman, R.R. 1992. Resistance monitoring of the sweetpotato whitefly (Homoptera: Aleyrodidae) in the Imperial Valley of California. J. Econ. Entomol. 85(4): 1063-1068.
- 3503.Prabhu, S.R., Chari, M.S., Ramakrishnayya, B.V., Rao, R.S.N., Kumar, D.G., Rao, B.V.K. and Murthy, P.S.N. 1993. Production and prospects of nicotine sulphate in pest management. pp. 297-305. In Botanical Pesticides and Integrated Pest Management. Indian Society of Tobacco Science, Rajahmundry, India.
- 3504.Prasad, G.S. and Logiswaran, G. 1997. Influence of weather factors on population fluctuation of insect pests on brinjal at Madurai, Tamil Nadu [India]. Indian J. Entomol. 59(4): 385-388.
- 3505.Prasad, M.S., Sarma, B.K. and Kumar, S. 1998. Transmission tests and varietal screening for urd leaf crinkle virus in black gram (*Vigna mungo* L. Hepper). Ann. Plant Prot. Sci. 6(2): 205-207.
- 3506.Prasad, S.M., Kudada, N., Mishra, B. and Dhar, V. 1992. Date of transplanting and incidence of leaf curl in chillies. J. Res., Birsra Agric Univ. [India] 4(1): 81-82.
- 3507.Prasad, V.D., Bharati, M. and Reddy, G.P.V. 1993. Relative resistance to conventional insecticides in three populations of cotton whitefly, *Bemisia tabaci* (Gennadius) in Andhra Pradesh. Indian J. Plant Prot. 21(1): 102-103.
- 3508.Price, J.F., Schuster, D.F. and Short, D.T. 1986. Managing sweetpotato whitefly. Greenhouse Grower December: 55-57.
- 3509.Price, J.F. and Schuster, D.J. 1990. Responses of sweetpotato whitefly to azadirachtin extracted from neem tree seeds (*Azadirachta indica*). pp. 85-90. In Neem's potential in pest management programs, Proceedings of the USDA neem workshop. USDA-ARS, ARS-86.
- 3510.Price, J.F. and Schuster, D.J. 1991. Effects of natural and synthetic insecticides on sweetpotato whitefly *Bemisia tabaci* (Homoptera: Aleyrodidae) and its hymenopterous parasitoids. Florida Entomol. 74(1): 60-68.
- 3511.Price, J.F., Schuster, D.J. and McClain, P.M. 1991. Azadirachtin from neem tree (*Azadirachta indica* A. Juss.) seeds for management of sweetpotato whitefly (*Bemisia tabaci* (Gennadius)) on ornamentals. Proc. Florida State Hort. Soc. 103: 186-188.
- 3512.Price, J.F. and Taborsky, D. 1992. Movement of immature *Bemisia tabaci* (Homoptera, Aleyrodidae) on poinsettia leaves. Florida Entomol. 75: 151-153.
- 3513.Priesner, H. and Hosny, M. 1934. Contributions to a knowledge of white flies (Aleyrodidae) of Egypt (II). Bull. Ministry Agric. Egypt Tech. Sci. Serv. 139: 1-21. [Cock (1986)]
- 3514.Priesner, H. and Hosny, M. 1940. Notes on parasites and predators of Coccidae and Aleyrodidae in Egypt. Bull. Soc. Fouad Entomol. 24: 58-70. [Cock (1986)]
- 3515.Proctor, J.H. 1974. A review of cotton entomology. Outlook Agric. 8(1): 15-22. [Cock (1986)]
- 3516.Prophete, E. and Donis, J. 2000. Historia y situacion de la produccion de frijol en los paises latinoamericanos afectados por geminivirus transmitidos por mosca blanca: Region caribe: Haiti. pp. 59-64. In Bean Golden Mosaic and Other Diseases of Common Bean Caused by Whitefly-Transmitted Geminiviruses in Latin America. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. [Spanish]
- 3517.Pruthi, H.S. 1937. Report of the Imperial Entomologist. Sci. Rep., Agric. Res. Inst. (New Delhi) 1935-36: 123-137. [Cock (1986)]
- 3518.Pruthi, H.S. 1937. Report of the Imperial Entomologist. Sci. Rep., Agric. Res. Inst. (New Delhi) 1936-37: 159-174. [Cock (1986)]
- 3519.Pruthi, H.S. 1939. Report of the Imperial Entomologist. Sci. Rep., Agric. Res. Inst. (New Delhi) 1937-38: 113-128. [Cock (1986)]
- 3520.Pruthi, H.S. 1940. Report of the Imperial Entomologist. Sci. Rep., Agric. Res. Inst. (New Delhi) 1938-39: 116-133. [Cock (1986)]
- 3521.Pruthi, H.S. 1941. Report of the Imperial Entomologist. Sci. Rep., Agric. Res. Inst. (New Delhi) 1939-40: 102-114. [Cock (1986)]
- 3522.Pruthi, H.S. 1942. Report of the Imperial Entomologist. Sci. Rep., Agric. Res. Inst. (New Delhi) 1940-41: 57-63. [Cock (1986)]
- 3523.Pruthi, H.S. 1946. Report of the Imperial Entomologist. Abridged Sci. Rep. Agric. Res. Inst. New Delhi. 1941-44: 64-71. [Cock (1986)]

3524. Pruthi, H.S. and Samuel, C.K. 1937. Entomological investigations of the leaf-curl disease of tobacco in North Bihar. I. Transmission experiments with some suspected insect vectors. II. An alternative host of the virus and the insect transmitter. *Indian J. Agric. Sci.* 7: 659-670. [Cock (1986)]
3525. Pruthi, H.S. and Samuel, C.K. 1939. Entomological investigations of the leafcurl disease of tobacco in northern India. III. The transmission of leaf-curl by white-fly, *Bemisia gossypiperda*, to tobacco, sann-hemp and a new alternate host of the leaf-curl virus. *Indian J. Agric. Sci.* 9: 223-275. [Cock (1986)]
3526. Pruthi, H.S. and Samuel, C.K. 1941. Entomological investigations of the leaf-curl disease of tobacco in northern India. IV. Transmission of the disease by white-fly (*Bemisia gossypiperda*) from some new alternate hosts. *Indian J. Agric. Sci.* 11: 387-409. [Cock (1986)]
3527. Pruthi, H.S. and Samuel, C.K. 1942. Entomological investigations on the leaf-curl disease of tobacco in northern India. V. Biology and population of the white-fly vector [*Bemisia tabaci* (Gen.)] in relation to the incidence of the diseases. *Indian J. Agric. Sci.* 12: 35-57. [Cock (1986)]
3528. Pun, K.B. and Doraiswamy, S. 2000. Host range and host preference of *Bemisia tabaci* Gen. for oviposition and development. *JASS* 13(1): 88-90.
3529. Pun, K.B., Doraiswamy, S. and Jeyarajan, R. 1999. Immunological detection of okra yellow vein mosaic virus. *Indian J. Virol.* 15(2): 93-96.
3530. Pun, K.B., Doraiswamy, S. and Jeyarajan, R. 2000. Screening of virus inhibitory chemicals and neem products against okra yellow vein mosaic virus. *Indian Phytopathol.* 53(1): 95-96.
3531. Pun, K.B. and Saud, B.K. 1998. Occurrence of Dolichos yellow mosaic virus disease in Assam. *Indian J. Virol.* 14(2): 131-132.
3532. Pundt, L.S. 1993. Tips for managing whiteflies on poinsettias. *Connecticut Greenhouse Newsl.* 174(June/July): 7-10.
3533. Punjab. 1935. Entomology. Report. Department of Agriculture, (Punjab) 1933-34: 48-56. [Cock (1986)]
3534. Punjab. 1937. Entomology. Rep. Dept. Agric. (Punjab) 1935-36: 51-55. [Cock (1986)]
3535. Purcell, M.F., Johnson, M.W. and Tabashnik, B.E. 1995. Effects of insecticide use on abundance and diversity of tomato pests and associated natural enemies in Hawaii. *Proc. Hawaiian Entomol. Soc.* 32: 45-59.
3536. Puri, S.N., Ansingkar, A.S., Ajankar, V.N., Lavekar, R.C., Butler, G.D., Jr. and Henneberry, T.J. 1993. Effect of cotton leaf morphology on incidence of *Bemisia tabaci* Genn. on cotton. *J. Appl. Zool. Res.* 4(1): 41-44.
3537. Puri, S.N., Bhosle, B.B., Borikar, P.S., Fartade, M.K., Kolhal, R.N., Ilyas, M., Kawthekar, B.R., Butler, G.D., Jr. and Henneberry, T.J. 1996. Wild brinjal *Solanum khasianum* Clarke as a potential trap crop management tool for *Bemisia* in cotton. pp. 237-240. *In D. Gerling and R.T. Mayer (Eds.), Bemisia : 1995 Taxonomy, Biology, Damage, Control and Management.* Intercept Ltd., Andover, Hants, UK.
3538. Puri, S.N., Bhosle, B.B., Ilyas, M., Butler, G.D., Jr. and Henneberry, T.J. 1994. Detergents and plant-derived oils for control of the sweetpotato whitefly on cotton. *Crop Prot.* 13(1): 45-48.
3539. Puri, S.N., Butler, G.D., Jr. and Henneberry, T.J. 1991. Plant derived oils and soap solutions as control agents for the whitefly on cotton. *J. Appl. Zool. Res.* 1(2): 1-5.
3540. Puri, S.N., Murthy, K.S. and Sharma, O.P. 1998. Integrated management of cotton whitefly, *Bemisia tabaci* (Gennadius). pp. 286-296. *In G.S. Dhaliwal, N.S. Randhawa, R. Arora and A.K. Dhawan (Eds.), Ecological agriculture and sustainable development, Vols 1 and 2.* Punjab Agric. Univ., Ludhiana, India.
3541. Purohit, M.S. and Deshpande, A.D. 1991. Effect of inorganic fertilizers and insecticides on population density of cotton whitefly *Bemisia tabaci*. *Indian J. Agric. Sci.* 61(9): 696-698. [Cock (1993)]
3542. Qasem, A., Sodah, M., Trahm, M. and Assaf, A. 1986. [Insecticides evaluation of the control of whitefly, *Bemisia tabaci* on cucumber: II.- glass house, Hussein Agricultural Station 1986]. Dept. Plant Prot., Jordan Univ., Amman, 3 pp. [Arabic]
3543. Qiu, Y.T., Drost, Y.C., daGraca, O.S. and van Lenteren, J.C. 1999. Preliminary study on interplant movement and host location rate of five parasitoids of *Bemisia argentifolii* in small greenhouse. *Bull. OILB/SROP (IOBC/WPRS)* 22(1): 197-200.
3544. Quaintance, A.L. 1900. Contribution towards a monograph of the American Aleurodidae. USDA, Tech. Ser. Bur. Entomol. 8: 9-64. [Cock (1986)]
3545. Quaintance, A.L. and Baker, A.C. 1914. Classification of the Aleurodidae Part II. U.S. Dept. Agric., Tech. Ser. Bur. Entomol. 27: 95-109.
3546. Quezada, J.R. and Saunders, J.L. 1989. Whiteflies: proposals for a programme of integrated pest management. *Boletin Informativo, Manejo Integrado De Plagas* 12: 6-7.
3547. Quintela, E.D., Sanchez, S.E.M. and Yokoyama, M. 1992. Parasitism of *Encarsia* spp. on *Bemisia tabaci*. *Anais Soc. Entomol. Brasil* 21(3): 471-475.
3548. Quintero, C., Cardona, C., Ramirez, D. and Jimenez, N. 1998. First report of biotype B of *Bemisia tabaci* (Homoptera: Aleyrodidae) in Colombia. [Spanish, English summary]. *Rev. Colombiana Entomol.* 24(1-2): 23-28.
3549. Quiot, J.B., Labonne, G. and Marrou, J. 1982. Controlling seed and insect-borne viruses. pp. 96-116. *In K.F. Harris and K. Maramorosch (Eds.), Pathogens, Vectors, and Plant Diseases: Approaches to Control.* Academic Press, New York, NY.
3550. Quiros, C.A., Calvo, G. and Ramirez, O. 1995. Diagnostico de la problematica fitosanitaria del cultivo de tomate, con enfasis en mosca blanca, *Bemisia tabaci* (Gennadius). [Diagnosis of phytosanitary problems in tomatoes, with an emphasis on whiteflies *Bemisia tabaci* (Gennadius)]. *Manejo Integrado de Plagas (Costa Rica)* 38: 8-15. [Spanish, English summary]
3551. Quirós, C.A., Ramirez, O. and Hilje, L. 1994. Participacion de los agricultores en adaptar y evaluar tecnologias de semilleros contra la mosca blanca (*Bemisia tabaci*), en tomate. [Participation by the tomato growers in adapting and evaluating technologies against the whitefly]. *Manejo Integrado de Plagas (Costa Rica)* 34: 1-7. [Spanish, English summary]
3552. Quisumbing, D.A., Wolfenbarger, D.A., Hilje, L. and Guharay, F. 1995. Bioassays of a long-lasting, yellow "Lure N Kill" insecticidal device against the silverleaf whitefly. pp. 866-868. *In D.A. Richter and J. Armour (Eds.), Proceedings Beltwide Cotton Conferences.* National Cotton Council, Memphis, TN.
3553. Radhakrishnan-Nair, R. and Wilson, K.I. 1970. Leaf curl of *Jatropha curcas* L. in Kerala. *Sci. Culture* 36: 569. [Cock (1986)]
3554. Radhakrishnan-Nair, R. and Wilson, K.I. 1970. Studies on some whitefly transmitted plant virus diseases from Kerala. *Agric. Res. J. (Kerala)* 7(2): 123-126. [Cock (1986)]
3555. Radwan, H.S.A., Ammar, I.M.A., Eisa, A.A., Assal, O.M. and Omar, H.I.H. 1983. Development retardation and inhibition of adult emergence in cotton whitefly, *Bemisia tabaci* Genn. following immature stages treated with two molt inhibitors. *Bull. Entomol. Soc. Egypt Econ. Ser.* 13: 175-181.
3556. Radwan, H.S.A., Ammar, I.M.A., Eisa, A.A., Omar, H.I.H. and Moftah, E.A.M. 1984. Latent effects of certain bacillus preparations on the biology of the cotton whitefly, *Bemisia tabaci*. [Arabic summary]. *Minufiya J. Agric. Res. (Egypt)* 8: 417-429. [Cock (1993)]
3557. Radwan, H.S.A., El-Ghar, G.E.S.A., Rashwan, M.H. and El-Bermawy, Z.A. 1990. Impact of several insecticides and insect growth regulators against the whitefly, *Bemisia tabaci* (Gennadius), in [Egyptian] cotton fields. *Bull. Entomol. Soc. Egypt Econ. Ser.* 18: 81-92.



3558. Radwan, S.M.E., Abdel-Hamid, Z.H., El-Sadaany, G.B. and Romeilah, M.A. 2000. The triangular relationship between cotton transplanting, planting dates and the population density of sap sucking pests. *Egyptian J. Agric. Res.* 78(4): 1449-1476.
3559. Raffie, A., Diaz, J. and McLeod, P. 1999. Effects of forage groundnut in reducing the sweetpotato whitefly and associated gemini virus disease in bell pepper in Honduras. *Trop. Agric.* 76(3): 208-211.
3560. Ragupathi, A. and Narayanasamy, P. 1998. Transmission studies on leafcurl virus disease of tomato (*Lycopersicon esculentum* Mill). *South Indian Hortic.* 46(3-6): 216-219.
3561. Rahman, A.A.A. 1988. Selective insecticides for cotton pest management (integrated pest control). pp. 109-113. *In Annu. Rep. Gezira Res. Stn. Substns.* 1980/1981. Agric. Res. Corp., Sudan [Cock (1993)]
3562. Raj, S.K., Aslam, M., Srivastava, K.M. and Singh, B.P. 1989. Association of geminivirus-like particles with yellow mosaic disease of *Dolichos lablab* L. *Curr. Sci. (Bangalore)* 58(14): 813-815. [Cock (1993)]
3563. Raj, S.K., Chandra, G. and Singh, B.P. 2000. Detection of a whitefly-transmitted geminivirus in *Launaea* spp. in India. *Plant Dis.* 84(5): 593.
3564. Raj, S.K. and Singh, B.P. 1996. Association of geminivirus infestation with yellow green mosaic disease of *Cucumis sativus*. Diagnosis by nucleic acid probes. *Indian J. Exp. Biol.* 34(6): 603-605.
3565. Raj, S.K., Srivastava, K.M. and Singh, B.P. 1996. Evidence from nucleic acid hybridization tests for geminivirus infection of ornamental crotons in India. *Eur. J. Plant Pathol.* 102(2): 201-203.
3566. Rajak, R.L. and Diwakar, M.C. 1987. Resurgence of cotton whitefly in India and its integrated management. *Plant Prot. Bull. (Faridabad)* 39(3): 13-14.
3567. Rajam, B., David, B.V. and Peter, C. 1992. Influence of biochemical parameters and other physical features of some host plants on the biology of the whitefly *Bemisia tabaci* (Genn.). *J. Appl. Zool. Res.* 3(2): 170-173.
3568. Rajam, B., Peter, C. and David, B.V. 1988. Influence of host plants on the parasitism of *Bemisia tabaci* Gennadius (Homoptera: Aleyrodidae) by *Encarsia* sp. *Curr. Sci. (Bangalore)* 57(22): 1246-1247. [Cock (1993)]
3569. Rajapakse, R.H.S. and Jayasena, K.W. 1989. Field pest problems of mungbean *Vigna radiata* in southern regions of Sri Lanka. *Entomon* 14(12): 159-164. [Cock (1993)]
3570. Ramakrishnan, K., Kandaswamy, T.K., Subramanian, K.S., Janarthanan, R., Mariappan, V., Samuel, G.S. and Navaneethan, G. 1972. Investigations of virus disease of pulse crops in Tamil Nadu. Final Tech. Rep. Coimbatore, India; Tamil Nadu Agric. Univ. [Cock (1986)]
3571. Ramappa, H.K., Muniyappa, V. and Colvin, J. 1998. The contribution of tomato and alternative host plants to tomato leaf curl virus inoculum pressure in different areas of South India. *Ann. Appl. Biol.* 133(2): 187-198.
3572. Ramirez-Arredondo, J.A., Cardenas, I.A., Sanchez, F.D. and Garzon-Tiznado, J.A. 1998. Virus transmitted by whitefly (*Bemisia tabaci* Gennadius) on pepper and zucchini squash in the Mayo Valley Sonora, Mexico. *Agric. Tecnica Mexico* 24(1): 37-43. [Spanish, English summary]
3573. Ramirez-Choza, J.L., Diaz-Plaza, R. and Aviles-Baeza, W.I. 1993. Mosquita blanca *Bemisia tabaci* Gen. Avances de investigacion en Yucatan. [*Bemisia tabaci* Gen. Research advances in Yucatan State]. Campo Experimental Zona Henequenera Cir Sureste, Mococho, Yucatan, Mexico, 16 pp [Spanish]
3574. Ramirez, P. 1997. Los geminivirus [Whitefly geminiviruses]. Manejo Integrado de Plagas (Costa Rica) 43: 40-54. [Spanish, English summary]
3575. Ramirez, P. and Maxwell, D. 1995. Geminivirus transmitidos por moscas blancas. [Geminivirus transmission by whiteflies]. Manejo Integrado de Plagas (Costa Rica) 36: 22-27. [Spanish, English summary]
3576. Ramirez, P. and Rivera-Bustamante, R. 1996. Identificación de geminivirus. pp. 30-41. *In* L. Hilje (Ed.), Metodologias para el estudio y manejo de moscas blancas y geminivirus. Centro Agronomico Tropical de Investigacion y Ensenanza, Turrialba, Costa Rica.
3577. Ramos, E.Q., Alves, S.B., Tanzini, M.R. and Lopes, R.B. 2000. Susceptibilidad de *Bemisia tabaci* a *Beauveria bassiana* en condiciones de laboratorio. [Susceptibility of *Bemisia tabaci* to *Beauveria bassiana* under laboratory conditions]. Manejo Integrado de Plagas (Costa Rica) 56: 65-69. [Spanish, English summary]
3578. Ramos, L.J., Bharathan, N., McMillan, R.T. and Narayanan, K.R. 1995. Histopathological changes associated with silverleaf syndrome in squash. *Plant Pathol.* 44(2): 316-324.
3579. Rangarajan, A.V., Mahadevan, N.R. and Iyemperumal, S. 1977. Pest complex of sunflower (*Helianthus annuus* Linn.) in Tamil Nadu. *Indian J. Entomol.* 37: 188-191. [Cock (1986)]
3580. Rangaraju, R. and Chenulu, V.V. 1980. A new method for counting whitefly *Bemisia tabaci* (Genn.) population in mung bean (*Vigna radiata* (L.) Wilczek). *Curr. Sci. (Bangalore)* 49: 825-826. [Cock (1986)]
3581. Ranjith, A.M. and Mohanasundaram, M. 1992. A new method for rapid leaf screening of cotton germplasm for the whitefly, *Bemisia tabaci* Gennadius. *Madras Agric. J.* 79(4): 218-219.
3582. Ranjith, A.M., Pillay, V.S., Sasikumaran, S. and Mammooty, K.P. 1992. New record of whitefly *Bemisia tabaci* on black pepper *Piper nigrum*. *Indian J. Agric. Sci.* 62(2): 166-168.
3583. Rao, A.S. 1958. Notes on Indian Aleyrodidae (whiteflies), with special reference to Hyderabad. Proc. 10th Int. Congress Entomol., Montreal, 1956. 1: 331-336. [Cock (1986)]
3584. Rao, A.S., Prasada Rao, R.D.V.J. and Reddy, P.S. 1980. A whitefly transmitted yellow mosaic disease on groundnut (*Arachis hypogaea* L.). *Curr. Sci. (Bangalore)* 49: 160. [Cock (1986)]
3585. Rao, D.G. and Varma, P.M. 1961. Investigations on yellow vein mosaic of *Malvastrum coromandalianum*, a whitefly-transmitted virus in India. Proc. 48th Indian Sci. Congr. Roorkee : 499-500. [Cock (1986)]
3586. Rao, N.V. and Reddy, A.S. 1989. Natural enemies of cotton whitefly, *Bemisia tabaci* (Genn.), under Andhra Pradesh (India) situation. *Bull. Entomol. (New Delhi)* 34(1/2): 163-169.
3587. Rao, N.V. and Reddy, A.S. 1989. Seasonal influence on developmental duration of whitefly *Bemisia tabaci* in upland cotton *Gossypium hirsutum*. *Indian J. Agric. Sci.* 59(6): 383-385. [Cock (1993)]
3588. Rao, N.V. and Reddy, A.S. 1992. The natural enemies of cotton whitefly, *Bemisia tabaci* - A review. *Agric. Rev.* 13: 12-20.
3589. Rao, N.V. and Reddy, A.S. 1993. A note on the mechanism of resistance to whitefly, *Bemisia tabaci* Genn. in cotton. *Indian J. Entomol.* 55(4): 456-459.
3590. Rao, N.V. and Reddy, A.S. 1994. Incidence of whitefly, *Bemisia tabaci* Genn., in relation to other sucking pests of cotton. *Indian J. Entomol.* 56(1): 104-106.
3591. Rao, N.V., Reddy, A.S., Ankaiah, R. and Mukudan, S. 1989. Effects of whitefly *Bemisia tabaci* Genn. on cotton yield and associated components. *Insect Sci. Appl.* 10(5): 685-690.
3592. Rao, N.V., Reddy, A.S., Ankaiah, R., Rao, Y.N. and Khasim, S.M. 1990. Incidence of whitefly *Bemisia tabaci* in relation to leaf characters of upland cotton *Gossypium hirsutum* L. *Indian J. Agric. Sci.* 60(9): 619-624. [Cock (1993)]
3593. Rao, N.V., Reddy, A.S., Rao, B.R. and Satyanarayana, G. 1991. Intraplant distribution of whitefly *Bemisia tabaci* (Genn.) on cotton *Gossypium hirsutum* L. *J. Insect Sci.* 4(1): 32-36.
3594. Rao, N.V., Reddy, A.S. and Rao, K.T. 1989. A method to monitor whitefly *Bemisia tabaci* in cotton, *Gossypium hirsutum*. *Indian J. Agric. Sci.* 59(7): 459-461. [Cock (1993)]

- 3595.Rao, N.V., Reddy, A.S. and Rao, K.T. 1991. Monitoring of cotton whitefly, *Bemisia tabaci* with sticky traps. Madras Agric. J. 78(1-4): 1-7.
- 3596.Rao, N.V., Reddy, A.S. and Rao, K.T. 1991. Reaction of few cotton cultures to whitefly, *Bemisia tabaci* Genn. Madras Agric. J. 78(1-4): 72-73.
- 3597.Rao, N.V., Reddy, A.S. and Rao, T.K. 1989. Natural enemies of cotton whitefly *Bemisia tabaci* Gennadius in relation to host population and weather factors. J. Biol. Control 3(1): 10-12.
- 3598.Rao, N.V., Reddy, A.S. and Reddy, D.D.R. 1990. Effect of some insecticides on the parasitoids and predators of the cotton whitefly, *Bemisia tabaci* Genn. J. Biol. Control 4(1): 4-7.
- 3599.Rao, N.V., Reddy, A.S. and Reddy, D.D.R. 1990. Impact of some insecticides on *Bemisia tabaci* on cotton. J. Plant Prot. Trop. (Malaysia) 7(2): 77-86.
- 3600.Rao, N.V., Reddy, A.S. and Reddy, D.D.R. 1990. Relative toxicity of some insecticides to cotton whitefly, *Bemisia tabaci*. Indian J. Plant Prot. 18(1): 97-100. [ Cock (1993)]
- 3601.Rao, N.V., Reddy, A.S. and Reddy, P.S. 1990. Relative efficacy of some new insecticides on insect pests of cotton. Indian J. Plant Prot. 18(1): 53-58.
- 3602.Rao, R.D.V.J.P., Ragunathan, V. and Joshi, N.C. 1983. Occurrence of yellow mosaic disease on clusterbean, *Cyamopsis tetragonoloba*. Indian J. Plant Prot. 10(1-2): 100. [ Cock (1986)]
- 3603.Rao, R.S.N., Chari, M.S. and Rao, S.G. 1990. Further record of natural enemies on the insect pests of tobacco in Andhra Pradesh. J. Biol. Control 4(1): 65-66. [ Cock (1993)]
- 3604.Rapet, S. and Chauvel, G. 1994. Cultures ornamentales: la situation phytosanitaire en 1993. [Ornamental crops: the phytosanitary situation in 1993]. Phytoma (France) 459: 46-48. [ French]
- 3605.Rapisarda, C. 1990. [*Bemisia tabaci* vector of TYLCV in Sicily]. Informatore Fitopatol. 40(6): 27-31. [ Cock (1993), Italian]
- 3606.Rapisarda, C. and Patti, I. 1983. Stato attuale delle conoscenze sulla composizione dell'aleirofauna siciliana. pp. 327-332. In Atti 13 Congresso Nazionale Italiano di Entomologia. Istituto di Entomologia Agraria e Apicoltura, Univerisità Torino [Cock (1986)]
- 3607.Rasool, G., Hussain, A.M., Malik, N.A., Zia, M.A. and Ghaffar, A. 1988. Field performance of some formulations of dimethoate insecticide against cotton sucking pests. J. Agric. Res. (Pakistan) 26(3): 233-236.
- 3608.Rat-Morris, E. 1990. [Contribution to the control of the aleyrodid *Bemisia tabaci* on poinsettia. Phytoma (France) 421: 46-47. [ Cock (1993), French]
- 3609.Rat-Morris, E. and Travers, M. 1989. Effet de la Buprofezine sur l'aleurode *Bemisia tabaci*. [Effect of Buprofezin on the white fly *Bemisia tabaci*]. Ecole Nationale D'ingenieurs Des Travaux De L'horticulture Et Du Paysage D'angers Resultats D'experimentation Et D'essais (France) No. 13, 7 pp. [ French]
- 3610.Rataul, H.S. and Brar, J.S. 1989. Status of tomato leaf curl virus research in India. Trop. Sci. 29(2): 111-118. [ Cock (1993)]
- 3611.Rataul, H.S. and Butter, N.S. 1975. Effect of different systemic granular insecticides on the population of whitefly *Bemisia tabaci* Genn. (Aleyrodidae: Hemiptera), the vector of tomato leafcurl virus (TLCV). J. Res., (Punjab Agric. Univ.) 12: 382-386. [ Cock (1986)]
- 3612.Rataul, H.S. and Butter, N.S. 1977. Control of tomato leafcurl virus in tomatoes (*Lycopersicon esculentum* Miller) by suppressing the vector population of *Bemisia tabaci* Genn. with insecticidal sprays. J. Res., (Punjab Agric. Univ.) 13: 303-307. [ Cock (1986)]
- 3613.Rataul, H.S. and Singh, L. 1974. Control of soybean yellow mosaic virus in soybean *Glycine* Max L. by controlling the vector whitefly *Bemisia tabaci* Genn. in Punjab. J. Res. (Ludhiana) 11(1): 73-76. [ Cock (1986)]
- 3614.Rataul, H.S. and Singh, L. 1977. Field studies on the control of soybean yellow-mosaic virus, transmitted by whitefly *Bemisia tabaci* Genn. by using different granular systemic insecticides. J. Res., (Punjab Agric. Univ.) 13: 298-302. [ Cock (1986)]
- 3615.Rathi, Y.P.S. and Nene, Y.L. 1974. Sex of *Bemisia tabaci* (Genn.) in relation to the transmission of mung bean yellow mosaic virus. Acta Bot. Indica 2: 74-76.
- 3616.Rathi, Y.P.S. and Nene, Y.L. 1974. A technique for handling whitefly *Bemisia tabaci* adults in serial transmission of viruses. Indian Phytopathol. 27: 390-393.
- 3617.Rathi, Y.P.S. and Nene, Y.L. 1975. Some aspects of the relationship between mung bean yellow mosaic virus and its vector *Bemisia tabaci*. Indian Phytopathol. 27: 459-462. [ Cock (1986)]
- 3618.Rathore, G.S. and Agnthotri, J.P. 1985. Effect of insecticides on yellow mosaic of moth (*Vigna aconitifolia* (Jacq) Marechel). Indian J. Virol. 1(1): 92-94. [ Cock (1993)]
- 3619.Rathore, Y.S. and Lal, S.S. 1998. Rectangular cage a possible replacement of round split cage for trapping whiteflies and jassids. Indian J. Pulses Res. 11(1): 115-117.
- 3620.Rathore, Y.S., Lal, S.S. and Singhal, R.A. 1998. Population of whitefly and jassid as influenced by sampling time. Indian J. Pulses Res. 11(1): 120-122.
- 3621.Rathore, Y.S. and Tiwari, S.N. 1998. Influence of crops and cropping seasons on spatial distribution of *Bemisia tabaci* Genn. Indian J. Pulses Res. 11(2): 76-85.
- 3622.Raychaudhuri, S.P., Chatterjee, S.N. and Dhar, H.K. 1961. Preliminary note on the occurrence of the yellow net vein disease of mulberry. Indian Phytopathol. 14: 94-95. [ Cock (1986)]
- 3623.Razvi, S.A., Azam, K.M. and Al-Raeesi, A.A. 1999. Monitoring of sweet potato whitefly, *Bemisia tabaci* (Gennadius) with yellow sticky traps. Sultan-Qaboos Univ. J. Sci. Res., Agric. Sci. 4(1): 11-16.
- 3624.Razvi, S.A., Azam, K.M. and Zouba, A. 2000. Screening of tomato cultivars against whitefly, *Bemisia tabaci* (Gennadius) and tomato leaf curl virus. Shashpa 7(2): 143-149.
- 3625.Reckhaus, P. 1979. A virus disease of white yam (*Dioscorea rotundata*) in Togo. Zeitschrift für Pflanzenkrankheiten und Pflanzenschutz 86: 763-766. [ Cock (1986)]
- 3626.Redak, R.A. and Bethke, J.A. 1993. Control of the sweetpotato whitefly on poinsettia 1992. Insecticide Acaricide Tests 18: 337-338.
- 3627.Redak, R.A. and Bethke, J.A. 1993. Control of the sweetpotato whitefly on poinsettia using soil treatments and a parasitoid, summer 1992. Insecticide Acaricide Tests 18: 339.
- 3628.Redak, R.A. and Bethke, J.A. 1993. Control of the sweetpotato whitefly on poinsettia using soil treatments summer 1992. Insecticide Acaricide Tests 18: 338-339.
- 3629.Redak, R.A. and Bethke, J.A. 1994. Control of the silverleaf whitefly (SW) on poinsettia under greenhouse conditions, fall 1993. Arthropod Management Tests 19: 334.
- 3630.Redak, R.A. and Bethke, J.A. 1994. Control of the silverleaf whitefly (SW) on poinsettia using soil treatments, winter 1992. Arthropod Management Tests 19: 333.
- 3631.Redak, R.A. and Bethke, J.A. 1995. Control of adult whitefly on poinsettia under greenhouse conditions, winter 1992. Arthropod Management Tests 20: 300-301.
- 3632.Redak, R.A. and Bethke, J.A. 1995. Control of the silverleaf whitefly on lantana under greenhouse conditions, fall 1993. Arthropod Management Tests 20: 299-300.
- 3633.Redak, R.A. and Bethke, J.A. 1995. Control of the silverleaf whitefly on poinsettia under greenhouse conditions, winter 1993. Arthropod Management Tests 20: 300.
- 3634.Redak, R.A. and Bethke, J.A. 1997. Control of silverleaf whitefly on poinsettia under greenhouse conditions, Fall 1996. Arthropod Management Tests 22: 387.
- 3635.Redak, R.A., Bethke, J.A. and Costa, H.C. 1998. Control of silverleaf whitefly on poinsettia under greenhouse conditions, summer 1997. Arthropod Management Tests 23: 350.

- 3636.Reddy, A.S., Azam, K.M., Rosaiah, B., Rao, T.B., Rao, B.R. and Rao, N.V. 1989. Biology and management of whitefly *Bemisia tabaci* (Gennadius) on cotton. *Andhra Agric. J. (India)* 36(2-3): 99-103.
- 3637.Reddy, A.S. and Rao, N.V. 1989. Cotton whitefly (*Bemisia tabaci* Genn.): A review. *Indian J. Plant Prot.* 17: 171-179.
- 3638.Reddy, A.S. and Rao, N.V. 1993. Efficacy and selectivity of insecticides of natural origin on pests of cotton, *Gossypium hirsutum*. pp. 170-174. *In* Botanical Pesticides and Integrated Pest Management. Indian Society of Tobacco Science, Rajahmundry, India.
- 3639.Reddy, A.S., Reddy, O.C., Rosaiah, B. and Bhaskara-Rao, T. 1987. Studies on the resurgence of spider mites and whiteflies of cotton. pp. 174-179. *In* S. Jayaraj (Ed.), Resurgence of Sucking Pests: Proceedings of National Symposium. Centre For Plant Protection Studies, Tamil Nadu Agric. Univ.
- 3640.Reddy, A.S., Rosaiah, B. and Bhaskara-Rao, T. 1985. Control of cotton whitefly in Andhra Pradesh. *Indian Farming* 35(8): 19, 12-22.
- 3641.Reddy, A.S., Rosaiah, B. and Rao, T.B. 1989. Seasonal occurrence of whitefly (*Bemisia tabaci* (Genn.) on cotton and its control. *Andhra Agric. J. (India)* 36(4): 275-279.
- 3642.Reddy, G.P.V. and Krishnamurthy, M.M. 1989. Insect pest management in cotton. *Pesticides* 23(10): 18-19.
- 3643.Reddy, K.R. and Singh, D.P. 1993. Inheritance of resistance to mungbean yellow mosaic virus. *Madras Agric. J.* 80(4): 199-201.
- 3644.Reddy, K.S. and Yaraguntai, R.C. 1979. Assessment of loss in tomato (*Lycopersicon esculentum* Mill.) by leaf curl virus *Bemisia tabaci* vectors. *Food Farming Agric.* 10(7): 258-259. [CATIE (1992)]
- 3645.Reddy, K.S. and Yaraguntai, R.C. 1981. Virus vector relationship in leaf curl disease of tomato whitefly *Bemisia tabaci* on *Lycopersicon esculentum* disease transmission. *Indian Phytopathol.* 34(3): 310-313. [CATIE (1992)]
- 3646.Reddy, K.S., Yaraguntai, R.C. and Sastry, K.S. 1981. Strains of leaf curl virus of tomato in India. *Zeitschrift für Pflanzenkrankheiten und Pflanzenschutz* 88: 400-404. [Cock (1986)]
- 3647.Reddy, P.S.R., Bavaji, J.N., Reddy, P.S. and Reddy, A.S. 1991. Reaction of certain *hirsutum* cotton cultures/varieties to whitefly *Bemisia tabaci* (Genn.) under rice fallows conditions. *Andhra Agric. J. (India)* 38(1): 91-92.
- 3648.Regu, K. and David, B.V. 1991. A new species of *Bemisia* from India with a key to Indian species. *Entomon* 16: 77-81.
- 3649.Reiderne-Saly, K. 1992. [Survey on the spread of *Trialeurodes* spp. in Budapest in 1991]. *Novenyvedelem (Hungary)* 28(4): 145-148. [Hungarian, English summary]
- 3650.Reifman, V.G. and Polivanova, T.A. 1969. Virus diseases of soybean in the Soviet Far East. *Trudy biologo-Pochvennogo Inst. (Viral Agric. Dis. Far East.)* 1: 83-104. [Cock (1986), Russian]
- 3651.Reina, J. and Bejarano, E.R. 1996. Perspectivas en la obtención de resistencia transgénica para TYLCV. pp. 87-90. *In* J.L. Cenis (Ed.), *El Virus del Rizado Amirillo (Hoja en Cuchara) del Tomate (TYLCV) y Su Vector Bemisia tabaci*, Jornadas 8. Consejería de Medio Ambiente, Agricultura y Agua, Murcia, Spain.
- 3652.Reina, J., Morilla, G., Bejarano, E.R., Rodríguez, M.D. and Janssen, D. 1999. First report of *Capsicum annum* plants infected by tomato yellow leaf curl virus. *Plant Dis.* 83(12): 1176.
- 3653.Reis, P.R., De Souza, J.C. and Goncalves, N.P. 1998. Pragas da videira tropical. [Portuguese]. *Informe Agropecuario Belo Horizonte* 19(194): 92-95.
- 3654.Reisman, D., Ricciardi, R.P. and Goodman, R.M. 1979. The size and topology of single-stranded DNA from bean golden mosaic virus. *Virology* 97: 388-395. [Cock (1986)]
- 3655.Rektorik, R.J. and Wright, J.E. 1992. Application technology for whitefly control with NATURALIS, a biorational insecticide. pp. 858-859. *In* D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
- 3656.Renou, A. and Chenet, T. 1988. Efficacy of biphenthrin on cotton crop in north Cameroon. *Coton Fibres Trop.* 43(3): 227-233. [Cock (1993), English and French]
- 3657.Renou, A. and Chenet, T. 1988. Perfecting a technique for the rapid evaluation of cotton plant infestation by nymphal in stars of whitefly. *Coton Fibres Trop.* 63(4): 293-298. [French]
- 3658.Renou, A. and Chenet, T. 1989. Efficacite de matieres actives insecticides contre les stades fixes de l'aleurode *Bemisia tabaci* (Genn.) en culture cotonniere au Nord-Cameroun. [The efficacy of insecticidal active ingredients against the stationary stages of the aleyrodid *Bemisia tabaci* (Genn) in cotton crops in North Cameroon]. *Coton Fibres Trop.* 44(1): 21-29. [Cock (1993), French]
- 3659.Resendiz-Ruiz, M.E. 1993. A new predator on the whitefly. *Southwest. Entomol.* 18: 147-148.
- 3660.Rethwisch, M.D., McDaniel, C.W., Shaw, M. and Thiessen, J. 1993. Evaluation of systemic insecticides for sweetpotato whitefly control on seedling cauliflower 1991. *Insecticide Acaricide Tests* 18: 117-118.
- 3661.Rethwisch, M.D., McDaniel, C.W., Shaw, M. and Thiessen, J. 1993. Sweetpotato whitefly control on broccoli 1991. *Insecticide Acaricide Tests* 18: 88-89.
- 3662.Rethwisch, M.D., Shaw, M., Thiessen, J., McDaniel, C.W. and Zaccaria, J.J. 1993. Evaluation of potential controls of sweetpotato whitefly on cauliflower 1991-1992. *Insecticide Acaricide Tests* 18: 118-119.
- 3663.Retuerma, M.L., Pableo, G.O. and Price, W.C. 1974. Preliminary study of the transmission of Philippine tomato leaf curl virus by *Bemisia tabaci* (Genn.). *Philippine J. Plant Indus.* 37(1-2): 45-49. [Cock (1986)]
- 3664.Revington, G.N., Sunter, G. and Bisaro, D.M. 1989. DNA sequences essential for replication of the b genome component of tomato golden mosaic virus. *Plant Cell* 1: 985-992.
- 3665.Reyes-Chavez, E., Reyes-Guerrero, D. and Byerly-Murphy, K.F. 1998. Inmigracion y colonizacion de mosca blanca e incidencia y severidad de enfermedades virales en jitomate. [White fly immigration and settling and incidence of viral diseases in tomato]. pp. 209-210. *In* Proceedings 21st National Congress of Biological Control, Rio Bravo, Tampaulipas, Mexico, 5-6 Nov 1998. INIFAP. [Spanish]
- 3666.Reyes, G.M., Martinez, A.L. and Chinte, P.T. 1959. Three virus diseases of plants new to the Philippines. *FAO Plant Prot. Bull.* 7(11): 141-143. [Cock (1986)]
- 3667.Reyes, M., Abud-Antun, A.J., Sension, E., Dupuy, J. and Barreiro, F.A. 1991. Incidencia de *Bemisia tabaci* Gennadius (Homoptera: Aleyrodidae) y sus enemigos naturales en el cultivo del tomate (*Lycopersicon esculentum* Mill.), asociado con otros cultivos en el valle de Azua. [Incidence of *Bemisia tabaci* Gennadius (Homoptera: Aleyrodidae) and the natural enemies on tomato crops (*Lycopersicon esculentum* Mill.) associated with other crops in the Valley of Azua]. pp. 471-477. *In* Caribbean Meetings on Biological Control, Guadeloupe, French Antilles, 5-7 November 1990. Institut National De La Recherche Agronomique, Paris. [Spanish, English summary]
- 3668.Reynaud, P. 2000. *Bemisia tabaci*: Situation and possibilities of development in France. *Phytoma (France)* 527: 18-21. [French, English summary]
- 3669.Rezende, J.A.M., Brown, J.K., Lourencao, A.L. and Bedendo, I.P. 1997. Observations on the reduced incidence of powdery mildew on zucchini squash silvered leaves in Brazil. *Fitopatol. Brasileira* 22(4): 568.
- 3670.Rich, G.J. and Womble, A. 1992. Control of *Bemisia tabaci* (Gennadius) with fenpropathrin. pp. 698-700. *In* D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
- 3671.Richardson, J.M. and Lembright, H.W. 1990. Extending chlorpyrifos residual activity for desert cotton insect control. *Down to Earth* 46(1): 1-4. [Cock (1993)]

- 3672.Richter, M. and Linss, H. 1993. [Susceptibility of Poinsettia (cultivars) against *Bemisia tabaci*]. Gartenbau Magazin (Germany) 2(8): 50-52. [ German]
- 3673.Rigden, J.E., Dry, I.B., Mullineaux, P.M. and Rezaian, M.A. 1993. Mutagenesis of the virion-sense open reading frames of tomato leaf curl geminivirus. Virology 193: 1001-1005.
- 3674.Riis, L. 1999. [Whitefly as pest and vector: Successful pan-tropical cooperation to obtain more yield and less environmental impact]. Jord Og Viden (Denmark) 144(11): 12-14. [ Danish]
- 3675.Riley, D. 1994. Insecticide control of sweetpotato whitefly in South Texas. Subtrop. Plant Sci. 46: 45-49.
- 3676.Riley, D., Batal, D. and Wolff, D. 2001. Resistance in glabrous-type *Cucumis melo* L. to whiteflies (Homoptera: Aleyrodidae). J. Entomol. Sci. 36(1): 46-56.
- 3677.Riley, D., Nava-Camberos, U. and Allen, J. 1996. Population dynamics of *Bemisia* in agricultural systems. pp. 93-109. In D. Gerling and R.T. Mayer (Eds.), *Bemisia* : 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
- 3678.Riley, D. and Wolfenbarger, D. 1993. Cultivated hosts and population dynamics of sweetpotato whitefly in the Lower Rio Grande Valley of TX. pp. 667-670. In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3679.Riley, D.G. 1995. Control of whitefly in cotton with Capture and various additives, 1992. Arthropod Management Tests 20: 213-215.
- 3680.Riley, D.G. 1995. Control of whitefly in cotton with Danitol, Asana, Monitor, Orthene and combinations, 1992. Arthropod Management Tests 20: 215-216.
- 3681.Riley, D.G. 1995. Melon cultivar response to *Bemisia*. Subtrop. Plant Sci. 47: 39-45.
- 3682.Riley, D.G. and Ciomperlik, M.A. 1997. Regional population dynamics of whitefly (Homoptera: Aleyrodidae) and associated parasitoids (Hymenoptera: Aphelinidae). Environ. Entomol. 26(5): 1049-1055.
- 3683.Riley, D.G. and Palumbo, J.C. 1995. Action thresholds for *Bemisia argentifolii* (Homoptera: Aleyrodidae) in cantaloupe. J. Econ. Entomol. 88(6): 1733-1738.
- 3684.Riley, D.G. and Palumbo, J.C. 1995. Interaction of silverleaf whitefly (Homoptera: Aleyrodidae) with cantaloupe yield. J. Econ. Entomol. 88(6): 1726-1732.
- 3685.Riley, D.G., Sparks, A.N. and Norman, J. 1991. The sweetpotato whitefly in the lower Rio Grande Valley. Texas A&M Univ. (mimeograph)
- 3686.Riley, D.G. and Sparks, A.N., Jr. 1993. Managing the sweetpotato whitefly in the Lower Rio Grande Valley of Texas. Bull. Texas Agric. Exp. Stn. 5082: 1-12.
- 3687.Riley, D.G., Tan, W.J. and Wolfenbarger, D. 1999. Esterase isozymes associated with bifenthrin resistance in the silverleaf whitefly (Homoptera: Aleyrodidae). J. Entomol. Sci. 34(3): 295-304.
- 3688.Riley, D.G., Tan, W.J. and Wolfenbarger, D. 2000. Activities of enzymes associated with inheritance of bifenthrin resistance in the silverleaf whitefly, *Bemisia argentifolii*. Southwest. Entomol. 25(3): 201-211.
- 3689.Rimon, D. 1982. II. The whitefly as a factor in contamination by sugars and in fiber stickiness. Div. Industrial Crops, Volcani Center, Bet Dagan, Israel : 35 pp.
- 3690.Rimon, D. 1983. III. The effect of application of sugars on cotton fibers. Div. Industrial Crops, Volcani Center, Bet Dagan, Israel : 13 pp.
- 3691.Rimon, D., Kaganovskii, I. and Altachan, L. 1983. IV. *Bemisia tabaci* (Gennad.) as a factor in sugars contamination and stickiness of fibers in the 1982 season. Div. Industrial Crops, Volcani Center, Bet Dagan, Israel : 8pp.
- 3692.Rios-Labrada, H., Fernandez-Almirall, A. and Galarraga, E.C. 1997. Response of Cuban pumpkin (*Cucurbita moschata* Duch.) to abiotic and biotic stress interactions. Rep. Cucurbit Genetics Coop. 20: 50-52.
- 3693.Ripper, W.E. and George, L. 1965. Cotton pests of the Sudan: Their habits and control. pp. 90-106. In Blackwell Scientific Publications, Oxford, UK.
- 3694.Rivas, G.G. and Lastra, R. 1993. Deteccion no radiactiva de geminivirus en tomate mediante hibridacion de acidos nucleicos. [Non-radioactive detection of geminiviruses in tomatoes through hybridization of nucleic acids]. Manejo Integrado de Plagas (Costa Rica) 30: 7-10. [ Spanish, English summary]
- 3695.Rivas, G.G., Lastra, R. and Hilje, L. 1994. Retardo de la virosis transmitida por *Bemisia tabaci* (Gennadius) en tomate mediante semilleros cubiertos. [Retardation of the virus transmitted by *Bemisia tabaci* (Gennadius) in semi-covered seed-beds]. Manejo Integrado de Plagas (Costa Rica) 31: 12-16. [ Spanish, English summary]
- 3696.Rivas, G.G., Ramirez, P., Cubillo, D. and Hilje, L. 1995. Deteccion de virus en plantas silvestres asociadas con el tomate y chile dulce en Costa Rica. [Detection of viruses in wild plants associated with tomatoes and sweet peppers in Costa Rica]. Manejo Integrado de Plagas (Costa Rica) 38: 37-39. [ Spanish, English summary]
- 3697.Rivas, G.G., Ramirez, R., Cubillo, D. and Hilje, L. 1995. Translocacion y cuantificacion del adn viral de geminivirus asociados con el mosaico amarillo del tomate. [ Translocation and quantification of geminiviruses associated with the tomato yellow mosaic disease]. Manejo Integrado de Plagas (Costa Rica) 38: 20-24. [ Spanish, English summary]
- 3698.Rivnay, E. 1962. The tobacco whitefly, *Bemisia tabaci* Gennad. pp. 56-59. In W.W. Weisbach (Ed.), Field Crop Pests in the Near East. Dr. W. Junk, Den Haag, Monographie Biologicae Uilgeverij.
- 3699.Rivnay, T. and Gerling, D. 1987. Aphelinidae parasitoids (Hymenoptera: Chalcidoidea) of whiteflies (Homoptera: Aleyrodidae) in Israel, with description of three new species. Entomophaga 32(5): 463-475. [ Cock (1993)]
- 3700.Rizk, G.A. and Kamel, E.H. 1991. Evaluation of reldan bioencapsulated for the control of sucking insects, Homoptera, Aphididae and Aleyrodidae in cotton fields. Minia J. Agric. Res. Devel. (Egypt) 13(1): 233-243.
- 3701.Rizk, G.N. and Ahmed, K.G. 1981. Population dynamics of some insect pests attacking squash plants, *Cucurbita pepo* L. in Iraq. Res. Bull., Faculty Agric., Ain Shams Univ. 1653(8): 1-5. [ Cock (1986)]
- 3702.Rizvi, S.M.A. and Singh, A.K. 1994. Succession of insect pests infesting rajmash in Eastern Uttar Pradesh. Indian J. Plant Prot. 22(2): 226-227.
- 3703.Robb, K.L. 1991. Biology and control of whiteflies in floriculture greenhouses. Flower and Nursery Report for Commercial Growers, Agric. Ext. Serv., Univ. California, Berkeley, 8 pp.
- 3704.Roberson, R. 1996. Pakistan cotton production emerges from difficult growing conditions. pp. 459-460. In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3705.Roberts, C.W., Cheung, P.S.R. and Perkins, H.H., Jr. 1978. Implications of monosaccharides in sticky cotton processing. Part II: Effects of growing conditions on fiber contaminants. Textile Res. J. 48: 91-96.
- 3706.Roberts, C.W., Koenig, H.S., Merrill, R.G., Cheung, P.S.R. and Perkins, H.H., Jr. 1976. Implications of monosaccharides in sticky cotton processing. Textile Res. J. 46: 374-380.
- 3707.Roberts, I.M. 1989. Indian cassava mosaic virus: ultrastructure of infected cells. J. Gen. Virol. 70: 2729-2739.
- 3708.Roberts, I.M., Robinson, D.J. and Harrison, B.D. 1984. Serological relationships and genome homologies among geminiviruses. J. Gen. Virol. 65: 1723-1730. [ Cock (1986)]

- 3709.Roberts, P.M. and Tankersley, T.B. 2001. Silverleaf whitefly: an economic pest of cotton in Georgia ? p. 1133. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3710.Roberts, W. 1930. Note on the American cotton crop failure in the Punjab. *Agric. J. India* 24: 77-81.
- 3711.Robertson, I.A.D. 1987. The whitefly, *Bemisia tabaci* (Gennadius) as a vector of African cassava mosaic virus at the Kenya coast and ways in which the yield losses in cassava, *Manihot esculenta* Crantz caused by the virus can be reduced. [French summary]. *Insect Sci. Appl.* 8(4-6): 797-801. [Cock (1993)]
- 3712.Robinson, J.R.C. and Taylor, M.J. 1996. Economic case studies of whitefly management. pp. 225-236. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia : 1995 Taxonomy, Biology, Damage, Control and Management*. Intercept Ltd., Andover, Hants, UK.
- 3713.Robledo, C.T. and Sagahón, J.C.R. 1999. Campaña contra la mosquita (Homoptera: Aleyrodidae) Mexico. pp. 165-174. *In* 7th Taller Latinoamericano y del Caribe Sobre Moscas-Blancas y Geminivirus. IPA, Recife, PE, Brazil. [Spanish]
- 3714.Rochester, D.E., De Paulo, J.J., Fauquet, C.M. and Beachy, R.N. 1994. Complete nucleotide sequence of the geminivirus tomato yellow leaf curl virus, Thailand isolate. *J. Gen. Virol.* 75: 477-485.
- 3715.Rochester, D.E., Kosiratana, W. and Beachy, R.N. 1990. Systemic movement and symptom production following agroinoculation with a single DNA of tomato yellow leaf curl geminivirus (Thailand). *Virology* 178: 520-526.
- 3716.Rochester, D.E., Jr. 1991. Genetic analysis of tomato yellow leaf curl, a geminivirus. *Dissertation Abst. Int.* 53(05B): 2122.
- 3717.Rochow, W.F. and Ball, E.M. 1967. Serological blocking of aphid transmission of barley yellow dwarf virus. *Virology* 33: 359-362.
- 3718.Rodrigo, P.A. 1947. Soybean culture in the Philippines. *Philippine J. Agric.* 13: 1-22. [Cock (1986)]
- 3719.Rodrigues, F.D., Borges, A.C.F., DosSantos, M.R., Fernandes, J.J. and Junior, A.D. 1997. Whitefly population fluctuation and golden mosaic incidence in bean. *Pesquisa Agropecuaria Brasileira* 32(10): 1023-1027. [Portuguese]
- 3720.Rodríguez, E. 2000. Historia y situación de la producción de frijol en los países latinoamericanos afectados por geminivirus transmitidos por mosca blanca: América Central: Panamá. pp. 47-48. *In* Bean Golden Mosaic and Other Diseases of Common Bean Caused by Whitefly-Transmitted Geminiviruses in Latin America. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. [Spanish]
- 3721.Rodríguez, H., De los Angeles Pino, M., Leon, A. and Terry, E. 1998. Effect of tomato-maize association in the behavior of *Bemisia tabaci* (Gennadius) in optimal growing season. [Spanish, English summary]. *Rev. Protección Vegetal* 13(2): 97-102.
- 3722.Rodríguez, M.D. 1996. Incidencia de *B. tabaci* en un sistema de protección integrada: experiencia en Almería. pp. 71-74. *In* J.L. Cenis (Ed.), *El Virus del Rizado Amirillo (Hoja en Cuchara) del Tomate (TYLCV) y Su Vector Bemisia tabaci*, Jornadas 8. Consejería de Medio Ambiente, Agricultura y Agua, Murcia, Spain.
- 3723.Rodríguez, M.D. and Téllez, M.M. 2000. La paga principal de los cultivos hortícolas protegidos en los climas cálidos: La mosca blanca. *La Industria del Invernadero Extra*: 30-36. [Spanish]
- 3724.Rodríguez-Rodríguez, J.R. and Yoshii, K. 1990. Tolerancia varietal de frijol al "Mosaico dorado" y control químico del vector *Bemisia tabaci* Genn. en Papantla, Veracruz. [Tolerance variety to bean "Golden mosaic" virus and chemical protection of vector *Bemisia tabaci* Genn. at Papantla, Veracruz]. *Agric. Técnica México* 16(1/2): 19-32. [Spanish, English summary]
- 3725.Rodríguez-Rodríguez, M.D., Moreno, R., Tellez, M.M., Rodríguez-Rodríguez, M.P. and Fernández-Fernández, R. 1994. *Eretmocerus mundus* (Mercet), *Encarsia lutea* (Masi) y *Encarsia transvena* (Timberlake) (Hym., Aphelinidae) parasitoides de *Bemisia tabaci* (Homoptera: Aleyrodidae) en los cultivos hortícolas protegidos almerienses. [ *Eretmocerus mundus* (Mercet), *Encarsia lutea* (Masi) and *Encarsia transvena* (Timberlake) (Hym., Aphelinidae), parasitoids of *Bemisia tabaci* (Homoptera: Aleyrodidae) in protected vegetable crops in Almería]. *Boletín de Sanidad Vegetal, Plagas* 20(3): 695-702. [Spanish, English summary]
- 3726.Rogers, S.G., Bisaro, D.M., Horsch, R.B., Fraley, R.T., Hoffmann, N.L., Brand, L., Elmer, J.S. and Lloyd, A. 1986. Tomato golden mosaic virus A component DNA replicates autonomously in transgenic plants. *Cell* 45: 593-600.
- 3727.Rogers, S.G., Elmer, J.S., Sunter, G., Gardiner, W.E., Brand, L., Browning, C.K. and Bisaro, D.M. 1989. Molecular genetics of tomato golden mosaic virus. pp. 199-215. *In* Staskawicz, P. Ahlquist and Yoder (Eds.), *Molecular Biology of Plant-Pathogen Interactions*. A. R. Liss, New York, USA.
- 3728.Rojas, A. and Anderson, P. 1994. Situación actual del mosaico dorado del frijol en la América Central: Nicaragua. [Current situation of bean golden mosaic virus in Latin America: Nicaragua]. pp. 51-61. *In* F.J. Morales (Ed.), *Bean Golden Mosaic: Research Advances*. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. [Spanish]
- 3729.Rojas, A., Kvarnheden, A. and Valkonen, J.P.T. 2000. Geminiviruses infecting tomato crops in Nicaragua. *Plant Dis.* 84(8): 843-846.
- 3730.Rojas, M.R., Gilbertson, R.L., Russell, D.R. and Maxwell, D.P. 1993. Use of degenerate primers in the polymerase chain reaction to detect whitefly-transmitted geminiviruses. *Plant Dis.* 77(4): 340-347.
- 3731.Rojas, T., Pons, N. and Arnal, E. 1998. *Cladosporium herbarum* on whiteflies (Homoptera: Aleyrodidae), in Venezuela. [Spanish, English summary]. *Boletín Entomol. Venezolana, Serie Monografías* 13(1): 57-65.
- 3732.Romeis, J. and Zebitz, C.P.W. 1997. Searching behavior of *Encarsia formosa* as mediated by color and honeydew. *Entomol. Exp. Appl.* 82(3): 299-309.
- 3733.Romero, M.A. 1995. Aspectos básicos de la mosca blanca. [Basic aspects of white fly]. *Fitofilo (México)* 88: 11-23. [Spanish]
- 3734.Romero-Rosales, F. 1995. Resistencia varietal a la mosca blanca *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae). [Varietal resistance to white fly (*Bemisia tabaci*) (Gennadius) (Homoptera: Aleyrodidae)]. *Fitofilo (México)* 88: 127-141. [Spanish]
- 3735.Ronchi, L., Gervasini, E. and Sama, A. 1994. Poinsettia: biological control of aleyrodids. *Colture Protette (Italy)* 23(2): 53-58. [Italian]
- 3736.Rosaiah, B., Reddy, A.S., Rao, T.B., Reddy, B.M., Rao, N.V., Rao, B.R., Reddy, V.C. and Srinivasulu, R. 1989. Varietal reaction to cotton whitefly, *Bemisia tabaci* (Genn.). *Andhra Agric. J. (India)* 36(4): 325-328.
- 3737.Rose, M. and Zolnerowich, G. 1997. *Eretmocerus* Haldeman (Hymenoptera: Aphelinidae) in the United States, with descriptions of new species attacking *Bemisia (tabaci) complex* (Homoptera: Aleyrodidae). *Proc. Entomol. Soc. Washington* 99(1): 1-27.
- 3738.Rose, M., Zolnerowich, G. and Hunter, M.S. 1996. Systematics, *Eretmocerus*, and biological control. pp. 477-497. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia : 1995 Taxonomy, Biology, Damage, Control and Management*. Intercept Ltd., Andover, Hants, UK.
- 3739.Rosell, R.C., Bedford, I.D., Frohlich, D.R., Gill, R.J., Brown, J.K. and Markham, P.G. 1997. Analysis of morphological variation in distinct populations of *Bemisia tabaci* (Homoptera: Aleyrodidae). *Ann. Entomol. Soc. Am.* 90(5): 575-589.
- 3740.Rosell, R.C., Bedford, I.D., Markham, P.G., Frohlich, D.R. and Brown, J.K. 1996. Morphological variation in *Bemisia* populations. pp. 147-149. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia : 1995 Taxonomy, Biology, Damage, Control and Management*. Intercept Ltd., Andover, Hants, UK.

3741. Rosell, R.C., Lichty, J.E. and Brown, J.K. 1995. Ultrastructure of the mouthparts of adult sweetpotato whitefly, *Bemisia tabaci* Gennadius (Homoptera: Aleyrodidae). *Int. J. Insect Morph. Embryol.* 24(3): 297-306.
3742. Rosell, R.C., Torres Jerez, I. and Brown, J.K. 1999. Tracing the geminivirus-whitefly transmission pathway by polymerase chain reaction in whitefly extracts, saliva, hemolymph, and honeydew. *Phytopathology* 89(3): 239-246.
3743. Rosell, H.W. 1986. Rice yellow mottle and African soybean dwarf, newly discovered virus diseases of economic importance in West Africa. pp. 146-153. *In* International Symposium on Virus Diseases of Rice and Leguminous Crops in the Tropics. Tropical Agriculture Research Centre, Yatabe, Tsukuba, Ibaraki, Japan. [Cock (1993)]
3744. Rosset, P., Meneses, R., Lastra, R. and Gonzalez, W. 1990. Estimacion de perdidas e identificacion del geminivirus transmitido al tomate por la mosca blanca *Bemisia tabaci* Genn. (Homoptera: Aleyrodidae) en Costa Rica. [Estimations of losses and identification of the geminivirus transmitted to tomatoes by whitefly *Bemisia tabaci* Genn. (Homoptera: Aleyrodidae) in Costa Rica]. *Manejo Integrado de Plagas (Costa Rica)* 15: 24-34. [Spanish, English summary]
3745. Rosset, P.M. 1986. Ecological and economic aspects of pest management and polycultures of tomatoes in Central America. [University of Michigan]. Dissertation Abst. Int. 47(03B): 904.
3746. Rosset, P.M. 1988. [Control of insect pests in tomato: some considerations on the Central American experience]. *Manejo Integrado de Plagas (Costa Rica)* 7: 1-12. [Cock (1993), Spanish]
3747. Rossetto, D., Costa, A.S., Miranda, M.A.C., Nagai, V. and Abramides, E. 1977. Diferencias na oviposicao de *Bemisia tabaci* em variedades de soja. *Anais Soc. Entomol. Brasil* 6(2): 256-263. [Cock (1986)]
3748. Rote, N.B. and Puri, S.N. 1991. Population dynamics of whitefly on cotton and its relationship with weather parameters. *J. Cotton Res. Dev.* 5(2): 181-189.
3749. Rote, N.B. and Puri, S.N. 1992. Effects of fertilizer application on incidence of whitefly on different cotton cultivars. *J. Maharashtra Agric. Univ. (India)* 17(1): 45-48.
3750. Rote, N.B., Puri, S.N., Butler, G.D., Jr. and Henneberry, T.J. 1992. Whitefly population levels, fecundity and developmental period on cotton following insecticidal applications. *J. Appl. Zool. Res.* 3(1): 1-6.
3751. Rowland, M., Hackett, B. and Stribley, M. 1991. Evaluation of insecticides in field-control simulators and standard laboratory bioassays against resistant and susceptible *Bemisia tabaci* (Homoptera: Aleyrodidae) from Sudan. *Bull. Entomol. Res.* 81(2): 189-199. [Cock (1993)]
3752. Rowland, M., Pye, B., Stribley, M., Hackett, B., Denholm, I. and Sawicki, R.M. 1990. Laboratory apparatus and techniques for the rearing and insecticidal treatment of whitefly *Bemisia tabaci* (Homoptera: Aleyrodidae) under simulated field conditions. *Bull. Entomol. Res.* 80(2): 209-216.
3753. Rowland, M.W., Hackett, B., Stribley, M.F. and Sawicki, R.M. 1990. The cotton-whitefly field control simulator: laboratory apparatus for evaluating insecticides and resistance management strategies under simulated field conditions. pp. 1195-1200. *In* Brighton Crop Protection Conference: Pests and Diseases. British Crop Protection Council, Thornton Heath, UK. [Cock (1993)]
3754. Roychoudhury, R. and Jain, R.K. 1993. Effect of climatological factors on the population of aphids and whiteflies in Lucknow region of Uttar Pradesh (India). *Int. J. Trop. Plant Dis.* 11(2): 197-206.
3755. Roye, M.E., Henry, N.M., Burrell, P.D., McLaughlin, W.A., Nakhla, M.K. and Maxwell, D.P. 2000. A new tomato-infecting begomovirus in Barbados. *Plant Dis.* 84(12): 1342.
3756. Roye, M.E., McLaughlin, W.A., Spence, J.D. and Maxwell, D.P. 1999. The common weed *Macroptilium lathyroides* is not a source of crop-infecting geminiviruses from Jamaica. *Trop. Agric.* 76(4): 256-262.
3757. Roye, M.E., Wernecke, M.E., McLaughlin, W.A., Nakhla, M.K. and Maxwell, D.P. 1999. Tomato dwarf leaf curl virus, a new bipartite geminivirus associated with tomatoes and peppers in Jamaica and mixed infection with tomato yellow leaf curl virus. *Plant. Pathol.* 48(3): 370-378.
3758. Rubeiz, I.G., Aslam, M., Chahine, H. and Alassir, I.A. 1995. Whitefly populations in greenhouse cantaloupe as affected by poultry manure. *Biol. Agric. Hortic.* 12(3): 277-281.
3759. Rubinstein, G. and Czosnek, H. 1997. Long-term association of tomato yellow leaf curl virus with its whitefly vector *Bemisia tabaci*: effect on the insect transmission capacity, longevity and fecundity. *J. Gen. Virol.* 78(10): 2683-2689.
3760. Rubinstein, G., Morin, S. and Czosnek, H. 1999. Transmission of tomato yellow leaf curl geminivirus to imidacloprid treated tomato plants by the whitefly *Bemisia tabaci* (Homoptera: Aleyrodidae). *J. Econ. Entomol.* 92(3): 658-662.
3761. Rubio, L., Soong, J., Kao, J. and Falk, B. W. 1999. Geographic distribution and molecular variation of isolates of three whitefly-borne closteroviruses of cucurbits: lettuce infectious yellows virus, cucurbit yellow stunting disorder virus, and beet pseudo-yellows virus. *Phytopathology* 89(8): 707-711.
3762. Ruisinger, M. and Backhaus, G.F. 1994. Control of *Trialeurodes vaporariorum* and *Bemisia tabaci* with *Encarsia formosa* in stock cultures of *Euphorbia pulcherrima*. *Gesunde Pflanzen* 46: 122-127. [German, English summary]
3763. Ruiz, J. and Medina Z. J. 2001. Avances en el manejo integrado de *Bemisia tabaci* en tomate y chile en Oaxaca, México. [Advances in the integrated management of *Bemisia tabaci* on tomato and pepper in Oaxaca, Mexico]. *Manejo Integrado de Plagas (Costa Rica)* 59: 34-40. [Spanish, English summary]
3764. Ruiz-Vega, J. and Aquino-Bolaños, T. 1999. Manejo de *Bemisia tabaci* mediante barreras vivas y *Paecilomyces* en Oaxaca, México. [Management of *Bemisia tabaci* with living barriers and *Paecilomyces* in Oaxaca, Mexico]. *Manejo Integrado de Plagas (Costa Rica)* 52: 68-73. [Spanish, English summary]
3765. Ruiz-Vega, J., Arce-Gonzalez, F., Aquino, B.T., Garcia-Garcia, J. and Martinez, D. 1998. Control biológico integrado de mosquita blanca *Bemisia tabaci* Genn. (Homoptera: Aleyrodidae) en chile de agua. [Integrated biological control of white fly *Bemisia tabaci* Genn. (Homoptera: Aleyrodidae) in water chili]. pp. 203-205. *In* Proceedings 21st National Congress of Biological Control, Rio Bravo, Tampaulipas, Mexico, 5-6 Nov 1998. INIFAP. [Spanish]
3766. Rumei, X. 1991. Improvements of the plant-pest-parasitoid PPP model and its application on whitefly *Encarsia* population dynamics under different release methods. *J. Appl. Entomol.* 112(3): 274-287.
3767. Rumei, X. 1996. The occurrence and distribution of *Bemisia* in China. pp. 125-131. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia*: 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
3768. Rushing, A., Sunter, G., Gardiner, W., Dute, R. and Bisaro, D.M. 1987. Ultrastructural aspects of tomato golden mosaic virus infection of tobacco. *Phytopathology* 77(8): 1231-1236.
3769. Rushtapakornchai, W. and Petchwicht, P. 1996. Efficiency of some insecticides for controlling tobacco whitefly *Bemisia tabaci* and leaf miner *Liriomyza trifolii* on tomato. [Thai, English summary]. *Khon Kaen Agric. J.* 24(4): 184-189.
3770. Russell, L.M. 1948. The North American species of whiteflies of the genus *Trialeurodes*. USDA, Misc. Publ. No. 635, 85 pp.
3771. Russell, L.M. 1957. Synonyms of *Bemisia tabaci* (Gennadius) (Homoptera, Aleyrodidae). *Bull. Brooklyn Entomol. Soc.* 52: 122-123. [Cock (1986)]
3772. Russell, L.M. 1975. Collection records of *Bemisia tabaci* (Gennadius) in the United States. USDA, Coop. Econ. Insect Rep. 25: 229-230.
3773. Russell, L.M. and Etienne, J. 1985. A list of the Aleyrodidae of the Island of Reunion. *Proc. Entomol. Soc. Washington* 87: 202-206. [Cock (1986)]

3774. Russell, T.E. 1982. Effect of cotton leaf crumple (CLC) disease on stub and planted cotton. pp. 43-47. *In* Cotton, A College of Agriculture Report, Series P-56. University of Arizona, Tucson.
3775. Russo, G. 1942. I parassiti animali dannosi alle coltivazioni di cotone. Ricerche ed osservazioni eseguite in Italia nel 1941. *Ann. Facolta Agric. Univ. Pisa* 5(N.S.): 436-480. [Cock (1986), Italian with English summary]
3776. Russo, M., Cohen, S. and Martelli, G.P. 1980. Virus-like particles in tomato plants affected by the yellow leafcurl disease. *J. Gen. Virol.* 49: 209-213. [Cock (1986)]
3777. Rustamani, M.A., Hussain, T., Baloch, H.B., Talpur, M.A. and Mal, K. 1994. Comparative effectiveness of different insecticides in controlling whitefly and mosaic disease on chilies. *Proc. Pakistan Congress Zool.* 14: 61-64.
3778. Rustamani, M.A., Memon, N., Leghari, M.H., Dhaunroo, M.H. and Sheikh, S.A. 1999. Impact of various fertilizer levels on the incidence of sucking complex in cotton. *Pakistan J. Zool.* 31(4): 323-326.
3779. Ryckewaert, P. and Alauzet, C. 2001. Characterization of *Bemisia* (Hom., Aleyrodidae) from the Lesser Antilles by electrophoresis. *J. Appl. Entomol.* 125(5): 263-266.
3780. Rylski, I., Berlinger, M.J., Dahan, R. and Spiegelman, M. 1984. The effect of plastic covering and of removing one or two flower clusters, on the yield of glasshouse tomatoes. *Hassadeh* 64: 2008-2010. [Hebrew, English summary]
3781. Sabillon, A. and Bustamante, M. 1995. Evaluacion de extractos botanicos para el control de plagas del tomate (*Lycopersicon esculentum* Mill.) [Evaluation of botanical extracts for the control of tomato pests (*Lycopersicon esculentum* Mill)]. *CEIBA* 36(2): 179-187. [Spanish, English summary]
3782. Saborio, M.M. 1994. Control fitogenetico del complejo mosca blanca virus. [Phylogenetic control of the whitefly-virus complex]. *Manejo Integrado de Plagas (Costa Rica)* 34: 36-40. [Spanish, English summary]
3783. Sadof, C. and Foster, R. 1992. Whitefly Q and A for breeding plant producers. *Floriculture Indiana, Coop. Ext. Serv., Purdue Univ.* 6(2): 1, 3-4.
3784. Sahoo, B.K. and Patnaik, N.C. 1994. Insect pests in greengram and blackgram in the South Coastal Region of Orissa with notes on their seasonal activity. *Orissa J. Agric. Res.* 7(suppl.): 74-76.
3785. Sahoo, B.K. and Sahu, P.N. 1991. Evaluation of promising blackgram varieties against whitefly (*Bemisia tabaci* Genn.) and yellow mosaic. *Madras Agric. J.* 78(1-4): 93-94.
3786. Sahoo, B.K., Sahu, P.N. and Mishra, M.R. 1989. Field evaluation of greengram varieties against whitefly and yellow mosaic virus disease. *Orissa J. Agric. Res.* 2(2): 136-137. [Cock (1993)]
3787. Saikia, A.K. and Muniyappa, V. 1989. Epidemiology and control of tomato leaf curl virus in southern India. *Trop. Agric.* 66: 350-354.
3788. Saikia, A.K. and Muniyappa, V. 1989. Ultrastructural changes in phloem cells of leaf curl-affected tomato from India. *J. Phytopathol.* 124: 1-6.
3789. Saito, T. 1992. Control of *Thrips palmi* and *Bemisia tabaci* by mycoinsecticidal preparation of *Verticillium lecanii*. *Proc. Kanto-Tosan Plant Prot. Soc.* 39: 209-210. [Japanese, English summary]
3790. Saito, T. 1993. Control of two whitefly species, *Bemisia tabaci* and *Trialeurodes vaporariorum*, by a preparation of *Verticillium lecanii*. *Proc. Kanto-Tosan Plant Prot. Soc.* 40: 221-222. [Japanese, English summary]
3791. Saklani, U.D. and Mathai, P.J. 1978. Effect of insecticides on leaf curl incidence of tomato. *Pesticides* 12(8): 17-20, 25. [Cock (1986)]
3792. Saladin, F., Mendez, R.M., Nin, J.C., Angeles, R., Herrera, M. and Martinez, T. 1994. Alternativa para el manejo integrado en el control del virus del mosaico dorado del frijol en Republica Dominicana. [Alternatives for the integrated control of bean golden mosaic virus in the Dominican Republic]. pp. 183-189. *In* F.J. Morales (Ed.), *Bean Golden Mosaic: Research Advances*. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. [Spanish, English summary]
3793. Saladin-Garcia, F. and Morales, F.J. 2000. Historia y situacion de la produccion de frijol en los paises latinoamericanos afectados por geminivirus transmitidos por mosca blanca: Region caribe: Republica Dominicana. pp. 55-58. *In* *Bean Golden Mosaic and Other Diseases of Common Bean Caused by Whitefly-Transmitted Geminiviruses in Latin America*. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. [Spanish]
3794. Salama, A.E., Adam, F.A., El-Nawawy, A., Abbassy, M. and Abo-Salem, M. 1984. Sequential insecticide treatments for the control of sucking pests with regards to some of their predators. *Med. Fac. Landbouww. Univ. Gent* 49(3a): 885-891. [Cock (1986)]
3795. Salama, E.A., Al-Beshr, A.A. and Abou-Ghadir, M.F. 1994. Identification and characterization of tomato viruses in the Eastern Province of Saudi Arabia (1). *Egyptian J. Phytopathol.* 22(1): 125-141.
3796. Salas, F.J.S., Barradas, M.M. and Parra, J.R.P. 1999. Transmission trials of a cowpea severe mosaic virus isolate (CpSMV-SP) by arthropods under laboratory conditions. [Portuguese, English summary]. *Scientia Agric.* 56(2): 413-420.
3797. Salas, J. 1995. Trampas amarillas en la captura de *Bemisia tabaci* y sus parasitoides *Encarsia* y *Eretmocerus* [Yellow traps and the capture of *Bemisia* and its parasites *Encarsia* and *Eretmocerus*]. *Manejo Integrado de Plagas (Costa Rica)* 37: 39-42. [Spanish, English summary]
3798. Salas, J., Arnal, E. and Mendoza, O. 1994. Integrated management of whitefly in Venezuela. *FONAIAP Divulga* 11(45): 21-24. [Spanish]
3799. Salas, J. and Mendoza, O. 1995. Biology of the sweetpotato whitefly (Homoptera: Aleyrodidae) on tomato. [Spanish summary]. *Florida Entomol.* 78(1): 154-160.
3800. Salavatian, M. 1987. Cotton whitefly and its control. *Ministry Agric. Natural Resources, Tehran, Ext. Bull.*, 52 pp. [Persian]
3801. Salazar, E., Cubillo, D., Ramirez, P., Rivas-Platero, G. and Hilje, L. 1998. Severidad del moteado amarillo del tomate y reduccion del rendimiento del cultivo en respuesta a la densidad de adultos viruliferos de *Bemisia tabaci* [Effect of viruliferous adult *Bemisia tabaci* density on tomato yellow mottle severity and crop yield]. *Manejo Integrado De Plagas (Costa Rica)* 50: 42-50. [Spanish, English summary]
3802. Salem, I.E.A. 1995. Trichomal exudate extracts from *Hyoscyamus muticus* leaf surface highly active against the cotton whitefly *Bemisia tabaci* Genn (Aleyrodidae). *Med. Fac. Landbouww. Univ. Gent* 60(3B): 991-994.
3803. Salem, M. 1993. Distribution pattern of infestation and control of white fly *Bemisia tabaci* (Genn) on cucumber plants in the greenhouses. *Ain-Shams Univ. Ann. Agric. Sci. (Egypt)* 38(2): 783-793.
3804. Salem, R.M. 1997. Population dynamics of three piercing sucking pests and associated predators as influenced by preceding crop and the cotton chemical control programme. *Tanta J. Agric. Res. (Egypt)* 23: 219-228.
3805. Salem, R.M. 1998. Population density of some sucking pests infesting Giza 86 cotton variety in relation to associated predators under certain climatic factors in Kafr El-Sheikh. *Alexandria Sci. Exchange (Egypt)* 19(2): 197-207.
3806. Salguero, V. 1992. Perspectivas para el manejo del complejo mosca blanca-virosis. [Perspectives on the management of the whitefly-virus complex. pp. 20-26. *In* L. Hilje and O. Arboleda (Eds.), *Las moscas blancas (Homoptera: Aleyrodidae) en America Central y el Caribe*. CATIE, Turrialba, Costa Rica. [Spanish]

- 3807.Salguero, V. and Morales, J. 1994. Eficiencia de insecticidas para el control de *Bemisia tabaci* (Gennadius) en tomate. [Efficiency of insecticides for the control of *Bemisia tabaci* (Gennadius) on tomato]. Manejo Integrado de Plagas (Costa Rica) 31: 25-28. [ Spanish, English summary]
- 3808.Salih, H.S.A., Abou-Jawdah, Y.A., Nounawar, M.A. and Shougaidef, W.N. 1991. [Studies on the yellowing disease of watermelon in Jizan]. Arab J. Plant Prot. 9(1): 27-31. [ Cock (1993), Arabic, English summary]
- 3809.Salim, M., Masud, S.A. and Khan, A.M. 1987. *Orius albidipennis* (Reut.) (Hemiptera: Anthoridae) - a predator of cotton pests. Philippine Entomol. 7(1): 37-42. [ Cock (1993)]
- 3810.Salinas-Perez, R.A. 2000. Historia y situacion de la produccion de frijol en los paises latinoamericanos afectados por geminivirus transmitidos por mosca blanca: Mexico: Mexico-noroeste. pp. 13-18. *In* Bean Golden Mosaic and Other Diseases of Common Bean Caused by Whitefly -Transmitted Geminiviruses in Latin America. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. [Spanish]
- 3811.Salman, A.G.A. and Abd-El-Raof, T.K. 1983. Effect of certain pesticides used against cotton pests on three predaceous insects and honey bee workers. Bull. Entomol. Soc. Egypt Econ. Ser. 11(1978-79): 155-162. [ Cock (1986)]
- 3812.Salvucci, M.E. 2000. Effect of the alpha-glucosidase inhibitor, bromoconduritol, on carbohydrate metabolism in the silverleaf whitefly, *Bemisia argentifolii*. Arch. Insect Biochem. Physiol. 45(3): 117-128.
- 3813.Salvucci, M.E. 2000. Sorbitol accumulation in whiteflies: Evidence for a role in protecting proteins during heat stress. J. Thermal Biol. 25(5): 353-361.
- 3814.Salvucci, M.E. and Crafts-Brandner, S.J. 2000. Effects of temperature and dietary sucrose concentration on respiration in the silverleaf whitefly, *Bemisia argentifolii*. J. Insect Physiol. 46(11): 1461-1467.
- 3815.Salvucci, M.E., Hendrix, D.L. and Wolfe, G.R. 1999. Effect of high temperature on the metabolic process affecting sorbitol synthesis in the silverleaf whitefly, *Bemisia argentifolii*. J. Insect Physiol. 45(1): 21-27.
- 3816.Salvucci, M.E., Rosell, R.C. and Brown, J.K. 1998. Uptake and metabolism of leaf proteins by the silverleaf whitefly. Arch. Insect Biochem. Physiol. 39: 155-165.
- 3817.Salvucci, M.E., Stecher, D.S. and Henneberry, T.J. 2000. Heat shock proteins in whiteflies, an insect that accumulates sorbitol in response to heat stress. J. Thermal Biol. 25(5): 363-371.
- 3818.Salvucci, M.E., Wolfe, G.R. and Hendrix, D.L. 1997. Effect of sucrose concentration on carbohydrate metabolism in *Bemisia argentifolii*. Biochemical mechanism and physiological role for trehalulose synthesis in the silverleaf whitefly. J. Insect Physiol. 43(5): 457-464.
- 3819.Salvucci, M.E., Wolfe, G.R. and Hendrix, D.L. 1998. Purification and properties of an unusual NADPH-dependent ketose reductase from the silverleaf whitefly. Insect Biochem. Mol. Biol. 28(5-6): 357-363.
- 3820.Sammour, E.A., Abdalla, E.F. and Abdallah, S.A. 1993. Field evaluation of different insecticidal groups for the control of all stages of the cotton whitefly, *Bemisia tabaci*, Genn on tomato plants. Bull. Faculty Agric. (Univ. Cairo) 44(4): 931-944.
- 3821.Sampson, A.C. and King, V.J. 1996. *Macrolophus caliginosus* field establishment and pest control effect in protected tomatoes. Bull. OILB/SROP (IOBC/WPRS) 19(1): 143-146.
- 3822.Samretwanich, K., Chiemsombat, P., Kittipakorn, K. and Ikegami, M. 2000. Tomato leaf curl geminivirus associated with cucumber yellow leaf disease in Thailand. J. Phytopathol. 148(11-12): 615-617.
- 3823.Samretwanich, K., Chiemsombat, P., Kittipakorn, K. and Ikegami, M. 2000. Yellow leaf disease of cantaloupe and wax gourd from Thailand caused by tomato leaf curl virus. Plant Dis. 84(2): 200.
- 3824.Samretwanich, K., Chiemsombat, P., Kittipakorn, K. and Ikegami, M. 2000. Yellow leaf disease of muskmelon from Thailand caused by Tomato leaf curl virus. Plant Dis. 84(6): 707.
- 3825.Samretwanich, K., Sakuragaoka, Chiemsombat, P., Kittipakorn, K. and Ikegami, M. 2000. A new geminivirus associated with a yellow leaf curl disease of pepper in Thailand. Plant Dis. 84(9): 1047.
- 3826.Samuel, C.K. 1950. Parasites and parasitism of the white-fly, *Bemisia tabaci* (Gen.), vector of tobacco leaf-curl in Northern India. Indian J. Entomol. 12: 248-250. [ Cock (1986)]
- 3827.Sanap, M.M., Aher, R.P. and Deshmukh, R.B. 1995. Incidence of major insect pests associated with pigeonpea at Rahuri [India]. Indian J. Pulses Res. 8(2): 192-194.
- 3828.Sanchez, A., Geraud-Pouey, F. and Esparza, D. 1997. Bionomics of the tobacco whitefly, *Bemisia tabaci* (Homoptera: Aleyrodidae) and potential for population increase on five host plant species. Rev. Facultad Agron., Univ. Zulia (Venezuela) 14(2): 193-206. [ Spanish, English summary]
- 3829.Sanchez-Campos, S., Navas-Castillo, J., Camero, R., Soria, C., Diaz, J.A. and Moriones, E. 1999. Displacement of tomato yellow leaf curl virus (TYLCV)-Sr by TYLCV-Is in tomato epidemics in Spain. Phytopathology 89(11): 1038-1043.
- 3830.Sanchez-Campos, S., Navas-Castillo, J., Monci, F., Diaz, J.A. and Moriones, E. 2000. *Mercurialis ambigua* and *Solanum luteum*: Two newly discovered natural hosts of tomato yellow leaf curl geminiviruses. Eur. J. Plant Pathol. 106(4): 391-394.
- 3831.Sanderson, J. 1991. "Sweetpotato" whitefly vs. "poinsettia" whitefly: significance to New York State greenhouses. Long Island Hortic. News (December): 1.
- 3832.Sanderson, J. 1992. Identification of greenhouse and sweetpotato whiteflies. The Grower: Vegetable and Small Fruit Newsletter 92(12): 4-5.
- 3833.Sanderson, J. and Ferrentino, G. 1989. Whitefly biology and management in the greenhouse. Long Island Hortic. News (November): 1-4.
- 3834.Sanderson, J.P. 1987. Sweetpotato whitefly in New York greenhouses. Long Island Hortic. News (November): 1-2.
- 3835.Sanderson, J.P. 1992. Planning ahead for sweetpotato and greenhouse whiteflies. pp. 26-36. *In* Proceedings Eighth Conference on Insect and Disease Management on Ornamentals. Growers Council Society of American Florists, Alexandria, VA.
- 3836.Sanderson, J.P. and Ferrentino, G.W. 1993. Tracking whitefly populations. pp. 30-36. *In* J.A. Martens and K. Pyle (Eds.), Poinsettias: growing & marketing. Ball Publishing, Batavia, IL.
- 3837.Sandhu, T.S. 1978. Breeding for yellow mosaic virus resistance in mungbean. pp. 176-179. *In* R. Cowell (Ed.), 1st International Mungbean Symposium. Office of Information Services, Asian Vegetable Research and Development Center, Taiwan.
- 3838.Sandu, Z. and Schneider, R. 1989. [Control of *Bemisia tabaci* on poinsettia]. Hassadeh 69(6): 1047. [ Cock (1993), Hebrew]
- 3839.Saranga, Y., Lin, T. and Schwartz, A. 1997. Effect of whitefly on cotton productivity. p. 1432. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 3840.Sardana, H.R. and Verma, S. 1987. Effect of fertilizers and insecticides on the incidence of yellow mosaic disease of greengram *Vigna radiata* Wilezeck. Plant Prot. Bull. (India) 39(4): 3-6. [ Cock (1993)]
- 3841.Sarita, V. 1993. Evaluation of processing tomato (*Lycopersicon esculentum*) cultivars in the Azua Valley. Boletín FDA 6(3): 6-7. [ Spanish]
- 3842.Sartorato, A. and Seijas, C.A.R. 1985. An approach to control of bean golden mosaic virus in dry beans (*Phaseolus vulgaris* L.). Annu. Rep. Bean Improvement Coop. 28: 26-27.
- 3843.Sastry, K.S. 1989. Tomato leaf curl virus management by carbofuran plus oil combination. [Turkish summary]. J. Turkish Phytopathol. 18(1-2): 11-16. [ Cock (1993)]
- 3844.Sastry, K.S.M. 1966. Yellow vein mosaic of *Rosa bourbiana*. Indian Phytopathol. 19: 316-317. [ Cock (1986)]



- 3845.Sastry, K.S.M. and Singh, S.J. 1974. Control of the spread of the tomato leaf curl virus by controlling the whitefly (*Bemisia tabaci* Gen.) population. *Indian J. Hortic.* 31: 178-181. [Cock (1986)]
- 3846.Sastry, K.S.M., Singh, S.J. and Sastry, K.S. 1977. Effect of border cropping and the use of insecticide in relation to the incidence of tomato leaf curl virus (TLCV). *Indian J. Hortic.* 34: 319-322.
- 3847.Satoh, G.T. and Plapp, F.W., Jr. 1993. Use of juvenoid insect growth regulators for management of cotton aphid and sweetpotato whitefly populations. pp. 751-757. *In* D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
- 3848.Satpute, U.S. and Subramanian, T.R. 1983. A note on the secondary outbreak of whitefly (*Bemisia tabaci*) on cotton, with phosalone treatment. *Pestology* 7: 4.
- 3849.Sattar, A., Ullah, K., Ahad, A. and Yousaf, M. 1989. Insect pests of sunflower in N.W.F.P., Pakistan. *Pakistan J. Agric. Res.* 5(4): 239-240. [Cock (1993)]
- 3850.Satyanarayana, J., Singh, K.M. and Singh, R.N. 1995. Insect pest succession in rice bean. *Bull. Entomol. (New Delhi)* 36(1/2): 78-83.
- 3851.Saunders, K., Lucy, A. and Stanley, J. 1991. DNA forms of the geminivirus African cassava mosaic virus consistent with a rolling circle mechanism of replication. *Nucleic Acids Res.* 19(9): 2325-2330.
- 3852.Sauti, R.F.N. 1982. Malawi. pp. 104-106. *In* Root Crops in Eastern Africa. *Proceedings of Workshop Held at Kigali, Rwanda, 23-27 November 1980*. International Development Research Centre, Ottawa, Canada. [Cock (1986)]
- 3853.Sauti, R.F.N. 1984. Distribution, utilization and production constraints of cassava in Malawi. pp. 81-86. *In* A.H. Greathead, R.H. Markham, R.J. Murphy, S.T. Murphy and I.A.D. Robertson (Eds.), *Integrated Pest Management of Cassava Green Mite: Proceedings of a Regional Training Workshop in East Africa, 30 April - 4 May 1984*. Commonwealth Institute of Biological Control, Ascot, UK. [Cock (1986)]
- 3854.Sauvion, N., Pavis, C., Huc, A., Rousseau, M. and Boissot, N. 1999. Caractérisation de *Bemisia tabaci* biotype B (Hemiptera: Aleyrodidae) en Guadeloupe. [Characterization of the biotype B of *Bemisia tabaci* (Hemiptera: Aleyrodidae) in Guadeloupe, French West Indies]. *Ann. Soc. Entomol. Fr.* 35: 46-52.
- 3855.Sauvion, N., Pavis, C., Huc, A., Rousseau, M. and Boissot, N. 1999. Characterization of the B biotype of *Bemisia tabaci* Gennadius (Hemiptera : Aleyrodidae) in Guadeloupe, French West Indies. *Ann. Soc. Entomol. Fr.* 35: 46-53.
- 3856.Sauvion, N. and Rahbé, Y. 1999. Etude du comportement alimentaire des Hémiptères par la méthode EPG: revue sur le principe, la mise en oeuvre et les domaines d'application. [Recording feeding behaviour of Hemiptera with the EPG method: a review]. *Ann. Soc. Entomol. Fr.* 35: 175-183.
- 3857.Sawicki, R.M., Rowland, M.W., Byrne, F.J., Pye, B.J., Devonshire, A.L., Denholm, I., Hackett, B.S. and Stribley, M.F. 1989. The tobacco whitefly field control simulator – a bridge between laboratory assays and field evaluation. *Assoc. Appl. Biol.* 21: 121-122.
- 3858.Saxena, R.C. and Barrion, A.A. 1987. Biotypes of insect pests of agricultural crops. *Insect Sci. Appl.* 8: 453-458.
- 3859.Saxena, S., Hallan, V., Singh, B.P. and Sane, P.V. 1998. Evidence from nucleic acid hybridization tests for a geminivirus infection causing leaf curl disease of papaya in India. *Indian J. Exp. Biol.* 36(2): 229-232.
- 3860.Schaefer, P.S., Barth, S.E. and White, H.B. 1996. Predation by *Enallagma civile* (Odonata: Coenagrionidae) on adult sweetpotato whitefly, *Bemisia tabaci* (Homoptera: Aleyrodidae). *Entomol. News* 107(5): 275-276.
- 3861.Schaefer, G.A. and Terry, E.R. 1976. Insect transmission of sweet potato disease agents in Nigeria. *Phytopathology* 66: 642-645. [Cock (1986)]
- 3862.Schauff, M.E. and Evans, G.A. 1996. A pictorial guide to the species of *Encarsia* (Hymenoptera: Aphelinidae) parasitic on whiteflies (Homoptera: Aleyrodidae) in North America. *Proc. Entomol. Soc. Washington* 98: 1-35.
- 3863.Scheel, C. 1996. Status vedr. Danmark som beskyttet zone for *Bemisia tabaci* og tomatbronzetopvirus. [Status concerning Denmark as a protected zone for *Bemisia tabaci* and tomato spotted wilt virus]. pp. 259-263. *In* 13th Danish Plant Prot. Conf. Pests and Diseases. SP Rapport Statens Planteavlsvforsog. [Danish, English Summary]
- 3864.Schmalstig, J.G. and McAuslane, H.J. 2001. Developmental anatomy of zucchini leaves with squash silverleaf disorder caused by the silverleaf whitefly. *J. Am. Soc. Hortic. Sci.* 126(5): 544-554.
- 3865.Schmidt, S., Naumann, I.D. and De Barro, P.J. 2001. *Encarsia* species (Hymenoptera: Aphelinidae) of Australia and the Pacific Islands attacking *Bemisia tabaci* and *Trialeurodes vaporariorum* (Hemiptera: Aleyrodidae) - a pictorial key and descriptions of four new species. *Bull. Entomol. Res.* 91(5): 369-387.
- 3866.Schulten, G.G.M. 1997. Overview of the whitefly-virus problem: objectives of the workshop. pp. 7-10. *In* N. Ioannou (Ed.), *Management of the whitefly-virus complex*. FAO Plant Production and Protection Paper 143, Rome, Italy.
- 3867.Schultz, L.R., Jackson, J.E. and Faulkner, R.C. 1967. Studies on the sowing date of cotton in the Sudan Gezira. II. The relationship between sowing date of cotton and the incidence of insect pests. *J. Agric. Sci. (Cambridge)* 69: 317-327. [Cock (1986)]
- 3868.Schuster, D.J. 1991. Evaluation of the insect growth regulator teflubenzuron for control of the sweetpotato whitefly on fresh market tomatoes in west-Central Florida USA Fall 1989. *Insecticide Acaricide Tests* 16: 118-119.
- 3869.Schuster, D.J. 1991. Sweetpotato whitefly management on fresh market tomatoes in west-Central Florida USA Fall 1989. *Insecticide Acaricide Tests* 16: 119-120.
- 3870.Schuster, D.J. 1991. Sweetpotato whitefly management on fresh market tomatoes in west-Central Florida USA Spring 1989. *Insecticide Acaricide Tests* 16: 114-115.
- 3871.Schuster, D.J. 1992. Evaluation of insecticides for management of the sweetpotato whitefly and geminivirus on fresh market tomatoes in west-central Florida, Fall 1990. *Insecticide Acaricide Tests* 17: 166-167.
- 3872.Schuster, D.J. 1992. Evaluation of petroleum oils for management of the sweetpotato whitefly and geminivirus on fresh market tomatoes in west-central Florida, Spring 1990. *Insecticide Acaricide Tests* 17: 162.
- 3873.Schuster, D.J. 1992. Insect growth regulators and thiodan for management of the sweetpotato whitefly on fresh market tomatoes in west-central Florida, Spring 1990. *Insecticide Acaricide Tests* 17: 159-160.
- 3874.Schuster, D.J. 1992. Insecticides for management of the sweetpotato whitefly on fresh market tomatoes in west-central Florida, Spring 1990. *Insecticide Acaricide Tests* 17: 163.
- 3875.Schuster, D.J. 1992. Integration of natural enemies for management of the sweetpotato whitefly and associated disorders on mixed-cropped vegetables. *Sustain. Agric. Res. Education (SARE) or Agric. in Concert with the Environment (ACE) Res. Projects*, 29 pp.
- 3876.Schuster, D.J. 1992. Management of the sweetpotato whitefly and geminivirus on fresh market tomatoes in west-central Florida, Spring 1991. *Insecticide Acaricide Tests* 17: 160-161.
- 3877.Schuster, D.J. 1992. Monitor/pyrethroid combinations for management of the sweetpotato whitefly on fresh market tomatoes in west-central Florida, Spring 1990. *Insecticide Acaricide Tests* 17: 163-164.
- 3878.Schuster, D.J. 1992. Petroleum oils for management of the sweetpotato whitefly and geminivirus on fresh market tomatoes in west-central Florida, Fall 1990. *Insecticide Acaricide Tests* 17: 164-165.

- 3879.Schuster, D.J. 1992. Pyrethroid/organophosphate combinations for management of the sweetpotato whitefly and geminivirus on fresh market tomatoes in west-central Florida, Spring 1991. *Insecticide Acaricide Tests* 17: 167-168.
- 3880.Schuster, D.J. 1993. Management of the sweetpotato whitefly and geminivirus on fresh market tomatoes in west-central Florida fall 1991. *Insecticide Acaricide Tests* 18: 180-181.
- 3881.Schuster, D.J. 1993. Management of the sweetpotato whitefly and geminivirus on fresh market tomatoes in west-central Florida spring 1992. *Insecticide Acaricide Tests* 18: 177-179.
- 3882.Schuster, D.J. 1993. Sweetpotato whitefly and geminivirus control on fresh market tomatoes in west-Central Florida spring 1992. *Insecticide Acaricide Tests* 18: 182-183.
- 3883.Schuster, D.J. 1994. Insect control on fresh market tomatoes in west-central Florida, spring 1993. *Arthropod Management Tests* 19: 150-152.
- 3884.Schuster, D.J. 1995. Control of insects on fresh market tomatoes in west-Central Florida, spring 1994. *Arthropod Management Tests* 20: 134-136.
- 3885.Schuster, D. J. 1995. Insect control on bell pepper in west-Central Florida, spring 1994. *Arthropod Management Tests* 20: 103-105.
- 3886.Schuster, D.J. 1996. Insect management on fresh market tomatoes in West-Central Florida, Spring 1995. *Arthropod Management Tests* 21: 185.
- 3887.Schuster, D.J. 1997. Evaluation of biorational pesticides for management of the silverleaf whitefly on fresh market tomatoes, Spring 1994. *Arthropod Management Tests* 22: 179.
- 3888.Schuster, D.J. 1997. Management of insects on fresh market tomatoes, spring 1996C. *Arthropod Management Tests* 22: 184-185.
- 3889.Schuster, D.J. 1998. Intraplant distribution of immature lifestages of *Bemisia argentifolii* (Homoptera: Aleyrodidae) on tomato. *Environ. Entomol.* 27(1): 1-9.
- 3890.Schuster, D.J. 2000. Control of the silverleaf whitefly on fresh market tomatoes in fall 1997. *Arthropod Management Tests* 25: E99.
- 3891.Schuster, D.J. 2000. Insecticides applied on demand for managing the silverleaf whitefly on fresh market tomatoes in fall 1998. *Arthropod Management Tests* 25: E101.
- 3892.Schuster D. J. 2000. Insecticides applied on demand for managing the silverleaf whitefly on fresh market tomatoes in spring 1998. *Arthropod Management Tests* 25: E100.
- 3893.Schuster, D.J. 2001. Application of insecticides following imidacloprid for managing the silverleaf whitefly on fresh market tomatoes, spring 1999. *Arthropod Management Tests* 26: E90.
- 3894.Schuster, D.J. 2001. Relationship of silverleaf whitefly population density to severity of irregular ripening of tomato. *Hortscience* 36(6): 1089-1090.
- 3895.Schuster, D.J. 2001. Soil and foliar applied insecticides for managing the silverleaf whitefly on fresh market tomatoes, spring 2000. *Arthropod Management Tests* 26: E92.
- 3896.Schuster, D.J. and Arnold, C.E. 1991. Sweetpotato whitefly and leafminer management on fresh market tomatoes in southwest Florida USA Spring 1989. *Insecticide Acaricide Tests* 16: 116.
- 3897.Schuster, D.J., Evans, G.A., Bennett, F.D., Stansly, P.A., Jansson, R.K., Leibe, G.L. and Webb, S.E. 1998. A survey of parasitoids of *Bemisia* spp. whiteflies in Florida, the Caribbean, and Central and South America. *Int. J. Pest Manage.* 44(4): 255-260.
- 3898.Schuster, D.J., Everett, P.H., Price, J.F. and Kring, J.B. 1990. Suppression of the sweetpotato whitefly on commercial fresh market tomatoes. *Proc. Florida State Hortic. Soc.* 102: 374-379.
- 3899.Schuster, D.J., Funderburk, J.E. and Stansly, P.A. 1996. IPM in tomatoes. pp. 387-411. *In* D. Rosen, F.D. Bennett and J.L. Capinera (Eds.), *Pest management in the subtropics: Integrated pest management: A Florida perspective*. Intercept Ltd., Andover, UK.
- 3900.Schuster, D.J., Kring, J.B. and Price, J.F. 1991. Association of the sweetpotato whitefly with a silverleaf disorder of squash. *HortScience* 26(2): 155-156. [Cock (1993)]
- 3901.Schuster, D.J., Mueller, T.F., Kring, J.B. and Price, J.F. 1990. Relationship of the sweetpotato whitefly, *Bemisia tabaci* (Genn.) to a new fruit disorder of tomato. *HortScience* 25(12): 1618-1620. [Cock (1993)]
- 3902.Schuster, D.J. and Pitts, D.J. 1991. Tomato, *Lycopersicon esculentum* Mill., 'Sunny' sweetpotato whitefly *Bemisia tabaci* sweetpotato whitefly management on fresh market tomatoes in southwest Florida USA Fall 1988. *Insecticide Acaricide Tests* 16: 117-118.
- 3903.Schuster, D.J. and Polston, J.E. 1997. Management of insect on fresh market tomatoes spring 1996B. *Arthropod Management Tests* 22: 183-184.
- 3904.Schuster D. J. and Polston, J.E. 1997. Management of whiteflies and armyworm on fresh market tomatoes Fall 1995. *Arthropod Management Tests* 22: 180-181.
- 3905.Schuster, D.J. and Polston, J.E. 1998. Insect management on fresh market tomatoes spring 1997B. *Arthropod Management Tests* 23: 159-160.
- 3906.Schuster, D.J. and Polston, J.E. 1998. Management of whiteflies, bugs, and leafminers on fresh market tomatoes, fall 1996. *Arthropod Management Tests* 23: 156-157.
- 3907.Schuster, D.J., Polston, J.E. and Price, J.F. 1993. Reservoirs of the sweetpotato whitefly for tomatoes in West -Central Florida. *Proc. Florida State Hortic. Soc.* 105: 311-314.
- 3908.Schuster, D.J. and Price, J.F. 1991. *Poinsettia Euphorbia pulcherrima* Wild sweetpotato whitefly *Bemisia tabaci* (Genn.) insecticide induced lifestage specific mortality on the sweetpotato whitefly in the greenhouse Summer 1989. *Insecticide Acaricide Tests* 16: 257.
- 3909.Schuster, D.J. and Price, J.F. 1996. Parasitization of *Bemisia argentifolii* (Hom: Aleyrodidae) by *Encarsia pergandiella* (Hym: Aphelinidae). *Entomophaga* 41(1): 95-103.
- 3910.Schuster, D.J., Price, J.F., Kring, J.B. and Everett, P.H. 1989. Integrated management of the sweetpotato whitefly on commercial tomato. Bradenton GCREC Res. Rep. BRA1989-12. Gulf Coast Res. Educ. Center, IFAS, Univ. Florida, Bradenton, Florida, 23 pp.
- 3911.Schuster, D.J. and Stansly, P.A. 2000. Response of two lacewing species to biorational and broad-spectrum insecticides. *Phytoparasitica* 28(4): 297-304.
- 3912.Schuster, D.J., Stansly, P.A. and Polston, J.E. 1996. Expressions of plant damage by *Bemisia*. pp. 153-165. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia* : 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
- 3913.Schuster, M.F. 1964. A whitefly-transmitted mosaic virus of *Wissadula amplissima* Plant Dis. Rep. 48: 902-905.
- 3914.Schutte, F. and Bruno, G.O.A. 1976. Migracion de los insectos en el cultivo del algod6n. [Insect migration in cotton culture]. *Soc. Ingenieros Agron. (El Salvador)* 5(1): 2, 4-8, 10. [Spanish]
- 3915.Schweizer, J. 1939. Jaarverslag tabak over Juli 1938 t/m Juni 1939. Mededelingen van het Besoekisch proefstation 64: 29-50.
- 3916.Scott, J.W., Stevens, M.R., Barten, J.H.M., Thome, C.R., Polston, J.E., Schuster, D.J. and Serra, C.A. 1996. Introgression of resistance to whitefly-transmitted geminiviruses from *Lycopersicon chilense* to tomato. pp. 357-367. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia* : 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
- 3917.Seal, D.R. 1994. Control of sweetpotato whitefly in squash, 1992. *Arthropod Management Tests* 19: 144.
- 3918.Seal, D.R. 1994. Control of the sweetpotato whitefly in bush bean, 1992. *Arthropod Management Tests* 19: 53.
- 3919.Seal, D.R. 1994. Effectiveness of different insecticides for the control of sweetpotato whitefly, *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae) on vegetable crops in Southern Florida. *Proc. Florida State Hortic. Soc.* 106: 224-228.

3920. Seal, D.R. 1996. Control of silverleaf whitefly in squash, 1995. *Arthropod Management Tests* 21: 176.
3921. Seal, D.R. 1999. Control of silverleaf whitefly on cabbage, spring 1998. *Arthropod Management Tests* 24: 109-110.
3922. Seal, D.R. 1999. Control of silverleaf whitefly on spring cabbage, 1998. *Arthropod Management Tests* 21: 108-109.
3923. Seal, D.R. 2001. Control of the silverleaf whitefly and its associated bean golden mosaic virus in beans, 2000. *Arthropod Management Tests* 26: E8.
3924. Seal, D.R. 2001. Effectiveness of Neemix in managing silverleaf whitefly and tomato yellow leaf curl virus in tomatoes, 2000. *Arthropod Management Tests* 26: E93.
3925. Seal, D.R. and Baranowski, R.M. 1995. Control of silverleaf whitefly in squash, 1994. *Arthropod Management Tests* 20: 132.
3926. Seal, D.R., Bryan, J.H. and Bryan, H.D. 1995. Abundance and management of silverleaf whitefly, *Bemisia argentifolii* (Homoptera, Aleyrodidae), on ornamentals. *Proc. Florida State Hort. Soc.* 107: 222-226.
3927. Sechser, B., Bourgeois, F., Reber, B. and Wesiak, H. 1994. The integrated control of whiteflies and aphids on tomatoes in glasshouses with pymetrozine. *Med. Fac. Landbouww. Univ. Gent* 59(2b): 579-583.
3928. Secker, A.E., Bedford, I.D., Markham, P.G. and De Courcy-Williams, M.E. 1998. Squash, a reliable field indicator for the presence of the B biotype of tobacco whitefly, *Bemisia tabaci*. pp. 837-842. *In* Brighton Crop Protection Conference: Pests and Diseases. British Crop Protection Council, Farnham, UK.
3929. Sedrak, R.A. 1998. Residual effect of certain pesticides on the spider mite, *Tetranychus urticae* Koch, *Aphis gossypii* Glover and *Bemisia tabaci*, Genn under semi-field conditions. *Menofiya J. Agric. Res. (Egypt)* 23(2): 341-347.
3930. Seedco Australia Co-operative Ltd. 2000. Variety: 'Rapide'. Application no: 1997/294. *Plant Varieties J.* 13(2): 40-41.
3931. Segarra-Carmona, A.E., Bird, J. and Escudero, J. 1990. Silvering of *Cucurbita moschata* (Duchesne Poir) associated with *Bemisia tabaci* Genn. (Homoptera: Aleyrodidae) in Puerto Rico. *J. Agric. (Univ. Puerto Rico)* 74: 477-478.
3932. Seif, A.A. 1981. Seasonal fluctuations of adult population of the whitefly, *Bemisia tabaci*, on cassava. *Insect Sci. Appl.* 1: 363-364. [Cock (1986)]
3933. Seif, A.A. 1981. Transmission of cassava mosaic virus by *Bemisia tabaci*. *Plant Dis.* 65: 606-607. [Cock (1986)]
3934. Sekeroglu, E. 1999. IPM for cotton in Cukurova Region of Turkey. pp. 206-214. *In* Proceedings ICAC-CCRI Regional Consultation, Insecticide Resistance Management in Cotton. Central Cotton Research Institute, Multan, Pakistan.
3935. Sekeroglu, E. and Ozgur, A.F. 1988. *Bemisia tabaci*: population increases on cotton cultivars in Turkey. [Turkish summary]. *Türkiye Entomol. Dergisi* 12(4): 195-200. [Cock (1993)]
3936. Sela, I., Assouline, I., Tanne, E., Cohen, S. and Marco, S. 1980. Isolation and characterization of a rod-shaped, whitefly-transmissible, DNA-containing plant virus. *Phytopathology* 70: 226-228. [Cock (1986)]
3937. Sengonca, Ç. 1975. Occurrence of whitefly, *Bemisia tabaci* (Genn.), on cotton plants in Southern Anatolia (Homoptera: Aleyrodidae). *Anz. Schädlingkunde, Pflanzenschutz, Umweltschutz* 48: 140-144. [Cock (1986)]
3938. Sengonca, Ç. 1982. The principal cotton pests and their economic thresholds in the Kilikien Plain in southern Turkey. *Entomophaga* 27(Special Issue): 51-56. [Cock (1986)]
3939. Sequeira, J.C. and Harrison, B.D. 1982. Serological studies on cassava latent virus. *Ann. Appl. Biol.* 101: 33-42. [Cock (1986)]
3940. Sergeev, G.E., Velikan', V.S., Sukhoruchenko, G.I. and Evdokarova, T.G. 1996. Cotton whitefly *Bemisia tabaci* Genn. (Homoptera, Aleyrodidae): A new cotton plant pest in Turkmenistan. 2. Deleterious effects and prognosis for crop losses and cotton fiber quality. *Entomol. Obozrenie* 75(1): 16-24. [Russian]
3941. Serra, C.A. 1996. Biología de moscas blancas. pp. 11-21. *In* L. Hilje (Ed.), *Metodologías para el estudio y manejo de moscas blancas y geminivirus*. Centro Agronomico Tropical de Investigacion y Enseñanza, Turrialba, Costa Rica.
3942. Serra, C.A. 1996. Muestreo de moscas blancas. pp. 22-29. *In* L. Hilje (Ed.), *Metodologías para el estudio y manejo de moscas blancas y geminivirus*. Centro Agronomico Tropical de Investigacion y Enseñanza, Turrialba, Costa Rica.
3943. Serra, C.A. and Schmutterer, H. 1993. Control of the sweetpotato whitefly, *Bemisia tabaci* Genn. with neem extracts in tomato fields in the Dominican Republic. *Mitteilungen der Deutschen Gesellschaft für Allgemeine und Angewandte Entomol.* 8(4-6): 795-801. [German, English summary]
3944. Serrano, C.L., Sermeño Ch, J.M. and Larios, J.F. 1992. Las moscas blancas en el Salvador. [The whiteflies in El Salvador]. pp. 42-49. *In* L. Hilje and O. Arboleda (Eds.), *Las moscas blancas (Homoptera: Aleyrodidae) en America Central y el Caribe*. CATIE, Turrialba, Costa Rica. [Spanish]
3945. Servian de Cardozo, J.F. and Matsui, M. 1992. A search for effective granular insecticides against the sweetpotato whitefly, *Bemisia tabaci*. [Japanese, English summary]. *Proc. Kanto-tosan Plant Prot. Soc.* 39: 211-213.
3946. Servin, R., Martinez-Carrillo, J.L. and Hiales, L. 1999. Weeds and cultivated hosts of the silverleaf whitefly *Bemisia argentifolii* Bellows and Perring in Baja California Sur, Mexico. *Southwest. Entomol.* 24(1): 31-36.
3947. Servin-Villegas, R., Martinez, J.L., Troyo-Dieguez, E. and Ortega-Rubio, A. 1997. Susceptibilidad de adultos de *Bemisia argentifolii* (Bellows & Perring), a insecticidas de uso comun en Baja California Sur, Mexico. [Susceptibility of adults of *Bemisia argentifolii* (Bellows and Perring) to commonly used insecticides in Baja California Sur, Mexico]. *Southwest. Entomol.* 22(1): 91-101. [Spanish]
3948. Servin-Villegas, R., Troyo-Dieguez, E. and Martinez-Carrillo, J.L. 2001. Wild hosts of *Bemisia argentifolii* (Homoptera: Aleyrodidae) Bellows & Perring in semiarid northwest Mexico. *Southwest. Entomol.* 26(3): 239-244.
3949. Sese, A.I.L., Gomez-Guillamon, M.L. and Diaz-Ruiz, J.R. 1994. Appearance of a possible new melon yellowing disease in Spain. *Cucurbit Genet. Coop.* 17: 72-73.
3950. Sethi, G.R., Prasad, H. and Singh, K.M. 1978. Incidence of insect pests on different varieties of sunflower, *Helianthus annuus* Linnaeus. *Indian J. Entomol.* 40: 101-103. [Cock (1986)]
3951. Severson, R.F., Chorlyk, O.T., Stephenson, M.G., Akey, D.H., Neal, J.W., Jr., Pittarelli, G.W., Jackson, D.M. and Sisson, V.A. 1994. Characterization of natural pesticide from *Nicotiana glauca*. pp. 109-121. *In* P.A. Hedin (Ed.), *ACS Symposium Series No. 557, Bioregulators for Crop Protection and Pest Control*. American Chemical Society
3952. Sewify, G.H., Abd-Rabou, S.H., Ahmed, N.H. and Elnagar, S. 1999. Seasonal abundance of whitefly *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae) on wild plants in Giza, Egypt. *Bull. Faculty Agric. (Univ. Cairo)* 50(4): 755-766.
3953. Sewify, G.H., El-Arnaouty, S.A. and Belal, M.H. 1996. The effect of cotton late planting on population densities of sucking insects and their associated predators in Giza region, Egypt. *Bull. Faculty Agric. (Univ. Cairo)* 47(4): 665-675.
3954. Shafee, S.A. 1973. Indian species of the genus *Prospaltella* Ashmead (Hym.: Aphelinidae). *Entomophaga* 18: 251-258. [Cock (1986)]
3955. Shafee, S.A. and Rizvi, S. 1982. A new species of *Encarsia* Foerster (Hymenoptera: Aphelinidae) from Pakistan. *J. Entomol. Res. (New Delhi)* 6: 157-158. [Cock (1986)]

3956. Shafik, H.L. and Aref, A.S. 1985. [Studies on okra yellow leaf curl disease in Iraq]. Arab J. Plant Prot. 3(2): 94-97. [Cock (1993), Arabic, English summary]
3957. Shah, H., Khalid, S. and Hameed, S. 1999. Response of cotton germplasm to cotton leaf curl virus. pp. 250-256. In Proceedings ICAC-CCRI Regional Consultation, Insecticide Resistance Management in Cotton. Central Cotton Research Institute, Multan, Pakistan.
3958. Shaheen, A.H. 1976. Survey and chemical control of insects attacking *Luffa aegyptiaca*. Agric. Res. Rev. 54(1): 143-152. [Cock (1986)]
3959. Shaheen, A.H. 1977. Control of white fly *Bemisia tabaci* Genn. the main insect factor transmitting tomato leaf curl virus diseases. Moshtohor Ann. Agric. Sci. (Egypt) 6: 235-245.
3960. Shaheen, A.H. 1977. Survey of pests attacking soybean plants in Egypt with some ecological notes. Agric. Res. Rev. 55(1): 59-65. [Cock (1986)]
3961. Shaheen, A.H. 1977. Survey of pests attacking tomato in Egypt with some ecological notes. Agric. Res. Rev. 55(1): 49-57. [Cock (1986)]
3962. Shaheen, A.H. 1983. Some ecological aspects of the whitefly, *Bemisia tabaci* Genn., on tomato. Bull. Entomol. Soc. Egypt 62: 83-87. [Cock (1986)]
3963. Shaheen, A.H. 1983. Some ecological aspects on the white fly *Bemisia tabaci* Genn., the main insect vector transmitting tomato leaf curl virus diseases. Moshtohor Ann. Agric. Sci. (Egypt) 19: 453-457.
3964. Shaheen, A.H., El-Ezz, A.A. and Assem, M.A. 1973. Chemical control of cucurbit pests at Komombo. Agric. Res. Rev. 51: 103-107. [Cock (1986)]
3965. Shaheen, A.H., Samhan, M. and El-Ezz, A.A. 1973. Cucurbit pests at Komombo. Agric. Res. Rev. 51: 97-101. [Cock (1986)]
3966. Shahriary, D. and Bananej, K. 1997. Occurrence of tomato yellow leaf curl virus (TYLCV) in tomato fields of Varamin. [Iran]. Appl. Entomol. Phytopathol. 65(1): 28-29 (English), 109-110 (Persian).
3967. Shalaby, A.A., Nakhla, M.K., Shafie, M.S., Mazyad, H.M. and Maxwell, D.P. 1997. Molecular characterization of tomato yellow leaf curl geminivirus (TYLCV) isolated from pepper collected in Egypt. Moshtohor Ann. Agric. Sci. (Egypt) 35(2): 819-831.
3968. Shalaby, F.F., Abdel-Gawaad, A.A., El-Sayed, A.M. and Abo-El-Ghar, M.R. 1990. Natural role of *Eretmocerus mundus* Mercet and *Prospaltella lutea* Masi on populations of *Bemisia tabaci* Genn. Agric. Res. Rev. 68: 197-208.
3969. Shalaby, F.F., El-Khayat, E.F., El-Sayed, A.M. and Hady, S.A. 1994. Relationships between some physical and phytochemical properties of host leaves and the rate of infestation with *Bemisia tabaci* (Genn.). Moshtohor Ann. Agric. Sci. (Egypt) 32(1): 567-575.
3970. Shanab, L.M. and Awad-Allah, S.S. 1982. Studies on the whitefly (*Bemisia tabaci* Genn.) infesting tomato at Mansoura District, Egypt. Acta Phytopathologica Academiae Scientiarum Hungaricae 17(1-2): 147-155. [Cock (1986)]
3971. Shannon, P.J. 1996. Hongos entomopatógenos. pp. 60-68. In L. Hilje (Ed.), Metodologías para el estudio y manejo de moscas blancas y geminivirus. Centro Agronómico Tropical de Investigación y Enseñanza, Turrialba, Costa Rica.
3972. Shanthi, P. and Natarajan, K. 1991. Population dynamics, behavior and development of *Encarsia shafeei*, a parasitoid of *Bemisia tabaci*. pp. 55-58. In T.N. Ananthakrishnan (Ed.), Emerging Trends in Biological Control of Phytophagous Insects: National Symposium, Madras, India. Oxford and IBH Publications, New Delhi, India.
3973. Shapiro, J.P. 1996. Insect-plant interactions and expression of disorders induced by the silverleaf whitefly, *Bemisia argentifolii*. pp. 167-177. In D. Gerling and R.T. Mayer (Eds.), *Bemisia*: 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
3974. Sharaf-El-Din, A.A.A., Ismail, I.I. and Semeada, A.M. 1993. Population density of *Bemisia tabaci* (Genn.) on various crops as affected by certain weather factors at Fayoum Governorate, Egypt. Mansoura Univ. J. Agric. Sci. (Egypt) 18(8): 2427-2435.
3975. Sharaf, N. 1986. Chemical control of *Bemisia tabaci*. Agric. Ecosystems Environ. 17(1-2): 111-127. [Cock (1993)]
3976. Sharaf, N., Ahmad-Bitar, R.N. and Bulos, B.R. 1982. Parasitization of the tobacco whitefly, *Bemisia tabaci* Genn. (Hom., Aleyrodidae) on *Lantana camara* L. in the Jordan Valley. Z. Angew. Entomol. 94: 263-271.
3977. Sharaf, N., Al-Musa, A. and Nazer, I. 1983. Impact of three irrigation regimes on white fly (*Bemisia tabaci* Genn.) population and incidence of tomato yellow leaf curl on tomatoes in Jordan. Dept. Plant Prot., Jordan Univ., Amman, 11 pp.
3978. Sharaf, N., Al-Musa, A. and Nazer, I. 1984. The impact of three irrigation methods on the whitefly (*Bemisia tabaci* Genn.) population and the incidence of tomato leaf curl disease in Jordan. Dirasat 11: 109-119.
3979. Sharaf, N., Al-Musa, A.M. and Nazer, I.K. 1982. Control of tomato yellow leaf curl virus in Jordan. I. Effect of two irrigation regimes alone or in combination with pirimiphos-methyl on whitefly (*Bemisia tabaci* Genn.) population and the incidence of tomato yellow leaf curl virus. Dept. Plant Prot., Jordan Univ., Amman, 7 pp.
3980. Sharaf, N. and Batta, Y. 1984. Effect of some factors on the relationship between the whitefly *Bemisia tabaci* Genn. (Homopt., Aleyrodidae) and the parasitoid *Eretmocerus mundus* Mercet (Hymenopt., Aphelinidae). Dept. Plant Prot., Jordan Univ., Amman, 10 pp.
3981. Sharaf, N. and Batta, Y. 1985. Effect of some factors on the relationship between the whitefly *Bemisia tabaci* Genn. (Homoptera: Aleyrodidae) and the parasitoid *Eretmocerus mundus* Mercet (Hymenoptera: Aphelinidae) in Jordan. Z. Angew. Entomol. 99: 267-276. [Cock (1986)]
3982. Sharaf, N. and Batta, Y. 1996. Effect of temperature on life history of *Eretmocerus mundus* Mercet (Hymenoptera: Aphelinidae). Dirasat 23(3): 214-219.
3983. Sharaf, N. and Nazer, I.K. 1982. Effect of N P and K soil fertilizers on population trends of the tobacco whitefly (*Bemisia tabaci* Genn., Homoptera: Aleyrodidae) and the incidence of tomato leaf curl virus in tomatoes in the Jordan Valley. Dirasat 9: 13-25.
3984. Sharaf, N.S. 1981. Restriction of the spread of the tomato yellow leaf curl virus in tomato by controlling the vector, whitefly *Bemisia tabaci* Genn. (Homoptera: Aleyrodidae). Dirasat 8: 71-77.
3985. Sharaf, N.S. 1981. Studies on whiteflies on tomatoes in the Jordan Valley. II. Seasonal abundance of the immature stages of the tobacco whitefly, *Bemisia Tabaci* Genn. (Homoptera: Aleyrodidae.). Dirasat 8: 127-146.
3986. Sharaf, N.S. 1982. Determination of the proper height, direction, position, and distance of a yellow sticky trap for monitoring adult sweetpotato whitefly populations (*Bemisia tabaci* Genn., Homoptera: Aleyrodidae). Dirasat 9: 169-182.
3987. Sharaf, N.S. 1982. Factors limiting the abundance of the tobacco whitefly (*Bemisia tabaci* Genn., Homoptera: Aleyrodidae) on tomatoes during the spring season in the Jordan Valley. Dirasat 9: 97-104.
3988. Sharaf, N.S. 1982. Reduction in toxicity of certain insecticides to the aphelinid parasite, *Encarsia formosa* Gahan. I. in the host. Dirasat 9: 53-63.
3989. Sharaf, N.S. 1984. Ecology and control of the sweetpotato whitefly (*Bemisia tabaci* Genn., Homoptera: Aleyrodidae) in Jordan. Dirasat 11: 45-56. [Cock (1986)]
3990. Sharaf, N.S., Ahmad-Bitar, R.N. and Bulos, B.R. 1982. Phototactic response of the tobacco whitefly, (*Bemisia tabaci* Genn., Homoptera: Aleyrodidae). Dirasat 9(1): 135-144.
3991. Sharaf, N.S., Al-Musa, A.M. and Batta, Y. 1985. Effect of different host plants on population development of the sweet potato whitefly (*Bemisia tabaci* Genn., Homoptera: Aleyrodidae). Dirasat 12(6): 89-100.
3992. Sharaf, N.S. and Allawi, T.F. 1980. Studies on whiteflies on tomato in the Jordan Valley. I. Host range of the tobacco whitefly, *Bemisia tabaci* Genn. (Homoptera Aleyrodidae). Dirasat 7: 53-63.

- 3993.Sharaf, N.S. and Allawi, T.F. 1980. Studies on whiteflies on tomato in the Jordan Valley. III. Laboratory and field experiments on the control of whitefly (*Bemisia tabaci* Genn., Homoptera: Aleyrodidae) populations with organophosphorus insecticides and the incidence of tomato yellow leaf curl virus. Zeitschrift für pflanzenkrankheiten und Pflanzenschutz 87(3): 176-184. [ Cock (1986)]
- 3994.Sharaf, N.S. and Allawi, T.F. 1981. Control of *Bemisia tabaci* Genn., a vector of tomato yellow leaf curl virus disease in Jordan. J. Plant Dis. Prot. 87: 123-131.
- 3995.Sharaf, N.S. and Allawi, T.F. 1981. Morphological studies on three aphelinid parasites of whiteflies with observation on their host insects in Jordan. Dirasat 8: 59-70.
- 3996.Sharaf, N.S., Musa, A.M. and Nazer, I.K. 1984. Control of tomato yellow leaf curl virus in Jordan. Effect of two irrigation regimes alone or in combination with pirimiphosmethyl on whitefly (*Bemisia tabaci* Genn.) population and the incidence of tomato yellow leaf curl virus. J. Plant Dis. Prot. 91(4): 404-410. [ CATIE (1992)]
- 3997.Sharaf, N.S., Nazer, I.K. and Al-Musa, A. 1983. Control of tomato yellow leaf curl disease in Jordan; III. Effect of sticky color traps alone or in combination with pirimiphos-methyl on white fly (*Bemisia tabaci* Genn.) population and the incidence of tomato yellow leaf curl. Dirasat 10(1): 45-55.
- 3998.Sharma, A., Murayama, A., Osaki, T. and Ikegami, M. 1998. Characterization of virus specific DNA forms from tomato tissues infected by tobacco leaf curl virus: evidence for a single genomic component producing defective DNA molecules. Plant Pathol. 47(6): 787-793.
- 3999.Sharma, A.N. 1991. Effect of dust formulations on incidence of insects and grain yields of soybean (*Glycine max* (L.) Merr.) in Madhya Pradesh. J. Oilseeds Res. 8(2): 226-230.
- 4000.Sharma, A.N. 1994. Control of stem fly and sucking insects of soybean through seed treatment. J. Maharashtra Agric. Univ. [India] 19(2): 310-311.
- 4001.Sharma, D., Bagmare, A. and Gupta, A. 1997. Effect of weather parameters on population build-up of key pests of soybean. J. Insect Sci. 10(2): 120-124.
- 4002.Sharma, S.K., Mathur, S.D., Khan, R.M. and Mathur, B.N. 1971. Evaluation of some modern insecticides for the control of insect pests of cotton by means of aerial spraying and their effect on parasites and predators. Zeitschrift für Pflanzenkrankheiten und Pflanzenschutz 78: 286-295. [ Cock (1986)]
- 4003.Sharma, S.R. and Varma, A. 1982. Control of yellow mosaic of mungbean through insecticides and oil. J. Entomol. Res. (New Delhi) 6: 130-136. [ Cock (1986)]
- 4004.Sharma, S.S. and Batra, G.R. 1995. Whitefly outbreak and failure of insecticides in its control in Haryana State [India] - a note. Haryana J. Hortic. Sci. 24(2): 160-161.
- 4005.Shaw, M.J.P. 1979. Insect -borne diseases of tobacco in Rhodesia and the role of the tobacco-free period. Rhodesia Agric. J. 76(2): 87-90. [ Cock (1986)]
- 4006.Shchukin, A. and Wool, D. 1994. Pyrethroid resistance and esterase activity in selected laboratory populations of sweetpotato whiteflies *Bemisia tabaci* (Homoptera: Aleyrodidae). Eur. J. Entomol. 91(3): 285-295.
- 4007.Sheetz, R., Goolsby, J. and Poprawski, T. 1997. Antibiotic treatment of a *Nosema* sp. (Protozoa: Microsporida) infecting the ovaries of a parasitic *Encarsia* wasp (Hymenoptera: Aphelinidae). Subtrop. Plant Sci. 49: 50-52.
- 4008.Sheffield, F.M.L. 1957. Virus diseases of sweet potato in East Africa. I. Identification of the viruses and their insect vectors. Phytopathology 47: 582-590. [ Cock (1986)]
- 4009.Sheffield, F.M.L. 1957. Virus diseases of sweet potato in East Africa. II. Transmission to alternative hosts. Phytopathology 48: 1-6. [ Cock (1986)]
- 4010.Shelke, S.S., Mali, A.R. and Ajri, D.S. 1987. Effect of different schedules of insecticidal sprays on pest incidence, yield of seed cotton and quality of seed in laxmi cotton. Curr. Res. Reporter 3(2): 39-45. [ Cock (1993)]
- 4011.Shimada, T. 1994. Control of the sweetpotato whitefly, *Bemisia tabaci* (Gennadius, using vinyl films that absorb ultraviolet. Proc. Kanto-tosan Plant Prot. Soc. 41: 213-216. [ Japanese, English summary]
- 4012.Shimron, O., Hefetz, A. and Gerling, D. 1992. Arrestment responses of *Eretmocerus* species and *Encarsia deserti* (Hymenoptera, Aphelinidae) to *Bemisia tabaci* honeydew. J. Insect Behav. 5: 517-526.
- 4013.Shiohara, K., Tsuboi, S., Iwaya, K. and Moriya, K. 1994. Development of chloronicotynyl insecticide, imidacloprid. J. Pestic. Sci. 19(4): 329-332. [ Japanese, English summary]
- 4014.Shires, S., Murray, A. and Sadig, S. 1983. Stickiness in cotton: progress in solving the problem. Shell Agric. May: 1-2. [ Cock (1986)]
- 4015.Shires, S.W., Inglesfield, C. and Tipton, J.D. 1987. Effects of chlorfenvinphos on *Bemisia tabaci* Genn. and its parasites on cotton in the Sudan Gezira. Crop Prot. 6(2): 109-116.
- 4016.Shivanathan, P. 1983. The epidemiology of three diseases caused by whitefly-borne pathogens. pp. 323-330. In R.T. Plumb and J.M. Thresh (Eds.), Plant Virus Epidemiology of Three Diseases Caused by Whitefly -borne Pathogens. Blackwell Scientific Publications, Oxford, UK. [Cock (1986)]
- 4017.Shreni, V.C.D., Sivastava, K.M. and Singh, B.P. 1980. Source and spread of zinnia yellow-net disease *Bemisia tabaci*. Indian J. Hortic. 37(2): 192-194. [ CATIE (1992)]
- 4018.Shrestha, K., Shrestha, S.K., Timila, R.D. and Joshi, S. 1997. Tomato leaf curl disease: a white fly transmitted gemini virus of tomato in Nepal. Nepalese J. Agric. 18/28: 107-112.
- 4019.Siddig, S.A. 1982. Major pests of faba beans in Sudan. pp. 277-283. In Proceedings Faba Bean Conference, Cairo, Egypt, 7-11 Mar 1981. Martinus Nijhoff Publishers, Den Haag, Netherlands.
- 4020.Siddig, S.A. 1984. Entomology. pp. 248-254. In Annu. Rep. Gezira Res. Stn. Substns. 1976/1977. Agric. Res. Corp., Sudan [Cock (1986)]
- 4021.Siddig, S.A. 1987. Entomology. pp. 310-317. In Annu. Rep. Gezira Res. Stn. Substns. 1978/1979. Agric. Res. Corp., Sudan [Cock (1993)]
- 4022.Siddig, S.A. 1987. A proposed pest management program in cluding neem treatments for combating potato pests in the Sudan. pp. 449-459. In H. Schmutterer and K.R.S. Ascher (Eds.), Natural Pesticides from the Neem Tree (*Azadirachta indica* A. Juss) and Other Tropical Plants. Eschborn, German Federal Republic; Deut che Gesellschaft für Technische Zusammenarbeit. [Cock (1993)]
- 4023.Siddiqui, K.H. and Trimohan. 1999. Resistant sources amongst soybean genotypes against yellow mosaic disease transmitted by whitefly. Shashpa 6(2): 153-166.
- 4024.Siddiqui, K.H. and Trimohan. 2000. Evaluation of some insecticidal formulations against major insect pests (*Melanagromyza sojae* Zehnt. and *Bemisia tabaci* Genn.) of soybean. Shashpa 7(2): 167-170.
- 4025.Siddiqui, K.H., Trimohan, Rana, V.K.S. and Lal, S.K. 1999. Location of sources of resistance amongst soybean genotypes to yellow mosaic virus disease due to whitefly, *Bemisia tabaci* Genn. under natural conditions. Shashpa 6(1): 37-40.
- 4026.Sidhu, A.S. and Bhalla, J.S. 1975. Comparative efficacy of some new insecticides as aerial sprays for the control of cotton pests. Plant Prot. Bull. (India) 23(4; 1971): 44-47.
- 4027.Sidhu, A.S. and Dhawan, A.K. 1977. Testing of new insecticides for the control of cotton pests, *Amrasca devastans*, *Pectinophora gossypiella*, *Bemisia tabaci*. Pesticides 11(7): 16-18.
- 4028.Sidhu, A.S. and Dhawan, A.K. 1981. Evaluation of some new insecticides against foliage feeding pests of cotton. Pesticides 15(6): 20-22. [ Cock (1986)]

- 4029.Sidhu, A.S. and Dhawan, A.K. 1981. Seasonal abundance of different insect pests on desi cotton (*Gossypium arboreum* L.). J. Res. (Punjab Agric. Univ.) 17(1980): 275-281. [ Cock (1986)]
- 4030.Sidhu, A.S. and Dhawan, A.K. 1987. Testing of phosalone and phenthoate for control of sucking pests and pink bollworm on cotton. Pesticides 21(3): 37-39. [ Cock (1993)]
- 4031.Sieburth, P.J., Schroeder, W.J. and Mayer, R.T. 1998. Effects of oil and oil-surfactant combinations on silverleaf whitefly nymphs (Homoptera: Aleyrodidae) on collards. Florida Entomol. 81(3): 446-450.
- 4032.Silberschmidt, K. and Tommasi, L.R. 1956. A solanaceous host of the virus of 'infectious chlorosis' of Malvaceae. Ann. Appl. Biol. 44: 161-165. [ Cock (1986)]
- 4033.Silva-Vara, S., Rodriguez-Montessoro, R., Delgadillo -Sanchez, F. and Cardenas-Soriano, E. 1993. Etiologia y caracterizacion de una nueva enfermedad viral en calabaza (*Cucurbita* spp.) en el Estado de Sinaloa, Mexico. [Etiology and characterization of a new viral squash (*Cucurbita* spp) disease at Sinaloa State, Mexico]. Agrociencia (Mexico), Serie Proteccion Vegetal. 4(2): 231-246. [ Spanish, English summary]
- 4034.Silva-Vara, S., Rodriguez-Montessoro, R., Garzon-Tiznado, A., Delgadillo -Sanchez, F. and Cardenas-Soriano, E. 1994. Efecto de una variante del virus enrollamiento de la hoja de la calabaza (VEHC, squash leaf curl virus, SqLCV) en genotipos de cucurbitaceas. [Effect of a strain of the squash leaf curl virus (SqLCV) in cucurbit genotypes]. Rev. Mexicana Fitopatologia 12(1): 15-20. [ Spanish, English summary]
- 4035.Silvestri, F. 1927. Contribuzione alla conoscenza degli Aleurodidae (Insecta: Hemiptera) viventi su *Citrus* in extremo oriente e dei loro parassiti. Bull. Lab. Zool. General Agraria Della Facolta Agraria Portici 21: 1-60. [ Cock (1986)]
- 4036.Silvie, P., Delvare, G. and Maldes, J.M. 1989. [Arthropods associated with the cotton crop in Chad: pests, predators and parasitoids]. Coton Fibres Trop. 44(4): 275-290. [ Cock (1993), French, English summary]
- 4037.Silvie, P. and Papierok, B. 1991. Natural enemies of insect pests on cotton in Chad: preliminary data on entomopathogenic fungi. Coton Fibres Trop. 46: 304-308.
- 4038.Silvie, P. and Papierok, B. 1991. Natural enemies of insect pests on cotton in Chad: preliminary data on entomophthoralean fungi. Coton Fibres Trop. 46(4): 293-308.
- 4039.Sim, J., Valverde, R.A. and Clark, C.A. 2000. Whitefly transmission of Sweetpotato chlorotic stunt virus. Plant Dis. 84(11): 1250.
- 4040.Simmons, A.L. and Dennehy, T.J. 1996. Contrasts of three insecticide resistance monitoring methods for whitefly. pp. 140-144. In P. Dugger and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 4041.Simmons, A.L., Williams, L., III and Dennehy, T.J. 1997. Investigations of two insect growth regulators against Arizona whitefly populations. pp. 1248-1252. In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 4042.Simmons, A.M. 1994. Oviposition on vegetables by *Bemisia tabaci* (Homoptera, Aleyrodidae) - Temporal and leaf surface factors. Environ. Entomol. 23(2): 381-389.
- 4043.Simmons, A.M. 1998. Survey of the parasitoides of *Bemisia argentifolii* (Homoptera: Aleyrodidae) in coastal South Carolina using yellow sticky traps. J. Entomol. Sci. 33(1): 7-14.
- 4044.Simmons, A.M. 1999. Nymphal survival and movement of crawlers of *Bemisia argentifolii* (Homoptera: Aleyrodidae) on leaf surfaces of selected vegetables. Environ. Entomol. 28(2): 212-216.
- 4045.Simmons, A.M. and Elsey, K.D. 1995. Overwintering and cold tolerance of *Bemisia argentifolii* (Homoptera: Aleyrodidae) in coastal South Carolina. J. Entomol. Sci. 30(4): 497-506.
- 4046.Simmons, A.M. and Jackson, D.M. 1999. An ultrasonic fogging device for managing *Bemisia argentifolii* (Homoptera: Aleyrodidae) in greenhouse vegetables. J. Entomol. Sci. 34(4): 494-496.
- 4047.Simmons, A.M. and Jackson, D.M. 2000. Evaluation of foliar-applied insecticides on abundance of parasitoids of *Bemisia argentifolii* (Homoptera : Aleyrodidae) in vegetables. J. Entomol. Sci. 35(1): 1-8.
- 4048.Simmons, A.M. and McCreight, J.D. 1996. Evaluation of melon for resistance of *Bemisia argentifolii* (Homoptera: Aleyrodidae). J. Econ. Entomol. 89(6): 1663-1668.
- 4049.Simmons, A.M. and McCutcheon, G.S. 2001. Daily foraging incidence of *Encarsia pergandiella* (Hymenoptera: Aphelinidae) on cowpea. J. Entomol. Sci. 36(2): 218-221.
- 4050.Simmons, A.M., McCutcheon, G.S., Dufault, R.J., Hassell, R.L. and Rushing, J.W. 2000. *Bemisia argentifolii* (Homoptera : Aleyrodidae) attacking species of medicinal herbal plants. Ann. Entomol. Soc. Am. 93(4): 856-861.
- 4051.Simmons, G.S. 2000. Studies on dispersal of a native parasitoid *Eretmocerus eremicus* and augmentative biological control of *Bemisia tabaci* infesting cotton. [University of Arizona]. Dissertation Abst. Int. 61(03B): 1189.
- 4052.Simmons, G.S. and Minkenber, O.P.J.M. 1994. Field-cage evaluation of augmentative biological control of *Bemisia argentifolii* (Homoptera: Aleyrodidae) in Southern California cotton with the parasitoid *Eretmocerus* nr. *californicus* (Hymenoptera: Aphelinidae). Environ. Entomol. 23(6): 1552-1557.
- 4053.Simon, C., Frati, F., Beckenbaach, A., Crespi, B., Liu, H. and Flook, P. 1994. Evolution, weighing, and phylogenetic utility of mitochondrial gene sequences and a compilation of conserved polymerase chain reaction primers. Ann. Entomol. Soc. Am. 87(6): 651-701.
- 4054.Simwat, G.S. and Dhawan, A.K. 1995. Population distribution of cotton jassid (*Amrasca biguttula*) and whitefly (*Bemisia tabaci*) on upland cotton (*Gossypium hirsutum*) at different times during the day and levels of plant canopy. Indian J. Agric. Sci. 65(1): 84-86.
- 4055.Simwat, G.S. and Dhawan, A.K. 1996. Efficacy of dimethoate for controlling sucking pests of upland cotton (*Gossypium hirsutum*) under irrigation. Indian J. Agric. Sci. 66(4): 263-264.
- 4056.Simwat, G.S. and Gill, B.S. 1992. Population build-up of cotton jassid and white fly on *Gossypium arboreum* and its correlation with weather factors. J. Res. Punjab Agric. Univ. 29(1): 70-76.
- 4057.Simwat, G.S. and Kaler, D. 2001. Impact of insecticidal treatments on nymphal growth index of whitefly (*Bemisia tabaci*) infesting upland cotton (*Gossypium hirsutum*). Indian J. Agric. Sci. 71(5): 353-354.
- 4058.Singh, A. and Sirohi, A. 1997. Effect of various insecticides for the control of yellow mosaic of black gram. Plant Dis. Res. 12(1): 37-38.
- 4059.Singh, B. and Chahal, B.S. 1976. Effect of Cycocel treatment in the incidences of insect pests on the American cotton, *Gossypium hirsutum* L. Plant Prot. Bull. (India) 24(1-2 ;1972): 49-54. [ Cock (1986)]
- 4060.Singh, B.P. and Misra, A.K. 1971. Occurrence of hollyhock yellow mosaic virus in India. Indian Phytopathol. 24: 213-214. [ Cock (1986)]
- 4061.Singh, B.R. and Singh, M. 1989. Control of yellow-vein mosaic of okra by checking its vector whitefly through adjusting dates of sowing, insecticidal application and crop barrier. Indian J. Virol. 5(1-2): 61-66. [ Cock (1993)]
- 4062.Singh, D. 1995. Development of suitable sampling methods for the arthropods of four mint species. Insect Sci. Appl. 16(3-4): 333-337.
- 4063.Singh, D., Verma, N. and Naqvi, Q.A. 1999. Oxalis leaf curl-A new report from Aligarh, India. Indian J. Virol. 15(2): 119-120.
- 4064.Singh, G., Simwat, G.S. and Sidhu, A.S. 1974. Comparative efficacy of some new insecticides for the control of cotton pests. Indian J. Agric. Sci. 43(1973): 653-658. [ Cock (1986)]
- 4065.Singh, H. and Chhuneja, P.K. 1987. Performance of high-volume, low-volume and ultra-low-volume sprays for the control of *Amrasca biguttula biguttula* (Ishida) and *Bemisia tabaci* Genn. infesting cotton. Indian J. Agric. Sci. 57(5): 360-364. [ Cock (1993)]

- 4066.Singh, H. and Khangura, J.S. 1973. Efficacy of dilute and concentrated sprays for the control of cotton pests. *Cotton Growing Rev.* 50: 72-78. [Cock (1986)]
- 4067.Singh, H., Sandhu, G.S. and Mavi, G.S. 1972. Control of yellow mosaic virus in soybean, *Glycine max* (L.) Merrill by the use of granular insecticides. *Indian J. Entomol.* 33(1971): 272-278. [Cock (1986)]
- 4068.Singh, J., Arora, R. and Sidhu, A.S. 1987. Impact of cotton off-types on the incidence of different insect pests and their management. *Indian J. Ecol.* 14(2): 254-260. [Cock (1993)]
- 4069.Singh, J. and Butter, N.S. 1985. Influence of climatic factors on the build up of whitefly *Bemisia tabaci* Genn. on cotton. *Indian J. Entomol.* 47: 359-360.
- 4070.Singh, J., Dhaliwal, Z.S., Sandhu, S.S. and Sidhu, A.S. 1990. Temporal changes in the dispersion of populations of three homopterous insect pests of upland cotton. *Insect Sci. Appl.* 11: 73-77.
- 4071.Singh, J., Gill, R.S., Brar, A.P.S. and Sandhu, B.S. 1996. Evaluation of some exotic cotton hybrids against insect pests. *Crop Improvement* 23(2): 275-277.
- 4072.Singh, J., Mahal, M.S., Singh, R., Brar, D.S., Dhaliwal, Z.S. and Singh, B. 1995. A plan for sampling cotton jassid and whitefly populations on hirsutum cotton. *J. Res. Punjab Agric. Univ.* 32(1): 46-50.
- 4073.Singh, J., Singh, D., Sohi, A.S., Brar, D.S., Kapoor, S.K. and Russell, D. 1999. Management of resistant insect pests of cotton in North India - an analysis. pp. 131-142. *In Proceedings ICAC-CCRI Regional Consultation, Insecticide Resistance Management in Cotton.* Central Cotton Research Institute, Multan, Pakistan.
- 4074.Singh, J., Sohi, A.S., Brar, D.S., Denholm, I., Russell, D. and Briddon, R. 1999. Management of cotton leaf curl viral disease in India. pp. 277-284. *In Proceedings ICAC-CCRI Regional Consultation, Insecticide Resistance Management in Cotton.* Central Cotton Research Institute, Multan, Pakistan.
- 4075.Singh, J., Sohi, A.S., Dhaliwal, Z.S. and Mann, H.S. 1993. Comparative incidence of *Helicoverpa armigera* (Hub.) and other insect pests on okra and sunflower intercrops in cotton under Punjab conditions. *J. Insect Sci.* 6(1): 137-138.
- 4076.Singh, J., Sohi, A.S. and Mann, H.S. 1997. Screening of cotton germplasm against cotton leaf curl viral disease using its vector *Bemisia tabaci* (Genn.). *J. Res. (Punjab Agric. Univ.)* 34(3): 294-298.
- 4077.Singh, J., Sohi, A.S., Mann, H.S. and Kapur, S.P. 1994. Studies on whitefly, *Bemisia tabaci* (Genn.) transmitted cotton leaf curl diseases in Punjab. *J. Insect Sci.* 7(2): 194-198.
- 4078.Singh, J.P. and Gupta, G.P. 1993. Impact of various insecticides on intermittent population of jassid and whitefly infesting American cotton during different spray schedules followed for the control of bollworm complex. *J. Entomol. Res. (New Delhi)* 17(4): 297-303.
- 4079.Singh, K. 1931. A contribution towards our knowledge of the Aleyrodidae (whiteflies) of India. *Memoirs Dep. Agric. (India Entomol. Ser.)* 12: 1-98. [Cock (1986)]
- 4080.Singh, M.P., Vir, S. and Lodha, S. 1984. Insect pest and disease management in arid zone. *Indian Farming* 34(7): 71, 73, 79. [Cock (1986)]
- 4081.Singh, O.P., Singh, K.J. and Singh, P.P. 1988. [Effect of dates of sowing and varieties on the incidence of major insect pests of soybean in Madhya Pradesh]. *Bhartiya Krishi Anusandhana Patrika* 3(1): 47-52. [Cock (1993), Hindi, English summary]
- 4082.Singh, R. and Gupta, G.P. 1993. Effect of insecticidal schedules on intermittent population of jassid and whitefly and yield of cotton. *Pestic. Res. J.* 5(2): 179-185.
- 4083.Singh, R. and Kalra, V.K. 1995. Studies on the insect-pest complex associated with summer mungbean, *Vigna radiata* (L.) Wilczek and urdbean, *Vigna mungo* (L.) Hepper in Haryana. [India]. *J. Insect Sci.* 8(2): 181-184.
- 4084.Singh, R.A. and Gurha, S.N. 1994. Influence of cropping seasons on the incidence of yellow mosaic disease in mungbean genotypes. *Indian J. Pulses Res.* 7(2): 206-208.
- 4085.Singh, R.A., Gurha, S.N., Misra, D.P. and Gangal, L.L. 1981. Role of systemic insecticides in augmenting yields with reduced yellow mosaic virus incidence in mung bean. *Indian J. Plant Prot.* 8: 167-169. [Cock (1986)]
- 4086.Singh, R.N. 1969. Studies on virus diseases and their insect vectors - I.A. survey for virus diseases and their insect vectors on kitchen garden plants in Uttar Pradesh. *Labdev J. Sci. Technol.* 7: 205-206. [Cock (1986)]
- 4087.Singh, R.P., Singh, V.P., Singh, M., Gupta, G.P. and Lal, C.B. 2001. Selection of narrow leaf upland cotton (*Gossypium hirsutum*) lines with better fibre quality and tolerance to whitefly (*Bemisia tabaci*). *Indian J. Agric. Sci.* 71(2): 131-133.
- 4088.Singh, S., Sidhu, A.S. and Sidhu, H.S. 1958. Field control of cotton jassid and whitefly in the Punjab. *Indian Cotton Growing Rev.* 12: 391-405. [Cock (1986)]
- 4089.Singh, S.J. 1990. Etiology and epidemiology of whitefly-transmitted virus diseases of okra in India. *Plant Dis. Res.* 5(1): 64-70. [Cock (1993)]
- 4090.Singh, S.J. and Dutta, O.P. 1986. Enation leaf curl of okra - a new virus disease. *Indian J. Virol.* 2(1): 114-117. [Cock (1993)]
- 4091.Singh, S.J., Sastry, K.S.M. and Sastry, K.S. 1974. Effect of oil spray on the control of tomato leaf-curl virus in field. *Indian J. Agric. Sci.* 43(1973): 669-672. [Cock (1986)]
- 4092.Singh, S.P. 1999. Biological control in India. *Indian J. Plant Prot.* 27(1-2): 126-138.
- 4093.Singh, S.P., Morales, F.J., Miklas, P.N. and Terán, H. 2000. Section for bean golden mosaic resistance in intra- and interracial bean populations. *Crop Sci.* 40: 1565-1572.
- 4094.Singh, T.V.K., Singh, K.M. and Singh, R.N. 1990. Groundnut pest complex: III. Incidence of insect pests in relation to agroclimatic conditions as determined by graphical super imposition technique. *Indian J. Entomol.* 52(4): 686-692. [Cock (1993)]
- 4095.Singh, T.V.K., Singh, K.M. and Singh, R.N. 1991. Influence of intercropping II: Incidence of hemipteran and homopteran pests in groundnut. *Indian J. Entomol.* 53(2): 190-209.
- 4096.Singh, T.V.K., Venkateswarlu, O. and Sudhakar, R. 1997. Insect pests of horsegram, *Macrotylona uniflorum* (Linn) in Andhra Pradesh. *Insect Environ.* 3(2): 53-54.
- 4097.Singh, U.C., Singh, R. and Nagaich, K.N. 1998. Reaction of some promising chilli varieties against major insect pests and leaf curl disease. *Indian J. Entomol.* 60(2): 181-183.
- 4098.Singh, U.C., Singh, R. and Nagaich, K.N. 1999. Evaluation of tomato varieties against jassid (*Empoasca devastans*), whitefly (*Bemisia tabaci*) and leaf curl. *Indian J. Entomol.* 61(2): 173-176.
- 4099.Singh, U.C., Singh, R. and Sharma, U. 1998. Distribution of soybean stem fly (*Melanagromyza sojae* Z.), white fly (*Bemisia tabaci* C.) and YMV incidence in Gwalior. *Crop Res. (Hisar)* 16(1): 112-114.
- 4100.Sinha, S.N. and Chakrabarti, A.K. 1982. Studies on the control of yellow vein mosaic in okra seed crop. *Vegetable Sci.* 9(1): 64-69. [Cock (1986)]
- 4101.Sinisterra, X.H., Polston, J.E., Abouzid, A.M. and Hiebert, E. 1999. Tobacco plants transformed with a modified coat protein of tomato mottle begomovirus show resistance to virus infection. *Phytopathology* 89(8): 701-706.
- 4102.Sippell, D.W., Bindra, O.S. and Khalifa, H. 1982. A preliminary study of the relationship of leaf-lobing and leaf-hair density in cotton with whitefly (*Bemisia tabaci*) populations and proposal for further investigations. *FAO Working Paper* 9, 10 pp.
- 4103.Sippell, D.W., Bindra, O.S. and Khalifa, H. 1983. Development and Application of Integrated Pest Control in Cotton and Rotational Food Crops. Resistance to whitefly (*Bemisia tabaci*) in cotton. *FAO Working Paper* 10, 22 pp.

4104. Sippell, D.W., Bindra, O.S. and Khalifa, H. 1987. Resistance to whitefly (*Bemisia tabaci*) in cotton in the Sudan. *Crop Prot.* 6(3): 171-178.
4105. Sivaprasad, V. and Sreenivasulu, P. 1996. Characterization of two strains of cowpea mild mottle virus naturally infecting groundnut (*Arachis hypogaea* L.) in India. *J. Phytopathol.* (Berlin) 144(1): 19-23.
4106. Sivasupramaniam, S., Dennehy, T.J. and Williams, L., III. 1997. Management of pyrethroid-resistant whiteflies in Arizona cotton: selection, cross-resistance, and dynamics. pp. 306-317. *In* Cotton, A College of Agriculture Report, Series P-108. University of Arizona, Tucson.
4107. Sivasupramaniam, S., Johnson, S., Watson, T.F., Osman, A.A. and Jassim, R. 1997. A glass-vial technique for monitoring tolerance of *Bemisia argentifolii* (Homoptera: Aleyrodidae) to selected insecticides in Arizona. *J. Econ. Entomol.* 90(1): 66-74.
4108. Sivasupramaniam, S. and Watson, T.F. 2000. Selection for fenpropathrin and fenpropathrin plus acephate resistance in the silverleaf whitefly (Homoptera: Aleyrodidae). *J. Econ. Entomol.* 93(3): 949-954.
4109. Sivasupramaniam, T.J., Dennehy, T.J. and Williams, L., III. 1997. Management of pyrethroid-resistant whiteflies in Arizona cotton: selection, cross-resistance and dynamics. pp. 1252-1259. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
4110. Siviero, P. 1997. L'andamento della campagna del pomodoro da mensa. [Trends in the cultivation of table tomatoes]. *Informatore Agrario* 53(47): 61-63. [Italian]
4111. Siviero, P. and Motton, M.S. 1997. Avversita del pomodoro da mensa coltivato in pieno campo. [Adversities of field-grown table tomatoes]. *Informatore Agrario* 53(47): 51-59. [Italian]
4112. Skinner, R.H. 1996. Leaf temperature effects on *Bemisia argentifolii* (Homoptera: Aleyrodidae) oviposition. *Environ. Entomol.* 25(6): 1371-1375.
4113. Skinner, R.H. 1996. Response of *Bemisia argentifolii* (Homoptera: Aleyrodidae) to water and nutrient stressed cotton. *Environ. Entomol.* 25(2): 401-406.
4114. Skinner, R.H. and Cohen, A.C. 1994. Phosphorus nutrition and leaf age effects on sweetpotato whitefly (Homoptera: Aleyrodidae) host selection. *Environ. Entomol.* 23(3): 693-698.
4115. Sleat, D.E., Hull, R., Turner, P.C. and Wilson, M.A. 1988. Studies on the mechanism of translational enhancement by the 5'-leader sequence of tobacco mosaic virus RNA. *Eur. J. Biochem.* 175: 75-86.
4116. Smart, J.R., Bradford, J.M., Rektorik, R.J., Robinson, J. and Wolfenbarger, D.A. 1994. Tillage alternatives for cotton production in the lower Rio Grande Valley. pp. 1591-1594. *In* D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
4117. Smee, C. 1933. Report of the Entomologist. Nyasaland Dep. Agric. 1932: 48-52. [Cock (1986)]
4118. Smee, C. 1934. Report of the Entomologist. Nyasaland Dep. Agric. 1933: 46-53. [Cock (1986)]
4119. Smee, C. 1945. Notes on plant virus diseases with particular reference to tobacco. Nyasaland Agric. Quart. J. 5(4): 73-89. [Cock (1986)]
4120. Smith, C.W. 1998. Tamcot 8104: new mid season cotton cultivar for central and south Texas. pp. 607-609. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
4121. Smith, C.W. 2001. Registration of TAM 88G-104 high-yielding upland cotton germplasm. *Crop Sci.* 41(4): 1369-1370.
4122. Smith, C.W. 2001. Registration of TAM 90C-19s and TAM 90J-57s silverleaf whitefly resistant upland cotton germplasm. *Crop Sci.* 41(4): 1372-1373.
4123. Smith, H.A. 1999. Intercropping and whitefly (Homoptera: Aleyrodidae) management (Pest management). [University of Florida]. Dissertation Abst. Int. 60(12B): 5890.
4124. Smith, H.A., Evans, G.A. and McSorley, R. 2000. A survey of parasitoids of *Trialeurodes vaporariorum* and *Bemisia tabaci* (Homoptera: Aleyrodidae) in eastern Guatemala. *Florida Entomol.* 83(4): 492-496.
4125. Smith, H.A., Koenig, R.L., McAuslane, H.J. and McSorley, R. 2000. Effect of silver reflective mulch and a summer squash trap crop on densities of immature *Bemisia argentifolii* (Homoptera: Aleyrodidae) on organic bean. *J. Econ. Entomol.* 93(3): 726-731.
4126. Smith, H.A. and McSorley, R. 2000. Potential of field corn as a barrier crop and eggplant as a trap crop for management of *Bemisia argentifolii* (Homoptera: Aleyrodidae) on common bean in north Florida. *Florida Entomol.* 83(2): 145-158.
4127. Smith, H.A., McSorley, R. and Izaguirre, J.A.S. 2001. Effect of intercropping common bean with poor hosts and nonhosts on numbers of immature whiteflies (Homoptera: Aleyrodidae) in the Salama Valley, Guatemala. *Environ. Entomol.* 30(1): 89-100.
4128. Smith, I.M. 1999. Glasshouse quarantine pests for the EPPO region and measures recommended by EPPO and the EU to prevent their spread. *Bull. OEPP* 29(1-2): 23-27.
4129. Smith, I.M. 1999. Review of the status of glasshouse quarantine pests in EPPO countries. *Bull. OEPP* 29(1-2): 91-93.
4130. Smith, M.T., Lanham, D.J. and Hennessey, R.D. 1996. *Encarsia formosa* Gahan (Hymenoptera: Aphelinidae) parasitizing *Bemisia argentifolii* Bellows & Perring (Homoptera: Aleyrodidae): behavioral analysis of two geographic populations. pp. 703-706. *In* P. Dugger and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
4131. Smith, P.A. 1997. Oil and emulsion formulations of a microbial control agent [*Paecilomyces fumosoroseus*] increase the potency against a wider range of pest life stages. *Phytoparasitica* 25(SUPPL.): S93-S100.
4132. Smith, W.J., Smith, C.W. and Meagher, R.L. 1996. Abaxial surface and emulsified leaf pH of cotton, *Gossypium* spp. *Southwest. Entomol.* 21(4): 369-376.
4133. Smitley, D.R. and Davis, T.W. 1995. Whitefly control on poinsettia plants, 1994. *Arthropod Management Tests* 20: 301.
4134. Smitley, D.R. and Davis, T.W. 1999. Greenhouse whitefly control on zinnia, 1998. *Arthropod Management Tests* 24: 382-383.
4135. Snyder, J.C., Simmons, A.M. and Thacker, R.R. 1998. Attractancy and ovipositional response of adult *Bemisia argentifolii* (Homoptera: Aleyrodidae) to type IV trichome density on leaves of *Lycopersicon hirsutum* grown in three day-length regimes. *J. Entomol. Sci.* 33(3): 270-281.
4136. Soares, J.J., Ribeiro, O.R., Curvelo-Freire, E., Silveira-Carvalho, O. and Lima-Vasconcelo, O. 1997. Mosca branca *Bemisia* sp. una nova praga do algodoeiro no sudoeste Baiano. [Whitefly *Bemisia* sp. a new pest of cotton in southwestern Baiano, Brazil]. *Centro Nacional de Pesquisa do Algodao* No. 55, 7 pp. [Portuguese]
4137. Sobeiha, A.K., Zidan, Z.H., Mowafy, M.M. and Marzouk, A.S. 1997. Effect of certain insecticides regimes on some sucking insects and natural enemies inhabiting cotton plants. *Ain-Shams Univ. Ann. Agric. Sci. (Egypt)* 42(2): 665-674.
4138. Sohi, A.S., Singh, J. and Mann, H.S. 1995. Impact of plant spacing on incidence of insect pests and seed cotton yield of American cotton. *Insect Environ.* 1(2): 15-16.
4139. Soliman, Z.R., Shehata, K.K. and Gomaa, E.A. 1976. On the food range and economic importance of the predatory mite *Agistemus exsertur* Gonz. (Acari, Prostigmata). *Anz. Schadlingskunde, Pflanzenschutz, Umweltschutz* 49(6): 87-90. [Cock (1986)]
4140. Somasekhara, Y.M., Nateshan, H.M. and Miniyyappa, V. 1997. Evaluation of neem products and insecticides against whitefly (*Bemisia tabaci*, a vector of tomato leaf curl geminivirus disease). *Indian J. Plant Prot.* 25(1): 56-59.
4141. Sonalkar, V. 1999. *Bemisia tabaci* (Gennadius) control with acetamiprid on okra in Vidarbha region of Maharashtra. *Pest Manag. Econ. Zool.* 7(1): 87-89.



- 4142.Sonne, H., Aagesen, J. and Amsen, M. 1989. [The cotton white fly *Bemisia tabaci*]. Gartner Tidende 105(17): 399-401. [Cock (1993), Danish]
- 4143.Soomro, B.A., Baloch, A.A. and Kalroo, A.M. 1989. Population dynamics studies on white fly (*Bemisia tabaci*) in Sind Pakistan: 1. Population trend on cotton. Pakistan J. Agric. Res. 5(1/2): 25-34.
- 4144.Soria, C., Lopez-Sese, A.I. and Gomez-Guillamon, M.L. 1999. Resistance of *Cucumis melo* against *Bemisia tabaci* (Homoptera: Aleyrodidae). Environ. Entomol. 28(5): 831-835.
- 4145.Soria, C., Sese, A.I. and Gomez-Guillamon, M.L. 1995. Specificity of transmission of melon yellowing virus by *Trialeurodes vaporariorum* and *Bemisia tabaci*. Cucurbit Genet. Coop. 18: 44.
- 4146.Soto, J.J. and Rodriguez, R. 2000. Historia y situacion de la produccion de frijol en los paises latinoamericanos afectados por geminivirus transmitidos por mosca blanca: America Central: Guatemala. pp. 25-28. In Bean Golden Mosaic and Other Diseases of Common Bean Caused by Whitefly-Transmitted Geminiviruses in Latin America. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. [Spanish]
- 4147.Southern Rhodesia. 1933. Government Notice No. 367. Southern Rhodesia Government Gazette 1933: 353-354. [Cock (1986)]
- 4148.Soyer, D. 1939. La "rosette" de l'arachide. Recherches sur les vecteurs possibles de la maladie. pp. 21, 23. In Inst. Natl. Etude Agronomique (Congo Belge). [Cock (1986)]
- 4149.Sparks, A.N., Jr. 1993. Control of SPWF with pyrethroid organophosphate combinations 1991. Insecticide Acaricide Tests 18: 253.
- 4150.Sparks, A.N., Jr., Norman, J.W., Jr. and Riley, D.G. 1992. Management of sweetpotato whitefly in the Lower Rio Grande Valley. pp. 691-692. In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 4151.Sparks, A.N., Jr. and Riley, D.G. 1993. Evaluation of foliar applied insecticide for SPWF control 1991. Insecticide Acaricide Tests 18: 252.
- 4152.Sparks, B. 1994. Whitefly control in greenhouses and interior plantscapes. Bull. Univ. Georgia No. 1077
- 4153.Spencer, J. 1997. Integrated pest management for whitefly in cotton. pp. 5-6. In International Cotton Advisory Committee, 56th Plenary Meeting, Paraguay.
- 4154.Spencer, J., Scott, M.S., Forer, G., Niv, A., Horowitz, A.R., Navon, A., Levski, S. and Yablonski, S. 1999. Delt apine BOLLGARD variety response to the Israeli pest complex. pp. 1000-1002. In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 4155.Srinivasan, G. and Babu, P.C.S. 2001. Field evaluation of neem products against whitefly, *Bemisia tabaci* Gennadius on brinjal. Ann. Plant Prot. Sci. 9(1): 19-21.
- 4156.Srivastava, K.M., Aslam, M. and Rao, B.L.S. 1985. A whitefly transmitted yellow vein mosaic disease of *Cosmos sulphureus* Cav. Curr. Sci. (Bangalore) 54(21): 1126-1128. [Cock (1993)]
- 4157.Srivastava, K.M., Singh, B.P., Dwadash-Shreni, V.C. and Srivastava, B.N. 1977. *Zinnia* yellow net disease - transmission, host range, and agent-vector relationship. Plant Dis. Rep. 61: 550-554. [Cock (1986)]
- 4158.Srivastava, K.M. and Singh, L.N. 1976. A review of the pest complex of kharif pulses in Uttar Pradesh. Pest Abstracts and News Summaries 22: 333-335. [Cock (1986)]
- 4159.Sserubombwe, W.S., Thresh, J.M., Otim-Nape, G.W. and Osiru, D.S.O. 2001. Progress of cassava mosaic virus disease and whitefly vector populations in single and mixed stands of four cassava varieties grown under epidemic conditions in Uganda. Ann. Appl. Biol. 138(2): 161-170.
- 4160.Staetz, C.A., Boyler, K.A., Gage, E.V., Riley, D.G. and Wolfenbarger, D.A. 1992. Vial bioassay for contact insecticides for adult whiteflies, *Bemisia tabaci*. pp. 704-707. In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
- 4161.Stam, P.A., Abdelrahman, A.A. and Munir, B. 1994. Comparisons of control action thresholds for *Heliothis armigera*, *Bemisia tabaci* and *Aphis gossypii* on cotton in the Sudan Gezira and Rahad regions. Crop Prot. 13(7): 503-512.
- 4162.Stam, P.A. and Elmosa, H. 1990. The role of predators and parasites in controlling populations of *Earias insulana*, *Heliothis armigera* and *Bemisia tabaci* on cotton in the Syrian-Arab-Republic. Entomophaga 35: 315-327.
- 4163.Stanley, D. 1991. Whitefly causes bleak times for growers - producers face whole armies of tiny, disease-spreading marauders. Agric. Res. (USDA) 39(1): 16-17.
- 4164.Stanley, J. 1983. Infectivity of the cloned geminivirus genome requires sequences from both DNAs. Nature (London) 305: 643-645. [Cock (1986)]
- 4165.Stanley, J. 1985. The molecular biology of geminiviruses. Adv. Virus Res. 30: 139-177.
- 4166.Stanley, J. 1991. The molecular determinants of geminivirus pathogenesis. Seminars in Virology 2: 139-149.
- 4167.Stanley, J. and Davies, J.W. 1985. Structure and function of the DNA genome of geminiviruses. Nucleic Acids in Plants 2: 191-218.
- 4168.Stanley, J., Frischmuth, T. and Ellwood, S. 1990. Defective viral DNA ameliorates symptoms of geminivirus infection in transgenic plants. Proc. Natl. Acad. Sci. 87(16): 6291-6295.
- 4169.Stanley, J. and Gay, M.R. 1983. Nucleotide sequence of cassava latent virus DNA. Nature (London) 301: 260-262. [Cock (1986)]
- 4170.Stanley, J. and Latham, J.R. 1992. A symptom variant of beet curly top geminivirus produced by mutation of open reading frame C4. Virology 190: 506-509.
- 4171.Stanley, J., Latham, J.R., Pinner, M.S., Bedford, P.I. and Markham, P.G. 1992. Mutational analysis of the monopartite geminivirus beet curly top virus. Virology 191: 396-405.
- 4172.Stanley, J., Townsend, R. and Curson, S.J. 1985. Pseudorecombinants between cloned DNAs of two isolates of cassava latent virus. J. Gen. Virol. 66: 1055-1061.
- 4173.Stansly, P.A. 1989. Control of sweetpotato whitefly and associated disorders on staked tomato. Insecticide Acaricide Tests 16: 120-121.
- 4174.Stansly, P.A. 1991. Pest status and control of sweetpotato whitefly on cucurbits. pp. 35-48. In G. Hochmuth (Ed.), Proceedings of the Florida Watermelon Institute. IFAS, Univ. Florida, Gainesville, FL.
- 4175.Stansly, P.A. 1993. Management of the whitefly, *Bemisia tabaci*, and geminiviruses on tomato in Florida and the Dominican Republic. pp. 163-166. In CTA - Seminar Proceedings.
- 4176.Stansly, P.A. 1993. Steps toward integrated management of *Bemisia tabaci*. Proc. Congress Colombian Entomol. Soc. 20: 251-258.
- 4177.Stansly, P.A. 1995. Seasonal abundance of silverleaf whitefly in southwest Florida vegetable fields. Proc. Florida State Hortic. Soc. 108: 234-242.
- 4178.Stansly, P.A. and Avilez, M.C. 1998. Control of silverleaf whitefly on commercial eggplant, 1997. Arthropod Management Tests 23: 107-108.
- 4179.Stansly, P.A. and Cawley, B.M. 1992. Control of sweetpotato whitefly and geminivirus transmission on staked tomato. Insecticide Acaricide Tests 17: 171-172.
- 4180.Stansly, P.A. and Cawley, B.M. 1992. Control of sweetpotato whitefly and leafminers on staked tomato with insecticide, soap and oil sprays. Insecticide Acaricide Tests 17: 169-171.
- 4181.Stansly, P.A. and Cawley, B.M. 1993. Control of sweetpotato whitefly, tomato pinworm and American serpentine leafminer on staked tomato fall 1991. Insecticide Acaricide Tests 18: 184.

4182. Stansly, P.A. and Cawley, B.M. 1994. Control of adult sweetpotato whitefly (SPWF) and tomato mottle geminivirus (TMoV) transmission on staked tomato, spring, 1992. *Arthropod Management Tests* 19: 156.
4183. Stansly, P.A. and Cawley, B.M. 1994. Control of immature sweetpotato whitefly (SPWF) on staked tomato, spring, 1992. *Arthropod Management Tests* 19: 156-158.
4184. Stansly, P.A. and Cawley, B.M. 1995. Control of sweetpotato whitefly, geminivirus transmission, tomato pinworm, and leafminer on staked tomato, spring 1991. *Arthropod Management Tests* 20: 139.
4185. Stansly, P.A. and Conner, J.M. 1994. Control of immature sweetpotato whitefly (SPWF) and tomato pinworm (TPW) in staked tomato, spring, 1993. *Arthropod Management Tests* 19: 159-160.
4186. Stansly, P.A. and Conner, J.M. 1994. Suppression of adult sweetpotato whitefly (SPWF) and spread of tomato mottle geminivirus (TMoV) in staked tomato, spring, 1993. *Arthropod Management Tests* 19: 158-159.
4187. Stansly, P.A. and Conner, J.M. 1995. Control of silverleaf whitefly and tomato pinworm on staked tomato with chemical and biological insecticides, spring 1994. *Arthropod Management Tests* 20: 145-146.
4188. Stansly, P.A. and Conner, J.M. 1997. Control of silverleaf whitefly (SWF) on tomato using Sanmite applied with hydraulic and airboom sprayers, 1996. *Arthropod Management Tests* 22: 187-188.
4189. Stansly, P.A. and Conner, J.M. 1998. Control of silverleaf whitefly on staked tomato with foliar and soil-applied systemic insecticides. *Arthropod Management Tests* 23: 165-166.
4190. Stansly, P.A. and Conner, J.M. 1998. Control of silverleaf whitefly on tomato with sucrose esters, insect growth regulators, and synthetic pyrethroid/organophosphate tank mix, 1995. *Arthropod Management Tests* 23: 161-162.
4191. Stansly, P.A. and Conner, J.M. 2000. Impact of insecticides on silverleaf whitefly and tomato yellow leaf curl virus on staked tomato, 1999. *Arthropod Management Tests* 25: 104.
4192. Stansly, P.A., Conner, J.M. and Peach, D.R. 2001. Impact of insecticides on silverleaf whitefly and tomato yellow leafcurl virus (TYLCV) on staked tomato, 2000. *Arthropod Management Tests* 26: E96.
4193. Stansly, P.A., Conner, J.M. and Pomerinke, M.A. 1999. Control of silverleaf whitefly and turnip aphid on collards with foliar and soil-applied systemic insecticides, 1998. *Arthropod Management Tests* 24: 124-125.
4194. Stansly, P.A., Conner, J.M. and Pomerinke, M.A. 1999. Control of silverleaf whitefly on staked tomato with foliar insecticides, 1998. *Arthropod Management Tests* 24: 183-184.
4195. Stansly, P.A. and Liu, T.X. 1995. Activity of some biorational insecticides on silverleaf whitefly. *Proc. Florida State Hort. Soc.* 107: 167-171.
4196. Stansly, P.A. and Liu, T.X. 1997. Selectivity of insecticides to *Encarsia pergandiella* (Hymenoptera: Aphelinidae), an endoparasitoid of *Bemisia argentifolii* (Hemiptera: Aleyrodidae). *Bull. Entomol. Res.* 87(5): 525-531.
4197. Stansly, P.A., Liu, T.X., Schuster, D.J. and Dean, D.E. 1996. Role of biorational insecticides in management of *Bemisia*. pp. 605-615. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia* : 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
4198. Stansly, P.A., Liu, T.X. and Vavrina, C.S. 1998. Response of *Bemisia argentifolii* (Homoptera: Aleyrodidae) to imidacloprid under greenhouse, field, and laboratory conditions. *J. Econ. Entomol.* 91(3): 686-692.
4199. Stansly, P.A. and Schuster, D.J. 1990. Whitefly update. pp. 41-59. *In* W.M. Stall (Ed.), *Proceedings Florida Tomato Institute, Vegetable Crops Special Series SS-VEC-001*. IFAS, Univ. Florida, Gainesville, FL.
4200. Stansly, P.A. and Schuster, D.J. 1992. The sweetpotato whitefly and integrated pest management of tomato. pp. 54-73. *In* C.S. Vavrina (Ed.), *Proceedings Florida Tomato Institute, Vegetable Crops Special Series SS-HOS-001*. IFAS, Univ. Florida, Gainesville, FL.
4201. Stansly, P.A., Schuster, D.J. and Leibe, G.L. 1991. Management strategies for the sweetpotato whitefly. pp. 20-42. *In* C.S. Vavrina (Ed.), *Proceedings Florida Tomato Institute, Vegetable Crops Special Series SS-VEC-001*. IFAS, Univ. Florida, Gainesville, FL.
4202. Stansly, P.A., Schuster, D.J. and Liu, T.X. 1997. Apparent parasitism of *Bemisia argentifolii* (Homoptera: Aleyrodidae) by aphelinidae (Hymenoptera) on vegetable crops and associated weeds in south Florida. *Biol. Control* 9(1): 49-57.
4203. Stansly, P.A., Schuster, D.J. and McAuslane, H.J. 1994. Biological control of silverleaf whitefly: an evolving sustainable technology. pp. 484-491. *In* *Environmentally Sound Agriculture: Proc. Second Conf. American Society of Agricultural Engineers*, St. Joseph, Michigan.
4204. Stathopoulos, D.G. 1964. Studies on the identification and bio-ecology of *Aphis* spp., *Thrips tabaci* Lind., *Bemisia tabaci* Genn., *Empoasca* sp. and *Tetranychus telarius* L. cotton pests I. *Annu. Rep. Plant Prot. Agric. Res. Stn. (Thessaloniki)* 2: 39-47. [Cock (1986)]
4205. Stathopoulos, D.G. 1967. Studies on chemical control of *Tetranychus urticae* Koch (*T. telarius* L.) and *Bemisia tabaci* Genn. on cotton. *Annu. Rep. Plant Prot. Agric. Res. Stn. (Thessaloniki)* 3: 1965: 50-55. [Cock (1986), Greek with English summary]
4206. Stathopoulos, D.G. 1967. Studies on the identification and bio-ecology of *Aphis* spp., *Thrips tabaci* Lind., *Bemisia tabaci* Genn., *Empoasca* sp. and *Tetranychus urticae* Koch (*T. telarius* L.) cotton pests. II. *Annu. Rep. Plant Prot. Agric. Res. Stn. (Thessaloniki)* 3 :1965: 41-49. [Cock (1986), Greek with English summary]
4207. Stathopoulos, D.G., Mentzelos, I.A. and Savvidis, S.D. 1967. Survey of insects and other pests on crops in Macedonia and Thrace. II. *Annu. Rep. Plant Prot. Agric. Res. Stn. (Thessaloniki)* 3: 102-106. [Cock (1986), Greek, English summary]
4208. Steenberg, I. and Humber, R.A. 1999. Entomopathogenic potential of *Verticillium* and *Acremonium* species (Deuteromycotina: Hyphomycetes). *J. Invertebr. Pathol.* 73(3): 309-314.
4209. Stein, V.E., Coutts, R.H.A. and Buck, K.W. 1983. Serological studies on tomato golden mosaic virus, a geminivirus. *J. Gen. Virol.* 64: 2493-2498. [Cock (1986)]
4210. Steiner, M.Y. 1993. IPM practices in greenhouse poinsettia crops in Alberta, Canada. *Bull. OILB/SROP (IOBC/WPRS)* 16(8): 133-134.
4211. Stenger, D.C., Duffus, J.E. and Curson, S.J. 1990. Biological and genomic properties of a geminivirus isolated from pepper. *Phytopathology* 80(8): 704-709.
4212. Stenger, D.C., Revington, G.N., Stevenson, M.C. and Bisaro, D.M. 1991. Replicational release of geminivirus genomes from tandemly repeated copies: Evidence for rolling-circle replication of a plant viral DNA. *Proc. Natl. Acad. Sci.* 88(18): 8029-8033.
4213. Stenseth, C. 1990. [Whiteflies on ornamental plants in the greenhouse] Mellus på pryddplanter i vekstus. *Gartneryrket (Norway)* 80(3): 16-18. [Cock (1993), Norwegian]
4214. Stenseth, C. 1993. Biological control of cotton whitefly, *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae) by *Encarsia formosa* (Eulophidae: Hymenoptera) on *Euphorbia pulcherrima* and *Hypoestes phyllostachya*. *Bull. OILB/SROP (IOBC/WPRS)* 16(8): 135-140.
4215. Stenseth, S. and Singh, H.M. 1990. Buprofezin against the glasshouse whitefly and the cotton whitefly (Buprofezin mot veksthusmellus og bomullsmellus). *Gartneryrket (Norway)* 80(1): 18-19.
4216. Stevens, T.J., Kilmer, R.L. and Glenn, S.J. 2000. An economic comparison of biological and conventional control strategies for whiteflies (Homoptera : Aleyrodidae) in greenhouse poinsettias. *J. Econ. Entomol.* 93(3): 623-629.
4217. Stimmel, J.F. and Wheeler, A.G., Jr. 1988. The sweet potato whitefly in Pennsylvania. *Pennsylvania Flower Growers Bull.* 381: 1.

- 4218.Stoffella, P.J. and Powell, C.A. 1994. Sweet potato whitefly and tomato irregular ripening: a research review. Proc. Interamerican Soc. Trop. Hortic. 38: 240-243.
- 4219.Stolz, M. 1994. Side-effects of a pyridine azomethine (CGA 215'944), a new insecticide against aphids and whiteflies, on the parasitic wasp *Encarsia formosa* Gahan (Hymenoptera: Aphelinidae). [German summary]. Zeitschrift für pflanzenkrankheiten und Pflanzenschutz 101(6): 649-653.
- 4220.Stoner, A. and Butler, G.D., Jr. 1965. *Encarsia lutea* as an egg parasite of bollworm and cabbage looper in Arizona cotton. J. Econ. Entomol. 58: 1148-1150. [ Cock (1986)]
- 4221.Storey, H.H. 1932. Leafcurl of tobacco in Southern Rhodesia. Rhodesia Agric. J. 29(3): 186-192. [ Cock (1986)]
- 4222.Storey, H.H. 1932. Report of the Plant Pathologist. Rep. East African Agric. Res. Inst. (Amani) 4(1931-32): 8-13. [ Cock (1986)]
- 4223.Storey, H.H. 1935. Virus disease of East African plants: II - Leaf-curl disease of tobacco. East African Agric. Forestry J. 1: 148-153. [ Cock (1986)]
- 4224.Storey, H.H. 1938. Virus disease of plants. Rep. East African Agric. Res. Inst. (Amani) 10(1937): 9-13. [ Cock (1986)]
- 4225.Storey, H.H. 1939. Plant pathology. Virus disease of plants. Rep. East African Agric. Res. Inst. (Amani) 11(1938): 13-19. [ Cock (1986)]
- 4226.Storey, H.H. and Nichols, R.F.W. 1938. Studies of the mosaic disease of cassava. Ann. Appl. Biol. 25: 790-806. [ Cock (1986)]
- 4227.Streibert, H.P., Drabek, J. and Rindlisbacher, A. 1988. CGA 106630 - a new type of acaricide/insecticide for the control of the sucking pest complex in cotton and other crops. pp. 25-32. In Brighton Crop Protection Conference: Pests and Diseases. British Crop Protection Council, Surrey, UK.
- 4228.Suarez-Gomez, H.D. and Bolano-Amaya, R.E. 1999. Manejo de la mosca blanca, *Bemisia tabaci* G. (Homoptera: Aleyrodidae) en el cultivo del tomate en el Norte del Cesar. Corpoica Pronatta, Valledupar, Colombia, 21 pp. [ Spanish]
- 4229.Suazo, P.E. 1995. Effect of pruning and foliar fertilization on the severity of the tomato yellow mosaic virus. M. Sc. Thesis. CATIE. Turrialba, Costa Rica, 84 pp.
- 4230.Subbaratnam, G.V. and Butani, D.K. 1982. Chemical control of insect pest complex of brinjal. Entomon 7: 97-100. [ Cock (1986)]
- 4231.Sudhakar, K. and Paul, M.D. 1988. Comparative efficacy of insecticides against cotton whitefly and gram caterpillar. Pesticides 22: 29-31.
- 4232.Sudhakar, K. and Paul, M.D. 1991. Efficacy of conventional insecticides for control of cotton whitefly *Bemisia tabaci* and gram-podborer *Helicoverpa armigera* on cotton *Gossypium* sp. Indian J. Agric. Sci. 61(9): 685-687. [ Cock (1993)]
- 4233.Sudhakar, K., Punnaiah, K.C. and Krishnayya, P.V. 1998. Efficacy of certain selected insecticides on the sucking pest complex of brinjal (*Solanum melongena*). Indian J. Entomol. 60(3): 241-244.
- 4234.Sudhakar, K., Punnaiah, K.C. and Krishnayya, P.V. 1998. Influence of different fertilizers and selected insecticides on the incidence of sucking pests of brinjal. Indian J. Entomol. 60(3): 245-249.
- 4235.Sugiura, M., Bandaranayake, C.M. and Hemachandra, G.H. 1975. Chilli virus disease in Sri Lanka. Tech. Bull. Trop. Agric. Res. Center 8: 1-62. [ Cock (1986)]
- 4236.Suhail, A., Riaz, M., Rashid, M.A. and Akram, W. 1996. Effect of weather factors on sucking insect pests of NIAB-86 cotton and their chemical control. J. Anim. Plant Sci. (Pakistan) 6(1/2): 31-32.
- 4237.Suharto, S., Surjana, T., Kartaatmadja, S., Zainal, I. and Jumanto. 1987. Whitefly, *Bemisia tabaci* Genn (Aleyrodidae), as a vector of virus disease on soybean in Java. Media Penelitian Sukamandi (Indonesia) 4: 25-28. [ Indonesian, English summary]
- 4238.Sukhija, H.S., Butter, N.S., Kular, J.S. and Singh, T.H. 1989. Efficacy of triazophos (Hostathion, 40 EC) against sucking pests of upland cotton. J. Indian Soc. Cotton Improvement 14(2): 146-149.
- 4239.Sukhija, H.S., Butter, N.S. and Singh, J. 1986. Determination of the economic threshold of whitefly, *Bemisia tabaci* Genn., on American cotton in the Punjab. Trop. Pest Manage. 32(2): 134-136. 190, 194.
- 4240.Sukhija, H.S., Butter, N.S., Singh, J. and Singh, B. 1987. Assessment of losses due to important insect pests of cotton in the Punjab. Agric. Sci. Digest (India) 7(2): 115-118. [ Cock (1993)]
- 4241.Sukhoruchenko, G.I. 1996. Pesticide resistance of cotton plant pests in Central Asia and Azerbaijan: State of the problem in the early 90s. Entomol. Obozrenie 75(1): 3-15. [ Russian]
- 4242.Sukhoruchenko, G.I., Velikan, V.S. and Evdokarova, T.G. 1993. [Cotton whitefly]. Zashchita Rastenii (Moskva) 2: 26-27. [ Russian]
- 4243.Sukhoruchenko, G.I., Velikan, V.S., Evdokarova, T.G. and Mironov, V.G. 1997. Cotton whitefly *Bemisia tabaci* Genn. (Homoptera, Aleyrodidae) - a new pest of cotton plants in Turkmenia. 3. Assortment of the means of the cotton whitefly control. Entomol. Obozrenie 76(2): 278-289,492. [ Russian]
- 4244.Sukhoruchenko, G.I., Velikan', V.S., Niyazov, O.D. and Evdokarova, T.G. 1995. Cotton whitefly *Bemisia tabaci* Genn. (Homoptera, Aleyrodidae): A new cotton pest in Turkmenistan. I. Species composition and distribution of cotton whitefly in Middle Asia. Entomol. Obozrenie 74(3): 516-527. [ Russian]
- 4245.Sukhoruchenko, G.I., Velikan', V.S., Niyazov, O.D. and Evdokarova, T.G. 1996. The cotton whitefly *Bemisia tabaci* Genn. (Homoptera, Aleyrodidae), a new cotton pest in Turkmenistan. I. The species composition of whiteflies and the distribution of the cotton whitefly in Central Asia. Entomol. Rev. 75(8): 38-52.
- 4246.Sullivan, D.J. and Volkl, W. 1999. Hyperparasitism: multitrophic ecology and behavior. Ann. Rev. Entomol. 44: 291-315.
- 4247.Sultan, M.J., Sabri, M.A. and Tariq, M. 2000. Different control measures against the insect pests of bitter melon (*Momordica charantia* L.). Pakistan J. Biol. Sci. 3(6): 1054-1055.
- 4248.Sumalde, A.C. 1990. Whiteflies: emerging plant pests in the Philippines. 21st Pest Control Council of the Philippines Annual Convention, Bacolod City, 1 p.
- 4249.Summers, C.G. 1997. Phototactic behavior of *Bemisia argentifolii* (Homoptera: Aleyrodidae) crawlers. Ann. Entomol. Soc. Am. 90(3): 372-379.
- 4250.Summers, C.G., Elam, P. and Newton, A.S., Jr. 1995. Colonization of ornamental landscape plants by *Bemisia argentifolii* Bellows & Perring (Homoptera: Aleyrodidae). Pan-Pacific Entomol. 71(3): 190-198.
- 4251.Summers, C.G., Newton, A.S. and Hansen, K.R. 1995. Susceptibility of selected grape cultivars and tree fruit to silverleaf whitefly (*Bemisia argentifolii*) colonization. Hortscience 30(5): 1040-1042.
- 4252.Summers, C.G., Newton, A.S., Jr. and Estrada, D. 1996. Intraplant and interplant movement of *Bemisia argentifolii* (Homoptera: Aleyrodidae) crawlers. Environ. Entomol. 25(6): 1360-1364.
- 4253.Summy, K.R. and King, E.G. 1992. Cultural control of cotton insect pests in the United States. Crop Prot. 11: 307-319.
- 4254.Sundaram, M.K., Dhandapani, N., Swamiappan, M., Babu, P.C.S. and Jayaraj, S. 1994. A study on the management of some pests of groundnut (*Arachis hypogaea* L.) with biocontrol agents. J. Biol. Control 8(1): 1-4.
- 4255.Sundaramurthy, V.T. 1992. Upsurgence of whitefly *Bemisia tabaci* Gen. in the cotton ecosystem in India. Outlook Agric. 21(2): 109-115.
- 4256.Sundaramurthy, V.T. 1993. Whitefly upsurgence in Indian cotton ecosystems. Resist. Pest Manage. 5: 14.
- 4257.Sundaramurthy, V.T. 1998. Sugar distillery effluent activity against insect pests. Int. Sugar J. 100: 566-567.
- 4258.Sundararaju, D. and Rangarajan, A.V. 1987. Effect of insecticides in combination with fertilizers in controlling yellow mosaic disease and pod borer of green gram. Pesticides 21(12): 20-21. [ Cock (1993)]
- 4259.Sundararaju, D. and Rangarajan, A.V. 1987. Insecticidal control of pests of green gram and black gram. Pesticides 21(12): 18-19. [ Cock (1993)]

- 4260.Sunter, G. and Bisaro, D.M. 1991. Transactivation in a geminivirus: AL2 gene product is needed for coat protein expression. *Virology* 180: 416.
- 4261.Sunter, G. and Bisaro, D.M. 1992. Transactivation of geminivirus AR1 and BR1 gene expression by the viral AL2 gene product occurs at the level of transcription. *Plant Cell* 4: 1321-1331.
- 4262.Sunter, G., Gardiner, W.E. and Bisaro, D.M. 1989. Identification of tomato golden mosaic virus-specific RNAs in infected plants. *Virology* 170: 243-250.
- 4263.Sunter, G., Gardiner, W.E., Rushing, A.E., Rogers, S.G. and Bisaro, D.M. 1987. Independent encapsidation of tomato golden mosaic virus A component DNA in transgenic plants. *Plant Mol. Biol.* 8: 477-484.
- 4264.Sunter, G., Hartitz, M.D. and Bisaro, D.M. 1993. Tomato golden mosaic virus leftward gene expression: autoregulation of geminivirus replication protein. *Virology* 195: 275-280.
- 4265.Sunter, G., Hartitz, M.D., Hormudzi, S.G., Brough, C.L. and Bisaro, D.M. 1990. Genetic analysis of tomato golden mosaic virus: ORF AL2 is required for coat protein accumulation while ORF AL3 is necessary for efficient DNA replication. *Virology* 179: 69-77.
- 4266.Supriyatdi, D. 1998. The development of silverleaf whitefly/SLW (*Bemisia argentifolii* Bellows and Perring) at different temperatures and two cultivars of cotton (*Gossypium hirsutum* L.). *J. Penelitian Pertanian Terapan (Indonesia)* 2: 28-32. [ Indonesian]
- 4267.Supriyatin. 1999. The primary pests of taro in East Java (Indonesia). pp. 401-408. *In* R. Krisdiana, Trustinah, A. Taufiq and A. Winarto (Eds.), *Improvement of Component Technology to Increase Legume and Tuber Plants Productivity*, Malang, Indonesia, 23-24 Feb 1998. [Indonesian, English summary]
- 4268.Surendran, M., Shanmugam, V., Kamalakannan, A. and Srinivasan, R. 2000. Transmission studies of brinjal mosaic virus. *Int. J. Trop. Agric.* 18(2): 123-131.
- 4269.Surulivelu, T. 1991. Effect of insecticides on the occurrence and association of whitefly, aphid and parasites on cotton. *J. Biol. Control* 5(1): 4-7.
- 4270.Susa, A.M. and Aganon, T.M. 1993. Evaluation of cotton varieties/lines for resistance against whitefly (*Bemisia tabaci* Genn.) (Homoptera:Aleyrodidae). *Cotton Res. J. (Philippines)* 6(1/2): 35-49.
- 4271.Suszkiw, J. 2000. Lab diets for two pest insects. *Agric. Res. (USDA)* 48(4): 22.
- 4272.Sutabutra, T., Attathom, S. and Kositratana, W. 1990. Detection of tomato yellow leaf curl virus in whitefly with a non-radioactive probe. pp. 110-113. *In* *Agricultural Biotechnology Conference*, Bangkok, Thailand, 16-18 Aug 1990.
- 4273.Suteri, B.D. 1974. Occurrence of soybean yellow mosaic virus in Uttar Pradesh. *Curr. Sci. (Bangalore)* 43: 689-690. [ Cock (1986)]
- 4274.Suteri, B.D. 1975. Reduction in oil content of yellow mosaic infected soybean seeds. *Curr. Sci. (Bangalore)* 44: 287. [ Cock (1986)]
- 4275.Suwanpornskul, R., Anusonratchada, B. and Kadkao, S. 1994. Seasonality of insect pests on sunflower. pp. 333-339. *In* *Research Report 1992: Corn Sunflower Groundnut and Local Plants*. Chiang Mai Field Crops Research Center, Chiang Mai, Thailand. [Thai, English summary]
- 4276.Suwwan, M.A., Akkawi, M., Al-Musa, A.M. and Mansour, A. 1988. Tomato performance and incidence of tomato yellow leaf curl (TYLCV) virus as affected by type of mulch. *Sci. Hortic.* 37: 39-45.
- 4277.Swanson, M.M., Brown, J.K., Poulos, B.T. and Harrison, B.D. 1992. Genome affinities and epitope profiles of whitefly -transmitted geminiviruses from the Americas. *Ann. Appl. Biol.* 121: 285-296.
- 4278.Swanson, M.M. and Harrison, B.D. 1993. Serological relationships and epitope profiles of isolates of okra leaf curl geminivirus from Africa and the Middle East. *Biochimie (Paris)* 75(8): 707-711.
- 4279.Swanson, M.M. and Harrison, B.D. 1994. Properties, relationships and distribution of cassava mosaic geminiviruses. *Trop. Sci.* 34(1): 15-25.
- 4280.Swanson, M.M., Valand, G.B., Muniyappa, V. and Harrison, B.D. 1998. Serological detection and antigenic variation of two whitefly - transmitted geminiviruses: tobacco leaf curl and croton yellow vein mosaic viruses. *Ann. Appl. Biol.* 132(3): 427-435.
- 4281.Swanson, M.M., Varma, A., Muniyappa, V. and Harrison, B.D. 1992. Comparative epitope profiles of the particle proteins of whitefly - transmitted geminiviruses from nine crop legumes in India. *Ann. Appl. Biol.* 120(3): 425-433.
- 4282.Swearingen, M., Headrick, D. and Bellows, T. 1997. Comparison of fixation and drying procedures for scanning electron microscopy among insect body types. *Proc. Entomol. Soc. Washington* 99(3): 513-522.
- 4283.Swirski, E., Amital, S. and Dorzia, N. 1967. Laboratory studies on the feeding development and reproduction of the predaceous mite *Amblyseius rubini* Swirski and Amital and *Amblyseius swirski* Athias (Acarina: Phytoseiidae) on various kinds of food substances. *Israel J. Agric. Res.* 17: 101-119. [ Cock (1986)]
- 4284.Swirski, E., Amital, S. and Dorzia, N. 1970. Laboratory studies on the feeding habits, post -embryonic survival and oviposition on the predaceous mites *Amblyseius chilensis* Dosse and *Amblyseius hibisci* Chant (Acarina: Phytoseiidae) on various kinds of food substances. *Entomophaga* 15: 93-106. [ Cock (1986)]
- 4285.Swirski, E. and Dorzia, N. 1968. Studies on the feeding, development and oviposition of the predaceous mite *Amblyseius limonicus* Garman and McGregor (Acarina: Phytoseiidae) on various kinds of food substances. *Israel J. Agric. Res.* 18: 71-75. [ Cock (1986)]
- 4286.Swirski, E. and Dorzia, N. 1969. Laboratory studies on the feeding, development and fecundity of the predaceous mite *Typhlodromus occidentalis* Nesbitt (Acarina: Phytoseiidae) on various kinds of food substances. *Israel J. Agric. Res.* 19(3): 143-145. [ Cock (1986)]
- 4287.Symondson, W.O.C., Gasull, T. and Liddell, J.E. 1999. Rapid identification of adult whiteflies in plant consignments using monoclonal antibodies. *Ann. Appl. Biol.* 134(3): 271-276.
- 4288.Szabo, P. and Hatalane-Zseller, I. 1991. [Appearance of *Bemisia tabaci* (Gennadius) in Hungary]. *Novenyvedelem (Hungary)* 27(6): 262-264. [ Hungarian, English summary]
- 4289.Szabo, P., van Lenteren, J.C. and Huisman, P.W.T. 1993. Development time, survival and fecundity of *Encarsia formosa* on *Bemisia tabaci* and *Trialeurodes vaporariorum*. *Bull. OILB/SROP (IOBC/WPRS)* 16: 173-176.
- 4290.Szabo, P., van Lenteren, J.C. and Huisman, P.W.T. 1995. The development, lifespan, fecundity and mortality of the whitefly parasitoid *Encarsia formosa* in the hosts *Bemisia tabaci* and *Trialeurodes vaporariorum*. *Novenyvedelem (Hungary)* 31(2): 67-70. [ Hungarian, English summary]
- 4291.Tag, E.E. 1989. [Effectiveness of pesticides on cotton in the Sudan (Gezira region)]. *Agrokhimiya* 12: 103-105. [ Cock (1993), Russian]
- 4292.Taha, H.A., Soliman, S.M., Abdel-Haleem, A. and El-Raies, S.A. 2001. Field studies on the main pests infesting cotton with refer to their natural enemies. pp. 885-888. *In* P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
- 4293.Tahir, M., Hussain, T. and Mahmood, T. 1997. Cotton leaf crumple disease: A review. *J. Agric. Res. (Pakistan)* 35(5): 337-341.
- 4294.Takahashi, R. 1931. Some white-flies of Formosa. *Trans. Nat. Hist. Soc. Formosa* 21(115): 203-209. [ Cock (1986)]
- 4295.Takahashi, R. 1933. Aleyrodidae of Formosa, Part II. Observations on the coccidae of formosa, Part III. *Rep. Gov. Res. Inst., Dept Agric. (Formosa)* 60: 1-24. [ Cock (1986)]
- 4296.Takahashi, R. 1936. Some Aleyrodidae, Aphididae, Coccidae (Homoptera), and Thysanoptera from Micronesia. *Tenthredo* 1(2): 109-120. [ Cock (1986)]
- 4297.Takahashi, R. 1940. Some Aleyrodidae from Mauritius (Homoptera). *Insecta matsumurana* 14(1): 1-5. [ Cock (1986)]
- 4298.Takahashi, R. 1940. Some foreign Aleyrodidae (Hemiptera) I. *Trans. Nat. Hist. Soc. Formosa* 30: 43-47. [ Cock (1986)]

- 4299.Takahashi, R. 1941. Some foreign Aleyrodidae (Hemiptera) III. Species from Hong Kong and Mauritius. Trans. Nat. Hist. Soc. Formosa 31: 351-357. [Cock (1986)]
- 4300.Takahashi, R. 1955. *Bemisia* and *Acanthobemisia* of Japan (Aleyrodidae, Homoptera). Kontyu 23(1): 1-5. [Cock (1986)]
- 4301.Takahashi, R. 1956. Insects of Micronesia: Homoptera: Aleyrodidae. Insects Micronesia 6: 1-13. [Cock (1986)]
- 4302.Takahashi, R. 1957. Some Aleyrodidae from Japan (Homoptera). Insecta Matsumurana 21: 12-21. [Cock (1986)]
- 4303.Takahashi, R. and Mamet, R. 1952. Some species of Aleyrodidae from Madagascar (Homoptera) II. Mem. Instit. Sci. Madagascar (E) 1: 111-113.
- 4304.Takeuchi, J., Horie, H., Takeda, N., Watanabe, K. and Habu, N. 1994. Occurrence of discoloration of garland chrysanthemum, *Chrysanthemum coronarium* caused by sweetpotato whitefly, *Bemisia tabaci* (Gennadius), in Tokyo. Proc. Kanto-Tosan Plant Prot. Soc. 41: 295-297. [Japanese, English summary]
- 4305.Talebi, A., Kamali, K., Esmaili, M., Khalghani, J. and Sahragard, A. 2000. Comparison of the length of the developmental period, adult longevity and fecundity of parasitoid wasps, *Encarsia lutea* and *Eretmocerus mundus* (Hym.: Aphelinidae), on *Bemisia tabaci* (Hom.: Aleyrodidae) nymphs. Appl. Entomol. Phytopathol. 67(1/2): 1-15. [Farsi]
- 4306.Tan, H.N.P. and Wong, S.M. 1993. Some properties of Singapore ageratum yellow vein virus (SAYVV). J. Phytopathol. 139: 165-176.
- 4307.Tan, P.H.N., Wong, S.M., Wu, M., Bedford, I.D., Saunders, K. and Stanley, J. 1995. Genome organization of ageratum yellow vein virus, a monopartite whitefly-transmitted geminivirus isolated from a common weed. J. Gen. Virol 76(12): 2915-2922.
- 4308.Tan, W.J. and Riley, D.G. 2000. Effects on reproduction of silverleaf whitefly, *Bemisia argentifolii* due to bifenthrin resistance. Resistant Pest Manage. 11(1): 19-21.
- 4309.Tan, W.J., Riley, D.G. and Wolfenbarger, D.A. 1996. Quantification and genetic analysis of bifenthrin resistance in the silverleaf whitefly. Southwest. Entomol. 21(3): 265-275.
- 4310.Tapia, E.A. 1968. El girasol, nuevo hospedador para un homoptero conocido. Hoja Informativa. Inst. Patologia Vegetal (Argentina) 25: 2. [Cock (1986)]
- 4311.Tappertzhofen, S. 1995. On the effect of neem products on *Bemisia tabaci* using different adjuvants. Anz. Schadlingskunde, Pflanzenschutz, Umweltschutz 68(6): 137-139. [German]
- 4312.Tappertzhofen, S. 1996. Zur Populationsentwicklung von *Bemisia argentifolii* im Sudwesten der Dominikanischen Republik. [Population dynamics of *Bemisia argentifolii* in the south west of the Dominican Republic. Anz. Schadlingskunde, Pflanzenschutz, Umweltschutz 69(7): 153-156. [German]
- 4313.Tarczynski, M.C., Byrne, D.N. and Miller, W.B. 1992. High performance liquid chromatography analysis of carbohydrates of cotton-phloem sap and of honeydew produced by *Bemisia tabaci* feeding on cotton. Plant Physiol. 98: 753-756.
- 4314.Tariq, M., Sabri, M.A. and Sultan, M.J. 2000. Comparative efficacy of insecticides, insect growth and M.Y. strategy against insect pests of okra (*Abelmoschus esculentus*). Pakistan J. Biol. Sci. 3(7): 1166-1167.
- 4315.Tarr, S.A.J. 1951. Leafcurl disease of cotton. Commonwealth Mycol. Inst., 55 pp. [Cock (1986)]
- 4316.Tawfik, M.F.S., Awadallah, K.T., Hafez, M. and Sarhan, A.A. 1978. Biology of the aphelinid parasite *Eretmocerus mundus* Mercet. Bull. Entomol. Soc. Egypt 62: 33-48. [Cock (1986)]
- 4317.Tayeb, E.H.A., Hassan, N.A. and Kotob, F.K. 1999. Efficacy of some biorational insecticides to certain sucking insects attacking eggplant, *Solanum melongena* L. Minufiya J. Agric. Res. (Egypt) 24(2): 471-475.
- 4318.Taylor, D.E. 1981. White-flies. Zimbabwe Agric. J. 78(1): 25. [Cock (1986)]
- 4319.Taylor, R.A.J., Shalhevet, S., Spharim, I., Berlinger, M.J. and Lebiush-Mordechi, S. 2001. Economic evaluation of insect-proof screens for preventing tomato yellow leaf curl virus of tomatoes in Israel. Crop Prot. 20(7): 561-569.
- 4320.Teich, Y. 1966. Mites of the family Phytoseiidae as predators of the tobacco whitefly, *Bemisia tabaci* Gennadius. Israel J. Agric. Res. 16: 141-142. [Cock (1986)]
- 4321.Teitel, M. and Shklyar, M. 1998. Pressure drop across insect-proof screens. Trans. Am. Soc. Agric. Eng. 41(6): 1829-1834.
- 4322.Tengkanon, W., Roechan, M., Kartosuwondo, U. and Sakti, B. 1986. [Critical period of orba variety of soybean susceptible to viruses infected through *Bemisia tabaci* Genn]. pp. 89-96. In Seminar on Food Crops Research Results, Volume 1: Non Rice Food Crops, Sukamandi, Indonesia, 16-18 Jan 1986. Pusat Penelitian Dan Pengembangan Tanaman Pangan, Bogor, Indonesia. [Indonesian]
- 4323.Terry, E.R. and Hahn, S.K. 1982. Increasing and stabilizing cassava and sweet-potato productivity by disease resistance and crop hygiene. pp. 47-52. In Root Crops in Eastern Africa. Proceedings of a Workshop Held at Kigali, Rwanda, 23-27 November 1980. International Development Research Centre, Ottawa, Canada. [Cock (1986)]
- 4324.Teuber, L.R., Rupert, M.E., Gibbs, L.K. and Taggard, K.L. 1997. Breeding resistant alfalfa holds promise for silverleaf whitefly management. California Agric. 51(3): 25-29.
- 4325.Thakur, B.S., Dhanorkar, B.K. and Puri, S.N. 1991. Bioefficacy of some insecticides whitefly against infesting cotton. J. Maharashtra Agric. Univ. (India) 16(3): 432-433.
- 4326.Thanki, K.V. and Patel, J.R. 1991. Field evaluation of some brinjal varieties for resistance to insect pests and diseases in north Gujarat conditions. Gujarat Agric. Univ. Res. J. 16(2): 94-97.
- 4327.Thapliyal, P.N., Dubey, K.S. and Bhadula, H.K. 1987. Control of yellow mosaic of soybean with granular insecticides. Indian Phytopathol. 40(1): 110-111. [Cock (1993)]
- 4328.Thewys, G., Herve, J.J. and Larroque, M. 1979. Bilan de deux annees d' experimentation de la decamethrine sur cotonnier en milieu paysan au Senegal. pp. 115-126. In Congrès sur la lutte contre les insectes en milieu tropical. Chambre de Commerce et d'Industrie de Marseille, 13-16 Mars 1979. Compte rendu des travaux. Chambre de Commerce et d'Industrie, Marseilles, France. [Cock (1986)]
- 4329.Thomas, J.C., Adams, D.G., Keppenne, V.D., Wasmann, C.C., Brown, J.K., Kanost, M.R. and Bohnert, H.J. 1995. *Manduca sexta* encoded protease inhibitors expressed in *Nicotiana tabacum* provide protection against insects. Plant Physiol. Biochem. 33(5): 611-614.
- 4330.Thomas, J.C., Adams, D.G., Keppenne, V.D., Wasmann, C.C., Brown, J.K., Kanost, M.R. and Bohnert, H.J. 1995. Protease inhibitors of *Manduca sexta* expressed in transgenic cotton. Plant Cell Reports 14: 758-762.
- 4331.Thomas, J.C., Adams, D.G., Nessler, C.L., Brown, J.K. and Bohnert, H.J. 1995. Tryptophan decarboxylase, tryptamine, and reproduction of the whitefly. Plant Physiol. 109(2): 717-720.
- 4332.Thomas, J.C. and Bohnert, H.J. 1993. Expression of insecticidal protease inhibitors in Arizona cotton. pp. 264-267. In Cotton, A College of Agriculture Report, Series P-94. University of Arizona, Tucson.
- 4333.Thomas, K.M. and Krishnaswami, C.S. 1939. Leaf crinkle - a transmissible disease of papaya. Curr. Sci. (Bangalore) 8: 316. [Cock (1986)]
- 4334.Thomas, R. 1932. Periodic failure of the Punjab-American cotton crop. Agric. Live-stock (India) 2(3): 243-274. [Cock (1986)]
- 4335.Thome, C.R., Schuster, D.J. and Polston, J.E. 1996. Nocturnal feeding of silverleaf whitefly adults (Homoptera: Aleyrodidae). J. Agric. Entomol. 13(3): 275-278.
- 4336.Thömmes, P.A. and Buck, K.W. 1994. Synthesis of the tomato golden mosaic virus AL1, AL2, AL3, and AL4 proteins *in vitro*. J. Gen. Virol. 75: 1827-1834.

- 4337.Thompson, W.M.O. 2000. Development, morphometrics and other biological characteristics of the whitefly *Bemisia tabaci* (Gennadius) on cassava. *Insect Sci. Appl.* 20(4): 251-258.
- 4338.Thompson, W.R. 1944. A catalogue of the parasites and predators of insect pests. Section 1 Parasite host catalogue. Part 3 Parasites of the Hemiptera. 2nd Edition. pp. 22,132-133,143-144. *In* Commonwealth Agricultural Bureaux, Farnham Royal, UK.
- 4339.Thongmearkom, P., Honda, Y., Iwaki, M. and Deema, N. 1984. Ultrastructure of soybean leaf cells infected with cowpea mild mottle virus. *Phytopathol. Z.* 109: 74-79. [Cock (1986)]
- 4340.Thongmearkom, P., Honda, Y., Saito, Y. and Syamananda, R. 1981. Nuclear ultrastructural changes and aggregates of viruslike particles in mungbean cells affected by mungbean yellow mosaic disease. *Phytopathology* 71: 41-44. [Cock (1986)]
- 4341.Thottappilly, G. and Rossel, H.W. 1997. Identification and characterization of viruses infecting bambara groundnut (*Vigna subterranea*) in Nigeria. *Int. J. Pest Manage.* 43(3): 177-185.
- 4342.Thresh, J.M., Fargette, D. and Otim-nape, G.W. 1994. Effects of African cassava mosaic geminivirus on the yield of cassava. *Trop. Sci.* 34(1): 26-42.
- 4343.Thresh, J.M., Fishpool, L.D.C., Otim-nape, G.W. and Fargette, D. 1994. African cassava mosaic virus disease: an under-estimated and unsolved problem. *Trop. Sci.* 34(1): 3-14.
- 4344.Thung, T.H. 1932. De krul- en kroepoek-ziekten van tabak en de oorzaken van hare verbreiding. [The curl and crinkle diseases of the tobacco and the causes of their dissemination]. *Mededelingen, Proefstation voor vorstenlandsche Tabak* 72: 1-54. [Cock (1986), German, English summary]
- 4345.Thung, T.H. 1934. Bestrijding der krul- en kroepoek-ziekten van tabak. [The control of the curl and crinkle diseases of tobacco]. *Mededelingen, Proefstation voor vorstenlandsche Tabak* 78: 3-18. [Cock (1986), German, English summary]
- 4346.Tian, T., Klaassen, V.A., Soong, J., Wisler, G., Duffus, J.E. and Falk, B.W. 1996. Generation of cDNAs specific to lettuce infectious yellows closterovirus and other whitefly-transmitted viruses by RT-PCR and degenerate oligonucleotide primers corresponding to the closterovirus gene encoding the heat shock protein 70 homolog. *Phytopathology* 86(11): 1167-1173.
- 4347.Tian, T.Y., Rubio, L., Yeh, H.H., Crawford, B. and Falk, B.W. 1999. Lettuce infectious yellows virus: in vitro acquisition analysis using partially purified virions and the whitefly *Bemisia tabaci*. *J. Gen. Virol.* 80(5): 1111-1117.
- 4348.Tigano-Milani, M.S., Honeycutt, R.J., Lacey, L.A., Assis, R., McClelland, M. and Sobral, B.W.S. 1995. Genetic variability of *Paezilomyces fumosoroseus* isolates revealed by molecular markers. *J. Invertebr. Pathol.* 65(3): 274-282.
- 4349.Timofeeva, T.V. 1963. *Encarsia* - a parasite of the greenhouse whitefly. *Zaschita Rastenii of Vreditelei* 8(1): 44. [Cock (1986), Russian]
- 4350.Tochihara, H. 1980. New plant virus diseases observed in Japan in the last 10 years. *Rev. Plant Prot. Res.* 13: 122-132. [Cock (1986)]
- 4351.Tonhasca, A. and Byrne, D.N. 1994. The effects of crop diversification on herbivorous insects: a meta-analysis approach. *Ecol. Entomol.* 19: 239-244.
- 4352.Tonhasca, A., Palumbo, J.C. and Byrne, D.N. 1994. Aggregation patterns of *Bemisia tabaci* (Homoptera: Aleyrodidae) in response to insecticide applications. *Entomol. Exp. Appl.* 72(3): 265-272.
- 4353.Tonhasca, A., Palumbo, J.C. and Byrne, D.N. 1994. Binomial sampling plans for *Bemisia tabaci* populations in cantaloupes. *Res. Popul. Ecol.* 36(2): 181-186.
- 4354.Tonhasca, A., Palumbo, J.C. and Byrne, D.N. 1994. Distribution patterns of *Bemisia tabaci* (Homoptera: Aleyrodidae) in cantaloupe fields in Arizona. *Environ. Entomol.* 23(4): 949-954.
- 4355.Tonhasca, A., Jr., Palumbo, J.C. and Byrne, D.N. 1996. Evaluation of the power law and patchiness regressions with regression diagnostics. *J. Econ. Entomol.* 89(6): 1477-1484.
- 4356.Torres-Pacheco, I., Garzon-Tiznado, J.A., Brown, J.K., Becerra-Flora, A. and Rivera-Bustamante, R.F. 1996. Detection and distribution of geminiviruses in Mexico and the southern United States. *Phytopathology* 86(11): 1186-1192.
- 4357.Torres-Pacheco, I., Garzon-Tiznado, J.A., Herrera-Estrella, L. and Rivera-Bustamante, R.F. 1993. Complete nucleotide sequence of pepper huasteco virus: analysis and comparison with bipartite geminiviruses. *J. Gen. Virol.* 74: 2225-2231.
- 4358.Toscano, N., Henneberry T. and Castle, S. 1994. Population dynamics and pest status of silverleaf whitefly in the USA. *Arab J. Plant Prot.* 12(2): 137-142.
- 4359.Toscano, N.C., Blua, M., Ballmer, G. and Madore, M. 1992. The impact of sweetpotato whitefly, *Bemisia tabaci*, upon cotton quantity and quality in California. pp. 684-686. *In* D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
- 4360.Toscano, N.C., Castle, S.J., Henneberry, T.J. and Castle, N.P. 1998. Persistent silverleaf whitefly exploits desert crop systems. *California Agric.* 52(2): 29-33.
- 4361.Toscano, N.C. and Henneberry, T.J. 1997. Strategies to extend the effectiveness of chemicals needed for whitefly control. pp. 129-136. *In* N. Ioannou (Ed.), *Management of the whitefly-virus complex*. FAO Plant Production and Protection Paper 143, Rome, Italy.
- 4362.Toscano, N.C. and Henneberry, T.J. 1997. Whitefly management on agricultural crops. pp. 125-128. *In* N. Ioannou (Ed.), *Management of the whitefly-virus complex*. FAO Plant Production and Protection Paper 143, Rome, Italy.
- 4363.Toscano, N.C., Henneberry, T.J., Perring, T.M. and Giorgio, C. 1994. Identification, management and status of the silverleaf whitefly, *Bemisia argentifolii* in California. p. 152. *In* D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
- 4364.Toscano, N.C., Immaraju, J.A. and Georghiou, G.P. 1985. Resistance studies on the sweetpotato whitefly, *Bemisia tabaci* Genn. (Homoptera: Aleyrodidae) in the Imperial Valley, California. pp. 178-180. *In* J.M. Brown and T.C. Nelson (Eds.), *Proceedings Beltwide Cotton Production Research Conferences*. National Cotton Council, Memphis, TN.
- 4365.Toscano, N. C., Prabhaker, N., Castle, S. and Henneberry, T. 1999. Managing whiteflies (*Bemisia tabaci*), strain B, in an agricultural system. pp. 41-57. *In* I.M.C. Gomez (Ed.), *Current trends in epidemiology and virus control in horticultural crops*. Fundacion para la investigacion Agraria en la Provincia de Almeria, Aguadulce, Spain. [Spanish and English]
- 4366.Toscano, N.C., Prabhaker, N., Castle, S.J. and Henneberry, T.J. 2001. Inter-regional differences in baseline toxicity of *Bemisia argentifolii* (Homoptera: Aleyrodidae) to the two insect growth regulators, buprofezin and pyriproxyfen. *J. Econ. Entomol.* 94(6): 1538-1546.
- 4367.Toscano, N.C., Prabhaker, N., Zhou, S. and Ballmer, G. 1998. Toxicity of Applaud and Knack against silverleaf whiteflies from Southern California: implications for susceptibility monitoring. pp. 1093-1095. *In* P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
- 4368.Toscano, N.C., Trumble, J., Ting, I.P. and McCalley, N.F. 1984. Insect pest management of lettuce. pp. 69-89. *In* Iceberg Lettuce Research Program Annual Report (USA).
- 4369.Toscano, N.C., Yoshida, H.A. and Henneberry, T.J. 1997. Responses to azadirachtin and pyrethrum by two species of *Bemisia* (Homoptera: Aleyrodidae). *J. Econ. Entomol.* 90(2): 583-589.
- 4370.Tothill, J.D. 1948. *Agriculture in the Sudan*. Oxford University Press, London, UK, 974 pp. [Cock (1986), *Bemisia tabaci*; pp. 324-340, 549-555]
- 4371.Townsend, M.L., Oetting, R.D. and Braman, S.K. 1999. Whitefly and lace bug performance on *Lantana* spp. cultivars in the greenhouse. *J. Environ. Hortic.* 17(3): 99-102.

4372. Townsend, R., Stanley, J., Curson, S.J. and Short, M.N. 1985. Major polyadenylated transcripts of cassava latent virus and location of the gene encoding coat protein. *EMBO J.* 4(1): 33-37.
4373. Townsend, R., Watts, J. and Stanley, J. 1986. Synthesis of viral DNA forms in *Nicotiana plumbaginifolia* protoplasts inoculated with cassava latent virus (CLV): evidence for the independent regulation of one component of the CLV genome. *Nucleic Acids Res.* 14(3): 1253-1266.
4374. Traboulsi, R. 1994. *Bemisia tabaci*: a report on its pest status with particular reference to the Near East. *FAO Plant Prot. Bull.* 42(1/2): 33-58.
4375. Trehan, K.N. 1944. Distribution of white-fly in the Punjab. *Indian Farming* 5: 514-515. [Cock (1986)]
4376. Trehan, K.N. 1944. Further notes on the bionomics of *Bemisia gossypiperda* M. & L., the white-fly of cotton in the Punjab. *Indian J. Agric. Sci.* 14: 53-63. [Cock (1986)]
4377. Tremblay, L. 1959. Osservazione sulla simbiosi endocellulare di alcuni Aleyrodidae (*Bemisia tabaci* Gennad. *Aleurobolus olivinus* Silv. *Trialeurodes vaporariorum* West.). *Bull. Lab. Entomol. Agraria "Filippo Silvestri" Portici* 17: 210-246. [Cock (1986)]
4378. Trench, T.N., Martin, M.M. and Hemmes, E.A. 1985. An assessment of cassava African mosaic disease in South Africa and Swaziland. *South African J. Plant Soil* 2: 169-170. [Cock (1993)]
4379. Tripathy, M.K., Senapati, B. and Mishra, L.K. 1995. Effect of application of insecticidal schedules on the incidence of white flies (*Bemisia tabaci*) and jassids (*Amrasca biguttula biguttula*) and fruit yield in brinjal. *J. Appl. Biol.* 5(1/2): 91-92.
4380. Trisusilowati, E.B., Suseno, R., Sosromarsono, S., Barizi, Soedarmadi and Nur, M.A. 1990. Transmission, serological aspects and morphology of the tobacco krupuk virus. *Indonesian J. Trop. Agric.* 1(2): 75-79.
4381. Trottin-Caudal, Y., Millot, P. and Cheyrias, J.M. 1995. Methodes de lutte contre les aleurodes sur tomate sous abri. [Control methods against whiteflies in greenhouse tomatoes]. *Infos CTIFL (France)* 111: 24-28. [French, English summary]
4382. Tsai, J.H. and Wang, K.H. 1996. Development and reproduction of *Bemisia argentifolii* (Homoptera: Aleyrodidae) on five host plants. *Environ. Entomol.* 25(4): 810-816.
4383. Tsai, M.C., Liu, C.S. and Su, H.J. 1997. Poinsettia leaf curl, a new disease caused by a geminivirus. *J. Phytopathol. (Berlin)* 145(8-9): 347-350.
4384. Tsao, P.W. 1963. Intranuclear inclusion bodies in the leaves of cotton plants infected with leaf crumple virus. *Phytopathology* 53: 243-244.
4385. Tsering, K. and Patel, B.N. 1990. Simultaneous transmission of tobacco leaf curl virus and yellow-vein mosaic virus of *Abelmoschus esculentus* (L.) Moench by *Bemisia tabaci*. *Tobacco Res.* 16(2): 127-128. [Cock (1993)]
4386. Tsering, K. and Patel, B.N. 1991. Persistence of tobacco leaf curl virus (TLCV) in its vector, *Bemisia tabaci* Gennadius. *Tobacco Res.* 17(2): 89-92.
4387. Tsueda, H. and Tsuchida, K. 1998. Differences in spatial distribution and life history parameters of two sympatric whiteflies, the greenhouse whitefly (*Trialeurodes vaporariorum* Westwood) and the silverleaf whitefly (*Bemisia argentifolii* Bellows & Perring), under greenhouse and laboratory conditions. *Appl. Entomol. Zool.* 33(3): 379-383.
4388. Tsyplenkov, A.E. 1991. Viruses and whitefly. *Zashchita Rastenii (Moskva)* 8: 48-49. [Russian]
4389. Tu, Z., Byrne, D.N. and Hagedorn, H.H. 1997. Vitellin of the sweet potato whitefly, *Bemisia tabaci*: Biochemical characterization and titer changes in the adult. *Arch. Insect Biochem. Physiol.* 34(2): 223-237.
4390. Tufail, M., Rasheed, A. and Bari, S. 1996. The comparative efficacy of some latest spray schedules against the whitefly, *Bemisia tabaci* (Genn.) on FH-682, cotton. *Pakistan Entomol.* 18(1/2): 67-69.
4391. Tulmann-Neto, A. 1979. Obtenção de resistência ou tolerância ao vírus do mosaico dourado do feijão (Phaseolus vulgaris L.) a través de indução e mutação. Tese de Livre Docência Apresentada a ESALQ, Piracicaba, S.P. Brasil [Portuguese]
4392. Tunç, A., Turhan, N., Belli, A.H., Kismir, A., Tekin, T. and Kisakürek, N. 1983. Çukurova bölgesinde beyaz sinek (*Bemisia tabaci* Genn.) in kisi geçirme durumu ve konukçularinin tesbiti üzerinde araştırmalar. *Türkiye Bitki Koruma Bülteni* 23(1): 42-51. [Cock (1986)]
4393. Tunio, A.G., Munshi, G.H., Rizvi, S.N.H., Khuuro, R.D. and Khan, M.M. 1986. Safflower: the oil crop. *Pakistan Agric.* 8(11/12): 26-31.
4394. Turenne, H. 1981. Essayo de control químico de la mosca blanca (*Bemisia tabaci*) en frijol común (*P. vulgaris*). [A trial chemical test for the control of the whitefly (*Bemisia tabaci*) for the common bean (*P. vulgaris*)]. *Reunion Anual del PCCMCA* 271: 8. [CATIE (1992), Spanish]
4395. Turhan, N., Tunç, A., Belli, A., Kismir, A. and Kisakürek, N. 1983. Çukurova'da soya (*Glycine max* L.)'da böcek ve akar faunasinin tesbiti üzerinde çalışmalar. *Türkiye Bitki Koruma Bülteni* 23(3): 148-169. [Cock (1986)]
4396. Tzeng, C.C. and Kao, S.S. 1995. Toxicity of insecticides to *Eretmocerus orientalis* and *Encarsia transvena* -parasitoids of silver leaf white fly (*Bemisia argentifolii*). *Plant Prot. Bull. (Taichung)* 37(3): 271-279. [Chinese, English summary]
4397. Tzeng, C.C. and Kao, S.S. 1999. Toxicity of nine insecticides to *Eretmocerus orientalis* adults - a parasitoid of silverleaf whitefly (*Bemisia argentifolii*). [Chinese, English summary]. *Plant Prot. Bull. (Taipei)* 41(1): 83-86.
4398. Ucko, O., Cohen, S. and Ben-Joseph, R. 1998. Prevention of virus epidemics by a crop-free period in the Arava region of Israel. *Phytoparasitica* 26(4): 313-321.
4399. Ujvary, I., Matolcsy, G., Belai, I., Szurdoki, F., Bauer, K., Varjas, L. and Kramer, K.J. 1996. Projuvencoids: Synthesis and biological evaluation of sulfenylated, sulfinylated and sulfonylated carbamates. *Arch. Insect Biochem. Physiol.* 32(3-4): 659-669.
4400. Uk, S. 1978. Seasonal trend of the whitefly adult population in the CIBA-GEIGY Sudan Project Area. *Agric. Aviation Res. Unit, Progress Rep.* 77/78: 1-10.
4401. Uk, S. 1986. Insecticides and fungicides department. *Rep. Rothamsted Exp. Stn. (Harpenden, UK)* 1986: 94-104. [Cock (1993)]
4402. Uk, S. and Dittrich, V. 1986. The behaviour-modifying effect of chlordimeform and endosulfan on the adult whitefly *Bemisia tabaci* (Genn.) which attacks cotton in the Sudan. *Crop Prot.* 5(5): 341-347. [Cock (1993)]
4403. Ulgenturk, S. and Ulusoy, M.R. 1999. The whitefly species (Homoptera: Aleyrodidae) in Ankara. *Türkiye Entomol. Dergisi* 23(4): 259-268. [Turkish, English summary]
4404. Ullah, K., Imran, M., Yousaf, M., Sherin, M. and Shah, F. 1981. Relative abundance of white fly, *Bemisia tabaci* (Gen) on soybean cultivars and its chemical control. *J. Sci. Technol. (Pakistan)* 5(1/2): 112-114.
4405. Ulubilir, A. and Yabas, C. 1995. Population fluctuations, natural enemies and chemical control possibilities of cotton white fly (*Bemisia tabaci* Genn.) on vegetables in Cukurova. *Bitki Koruma Bulteni* 35(3/4): 191-210. [Turkish]
4406. Ulubilir, A., Yabas, C. and Zeren, O. 1996. Investigations on the chemical control of the whitefly (*Bemisia tabaci* Genn.) on vegetables in the Cukurova region of Turkey. [Turkish, English summary]. *Zirai Mucadele Arastirma Yilligi* 28-29: 29-30.
4407. Ulubilir, A., Yigit, A., Yucel, S. and Yabas, C. 1997. Biological control of *Bemisia tabaci* Genn. (Homoptera: Aleyrodidae) by *Deraeocoris pallens* Reut. (Hemiptera: Miridae) on eggplant in plastic houses in Adana. *Adv. Hortic. Sci.* 11(4): 202-204.
4408. Umaharan, P., Padidam, M., Phelps, R.H., Beachy, R.N. and Fauquet, C.M. 1998. Distribution and diversity of geminiviruses in Trinidad and Tobago. *Phytopathology* 88(12): 1262-1268.

4409. Unruh, B.L. and Silvertooth, J.C. 1997. Planting and irrigation termination timing effects on the yield of Upland and Pima cotton. *J. Prod. Agric.* 10(1): 74-79.
4410. Unseld, S., Ringel, M., Hofer, P., Hohnle, M., Jeske, H., Bedford, I.D., Markham, P.G. and Frischmuth, T. 2000. Host range and symptom variation of pseudorecombinant virus produced by two distinct bipartite geminiviruses. *Arch. Virol.* 145(7): 1449-1454.
4411. USDA. 1998. Management of the silverleaf whitefly; a success story in progress. U.S. Dept. Agric., 3 pp.
4412. Usin, C., Guirao, P., Cifuentes, D., Esteban, J. and Beitia, F. 1997. Inducción diferencial de plateado en variedades de calabacín, por diversas poblaciones de *Bemisia tabaci* (Gennadius) (Homoptera, Aleyrodidae). [Differential induction of leafsilvering in squash cultivars by several populations of *Bemisia tabaci* (Gennadius) (Homoptera, Aleyrodidae)]. *Boletín Sanidad Vegetal Plagas (Argentina)* 23(4): 551-556. [Spanish, English summary]
4413. Uthamasamy, S., Sivasubramanian, P. and Thangaraju, D. 1990. Monitoring of whitefly *Bemisia tabaci* on upland cotton *Gossypium hirsutum*. *Indian J. Agric. Sci.* 60(11): 744-746. [Cock (1993)]
4414. Uygun, N. and Özgür, F. 1980. İçel ve Adana illeri sera sebze zararlılarının saptanması, endosülfan Rooktablet ve primicarb' in *Myzus persicae* (Sulz.)'Ye etkiləri. [With German summary]. *Türkiye Bitki Koruma Dergisi* 4(3): 185-192. [Cock (1986)]
4415. Uygun, N., Ulusoy, M.R., Karaca, Y. and Kersting, U. 1997. Approaches to biological control of *Dialeurodes citri* (Ashmead) in Turkey. *Bull. OILB/SROP (IOBC/WPRS)* 20(7): 52-62.
4416. Uzcategui, R.C. and Lastra, R. 1978. Transmission and physical properties of the causal agent of mosaico amarillo del tomate (tomato yellow mosaic). *Phytopathology* 68: 985-988. [Cock (1986)]
4417. Vacante, V. 1995. New problems and acquisitions on the spreading of thrips and whiteflies. *Informatore Fitopatol.* 45(10): 49-55. [Italian, English summary]
4418. Vacante, V. 1997. Influnza di imidacloprid sull'impollinazione del pomodoro con *Bombus terrestris*. [The effect of imidacloprid on the pollination of tomato by *Bombus terrestris*]. *Informatore Agrario* 53(43): 68-70. [Italian]
4419. Vacante, V., Tropea-Garzia, G. and Onillon, J.C. 1994. Premières observations sur la dynamique des populations de *Trialeurodes vaporariorum* (Westwood) et de *Bemisia tabaci* (Gennadius) (Homopt. Aleyrodidae) en serre d'aubergine. [First observations on population dynamics of *Trialeurodes vaporariorum* (Westwood) and of *Bemisia tabaci* (Gennadius) (Homopt. Aleyrodidae) in greenhouse eggplant crops]. *Bull. OILB/SROP (IOBC/WPRS)* 17: 81-88. [French]
4420. Vacante, V., Tropea Garzia, G., Onillon, J.C. and Pucci, C. 1995. Observations on population dynamics of *Bemisia tabaci* (Gennadius) in an unheated protected pepper crop. *Med. Fac. Landbouww. Univ. Gent* 60(3A): 635-643. [French]
4421. Vadodaria, M.P. and Vyas, H.N. 1987. Control of whitefly, *Bemisia tabaci* (Gennadius) and its impact on yellow mosaic virus (YMV) in green gram *Vigna radiata* (L.) Wilczek and the grain yield. *Indian J. Agric. Res.* 21(1): 21-26. [Cock (1993)]
4422. Vaishampayan, S.M. and Kogan, M. 1980. Sampling whiteflies on soybean. pp. 305-311. *In* M. Kogan and D.C. Herzog (Eds.), *Sampling Methods in Soybean Entomology*. Springer-Verlag, New York.
4423. Vaissayre, M., Menozzi, P., Nibouche, S. and Deguine, J.P. 1998. Aleyrodidae in cotton farming systems: Biology and population management. *Agric. Devel.* 20: 4-12. [French, English summary]
4424. Vakili, N., Bird, J., Sanchez, J. and Woodbury, R. 1973. Wild hosts of whitefly-transmitted viruses in tropical America. pp. 42-44. *In* Reunion Annual Programa Coopertivo Centro Americano para el Mejoramiento de cultivos Alimenticios.
4425. Valand, G.B. and Desai, M.V. 1980. Studies on transmission of different types of tobacco leaf curl virus. *Gujarat Agric. Univ. Res. J.* 5(2): 20-24. [Cock (1986)]
4426. Valand, G.B. and Muniyappa, V. 1992. Epidemiology of tobacco leaf curl virus in India. *Ann. Appl. Biol.* 120: 257-267. [Cock (1993)]
4427. Valdez, J.A. and Wolfenbarger, D.A. 1995. Yellow traps and insecticides for control of a strain of sweet potato whitefly and associated virus incidence on pepper. *J. Entomol. Sci.* 30(2): 342-348.
4428. Vallejos, J.E. 1997. An expert system to evaluate the impact of the *Bemisia tabaci* - geminivirus complex on bean, tomato and sweet pepper for planning purposes. M. Sc. Thesis. CATIE. Turrialba, Costa Rica, 120 pp.
4429. Valverde, R.A., Lotrakul, P., Landry, A.D. and Boudreaux, J.E. 2001. First report of Tomato yellow leaf curl virus in Louisiana. *Plant Dis.* 85(2): 230.
4430. van Dam, N.M. and Hare, J.D. 1998. Biological activity of *Datura wrightii* glandular trichome exudate against *Manduca sexta* larvae. *J. Chem. Ecol.* 24(9): 1529-1549.
4431. van Dam, N.M. and Hare, J.D. 1998. Differences in distribution and performance of two sap-sucking herbivores on glandular and non-glandular *Datura wrightii*. *Ecol. Entomol.* 23(1): 22-32.
4432. van de Ven, W.T.G., LeVesque, C.S., Perring, T.M. and Walling, L.L. 2000. Local and systemic changes in squash gene expression in response to silverleaf whitefly feeding. *Plant Cell* 12(8): 1409-1423.
4433. van der Kamp, R.J. and van Lenteren, J.C. 1981. The parasite-host relationship between *Encarsia formosa* Gahan (Hymenoptera: Aphelinidae) and *Trialeurodes vaporariorum* (Westwood) (Homoptera: Aleyrodidae) XI. Do mechanical barriers of the host plant prevent successful penetration of the phloem by whitefly larvae and adults? *Z. Angew. Entomol.* 92: 149-159.
4434. van der Laan, P.A. 1940. Motschildluis en *Eupatorium* als oorzaken van pseudo-mozaiek. *Vlugschriften van het Deli-Proef Station te Medan* 67: 1-4.
4435. van der Laan, P.A. 1961. A quick field method of evaluating insecticides against whitefly. *Empire Cotton Growing Rev.* 38(3): 189-191. [Cock (1986)]
4436. van der Laan, P.A. 1961. Stimulating effect of DDT treatment of cotton on white flies (*Bemisia tabaci* Genn.; Aleyrodidae) in the Sudan Gezira. *Entomol. Exp. Appl.* 4: 47-53. [Cock (1986)]
4437. van der Laan, P.A. 1961. Toxaphene and Delnav as insecticides on cotton in the Sudan Gezira. *Empire Cotton Growing Rev.* 38(2): 111-118. [Cock (1986)]
4438. van Dorst, H.J.M., Huijberts, N. and Bos, L. 1983. Yellowings of glasshouse vegetables, transmitted by *Trialeurodes vaporariorum*. *Netherlands J. Plant Pathol.* 89: 171-184. [Cock (1986)]
4439. van Driesche, R.G., Hoddle, M.S., Lyon, S. and Sanderson, J.P. 1999. Use of insect growth regulators to reduce rates of *Eretmocerus eremicus* needed for biological control for whiteflies on poinsettia. *Bull. OILB/SROP (IOBC/WPRS)* 22(1): 61-64.
4440. van Driesche, R.G., Hoddle, M.S., Lyon, S. and Sanderson, J.P. 2001. Compatibility of insect growth regulators with *Eretmocerus eremicus* (Hymenoptera: Aphelinidae) for whitefly (Homoptera: Aleyrodidae) control on poinsettias II. Trials in commercial poinsettia crops. *Biol. Control* 20(2): 132-146.
4441. van Driesche, R.G., Hoddle, M.S., Roy, S., Lyon, S. and Sanderson, J.P. 2001. Effect of parasitoid release pattern on whitefly (Homoptera: Aleyrodidae) control in commercial poinsettia. *Florida Entomol.* 84(1): 63-69.
4442. van Driesche, R.G., Lyon, S.M., Hoddle, M.S., Roy, S. and Sanderson, J.P. 1999. Assessment of cost and performance of *Eretmocerus eremicus* (Hymenoptera: Aphelinidae) for whitefly (Homoptera: Aleyrodidae) control in commercial poinsettia crops. *Florida Entomol.* 82(4): 570-594.
4443. van Giessen, W.A., Mollema, C. and Elsey, K.D. 1995. Design and use of a simulation model to evaluate germplasm for antibiotic resistance to the greenhouse whitefly (*Trialeurodes vaporariorum*) and the sweetpotato whitefly (*Bemisia tabaci*). *Entomol. Exp. Appl.* 76(3): 271-286.



4444. van Iersel, M.W., Oetting, R.D. and Hall, D.B. 2000. Imidacloprid applications by subirrigation for control of silverleaf whitefly (Homoptera: Aleyrodidae) on poinsettia. *J. Econ. Entomol.* 93(3): 813-819.
4445. van Iersel, M.W., Oetting, R.D., Hall, D.B. and Kang, J.G. 2001. Application technique and irrigation method affect imidacloprid control of silverleaf whiteflies (Homoptera: Aleyrodidae) on poinsettias. *J. Econ. Entomol.* 94(3): 666-672.
4446. van Lenteren, J.C. 1999. Fundamental knowledge about insect reproduction: essential to develop sustainable pest management. *Invert. Reprod. Devel.* 36(1-3): 1-15.
4447. van Lenteren, J.C. 2000. A greenhouse without pesticides: fact or fantasy? *Crop Prot.* 19(6): 375-384.
4448. van Lenteren, J.C. and Brasch, K. 1994. Variation in acceptance and parasitization of *Bemisia tabaci* by *Encarsia formosa*. *Bull. OILB/SROP (IOBC/WPRS)* 17: 96-103.
4449. van Lenteren, J.C., Drost, Y.C., van Roermund, H.J.W. and Postuma-Doodeman, C.J.A.M. 1997. Aphelinid parasitoids as sustainable biological control agents in greenhouses. *J. Appl. Entomol.* 121(9-10): 473-485.
4450. van Lenteren, J.C. and Martin, N.A. 1999. Biological control of whiteflies. pp. 202-216. *In* R. Albajes, M.L. Gullino, J.C. van Lenteren and Y. Elad (Eds.), *Integrated Pest and Disease Management in Greenhouse Crops*. Kluwer, Dordrecht, The Netherlands.
4451. van Lenteren, J.C., Meekes, E. and Qiu, Y.T. 1999. Management of whiteflies: new natural enemies and host-plant resistance. *Bull. OILB/SROP (IOBC/WPRS)* 22(1): 145-148.
4452. van Lenteren, J.C. and Noldus, L.P.J. 1990. Whitefly-plant relationships behavioral and ecological aspects. pp. 47-90. *In* D. Gerling (Ed.), *Whiteflies: their Bionomics, Pest Status, and Management*. Intercept, Andover, UK. [Cock (1993)]
4453. van Looy, L., Sneijders, B. and Stepman, W. 1999. Plant protection in greenhouses. 'Pest in first': new prospects for biological control in tomatoes. [Dutch, English summary]. *Proeftuinnieuws* 8: 32-33.
4454. van Rheenen, H.A. 1973. Major problems of growing sesame (*Sesamum indicum* L.) in Nigeria. *Med. Landbouww. Wageningen* 12: 1-130. [Cock (1986)]
4455. van Roermund, H.J.W., Hemerik, L. and van Lenteren, J.C. 1994. Influence of intrapatch experiences and temperature on the time allocation of the whitefly parasitoid *Encarsia formosa* (Hymenoptera: Aphelinidae). *J. Insect Behav.* 7(4): 483-501.
4456. van Roermund, H.J.W., van Lenteren, J.C. and Rabbinge, R. 1997. Analysis of foraging behavior of the whitefly parasitoid *Encarsia formosa* on a leaf: a simulation study. *Biol. Control* 8(1): 22-36.
4457. van Schaik, P.H., Erwin, D.C. and Garber, M.J. 1962. Effects of time of symptom expression of the leaf-crumple virus on yield and quality of fiber of cotton. *Crop Sci.* 2: 275-277.
4458. van Schelt, J., Klapwijk, J., Letard, M. and Aucouturier, C. 1996. The use of *Macrolophus caliginosus* as a whitefly predator in protected crops. pp. 515-521. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia: 1995 Taxonomy, Biology, Damage, Control and Management*. Intercept Ltd., Andover, Hants, UK.
4459. van Velsen, R.J. 1967. Little leaf, a virus disease of *Ipomoea batatas* in Papua and New Guinea. *Papua New Guinea Agric. J.* 18(4): 126-128. [Cock (1986)]
4460. van Velsen, R.J. 1967. A mosaic disease of *Hibiscus manihot* in Papua and New Guinea. *Papua New Guinea Agric. J.* 19: 10-12. [Cock (1986)]
4461. van Wezel, R., Liu, H.T., Tien, P., Stanley, J. and Hong, Y.G. 2001. Gene C2 of the monopartite geminivirus Tomato yellow leaf curl virus-China encodes a pathogenicity determinant that is localized in the nucleus. *Mol. Plant-Microbe Interactions* 14(9): 1125-1128.
4462. Vandenberg, J.D., Shelton, A.M. and Wraight, S.P. Application and evaluation of entomopathogens in crucifers and cucurbits. pp. 389-403. *In* L.A. Lacey and H.K. Kaya (Eds.), *Field Manual of Techniques in Invertebrate Pathology*. Kluwer Academic Publishers, Dordrecht.
4463. Vargas-Camplis, J. 1996. Natural enemies of bollworm complex and other foliage feeding worms in northern Tamaulipas and their role in cotton production [Mexico]. pp. 710-712. *In* P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
4464. Varma, A., Dhar, A.K. and Mandal, B. 1992. MYMV (Mungbean Yellow Mosaic Disease) transmission and control in India. pp. 8-27. *In* S.K. Green and D. Kim (Eds.), *Mungbean Yellow Mosaic Disease: Proceedings International Workshop*, Bangkok, Thailand 2-3 July 1991. Asian Vegetable Research and Development Center, Taipei, Taiwan.
4465. Varma, P.M. 1952. Studies on the relationship of the *Bhendi* yellow vein-mosaic virus and its vector, the whitefly (*Bemisia tabaci* Gen.). *Indian J. Agric. Sci.* 22: 75-91. [Cock (1986)]
4466. Varma, P.M. 1955. Ability of the white-fly to carry more than one virus simultaneously. *Curr. Sci. (Bangalore)* 24: 317-318. [Cock (1986)]
4467. Varma, P.M. 1956. Persistence of yellow-vein mosaic virus of *Abelmoschus esculentus* (L.) Moench in its vector *Bemisia tabaci* (Gen.). *Indian J. Agric. Sci.* 25(1955): 293-302. [Cock (1986)]
4468. Varma, P.M. 1963. Transmission of plant viruses by whiteflies. *Bull. Natl. Inst. Sci. India* 24: 11-33. [Cock (1986)]
4469. Varma, P.M. and Capoor, S.P. 1959. Mosaic disease of cardamom and its transmission by the banana aphid *Pentalonia nigronervosa* Coq. *Indian J. Agric. Sci.* 28(1958): 97-107. [Cock (1986)]
4470. Vasudeva, R.S. 1953. Some important whitefly (*Bemisia tabaci* Gen.) transmitted viruses in India. *Raissunti delle Comunicazioni VI Congresso Int. di Microbiologia, Roma 6-12 Settembre 1952.* 2: 605-606. [Cock (1986)]
4471. Vasudeva, R.S. and Azad, R.N. 1948. Potato necrosis. *Curr. Sci. (Bangalore)* 17: 216-217. [Cock (1986)]
4472. Vasudeva, R.S. and Sahambi, H.S. 1955. Phyllody in sesamum (*Sesamum orientale* L.). *Indian Phytopathol.* 8: 124-129. [Cock (1986)]
4473. Vasudeva, R.S. and Sam Raj, J. 1948. A leaf curl disease of tomato. *Phytopathology* 38: 364-369. [Cock (1986)]
4474. Vavrina, C.S., Stansly, P.A. and Liu, T.X. 1995. Household detergent on tomato: phytotoxicity and toxicity to silverleaf whitefly. *Hortscience* 30(7): 1406-1409.
4475. Vazquez, L.L. 1999. Mosca blanca-geminivirus en el Caribe: estado actual y perspectivas. pp. 45-58. *In* 7th Taller Lationamericano y del Caribe Sobre Moscas-Blancas y Geminivirus. IPA, Recife, PE, Brasil. [Spanish]
4476. Vázquez, L.L., de la Iglesia, M., López, D., Jiménez, R., Mateo, A. and Vera, E.R. 1995. Mosca blancas (Homoptera: Aleyrodidae) detectadas en los principales cultivos agrícolas de Cuba. [Whiteflies detected in the main agricultural crops of Cuba]. *Manejo Integrado de Plagas (Costa Rica)* 36: 18-21. [Spanish, English summary]
4477. Vazquez, L.L., Jimenez, R., De La Iglesia, M., Mateo, A. and Borges, M. 1997. Planta hospedera de *Bemisia tabaci* (Homoptera: Aleyrodidae) en Cuba. *Rev. Biol. Trop.* 44(3/45)(1): 143-148.
4478. Veenstra, K.H. and Byrne, D.N. 1998. The effects of physiological factors and host plant experience on the ovipositional activity of the sweet potato whitefly, *Bemisia tabaci*. *Entomol. Exp. Appl.* 89(1): 15-23.
4479. Veenstra, K.H. and Byrne, D.N. 1998. Effects of starvation and oviposition activity on the reproductive physiology of the sweet potato whitefly, *Bemisia tabaci*. *Physiol. Entomol.* 23(1): 62-68.

- 4480.Veenstra, K.H. and Byrne, D.N. 1999. Does dispersal affect the reproductive physiology of the sweet potato whitefly, *Bemisia tabaci*? *Physiol. Entomol.* 24(1): 72-75.
- 4481.Vega, F.E., Jackson, M.A. and McGuire, M.R. 1999. Germination of conidia and blastospores of *Paecilomyces fumosoroseus* on the cuticle of the silverleaf whitefly, *Bemisia argentifolii*. *Mycopathologia* 147(1): 33-35.
- 4482.Vega, J., Munoz, R. and Pitty, A. 1992. Evaluacion de plagas, factores agronomicos y economicos del maiz y frijol en relevo bajo dos sistemas de labranza. [Evaluation of pests, economic and agronomic factors of the maize in rotation with beans under two tillage systems]. *Manejo Integrado De Plagas (Costa Rica)* 26: 13-20. [ Spanish, English summary]
- 4483.Veierov, D. 1996. Physically and behaviorally active formulations for control of *Bemisia*. pp. 557-576. In D. Gerling and R.T. Mayer (Eds.), *Bemisia* : 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK.
- 4484.Veierov, D., Fenigstein, A., Melamed-Madjar, V. and Klein, M. 1988. Effects of concentration and application method on decay and residual activity of foliar chlorpyrifos. *J. Econ. Entomol.* 81: 621-627.
- 4485.Velez, J.J., Bassett, M.J., Beaver, J.S. and Molina, A. 1998. Inheritance of resistance to bean golden mosaic virus in common bean. *J. Am. Soc. Hortic. Sci.* 123(4): 628-631.
- 4486.Venkatesan, S., Balasubramanian, G., Sivaprakasam, N., Narayanan, A. and Gopalan, M. 1987. Effect of intercropping of pulses and sunflower on the incidence of sucking pests of rainfed cotton. *Madras Agric. J.* 74(8-9): 364-368. [ Cock (1993)]
- 4487.Venkatesan, T. and Kundu, G.D. 1994. Bio-efficacy of insecticides for the control of stemfly and white fly infesting the soybean crop. *Indian J. Entomol.* 56(4): 418-421.
- 4488.Venkatesh, H.M., Muniyappa, V., Ravi, K.S., Krishnaprasad, P.R. and Verghese, A. 1998. Management of chilli leaf curl complex. pp. 111-117. In P.P. Reddy, N.K.K. Kumar and A. Varghese (Eds.), *Advances in IPM for Horticultural Crops. Proceedings of the First National Symposium on Pest Management in Horticultural Crops: Environmental Implications and Thrusts.* Indian Institute of Horticultural Research, Bangalore, India.
- 4489.Verma, A., Basu, D., Nath, P.S., Das, S., Ghatak, S.S. and Mukhopadhyay, S. 1989. Some ecological considerations of whitefly and whitefly transmitted virus diseases of vegetables in West Bengal. *Indian J. Virol.* 5: 79-87.
- 4490.Verma, A.K., Basu, D., Nath, P.S., Das, S., Ghatak, S.S. and Mukhopadhyay, S. 1989. Relationship between the population of whitefly, *Bemisia tabaci* Genn (Homoptera: Aleyrodidae) and the incidence of tomato leaf-curl virus disease. *Indian J. Mycol. Res.* 27(1): 49-52. [ Cock (1993)]
- 4491.Verma, A.K., Ghatak, S.S. and Mukhopadhyay, S. 1990. Effect of temperature on development of whitefly *Bemisia tabaci* (Homoptera: Aleyrodidae) in West Bengal India. *Indian J. Agric. Sci.* 60(5): 332-336. [ Cock (1993)]
- 4492.Verma, H.N., Srivastava, K.M. and Mathur, A.K. 1975. A whitefly - transmitted yellow mosaic virus disease of tomato from India. *Plant Dis. Rep.* 59: 494-498. [ Cock (1986)]
- 4493.Verma, M.M. and Sandhu, S.S. 1992. Breeding for resistance/tolerance to MYMV and its vector in India. pp. 28-40. In S.K. Green and D. Kim (Eds.), *Mungbean Yellow Mosaic Disease: Proceedings International Workshop, Bangkok, Thailand 2-3 July 1991.* Asian Vegetable Research and Development Center, Taipei, Taiwan.
- 4494.Verma, R.P.S. and Singh, D.P. 1989. Inheritance of resistance to mungbean yellow mosaic virus in blackgram. *Indian J. Genet. Plant Breed.* 49(3): 321-324. [ Cock (1993)]
- 4495.Verma, R.S., Das, S.B., Shaw, S.S., Mandloi, K.C. and Badaya, A.K. 1989. Chemical control of whitefly (*Bemisia tabaci*) on cotton. *J. Cotton Res. Dev.* 3(1): 49-51.
- 4496.Verma, V.S. 1974. Lupin leaf curl virus. *Gartenbauwissenschaft* 39: 55-56. [ Cock (1986)]
- 4497.Verma, V.S. 1974. Salvia yellow vein mosaic virus. *Gartenbauwissenschaft* 39: 565-566. [ Cock (1986)]
- 4498.Verma, V.S. 1974. Soapwort leaf curl virus. *Gartenbauwissenschaft* 39: 567-568. [ Cock (1986)]
- 4499.Verma, V.S. and Singh, S. 1973. Balsam leaf curl disease. *Hortic. Res.* 13: 55-56. [ Cock (1986)]
- 4500.Vetten, H.J. and Allen, D.J. 1983. Effects of environment and host on vector biology and incidence of two whitefly -spread diseases of legumes in Nigeria. *Ann. Appl. Biol.* 102: 219-227. [ Cock (1986)]
- 4501.Vicente, M., Kanthack, R.D., Noronha, A.B. and Stradioto, M.F.S. 1988. [Incidence of golden mosaic on bean grown in two planting seasons in the Presidente Prudente Region]. *Fitopatol. Brasileira* 13(4): 373-376. [ Cock (1993), Portuguese]
- 4502.Vicentini, S., Faria, M. and Oliveira, M.R.V. 2001. Screening of *Beauveria bassiana* (Deuteromycotina: Hyphomycetes) isolates against nymphs of *Bemisia tabaci* (Genn.) biotype B (Homoptera: Aleyrodidae) with description of a new bioassay method. *Neotrop. Entomol.* 30(1): 97-103.
- 4503.Vidal, C., Fargues, J. and Lacey, L.A. 1997. Intraspecific variability of *Paecilomyces fumosoroseus* : effect of temperature on vegetative growth. *J. Invertebr. Pathol.* 70(1): 18-26.
- 4504.Vidal, C., Fargues, J., Lacey, L.A. and Jackson, M.A. 1998. Effect of various liquid culture media on morphology, growth, propagule production, and pathogenic activity to *Bemisia argentifolii* of the entomopathogenic Hyphomycete, *Paecilomyces fumosoroseus*; propagule morphology. *Mycopathologia* 143(1): 33-46.
- 4505.Vidal, C., Lacey, L.A. and Fargues, J. 1997. Pathogenicity of *Paecilomyces fumosoroseus* (Deuteromycotina: Hyphomycetes) against *Bemisia argentifolii* (Homoptera: Aleyrodidae) with a description of a bioassay method. *J. Econ. Entomol.* 90(3): 765-772.
- 4506.Vidal, C., Osborne, L.S., Lacey, L.A. and Fargues, J. 1998. Effect of host plant on the potential of *Paecilomyces fumosoroseus* Deuteromycotina: Hyphomycetes) for controlling the silverleaf whitefly, *Bemisia argentifolii* (Homoptera: Aleyrodidae) in greenhouses. *Biol. Control* 12(3): 191-199.
- 4507.Vidavsky, F. and Czosnek, H. 1998. Tomato breeding lines resistant and tolerant to tomato yellow leaf curl virus issued from *Lycopersicon hirsutum*. *Phytopathology* 88(9): 910-914.
- 4508.Vidavsky, F., Leviatov, S., Milo, J., Rabinowitch, H.D., Kedar, N. and Czosnek, H. 1998. Response of tolerant breeding lines of tomato, *Lycopersicon esculentum* originating from three different sources (*L. peruvianum*, *L. pimpinellifolium* and *L. chilense*) to early controlled inoculation by tomato yellow leaf curl virus (TYLCV). *Plant Breeding* 117(2): 165-169.
- 4509.Videllet, P., Albajes, R. and Gabarra, R. 1997. Host-feeding activity of *Encarsia pergandiella* Howard on *Bemisia tabaci* (Gennadius). *Bull. OILB/SROP (IOBC/WPRS)* 20(4): 147-152.
- 4510.Vieira, M.R. and Correa, L.S. 2001. Whiteflies (Homoptera: Aleyrodidae) and the predator *Delphastus pusillus* (Le Conte) (Coleoptera: Coccinellidae) on papaya tree (*Carica papaya* L.) grown under screened conditions. *Neotrop. Entomol.* 30(1): 171-173. [ Portuguese, English summary]
- 4511.Viggiani, G. 1982. New species and host records of African aphelinids (Research on Hymenoptera Chalcidoidea LXX). *J. Entomol. Soc. Southern Africa* 45(1): 27-32. [ Cock (1986)]
- 4512.Viggiani, G. 1985. Notes on a few Aphelinidae, with descriptions of five new species of *Encarsia* Foerster (Hymenoptera, Chalcidoidea). *Bull. Lab. Entomol. Agraria 'Filippo Silvestri' Portici* 42: 81-94. [ Cock (1993)]
- 4513.Viggiani, G. 1986. Notes on some species of *Coccophagus* Westwood, *Coccophagoides* Girault, *Encarsia* Foerster and *Encarsiella* Hayst (Hymenoptera: Aphelinidae) mainly from the Nearctic and Neotropical Regions. *Bull. Lab. Entomol. Agraria 'Filippo Silvestri' Portici* 43: 59-78. [ Cock (1993)]

4514. Viggiani, G. 1987. A new species of *Encarsia* Foerster (*E. plaumanni* sp. n.) (Hymenoptera: Aphelinidae) from Brazil. Bull. Lab. Entomol. Agraria 'Filippo Silvestri Portici' 44: 77-80.
4515. Viggiani, G. 1987. New species of *Encarsia foerster* (Hymenoptera: Aphelinidae) parasitoids of whiteflies. Bull. Lab. Entomol. Agraria 'Filippo Silvestri' Portici 44: 33-42.
4516. Viggiani, G. 1987. Le specie italiane del genere *Encarsia* Foerster (Hymenoptera: Aphelinidae). Bull. Lab. Entomol. Agraria 'Filippo Silvestri' Portici 44: 25-31. [Cock (1993)]
4517. Viggiani, G. 1989. [Some novel insect pests]. Informatore Agrario 45(28): 63-64. [Cock (1993), Italian]
4518. Viggiani, G. 1991. Diversity of the Aphelinidae in agroecosystems. Bull. Lab. Entomol. Agraria 'Filippo Silvestri' Portici 48: 9-18.
4519. Viggiani, G. 1993. New species of *Encarsia* Foerster (Hymenoptera: Aphelinidae), parasitoids of whiteflies, from Hawaii and Yemen. Redia 76(1): 121-127.
4520. Viggiani, G. and Battaglia, D. 1983. Le specie Italiane del genere *Eretmocerus* Hald. (Homoptera: Aphelinidae). Bull. Lab. Entomol. Agraria 'Filippo Silvestri' Portici 40: 97-101. [Cock (1986)]
4521. Viggiani, G. and Evans, G.A. 1992. Descriptions of three new species of *Amitus* Haldeman (Hymenoptera: Platygasteridae), parasitoids of known whiteflies from the new world. Bull. Lab. Entomol. Agraria 'Filippo Silvestri' Portici 40: 189-194.
4522. Viggiani, G. and Laudonia, S. 1991. On the occurrence in Italy of *Encarsia meritoria* Gahan (Hymenoptera: Aphelinidae). Exotic parasitoid of whiteflies (Homoptera: Aleyrodidae). Redia 74(1): 135-140.
4523. Viggiani, G. and Mazzone, P. 1980. Le specie palaeartiche di *Encarsia* del gruppo *lutea* Masi (Hym. Aphelinidae), con descrizione di due nuove specie. Bull. Lab. Entomol. Agraria 'Filippo Silvestri', Portici. 37: 51-57. [Cock (1986)]
4524. Villalba, C. 1997. Multiple ToYMV-CR inoculations over tomato plants effect, treated with high dosages of phosphorus. M. Sc. Thesis. CATIE. Turrialba, Costa Rica, 54 pp.
4525. Villar, A., Gómez, E., Morales, F. and Anderson, P. 1998. Effect of legal measures to control *Bemisia tabaci* and geminiviruses in the Valley of Azua. National Integrated Pest Management Program, Santo Domingo, Dominican Republic Rep., 16 pp.
4526. Villeveille, M. and Lecoq, H. 1992. L'argenteure de la courgette, une maladie nouvelle en France, liée à un aleurode. [Squash silver leaf of the courgette (small marrow)]. Phytoma (France) 440: 35-36. [French]
4527. Vir, S. 1983. Efficacy of some important insecticides in the control of *Bemisia tabaci* (Genn.), a vector of the yellow mosaic disease on mothbean. Indian J. Plant Prot. 11(1-2): 31-33. [Cock (1986)]
4528. Vir, S. 1984. Assessment of yield loss due to yellow mosaic virus infection in mothbean. Pesticides 18(6): 33-34. [Cock (1986)]
4529. Viscarret, M.M. and Botto, E.N. 1996. Description and identification of *Trialeurodes vaporariorum* (Westwood) and *Bemisia tabaci* (Gennadius) (Hemiptera, Homoptera: Aleyrodidae). Rev. Chilena Entomol. 23: 51-58.
4530. Viscarret, M.M. and Botto, E.N. 1997. Numero, tiempo de desarrollo y supervivencia de estadios inmaduros de *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae, Aleyrodinae) en berenjena (*Solanum melanogeta*, Solanaceae). [Number, development time and survival of immature stages of *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae, Aleyrodinae) on berenjena (*Solanum melanogeta* Solanaceae)]. Boletín Sanidad Vegetal Plagas (Argentina) 23(4): 535-539. [Spanish, English summary]
4531. Viscarret, M.M., Botto, E.N. and Polaszek, A. 2000. Whiteflies (Hemiptera: Aleyrodidae) of economic importance and their natural enemies (Hymenoptera: Aphelinidae, Signiphoridae) in Argentina. Rev. Chilena Entomol. 26: 5-11.
4532. Visnya, A. 1941. Vorarbeiten zur Kenntnis der Aleruodiden-fauna von Ungarn, nebst systematischen Bemerkungen über die Gattungen *Aleurochiton*, *Pealius* und *Bemisia* (Homoptera). Fragm. Faun. Hung. 4 Suppl.: 1-19. [Hungarian]
4533. Vizgarra, O. 1995. Epidemiología y control integrado de las virosis del poroto transmitidas por mosca blanca en el NOA. [Epidemiology and integrated control of *Phaseolus vulgaris* virus diseases transmitted by whitefly in north east of Argentina]. Adv. Agroindustrial 16(62): 33-34. [Spanish]
4534. Vizgarra, O.N. and Morales, F.J. 2000. Historia y situación de la producción de frijol en los países latinoamericanos afectados por geminivirus transmitidos por mosca blanca: América del Sur: Bolivia. pp. 69-72. In Bean Golden Mosaic and Other Diseases of Common Bean Caused by Whitefly-Transmitted Geminiviruses in Latin America. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. [Spanish]
4535. Volker, D., Ernst, G.H., Ruesch, O. and Solang, U.K. 1990. Insecticide resistance in tropical and subtropical areas: experience in two decades. pp. 146-156. In Proceedings 3rd International Conference on Plant Protection in the Tropics, Genting Highlands, Pahang, Malaysia, 20-23 Mar 1990. Malaysian Plant Protection Society, Kuala Lumpur, Malaysia.
4536. von Arnim, A. and Stanley, J. 1992. Determinants of tomato golden mosaic virus symptom development located on DNA B. Virology 186: 286-293.
4537. von Arnim, A. and Stanley, J. 1992. Inhibition of African cassava mosaic virus systemic infection by a movement protein from the related geminivirus tomato golden mosaic virus. Virology 187: 555-564.
4538. von Arx, R., Baumgärtner, J. and Delucchi, V. 1983. A model to stimulate the population dynamics of *Bemisia tabaci* Genn. (Stern. Aleyrodidae) on cotton in the Sudan Gezira. Z. Angew. Entomol. 96: 341-363. [Cock (1986)]
4539. von Arx, R.V., Baumgärtner, J. and Delucchi, V. 1983. Developmental biology of *Bemisia tabaci* (Genn.) (Sternorrhyncha, Aleyrodidae) on cotton at constant temperatures. Bull. Swiss Entomol. Soc. 56: 389-399. [Cock (1986)]
4540. von Arx, R.V., Baumgärtner, J. and Delucchi, V. 1984. Sampling of *Bemisia tabaci* (Genn.) (Sternorrhyncha: Aleyrodidae) in Sudanese cotton fields. J. Econ. Entomol. 77: 1130-1136. [Cock (1986)]
4541. von Boguslawski, C. and Basedow, T. 2001. Studies in cotton fields in Egypt on the effects of pheromone mating disruption on *Pectinophora gossypiella* (Saund.) (Lep., Gelechiidae), on the occurrence of other arthropods, and on yields. J. Appl. Entomol. 125(6): 327-331.
4542. Vovlas, C., Sialer, M.F., Di Franco, A. and Gallitelli, D. 2001. Tomato yellow leaf curl [geminivirus] comes to Apulia. Informatore Agrario 57(6): 61-63. [Italian, English summary]
4543. Wagner, T.L. 1993. Temperature-dependent development of sweetpotato whitefly, *Bemisia tabaci* (Homoptera: Aleyrodidae), Biotype "B". pp. 714-718. In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
4544. Wagner, T.L. 1994. Temperature-dependent reproduction of sweetpotato whitefly, *Bemisia tabaci* (Homoptera: Aleyrodidae). pp. 885-886. In D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
4545. Wagner, T.L. 1995. Temperature-dependent development, mortality, and adult size of sweetpotato whitefly biotype B on cotton. Environ. Entomol. 24: 1179-1188.
4546. Wagner, T.L. and Willers, J.L. 1995. Field validation of a model describing development times of sweetpotato whitefly. pp. 840-843. In D.A. Richter and J. Armour (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.

4547. Wahla, M.A., Tufail, M. and Akbar, S. 1996. The comparative efficacy of different doses of Dimecron 100 Scw (Phosphamidon) against the whitefly, *Bemisia tabaci* (Genn.) on FH-682, cotton. *Pakistan Entomol.* 18(1/2): 45-46.
4548. Walker, G.P. and Gordh, G. 1989. The occurrence of apical labial sensilla in the Aleyrodidae and evidence for a contact chemosensory function. *Entomol. Exp. Appl.* 51(3): 215-224. [Cock (1993)]
4549. Walker, G.P. and Perring, T.M. 1994. Feeding and oviposition behavior of whiteflies (Homoptera: Aleyrodidae) interpreted from AC electronic feeding monitor waveforms. *Ann. Entomol. Soc. Am.* 87(3): 363-374.
4550. Wang, C.L. 1998. Two predacious *Orius* flower bugs (Hemiptera: Anthocoridae) in Taiwan. *Chinese J. Entomol.* 18(3): 199-202.
4551. Wang, H.L., Sudarshana, M.R., Gilbertson, R.L. and Lucas, W.J. 1999. Analysis of cell-to-cell and long-distance movement of a bean dwarf mosaic geminivirus-green fluorescent protein reporter in host and nonhost species: identification of sites of resistance. *Mol. Plant-Microbe Interactions* 12(4): 345-355.
4552. Wang, K.H. and Tsai, J.H. 1996. Temperature effect on development and reproduction of silverleaf whitefly (Homoptera: Aleyrodidae). *Ann. Entomol. Soc. Am.* 89(3): 375-384.
4553. Wangboonkong, S. 1981. Chemical control of cotton insect pests in Thailand. *Trop. Pest Manage.* 27: 495-500. [Cock (1986)]
4554. Warburg, O. 1894. Die Kulturpflanzen Usambaras. Mitteilungen von Forschungsreisenden und Gelehrten aus den deutschen Schutzgebieten 7: 131-199. [Cock (1986), German]
4555. Wardlow, L.R. 1989. Integrated pest management in poinsettias grown under glass. *Med. Fac. Landbouww. Univ. Gent* 54(3a): 867-872. [Cock (1993)]
4556. Wardlow, L.R. and van Lenteren, J.C. (Eds.). 1993. IOBC/WPRS workshop on IPM in greenhouse ornamentals. *Bull. OILB/SROP (IOBC/WPRS)* 16(8): 1-166.
4557. Watson, J.R. 1917. Florida truck and garden insects. *Bull. Florida Agric. Exp. Stn.* 134: 35-127. [Cock (1986)]
4558. Watson, J.R. 1918. Insects of a citrus grove. *Bull. Florida Agric. Exp. Stn.* 148: 165-267. [Cock (1986)]
4559. Watson, J.S., Hopper, B.L. and Tipton, J.D. 1982. Whitefly and the problem of sticky cotton. *Sp an* 25(2): 71-73. [Cock (1986)]
4560. Watson, T.F. 1993. Chemical control of the sweetpotato whitefly in cotton. pp. 221-239. *In* Cotton, A College of Agriculture Report, Series P-94. University of Arizona, Tucson.
4561. Watson, T.F. 1994. Sweetpotato whitefly seasonal biology. pp. 142-143. *In* D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
4562. Watson, T.F. 1995. Silverleaf whitefly: keys to management. pp. 160-161. *In* D.A. Richter and J. Armour (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
4563. Watson, T.F. and Silvertooth, J.C. 1992. Seasonal dynamics of sweetpotato whitefly. pp. 657-664. *In* D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
4564. Watson, T.F., Silvertooth, J.C. and Brown, P.W. 1993. Host plants associated with outbreaks of sweetpotato whitefly as it relates to population management in cotton. pp. 671-672. *In* D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
4565. Watson, T.F., Silvertooth, J.C. and Tellez, A. 1994. Varietal and nitrogen-level effects on sweetpotato whitefly populations in cotton. pp. 352-356. *In* Cotton, A College of Agriculture Report, Series P-96. University of Arizona, Tucson.
4566. Watson, T.F., Silvertooth, J.C. and Tellez, A. 1994. Varietal and nitrogen-level effects on sweetpotato whitefly populations in cotton. pp. 868-869. *In* D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
4567. Watson, T.F., Silvertooth, J.C., Tellez, A. and Lastra, L. 1992. Seasonal dynamics of sweetpotato whitefly in Arizona. *Southwest Entomol.* 17: 149-167.
4568. Watson, T.F., Tellez, A. and Peña, M. 1994. Chemical control of the sweetpotato whitefly in cotton. pp. 326-343. *In* Cotton, A College of Agriculture Report, Series P-96. University of Arizona, Tucson.
4569. Watson, T.F., Tellez, A. and Peña, M. 1995. Chemical control of the sweetpotato whitefly in cotton - 1994. pp. 259-270. *In* Cotton, A College of Agriculture Report, Series P-99. University of Arizona, Tucson.
4570. Webb, S.E. 1992. Floating row covers exclude insects affecting fall-grown squash in Central Florida. *Proc. Florida State Hort. Soc.* 104: 272-275.
4571. Webb, S.E. 1994. Management of insect pests of squash. *Proc. Florida State Hort. Soc.* 106: 165-168.
4572. Webb, S.E. 1997. Control of aphids and whiteflies on watermelon, 1995. *Arthropod Management Tests* 22: 194.
4573. Webb, S.E. and Linda, S.B. 1992. Evaluation of spunbonded polyethylene row covers as a method excluding insects and viruses affecting fall-grown squash in Florida. *J. Econ. Entomol.* 85(6): 2344-2352.
4574. Webb, S.E. and Mann, W.K. 1999. Evaluation of two systemic insecticides for control of melon aphid and silverleaf whitefly on squash, 1998. *Arthropod Management Tests* 24: 171-172.
4575. Weber, H. 1935. Der bau der imago der Aleurodinen. *Zoologica Heft* 89: 1-71.
4576. Weber, H. 1995. The anatomy of adult aleyrodids. An English translation of *Zoologica*, Heft 89, Stuttgart 1935: Der Bau de Imago der Aleurodinen. Ein Beitrag zur vergleichenden Morphologie des Insektenkörpers. [F. Schulthess, Translator]. D.L. Hendrix, R.J. Gill and E.W. Davidson (Eds.), E. Schweizerbart'sche Verlagsbuchhandlung, Stuttgart, 60 pp.
4577. Wei, Y.A., Hendrix, D.L. and Nieman, R. 1996. Isolation of a novel tetrasaccharide, bemisiotetrose, and glycine betaine from silverleaf whitefly honeydew. *J. Agric. Food Chem.* 44(10): 3214-3218.
4578. Wei, Y.A., Hendrix, D.L. and Nieman, R. 1997. Diglucolezitose, a novel pentasaccharide in silverleaf whitefly honeydew. *J. Agric. Food Chem.* 45(9): 3481-3486.
4579. Weintraub, P.G., Arazi, Y. and Horowitz, A.R. 1996. Management of insect pests in celery and potato crops by pneumatic removal. *Crop Prot.* 15(8): 763-769.
4580. Weintraub, P.G. and Horowitz, A.R. 1999. Management of the whitefly *Bemisia tabaci* (Genn.) on melon by vacuum removal. *Insect Sci. Appl.* 19(2-3): 173-178.
4581. Weller, G.L. and Graver, J.E.V. 1998. Cut flower disinfestation: assessment of replacement fumigants for methyl bromide. *Postharvest Biol. Tech.* 14(3): 325-333.
4582. Wessel-Beaver, L. 1997. Screening for silverleaf resistance in *Cucurbita* accessions. *Rep. Cucurbit Genetics Coop.* 20: 54-56.
4583. Wessel-Beaver, L. and Paris, H.S. 2000. Inheritance of silverleaf resistance in *Cucurbita moschata*. *Acta Hort.* 510: 289-295.
4584. West, J. 1936. Leaf curl of tobacco in southern Nigeria. *Trop. Agric. (Trinidad)* 13: 242-244. [Cock (1986)]
4585. Wilhoit, L., Schoenig, S., Supkoff, D. and Johnson, B.A. 1994. A regional simulation model for silverleaf whitefly in the Imperial Valley. PM 94-01. *Pest Management Analysis and Planning Program*, California, 109 pp.
4586. Williams, C.B. 1934. Field studies on the relation of insect pests to climatic conditions, with special reference to cotton. *Rep. Conf. Cotton-growing Problems* 2: 111-125. [Cock (1986)]
4587. Williams, F.J., Grewal, J.S. and Amin, K.S. 1968. Serious and new diseases of pulse crops in India in 1966. *Plant Dis. Rep.* 52: 300-304. [Cock (1986)]

4588. Williams, J.R. 1977. Some features of sex-linked hyperparasitism in Aphelinidae (Hymenoptera). *Entomophaga* 22: 345-350. [Cock (1986)]
4589. Williams, L. and Dennehy, T.J. 1996. Whitefly control in Arizona: developing a resistance management program for imidacloprid. *Resist. Pest. Manage.* 8(1): 48-52.
4590. Williams, L.A.D., Mansingh, A. and Mansingh, A. 1996. The insecticidal and acaricidal actions of compounds from *Azadirachta indica* (A. Juss.) and their use in tropical pest management. *Integrated Pest Manage. Rev.* 1(3): 133-145.
4591. Williams, L., III, Dennehy, T.J. and Palumbo, J.C. 1996. Whitefly control in Arizona: development of a resistance management program for imidacloprid. pp. 752-755. *In* P. Dugger and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
4592. Williams, L., III, Dennehy, T.J. and Palumbo, J.C. 1997. Defining the risk of resistance to imidacloprid in Arizona populations of whitefly. pp. 1242-1246. *In* P. Dugger and D. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
4593. Williams, L. III, Dennehy, T.J. and Palumbo, J.C. 1998. Can resistance to chloronicotinyl insecticides be averted in Arizona field crops? pp. 341-351. *In* Cotton, A College of Agriculture Report, Series P-112. University of Arizona, Tucson.
4594. Williams, M.L. and Miller, G.L. 1989. Chemical control of sweetpotato whitefly on hibiscus. *Res. Rep. Ser. - Alabama Agric. Exp. Stn.* ((6)): 9-10.
4595. Williams, R.J. 1976. A whitefly -transmitted golden mosaic of lima beans in Nigeria. *Plant Dis. Rep.* 60: 853-857. [Cock (1986)]
4596. Williams, T. 1995. The biology of *Encarsia tricolor*: an autoparasitoid of whitefly. *Biol. Control* 5(2): 209-217.
4597. Wilson, D. and Anema, B.P. 1988. Development of buprofezin for control of whitefly *Trialetrodes vaporariorum* and *Bemisia tabaci* on glasshouse crops in the Netherlands and the UK. pp. 175-180. *In* Brighton Crop Protection Conference: Pests and Diseases. British Crop Protection Council, Thornton Heath, UK. [Cock (1993)]
4598. Wilson, F.D. and Brown, J.K. 1991. Inheritance of response to cotton leaf crumple virus in cotton. *J. Hered.* 82: 508-509.
4599. Wilson, F.D., Brown, J.K. and Butler, G.D., Jr. 1988. Natural resistance of cotton to cotton leaf crumple virus. pp. 185-187. *In* Cotton, A College of Agriculture Report, Series P-72. University of Arizona, Tucson.
4600. Wilson, F.D., Brown, J.K. and Butler, G.D., Jr. 1989. Reaction of cotton cultivars and lines to cotton leaf crumple virus. *J. Arizona-Nevada Acad. Sci.* 23: 7-10.
4601. Wilson, F.D. and Butler, G.D., Jr. 1987. Whitefly adults in okra-leaf and normal-leaf cottons. pp. 163-165. *In* Cotton, A College of Agriculture Report, Series P-69. University of Arizona, Tucson.
4602. Wilson, F.D., Flint, H.M., Deaton, W.R., Fischhoff, D.A., Perlak, F.J., Armstrong, T.A., Fuchs, R.L., Berberich, S.A., Parks, N.J. and Stapp, B.R. 1992. Resistance of cotton lines containing a *Bacillus thuringiensis* toxin to pink bollworm (Lepidoptera: Gelechiidae) and other insects. *J. Econ. Entomol.* 85(4): 1516-1521.
4603. Wilson, F.D., Flint, H.M., Stapp, B.R. and Parks, N.J. 1993. Evaluation of cultivars, germplasm lines, and species of *Gossypium* for resistance to biotype "B" of sweetpotato whitefly (Homoptera: Aleyrodidae). *J. Econ. Entomol.* 86(6): 1857-1862.
4604. Wilson, F.D. and George, B.W. 1986. Smoothleaf and hirsute cottons: response to insect pests and yield in Arizona. *J. Econ. Entomol.* 79(1): 229-232. [Cock (1993)]
4605. Wilson, K.I. 1972. Chlorotic ring spot of Jasmine. *Indian Phytopathol.* 25: 157-158. [Cock (1986)]
4606. Wilson, K.I. and Potty, V.P. 1972. Yellow vein mosaic of *Blumea neilgherrensis* Hook. *Agric. Res. J. (Kerala)* 10: 68. [Cock (1986)]
4607. Wilson, T.M.A. and Davies, J.W. 1994. New roads to crop protection against viruses. *Outlook Agric.* 23(1): 33-39.
4608. Winter, S., Purac, A., Leggett, F., Frison, E.A., Rossel, H.W. and Hamilton, R.I. 1992. Partial characterization and molecular cloning of a Closterovirus from sweet potato infected with the sweet potato virus disease complex from Nigeria. *Phytopathology* 82(8): 869-875.
4609. Wisler, G.C. and Duffus, J.E. 2001. Transmission properties of whitefly-borne criniviruses and their impact on virus epidemiology. pp. 293-308. *In* K.F. Harris, O.P. Smith and J.E. Duffus (Eds.), *Virus-Insect-Plant Interactions*. Academic Press, San Diego, CA.
4610. Wisler, G.C., Duffus, J.E., Liu, H.Y., Li, R. and Falk, B.W. 1996. New whitefly-transmitted closterovirus identified in tomatoes. *California Agric.* 51(2): 24-26.
4611. Wisler, G.C., Duffus, J.E., Liu, H.Y. and Li, R.H. 1998. Ecology and epidemiology of whitefly-transmitted closteroviruses. *Plant Dis.* 82(3): 270-280.
4612. Wisler, G.C., Li, R.H., Liu, H.Y., Lowry, D.S. and Duffus, J.E. 1998. Tomato chlorosis virus: A new whitefly-transmitted, phloem-limited, bipartite closterovirus of tomato. *Phytopathology* 88(5): 402-409.
4613. Withthayarak, W., Bansit, K. and Chawalitwongphon, P. 1983. Field evaluation of the effectiveness and intervals of insecticides for controlling white fly. pp. 115-126. *In* Research Report 1983: Vegetables and Ornamental Plants Insect Pests Research Group. Department of Agriculture, Entomology and Zoology Division, Bangkok, Thailand. [Thai, English summary]
4614. Wolf, F.A., Whitcomb, W.H. and Mooney, W.C. 1949. Leafcurl of tobacco in Venezuela. *J. Elisha Mitchell Sci. Soc.* 65: 38-47. [Cock (1986)]
4615. Wolfe, G.R., Hendrix, D.L. and Salvucci, M.E. 1998. A thermoprotective role for sorbitol in the silverleaf whitefly, *Bemisia argentifolii*. *J. Insect Physiol.* 44(7-8): 597-603.
4616. Wolfe, G.R., Smith, C.A., Hendrix, D.L. and Salvucci, M.E. 1999. Molecular basis for thermoprotection in *Bemisia*: structural differences between whitefly ketose reductase and other medium-chain dehydrogenases/reductases. *Insect Biochem. Mol. Biol.* 29(2): 113-120.
4617. Wolfenbarger, D.A. and Cook, C.G. 1996. Response of whitefly populations to imidacloprid in stressed and unstressed irrigated cotton. pp. 1140-1142. *In* P. Dugger and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
4618. Wolfenbarger, D.A. and Riley, D.G. 1994. Toxicity of mixtures of insecticides and insecticides alone against B-strain sweetpotato whitefly. pp. 1214-1216. *In* D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
4619. Wolfenbarger, D.A., Riley, D.G., Akey, D.H. and Jones, W.A. 1994. Endosulfan and bifenthrin in alternate applications and mixture against B-strain whitefly. pp. 901-903. *In* D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
4620. Wolfenbarger, D.A., Riley, D.G., Staetz, C.A., Leibe, G.L., Herzog, G.A. and Gage, E.V. 1998. Responses of silverleaf whitefly (Homoptera: Aleyrodidae) to bifenthrin and endosulfan by vial bioassay in Florida, Georgia and Texas. *J. Entomol. Sci.* 33(4): 412-420.
4621. Wolny, W. and Godawa, H. 1994. Quarantine agrophages determined by EPP0. Supplement No. 14. *Ochroa Roslin* 38(5): 20-21. [Polish]
4622. Womac, A.R., Mulrooney, J.E., Howard, K.D. and Sumner, H.R. 1993. Advancements in sprayer technology for sweetpotato whitefly control. pp. 1621-1622. *In* D.J. Herber and D.A. Richter (Eds.), *Proceedings Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN.
4623. Wong, S.M., Swanson, M.M. and Harrison, B.D. 1993. A geminivirus causing vein yellowing of *Ageratum conyzoides* in Singapore. *Plant Pathol.* 42: 137-139.

4624. Wood, J.P. and Godfrey, L.D. 1998. Effects of whitefly insect growth regulators Knack and Applaud on cotton aphid reproduction and survival. pp. 1278-1281. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
4625. Wood, J.P., Godfrey, L.D., Goodell, P.B. and Eckert, J. 1997. Spatial and temporal distribution of the silverleaf whitefly in the San Joaquin Valley 1994-1996. pp. 1240-1241. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
4626. Wood, M. 1988. Culprit in winter lettuce price rise. *Agric. Res. (USDA)* 36(2): 4-5.
4627. Wood, M. 1988. Scientists take aim on lettuce menaces. *Agric. Res. (USDA)* 36(8): 10-12. [Cock (1993)]
4628. Wood, M. 2001. Protecting poinsettias. *Agric. Res. (USDA)* 49(12): 6-7.
4629. Woods, C. 1989. Growers fear repeat of last year's whitefly outbreak. *Citrus & Vegetable Mag.* 52(9): 32-34. [Cock (1993)]
4630. Woodward, T.E., Evans, J.W. and Easthop, V.F. 1970. Hemiptera. pp. 387-457. *In* Commonwealth Scientific and Industrial Research Organization (CSIRO) The insects of Australia. Melbourne Univ. Press, Melbourne, Australia. [Cock (1986)]
4631. Wool, D., Calvert, L., Constantino, L.M., Bellotti, A.C. and Gerling, D. 1994. Differentiation of *Bemisia tabaci* (Genn) (Hom., Aleyrodidae) populations in Columbia. *J. Appl. Entomol.* 117(2): 122-134.
4632. Wool, D., Gerling, D., Bellotti, A., Morales, F. and Nolt, B. 1991. Spatial and temporal genetic variation in populations of the whitefly *Bemisia tabaci* (Genn.) in Israel and Colombia: An interim report. [French summary]. *Insect Sci. Appl.* 12(1-3): 225-230. [Cock (1993)]
4633. Wool, D., Gerling, D., Bellotti, A.C. and Morales, F.J. 1993. Esterase electrophoretic variation in *Bemisia tabaci* (Genn) (Hom., Aleyrodidae) among host plants and localities in Israel. *J. Appl. Entomol.* 115: 185-196.
4634. Wool, D., Gerling, D. and Cohen, I. 1984. Electrophoretic detection of two endoparasite species, *Encarsia lutea* and *Eretmocerus mundus* in the whitefly, *Bemisia tabaci* (Genn.) (Hom., Aleyrodidae). *Z. Angew. Entomol.* 98: 276-279. [Cock (1986)]
4635. Wool, D., Gerling, D., Nolt, B.L., Constantino, L.M., Bellotti, A.C. and Morales, F.J. 1989. The use of electrophoresis for identification of adult whiteflies (Homoptera: Aleyrodidae) in Israel and Colombia. [German summary]. *J. Appl. Entomol.* 107(4): 344-350. [Cock (1993)]
4636. Wool, D. and Greenberg, S. 1990. Esterase activity in whiteflies (*Bemisia tabaci*) in Israel in relation to insecticide resistance. [French summary]. *Entomol. Exp. Appl.* 57(3): 251-258. [Cock (1993)]
4637. Woolley, J.B. 1997. Aphelinidae. pp. 134-150. *In* G.A.P. Gibson, J.T. Huber and J.B. Woolley (Eds.), Annotated Keys to the Genera of Nearctic Chalcidoidea (Hymenoptera). National Research Council of Canada, Ottawa, Canada.
4638. Woolley, J.M. and Heraty, J.M. 1998. *Encarsia* species of the world: a searchable database. [a catalogue of about 313 species of *Encarsia* with information on types, distribution and hosts]. URL <http://chalcidoids.tamu.edu/Encarsia.htm>
4639. Wraight, S.P. 1997. Evaluation of entomopathogenic fungi in field crops: methods for research and development. pp. 241-243. *In* Hacia un Futuro Agrícola Sostenible, Memoria: XX Congreso Nacional de Control Biológico. Universidad de Guadalajara, Guadalajara, Mexico.
4640. Wraight, S.P. and Bradley, C.A. 1996. Production, formulation, and application technologies for use of entomopathogenic fungi to control field crop pests. pp. 170-177. *In* 5th Simpósio de Controle Biológico, Anais Conferências e Palestras. EMBRAPA-CNPQSO, Foz do Iguaçu, Brasil.
4641. Wraight, S.P. and Carruthers, R.I. 1999. Production, delivery, and use of mycoinsecticides for control of insect pests of field crops. pp. 233-269. *In* F.R. Hall and J.J. Menn (Eds.), Methods in Biotechnology, Vol. 5: Biopesticides: Use and Delivery. Humana Press, Totowa, NJ.
4642. Wraight, S.P., Carruthers, R.I. and Bradley, C.A. 1996. Development of entomopathogenic fungi for microbial control of whiteflies of the *Bemisia tabaci* complex. pp. 28-34. *In* 5th Simpósio de Controle Biológico, Anais: Conferências e Palestras. EMBRAPA-CNPQSO, Foz do Iguaçu, Brasil.
4643. Wraight, S.P., Carruthers, R.I., Bradley, C.A., Jaronski, S.T., Lacey, L.A., Wood, P. and Galaini-Wraight, S. 1998. Pathogenicity of the entomopathogenic fungi *Paecilomyces* spp. and *Beauveria bassiana* against the silverleaf whitefly, *Bemisia argentifolii*. *J. Invertebr. Pathol.* 71(3): 217-226.
4644. Wraight, S.P., Carruthers, R.I., Jaronski, S.T., Bradley, C.A., Garza, C.J. and Galaini-Wraight, S. 2000. Evaluation of the entomopathogenic fungi *Beauveria bassiana* and *Paecilomyces fumosoroseus* for microbial control of the silverleaf whitefly, *Bemisia argentifolii*. *Biol. Control* 17(3): 203-217.
4645. Wright, J.E. 1992. Whiteflies: Development of NATURALIS, a biorational mycoinsecticide for control. pp. 887-888. *In* D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
4646. Wright, J.E., Bouse, L.F., Kirk, I.W., Carlton, J.B., Franz, E., Latheef, M.A. and Rektorik, R. 1993. Full season control of cotton insects in the Rio Grande Valley of Texas. pp. 849-855. *In* D.J. Herber and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
4647. Wright, J.E. and Knauf, T.A. 1994. Evaluation of Naturalis-L for control of cotton insects. pp. 45-52. *In* Brighton Crop Protection Conference: Pests and Diseases. British Crop Protection Council, Farnham, UK.
4648. Wrona, A.F., Kerby, T. and Shouse, P. 1995. Effect of irrigation timing on yield and earliness of five cotton varieties. pp. 1108-1109. *In* D.A. Richter and J. Armour (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
4649. Wu, Z.C. and Hu, J.S. 1995. Comparison of ELISA, dot-blot, and PCR assays for detection of whitefly-transmitted geminivirus. *Int. J. Trop. Plant Dis.* 13(2): 205-211.
4650. Wu, Z.C., Hu, J.S., Polston, J.E., Ullman, D.E. and Hiebert, E. 1996. Complete nucleotide sequence of a nonvector-transmissible strain of *Abutilon* mosaic geminivirus in Hawaii. *Phytopathology* 86(6): 608-613.
4651. Wyatt, B.G. and Heintz, C.E. 1982. Capsule-producing coryneform bacteria associated with stickiness in cotton. *Textile Res. J.* 52: 518-523.
4652. Wyatt, S.D. and Brown, J.K. 1996. Detection of subgroup III geminivirus isolates in leaf extracts by degenerate primers and polymerase chain reaction. *Phytopathology* 86(12): 1288-1293.
4653. Wyman, A.M., Steffens, R.J., Elbert, A. and Kuehnhold, J. 2001. Calypso: a new neonicotinoid insecticide for cotton and several other crops. pp. 47-49. *In* P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.
4654. Wysoki, M. and Cohen, M. 1983. Mites of the family Phytoseiidae (Acarina, Mesostigmata) as predators of the Japanese bayberry whitefly, *Parabemisia myricae* Kuwana (Hom., Aleyrodidae). *Agronomie (Paris)* 3: 823-825. [Cock (1986)]
4655. Yadav, D.N., Patel, R.C., Jo shi, A.D. and Patel, A.K. 1987. Resurgence of insect and mite pests on hybrid cotton in Gujarat. pp. 191-196. *In* S. Jayaraj (Ed.), Resurgence of Sucking Pests: Proceedings of National Symposium. Centre For Plant Protection Studies, Tamil Nadu Agric. Univ.
4656. Yadav, G.S. and Dahiya, B. 2000. Screening of some mung bean genotypes against major insect pests and yellow mosaic virus. *Ann. Agric. Bio. Res.* 5(1): 71-73.

4657. Yadav, L.B. and Rizvi, S.M.A. 1994. Studies on the insect pests of wasteland plantations at Faizabad (India). *J. Entomol. Res. (New Delhi)* 18(2): 115-120.
4658. Yadav, L.B. and Rizvi, S.M.A. 1995. Studies on the insect-pests of lime, *Citrus aurantifolia* Swinh., in Eastern Uttar Pradesh. *Ann. Plant Prot. Sci.* 3(1): 34-37.
4659. Yadav, L.S. and Yadav, P.R. 1983. Pest complex of cowpea (*Vigna sinensis* Savi) in Haryana. *Bull. Entomol. (New Delhi)* 24: 57-58. [Cock (1986)]
4660. Yamashita, S., Doi, Y., Yora, K. and Yoshino, M. 1979. Cucumber yellows virus: its transmission by the greenhouse whitefly, *Trialeurodes vaporariorum* (Westwood), and the yellowing disease of cucumber and muskmelon caused by the virus. *Ann. Phytopathol. Soc. Japan* 45: 484-496. [Cock (1986)]
4661. Yan, F.M. 2001. Application of non-morphological characters in taxonomy of whiteflies (Homoptera: Aleyrodidae). *Entomotaxonomia* 23(2): 107-113. [Chinese]
4662. Yanez-Morales, M.J. 1992. Características de la población alada de "mosquita blanca" (Homoptera: Aleyrodidae) en Chile serrano en el sur de Tamaulipas, Mexico. [Alate population characteristics of whitefly (Homoptera: Aleyrodidae) in "serrano" pepper in the south of Tamaulipas, Mexico]. *Agrociencia (Mexico). Serie Proteccion Vegetal.* 3(2): 7-18. [Spanish, English summary]
4663. Yano, E., van Lenteren, J.C., Rabbinge, R. and Hulspar-Jordan, P.M. 1989. The parasite-host relationship between *Encarsia formosa* (Hymenoptera: Aphelinidae), and *Trialeurodes vaporariorum* (Homoptera: Aleyrodidae) XXXI. Simulation studies of population growth of greenhouse whitefly on tomato. *Agric. Univ. Wageningen Papers* 89(2): 55-73.
4664. Yano, E., van Lenteren, J.C., Rabbinge, R., van Vianen, A., Dorsman, R. and Hulspar-Jordan, P.M. 1989. The parasite-host relationship between *Encarsia formosa* and *Trialeurodes vaporariorum*. XXXII. Simulation studies of the population growth of greenhouse whitefly on egg plant, cucumber, sweet pepper and gerbera. *Agric. Univ. Wageningen Papers* 89(2): 75-99.
4665. Yaranguntian, R.C. and Govindu, H.C. 1964. Virus disease of *Dolichos lablab* var. *typicum* from Mysore. *Curr. Sci. (Bangalore)* 33: 721-722. [Cock (1986)]
4666. Yarakinci, N. and Hincal, P. 1996. The research on the population development of the major pests (*Trialeurodes vaporariorum* (Westw.), *Bemisia tabaci* Gen. (Homoptera: Aleyrodidae) and *Liriomyza* Spp. (Diptera: Agromyzidae)) in the tomato glasshouses in Izmir. *Plant Protection Research Institute, Izmir, Turkey*, 8 pp. [Turkish, English summary]
4667. Yarakinci, N., Hincal, P. and Tuncay, O. 1999. The population growth of pests and their beneficials in cucumber plastic tunnels in the Izmir region of Turkey. *Acta Hort.* 491: 461-468.
4668. Yasnosh, V.A. 1989. Species of the genus *Encarsia* (Hymenoptera: Aphelinidae) - parasites of Aleyrodids in the USSR. *Proc. Zool. Inst., Leningrad* 191: 109-121. [Cock (1993), Russian]
4669. Yasnosh, V.A. 1991. Whiteflies and their natural enemies. *Zashchita Rastenii (Moskva)* 2: 24-26. [Russian]
4670. Yassin, A.M. 1975. Epidemics and chemical control of leaf curl virus disease of tomato in the Sudan. *Exp. Agric.* 11(3): 161-165. [Cock (1986)]
4671. Yassin, A.M. 1978. Whitefly - borne virus diseases of important crop plants in the Sudan. pp. 107-114. *In Proc. Fourth Conference of Pest Control, September 30-October 3, 1978. Academy of Scientific Research and Technology and National Research Center, Cairo, Egypt.* [Cock (1986)]
4672. Yassin, A.M. 1979. Leaf curl of tomato. pp. 85-89. *In Annu. Rep. Gezira Res. Stn. Substns. 1972/1973. Agric. Res. Corp., Sudan* [Cock (1993)]
4673. Yassin, A.M. 1983. A review of factors influencing control strategies against tomato leaf curl virus disease in the Sudan. *Trop. Pest Manage.* 29: 253-256. [Cock (1986)]
4674. Yassin, A.M. and Bendixen, L.E. 1982. Weed hosts of the cotton whitefly (*Bemisia tabaci* Genn.) Homoptera: Aleyrodidae. *Res. Bull., Ohio Agric. Res. Dev. Cent.* 1144: 10 pp. [Cock (1986)]
4675. Yassin, A.M. and Dafalia, G.D. 1982. Cotton reddening in the Gezira. *Trop. Pest Manage.* 28: 312-313.
4676. Yassin, A.M. and El-Nur, E. 1970. Transmission of cotton leaf curl virus by single insects of *Bemisia tabaci*. *Plant Dis. Rep.* 54(6): 528-531.
4677. Yassin, A.M. and Nour, M.A. 1965. Tomato leaf curl diseases in the Sudan and their relation to tobacco leaf curl. *Ann. Appl. Biol.* 56: 207-217. [Cock (1986)]
4678. Yassin, K.M., Bashir, N.H.H. and Gadalla, B.H. 1990. Effects of endosulfan, chlorpyrifos and their mixtures on *Bemisia tabaci* of Sudan Gezira. *Trop. Pest Manage.* 36(3): 230-233. [Cock (1993)]
4679. Yasui, M., Ellsworth, P.C., Lublinkhof, J. and Comer, D. 1997. Monitoring whitefly susceptibility to Applaud. pp. 257-264. *In Cotton, A College of Agriculture Report, Series P-108. University of Arizona, Tucson.*
4680. Yee, W.L., Hendrix, D.L., Toscano, N.C., Chu, C.C. and Henneberry, T.J. 1995. Honeydew production by *Bemisia argentifolii* on cotton and its relationship to leaf water potentials and temperature. pp. 870-873. *In D.A. Richter and J. Armour (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
4681. Yee, W.L., Hendrix, D.L., Toscano, N.C., Chu, C.C. and Henneberry, T.J. 1996. Diurnal field patterns of honeydew sugar secretion by *Bemisia argentifolii* (Homoptera: Aleyrodidae) nymphs on cotton. *Environ. Entomol.* 25(4): 776-782.
4682. Yee, W.L. and Toscano, N.C. 1996. Ovipositional preference and development of *Bemisia argentifolii* (Homoptera: Aleyrodidae) in relation to alfalfa. *J. Econ. Entomol.* 89(4): 870-876.
4683. Yee, W.L., Toscano, N.C., Chu, C.C., Henneberry, T.J. and Nichols, R.L. 1996. *Bemisia argentifolii* (Homoptera: Aleyrodidae) action thresholds and cotton photosynthesis. *Environ. Entomol.* 25(6): 1267-1273.
4684. Yee, W.L., Toscano, N.C., Chu, C.C., Henneberry, T.J. and Nichols, R.L. 1996. Photosynthesis and stomatal conductance of cotton infested by different whitefly threshold levels. pp. 1011-1017. *In P. Dugger and D.A. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
4685. Yee, W.L., Toscano, N.C., Hendrix, D.L. and Henneberry, T.J. 1997. Effects of insecticide applications on *Bemisia argentifolii* (Homoptera: Aleyrodidae) densities, honeydew production, and cotton yields. pp. 907-915. *In P. Dugger and D. Richter (Eds.), Proceedings Beltwide Cotton Conferences. National Cotton Council, Memphis, TN.*
4686. Yee, W.L., Toscano, N.C., Hendrix, D.L. and Henneberry, T.J. 1998. Effects of insecticide applications on *Bemisia argentifolii* (Homoptera: Aleyrodidae) densities and honeydew production. *Environ. Entomol.* 27(1): 22-32.
4687. Yee, W.L., Toscano, N.C., Palumbo, J.C., Blua, M.J. and Yoshida, H.A. 1997. Seasonal population trends of *Bemisia argentifolii* (Homoptera: Aleyrodidae) on alfalfa in southern California and Arizona. *Environ. Entomol.* 26(2): 241-249.
4688. Yein, B.R. 1981. Relative efficacy of certain insecticides against cotton pests. *J. Res. (Assam Agric. Univ.)* 2(2): 196-201. [Cock (1986)]
4689. Yein, B.R. 1983. Efficacy of certain insecticides against *Bemisia tabaci* (Genn.) and *Pagria signata* (Motsch.) on blackgram. *J. Res. (Assam Agric. Univ.)* 4(1): 45-49. [Cock (1993)]
4690. Yein, B.R. 1983. Relative susceptibility of some cotton cultivars to insect pests. *J. Res. (Assam Agric. Univ.)* 4(2): 141-147. [Cock (1993)]
4691. Yein, B.R. and Singh, H. 1982. Effect of pesticides and fertilizers on the population of whitefly and incidence of yellow-mosaic virus in greengram. *Indian J. Agric. Sci.* 52: 852-855. [Cock (1986)]

4692. Yen, D.F. and Tsai, Y.T. 1969. Entomogenous fungi of citrus Homoptera in Taiwan. *Plant Prot. Bull. (Taiwan)* 11(1): 1-10. [Cock (1986)]
4693. Yepiz-Plascencia, G.M., Vallejo-Cohen, S. and Valenzuela-Cornejo, P. 1998. Distribution of silverleaf whitefly, *Bemisia argentifolii*, Bellows and Perring (Homoptera: Aleyrodidae) in Sonora, Mexico. *Southwest. Entomol.* 23(1): 83-88.
4694. Yigit, A. 1992. Method for culturing *Serangium parcesetosum* Sicard (Coleoptera: Coccinellidae) on *Bemisia tabaci* Genn. (Homoptera: Aleyrodidae). *Zeitschrift für pflanzenkrankheiten und Pflanzenschutz* 99(5): 525-527. [English, German summary]
4695. Yokomi, R.K., Hoelmer, K.A. and Osborne, L.S. 1990. Relationships between the sweetpotato whitefly and the squash silverleaf disorder. *Phytopathology* 80(10): 895-900. [Cock (1993)]
4696. Yokomi, R.K., Jimenez, D.R., Osborne, L.S. and Shapiro, J.P. 1995. Comparison of silverleaf whitefly-induced and chlormequat chloride-induced leaf silencing in *Cucurbita pepo*. *Plant Dis.* 79(9): 950-955.
4697. Yoldas, Z. 1995. Hiyar seralarinda zarali *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae)'ye karsi biyolojik savasta *Encarsia formosa* (Gahan) (Hymenoptera, Aleyrodidae) nin etkinligi uzerinde bir arastirma. [Investigations on the effectiveness of *Encarsia formosa* (Gahan) (Hymenoptera, Aphelinidae) in the biological control of *Bemisia tabaci* (Genn.) (Homoptera, Aleyrodidae) in cucumbers in the greenhouse]. *Türkiye Entomol. Dergisi* 19(2): 95-100. [Turkish, English summary]
4698. Yoldas, Z. and Koclu, T. 1994. Study on the biology of *Encarsia formosa* (Gahan) (Hymenoptera: Aphelinidae) on cotton whitefly *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae). pp. 381-385. In *Türkiye III. Biyolojik Mucadele Kongresi Bildirileri*, Oct 25-28, 1994, Ege Universitesi Ziraat Fakultesi, Bitki Koruma Bolumu, Izmir, Bornova, Turkey. [Turkish, English summary]
4699. Yoldas, Z., Madanlar, N., Gul, A., Onogur, E. and Tuncay, O. 1999. Investigations on integrated control practices in vegetable glasshouses in Izmir. *Acta Hort.* 491: 453-460.
4700. Yoshii, K. 1975. Una nueva enfermedad de la soya (*Glycine max*) en el Valle del Cauca. [A new soybean (*Glycine max*) disease in the Cauca Valley]. *Noticias Fitopatol. (Colombia)* 4(1): 33-41. [Spanish, English summary]
4701. Younes, A.A. 1998. Studies on certain pests associated with some field crops. Ph.D. Dissertation, Monoufeya University, Shebin -El-Kom, Egypt, 276 pp.
4702. Young, B. 1944. Aleyrodidae from Szechwan, I. *Sinensia (Shanghai)* 13: 129-139. [Cock (1986)]
4703. Youngman, R.R., Toscano, N.C., Jones, V.P., Kido, K. and Natwick, E.T. 1986. Correlations of seasonal trap counts of *Bemisia tabaci* (Homoptera: Aleyrodidae) in Southeastern California. *J. Econ. Entomol.* 79(1): 67-70. [Cock (1993)]
4704. Yousaf, M. 1996. A word of caution for the chemical control of cotton whitefly. *Pakistan Entomol.* 18(1/2): 115-116.
4705. Yu, G., Rumei, X., Stouthamer, R. and van Lenteren, J.C. 1999. A multiplex PCR to detect *Encarsia formosa* host-feeding on *Bemisia argentifolii*. *J. Beijing Normal Univ. (Natural Science)* 35(3): 415-418. [Chinese, English summary]
4706. Yucel, S., Pala, H., Ulubilir, A. and Yigit, A. 1995. Effect of *Verticillium lecanii* (Zimm.) Viegas on cotton whitefly, *Bemisia tabaci* Genn. in greenhouses. pp. 211-215. In *S. Baloglu, H. Saygili, S. Cali, A. Erkilic, M. Ozaslan, S. Yucel and N.C. Posluoglu (Eds.), 7th Congress of Phytopathology in Turkey, Adana, 26-29 Sep 1995. Plant Protection Research Institute, Adana, Turkey.* [Turkish, English summary]
4707. Yuki, V.A., Lourenco, A.L., Kuniyuki, H. and Betti, J.A. 1998. Experimental transmission of bean golden mosaic virus by *Bemisia argentifolii* Bellows & Perring. [Portuguese, English summary]. *Anais Soc. Entomol. Brasil* 27(4): 675-678.
4708. Zachrisson, B. and Poveda, J. 1992. Las moscas blancas en Panama. [The whiteflies in Panama]. pp. 64-66. In *L. Hilje and O. Arboleda (Eds.), Las Moscas Blancas (Homoptera: Aleyrodidae) en America Central y el Caribe. CATIE, Turrialba, Costa Rica.* [In Spanish]
4709. Zakay, Y., Navot, N., Zeidan, M., Kedar, N., Rabinowitch, H., Czosnek, H. and Zamir, D. 1991. Screening *Lycopersicon* accessions for resistance to tomato yellow leaf curl virus: presence of viral DNA and symptom development. *Plant Dis.* 75(3): 279-281.
4710. Zaki, F.N. 1998. Efficiency of the entomopathogenic fungus, *Beauveria bassiana* (Bals), against *Aphis craccivora* Koch and *Bemisia tabaci*, Gennadius. *J. Appl. Entomol.* 122(7): 397-399.
4711. Zaki, F.N., El-Shaarawy, M.F. and Farag, N.A. 1999. Release of two predators and two parasitoids to control aphids and whiteflies. *J. Pest Sci.* 72(1): 19-20.
4712. Zalom, F.G., Castane, C. and Gabarra, R. 1995. Selection of some winter-spring vegetable crop hosts by *Bemisia argentifolii* (Homoptera: Aleyrodidae). *J. Econ. Entomol.* 88(1): 70-76.
4713. Zalom, F.G., Castane, C. and Gabarra, R. 1996. Effects of chilling of *Bemisia argentifolii* (Homoptera: Aleyrodidae) infesting cabbage. *J. Entomol. Sci.* 31(1): 39-51.
4714. Zalom, F.G. and Natwick, E.T. 1987. Developmental time of sweetpotato whitefly (Homoptera: Aleyrodidae) in small field cages on cotton plants. [Spanish summary]. *Florida Entomol.* 70: 67-70. [Cock (1993)]
4715. Zalom, F.G., Natwick, E.T. and Toscano, N.C. 1985. Temperature regulation of *Bemisia tabaci* (Homoptera: Aleyrodidae) populations in Imperial Valley cotton. *J. Econ. Entomol.* 78(1): 61-64. [Cock (1993)]
4716. Zaman, M. 1986. Effect of different insecticides on some sucking insect pests of cotton. *Pakistan Cotton* 30(2): 47-54.
4717. Zaman, M. 1987. Abundance and distribution of some sucking pests of five cultivars of jute. *Pakistan J. Agric. Res.* 8(2): 168-175.
4718. Zaman, M. 1988. Incidence of the cotton whitefly, *Bemisia tabaci* (Gen.), at different levels of plants of jute cultivars in Peshawar. *Sarhad J. Agric. (Pakistan)* 4(2): 153-158.
4719. Zaman, M. 1990. Entomophagous insects and mites found in jute fields at Tarnab, Peshawar. *J. Insect Sci.* 3(2): 133-135.
4720. Zaman, M. and Karimullah. 1985. Relative abundance of whitefly, *Bemisia tabaci* (Gen.) on six cultivars of jute in Peshawar. *Pakistan Entomol.* 7(1/2): 71-76.
4721. Zaman, M. and Karimullah. 1987. Evaluation of granular systemic pesticides against the major sucking pests of jute in Peshawar. *Pakistan J. Agric. Res.* 8(1): 61-66. [Cock (1993)]
4722. Zaman, M. and Karimullah. 1987. Relative abundance of white fly and yellow mite on six cultivars of jute in Peshawar. *Pakistan J. Agric. Res.* 8(2): 218-227.
4723. Zamir, D., Ekstein-Michelson, I., Zakay, Y., Navot, N., Zeidan, M., Sarfatti, M., Eshed, Y., Harel, E., Pleban, T., van Oss, H., Kedar, N., Rabinowitch, H.D. and Czosnek, H. 1994. Mapping and introgression of a tomato yellow leaf curl virus tolerance gene, TY-1. *Theor. Appl. Genet.* 88(2): 141-146.
4724. Zamora, M.E. 1996. Identificación de plantas silvestres como reservorios de los virus del mosaico dorado del frijol (BGMV) y del mosaico enano del frijol (BDMV), en el Valle de Pueblo Nuevo, Nicaragua. [Identification of wild plants as possible hosts of bean golden mosaic virus (BGMV) and bean dwarf mosaic virus (BDMV) in the Pueblo Nuevo Valley, Nicaragua]. *M. Sc. Thesis. CATIE, Turrialba, Costa Rica*, 83 pp.
4725. Zeidan, M. and Czosnek, H. 1991. Acquisition of tomato yellow leaf curl virus by the whitefly *Bemisia tabaci*. *J. Gen. Virol.* 72(11): 2607-2614. [Cock (1993)]
4726. Zeidan, M. and Czosnek, H. 1994. Acquisition and transmission of *Agrobacterium* by the whitefly *Bemisia tabaci*. *Mol. Plant-Microbe Interactions* 7(6): 792-798.



- 4727 Zeidan, M., Green, S.K., Maxwell, D.P., Nakhla, M.K. and Czosnek, H. 1998. Molecular analysis of whitefly-transmitted tomato geminiviruses from Southeast and East Asia. *Trop. Agric. Res. Ext.* 1(2): 107-115.
- 4728 Zhang, X.S., Holt, J. and Colvin, J. 2000. A general model of plant-virus disease infection incorporating vector aggregation. *Plant Pathol.* 49(4): 435-444.
- 4729 Zheng, B.Z. and Goa, X.W. 1995. Monitoring insecticide resistance in greenhouse whitefly adults in Beijing, China, 1991 to 1995. *Resistant Pest Manage.* 7(2): 23-24.
- 4730 Zhou, X., Liu, Y., Calvert, L., Munoz, C., Otim-Nape, G.W., Robinson, D.J. and Harrison, B.D. 1997. Evidence that DNA-A of a geminivirus associated with severe cassava mosaic disease in Uganda has arisen by interspecific recombination. *J. Gen. Virol.* 78: 2101-2111.
- 4731 Zhou, X.P., Xie, Y. and Zhang, Z.K. 2001. Molecular characterization of a distinct begomovirus infecting tobacco in Yunnan, China. *Arch. Virol.* 146(8): 1599-1606.
- 4732 Zhou, X.P., Xie, Y., Zhang, Z.K., Qi, Y.J. and Wu, J.J. 2001. Molecular characterization of a novel defective DNA isolated from tobacco tissues infected with tobacco leaf curl virus. *Acta Virol.* 45(1): 45-50.
- 4733 Zidan, Z.H., Afifi, F.A., Moawad, A.G., Wahed, M.S.A. and Emam, A.K. 1993. Relative susceptibility of tomato varieties by whitefly, aphid, and TYLCV virus at Fayoum Governorate. *Arab Universities J. Agric. Sci.* 1(2): 191-200. [Arabic, English Summary]
- 4734 Zidan, Z.H., Afifi, F.A., Sobeiha, A.K., El-Hamaky, M.A. and Moawad, A.G. 1994. Role of pesticides in controlling tomato pests in normal and sustainable agriculture in Fayoum Governorate. *Arab Universities J. Agric. Sci.* 2(1): 165-178.
- 4735 Zidan, Z.H., Afifi, F.A., Sobeiha, A.K., Moawad, A.G. and El-Malky, K. 1994. Efficiency of juvenile hormone analogues pyriproxyfen, fenocarb and their binary mixtures against whitefly, virus infestation and tomato production. *Arab Universities J. Agric. Sci.* 2(1): 179-185. [Arabic, English summary]
- 4736 Zidan, Z.H., Sobeiha, A.K., Dahroug, S.M.A., Abdel-Moaty, M. and Emam, A. 1994. Bio-residual activity of certain insecticides against the population density of whitefly, *Bemisia tabaci*, infesting cucumber plants in plastic houses. *Ain-Shams Univ. Ann. Agric. Sci. (Egypt)* 39(2): 815-821.
- 4737 Ziegler, A., Mayo, M.A. and Torrance, L. 1998. Synthetic antigen from a peptide library can be an effective positive control in immunoassays for the detection and identification of two geminiviruses. *Phytopathology* 88(12): 1302-1305.
- 4738 Zil'bermint, I.V., Abramova, T.L. and Yakovleva, I.N. 1984. Inhibiting resistance development in greenhouse whitefly by alternating the insecticides. *Khimiya v Sel'skom Khozyaistve* 22(7): 29-32. [Cock (1986), Russian]
- 4739 Zilberstein, A., Navot, N., Ovadia, S., Reinhartz, A., Herzberg, M. and Czosnek, H. 1989. Field-usable assay for diagnosis of the tomato yellow leaf curl virus in squashes of plants and insects by hybridization with a chromogenic DNA probe. *Technique, J. Meth. Cell Mol. Biol.* 1(2): 118-124. [Cock (1993)]
- 4740 Zipori, I., Berlinger, M.J., Dayan, E., Dahan, R., Shmuel, D., Mordechi, S. and Aharon, Y. 1988. [Integrated control of *Bemisia tabaci* in greenhouse tomatoes planted early in the season. *Hassadeh* 68(9): 1711-1713. [Cock (1993), Hebrew, English summary]
- 4741 Zolnerowich, G. and Rose, M. 1998. *Eretmocerus Haldehan* (Hymenoptera: Aphelinidae) imported and released in the United States for control of *Bemisia (tabaci) complex* (Homoptera: Aleyrodidae). *Proc. Entomol. Soc. Washington* 100(2): 310-323.
- 4742 Zouba, A. 1995. A geminivirus causing tomato leaf curl disease in the Sultanate of Oman. *Indian J. Mycol. Plant Pathol.* 25(3): 302-303.
- 4743 Zouba, A., Azam, K.M., Razvi, S.A. and Ai-Wahaibi, A.K. 1993. Temporal increase of tomato leaf curl virus on staggered plantings of tomato in the Sultanate of Oman. *South Indian Hortic.* 41(1): 28-32.
- 4744 Zouba, A.A., Lopez, M.V. and Anger, H. 1998. Squash yellow leaf curl virus: a new whitefly-transmitted poty-like virus. *Plant Dis.* 82(5): 475-478.
- 4745 Zubiatur, Y.M., De Blas, C., Quinones, M., Castellanos, C., Peralta, E.L. and Romero, J. 1998. Havana tomato virus, a new bipartite geminivirus infecting tomatoes in Cuba. *Arch. Virol.* 143(9): 1757-1772.

## ABSTRACTS

1. Abdel-Megeed, M., Zidan, Z.H., Abdel-Wahed, M., Hegazy, G. and Subiha, A. 1994. Ecological studies on whitefly, *Bemisia tabaci*, on cucumber cultivated in plastic houses in Egypt. p. 115. *In B. Ezzahiri and M. Bouhache (Eds.), Fifth Arab Congress of Plant Protection, November 27 - December 2, 1994, Fez, Morocco. Actes Editions, Rabat, Maroc. [Abstract]*
2. Abou-Jawdah, Y., Soubra, K.H., Maalouf, R. and Shebaro, W.A. 1996. Reaction of tomato genotypes to infection by tomato yellow leaf curl geminivirus in Lebanon. *Phytopathology* 86(11 SUPPL.): S71. [Abstract]
3. Abouzid, A.M. and Hiebert, E. 1994. A comparison of partial sequences from selected geminiviruses naturally infecting weeds and crops in Florida. p. 36. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]*
4. Abouzid, A.M., Polston, J.E. and Hiebert, E. 1995. Adaptability of *Sida* mosaic geminivirus during infection in new hosts. *Phytopathology* 85(10): 1139. [Abstract]
5. Adams, C. and Rice, R.P., Jr. 2001. Evaluation of pesticidal control of whitefly on greenhouse poinsettias. *Hortscience* 36(3): 515. [Abstract]
6. Aharonson, N. 1989. Agricultural and environmental aspects of the use of aldicarb for the control of *Bemisia tabaci* in cotton. *Phytoparasitica* 17: 230. [Abstract]
7. Aharonson, N. 1989. Agricultural and environmental aspects of the use of aldicarb for the control of *Bemisia tabaci* on cotton. *Phytoparasitica* 17(3): 230. [Abstract]
8. Ahmad, M. and Ahmad, Z. 1996. Development of insecticide resistance in whitefly and its management in Pakistan. pp. 50-51. *In M.J. Iqbal and A. Kasana (Eds.), Second International Congress of Entomological Sciences, March 19-21, 1996 Islamabad, Pakistan. [Abstract]*
9. Ahmadi, A.A. and Al-Mansoor, H. 1995. The potential of *Eretmocerus mundus* and *Encarsia lutea* as parasitoids of the Cotton Whitefly, *Bemisia tabaci* (Gennadii) in Fars province. p. 103. *In Proceedings 12th Iranian Plant Protection Congress, 2-7 September 1995, Karadj, Iran. [Persian, Abstract]*
10. Ahmed, K.M. 1985. Symptoms and vector of mungbean yellow mosaic virus in Bangladesh. p. 7. *In National Plant Pathology Conference, Joydebpur, Bangladesh, 13-14 April 1985. Bangladesh Agricultural Research Institute, Joydebpur. [Abstract]*
11. Akey, D.H., Chortyk, O.T., Pittarelli, G.W., Stevenson, M.G. and Henneberry, T.J. 1997. Natural and sucrose esters as biorational insecticides in field trials against the silverleaf whitefly *Bemisia argentifolii*. p. 130. *In Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]*
12. Akey, D.H., Chortyk, O.T., Stephenson, M.G. and Henneberry, T.J. 1996. Sucrose esters as biorational insecticides in field trials against silverleaf whitefly *Bemisia argentifolii*. p. 57. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]*

13. Akey, D.H., Chortyk, O.T., Stevenson, M.G. and Henneberry, T.J. 1995. *Nicotiana glauca* extract activity against silverleaf in small plot trials. p. 55. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
14. Akey, D.H. and Chu, C.C. 1999. Acetamiprid as NI 25 against the silverleaf whitefly, *Bemisia argentifolii*, in field trials in upland cotton in Arizona. p. 48. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
15. Akey, D.H., Chu, C.C. and Henneberry, T.J. 1998. NI-25(Rhone-Poulenc EXP 80667A) - an experimental systemic insecticide for silverleaf whitefly control on cotton. p. 36. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
16. Akey, D.H., Dennehey, T.J. and Henneberry, T.J. 1995. Insecticide rotation against SLWF for IPM programs in cotton. p. 56. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
17. Akey, D.H., Ellsworth, P.C., Dennehey, T.J. and Henneberry, T.J. 1997. Insecticide resistance management: strategies of using IGRs vs pyrethroids for initial applications against silverleaf whitefly. p. 109. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
18. Akey, D.H., Goodwin, J. and Kirk, I.W. 1996. Comparison of day versus night applications for control of silverleaf whitefly and pink bollworm. p. 54. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
19. Akey, D.H. and Henneberry, T.J. 1994. Plot trials with amitraz for chemical control of SPWF in cotton. p. 75. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
20. Akey, D.H. and Henneberry, T.J. 1994. Plot trials with buprofezin for chemical control of SPWF in cotton. p. 74. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
21. Akey, D.H. and Henneberry, T.J. 1994. Small plot trial with candidates for chemical control of SPWF in cotton. p. 73. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
22. Akey, D.H. and Henneberry, T.J. 1994. Use of hydraulic sprayers for ground control of SPWF in cotton. p. 76. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
23. Akey, D.H. and Henneberry, T.J. 1995. Azadirachtin (Azatin) and KDF-0520 (Agridyne Technologies Inc.) activity against silverleaf whitefly in small plot trials. p. 57. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
24. Akey, D.H. and Henneberry, T.J. 1995. Buprofezin activity against silverleaf whitefly in small plot trials. p. 58. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
25. Akey, D.H. and Henneberry, T.J. 1995. Comparisons of buprofezin formulations against silverleaf whitefly in medium plot trials. p. 59. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
26. Akey, D.H. and Henneberry, T.J. 1995. Interaction with bifenthrin and fenoxycarb for chemical control of silverleaf whitefly in cotton. p. 60. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
27. Akey, D.H. and Henneberry, T.J. 1995. Pymetrozine (CGA 215'944, Ciba) activity against silverleaf whitefly in small plot trials. p. 61. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
28. Akey, D.H. and Henneberry, T.J. 1995. Pyriproxyfen activity against silverleaf whitefly in small plot trials. p. 62. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
29. Akey, D.H. and Henneberry, T.J. 1995. Use of a 400 psi hydraulic sprayer for ground control of SPWF in cotton. p. 63. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
30. Akey, D.H. and Henneberry, T.J. 1996. Evaluation of hot water treatment for control of silverleaf whitefly nymphs on upland cotton in a laboratory and field trial in Arizona. p. 55. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
31. Akey, D.H. and Henneberry, T.J. 1996. Fexoxycarb, Pymetrozine (C G A-215944), and Fenpropathrin/Acephate: rotational use studies for silverleaf whitefly control. p. 56. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
32. Akey, D.H. and Henneberry, T.J. 1998. Use of a sugar ester, produced by AVA Chemical Ventures, as a biorational agent against the silverleaf whitefly, *Bemisia argentifolii*, in field trials in upland cotton. p. 38. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
33. Akey, D.H. and Henneberry, T.J. 1998. Use of entomopathogenic fungi, *Beauveria bassiana* and *Paecilomyces fumosoroseus*, as biorational agents against the silverleaf whitefly, *Bemisia argentifolii*, in field trials in upland cotton. p. 54. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
34. Akey, D.H. and Henneberry, T.J. 1998. Use of the azadirachtin product, Bollwhip, as a biorational agent against the silverleaf whitefly, *Bemisia argentifolii*, in field trials in upland cotton. p. 37. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
35. Akey, D.H. and Henneberry, T.J. 1999. Azadirachtin (as Bollwhip R), a biorational agent against the silverleaf whitefly, *Bemisia argentifolii*, in field trials in Upland cotton in Arizona. p. 47. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
36. Akey, D.H. and Henneberry, T.J. 1999. Progress in development of IPM for upland cotton in Arizona using biorational and biopesticide agents for control of silverleaf whitefly (SLWF) *Bemisia argentifolii* and other cotton pests. p. 115. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]

37. Akey, D.H. and Henneberry, T.J. 1999. A sugar ester (AVA Chemical Ventures, L.L.C.) as a biorational agent against the silverleaf whitefly, *Bemisia argentifolii*, in field trials in upland cotton in Arizona. p. 46. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
38. Akey, D.H. and Henneberry, T.J. 1999. Use of the entomopathogenic fungi, *Beauveria bassiana* and *Paecilomyces fumosoroseus* as biorational agents against the silverleaf whitefly (SLWF), *Bemisia argentifolii*, in field trials in upland cotton. p. 68. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
39. Akey, D.H. and Henneberry, T.J. 2000. Azadirachtin (as Bollwhip), a biorational agent against the silverleaf whitefly, *Bemisia argentifolii*, in field trials in upland cotton in Arizona. p. 60. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
40. Akey, D.H. and Henneberry, T.J. 2000. Effect on beneficial arthropods of biorationals (insect growth regulators and entomopathogenic fungi) used for control of silverleaf whitefly, *Bemisia argentifolii*, in upland cotton in Arizona. p. 80. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
41. Akey, D.H. and Henneberry, T.J. 2000. Effect on *Lygus* of biorationals (insect growth regulators and entomopathogenic fungi) used for control of the whitefly *Bemisia argentifolii*, in field trials in upland cotton in Arizona. p. 61. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
42. Akey, D.H. and Henneberry, T.J. 2000. Progress in development of IPM for upland cotton in Arizona using biorational and biopesticide agents for control of silverleaf whitefly (SLWF) *Bemisia argentifolii* and other pests. p. 134. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
43. Akey, D.H. and Henneberry, T.J. 2000. Use of the entomopathogenic fungi, *Beauveria bassiana*, and *Paecilomyces fumosoroseus* as biorational agents against the silverleaf whitefly, *Bemisia argentifolii*, in field trials in upland cotton. p. 81. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
44. Akey, D.H., Henneberry, T.J. and Chu, C.C. 1993. [chemical control, cotton, large plots]. p. 53. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
45. Akey, D.H., Henneberry, T.J. and Chu, C.C. 1993. [chemical control, cotton, small plots]. p. 53. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
46. Akey, D.H., Henneberry, T.J. and Chu, C.C. 1999. Ground application techniques that improved under-leaf and inner canopy coverage cotton: development and testing of hydrostatic sprayers-transition from experimental to farm equipment. p. 45. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
47. Akey, D.H., Henneberry, T.J. and Hernández, M. 1993. [movement into cotton from cantaloupe]. p. 7. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
48. Akey, D.H., Henneberry, T.J. and Wuertz, D.A. 1994. Use of the fungus, *Beauveria bassiana*, as *Naturalis L* against the sweetpotato whitefly (SPWF), *Bemisia tabaci* in furrow and sub-drip irrigated cotton. p. 121. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
49. Akey, D.H., Henneberry, T.J. and Wuertz, D.A. 1994. Whole season rotational pesticide system for control of sweetpotato whitefly in cotton. p. 77. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
50. Akey, D.H., Neal, J.W., Severson, R., Stephenson, M., Pittarelli, G. and Buta, G. 1993. [chemical control with *Nicotiana glauca* extract]. p. 54. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
51. Akey, D.H., Wright, J.E., Palumbo, J. and Tollefson, S. 1993. Distribution of the fungus, *Beauveria bassiana*, as the product *Naturalis L* against the sweetpotato whitefly (SPW), *Bemisia tabaci*. p. 92. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
52. Al-Ezabi, F.A. 1994. Polypropylene covers as a new technology to control tomato yellow leaf curl virus. p. 120. *In* B. Ezzahiri and M. Bouhache (Eds.), Fifth Arab Congress of Plant Protection, November 27 - December 2, 1994, Fez, Morocco. Actes Editions, Rabat, Maroc. [Abstract]
53. Al-Mansoor, H. and Ahmadi, A.A. 1995. The evaluation of parasitism capacity of *Eretmocerus mundus* Mercet on two cotton cultivars (hairy and hairless). p. 104. *In* Proceedings 12th Iranian Plant Protection Congress, 2-7 September 1995, Karadj, Iran. [Abstract]
54. Al-Saggaf, S.M. 1994. Influence of different methods of tomato seedling protection on subsequent infection with tomato leaf-curl virus disease. p. 195. *In* B. Ezzahiri and M. Bouhache (Eds.), Fifth Arab Congress of Plant Protection, November 27 - December 2, 1994, Fez, Morocco. Actes Editions, Rabat, Maroc. [Abstract]
55. Aldana, L.F., Masaya, P. and Yoshii, K. 1981. La tolerancia al mosaico dorado del frijol y el combate quimico del vector (*Bemisia tabaci*) como medico de control. Memoria de la XXVII Reunion Anual PCCMCA. Santo Domingo 3: 19. [Abstract]
56. Allen, J.C., Brewster, C.C., Paris, J.F., Summers, C.G. and Riley, D.G. 1994. Spatio-temporal modelling of silverleaf whitefly dynamics in a regional cropping system using satellite data. *Phytoparasitica* 22(4): 316. [Abstract]
57. Allen, J.C., Byrne, D.N., Paris, J.F. and Riley, D.G. 1997. Large scale dynamics of the silverleaf whitefly in the Imperial Valley California. p. 191. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
58. Allen, J.C., Carruthers, R.I., Naranjo, S.E. and Wagner, T. 1993. Spatial and temporal modeling of sweetpotato whitefly population dynamics. p. 8. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
59. Allen, J.C., Fasulo, T.R., Schuster, D.J., Stansly, P.A., Byrne, D., Paris, J.F., Perring, T.M., Riley, D.G. and Summers, C.G. 1994. Modeling the movement and reproduction of the sweetpotato whitefly. p. 175. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]

60. Allen, J.C., Fasulo, T.R., Schuster, D.J., Stansly, P.A., Byrne, D., Paris, J.F., Perring, T.M., Riley, D.G. and Summers, C.G. 1995. Large scale cropping patterns in relation to reproduction and movement of silverleaf whitefly. p. 161. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
61. Allen, J.C., Stansly, P.A., Schuster, D.J., Riley, D.G. and Perring, T.M. 1993. [computer display of data and simulation of movement]. p. 135. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
62. Anderson, P.K. 1999. Use of a mathematical model as an analytical tool to prioritize IPM research and interventions for whitefly - transmitted geminiviruses in Latin America. p. 97. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce (Almeria), Spain. [Abstract]
63. Anderson, P.K. 2000. The CGIAR systemwide IPM project on sustainable integrated management of whiteflies as pests and vectors of plant viruses in the tropics. p. 135. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
64. Andress, E., Gould, J. and Quinn, M.A. 2000. Assessing the impact of established whitefly parasitoids in the Imperial Valley using multivariate techniques. p. 82. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
65. Ansolabehere, M.J. 1996. Silverleaf whitefly control in cotton with KNACK insect growth regulator. p. 58. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
66. Ansolabehere, M.J., Chernicky, J.P. and West, S. 1995. SLWF control in cotton with pyriproxyfen, an IGR. p. 64. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
67. Ansolabehere, M.J. and De Witt, T.C. 1994. Pyriproxyfen (S-71639), an IGR for whitefly control. p. 78. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
68. Ansolabehere, M.J., Lindemann, R.H. and Welch, L.L. 1994. Fenprothrin plus acephate for whitefly control. p. 79. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
69. Anthony, N., Brown, J., Markham, P. and French-Constant, R. 1995. Molecular analysis of cyclodiene resistance associated mutations among populations of the sweetpotato whitefly *Bemisia tabaci*. p. 65. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
70. Anthony, N.M., Brown, J.K., Markham, P.G. and French-Constant, R.H. 1994. Cyclodiene insecticide resistance in strains of *Bemisia tabaci* is correlated with presence in crop systems and not with whitefly biotype. *Phytoparasitica* 22(4): 347. [Abstract]
71. Antignus, Y., Adler, O., Perlman, M., Ben-Joseph, R. and Cohen, S. 1994. Association of the nucleic acid and coat protein of tomato yellow leaf curl virus with the whitefly vector *Bemisia tabaci*. *Phytoparasitica* 22(4): 326. [Abstract]
72. Antignus, Y., Czosnek, H., Cohen, S., Ben, R., Ben-Joseph, R., Navot, N. and Zamir, D. 1987. Isolation and partial characterization of tomato yellow leaf curl, a whitefly-transmitted geminivirus. *Phytoparasitica* 15: 262. [Abstract]
73. Antignus, Y., Lapidot, M. and Cohen, S. 1999. Optical barriers: an innovative IPM tool for the control of insect pests and virus diseases in protected crops. p. 151. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce(Almeria), Spain. [Abstract]
74. Antignus, Y., Perlman, M., Ben-Joseph, R. and Cohen, S. 1993. The interaction of tomato yellow leaf curl virus with its whitefly vector, *Bemisia tabaci*. *Phytoparasitica* 21(2): 174-175. [Abstract]
75. Arnal, E., Ramos, F. and Notz, A. 1997. Aspectos fitosanitarios y economicos del proyecto mosca blanca en tomate periodo 1996-1997 en Guanayen, estado Aragua. pp. 63-64. *In* A. Arcia and A. Romero (Eds.), Sociedad Venezolana De Ingenieros Agronomos, Maracay (Venezuela). [Spanish, Abstract]
76. Arnó, J. and Gabarra, R. 1994. Potential for biological control of mixed *Trialetrodes vaporariorum* and *Bemisia tabaci* populations in winter tomato crops grown in greenhouses. *Phytoparasitica* 22(4): 340-341. [Abstract]
77. Arsénio, A.F., Neto, E., Ramos, N., Mangerico, S., Fernandes, J.E., Lavadinho, A.M.P., Lopes, A., Guimaraes, J.M. and Louro, D. 1999. The use of nets and insecticide treatments in the control on *Bemisia tabaci* Genn. populations and tomato yellow leaf curl virus spread in protected tomato crops in Portugal. p. 186. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce(Almeria), Spain. [Abstract]
78. Ausher, R. 1994. Implementation of integrated pest management programs in Israel. *Phytoparasitica* 22(4): 353. [Abstract]
79. Bahamish, H. and Al-Segaf, S.M. 1994. Effect of leaf curl virus disease on tomato varieties in Yemen. p. 184. *In* B. Ezzahiri and M. Bouhache (Eds.), Fifth Arab Congress of Plant Protection, November 27 - December 2, 1994, Fez, Morocco. Actes Editions, Rabat, Maroc. [Abstract]
80. Ball, J.C. and Weddle, D. 1993. [SPW and parasitoids: population followed on hibiscus, orchid tree, snail vine]. p. 93. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
81. Bar, H. and Weinberg, A. 1993. Methods of testing and preventing cotton stickiness. *Phytoparasitica* 21(2): 173-174. [Abstract]
82. Bar, H. and Weinberg, A. 1994. A new approach to assessment of the honeydew content of a cotton crop. *Phytoparasitica* 22(4): 320-321. [Abstract]
83. Barash, I., Mor, H., Gindin, G., Raccach, B. and Ben-Ze'ev, I. 1994. Selection and possible genetic manipulation of entomopathogenic fungi for biocontrol. *Phytoparasitica* 22(4): 343-344. [Abstract]
84. Barten, J.H.M., Thome, C.H., Stevens, M.R., Schuster, D.J., Scott, J.W. and Chambliss, O.L. 1994. Evaluating resistance in tomato to the silverleaf whitefly, *Bemisia argentifolii*. *Phytoparasitica* 22(4): 330-331. [Abstract]
85. Bartlett, A.C. and Lewis, L.J. 1996. Genetic analysis of whitefly populations. p. 16. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
86. Beehler, L.L., Toscano, N.C. and Coates, W. 1995. Evaluation of insecticide application equipment for spray deposition and efficacy against *Bemisia argentifolii* on tomatoes. p. 66. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
87. Beehler, L.L., Toscano, N.C., Coates, W. and Ballmer, G.R. 1994. Evaluation of insecticide application equipment for spray deposition and efficacy against *Bemisia tabaci* on tomatoes. p. 80. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
88. Bell, M.L., Bailey, D.A. and Baker, J.R. 1996. Application technique influences imidacloprid efficacy on poinsettia. *Hortscience* 31(5): 752. [Abstract]

89. Bell, M.L., Baker, J.D. and Bailey, D.A. 1996. Efficacy against silverleaf whitefly of insecticidal dips for rooted poinsettia cuttings. *Hortscience* 31(4): 584-585. [ Abstract]
90. Bell, M.L. and Baker, J.R. 1997. Greenhouse screens and their efficacy against whiteflies and thrips. *Hortscience* 32(3): 479. [ Abstract]
91. Bellows, T.S., Jr., Perring, T.M., Gill, R.J. and Headrick, D.H. 1994. Description of a species of *Bemisia*. p. 37. In *Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan*. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
92. Bellows, T.S., Jr., Perring, T.M. and Headrick, D.H. 1994. Parasitism of silverleaf whitefly in California crops and weeds. p. 122. In *Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan*. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
93. Beltran, R.J.F., Gomez, C.O., Brown, J.K. and Lambe, R.C. 1988. Perforated floating row cover insect barrier to control two plant viruses of squash. *Biological and Cultural Tests for Control of Plant Diseases*. Am. Phytopathol. Soc. 3: 26. [ Abstract]
94. Ben-Ze'ev, I.S., Gindin, G., Barash, I. and Raccach, B. 1994. Identification of entomopathogenic fungi attacking *Bemisia tabaci* in Israel. *Phytoparasitica* 22(4): 344. [ Abstract]
95. Benmessaoud, H., Sayoud, H.H. and Keroui, W. 1994. Experiments in biocontrol of *Bemisia tabaci* (Homoptera Aleyrodidae) using *Encarsia formosa* (Hymenoptera Aphelinidae). p. 165. In B. Ezzahiri and M. Bouhache (Eds.), *Fifth Arab Congress of Plant Protection*, November 27 - December 2, 1994, Fez, Morocco. Actes Editions, Rabat, Maroc. [Abstract]
96. Berlinger, M.J. 1984. Host plant resistance to *Bemisia tabaci*. p. 569. In *XVII Int. Congress Entomol.* [Abstract]
97. Berlinger, M.J., Dahan, D. and Urkin -Shevach, E. 1983. The effect of light on the resistance of wild species of Solanaceae to *Bemisia tabaci*. *Phytoparasitica* 11: 63. [ Abstract]
98. Berlinger, M.J. and Dahan, R. 1982. Control of *Bemisia tabaci*, the vector of tomato yellow leaf curl virus, in glasshouse tomatoes. *Phytoparasitica* 10: 297. [ Abstract]
99. Berlinger, M.J. and Dahan, R. 1983. Integrated control of the tobacco whitefly *Bemisia tabaci* in greenhouse tomatoes in Israel. p. 1109. In *10th International Congress of Plant Protection; Proceedings of a Conference Held at Brighton, England 20-25 November 1983*. Plant Protection for Human Welfare. British Crop Protection Council, Croydon, UK. [Abstract, Cock (1986)]
100. Berlinger, M.J. and Dahan, R. 1986. Flight patterns of *Bemisia tabaci*, a vector of plant viruses. p. 23. In *Proc. International Conference on Tropical Entomology*, 31 August- 5 September 1986, Nairobi, Kenya. [Abstract]
101. Berlinger, M.J., Dahan, R. and Cohen, S. 1984. Phenology of the tobacco whitefly, *Bemisia tabaci*, in glasshouse tomatoes in Israel. *Phytoparasitica* 2: 141. [ Abstract]
102. Berlinger, M.J., Dahan, R. and Mordechi, S. 1986. Breeding tomatoes resistant to the transmission of tomato yellow leaf curl virus by *Bemisia tabaci*. *Phytoparasitica* 14: 158-159. [ Abstract]
103. Berlinger, M.J., Dahan, R. and Mordechi, S. 1987. Has the tobacco whitefly, *Bemisia tabaci*, developed resistance to synthetic pyrethroids? *Phytoparasitica* 15: 264. [ Abstract]
104. Berlinger, M.J., Dahan, R., Mordechi, S. and Oren, R. 1985. Photoperiodism in the tobacco whitefly, *Bemisia tabaci*. *Phytoparasitica* 13: 74. [ Abstract ]
105. Berlinger, M.J. and Lebiush-Mordechi, S. 1994. Physical means for the control of *Bemisia tabaci*. *Phytoparasitica* 22(4): 352. [ Abstract]
106. Berlinger, M.J. and Lebiush-Mordechi, S. 1995. Physical means for the control of *Bemisia tabaci*. p. 146. In *Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan*. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
107. Berlinger, M.J. and Lehmann-Sigura, N. 1986. The ability of *Bemisia tabaci*, the vector of tomato yellow leaf curl virus, to survive climatic conditions. *Phytoparasitica* 14: 152. [ Abstract]
108. Berlinger, M.J., Leibush-Mordechai, S., Mor, N. and Fridja, D. 1993. Insecticide efficiency in controlling the whitefly *Bemisia tabaci* and the tomato yellow leaf curl virus. *Phytoparasitica* 21(2): 176. [ Abstract]
109. Berlinger, M.J. and Nir, L.B. 1989. A population growth model for *Bemisia tabaci*. *Phytoparasitica* 17(3): 228-229. [ Abstract]
110. Bharathan, N., Graves, W.R., Narayanan, K.R., Bryan, H.H. and McMillan, R.T., Jr. 1989. Whitefly-mediated silvering of squash leaves. *Phytopathology* 79: 1213. [ Abstract]
111. Bi, J.L., Ballmer, G.R. and Toscano, N.C. 1999. Effect of nitrogen fertility on cotton-whitefly interactions. p. 96. In *Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan*. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
112. Bindra, O.S. and Rahman, A.A.A. 1983. Cotton integrated-pest-control in the Sudan. p. 937. In *10th International Congress of Plant Protection; Proceedings of a Conference Held at Brighton, England 20-25 November 1983*. Plant Protection for Human Welfare. British Crop Protection Council, Croydon, UK. [Abstract, Cock (1986)]
113. Bird, J. 1962. A whitefly-transmitted mosaic of *Rhynchosia minima* and its relation to tobacco leaf curl and other virus diseases of plants in Puerto Rico. *Phytopathology* 52: 286. [ Abstract, Cock (1986)]
114. Bird, J., Kimura, M., Monllor, A.C., Rodriguez, R.L., Sanchez, J. and Maramorosch, K. 1975. Mosaico de *Euphorbia prunifolia* Jacq. en Puerto Rico; transmision, hospederas y etiologia. [Mosaic of *Euphorbia prunifolia* Jacq. in Puerto Rico; transmission, hosts and ethiology]. pp. 233-234. In *Memoir 21. Annual Meeting of the Central American Cooperative Program for Improving Food Crops*, Santa Tecla, El Salvador, 7 April 1975. [Spanish, Abstract]
115. Bird, J., Sanchez, J. and Vakili, N.G. 1973. Golden yellow mosaic of beans (*Phaseolus vulgaris*) in Puerto Rico. *Phytopathology* 63: 1435. [ Abstract, Cock (1986)]
116. Birdsall, S.L., Ritter, D. and Cason, P.L. 1995. Economic impact of the silverleaf whitefly in Imperial Valley, California. p. 162. In *Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan*. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
117. Birdsall, S.L., Ritter, D. and Cason, P.L. 1996. Economic impact of the silverleaf whitefly in Imperial Valley, California, from 1991 to 1995. p. 176. In *Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan*. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
118. Bivins, D., Ellsworth, P., Brown, P. and Chapman, R. 1996. Low temperature mortality and overwintering ecology of *Bemisia tabaci* (= *Bemisia argentifolii*). p. 1. In *Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan*. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
119. Bivins, D., Ellsworth, P.C., Brown, P. and Chapman, R.F. 1997. Overwintering ecology of *Bemisia*. p. 63. In *Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001)*. U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
120. Black, L.M. 1953. Loss of vector transmissibility by viruses normally insect transmitted. *Phytopathology* 43: 466. [ Abstract]
121. Blackmer, J. and Byrne, D.N. 1994. Host-plant effects on life-history traits and flight behavior of *Bemisia tabaci*. p. 10. In *Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan*. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]

122. Blackmer, J.L. and Byrne, D.N. 1999. Developmental and behavioral effects of dietary constituents on *Bemisia tabaci*. p. 1. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
123. Blackmer, J.L., Davidson, E.W. and Lee, L.L. 2000. Evaluation of artificial diets for rearing *Bemisia tabaci* (Biotype B) (Homoptera: Aleyrodidae). p. 9. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
124. Bloch, G. and Wool, D. 1993. Quantitative genetics of esterase activity and resistance to methidathion in the whitefly *Bemisia tabaci* in Israel. *Phytoparasitica* 21(2): 180. [ Abstract]
125. Blua, M.J., Toscano, N.C., Henneberry, T.J., Ballmer, G.R. and Beehler, L.L. 1994. Urea, aleitte and organic fertilizer effects on silverleaf whitefly populations. p. 81. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
126. Bográn, C.E., Heinz, K.M., Ciomperlik, M. and Wendel, L. 1997. Interspecific interactions among *Bemisia argentifolii* parasitoids. p. 134. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
127. Boissot, N., Lafortune, D., Ano, G., Pavis, C. and Sauvion, N. 1999. Resistance against geminivirus and *Bemisia tabaci* B in tomato and melon [French West Indies]. p. 98. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
128. Boissot, N. and Pavis, C. 1999. Resistance against *Bemisia tabaci* B biotype in melon [French West Indies]. p. 97. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
129. Bouse, L.F., Carlton, J.B., Franz, E. and Kirk, I.W. 1994. Aircraft spray nozzles for minimizing spray drift and optimizing spray deposition on cotton. p. 82. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
130. Bouse, L.F., Carlton, J.B., Franz, E., Kirk, I.W. and Latheef, M.A. 1994. Aerial spray deposition studies for sweetpotato whitefly control in cotton. p. 83. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
131. Bradley, C., Wraight, S., Carruthers, R., Staten, R. and Jaronski, S. 1996. Progress in developing a myco-insecticide for whitefly control. p. 59. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
132. Brazzle, J.R., Fien, B. and Toscano, N. 1998. Silverleaf whitefly in the Southern San Joaquin Valley: an areawide management project in progress. p. 106. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
133. Brazzle, J.R., Heinz, K.M., Parella, M.P. and Pickett, C.H. 1994. Field evaluations of *Delphastus pusillus* for control of *Bemisia tabaci* infesting upland cotton. p. 123. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
134. Brazzle, J.R., Heinz, K.M., Parrella, M.P. and Wrona, A.F. 1995. Pattern analysis of *Bemisia argentifolii* infesting Imperial Valley [CA] cotton. p. 3. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
135. Brazzle, J.R., Toscano, N. and Goodell, P. 1999. Implementing a resistance management program for *Bemisia argentifolii*: Building the necessary bridges. p. 2. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
136. Briddon, R.W. 2000. Diagnosing tomato yellow leaf curl virus. *Phytopathology* 90(6 Suppl.): S9. [ Abstract]
137. Briddon, R.W., Farquhar, J.A., Roussot, C., Banks, G.K., Bedford, I.D., Legg, J. and Markham, P.G. 1999. Geminiviruses and cassava whiteflies across Africa. pp. 75-76. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce (Almeria), Spain. [Abstract]
138. Brower, A.G. and Byrne, D.N. 1995. Population dynamics of whitefly predators occurring naturally in cotton. p. 4. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
139. Brown, J.K. 1994. Detection and identification of whitefly-transmitted geminiviruses by polymerase chain reaction (PCR). p. 38. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
140. Brown, J.K. 1997. A DNA sequence database for Subgroup III Geminiviruses. p. 79. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
141. Brown, J.K. 2000. Biological, genetic, and molecular characterization of begomoviruses from Puerto Rico and the Caribbean Region. *Phytopathology* 90(6 Suppl.): S122. [ Abstract]
142. Brown, J.K. 2001. Molecular tracking of new and emergent begomoviruses and genetic variation of the whitefly vector: *Bemisia tabaci*. *South African J. Sci.* 97(1-2): ii. [ Abstract]
143. Brown, J.K. and Banks, G.K. 1995. Analysis of the capsid protein gene of Subgroup II Geminiviridae. p. 26. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
144. Brown, J.K. and Bird, J. 1994. Introduction of an exotic whitefly (*Bemisia*) vector facilitates secondary spread of jatropha mosaic virus, a geminivirus previously vectored exclusively by the jatropha biotype. *Phytoparasitica* 22(4): 325. [ Abstract]
145. Brown, J.K., Bird, J., Banks, G., Sosa, M., Kiesler, K., Cabrera, I. and Fornaris, G. 1995. First report of an epidemic in tomato caused by two whitefly-transmitted geminiviruses in Puerto Rico. *Plant Dis.* 79(12): 1250. [ Abstract]
146. Brown, J.K., Bird, J., Fletcher, D., Goldberg, N.P. and Coats, S.A. 1993. A new virus disease of passionvine (*Passiflora edulis* var. *flavicarpa*) caused by a whitefly-transmitted geminivirus. *Phytopathology* 83 : 698. [ Abstract]
147. Brown, J.K., Bird, J. and Fletcher, D.C. 1993. First report of passiflora leaf mottle disease caused by a whitefly-transmitted geminivirus in Puerto Rico. *Plant Dis.* 77(12): 1264. [ Abstract]
148. Brown, J.K., Butler, G.D., Jr and Nelson, M.R. 1983. Occurrence of cotton leaf crumple associated with severe whitefly infestations in Arizona. *Phytopathology* 73: 787. [ Abstract]
149. Brown, J.K., Campodonico, O.P. and Nelson, M.R. 1989. A whitefly-transmitted geminivirus from peppers with tigré disease. *Plant Dis.* 73(7): 610. [ Abstract, Cock (1993)]

150. Brown, J.K., Chapman, M.A. and Nelson, M.R. 1990. Bean calico mosaic, a new disease of common bean caused by a whitefly - transmitted geminivirus. *Plant Dis.* 74(1): 81. [ Abstract, Cock (1993)]
151. Brown, J.K., Coats, S., Bedford, I.D., Markham, P.G. and Bird, J. 1992. Biotypic characterization of *Bemisia tabaci* populations based on esterase profiles, DNA fingerprinting, virus transmission, and bioassay to key host plant species. *Phytopathology* 82: 1104. [ Abstract]
152. Brown, J.K., Costa, H.S. and Bird, J. 1991. Variation on *Bemisia tabaci* populations based on geographic origin, silverleaf symptom inductions, and esterase banding patterns.(abstr.). *Phytopathology* 81: 1157. [ Abstract]
153. Brown, J.K., Costa, H.S. and Laemmlen, F. 1992. First report of whitefly-associated squash silverleaf (SSL) of *Cucurbita*, and of white streaking (WSt) disorder of cole crops in Arizona and California. *Plant. Dis.* 76(4): 426. [ Abstract, Cock (1993)]
154. Brown, J.K., Fletcher, D.C., Goldberg, N.P. and Coats, S.A. 1993. [Surveys of 'A' and 'B' biotype , crossing, geminivirus]. p. 26. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
155. Brown, J.K. and Frohlich, D.R. 1995. Mitochondrial 16S ribosomal subunit as a subspecies/species marker for *Bemisia tabaci* (Gennadius): evidence for a species complex. p. 27. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
156. Brown, J.K., Hartitz, M.D., Rosell, R.C. and Bisaro, D.M. 1996. Subgroup III geminivirus coat protein plays a direct role in whitefly-mediated virus transmission. *Phytopathology* 86(11 SUPPL.): S43. [ Abstract]
157. Brown, J.K., Idris, A.M. and Fletcher, D.C. 1993. Sinaloa tomato leaf curl virus, a newly described geminivirus of tomato and pepper in West Coastal Mexico. *Plant Dis.* 77(12): 1262. [ Abstract]
158. Brown, J.K., Idris, A.M., Torres, J.I. and Bird, J. 2000. *Jatropha* mosaic begomovirus variants from weed and cultivated hosts in Puerto Rico. *Phytopathology* 90(6 Suppl.): S122. [ Abstract]
159. Brown, J.K., Idris, A.M., Torres-Jerez, I. and Wyatt, S.D. 1999. The core region of the coat protein gene of viruses of the *Geminiviridae* is phylogenetically informative: Introducing an interactive website for *Begomovirus* identification using the core CP sequence. p. 80. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce(Almeria), Spain. [Abstract]
160. Brown, J.K., Jimenez-Garcia, E. and Nelson, M.R. 1988. Bean calico mosaic virus, a newly described geminivirus of bean. *Phytopathology* 78: 1579. [ Abstract]
161. Brown, J. K., Lastra, R. and Bird, J. 1991. First documentation of whitefly-transmitted gemini viruses causing widespread disease of cotton, tobacco, and tomato in Dominican Republic and in tomato in Puerto Rico. *Fitopatologia* 26: 47. [ Abstract]
162. Brown, J.K., Nelson, M.R. and Lambre, R.C. 1986. Cotton leaf crumple virus transmitted from naturally infected bean from Mexico. *Plant Dis.* 70(10): 981. [ Abstract, Cock (1993)]
163. Brown, J.K. and Poulos, B.T. 1990. Serrano golden mosaic virus a newly identified whitefly-transmitted geminivirus of pepper and tomato in the United States and Mexico. *Plant Dis.* 74(9): 720. [ Abstract, Cock (1993)]
164. Brown, J.K., Rosell, R.C. and Torres-Jerez, I. 1996. Morphological and genetic analysis of the *Bemisia tabaci* species complex. p. 18. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
165. Brown, J.K., Sosa, M. and Bird, J. 1998. Biological and molecular characteristics of tomato-infecting geminiviruses transmitted by the *Bemisia tabaci* (Genn.) species complex in Puerto Rico. *Phytopathology* 88(9 suppl.): S120. [ Abstract]
166. Brown, J.K. and Stanghellini, M.E. 1988. Lettuce infectious yellows virus in hydroponically grown lettuce in Pennsylvania. *Plant Dis.* 72(5): 453. [ Abstract, Cock (1993)]
167. Brown, J.K. and Torres, J.I. 1999. The mitochondria COI gene as an informative molecular marker and diagnostic sequence for identification of *Bemisia tabaci* (Genn.), the whitefly vector of geminiviruses. *Phytopathology* 89(6 Suppl.): S91. [ Abstract]
168. Brown, J.K. and Torres-Jerez, I. 1999. The mitochondria COI gene as an informative molecular marker for phylogenetic analysis and identification of *Bemisia tabaci* (Genn.), the whitefly vector of geminiviruses. p. 79. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce(Almeria), Spain. [Abstract]
169. Brown, J.K., Wyatt, S.D., Banks, G.K., McGrath, P.F., Idris, A.M., Parker, P. and Bird, J. 1996. DNA sequence database for whitefly-transmitted geminiviruses from the U. S., Mexico, and the Caribbean Basin. p. 17. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
170. Brown, J.K., Wyatt, S.D. and Frohlich, D.R. 1994. The capsid protein - a determinant of host plant affiliations of whitefly-transmitted geminiviruses ? *Phytoparasitica* 22(4): 325-326. [ Abstract]
171. Brown, J.K., Wyatt, S.D. and Frohlich, D.R. 1994. The coat protein gene of whitefly-transmitted geminiviruses: preliminary analysis. *Phytopathology* 84: 1132. [ Abstract]
172. Brownbridge, M., Parker, B.L. and Skinner, M. 1993. Fungal pathogens for biocontrol of insects in greenhouses. *HortScience* 28(4): 260. [ Abstract]
173. Buckner, J., Nelson, D., Cohen, A., Freeman, T. and Ruud, R. 1997. Whiteflies on an artificial membrane and their interactions with predators. p. 80. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
174. Buckner, J.S., Freeman, T.P., Chu, C.C., Nelson, D.R. and Henneberry, T.J. 1999. Whitefly egg pedicel morphology and penetration characteristics. p. 99. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
175. Buckner, J.S., Freeman, T.P. and Nelson, D.R. 2000. The physiology of the whitefly egg pedicel. p. 113. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
176. Buckner, J.S. and Nelson, D.R. 1994. Identification and occurrence of the surface lipids and wax particles of the adult whiteflies, *Bemisia tabaci* and *Trialeurodes vaporariorum*. p. 39. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
177. Buckner, J.S., Nelson, D.R. and Mardaus, M.C. 1996. The surface lipids of *Bemisia argentifolii* nymphs and exuviae. p. 19. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
178. Buckner, J.S., Nelson, D.R. and Riemann, J.G. 1993. [SPW and *Trialeurodes vaporariorum* : wax glands]. p. 27. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
179. Buckner, J.S., Poprawski, T.J., Jones, W.A. and Nelson, D.R. 2000. The effects of *Eretmocerus mundus* parasitism on the cuticular lipids of *Bemisia argentifolii* nymphs. p. 83. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]

180. Buta, J.G. and Pittarelli, G.W. 1993. [Nicotiana activity]. p. 55. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
181. Butani, P.G. and Kapadia, M.N. 1995. Management of insect vectors of yellow mosaic virus (YMV) and crinkling diseases of green gram. p. 50. *In* Symposia on National Water Policy, Vector Biology and Integrated On-farm and Off-farm Employment, 2nd Agricultural Science Congress 19-21 January. National Academy of Agricultural Sciences, Andhra Pradesh Agricultural University, Rajendranagar, Hyderabad, India. [Abstract]
182. Butler, G.D., Jr. and Henneberry, T.J. 1993. [biological control, cotton, *Drapetis* (Empididae)]. p. 94. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
183. Butler, G.D., Jr. and Henneberry, T.J. 1993. [biological control on ornamentals, lantana, hibiscus, cotton, *Eretmocerus*]. p. 95. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
184. Butler, G.D., Jr. and Henneberry, T.J. 1993. [host plants]. p. 9. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
185. Butler, G.D., Jr., Henneberry, T.J., Stansly, P.A. and Schuster, D.J. 1993. [control, insecticidal soap, oils, detergents], p. 56. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
186. Byrne, D.N. and Blackmer, J. 1994. Behavioral, morphological and physiological traits associated with migratory *Bemisia tabaci*. p. 11. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
187. Byrne, D.N., Blackmer, J.L. and Rathman, R.J. 1994. Migration and dispersal by the sweetpotato whitefly. *Phytoparasitica* 22(4): 314. [Abstract]
188. Byrne, D.N., Blackmer, J.L., Rathman, R.J. and Tonhasca, A. 1993. [flight]. p. 10. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
189. Byrne, D.N., Isaacs, R. and Veenstra, K.H. 1996. Flight behavior by *Bemisia tabaci*, the virus bus. *Phytopathology* 86(11 SUPPL.): S113. [Abstract]
190. Byrne, D.N. and Rathman, R.J. 1996. Localized migration by the sweet potato whitefly, *Bemisia tabaci*. p. 2. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
191. Byrne D. N., Rathman, R.J., Orum, T.V. and Palumbo, J.C. 1995. Localized migration by sweet potato whiteflies. p. 5. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
192. Byrne, F.J., Prabhaker, N., Toscano, N.C., Nauen, R. and Castle, S. 2000. Studies on imidacloprid resistance in *Bemisia* whiteflies. p. 62. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
193. Byrne, F.J. and Toscano, N.C. 1999. *In vitro* acetylcholinesterase inhibition in *Bemisia tabaci* and its relevance to organophosphorus resistance expressed in bioassays. p. 49. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
194. Caballero, R. 1994. Whiteflies in Central America: economic importance, management and present research status. *Phytoparasitica* 22(4): 357. [Abstract]
195. Caballero, R., Anthony, N.M., French-Constant, R. and Brown, J.K. 1997. Monitoring endosulfan resistance and frequency of geminivirus DNA in individual whiteflies by polymerase chain reaction (PCR). p. 81. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
196. Cahill, M. and Denholm, I. 1997. Managing resistance to the chloronicotinyls: rhetoric or reality? *Amer. Chem. Soc.* 214(1-2): 20. [Abstract]
197. Campbell, B.C. 1993. [eubacterial endosymbionts, genes, rRNA transcript]. p. 28. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
198. Cardoza, Y.J., McAuslane, H.J. and Webb, S.E. 1998. Tolerance in zucchini, *Cucurbita pepo* L., to whitefly-induced squash silverleaf disorder. p. 88. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
199. Carlton, J.B. 1995. Retention phenomenon of agricultural liquid formulations on plant leaves. p. 67. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
200. Carlton, J.B., Bouse, L.F. and Kirk, I.W. 1995. Electrostatic charging of aerial spray over cotton. p. 68. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
201. Carlton, J.B., Bouse, L.F., Kirk, I.W. and Latheef, M.A. 1994. Aerial spray application systems/performance for sweetpotato whitefly control. p. 84. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
202. Carlton, J.B., Kirk, I.W. and Latheef, M.A. 1996. Aerial electrostatic charged sprays for control of sweetpotato whitefly in cotton. p. 60. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
203. Carnero, A., Díaz, S., Amador, S. and Hernández, E. 2000. Impact of *Nesidiocoris tenuis* (Hemiptera: Miridae) on whitefly populations in protected tomato crops. *Bull. OILB/SROP (IOBC/WPRS)* 23: 259. [Abstract]
204. Carnero, A. and González-Andujar, J.L. 1994. Spatial and temporal distribution of fourth-instar larvae of *Trialeurodes vaporariorum* and *Bemisia tabaci* in tomato plants. *Phytoparasitica* 22(4): 317. [Abstract]
205. Carruthers, R., Bradley, C., Wraight, S. and Jaronski, S. 1994. Development of fungal pathogens for control of sweetpotato whitefly. p. 124. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
206. Carruthers, R., Ciomperlik, M., Esau, K., Goolsby, J., Bradley, C., Jones, W., Legaspi, B., Parker, P., Poprawski, T., Taylor, M., Vacek, D., Wendel, L. and Wraight, S. 1996. Demonstration of biological control based IPM of sweetpotato whitefly. p. 104. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
207. Carruthers, R. and Wraight, S. 1993. [biological control, fungi, *Beauveria*, *Paecilomyces*, stains for spores]. p. 97. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
208. Carruthers, R.I. 1993. [sampling]. p. 11. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]



209. Carson, A.C., Freeman, T., Henneberry, T.J., Nelson, D., Buckner, J. and Chu, C.C. 1996. Microstructure of whitefly feeding: scanning electron microscopy of salivary sheaths. p. 27. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
210. Castle, S., Creamer, R. and Henneberry, T. 1994. Partial characterization and transmission by the silverleaf whitefly of a new disease of lettuce. p. 40. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
211. Castle, S. and Henneberry, T. 1995. Seasonal sex ratio dynamics of whiteflies in Imperial Valley, CA. p. 6. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
212. Castle, S. and Henneberry, T. 1996. Temporal patterns and host plant effects on responses of *Bemisia tabaci* to insecticides. p. 62. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
213. Castle, S., Henneberry, T. and Toscano, N. 1993. [irrigation, rainfall, sprinkler]. p. 117. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
214. Castle, S., Henneberry, T. and Weddle, D. 1996. Responses of *Bemisia tabaci* populations in Imperial Valley to bifenthrin and endosulfan in a vial bioassay. p. 61. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
215. Castle, S. and Henneberry, T.J. 1997. Factors contributing to increases of *Bemisia tabaci* populations: defining the outbreak pyramid. p. 192. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
216. Castle, S. and Henneberry, T.J. 1997. Progeny sex ratios and production by wild-type single pairs. p. 64. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
217. Castle, S.J. 1998. Primary pest or synthetically induced? The role of insecticides and other factors in the pest status of *Bemisia tabaci*. p. 107. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
218. Castle, S.J. 1999. Concentration and management of *Bemisia tabaci* in melons as a trap crop for cotton. p. 116. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
219. Castle, S.J. 2000. Reduced whitefly infestations in cotton using a melon trap crop. p. 136. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
220. Castle, S.J., Ellsworth, P.C., Diehl, J.W. and Henneberry, T.J. 1998. Within season shift in adult whitefly responses to Danitol + Orthene according to insecticide regimen. p. 40. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
221. Castle, S.J., González-Loc, M., León-López, R., Prabhaker, N., Toscano, N.C. and Henneberry, T.J. 1998. Regional comparison of whitefly responses to Danitol + Orthene: reduced susceptibility in Mexicali Valley. p. 39. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
222. Castle, S.J., Prabhaker, N., Henneberry, T.J., Toscano, N.C., León-López, R., Sánchez, B. and Gonzalez-Loc, M. 1997. Insecticide resistance differences in whiteflies from four regions: agricultural, operational, and ecological components. p. 111. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
223. Cebrian, R., Camero, A. and Pérez-Padrón, F. 1994. Some aspects of the biological cycle of *Bemisia tabaci* in the Canary Islands. *Phytoparasitica* 22(4): 313. [Abstract]
224. Chaabra, K.S. and Kooner, B.S. 1995. Varietal resistance as component of IPM of whitefly, *Bemisia tabaci*, and MYMV in blackgram. p. 48. *In* Symposia on National Water Policy, Vector Biology and Integrated On-farm and Off-farm Employment, 2nd Agricultural Science Congress 19-21 January. National Academy of Agricultural Sciences, Andhra Pradesh Agricultural University, Rajendranagar, Hyderabad, India. [Abstract]
225. Chalfant, R.B. 1994. Evaluation of insecticides by chemigation and ground application to control sweetpotato whitefly on cucurbits and tomatoes. p. 85. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
226. Chalfant, R.B. and Summer, H.R. 1993. Control of the sweetpotato whitefly on bell pepper, tomato, watermelons, and squash with insecticides, biorationals and new equipment technology. p. 57. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
227. Chalfant, R.B. and Summer, H.R. 1996. Performance of three sprayers on summer squash to control whitefly. p. 63. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
228. Chalfant, R.B. and Sumner, H.R. 1995. Surface application equipment and new pesticides to control the silverleaf whitefly and silverleaf on summer squash in Georgia. p. 69. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
229. Chandler, L.D., Sumner, H.R. and Herzog, G.A. 1993. [peanuts, Berthoud, Cannon air boom sprayer, chemigation]. p. 58. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
230. Chandrasegaram, S., Athulathmudali, D.P. and De Seneviratne, S.N. 1981. Studies on a virus disease of pumpkin (*Cucurbita maxima*). p. 26. *In* Proceedings 37th Annual Session. Part 1: Sectional Summaries. Sri Lanka Association for the Advancement of Science, Colombo, Sri Lanka. [Abstract]
231. Channarayappa, G., Shivashankar, G., Muniyappa, V. and Frist, R.H. 1990. Resistance to *Lycopersicon* species to the tomato leaf curl virus vector *Bemisia tabaci*. *Phytopathology* 80: 670. [Abstract]
232. Charoenying, S. 1982. Cotton insect pest management: screening of selected cotton varieties resistant to the cotton jassid, the cotton whitefly, the American bollworm and the cotton aphid. pp. 67-68. *In* Research and Development Institute, Research Reports 1982. Kasetsart University, Bangkok, Thailand. [Abstract]

233. Chavarria, A., Goolsby, J.A. and Wendel, L.E. 1996. Field cage rearing of exotic parasitoids. p. 105. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
234. Chavarria, A., Goolsby, J.A. and Wendel, L.E. 1997. Field cage rearing of exotic parasitoids. p. 135. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
235. Cheek, S. 1994. *Bemisia tabaci* - the United Kingdom protected zone. *Phytoparasitica* 22(4): 332-333. [ Abstract]
236. Chirachanya, K., Chiamkanokchai, C. and Chansukho, P. 1982. Study on the relationship of population densities between larva and adult whitefly *Bemisia tabaci* Gennadius. p. 137. *In* Research Reports 1982. Department of Agriculture, Bangkok, Thailand. [Thai, Abstract]
237. Chopra, B.L., Randhawa, L.S., Sekhon, P.S. and Singh, J. 1999. Integrated management of cotton leaf curl virus in North India. p. 31. *In* Proceedings ICAC-CCRI Regional Consultation, Insecticide Resistance Management in Cotton. Central Cotton Research Institute, Multan, Pakistan. [Abstract]
238. Chortyk, O.T. 1995. Synthesis and pesticidal activities of new sucrose esters. *Amer. Chem. Soc.* 210(1-2): 71. [ Abstract]
239. Chortyk, O.T. 1996. New pesticidal sucrose esters for whitefly control. p. 64. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
240. Chortyk, O.T. and Nottingham, S.F. 1995. Natural sugar esters as potent whitefly insecticides. *Amer. Chem. Soc.* 209(1-2): 71. [ Abstract]
241. Chu, C.C., Boykin, M.A., Jackson, C.G. and Henneberry, T.J. 1997. Dyne-amic increased the efficacy of EXP 80667A against silverleaf whiteflies on cotton. p. 113. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
242. Chu, C.C., Boykin, M.A., Jackson, C.G. and Henneberry, T.J. 1997. EXP 80667A - an experimental systemic insecticide for silverleaf whitefly control on cotton. p. 112. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
243. Chu, C.C., Cohen, A.C., Natwick, E.T., Simmons, G.S. and Henneberry, T.J. 1999. Silverleaf whitefly colonization and leaf morphology relationships in upland cotton cultivars. p. 100. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
244. Chu, C.C., Erickson, E.H., Crafts-Brandner, S.J. and Henneberry, T.J. 2000. Evaluation of leaf-clip cage and development of a single leaf plant technique to study photosynthetic traits of cotton under silverleaf whitefly infestation-stress conditions. p. 12. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
245. Chu, C.C., Freeman, T.P., Buckner, J.S., Henneberry, T.J. and Nelson, D.R. 2000. Do silverleaf whiteflies use leaf surface clues for feeding sites selection? p. 14. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
246. Chu, C.C., Freeman, T.P., Buckner, J.S., Henneberry, T.J., Nelson, D.R. and Natwick, E.T. 2000. Cotton leaf trichomes and silverleaf whitefly density relationship. p. 13. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
247. Chu, C.C., Freeman, T.P., Buckner, J.S., Henneberry, T.J., Nelson, D.R., Walker, G.P. and Natwick, T. 2000. Silverleaf whitefly colonization on upland cottons and relationships to leaf morphology and leaf age. p. 10. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
248. Chu, C.C., Freeman, T.P., Buckner, J.S., Natwick, E.T., Nelson, D.R. and Henneberry, T.J. 1999. Silverleaf whitefly oviposition on upland cotton cultivars. p. 101. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
249. Chu, C.C. and Henneberry, T.J. 1996. Endosulfan as a synergist to fenprothrin for silverleaf whitefly control. p. 65. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
250. Chu, C.C., Henneberry, T.J. and Akey, D.H. 1993. [cantaloupe, chemical control]. p. 59. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
251. Chu, C.C., Henneberry, T.J. and Akey, D.H. 1993. [cantaloupe, chemical control, planting dates]. p. 118. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
252. Chu, C.C., Henneberry, T.J. and Akey, D.H. 1993. [cotton, chemical control]. p. 60. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
253. Chu, C.C., Henneberry, T.J. and Boykin, M.A. 1998. Response of silverleaf whitefly adults to low intensity of white fluorescent light. p. 7. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
254. Chu, C.C., Henneberry, T.J. and Boykin, M.A. 1999. Attraction of silverleaf whiteflies to white fluorescent and incandescent light under laboratory conditions. p. 8. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
255. Chu, C.C., Henneberry, T.J. and Cohen, A.C. 1994. Sweetpotato whitefly host preference and leaf habitat orientation. p. 41. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
256. Chu, C.C., Henneberry, T.J. and Cohen, A.C. 1996. Development and evaluation of a new whitefly trap. p. 20. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
257. Chu, C.C., Henneberry, T.J., Mackey, B.E. and Perkins, H.H. 2000. Effects of silverleaf whitefly infestations on upland cotton yield and honeydew lint contamination and establishment of action threshold in the Imperial Valley, California. p. 63. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]

258. Chu, C.C., Henneberry, T.J. and Natwick, E.T. 1998. Evaluation of CC trap color and placement in various crops. p. 6. *In Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
259. Chu, C.C., Henneberry, T.J. and Natwick, E.T. 1999. Silverleaf whitefly adults caught in CC traps at different trap heights and trap relationships to leaf-turn counts on cotton. p. 6. *In Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
260. Chu, C.C., Henneberry, T.J., Natwick, E.T. and Cohen, A.C. 1997. Selection of cotton varieties to reduce insecticide applications for silverleaf whitefly control. p. 193. *In Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001).* U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
261. Chu, C.C., Henneberry, T.J., Natwick, E.T., Ritter, D. and Birdsall, S.L. 2000. Seasonal activity of adult silverleaf whiteflies in the Imperial and Palo Verde Valleys, California. p. 11. *In Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan.* U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
262. Chu, C.C., Henneberry, T.J., Perkins, H.H., Mackey, B.E., Prabhaker, N., Naranjo, S.E. and Akey, D.H. 1995. Control action thresholds for silverleaf whitefly for cotton yields and link quality. p. 70. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
263. Chu, C.C., Henneberry, T.J., Perkins, H.H., Naranjo, S.E., Prabhaker, N., Akey, D.H. and Mackey, B.E. 1995. Relationships between silverleaf whitefly populations and sticky cotton. p. 71. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
264. Chu, C.C., Henneberry, T.J., Perkins, H.H., Jr., Hendrix, D.L. and Steele, T. 1996. Sticky cotton and reduction of lint stickiness. p. 21. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
265. Chu, C.C., Henneberry, T.J., Prabhaker, N., Perkins, H.H. and Akey, D.H. 1994. Sweetpotato whitefly action and economic thresholds. p. 87. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
266. Chu, C.C., Jackson, C.G., Natwick, E.T., Henneberry, T.J. and Simmons, G.S. 1999. Selectivity of CC trap catches of whitefly adults and whitefly parasites *Eretmocerus eremicus* and *Eretmocerus emiratus*. p. 5. *In Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
267. Chu, C.C., Natwick, E.T., Brushwood, D.E., Henneberry, T.J., Castle, S.J. and Cohen, A.C. 1999. Upland cotton susceptibility to silverleaf whitefly infestations. p. 117. *In Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
268. Chu, C.C., Natwick, E.T., Cohen, A.C., Simmons, G.S. and Henneberry, T.J. 1998. Relationship between morphology and silverleaf whitefly densities on deltapine cotton cultivars. p. 91. *In Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
269. Chu, C.C., Natwick, E.T. and Henneberry, T.J. 1999. Effects of aldicarb on cotton insects and plant growth and yield. p. 118. *In Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
270. Chu, C.C., Natwick, E.T., Henneberry, T.J. and Cohen, A.C. 1998. Choice study of cotton cultivars susceptible to silverleaf whiteflies in fields. p. 90. *In Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
271. Chu, C.C., Natwick, E.T., Henneberry, T.J. and Cohen, A.C. 1998. No-choice study of cotton cultivar susceptibility to silverleaf whiteflies in the greenhouse. p. 89. *In Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
272. Chu, C.C., Natwick, E.T., Henneberry, T.J. and Lee, R. 1999. Effects of pyrethroid insecticides alone and in mixtures on silverleaf whitefly and cotton, cauliflower, and broccoli yields. p. 119. *In Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
273. Chu, C.C., Natwick, E.T., Perkins, H.H., Henneberry, T.J. and Cohen, A.C. 1996. Susceptibility of upland cotton cultivars to silverleaf whitefly under low desert conditions. p. 177. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
274. Chu, C.C., Natwick, E.T., Ritter, D., Henneberry, T.J. and Birdsall, S.L. 1999. Silverleaf whitefly adult catches in CC and suction traps above bare ground and implementation of CC traps for monitoring whitefly adult populations in Imperial and Palo Verde Valleys in California. p. 7. *In Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
275. Chu, C.C., Pinter, P.J., Jr., Henneberry, T.J., Umeda, K., Natwick, E.T., Wei, Y., Reddy, V.R. and Shrepatis, M. 1999. Catches of silverleaf whiteflies, thrips and leafhoppers with different trap base colored CC traps. p. 9. *In Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
276. Cicero, J., Hiebert, E. and Webb, S.E. 1994. Anatomy of *Bemisia tabaci*. p. 42. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
277. Ciomperlik, M., Goolsby, J., Hennessy, R., Meyerdirk, D., Parker, P., Vacek, D. and Wendel, L. 1995. APHIS biological control program against *Bemisia tabaci*. p. 101. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
278. Ciomperlik, M., Goolsby, J., Poprawski, T., Wendel, L. and Wraight, S. 1997. Demonstration of biological control based IPM of sweetpotato whitefly. p. 136. *In Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001).* U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
279. Ciomperlik, M.A. and Goolsby, J.A. 1996. Field impact evaluations of exotic parasitoids in the Lower Rio Grande Valley [TX] on selected crops. p. 107. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
280. Ciomperlik, M.A. and Hagler, J.R. 1995. Survey of native predators for SPWF egg predation in cotton in the Lower Rio Grande Valley, Texas. p. 102. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]

281. Ciomperlik, M.A., Rodriguez, J.M. and Wendel, L.E. 1995. Assessment of sweetpotato whitefly (*Bemisia tabaci*, Biotype B) and indigenous parasite populations in agrosystems of the Lower Rio Grande Valley, Texas. p. 103. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
282. Ciomperlik, M.A., Rodriguez, J.M. and Wendel, L.E. 1996. Assessments of sweetpotato whitefly (*Bemisia tabaci*, biotype B) and indigenous parasite populations in agrosystems of the Lower Rio Grande Valley, TX. p. 106. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
283. Ciomperlik, M.A. and Wendel, L.E. 1998. Regional population dynamics of sweetpotato whitefly (*Bemisia tabaci*, biotype B) in the Lower Rio Grande Valley of Texas. p. 55. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
284. Coates, W. 1994. Field trials of electrostatic sprayers on tomatoes, cotton and cauliflower. p. 86. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
285. Coates, W., Palumbo, J. and Watson, T. 1993. [chemical control, electrostatic sprayer, cotton, cauliflower, lettuce]. p. 61. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
286. Coates, W.E. and Palumbo, J.C. 1996. Deposition, off-target movement and efficacy of Capture and Thiodan applied to melons using several application technologies. p. 66. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
287. Coats, S.A., Frohlich, D.R., Henneberry, T.J. and Brown, J.K. 1994. The kinetics and inhibition of whitefly (*Bemisia tabaci*) acetylcholinesterases and host plant influences on the kinetic parameters of acetylcholinesterase activity in the 'A' and 'B' biotypes. p. 43. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
288. Coats, S.A., Hendrix, D.L. and Brown, J.K. 1993. [esterase banding patterns]. p. 29. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
289. Coats, S.A., Henneberry, T.J. and Brown, J.K. 1994. Assessment of sex ratios in the 'A' and 'B' biotypes of *Bemisia tabaci* Genn. p. 44. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
290. Coats, S.A., Lacey, L.A., Kirk, A.A. and Brown, J.K. 1994. Biotypes of *Bemisia tabaci* in Eurasia and the detection of the 'B' biotyped in the Mediterranean Basin by esterase profile analysis. p. 45. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
291. Cohen, A.C. and Brummett, D. 1996. Alkaline proteinases (serine proteinases) are not present in silverleaf or sweetpotato whiteflies. p. 22. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
292. Cohen, A.C., Chu, C.C., Henneberry, T.J., Freeman, T. and Margosan, D. 1996. Microstructure of feeding in nymphal *Bemisia argentifolii* in cotton and cantaloupe. p. 26. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
293. Cohen, A.C., Chu, C.C., Henneberry, T.J., Freeman, T. and Nelson, D. 1996. Cotton leaf surface features serve as behavioral cues to silverleaf whiteflies. p. 23. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
294. Cohen, A.C., Chu, C.C., Henneberry, T.J., Freeman, T.P., Nelson, D.R., Buckner, J., Margosan, D., Vail, P. and Aung, L.H. 1999. Studies of silverleaf whitefly nymphs feeding behavior. p. 10. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
295. Cohen, A.C., Chu, C.C., Henneberry, T.J., Nafziger, K. and Percy, T. 1996. Characteristics of cotton leaves related to whitefly feeding. p. 158. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
296. Cohen, A.C., Freeman, T., Chu, C.C., Henneberry, T.J., Buckner, J., Nelson, D. and Vail, P. 1997. Characteristics of leaves of cotton varieties in relation to whitefly feeding biology. p. 178. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
297. Cohen, A.C., Freeman, T., Chu, C.C., Henneberry, T.J., Buckner, J., Nelson, D. and Vail, P. 1997. *In Situ* studies of whitefly feeding biology. p. 83. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
298. Cohen, A.C., Henneberry, T.J. and Chu, C.C. 1995. Basic feeding relationships between silverleaf whiteflies and host plant vascular bundles by period covered: 1994. p. 28. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
299. Cohen, A.C., Henneberry, T.J. and Chu, C.C. 1995. Feeding and oviposition in silverleaf whiteflies on cantaloupe, cotton and lettuce leaves. p. 29. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
300. Cohen, A.C., Henneberry, T.J. and Chu, C.C. 1996. Geometric relationships between whitefly feeding behavior and vascular bundle arrangements. p. 25. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
301. Cohen, A.C., Henneberry, T.J., Hendrix, D. and Staten, R. 1993. [water loss, oxygen consumption, thermal tolerances, feeding studies, artificial diet, honeydew]. p. 30. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
302. Cohen, A.C., Henneberry, T.J. and Staten, R. 1993. [predators, *Geocoris*, *Chrysoperla*]. p. 96. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
303. Cohen, A.C., Nelson, D., Buckner, J. and Brummett, D. 1996. Nutrient rewards for predators of whiteflies: generalists' requirements. p. 28. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
304. Cohen, A.C., Smith, L. and Brummett, D. 1997. Development of an artificial diet for *Chrysoperla carnea*. p. 137. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]

305. Cohen, A.C., Smith, L. and Brummett, D. 1997. Measurement of nutrient rewards for predators consuming whiteflies: sterols from *Bemisia argentifolii*. p. 138. In Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
306. Cohen, A.C. and Smith, L.K. 1996. Development of artificial diet for whitefly predators: *Serangium parcesetosum*. p. 24. In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
307. Cohen, A.C., Staten, R.T. and Brummett, D. 1995. Why a generalist (*Geocoris punctipes*) cannot thrive on whiteflies and why a specialist (*Serangium parcesetosum*) can: or, whiteflies as junk food. p. 30. In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
308. Cohen, J., Gera, A., Ecker, R., Ben-Joseph, R., Perlman, M., Gokkes, M. and Antignus, Y. 1994. Lisianthus leaf curl, a new disease of lisianthus (*Eustoma grandiflorum*) caused by tomato yellow leaf curl virus. *Phytoparasitica* 22(4): 328. [ Abstract]
309. Cohen, M., Ziegweid, K. and Minkenberg, O.P.J.M. 1994. Use of genetic fingerprinting for identification of whitefly parasitoids. p. 46. In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
310. Cohen, S. 1993. Sweet potato whitefly biotypes and their connection with squash silver leaf. *Phytoparasitica* 21(2): 174. [ Abstract]
311. Cohen, S. 1994. Plant resistance of *Bemisia tabaci* viruses. *Phytoparasitica* 22(4): 328-329. [ Abstract]
312. Cohen, S., Antignus, Y. and Ben-Joseph, R. 1984. Whitefly-borne viruses in Israel. *Phytoparasitica* 2: 140. [ Abstract]
313. Cohen, S. and Ben-Joseph, R. 1986. Preliminary studies of the distribution of whiteflies (*Bemisia tabaci*), using florescent dust to mark the insects. *Phytoparasitica* 14: 152-153. [ Abstract]
314. Cohen, S., Ben-Joseph, R., Mor, N. and Antignus, Y. 1995. Unique effects of certain polyethylene sheets on the behavior of *Bemisia tabaci* (Gennadius) and spread of viruses vectored by this insect. p. 31. In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
315. Cohen, S., Duffus, J.E., Liu, H.Y. and Perry, R. 1991. Induction of silverleaf of squash by *Bemisia* whitefly from California desert whitefly populations. *Plant Dis.* 75(8): 862. [ Abstract, Cock (1993)]
316. Cohen, S., Duffus, J.E., Liu, H.Y. and Perry, R. 1992. Sweet potato whitefly *Bemisia tabaci* biotypes. 13th Congress Israeli Phytopathol. Soc. 20(3): 248. [ Abstract]
317. Cohen, S., Keren, J., Harpaz, I. and Bar-Joseph, R. 1986. Studies of the epidemiology of a whitefly-borne virus, tomato yellow leaf curl virus, in the Jordan Valley. *Phytoparasitica* 14: 158. [ Abstract]
318. Cohen, S. and Veierov, D. 1989. Effect of various insect control agents on the landing behavior of *Bemisia tabaci*. *Phytoparasitica* 17: 229-230. [ Abstract]
319. Colvin, J., Muniyappa, V., Ramappa, H.K., Cherian, A.K., Venkatesh, H.M., Nagaraju, N., Maruthi, M.N. and Green, S.K. 1999. The epidemiology and management of tomato leaf curl virus and *Bemisia tabaci* in southern India. pp. 72-73. In *Plant virus epidemiology: Current status and future prospects*. VII International Plant Virus Epidemiology Symposium, Aguadulce(Almeria), Spain. [Abstract]
320. Colvin, J., Otim -Nape, G.W., Holt, J., Omongo, C., Seal, S., Stevenson, P., Gibson, G., Cooter, R.J. and Thresh, J.M. 1999. Factors driving the current epidemic of severe cassava mosaic disease in East Africa. pp. 76-77. In *Plant virus epidemiology: Current status and future prospects*. VII International Plant Virus Epidemiology Symposium, Aguadulce(Almeria), Spain. [Abstract]
321. Conti, M. 1994. Whiteflies other than *Bemisia tabaci* as vectors of plant viruses. p. 15. In B. Ezzahiri and M. Bouhache (Eds.), Fifth Arab Congress of Plant Protection, November 27 - December 2, 1994, Fez, Morocco. Actes Editions, Rabat, Maroc. [Abstract]
322. Cook, C.G. 1996. Mechanisms of resistance to the silverleaf whitefly: applications to cotton breeding. p. 158. In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
323. Cook, C.G. 1997. Mechanisms of resistance to the silverleaf whitefly and its interactions with the reniform nematode: applications to cotton breeding. p. 179. In Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
324. Corbett, A., Rosenheim, J.A., Roltsch, W., Pickett, C. and Stimmann, M. 1996. Quantifying the effect of early -season refugia on aphelinid populations through elemental labeling. p. 108. In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
325. Costa, A.S. 1974. Molestias de tomateiro no Brasil transmitidas pela mosca branca *Bemisia tabaci*. [Tomato diseases transmitted by the white-fly *Bemisia tabaci* in Brazil]. *Fitopatologia* 9(2): 47. [ Portuguese, Abstract]
326. Costa, A.S. 1975. Comparacao de machos e femeas de *Bemisia tabaci* na transmissao do mosaico dourado do feijoeiro. [Comparison of male and female *Bemisia tabaci* in the transmission of the bean golden mosaic]. *Fitopatologia* 10(2): 51. [ Portuguese, Abstract]
327. Costa, A.S. 1975. Feijoeiro como modelo experimental em testes de fitoviricidas in vitro. [Bean plants as an experimental model to test phytoviricidas in vitro]. *Fitopatologia* 10(2): 51. [ Portuguese, Abstract]
328. Costa, A.S. 1975. Plantas teste para o mosaico dourado do feijoeiro. [Test plants for the bean golden mosaic]. *Fitopatologia* 10(2): 51. [ Portuguese, Abstract]
329. Costa, A.S., Oliveira, A.R. and Silva, D.M. 1975. Transmissao mecanica do mosaico dourado do tomateiro. [Mechanical transmission of tomato, golden mosaic, a whitefly -transmitted disease.]. *Fitopatologia* 10(2): 51-52. [ Portuguese, Abstract]
330. Costa, C.L. and Cupertino, F.P. 1975. Reacao de *Bemisia tabaci*, importante vector de virus, a tres diferentes cores. [Reaction of *Bemisia tabaci*, important vector of plant viruses, to three different colors]. *Fitopatologia* 10(2): 53. [ Portuguese, Abstract]
331. Costa, C.L. and Cupertino, F.P. 1976. Disseminacao do mosaico dourado, pelo vector, de uma fonte de inoculo proxima ao feijoal. [Dissemination of the bean golden mosaic virus, through the vector from a source of inoculum close to the bean field]. *Fitopatologia* 11(1/2): 12. [ Portuguese, Abstract]
332. Costa, C.L., Cupertino, F.P., Vieira, C. and Kitajima, E.W. 1975. Incidencia do mosaico dourado em feijoais do Triangulo Mineiro. [Occurrence of bean golden mosaic at the Triangulo Mineiro]. *Fitopatologia* 10(2): 53. [ Portuguese, Abstract]
333. Costa, H., Ullman, D.E., Johnson, M.W. and Tabashnik, B.E. 1994. Association between *Bemisia tabaci* density and reduced growth, yellowing and stem blanching of lettuce and kai choy. p. 12. In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
334. Costa, H., Ullman, D.E., Johnson, M.W., Westcot, D.M. and Tabashnik, B.E. 1994. Whitefly endosymbionts. p. 47. In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]

335. Costa, H.S. 1998. Interaction of *Beauveria bassiana* with various fungicides under exposed conditions. p. 108. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
336. Costa, H.S. and Brown, J.K. 1990. Variability in biological characteristics, isozyme patterns and virus transmission among populations of *Bemisia tabaci* Genn. in Arizona (Abstr.). *Phytopathology* 80(9): 888. [Abstract]
337. Costa, H.S., Hendrix, D.L., Davidson, E.W., Henneberry, T.J. and Toscano, N. 1996. Effects of antibacterial materials on silverleaf whitefly oviposition growth, survival, sex ratio and honeydew content. p. 29. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
338. Costa, H.S. and Robb, K.L. 1998. Effects of UV-blocking plastics on insect flight behavior. p. 109. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
339. Costa, H.S., Toscano, N.C., Hendrix, D.L. and Henneberry, T.J. 1997. Periodic pattern of honeydew production by immature *Bemisia argentifolii*. p. 84. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
340. Costa, H.S., Toscano, N.C. and Henneberry, T.J. 1996. Mycetocyte inclusion in the oocytes of silverleaf whitefly, *Bemisia argentifolii* (Homoptera: Aleyrodidae). p. 30. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
341. Costa, H.S., Westcot, D.M., Ullman, D.E., Rosell, R.C., Brown, J.K. and Johnson, M.W. 1995. Morphological variation in the bacterial endosymbionts within the whitefly genus *Bemisia*. p. 32. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
342. Costa, H.S., Westcot, D.M., Ullman, D.E., Rosell, R.C., Brown, J.K. and Johnson, M.W. 1995. Observations of virus-like particles in the mycetocytes of *Bemisia tabaci*. p. 33. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
343. Cottage, E.L.A. and Gunning, R.V. 1998. The effect of organophosphate insecticides on the inhibition of esterase mediated hydrolysis of 1-naphthyl butyrate in the B-type *Bemisia tabaci* (Homoptera: Aleyrodidae) in Australia. p. 204. *In* U. Kechagia (Ed.), World Cotton Research Conference 2. Sept. 6-12, 1998. Athens Greece. [Abstract]
344. Crafts-Brandner, S.J. 2000. Effect of N nutrition of free amino acid levels in silverleaf whiteflies. p. 15. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
345. Craig, W., McGrath, P.F., Torres-Jerez, I., Wyatt, S.D. and Brown, J.K. 1997. SqLCV [Squash leaf curl geminivirus]: evaluation of strains in homologous and heterologous combinations. p. 85. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
346. Creamer, R. 1993. [geminivirus, carrot]. p. 30. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
347. Crosslin, J.M., Brown, J.K. and Johnson, D.A. 1988. First report of zucchini yellow mosaic virus in *Cucurbita pepo* in the Pacific Northwest. *Plant Dis.* 72: 362. [Abstract]
348. Csizinszky, A.A. and Schuster, D.J. 1994. The influence of sweetpotato whitefly management and potassium fertilization on yield and irregular ripening of tomato. *HortScience* 29(5): 460. [Abstract]
349. Csizinszky, A.A. and Schuster, D.J. 1996. Effect of UV-reflective mulches on tomato yields and on the silverleaf whitefly, *Bemisia argentifolii* (Bellows and Perring). *Hortscience* 31(4): 570. [Abstract]
350. Csizinszky, A.A., Schuster, D.J. and Kring, J.B. 1990. Effect of mulch color on tomato yields and on insect vectors. *HortScience* 25: 1131. [Abstract]
351. Csizinszky, A.A., Schuster, D.J. and Kring, J.B. 1995. Evaluation of color mulches and oil sprays for yield and silverleaf whitefly control on tomatoes. *Hortscience* 30(4): 755. [Abstract]
352. Czosnek, H. and Ghanim, M. 1999. Sex-mediated transmission and propagation of tomato yellow leaf curl geminivirus by whiteflies. pp. 37-38. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce(Almeria), Spain. [Abstract]
353. Czosnek, H., Morin, S., Rubinstein, G., Fridman, V., Zeidan, M. and Ghanim, M. 2000. Tomato yellow leaf curl geminivirus (TYLCV-Is), a sexually transmitted disease of the whitefly *Bemisia tabaci* (*B. argentifolii*). pp. 4-5. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
354. Dafalla, G.A., Lecoq, H., Kheyr-Pour, A. and Gronenborn, B. 1994. A whitefly-transmitted geminivirus associated with a yellowing disease of watermelons in Sudan. p. 173. *In* B. Ezzahiri and M. Bouhache (Eds.), Fifth Arab Congress of Plant Protection, November 27 - December 2, 1994, Fez, Morocco. Actes Editions, Rabat, Maroc. [Abstract]
355. Dahan, R., Berlinger, M.J., Mordechi, S. and Cohen, S. 1985. Screening of insecticides to prevent the transmission of tomato yellow leaf curl virus by the tobacco whitefly, *Bemisia tabaci*. *Phytoparasitica* 13: 77. [Abstract]
356. Davidson, E.W., Cirillo, E., Patron, B.R., Sequra, B.J., Lacey, L.A., Steele, T. and Hendrix, D. 1995. The relationship between *Bemisia* microorganisms and honeydew formation. p. 34. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
357. Davidson, E.W., Jancovich, J.K., Lavine, M. and Hendrix, D. 1997. Feeding chamber and diet for culture of nymphal silverleaf whitefly, *Bemisia argentifolii* (Homoptera: Aleyrodidae). p. 86. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
358. Davidson, E.W., Jancovich, J.K., Lavine, M. and Hendrix, D.L. 1996. Feeding and behavior of *Bemisia argentifolii* larvae using an artificial feeding system. p. 32. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
359. Davidson, E.W. and Jones, W. 1999. Successful rearing of parasitoid wasps on *Bemisia argentifolii* cultured on artificial diet. p. 69. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
360. Davidson, E.W. and Jones, W.A. 1997. Suitability of *Bemisia argentifolii* reared on artificial diet as hosts for parasitoids. p. 114. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]

361. Davidson, E.W., Lavine, M.D., Mathews, M. and Hendrix, D.L. 1999. Improvements to the artificial feeding system for *Bemisia argentifolii*. p. 11. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
362. Davidson, E.W., Patron, B. R., Lacey, L.A., Frutos, R., Vey, A. and Hendrix, D.L. 1995. Activity of natural and semi-synthetic toxins on the silverleaf whitefly *Bemisia argentifolii*, using a novel feeding bioassay system. p. 35. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
363. Davidson, E.W., Patron, R., Vey, A., Frutos, R., St. Leger, R., Lacey, L.A. and Hendrix, D.L. 1996. Effects of destruxins from *Metarhizium anisopliae* and *Bacillus thuringiensis* delta-endotoxins on adult silverleaf whitefly, *Bemisia argentifolii*. p. 31. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
364. Davidson, E.W., Patron, R.B.R., Mitich, D. and Hendrix, D.L. 1994. A simplified feeding bioassay system for adult silverleaf whitefly, *Bemisia argentifolii*. *Phytoparasitica* 22(4): 350. [ Abstract]
365. Davis, M.J., Ying, Z.N. and McMillan, R.T., Jr. 1998. Occurrence of tomato yellow leaf curl geminivirus in the United States. p. 29. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
366. De Uzcategui, R.C. 1996. Properties of tomato yellow mosaic geminivirus in Venezuela. *Phytopathology* 86(11 SUPPL.): S114. [ Abstract]
367. Dean, D.E. and Schuster, D.J. 1994. Predaceous arthropods of *Bemisia tabaci* on tomatoes in Florida. *Phytoparasitica* 22(4): 334. [ Abstract]
368. Dean, D.E. and Schuster, D.J. 1998. Mortality factors affecting *Bemisia argentifolii* (Homoptera: Aleyrodidae) on tomatoes in Florida. p. 8. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
369. DeGrandi-Hoffman, G. and Naranjo, S.E. 1996. Biocontrol-whitefly: A whitefly, parasite, and predator population dynamics model. p. 3. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
370. DeGrandi-Hoffman, G., Naranjo, S.E. and Gould, J. 1997. Insights on *Bemisia* population dynamics from simulations with the BIOCONTROL-WHITEFLY model. p. 65. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
371. Degrandi-Hoffman, G., Naranjo, S.E., Li, D. and Henneberry, T.J. 1994. Development of BIOCONTROL-WHITEFLY: a whitefly/parasitoid/predator population dynamics model. p. 17. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
372. Denholm, I., Cahill, M.R., Byrne, F.J. and Devonshire, A.L. 1994. Progress with documenting and combating insecticide resistance in *Bemisia tabaci*. *Phytoparasitica* 22(4): 346. [ Abstract]
373. Dennehy, T.J. and Antilla, L. 1996. Whitefly resistance to insecticides in Arizona. p. 68. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
374. Dennehy, T.J., Williams, L., III, Li, X., Wigert, M. and Birdwell, E. 1997. Whitefly resistance in insecticides in Arizona cotton. p. 139. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
375. Dennehy, T.J., Williams, L., III, Russell, J.S., Li, X. and Wigert, M. 1996. Monitoring and management of whitefly resistance to insecticides in Arizona. p. 67. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
376. Dhingra, K.L. and Nariani, T.K. 1961. Yellow net virus disease of tobacco plant. *Indian J. Microbiol.* 1: 98. [ Abstract, Cock (1986)]
377. Diaz-Plaza, R. 1995. New disease in horticulture crops in Yucatán, Mexico. p. 36. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
378. Diaz-Plaza, R., Ramirez, C.J.L. and Aviles, B.W. 1995. Integrated control of virus transmitted by whitefly in tomato. p. 163. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
379. Diehl, J., Ellsworth, P., Umeda, K. and Husman, S. 1996. Transfer of integrated pest management technology in a community-based action program for sweetpotato whitefly management. p. 178. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
380. Dittmar, S.H., Ellsworth, P.C., Hartman, P.M., Martin, E.C., McCloskey, W.B., Olsen, M.W., Roth, R.L., Silvertooth, J.C. and Tronstad, R.E. 1999. Interdisciplinary demonstration of Arizona irrigated cotton production. p. 120. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
381. Dolores, L.M. and Bajet, N.B. 1995. Occurrence of tomato yellow leaf curl virus in the Philippines. *Philippine J. Crop Sci.* 20(Suppl. 1): 22. [ Abstract]
382. Dominguez-Rubio, Y. 1988. Extractos e infusiones vegetales y entomopatogenos como alternativa ecologica en la regulacion poblacional de mosquita blanca. [Vegetable extracts, infusions and entomopathogens as ecological alternative in the population regulation of white fly]. p. 596. *In* Proceedings National Entomological Congress. 33, Acapulco, Mexico, 24-27 May 1998. [Spanish, Abstract]
383. Doostdar, H., Inbar, M. and Mayer, R.T. 1999. Effects of tomato mottle virus infection on tomato root development. p. 31. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
384. Drees, B.M. 1993. [chemical control, insect growth regulator, *Beauveria*, *Paccilomyces*]. p. 61. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
385. Duffus, J.E. 1994. Diseases vectored by whiteflies: etiology, ecology, geographical distribution and possible control measures. p. 13. *In* B. Ezzahiri and M. Bouhache (Eds.), Fifth Arab Congress of Plant Protection, November 27 - December 2, 1994, Fez, Morocco. Actes Editions, Rabat, Maroc. [Abstract]
386. Duffus, J.E. 1994. Whitefly -borne viruses. *Phytoparasitica* 22(4): 323. [ Abstract]
387. Duffus, J.E., Caciagli, P., Liu, H.Y., Wisler, G.C. and Li, T. 1996. Occurrence of tomato infectious chlorosis in Europe. p. 33. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]

388. Duffus, J.E. and Johns, M.R. 1985. Melon leaf curl virus - a new gemini virus with host and serological variations from squash leaf curl virus. *Phytopathology* 75: 1312. [Abstract, Cock (1986)]
389. Duffus, J.E. and Liu, H.Y. 1994. The effects of whitefly population changes on lettuce infectious yellows virus epidemiology. p. 48. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
390. Duffus, J.E., Liu, H.Y. and Cohen, S. 1994. Partial characterization of a new closterovirus, the causal agent of cucurbit yellow stunting disorder. p. 49. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
391. Duffus, J.E., Liu, H.Y. and Wisler, G.C. 1995. Distribution of tomato infectious chlorosis virus in California. p. 37. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
392. El-Ghany, A., El-Sayed, M. and Moawad, G.M. 1995. Ecological and biological studies on *Encarsia lutea* a primary parasite of *Bemisia tabaci* in Egypt. p. 104. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
393. El-Lissy, O. and Antilla, L. 1993. [chemical control, field edge treatments, cotton]. p. 62. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
394. El-Lissy, O., Antilla, L., Leggett, J.E. and Staten, R.T. 1994. Areawide control of sweetpotato whitefly, *Bemisia tabaci*, on cotton in Paloma, AZ. p. 90. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
395. El-Serwiy, S. and El-Haidari, H. 1983. Population density of *Bemisia tabaci* (Gennadius) on cucumber. p. 61. *In* 1st Scientific Meeting, Arab Society for Plant Protection, Amman, Jordan, 22-25 Nov 1982. Arab Society of Plant Protection, Beirut, Lebanon. [Abstract]
396. Ellington, J.J. and Carrillo, T. 1995. A computer-assisted recognition system for whiteflies. p. 7. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
397. Ellis, D.R., McAvoy, R.J., Ayyash, L.A., Flanagan, M. and Ciomperlik, M.A. 2000. Evaluation of *Serangium parcesetosum* (Coleoptera: Coccinellidae) for biological control of silverleaf whitefly, *Bemisia argentifolii* (Homoptera: Aleyrodidae), on poinsettia. p. 84. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
398. Ellsworth, P., Dennehy, T., Diehl, J., Ansolobehere, M., Antilla, L., Howell, D., Hussman, S., Knowles, T., Minch, E., Palmer, A., Rowland, M., Stedman, S. and Thacker, G. 1997. Statewide training sessions for use of insect growth regulators in Arizona. p. 195. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
399. Ellsworth, P. and Diehl, J. 1996. Development & adoption of recommendations for whitefly management in cotton. p. 179. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
400. Ellsworth, P., Diehl, J., Dennehy, T. and Naranjo, S. 1995. Development and delivery of sampling plans for sweetpotato whiteflies in cotton. p. 164. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
401. Ellsworth, P., Diehl, J. and Husman, S. 1994. Organization of community-wide IPM in cotton. p. 176. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
402. Ellsworth, P., Diehl, J., Naranjo, S., Husman, S. and Dennehy, T. 1995. Establishment of integrated pest management infrastructure: A community-based action program for sweetpotato whitefly management. p. 165. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
403. Ellsworth, P. and Meade, D. 1994. Action thresholds for whiteflies in Arizona. p. 13. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
404. Ellsworth, P. and Meade, D. 1994. Chemical efficacy tests for sweetpotato whitefly control. p. 88. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
405. Ellsworth, P. and Meade, D. 1995. The effects of carbamates on sweetpotato whitefly population. p. 73. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
406. Ellsworth, P. and Meade, D. 1995. Field evaluation of novel chemistries for control of the sweetpotato whitefly. p. 72. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
407. Ellsworth, P., Meade, D., Byrne, D., Chernicky, J., Draeger, E. and Gibson, R. 1994. Progress on the use of trap crops for whitefly suppression. p. 160. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
408. Ellsworth, P., Meade, D., Husman, S., Ramsey, C., Silvertooth, J. and Malcuit, J. 1994. Sweetpotato whitefly preference and performance on medium maturity cotton varieties in Arizona. p. 159. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
409. Ellsworth, P., Meade, D. and Odom, P. 1994. Field evaluation of an insect growth regulator, Buprofezin, for control of the sweetpotato whitefly, *Bemisia tabaci*. p. 89. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
410. Ellsworth, P., Naranjo, S. and Diehl, J. 1997. Implementation and dissemination of sampling and threshold protocols for whitefly nymphs in cotton. p. 194. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
411. Ellsworth, P.C. 1999. Whitefly management in cotton - a historical perspective. p. 121. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]



412. Ellsworth, P.C., Akey, D.H., Dennehy, T.J., Williams, L., Kirk, I.W., Carleton, J., Henneberry, T.J., Coppedge, J. and Diehl, J.W. 1996. Understanding whitefly control: threshold, insecticide rotation and ground and air comparisons. p. 180. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
413. Ellsworth, P.C., Diehl, J.W. and Husman, S.H. 1994. Establishment of an integrated pest management infrastructure: a community-based action program for sweetpotato whitefly management. *Phytoparasitica* 22(4): 353-354. [ Abstract]
414. Ellsworth, P.C., Kirk, I.W., Diehl, J.W., Meade, D.L., Harris, C., Tom, H., Henneberry, T.J. and Coppedge, J.R. 1997. Large scale evaluation of insect growth regulators for whitefly control. p. 115. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
415. Ellsworth, P.C., Naranjo, S.E., Castle, S., Hagler, J. and Henneberry, T.J. 1998. Development of integrated whitefly management strategies. p. 110. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
416. Ellsworth, P.C., Silvertooth, J.C., McCloskey, W.B., Brown, P.W., Martin, E.C. and Moser, H.S. 1998. Integrated cotton management: incorporating whitefly management & multi-pest IPM. p. 111. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
417. Elmstrom, G. 1993. [squash, silvering]. p. 31. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
418. Elsey, K.D. 1993. [overwintering]. p. 12. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
419. Elsey, K.D. and Farnham, M. 1993. [*Brassica*, kale, broccoli, kohlrabi, cabbage, color]. p. 118. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
420. Elsey, K.D., Farnham, M.W. and van Giessen, W. 1994. Glossy brassica's resistance to *B. tabaci*. p. 158. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
421. Elsner, O. 1982. The quality of cotton lint contaminated by honeydew. *Phytoparasitica* 10: 295. [ Abstract]
422. Elsner, O., Stern, G. and Lubenskaya, G. 1983. The effect of honeydew on the quality of cotton lint. *Phytoparasitica* 11: 65. [ Abstract]
423. Elzen, G.W. 2000. Laboratory toxicity of selected insecticides to silverleaf whiteflies (preliminary results). p. 64. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
424. Espinosa, P. 1972. Estudios preliminares para el control de mosca blaca *Bemisia tabaci* (Genn.) del algodoner. p. 25. *In* G. Halffter (Ed.), Seventh National Congress Entomol., 13-16 October 1970, Mexico, D.F., *Folia Entomol. Mexicana* Vol. 23/24. [Abstract, Cock (1986)]
425. Esquer, T., Gomez, C.O., Brown, J.K., Golstein, D.E. and Lambe, R.C. 1987. Evaluation of spun-bonded polyester and perforated floating row covers to control two plant virus. Biological and cultural tests for control of plant diseases. *Am. Phytopathol. Soc.* 2: 29. [ Abstract]
426. Faria, J.C., Souza, J.A.C., Slack, S.A. and Maxwell, D.P. 1997. A new geminivirus associated with tomato in the state of Sao Paulo, Brazil. *Plant Dis.* 81(4): 423. [ Abstract]
427. Farias-Larios, J. and Michel-Rosales, A. 1998. Sustainable production of honeydew and muskmelon in Western Mexico. *Hortscience* 33(3): 495. [ Abstract]
428. Farias-Larios, J., Orozco, M., Guzman, S. and Gutierrez, A. 1997. Effectiveness of different applications of imidacloprid for the control of sweetpotato whitefly and muskmelon yield. *Hortscience* 32(3): 435-436. [ Abstract]
429. Farias-Larios, J., Orozco, M., Guzmán, S. and Pérez, J. 1996. Effect of plastic mulch floating rowcovers and microtunnels on insect populations and yield of muskmelon. *Hortscience* 31(4): 677. [ Abstract]
430. Farias-Larios, J., Orozco-Santos, M. and Guzman-Gonzalez, S. 1995. Yield of three cultivars of muskmelon growth on transparent mulch and floating rowcover in a tropical region. *Hortscience* 30(4): 890-891. [ Abstract]
431. Farias-Larios, J., Orozco-Santos, M. and Ramirez-Vazquez, N.R. 2000. Use of *Beauveria bassiana* for silverleaf whitefly (*Bemisia argentifolii* Bellows & Perring) management in muskmelon. *Hortscience* 35(3): 393. [ Abstract]
432. Fasulo, T.R. 1994. A hypertext knowledgebase of the sweetpotato whitefly. *Phytoparasitica* 22(4): 315. [ Abstract]
433. Fasulo, T.R., Allen, J.C., Bellows, T.S., Evans, G.A., Flint, M.L., Goodell, P.B., Liu, T.X., Nichols, R.L., Norman, J.W., Perring, T.M., Riley, D.G., Sparks, A.N., Stansly, P.A. and Toscano, N.C. 1995. A hypertext computer knowledgebase on whiteflies damaging to crops & ornamentals. p. 166. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
434. Fauquet, C.M., Sangare, A., Deng, D., Fux, C. and Beachy, R.N. 1994. Engineering plants for resistance to whitefly-borne viruses. *Phytoparasitica* 22(4): 329. [ Abstract]
435. Fehmy, M., Hegab, A.H. and Moawad, G.M. 1994. valuation of programs to control the cotton whitefly, *Bemisia tabaci*, in tomato and squash fields and reduce the spread of TYLCV in Egypt. *Phytoparasitica* 22(4): 348-349. [ Abstract]
436. Ferrentino, G.W. 1994. Integrated pest management of *Bemisia* in ornamental greenhouse production. *Phytoparasitica* 22(4): 356. [ Abstract]
437. Flanders, R., Orazo, M., Hancock, J. and Delfosse, E. 1993. [electronic bulletin board]. p. 136. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
438. Fletcher, D., Brown, J.K. and Wilson, F.D. 1993. Biolistic inoculation to evaluate cotton breeding lines for resistance to cotton leaf crumple virus and a whitefly-transmitted geminivirus affecting cotton in Guatemala. *Phytopathology* 83: 691. [ Abstract]
439. Flint, H.M. 1995. Garlic oil for whitefly control on cotton. p. 74. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
440. Flint, H.M., Hendrix, D., Salvucci, M. and Crafts-Brandner, S. 1997. The effects of daily removal of cotton blooms on plant growth and infestations of the silverleaf whitefly. p. 66. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
441. Flint, H.M., Naranjo, S.E., Henneberry, T.J. and Parks, N.J. 1994. Effects of cotton plant water stress on infestations by the sweetpotato whitefly. p. 161. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]

442. Flint, H.M., Naranjo, S.E., Leggett, J.E. and Henneberry, T.J. 1996. Effects of plant water stress on pest and beneficial insects infesting short and long staple cotton with emphasis on management of *Bemisia tabaci*. p. 159. In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
443. Flint, H.M., Wilson, F.D. and Naranjo, S. 1993. [cotton, water stress, pink bollworm]. p. 119. In Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
444. Flores, E. and Silberschmidt, K. 1963. Ability of single whiteflies to transmit concomitantly a strain of infectious chlorosis of Malvaceae and of *Leonurus* mosaic virus. Phytopathology 53: 238. [ Abstract, Cock (1986)]
445. Foltyn, S. and Gerling, D. 1984. Relationships among *Bemisia tabaci*, *Eretmocerus mundus* and *Encarsia lutea* under laboratory conditions. Phytoparasitica 12: 142. [ Abstract]
446. Forer, G. 1987. Development of *Bemisia tabaci* populations in the 1986 season. Phytoparasitica 15: 260. [ Abstract]
447. Forer, G. 1989. Whitefly populations in cotton - 1988. Phytoparasitica 17(3): 228. [ Abstract]
448. Forer, G. 1993. Control of *Bemisia tabaci* in cotton 1986-1992 and present status. Phytoparasitica 21(2): 171. [ Abstract]
449. Forer, G. and Gerling, D. 1984. Field activity of local parasitoids attacking *Bemisia tabaci* in the cotton field. Phytoparasitica 2: 143. [ Abstract]
450. Fox, C.A., Barioni, D., Trieber, T., Lochte, K., Tovar, M. and Pina, R. 1997. A demonstration of the effectiveness of 1% Detur for controlling WF pupae on containerized hibiscus. p. 116. In Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
451. Franco, M.I., Castillo-Garriga, A., Donoso, I. and Bejarano, E.R. 1999. Virus transmitted by whitefly: tomato yellow leaf curl virus (TYLCV), control strategies. p. 169. In Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce(Almeria), Spain. [Abstract]
452. Franz, E. and Carlton, J.B. 1993. [chemical control, spray deposits, airplane application, leaf washer, winglets]. p. 63. In Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
453. Freeman, T.P., Chu, C.C., Buckner, J.S., Nelson, D.R. and Henneberry, T.J. 1999. Variations in cotton leaf morphology related to whitefly feeding. p. 102. In Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
454. Freeman, T.P., Nelson, D.R., Buckner, J.S., Chu, C.C. and Henneberry, T.J. 2000. Determination of stylet length and extent of the stylet penetration of silverleaf whiteflies. p. 16. In Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
455. Freeman, T.P., Nelson, D.R., Buckner, J.S., Chu, C.C. and Henneberry, T.J. 2000. Mechanism and site of silverleaf whitefly stylet penetration. p. 17. In Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
456. Frisbie, R.E., Toscano, N.C., Stansly, P.A., Oetting, R.D. and Ellsworth, P.C. 1993. [sweetpotato whitefly action team, SWAT]. p. 137. In Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
457. Frohlich, D.R. and Brown, J.K. 1994. Development of molecular markers to facilitate population characterization of the whitefly, *Bemisia tabaci*. p. 50. In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
458. Frohlich, D.R. and Brown, J.K. 1994. Mitochondrial 16S Ribosomal subunit as a molecular marker in *Bemisia tabaci* and implications for population variability. Phytoparasitica 22(4): 311. [ Abstract]
459. Fu-Castillo, A.A. 1993. Analisis del uso de insecticidas por grupo toxicologico en cultivos hospedantes de mosquita blanca *Bemisia tabaci*, en la costa de Hermosillo, Son. [Crops attacked by the whitefly *Bemisia tabaci* and an analysis of the insecticides used according to toxicological group: Hermosillo coast, Sonora]. pp. 110-111. In J.M. Ramirez-Diaz (Ed.), Research Advances Ciano. Fall-Winter 1989-1990, Centro De Investigaciones Agricolas Del Noroeste. May 1993. [Spanish, Abstract]
460. Funk, C.J. and Salvucci, M.E. 2000. Localization of whitefly enzymes. p. 18. In Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
461. Galadima, A., Sanchez, C.A., Palumbo, J., Tickes, B., Matheron, M. and McGiffen, M. 2000. Preliminary evaluation of organic desert vegetable production systems. Hortscience 35(3): 508. [ Abstract]
462. Galves, G.E., Temple, S.R. and Yoshii, K. 1979. Lineas resistentes de frijol (*Phaseolus vulgaris* L.) al virus del mosaico dorado del frijol (BGMV). [Bean (*Phaseolus vulgaris*) resistant lines to golden mosaic virus (BGMV)]. Fitopatol. Colombiana 8(1): 27-28. [ Spanish, Abstract]
463. Gamez, R. 1974. Cambios en el desarrollo de plantas de frijol (*Phaseolus* spp.) asociados con la infeccion del virus del mosaico dorado. [Changes in development of bean plants (*Phaseolus* spp) associated with golden mosaic virus infection]. Fitopatologia 9(2): 38. [ Spanish, Abstract]
464. Gawel, N.J. and Bartlett, A.C. 1993. [randomly amplified polymorphic DNA (RAPD), RAPD-PCR, strains, DNA]. p. 31. In Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
465. Gawel, N. J. and Bartlett, A.C. 1994. DNA variation among populations of *Bemisia tabaci* Gennadius (Homoptera: Aleyrodidae). p. 51. In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
466. Gelman, D.B., Blackburn, M.B. and Hu, J.S. 2000. Ecdysteroid regulation of molting in 4th instars of the greenhouse (*Trialetrodes vaporariorum*) and silverleaf (*Bemisia argentifolii*) whiteflies. p. 19. In Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
467. Gelman, D.B., Blackburn, M.B., Hu, J.S. and Bentz, J. 1999. Characterization of fourth instar greenhouse whiteflies, *Trialetrodes vaporariorum* developmental markers and ecdysteroid fluctuations. p. 32. In Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
468. Geraud, F., Arcila, S. and Pena, M. 1991. Dinamica poblacional de la mosca blanca del tabaco *Bemisia tabaci* Gennadius (Homoptera: Aleyrodidae), en melon, en el Cebollal, Municipio Miranda, estado Falcon, Venezuela. [Population dynamics of the tobacco white fly *Bemisia tabaci* Gennadius (Homoptera: Aleyrodidae), on melons, in El Cebollal, Municipality Miranda, state Falcon, Venezuela]. Rev. Facultad Agron., Univ. Zulia (Venezuela) 8(4): 228. [ Spanish, Abstract]

469. Geraud, F., Chirinos, D., Rivero, G. and Sanchez, B. 1991. Avances en el manejo integrado de plagas del tomate en la Region Noroccidental del estado Zulia, Venezuela. [Advances in the integrated pest management of the tomato in the northwestern region of the state Zulia, Venezuela]. Rev. Facultad Agron., Univ. Zulia (Venezuela) 8(4): 224-225. [ Spanish, Abstract]
470. Geraud, F., Rivero, G., Chirinos, D. and Sanchez, B. 1991. Dinamica poblacional de la mosca blanca del tabaco *Bemisia tabaci* Gennadius en tomate, en la Region Noroccidental del estado Zulia, Venezuela. [Population dynamics of the tobacco white fly *Bemisia tabaci* Gennadius in tomatoes, in the northwestern region of the Zulia state, Venezuela]. Rev. Facultad Agron., Univ. Zulia (Venezuela) 8(4): 225-226. [ Spanish, Abstract]
471. Gerling, D. 1983. Overwintering of *Bemisia tabaci* in Israel. Phytoparasitica 11(1): 65. [ Abstract]
472. Gerling, D. 1989. Effects of the use of insect growth regulators on percent parasitism of *Bemisia tabaci* in cotton. Phytoparasitica 17(3): 233. [ Abstract]
473. Gerling, D., Foltyn, S. and Horowitz, A.R. 1982. Biology and field ecology of *Bemisia tabaci* and its natural enemies. Phytoparasitica 10: 293. [ Abstract]
474. Gerling, D. and Fried, R. 1996. [*Eretmocerus mundus*]. p. 109. In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
475. Gerling, D. and Guershon, M. 1999. Whitefly progeny production vs. mortality factors: ideas for future research. p. 12. In Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
476. Gerling, D., Guershon, M., Orion, T., Namies, N. and Reese, S. 1998. Population dynamics of whiteflies on wild hosts in Israel. p. 9. In Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
477. Gerling, D. and Henneberry, T.J. 1998. The status of *Bemisia* as a cotton pest: past trends and future possibilities. p. 170. In U. Kechagia (Ed.), World Cotton Research Conference 2. Sept. 6-12, 1998. Athens Greece. [Abstract]
478. Gerling, D. and Horowitz, A.R. 1983. Flight of adult *Bemisia tabaci* as determined in yellow trap catches. Phytoparasitica 11: 64. [ Abstract]
479. Gerling, D. and Kravchenko, V. 1994. Integrated pest management for the control of *Bemisia* attacking field crops outdoors. Phytoparasitica 22(4): 354. [ Abstract]
480. Gerling, D., Kravchenko, V. and Guershon, M. 1997. *Deraeocoris pallens* Reuter, *Orius niger* Wolf and *Campilomma unicolor* Poppius (Heteroptera) as predators of *Bemisia* nymphs. p. 140. In Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
481. Gerling, D. and Or, R. 1984. Influence of the host plant on the oviposition strategy of *Bemisia tabaci*. Phytoparasitica 2: 142. [ Abstract]
482. Gilbertson, R.L. 2000. A new bipartite geminivirus (Begomovirus) causing cucurbit leaf curl and crumpling symptoms in the Imperial Valley of California. p. 40. In Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
483. Gilbertson, R.L., Hou, Y.M., Guzman, P. and Carvalho, M.G. 1995. A new geminivirus associated with chlorosis of tomatoes in northwestern Mexico. Phytopathology 85(10): 1138. [ Abstract]
484. Gill, R.J. 1993. [morphology]. p. 32. In Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
485. Godfrey, L.D. and Goodell, P.B. 1996. Sticky trap sampling of silverleaf whitefly adults in the San Joaquin Valley [CA]. p. 5. In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
486. Godfrey, L.D., Goodell, P.B., Perring, T.M., Bellows, T.S., Summers, C.G., Bentley, W.J., Prather, T. and Coviello, R. 1995. Incidence of parasitism of silverleaf whitefly in the San Joaquin Valley [CA]. p. 105. In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
487. Godfrey, L.D., Goodell, P.B., Summers, C.G., Bentley, W.J., Prather, T. and Coviello, R. 1994. Contributions of crop and weed hosts to silverleaf whitefly populations in the San Joaquin Valley. p. 15. In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
488. Godfrey, L.D., Goodell, P.B., Summers, C.G., Bentley, W.J., Prather, T., Coviello, R., Perring, T.M. and Bellows, T.S. 1995. Seasonal development of silverleaf whitefly populations on crop and weed hosts in the San Joaquin Valley [CA]. p. 8. In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
489. Godfrey, L.D., Goodell, P.B., Summers, C.G., Bentley, W.J., Prather, T., Coviello, R., Perring, T.M. and Bellows, T.S. 1996. Seasonal dynamics of silverleaf whitefly on crop and weed hosts in the San Joaquin Valley [CA]. p. 4. In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
490. Godfrey, L.D. and Wood, J.P. 1994. Intraplant distribution of silverleaf whitefly on acala cotton. p. 14. In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
491. Godfrey, L.D., Wood, J.P., Goodell, P.B. and Eckert, J. 1997. Spatial and temporal distribution of the silverleaf whitefly in the San Joaquin Valley. p. 67. In Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
492. Godfrey, L.D. and Wynholds, P.F. 1995. Susceptibility of San Joaquin Valley [CA] acala cotton varieties to silverleaf whitefly. p. 147. In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
493. Gokkes, M. 1993. *Bemisia tabaci* in floriculture - present status. Phytoparasitica 21(2): 170. [ Abstract]
494. Gonzalez, R.A., Goldman, G.E., Natwick, E.T., Rosenberg, H.R., Grieshop, J.I., Sutter, S.R., Funakoshi, T. and Davila-Garcia, S. 1993. [damage: dollar losses, agricultural employment]. p. 12. In Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
495. Goodell, P.B. 1996. Extension efforts in national whitefly outreach and delivery. p. 181. In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
496. Goodell, P.B., Godfrey, L.D., Bentley, W.J., Coviello, R., Summers, C.G., Toscano, N.C., Gilbertson, R.L., Pickel, C. and Flint, M.L. 1994. Coordinating whitefly extension activities in the San Joaquin Valley. p. 177. In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]

497. Goodell, P.B., Godfrey, L.D., Flint, M.L., Bentley, W.J., Coviello, R., Ellsworth, P.C. and Dennehy, T. 1995. Development and delivery of a whitefly train-the-trainer program. p. 157. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
498. Goolsby, J.A. and Camplis, J.V. 1998. Conservation of *Bemisia* natural enemies in Mexican okra fields. p. 56. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
499. Goolsby, J.A. and Ciomperlik, M.A. 1997. Novel method for field delivery of parasitoids in row crops. p. 144. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
500. Goolsby, J.A. and Ciomperlik, M.A. 1998. Field evaluation of banker plants for field delivery of parasitoids in cucurbit crops. p. 57. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
501. Goolsby, J.A. and Jones, W. 1998. Survey for exotic parasitoids in the Lower Rio Grande Valley of Texas using sentinel plants. p. 59. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
502. Goolsby, J.A., Kirk, A. and Wendel, L.E. 1998. Importation of exotic natural enemies for *Bemisia tabaci* (Biotype "B"). p. 58. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
503. Goolsby, J.A., Legaspi, J.C. and Legaspi, B.C., Jr. 1996. Quarantine screening of natural enemies. p. 112. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
504. Goolsby, J.A. and Wendel, L.E. 1996. Establishment evaluation of exotic parasitoids in the Lower Rio Grande Valley and Wintergarden of Texas. p. 110. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
505. Goolsby, J.A. and Wendel, L.E. 1996. Importation of exotic natural enemies for *Bemisia tabaci* (Biotype "B"). p. 111. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
506. Goolsby, J.A. and Wendel, L.E. 1997. Colonization of exotic parasitoids in the Lower Rio Grande Valley and Wintergarden of Texas. p. 141. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
507. Goolsby, J.A. and Wendel, L.E. 1997. Importation of exotic natural enemies for *Bemisia tabaci* (Biotype "B"). p. 143. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
508. Gouge, D.H., Henneberry, T.J. and Reaves, L.L. 1996. Parasitism of *Bemisia argentifolii* (Homoptera: Aleyrodidae) by entomopathogenic nematodes in the families Steinernematidae and Heterorhabditidae. p. 113. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
509. Gould, J. 1999. Evaluating the impact of whitefly natural enemies established during the classical biological control program. pp. 2-3. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
510. Gould, J., Antilla, L. and Whitlow, M. 1998. Evaluation of the compatibility between insect growth regulators and natural enemies in controlling whitefly populations. p. 112. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
511. Gould, J., Antilla, L. and Whitlow, M. 1998. Integration of IPM and biocontrol in a multiple cropping system. p. 113. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
512. Gould, J. and Merten, P. 2000. Comparison of four methods of releasing whitefly parasitoids: Emergence, survival, dispersal, and mating success. p. 85. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
513. Gould, J. and Naranjo, S. 1996. Distribution and sampling of *Bemisia argentifolii* and *Eretmocerus* parasites on cantaloupe vines. p. 6. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
514. Gould, J., Waldner, D., Colletto, N., Antilla, L. and Santangelo, R. 1998. Release of exotic parasitoids for establishment in Arizona. p. 60. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
515. Granillo, C.R., Anaya, M. and Diaz, A. 1974. Virus diseases of sweet pepper in El Salvador. *Phytopathology* 64: 768. [Abstract, Cock (1986)]
516. Granillo, C.R., Diaz, A. and Anaya, M. 1974. The mosaic virus of kenaf (*Hibiscus cannabinus*) in El Salvador. *Phytopathology* 64: 768. [Abstract, Cock (1986)]
517. Greenberg, S.M., Jones, W.A., Legaspi, B.C., Jr. and Warfield, W.C. 1999. The effect of varying ratios of *Bemisia argentifolii* (Homoptera: Aleyrodidae) and *Eretmocerus mundus* (Hymenoptera: Aphelinidae) on parasitism. p. 71. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
518. Greenberg, S.M., Jones, W.A., Legaspi, B.C., Jr. and Warfield, W.C. 1999. Interaction between *Encarsia pergandiella* (Hymenoptera: Aphelinidae) and its host *Bemisia argentifolii* (Hymenoptera: Aleyrodidae): effects of parasitoid densities and host-parasitoid ratios. p. 70. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
519. Greenberg, S.M., Jones, W.A. and Liu, T.X. 2000. Tritrophic interactions between two species of whiteflies and two species of *Eretmocerus* (Hymenoptera: Aphelinidae) on tomato. p. 88. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
520. Greenberg, S.M., Jones, W.A. and Warfield, W.C. 1998. Comparative evaluation of host instar suitability of *Bemisia argentifolii* (Homoptera: Aleyrodidae) for the parasitoids *Eretmocerus mundus* and *Encarsia pergandiella* (Hymenoptera: Aphelinidae). p. 61. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]

521. Greenberg, S.M., Jones, W.A. and Warfield, W.C. 1998. Host instar suitability of *Bemisia argentifolii* (Homoptera: Aleyrodidae) for the parasitoid *Eretmocerus mundus* (Hymenoptera: Aphelinidae). p. 63. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
522. Greenberg, S.M., Jones, W.A. and Warfield, W.C. 1998. Host instar suitability of *Bemisia argentifolii* (Homoptera: Aleyrodidae) for the parasitoid *Encarsia pergandiella* (Hymenoptera: Aphelinidae). p. 62. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
523. Greenberg, S.M., Jones, W.A. and Warfield, W.C. 1999. Comparative host plant effects on the biologies of *Bemisia* and *Trialeurodes*. p. 13. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
524. Greenberg, S.M., Jones, W.A. and Warfield, W.C. 1999. Effects of host plant and whitefly species on parasitoid biology. p. 72. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
525. Greenberg, S.M., Legaspi, B.C. and Jones, W.A. 2000. Temperature effects on host mortality and parasitoid survival. p. 87. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
526. Greenberg, S.M., Legaspi, B.C., Jr. and Jones, W.A. 2000. Comparison of functional response and mutual interference between two aphelinid parasitoids of *Bemisia argentifolii* (Homoptera: Aleyrodidae). p. 86. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
527. Greenberg, S.M., Sappington, T., Liu, T.X. and Elzen, G.W. 2000. Preliminary data of the effects of cotton defoliant chemicals on *Bemisia argentifolii* mortality and its parasitoid survival. p. 137. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
528. Grinstein, A., Gan-Mor, S., Kletter, E., Spenser, J., Forer, G., Aharonson, N., Guershon, M., Gerling, D., Navo, D., Riven, Y. and Veierov, D. 1998. A new technology for improved pesticide coverage on cotton canopy: Part II - Field efficacy. p. 262. *In* U. Kechagia (Ed.), World Cotton Research Conference 2. Sept. 6-12, 1998. Athens Greece. [Abstract]
529. Gruenhagen, N.M., Perring, T.M. and Bellows, T.S., Jr. 1995. Searching behavior of *Eretmocerus* sp. on glabrous and hirsute melon varieties. p. 107. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
530. Gruenhagen, N.M., Perring, T.M., Bellows, T.S., Jr. and Farrar, C.A. 1995. Alteration of the silverleaf whitefly's host-plant environment to influence parasitism. p. 106. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
531. Guershon, M. and Gerling, D. 1993. The variable behavior of the coccinellid predator *Delphastus pusillus* resulting from exposure to nymphs of the whitefly *Bemisia tabaci* growing on hairy vs non-hairy leaves. *Phytoparasitica* 21(2): 172. [Abstract]
532. Guershon, M. and Gerling, D. 1994. Insect-plant interactions as related to phenotypic variation of *Bemisia tabaci* nymphs. *Phytoparasitica* 22(4): 312. [Abstract]
533. Guershon, M. and Gerling, D. 1995. Tritrophic relationships involving *Bemisia* and *Delphastus pusillus*. p. 108. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
534. Guirao, P., Cervera, M.T., Moya, A., Cabezas, J.A., Beitia, F., Martinez-Zapater, J.M. and Cenis, J.L. 1999. Genetic relationship of biotypes of *Bemisia tabaci* (Homoptera: Aleyrodidae) present in Spain based on RAPDS and AFLPS. p. 84. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce(Almeria), Spain. [Abstract]
535. Gupta, G.P. and Harris, M.K. 1994. Utilization of biopesticides in the management of whitefly populations. *Phytoparasitica* 22(4): 359. [Abstract]
536. Hagler, J. 1998. A laboratory study of the prey preference of five predator species on the various whitefly life stages. p. 64. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
537. Hagler, J., Jackson, G. and Gould, J. 1999. Evaluation of parasitoid dispersal patterns by mark-release-recapture. p. 73. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
538. Hagler, J., Jackson, G., Gould, J. and Ciomperlik, M. 1998. A simple protein marking ELISA to quantify parasitoid dispersal. p. 65. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
539. Hagler, J. and Naranjo, S. 1995. Characterizing and estimating the effect of the native predator complex of sweetpotato whitefly. p. 109. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
540. Hagler, J. and Naranjo, S. 1996. Evaluating the feeding behavior of commercial and indigenous *Hippodamia convergens* on sweetpotato whitefly: a laboratory and field study. p. 115. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
541. Hagler, J. and Naranjo, S. 1998. Feeding behavior of whitefly predators exposed to insect growth regulators and conventional insects. p. 66. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
542. Hagler, J., Palting, J. and Enriquez, J. 1996. A monoclonal antibody to *Bemisia tabaci* nymphal antigen: a tool for predator gut analysis. p. 114. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
543. Hagler, J.R. and Naranjo, S.E. 1993. Serological study of predators of sweetpotato whitefly eggs. p. 98. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
544. Hagler, J.R. and Naranjo, S.E. 1994. A serological analysis of predators of the sweetpotato whitefly egg stage. p. 125. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
545. Haider, M.S., Nadeem, A., Evans, A.A.F. and Markham, P.G. 2001. Serological relationships of whitefly-transmitted geminiviruses in and around cotton fields in Punjab, Pakistan. *Phytopathology* 91(Suppl. 6): S35. [Abstract]
546. Haji-Zadeh, J. and Kamali, K. 1995. Observation on mating and oviposition behaviours of *Stethorus gilvifrons* Mulsant. p. 299. *In* Proceedings 12th Iranian Plant Protection Congress, 2-7-September 1995, Karadj, Iran. [Persian, Abstract]

547. Hallan, V., Saxena, S. and Singh, B.P. 1998. Yellow net of *Triumffeta* is caused by a geminivirus: a first report. *Plant Dis.* 82(1): 127. [ Abstract]
548. Hameed, S., Naqvi, S.M.S. and Khalid, S. 1996. Genetic variation among whitefly population. p. 7. *In* M.J. Iqbal and A. Kasana (Eds.), Second International Congress of Entomological Sciences, March 19-21, 1996 Islamabad, Pakistan. [Abstract]
549. Hanif-Khan, S., Bullock, R.C., Stoffella, P.J., Brecht, J.K., Powell, C.A. and McAuslane, H.J. 1996. Symptom development and susceptibility of dwarf cherry tomato to tomato irregular ripening, a fruit disorder induced by silverleaf whitefly. *Hortscience* 31(4): 678. [ Abstract]
550. Hanif-Khan, S., Stoffella, P.J., Brecht, J.K., McAuslane, H.J., Bullock, R.C., Powell, C.A. and Yokomi, R. 1997. Plant growth regulators (GA-3 and CCC) and silverleaf whitefly effects on the induction of tomato irregular ripening in dwarf cherry tomato. *Hortscience* 32(3): 510. [ Abstract]
551. Harris, K., Pesic-Van Esbroeck, Z. and Duffus, J.E. 1994. A morphological study of *Bemisia* organ systems of known importance in homopteran virus transmission. *Phytoparasitica* 22(4): 323-324. [ Abstract]
552. Harris, K.F., Pesic-Van Esbroeck, Z. and Duffus, J.E. 1996. Morphological bases for whitefly transmission of viruses. p. 34. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
553. Harris, K.F., Pesic-Van Esbroeck, Z. and Duffus, J.E. 1996. Preparation of whole insects for combined light and electron microscopy and immunocytochemistry. p. 35. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
554. Harrison, B.D. 1994. Methods for the detection and identification of geminiviruses in plants and vector whiteflies. p. 15. *In* B. Ezzahiri and M. Bouhache (Eds.), Fifth Arab Congress of Plant Protection, November 27 - December 2, 1994, Fez, Morocco. Actes Editions, Rabat, Maroc. [Abstract]
555. Headrick, D., Bellows, T., Perring, T. and Orr, B. 1997. Natural enemy releases against the silverleaf whitefly in the Imperial Valley. p. 145. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
556. Headrick, D.H., Bellows, T.S. and Perring, T.M. 1995. Behavior of *Eretmocerus* sp. nr. *californicus* females attacking *Bemisia argentifolii* on two native California weeds. p. 110. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
557. Headrick, D.H., Bellows, T.S. and Perring, T.M. 1995. Behaviors of female *Eretmocerus* sp. nr. *californicus* (Hymenoptera: Aphelinidae) attacking *Bemisia argentifolii* (Homoptera: Aleyrodidae) on sweet potato, *Ipomoea batatas* (Convolvulaceae). p. 111. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
558. Headrick, D.H., Bellows, T.S. and Perring, T.M. 1995. Female behaviors of *Eretmocerus* sp. nr. *californicus* (Hymenoptera: Aphelinidae) attacking *Bemisia argentifolii* (Homoptera: Aleyrodidae) on cotton, *Gossypium hirsutum* (Malvaceae) and melon, *Cucumis melo* (Cucurbitaceae). p. 112. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
559. Headrick, D.H., Bellows, T.S., Jr. and Perring, T.M. 1994. Searching and parasitism behavior of silverleaf whitefly parasites. p. 126. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
560. Headrick, D.H., Orr, B., Bellows, T.S. and Perring, T.M. 1995. Introduction of natural enemies attacking *Bemisia argentifolii*. p. 113. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
561. Heilmann, L.J. 1997. Use of satellite DNA sequences as species-specific probes for *Eretmocerus* and *Encarsia* parasitoids. p. 87. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
562. Heilmann, L.J. 1998. Satellite DNAs as identification probes for *Encarsia* and *Eretmocerus* wasps. p. 67. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
563. Heinz, K.M. 1994. Predators and parasitoids as biological control agents of *Bemisia* in greenhouses. *Phytoparasitica* 22(4): 336. [ Abstract]
564. Heinz, K.M., Brazzle, J.R. and Pickett, C. 1995. Evaluations of field releases of *Eretmocerus* nr. *californicus* in San Joaquin Valley California cotton. p. 114. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
565. Heinz, K.M. and Parrella, M.P. 1994. Biological control of *Bemisia tabaci* infesting greenhouse poinsettia. p. 127. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
566. Heinz, K.M., Parrella, M.P., Hennessey, R.D. and Wendel, L.E. 1994. Behavioral comparisons of two strains of *Encarsia formosa* as potential biological control agents of *Bemisia tabaci*. p. 128. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
567. Heinz, K.M. and Zalom, F. 1994. The influence of tomato cultivar on *Bemisia tabaci* biological control. p. 129. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
568. Heinz, K.M. and Zalom, F.G. 1993. [tomato, breeding, natural enemy, predators], p. 13. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
569. Helman, S., Peterlin, O. and Contreras, M. 1994. Parasitoids of *Bemisia tabaci* in cotton in Santiago del Estero, Northwestern Argentina. *Phytoparasitica* 22(4): 337. [ Abstract]
570. Hendrix, D.L. 1993. [honeydew, sugars, enzyme]. p. 33. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
571. Hendrix, D.L., Blackledge, B., Steele, T. and Perkins, H.H. 1994. Sweetpotato whitefly honeydew analysis and enzyme degradation. p. 52. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
572. Hendrix, D.L., Salvucci, M.E. and Wolfe, G.R. 1998. Polyol metabolism in *Bemisia argentifolii*. p. 10. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
573. Hendrix, D.L. and Steele, T. 1994. *Bemisia* honeydew. *Phytoparasitica* 22(4): 320. [ Abstract]

574. Hendrix, D.L., Wei, Y., Steele, T. and Salvucci, M.E. 1996. Isolation and characterization of the large oligosaccharides in *Bemisia* honeydew. p. 36. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
575. Hendrix, D.L., Wei, Y.A. and Steele, T. 1997. Oligosaccharides in *Bemisia* honeydew. p. 88. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
576. Henneberry, T.J. 1999. Ecologically oriented management strategies for *Bemisia* in agricultural systems. pp. 78-79. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce(Almeria), Spain. [Abstract]
577. Henneberry, T.J., Bentley, W., Chu, C.C., Ellsworth, P., Goodell, P., Nichols, R.L., Naranjo, S.E., Riley, D.G., Toscano, N. and Watson, T. 1995. Progress in developing adult action thresholds for chemical control of silverleaf whitefly (SLW). p. 9. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
578. Henneberry, T.J., Blackledge, B., Perkins, H.H., Jr., Hendrix, D.L., Nichols, R.L. and Steele, T. 1997. Reducing cotton lint stickiness. p. 68. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
579. Henneberry, T.J., Forlow Jech, L., Hendrix, D. and Steele, T. 2000. Silverleaf whitefly honeydew production and honeydew sugar relationships to sticky cotton. p. 21. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
580. Henneberry, T.J., Forlow Jech, L., Hendrix, D.L. and Steele, T. 2000. Silverleaf whitefly honeydew, honeydew sugars and sticky cotton. p. 20. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
581. Henneberry, T.J., Forlow Jech, L. and Perkins, H.H., Jr. 1996. Silverleaf whitefly and sticky cotton relationships. p. 69. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
582. Henneberry, T.J., Hendrix, D.L., Perkins, H.H., Jr. and Flint, H.M. 1996. Silverleaf whitefly honeydew and sticky cotton relationships. p. 70. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
583. Henneberry, T.J., Hendrix, D. L., Perkins, H.H., Jr. and Naranjo, S.E. 1996. Silverleaf whitefly populations and relationships to sticky cotton and cotton yields. p. 71. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
584. Henneberry, T.J., Jech, L.F., Burke, R.A., Panter, M.J. and Faulconer, S.F. 1998. Honeydew produced by whitefly adults and nymphs sampled from untreated and insecticide-treated cotton plants. p. 13. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
585. Henneberry, T.J., Jech, L.F. and Hendrix, D.L. 1998. Seasonal distribution of *Bemisia argentifolii* (Homoptera: Aleyrodidae) honeydew sugars on cotton lint. p. 11. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
586. Henneberry, T.J., Jech, L.F., Hendrix, D.L. and Brushwood, D.E. 1998. Lint stickiness and *Bemisia argentifolii* (Homoptera: Aleyrodidae) populations. p. 12. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
587. Henneberry, T.J., Jech, L.F., Hendrix, D.L., Perkins, H.H., Jr. and Brushwood, D. 1997. Silverleaf whiteflies, cotton stickiness, and yields in long and short staple cottons. p. 117. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
588. Henneberry, T.J., Jech, L.F., Naranjo, S.E., Flint, H.M. and Akey, D.H. 1994. Sweetpotato whitefly populations and cotton lint yield. p. 16. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
589. Henneberry, T.J., Jech, L.F. and Perkins, H. 1993. [honeydew, minicard]. p. 13. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
590. Henneberry, T.J., Jones, W.A. and Faust, R. M. 2000. Brief history, research progress, and current pest status of *Bemisia* in the United States. pp. 1-2. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
591. Hennessey, R.D. 1993. [distribution, parasites, *Encarsia*, *Eretmocerus*]. p. 99. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
592. Hennessey, R.D. 1994. Whitefly parasite surveys. p. 130. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
593. Hernandez Hernandez, J. 1991. Evaluacion de barreras de sorgo para el manejo de insectos vectores en chile jalapeno. [Evaluation of sorghum barriers for vector insects management in chilli "Jalapeno"]. p. 140. *In* 4th Annual Meeting of the National Institute of Forestry, Husbandry and Agricultural Investigation at Veracruz State, Mexico. INIFAP. [Spanish, Abstract]
594. Hernandez-Jasso, A. and Pacheco-Covarrubias, J.J. 1998. Respuesta de cultivares de algodono a la mosquita blanca de la hoja plateada. [Response of cotton cultivars to silverleaf whitefly]. p. 140. *In* 17th Phytogetic Congress. Proceedings. Scientific Notes, Acapulco, Mexico, 5-9 Oct 1998. [Spanish, Abstract]
595. Herold, F. 1967. Investigation of a virus disease of *Anthurium andraeanum*. *Phytopathology* 57: 8. [Abstract, Cock (1986)]
596. Herzog, G.A., Sumner, H.R., Chandler, L.D., Severson, R.F. and Stephenson, M.G. 1993. [chemical control, Solonaceae]. p. 64. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
597. Herzog, G.A., Sumner, H.R., Chandler, L.D., Womac, A.R., Mulrooney, J.E. and Howard, K.D. 1993. [chemical control, electrostatic sprayer, hydraulic sprayer, Degania sprayer, Berthoud sprayer, Hydrapak sprayer]. p. 65. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
598. Hidayat, S.H., Gilbertson, R.L., Hanson, S.F., Morales, F.J., Ahlquist, P., Russell, D.R. and Maxwell, D.P. 1993. Complete nucleotide sequences of the infectious cloned DNAs of bean dwarf mosaic geminivirus. *Phytopathology* 83: 2. [Abstract]
599. Hiebert, E., Abouzid, A. and Polston, J.E. 1994. Whitefly-transmitted geminiviruses. *Phytoparasitica* 22(4): 324. [Abstract]

600. Hilje, L. 1996. Reduction of whitefly adult numbers and delay of geminivirus dissemination in tomatoes, by living soil covers, in Costa Rica. p. 160. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
601. Hilje, L., Cubillo, D. and Sanabria, G. 1997. Delay of geminivirus dissemination in tomato fields by interfering with *Bemisia tabaci* adults. p. 180. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
602. Hilje, L., Cubillo, D. and Sanabria, G. 1999. Bitterwood (*Quassia amara*) extracts kill *Bemisia tabaci* adults. p. 50. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
603. Hilje, L. and Stansly, P.A. 1999. Effectiveness of living ground covers for managing spread of geminiviruses in tomato by *Bemisia tabaci* in Costa Rica. p. 122. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
604. Hilje, L. and Stansly, P.A. 2000. Living ground covers are effective for managing whitefly-vectored geminiviruses in tomatoes. p. 138. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
605. Hoddle, M.S., van Driesche, R.G., Lyon, S.M. and Sanderson, J.P. 2000. Compatibility of selected insect growth regulating insecticides with the whitefly parasitoid *Eretmocerus eremicus* for control of *Bemisia argentifolii* on poinsettias. p. 139. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
606. Hoelmer, K., Gould, J. and Roltsch, W. 1996. Field cage evaluations of non-indigenous parasitoids in desert crops. p. 116. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
607. Hoelmer, K. and Hagler, J. 1996. Preliminary biological studies of *Semidalis* sp., a native neuropteran predator of *Bemisia*. p. 117. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
608. Hoelmer, K., Hagler, J. and Jackson, C.G. 1998. Continuing studies of *Semidalis* sp., a native predator of *Bemisia* in Desert AZ and CA. p. 69. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
609. Hoelmer, K.A. 1993. [*Delphastus*, *Encarsia*, *Eretmocerus*, *Nephaspis*]. p. 100. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
610. Hoelmer, K.A. 1994. Evaluation of indigenous and exotic natural enemies of *Bemisia* in southwestern desert agroecosystems. p. 131. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
611. Hoelmer, K.A. 1994. Parasitoids of whiteflies: their potential as controlling agents of outdoor populations of *Bemisia* spp. *Phytoparasitica* 22(4): 337-338. [Abstract]
612. Hoelmer, K.A. 1995. Introductions and evaluation of exotic parasitoids in Southwestern Deserts [USA]. p. 115. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
613. Hoelmer, K.A. 1997. 1996 field cage evaluations of non-indigenous parasitoids in desert crops. p. 146. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
614. Hoelmer, K.A. 1997. Parasitism of silverleaf whitefly in alfalfa. p. 148. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
615. Hoelmer, K.A. 1998. Comparative field cage evaluations of top-performing introduced parasitoids in desert cantaloupes. p. 68. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
616. Hoelmer, K.A. and Culver, G. 1997. Survey of desert host plants for whiteflies and parasitoids. p. 149. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
617. Hoelmer, K.A., Pickett, C.H. and Abel, W. 1999. Evaluation of citrus as an overwintering host of *Eretmocerus* parasitizing *Bemisia*. p. 74. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
618. Hoelmer, K.A. and Roltsch, W.J. 1994. Coniopterygids (Neuroptera) noted as predators of *Bemisia* in the Imperial Valley, CA. p. 132. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
619. Hoelmer, K.A., Roltsch, W.J. and Chu, C.C. 1997. Comparisons of the selectivity of whitefly traps for parasitoids of silverleaf whiteflies. p. 181. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
620. Hoelmer, K.A., Roltsch, W.J., Gould, J. and Simmons, G.S. 1997. Overwintering & dispersal of non-indigenous parasitoids released in CA & AZ desert valleys. p. 147. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
621. Hoelmer, K.A., Roltsch, W.J. and Simmons, G.S. 1998. Establishment of introduced *Eretmocerus* species in Imperial Valley, CA. p. 70. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
622. Hoelmer, K.A. and Simmons, A.M. 1999. Yellow sticky trap catches of *Bemisia* parasitoids and their relation to field populations. p. 75. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
623. Höfer, P., Höhnle, M., Bedford, I.D., Markham, P.G. and Frischmuth, T. 1999. Coat protein gene replacement results in whitefly-transmission of an insect non-transmissible geminivirus isolate. p. 37. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce (Almería), Spain. [Abstract]
624. Horowitz, A.R., Forer, G. and Ishaaya, I. 1996. [Controlling *Bemisia* with novel insecticides]. p. 72. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
625. Horowitz, A.R. and Ishaaya, I. 1994. Chemical control of *Bemisia tabaci* - management and application. *Phytoparasitica* 22(4): 345-346. [Abstract]



626. Horowitz, A.R., Mendelson, Z. and Ishaaya, I. 1993. Managing resistance to growth regulators in *Bemisia tabaci* in Israel. *Phytoparasitica* 21(2): 179. [ Abstract]
627. Horowitz, A.R., Motro, U. and Gerling, D. 1979. Natural mortality of the tobacco whitefly (*Bemisia tabaci*) in Israel cotton fields. p. 503. *In* Berger, H. (Ed.), Proceedings International Symposium of IOBC/WPRS on Integrated Control in Agriculture and Forestry, Vienna, Oct. 8-12, 1979. International Organization for Biological Control, Vienna, Austria. [Abstract, Cock (1986)]
628. Horowitz, A.R., Podoler, H. and Gerling, D. 1982. Population dynamics (life tables) of *Bemisia tabaci* under field conditions. *Phytoparasitica* 10: 294. [ Abstract]
629. Horowitz, A.R. and Toscano, N.C. 1989. Insecticide resistance in *Bemisia tabaci*: Synergism as a potential control approach. *Phytoparasitica* 17(3): 232. [ Abstract]
630. Houck, M.A. 1993. [morphometric, morphology], p. 34. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
631. Houk, M.S. and Hoefert, L.L. 1983. Ultrastructure of *Chenopodium* leaves infected by lettuce infectious yellows virus. *Phytopathology* 73: 790. [ Abstract, Cock (1986)]
632. Hu, J.S. and Gelman, D.B. 2000. Development of *Encarsia formosa* in the silverleaf whitefly, *Bemisia argentifolii*: Effect of host age. p. 89. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
633. Hu, J.S., Lius, S., Barry, K., Wu, Z.C., Wang, M. and Hamasaki, R.T. 1994. First report of a geminivirus in Hawaii. *Plant Dis.* 78(6): 641. [ Abstract]
634. Hu, J.S. and Ullman, D.E. 1995. Detection and characterization of geminiviruses in Hawaii. p. 38. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
635. Hunter, M.S., Collier, T.R. and Kelly, S.E. 1999. Interference competition between a primary parasitoid and an autoparasitoid on *Bemisia tabaci*. p. 76. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
636. Hunter, M.S. and Johnson, S.K. 1997. Interactions of an exotic autoparasitoid and a native primary parasitoid: preliminary investigations. p. 150. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
637. Hunter, W., Persad, A., Inbar, M., Doostdar, H. and Mayer, R.T. 1999. Geminivirus-mediated interspecific competition between whiteflies and other insects. p. 33. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
638. Hunter, W.B., Hiebert, E., Webb, S., Polston, J. and Tsai, J. 1995. Whitefly morphology (Homoptera: Aleyrodidae) and tomato mottle geminivirus. *Phytopathology* 85(10): 1181. [ Abstract]
639. Hunter, W.B. and Polston, J.E. 1996. Transmission of geminivirus by the whitefly, *Bemisia tabaci* (Genn.). *Phytopathology* 86(11 SUPPL.): S115. [ Abstract]
640. Idris, A.M., Banks, G. and Brown, J.K. 1994. Development of a diagnostic assay for whitefly-transmitted geminiviruses using PCR. *Phytopathology* 84: 1086. [ Abstract]
641. Idris, A.M. and Brown, J.K. 1995. PCR-based detection of geminiviruses in two whitefly vectors. *Phytopathology* 85(10): 1184. [ Abstract]
642. Idris, A.M. and Brown, J.K. 1997. Biological and molecular detection of subgroup III geminiviruses in plants and to whitefly vectors. *Phytopathology* 87(6 SUPPL.): S46. [ Abstract]
643. Idris, A.M. and Brown, J.K. 1997. Frequencies of whitefly-mediated transmission and PCR detection of geminivirus DNA in two whitefly biological types. p. 89. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
644. Idris, A.M., Fletcher, D.C. and Brown, J.K. 1993. Identification and partial characterization of Sinaloa tomato leaf curl virus (STLCV), a new whitefly-transmitted geminivirus affecting tomato and pepper from Sinaloa, Mexico. *Phytopathology* 83: 692. [ Abstract]
645. Idris, A.M., Rivas-Platero, G., Torres-Jerez, I. and Brown, J.K. 1999. First report of Sinaloa tomato leaf curl geminivirus in Costa Rica. *Plant Dis.* 83(3): 303. [ Abstract]
646. Inbar, M., Doostdar, H., Leibe, G.L. and Mayer, R.T. 1998. Asymmetric interspecific competition between whiteflies and leafminers. p. 14. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
647. Inbar, M., Doostdar, H. and Mayer, R.T. 1997. Artificial induction of plant defensive mechanisms may affect whitefly populations. p. 90. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
648. Inbar, M., Doostdar, H. and Mayer, R.T. 1998. Local and systemic effects of whiteflies on tomato photosynthesis and phytochemistry. p. 15. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
649. Isaacs, R. and Byrne, D.N. 1996. Testing the hypothesis that flight behavior of *Bemisia tabaci* is influenced by host plant water stress. p. 8. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
650. Isaacs, R. and Byrne, D.N. 1997. Host location flight behavior of *Bemisia*. p. 69. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
651. Isaacs, R. and Byrne, D.N. 1998. Aerial distribution of *Bemisia*. p. 16. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
652. Isaacs, R., Byrne, D.N. and Desi, S. 1999. Behavioral and developmental effects of trichome defenses in *Datura wrightii* on *Bemisia*. p. 103. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
653. Isaacs, R., Cahill, M. and Byrne, D.N. 1998. Host-evaluation behaviors of *Bemisia* and their modification by systemically-applied imidacloprid. p. 92. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
654. Isaacs, R., Hagler, J.R., Byrne, D.N. and Machtley, S.A. 1996. Whiteflies on the World Wide Web. p. 182. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
655. Isakeit, T., Robertson, N.L., Brown, J.K. and Gilbertson, R.L. 1994. First report of squash leaf curl virus on watermelon in Texas. *Plant Dis.* 78(10): 1010. [ Abstract]

656. Ishaaya, I. and Ascher, K.R.S. 1983. Synergized pyrethroids against *Bemisia tabaci*. *Phytoparasitica* 11: 67. [ Abstract]
657. Ishaaya, I. and Ascher, K.R.S. 1984. Synergized cypermethrin for controlling the whitefly, *Bemisia tabaci*, in cotton. *Phytoparasitica* 12: 139-140. [ Abstract]
658. Ishaaya, I., Ascher, K.R.S. and Casida, J.E. 1983. Pyrethroid synergism by esterase inhibitors in cotton pests. p. 230. *In* 10th International Congress of Plant Protection; Proceedings of a Conference Held at Brighton, England 20-25 November 1983. Plant Protection for Human Welfare. British Crop Protection Council, Croydon, UK. [Abstract, Cock (1986)]
659. Ishaaya, I., Mendelson, Z. and De Cock, A. 1989. Insect growth regulators for controlling the whitefly *Bemisia tabaci*: biological aspects and agricultural importance. *Phytoparasitica* 17(3): 232-233. [ Abstract]
660. Ishaaya, I., Mendelson, Z. and Horowitz, A.R. 1993. Buprofezin, pyriproxyfen and diafenthiuron suppress growth and development of the whitefly, *Bemisia tabaci*. *Phytoparasitica* 21(2): 176-177. [ Abstract]
661. Jackson, D.M. and Danehower, D.A. 1996. Integrated case study: Tobacco leaf surface components and their effects on insect pests and diseases. *J. Exp. Bot.* 47 (Suppl.): 53. [ Abstract]
662. Jackson, D.M. and Farnham, M.W. 1996. Field evaluation of collard genotypes for resistance to whiteflies. p. 161. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
663. Jackson, D.M., Farnham, M.W. and Simmons, A.M. 1999. Effects of interplanting of collard phenotypes on resistance to whiteflies. p. 104. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
664. Jackson, D.M., Simmons, A.M., Chortyk, O.T. and Stephenson, M.G. 1996. Production of biorational insecticides by *Nicotiana* species. p. 73. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
665. Jackson, D.M., van Giessen, W.A. and Farnham, M.W. 1997. Is breeding for resistance to silverleaf whitefly a viable pest management approach? p. 182. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
666. James, R.R. 2000. Effect of certain nutrients on germination of whitefly pathogen spores. p. 90. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
667. James, R.R. and Elzen, G. 1999. Integrating the biocontrol agent *Beauveria bassiana* with imidacloprid. p. 51. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
668. Jaronski, S.T. and Hoelmer, K. 1995. Impact of *Beauveria bassiana* Mycotech BB401 and *Paecilomyces fumosoroseus* Mycotech PFR612 on natural enemies of silverleaf whitefly in spring cantaloupe and cotton in Imperial, Valley, CA. p. 39. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
669. Jaronski, S.T., Lord, J.C. and Paden, R. 1996. *Bemisia argentifolii* control in melons with Mycotrol WP. p. 74. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
670. Jaronski, S.T., Rosinska, J., Brown, C., Osterlind, R. and Staten, R. 1997. Impact of *Beauveria bassiana* Mycotech strain GHA, buprofezin, and pyriproxyfen on whitefly predators in Arizona cotton. p. 118. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
671. Jaronski, S.T., Wood, P. and Underwood, N. 1995. Assessment of *Beauveria bassiana* Mycotech BB401 and *Paecilomyces fumosoroseus* Mycotech PFR612 efficacy against silverleaf whitefly in Imperial Valley, California. p. 75. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
672. Javan Moghaddam, H. 1993. Aspect of *Bemisia tabaci* Gennad. in Iran and world. p. 300. *In* Proceedings 11th Plant Protection Congress of Iran, 28-Aug.-2-Sept., 1993, Rasht, Iran. [Persian, Abstract]
673. Javan Moghaddam, H. and Noori, P. 1995. Comparative estimation of damage caused by *Bemisia tabaci* Gennad. in different varieties of cotton. p. 102. *In* Proceedings 12th Iranian Plant Protection Congress, 2-7-September 1995, Karadj, Iran. [Persian, Abstract]
674. Javan Moghaddam, H. and Noori, P. 1995. Population fluctuation of *Bemisia tabaci* Genn. on Varamin cotton Cultivar, in the Varamin region. p. 105. *In* Proceedings 12th Iranian Plant Protection Congress, 2-7-September 1995, Karadj, Iran. [Persian, Abstract]
675. Jayaraj, S., Uthamasamy, S., Thangaraju, D. and Rangarajan, A.V. 1988. Studies on monitoring key pests of cotton for integrated management. p. 52. *In* National Workshop on Pest and Disease Surveillance for IPM, 16-17 September. Indian Council of Agricultural Research. [Abstract]
676. Jech, L. 1999. Summary of standardized survey of whitefly in the Gila Basin near Gila Bend, AZ. p. 123. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
677. Jemsi, Gh. and Javanzadeh, M. 1995. Comparison of some insecticides for control of *Bemisia tabaci* in Khuzestan. p. 347. *In* Proceedings 12th Iranian Plant Protection Congress, 2-7-September 1995, Karadj, Iran. [Persian, Abstract]
678. Jiang, Y.X., De Blas, C., Barrios, L. and Fereres, A. 1999. Correlation between whitefly feeding behavior and tomato yellow leaf curl virus transmission. pp. 44-45. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce(Almeria), Spain. [Abstract]
679. Jiang, Y.X., De Blas, C. and Muñoz, M. 1999. Identification of some common weeds as reservoirs for tomato yellow leaf curl virus transmitted by *Bemisia tabaci* (Gennadius). pp. 82-83. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce(Almeria), Spain. [Abstract]
680. Jiang, Y.X., Nombela, G. and Muñoz, M. 2000. Analysis by DC electrical penetration graphs of the resistance to *Bemisia tabaci* (Homoptera: Aleyrodidae) on two near isogenic tomato lines. p. 114. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
681. Johnson, D.D. and Walker, G.P. 1997. Correlation of intracellular punctures produced by adult *Bemisia argentifolii* with an AC electronic feeding monitor waveform. p. 91. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]

682. Johnson, D.D., Walker, G.P. and Creamer, R. 1997. Identification of the *Bemisia argentifolii* feeding behavior that results in inoculation of lettuce chlorosis virus. p. 92. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
683. Johnson, M.W. 1995. Natural enemies associated with silverleaf whitefly in Hawaii. p. 116. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
684. Jones, W.A. 1993. [*Eretmocerus*, *Encarsia*, lantana, melon, cole, greenhouse, *Deraeocoris*, natural enemies]. p. 101. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
685. Jones, W.A. 1994. Biology and behaviour of *Eretmocerus* sp. nr. *californicus* from Texas. p. 133. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
686. Jones, W.A. 1995. Reproductive biology of *Eretmocerus* sp. from Texas. p. 117. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
687. Jones, W.A. and Bellows, T.S., Jr. 1995. A simple technique for using excised leaves for studying the biology, behavior and interactions of biological control agents of *Bemisia* spp. p. 118. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
688. Jones, W.A., Carruthers, R.I. and Nordlund, D.A. 1993. Biological control of the sweetpotato whitefly *Bemisia tabaci* (Homoptera: Aleyrodidae) in the Lower Rio Grande Valley, Texas. p. 222. *In* XXVIII Congreso Nacional de Entomología, Universidad de las Americas, Cholula, Puebla 23-26 May 1993. [Abstract]
689. Jones, W.A., Garcia, R. III, Goolsby, J.A., Legaspi, B.C., Jr., Warfield, W.C. and Talekar, N.S. 1997. Evaluation of *Eretmocerus* spp. on tomato. p. 152. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
690. Jones, W.A., Greenberg, S.M. and Warfield, W.C. 1998. Comparative evaluation of host instar suitability of *Bemisia argentifolii* (Homoptera: Aleyrodidae) for the parasitoids *Eretmocerus mundus* and *Encarsia pergandiella* (Hymenoptera: Aphelinidae). p. 71. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
691. Jones, W.A. and Monk, J.T. 1995. Fate of *Bemisia argentifolii* larvae following oviposition and host feeding by *Eretmocerus* sp. p. 119. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
692. Jones, W.A. and Poprawski, T.J. 1996. *Bemisia argentifolii* parasitized by *Eretmocerus* sp. is immune to infection by *Beauveria bassiana*. p. 118. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
693. Jones, W.A., Silva, P. and Snodgrass, G. 1997. Biology of *Deraeocoris nebulosus* (Heteroptera: Miridae) on *Bemisia*. p. 151. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
694. Jones, W.A., Warfield, W.C., Ciomperlik, M. and Rose, M. 1996. Temperature effects on foraging efficiency of four native and exotic parasitoids. p. 121. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
695. Jones, W.A., Warfield, W.C., Ciomperlik, M., Rose, M. and Goolsby, J. 1996. Comparative biology of a uniparental and biparental *Eretmocerus* spp. p. 120. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
696. Jones, W.A., Wolfenbarger, D.A. and Ciomperlik, M. 1995. Insecticide effects on immatures of native and imported *Eretmocerus* spp. p. 120. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
697. Jones, W.A., Wolfenbarger, D.A. and Kirk, A.A. 1994. Response of native and exotic parasitoid adults to insecticide residues. p. 134. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
698. Jones, W.A. and Wraight, S.P. 1996. Effects of fungal pathogen applications in vegetables on the foraging activity of native parasitoids. p. 119. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
699. Joyce, A. and Bellows, T. 1997. Field cage evaluation of *Amitus bennetti* (Hymenoptera: Platygasteridae), a parasitoid of *Bemisia argentifolii*. p. 153. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
700. Kasana, A. and Hashmi, A.A. 1996. Efficacy of some insecticides against *Bemisia tabaci*. p. 67. *In* M.J. Iqbal and A. Kasana (Eds.), Second International Congress of Entomological Sciences, March 19-21, 1996 Islamabad, Pakistan. [Abstract]
701. Kays, S.J. and Severson, R.F. 1993. [sweetpotato, plant breeding]. p. 119. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
702. Kern, J., Cohen, S., Harpaz, I. and Ben-Joseph, R. 1987. The influence of physical barriers on tomato yellow leaf curl virus epidemics and on plant populations of *Bemisia tabaci*. *Phytoparasitica* 15: 261-262. [Abstract]
703. Kerns, D.L. and Palumbo, J.C. 1996. Using Admire on desert vegetable crops. p. 73. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
704. Khambay, B.P.S., Batty, D., Cahill, M., Denholm, I.A., Niemeyer, H.M., Mead-Briggs, M., Vinall, S. and Simmonds, M.S.J. 1996. Naturally occurring resistance-defeating pesticides. *Proc. Am. Chem. Soc.* 212((1/2)): 21. [Abstract]
705. Kirk, A. and Lacey, L. 1995. Foreign exploration for natural enemies of *Bemisia tabaci*/*Bemisia argentifolii*. p. 121. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
706. Kirk, A. and Lacey, L. 1996. Efficacy of *Clitostethus arcuatus* a predator of *Bemisia argentifolii*. p. 122. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]

707. Kirk, A., Lacey, L., Goolsby, J., Ciomperlik, M., Able, W. and Pickett, C. 1996. Results of foreign exploration for parasitoids of SLWF. p. 124. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
708. Kirk, A., Lacey, L., Goolsby, J., Vacek, D., Schauff, M. and Nakahara, S. 1995. Drought adapted natural enemies of *Bemisia argentifolii* from Thailand. p. 122. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
709. Kirk, A., Lacey, L., Legaspi, S.C. and Carruthers, R. 1996. Foreign exploration for silverleaf whitefly natural enemies. p. 123. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
710. Kirk, A., Lacey, L. and Thistlewood, H. 1997. Exploration for natural enemies and evaluation of *Paecilomyces fumosoroseus* and the coccinellid *Clitostethus arcuatus*. p. 154. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
711. Kirk, A.A. and Lacey, L.A. 1994. Foreign exploration for natural enemies of *Bemisia tabaci*. *Phytoparasitica* 22(4): 335-336. [Abstract]
712. Kirk, I.W., Bouse, L.F., Carlton, J.B., Franz, E. and Latheef, M.A. 1993. [airplane application, cantaloupe, cotton]. p. 66. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
713. Klassen, V.A., Boeshore, M., Koonin, E.V., Tian, T. and Falk, B.W. 1994. Lettuce infectious yellows virus: a bipartite closterovirus transmitted by *Bemisia tabaci* and representative of a new genus of plant viruses. *Phytoparasitica* 22(4): 327. [Abstract]
714. Kletter, E. 1993. Threshold level for *Bemisia tabaci* on cotton. *Phytoparasitica* 21(2): 178-179. [Abstract]
715. Knauf, T.A. 1995. Control of silverleaf whitefly in greenhouse tomatoes with Naturalis-L. p. 123. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
716. Knauf, T.A. 1995. Control of whiteflies and mites in ornamentals with Troy Biosciences EXP 7744. p. 124. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
717. Knauf, T.A. and Wright, J.E. 1994. Evaluation of Naturalis\* L for control of sweetpotato whitefly in tomato fields in Florida. p. 135. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
718. Knauf, T.A. and Wright, J.E. 1994. Fermone Exp 7744: A biorational insecticide for whitefly control. p. 136. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
719. Knauf, T.A. and Wright, J.E. 1995. Evaluation of Naturalis-L for control of whitefly and other cotton insects. A synopsis of 1993 and 1994 trials in Arizona, Texas, Louisiana, and Mississippi. p. 125. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
720. Kring, J.B., Schuster, D.J., Price, J.F. and Simone, G.W. 1991. Sweetpotato whitefly -vecteded geminivirus on tomato in Florida. *Plant Dis.* 75(11): 1186. [Abstract, Cock (1993)]
721. Lacey, L. and Kirk, A. 1993. [natural enemies, *Encarsia*, *Eretmocerus*, *Paecilomyces*]. p. 102. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
722. Lacey, L., Kirk, A., Vey, A., Bolckman, K., Mercadier, G. and Vidal, C. 1995. The effect of host plant on activity of *Paecilomyces fumosoroseus*. p. 126. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
723. Lacey, L. and Kirk, A.A. 1994. Foreign exploration for natural enemies of *Bemisia tabaci* and related activities. p. 138. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
724. Lacey, L.A., Carruthers, R. and Fransen, J.J. 1994. Global distribution of naturally occurring fungi of *Bemisia tabaci* s.l. and their potential as natural and biological control agents. *Phytoparasitica* 22(4): 342. [Abstract]
725. Lacey, L.A. and Fransen, J.J. 1994. Fungi as biological control agents of *Bemisia tabaci* s.l. *Phytoparasitica* 22(4): 342-343. [Abstract]
726. Lacey, L.A. and Kirk, A.A. 1994. Foreign exploration for natural enemies of *Bemisia tabaci*. p. 137. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
727. Lacey, L.A., Kirk, A.A. and Akey, D.H. 1996. Experimental application of the fungus, *Paecilomyces fumosoroseus* using an irrigation system in a greenhouse for the control of *Bemisia argentifolii* on cabbage. p. 125. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
728. Lapidot, M., Ben-Joseph, R., Friedmann, M., Pilowsky, M. and Cohen, S. 2000. The effect of TYLCV-resistant tomato plants on virus epidemiology. p. 41. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
729. Lapidot, M., Friedmann, M., Cohen, S. and Pilowsky, M. 1999. The effect of tomato yellow leaf curl virus on new breeding lines with high levels of resistance to the virus. p. 74. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce(Almeria), Spain. [Abstract]
730. Larios, J.F., Fischner, F. and Bonilla, P. 1979. Interrelationship between the activity of whiteflies (*Bemisia tabaci* Genn.) and the incidence of infectious chlorosis virus in cotton in El Salvador. *Fitopatol. Colombiana* 8(1): 29. [Abstract]
731. Larios, J.F., Sandoval, O. and Lara, W.E. 1979. Analysis of the spread of infectious chlorosis virus from wild-weeds-host to cotton fields. *Fitopatol. Colombiana* 8(1): 28. [Abstract]
732. Latheef, M.A., Bouse, L.F. and Kirk, I.W. 1994. Aerial spray efficacy studies for sweetpotato whitefly control in cotton. p. 91. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
733. Latheef, M.A., Carlton, J.B. and Kirk, I.W. 1996. Aerial electrostatic charged sprays for control of sweetpotato whitefly in cotton. p. 76. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
734. Latheef, M.A. and Wolfenbarger, D. 1995. Response of silverleaf whitefly to spray droplet characteristics of Danitol plus Orthene on cotton. p. 76. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]

735. Latheef, M.A. and Wolfenbarger, D.A. 1996. Toxicity of mixtures of fenpropathrin and bifenthrin with Acephate against resistant and susceptible strains of sweetpotato whitefly. p. 77. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
736. Lazare, M. and Gerling, D. 1993. The population dynamics of natural enemies of *Bemisia tabaci* in cotton fields and the influence of insecticide sprays. *Phytoparasitica* 21(2): 171-172. [Abstract]
737. Lebiush-Mordechi, S., Frigida, D. and Berlinger, M.J. 1994. Proposed IPM strategy for greenhouses in warm countries. *Phytoparasitica* 22(4): 355-356. [Abstract]
738. Legaspi, B.C., Jr., Smits, N., Carruthers, R.I. and Hunter, M.S. 1994. Object-oriented simulation modeling in the biological control of *Bemisia tabaci*. p. 178. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
739. Legaspi, J.C., Carruthers, R.I., Nordlund, D.A., Correa, J.A., Cohen, A.C. and Holbrook, D. 1994. Effect of inundative releases of the predator, *Chrysoperla rufilabris* (Neuroptera) to control sweetpotato whitefly in an organic field crop. p. 139. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
740. Legaspi, J.C., Hadman, J., Carruthers, R.I., Legaspi, B.C., Everitt, J., Escobar, D., Anderson, G., Wendel, L., Davidson, J., Riley, D. and Murden, D. 1994. Cooperative research, implementation, and assessment project for sweetpotato whitefly biological control in the Lower Rio Grande Valley of Texas. p. 18. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
741. Legaspi, J.C., Legaspi, B.C., Jr., Correa, J.A. and Meagher, R.L., Jr. 1996. Evaluation of *Serangium parcesetosum* (Coccinellidae) as a biological control agent of the silverleaf whitefly. p. 126. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
742. Legaspi, J.C., Nordlund, D.A. and Carruthers, R.I. 1993. [*Chrysoperla*]. p. 103. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
743. Legaspi, S.C., Carruthers, R.I., Legaspi, B.C., Jr. and Poprawski, T.J. 1995. Foreign exploration and evaluation of some natural enemies of *Bemisia argentifolii* from Southeast Asia. p. 127. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
744. Legg, J. and Fishpool, L. 1994. Yellow sticky traps to monitor *Bemisia tabaci*, vector of African cassava mosaic geminiviruses in Uganda. *Phytoparasitica* 22(4): 319. [Abstract]
745. Legg, J.P. and Okao-Okuja, G. 1999. Progress in the diagnosis and epidemiological characterisation of cassava mosaic geminiviruses in East Africa. pp. 74-75. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce(Almeria), Spain. [Abstract]
746. Leggett, J.E. 1993. [mating, biotypes]. p. 34. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
747. Leggett, J.E. and Antilla, L. 1995. The effectiveness of mist blower in reducing whitefly numbers. p. 77. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
748. Leggett, J.E., El-Lissy, O., Antilla, L. and Dobratz, L. 1994. Factors influencing whitefly distribution and comparison of sampling methods in cotton fields. p. 19. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
749. Leggett, J.E. and Elhoff, L. 1997. The use of suction traps to sample whitefly populations. p. 70. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
750. Leggett, J.E. and Elhoff, L.K. 1996. Comparative mating behavior of three whitefly species. p. 37. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
751. Leibe, G.L., Schuster, D.J. and Stansly, P.A. 1993. [insecticide resistance, tomato, cantaloupe, watermelon]. p. 67. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
752. Leija-Chapman, V., Ciomperlik, M. and Wendel, L. 1998. Biology and predation of *Serangium parcesetosum* (Coleoptera: Coccinellidae) on *Bemisia tabaci* (Gennadius) (biotype B) (Homoptera: Aleyrodidae). p. 72. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
753. Leon-Espinosa, F. 1978. Comportamiento de líneas de algodónero al ataque de *Heliothis zea* (Boddie) y *Bemisia tabaci* (Gennadius), en el Soconusco, Chis. [Behavior of cotton plants lines to the attack of *Heliothis zea* (Boddie) and *Bemisia tabaci* (Gennadius) in Soconusco, Chis]. *Folia Entomol. Mexicana* 39/40: 75-76. [Spanish, Abstract]
754. LeVesque, C.S., Perring, T.M., Moore, B.K., Cooper, A. and Walling, L.L. 1998. Impact of silverleaf whitefly feeding on tomato fruit physiology. p. 93. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
755. LeVesque, C.S., Perring, T.M. and Walling, L.L. 1994. Silverleaf whitefly-induced tomato irregular ripening. p. 53. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
756. LeVesque, C.S., Perring, T.M. and Walling, L.L. 1997. Differential gene expression: a comparison of the silverleaf whitefly and sweetpotato whitefly using differential RNA display. p. 93. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
757. LeVesque, C.S., Perring, T.M., Walling, L.L. and James, A.A. 1995. Characterization of sex-specific gene expression in silverleaf and sweetpotato whiteflies by differential display. p. 40. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
758. LeVesque, C.S., Perring, T.M., Walling, L.L. and Polek, M. 1995. Induction of tomato irregular ripening by silverleaf whitefly feedings. p. 41. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
759. Li, T.X. and Stansly, P.A. 1995. Susceptibility of *Encarsia pergandiella* adults (Hymenoptera: Aphelinidae), parasitoid of *Bemisia argentifolii* (Homoptera: Aleyrodidae), to some selected insecticides on tomato and sweet potato leaves. p. 129. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]

760. Liedl, B.E., Lawson, D.M., White, K.K., Shapiro, J.A., Carson, W.G., Trumble, J.T. and Mutschler, M.A. 1995. Acylsugars of the wild tomato (*Lycopersicon pennellii* (Corr.) D'Arcy) alters settling and reduces oviposition of *Bemisia argentifolii* (silverleaf whitefly). *Hortscience* 30(4): 827. [Abstract]
761. Lima, L.H.C., Navia, D. and Oliveira, M.R.V. 1999. Occurrence and evaluation of *Bemisia tabaci* (Gennadius) (Hemiptera: Aleyrodidae) strains in Brazil using PCR-RAPD. p. 14. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
762. Lindquist, R.K. 1993. [biological control, *Encarsia*, poinsettia, *Paecilomyces*]. p. 104. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
763. Lindquist, R.K. 1993. [chemical control, insect growth regulator, biorational, poinsettia, chrysanthemum]. p. 68. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
764. Liu, H.Y., Duffus, J.E. and Cohen, S. 1993. [mating, isozyme, sweetpotato, broccoli, melon]. p. 35. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
765. Liu, T.X. 1999. Management of *Bemisia argentifolii* with application of biorational insecticides and imidacloprid on cantaloupe in spring in South Texas. p. 52. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
766. Liu, T.X. 2000. Efficacy and persistence of Platinum, Actara and Admire for management of silverleaf whitefly on cantaloupe in South Texas. p. 65. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
767. Liu, T.X. 2000. Population dynamics of silverleaf whitefly on spring collard and relationship to yield in the Lower Rio Grande Valley of Texas. p. 140. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
768. Liu, T.X. 2000. Screening cantaloupe varieties for whitefly resistance. p. 115. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
769. Liu, T.X., Crosby, K., Miller, M., Gregg, L. and Hernandez, R. 2000. Cucurbit yellow stunting disorder virus on melon in the Lower Rio Grande Valley of Texas: Blame to *Bemisia*? p. 42. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
770. Liu, T.X. and Stansly, P.A. 1994. Repellency of biorational insecticides to *Bemisia tabaci*. p. 93. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
771. Liu, T.X. and Stansly, P.A. 1994. Toxicity of biorational insecticides to *Bemisia tabaci*. p. 92. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
772. Liu, T.X. and Stansly, P.A. 1995. Oviposition by silverleaf whitefly on tomato: effects of leaf factors and insecticide residues. p. 42. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
773. Liu, T.X. and Stansly, P.A. 1995. Susceptibility of *Delphastus pusillus* (Coleoptera: Coccinellidae), a predator of *Bemisia argentifolii* (Homoptera: Aleyrodidae), to some selected biorational insecticides. p. 128. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
774. Liu, T.X. and Stansly, P.A. 1996. Development of *Encarsia pergandiella* (Hymenoptera: Aphelinidae) in four instars of *Bemisia argentifolii* (Homoptera: Aleyrodidae). p. 128. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
775. Liu, T.X. and Stansly, P.A. 1996. Oviposition and survivorship of *Encarsia pergandiella* (Hymenoptera: Aphelinidae) in four instars of *Bemisia argentifolii* (Homoptera: Aleyrodidae). p. 127. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
776. Liu, T.X. and Stansly, P.A. 1996. Pupal orientation and emergence of some aphelinid parasitoids of *Bemisia argentifolii* (Homoptera: Aleyrodidae). p. 129. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
777. Liu, T.X. and Stansly, P.A. 1997. Deposition of tracer dye on paper labels for evaluation of spray coverage. p. 119. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
778. Liu, T.X. and Stansly, P.A. 1997. Functional response of *Nephaspis oculatus* (Coleoptera: Coccinellidae) to egg density of *Bemisia argentifolii*. p. 156. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
779. Liu, T.X. and Stansly, P.A. 1997. Insecticidal effects of surfactants on *Bemisia argentifolii*. p. 120. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
780. Liu, T.X. and Stansly, P.A. 1997. Use of *Beauveria bassiana*, an entomopathogen to control nymphs of *Bemisia argentifolii* under field, laboratory and greenhouse conditions. p. 155. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
781. Liu, T.X. and Stansly, P.A. 1998. Efficacy of *Beauveria bassiana*, entomopathogen of *Bemisia argentifolii* nymphs on hibiscus under two humidity regimes. p. 73. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
782. Liu, T.X. and Stansly, P.A. 1999. Searching and feeding behaviour of *Nephaspis oculatus* and *Delphastus catalinae* (Coleoptera: Coccinellidae), predators of *Bemisia argentifolii* (Homoptera: Aleyrodidae). p. 77. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
783. Liu, T.X., Stansly, P.A. and Chortyk, O.T. 1996. Bioassay of insecticidal activity of natural and synthetic sugar esters against *Bemisia argentifolii* (Homoptera: Aleyrodidae). p. 78. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]

784. Liu, T.X., Stansly, P.A. and Chortyk, O.T. 1996. Insecticidal activity of natural and synthetic sugar esters against *Bemisia argentifolii* (Homoptera: Aleyrodidae) on field tomato plants. p. 79. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
785. Liu, T.X., Stansly, P.A., Sparks, A.N., Jr., Knowles, T.C. and Chu, C.C. 2000. Managing *Bemisia argentifolii* using Mycotrol and Naturalis-L (*Beauveria bassiana*) on vegetables, cotton and ornamentals in southern United States. p. 91. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
786. Loniello, A.O., Martinez, R.T., Rojas, M.R., Gilbertson, R.L., Brown, J.K. and Maxwell, D.P. 1992. Molecular characterization of bean calico mosaic geminivirus. *Phytopathology* 82: 1149. [Abstract]
787. Lopez, R.L., Cervantes, M. and Sánchez, B. 1995. IPM actions and practices in cotton. p. 168. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
788. López, R.L., Cervantes, M., Sánchez, B. and Hoyos, F. 1996. IPM actions and practices in cotton (2nd year). p. 183. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
789. López, R.L., Sánchez, B. and Hoyos, F. 1997. IPM actions and practices in cotton (3rd year). p. 121. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
790. López, R.L. and Tafuya, R. 1997. Rescate 200 (Acetamiprid) performance against silverleaf whitefly in Mexicali Valley, Mexico. p. 122. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
791. Lotrakul, P., Valverde, R.A., Clark, C.A., Sim, J. and De La Torre, R. 1998. Identification of a geminivirus infecting sweetpotato in the United States. *Phytopathology* 88(9 suppl.): S55. [Abstract]
792. Louro, D., Noris, E., Veratti, F. and Accotto, G.P. 1996. First report of tomato yellow leaf curl virus in Portugal. *Plant Dis.* 80(9): 1079. [Abstract]
793. Lublinkhof, J. and Odom, P.N. 1996. Buprofezin (Applaud), a new IGR for whitefly control. p. 80. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
794. Lynch, R.E. and Chamberlin, J.R. 1993. [peanut]. p. 120. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
795. Lynch, R.E. and Chamberlin, J.R. 1993. [peanut, within plant distribution]. p. 14. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
796. Magal, Z., Berlinger, M.J. and Benzioni, A. 1982. Influence of pH and sucrose content on the attraction of *Bemisia tabaci* in vivo and in vitro. *Phytoparasitica* 10: 294-295. [Abstract]
797. Mandal, B., Pappu, H.R. and Muniyappa, V. 2001. Characterization of Croton yellow vein mosaic virus, family Geminiviridae. *Phytopathology* 91(6 Suppl.): S57-S58. [Abstract]
798. Mansoor, S., Khan, S.H., Saeed, M., Bashir, A., Zafar, Y., Malik, K.A. and Markham, P.G. 1997. Evidence for the association of a bipartite geminivirus with tomato leaf curl disease in Pakistan. *Plant Dis.* 81(8): 958. [Abstract]
799. Maramorosch, K. 1996. Control of whitefly vectors and pests with neem tree natural products. *Phytopathology* 86(11 SUPPL.): S115. [Abstract]
800. Markham, P.G., Bedford, I.D., Briddon, R.W., Brown, J.K. and Costa, H.S. 1992. Biological diversity between *Bemisia tabaci* from different continents. Proc. XIX Int. Congress Entomol., Beijing, China June 28-July 4, 1992. [Abstract]
801. Markham, P.G., Bedford, I.D., Liu, S., Pinner, M. and Briddon, R.W. 1994. The transmission of geminiviruses with different biotypes of the *Bemisia tabaci* complex. *Phytoparasitica* 22(4): 324-325. [Abstract]
802. Martinez-Carrillo, J.L. 1995. Monitoring for resistance to insecticides in whitefly populations from the Yaqui Valley, Sonora, Mexico. p. 78. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
803. Martinez-Carrillo, J.L. 1996. Trends in resistance to insecticides in whitefly populations from the Yaqui Valley, Sonora, Mexico. p. 81. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
804. Martínez-Carrillo, J.L. 1996. Monitoreo de la resistencia a insecticidas en mosquita blanca del Valle del Yaqui, Sonora, México. p. 214. *In* Memorias 6th Congreso Internacional de Manejo Integrado de Plagas y 5th Taller Latinoamericano Sobre Moscas Blancas y Geminivirus, Acapulco, Mexico. [Abstract]
805. Martinez-Carrillo, J.L. and Pacheco-Covarrubias, J.J. 1997. Research review on silverleaf whitefly in Northwestern Mexico. p. 196. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
806. Martinez, Y., De Blas, C., Zabalgoceazcoa, I., Quinones, M., Castellanos, C., Peralta, E.L. and Romero, J. 1997. A bipartite geminivirus infecting tomatoes in Cuba. *Plant Dis.* 81(10): 1215. [Abstract]
807. Marutani, M., Yudin, L., Nafus, D., Cruz, F. and Santos, V. 1995. An outbreak of a new whitefly in Guam. *Hortscience* 30(4): 829. [Abstract]
808. Maruthi, M.N., Colvin, J. and Seal, S. 1999. The geographic distribution of cassava *Bemisia tabaci* biotypes in relation to the current epidemic of cassava mosaic disease in East Africa. p. 88. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce (Almeria), Spain. [Abstract]
809. Matyis, J.C., Silva, D.M., Oliveira, A.R. and Costa, A.S. 1976. Morphology of three white fly transmitted viruses.: Morfologia de tres virus transmitidos por *Bemisia tabaci*. *Fitopatologia* 11(1/2): 22. [Portuguese, Abstract]
810. Maw, B.W., Stephenson, M.G., Severson, R.F. and Eiteman, M.A. 1993. [Nicotiana]. p. 69. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
811. Mayberry, K.S. and Perring, T.M. 1992. The whitefly upsurge impact on California vegetable production. *HortScience* 27(6): 628. [Abstract]
812. Mayer, R.T. 2001. Whitefly-pathogen-host plant interactions: Possible involvement of plant defensive systems. *Am. Chem. Soc.* 222(1-2): Agro80. [Abstract]
813. Mayer, R.T., Inbar, M. and Doostdar, H. 1997. Pathogenesis-related proteins in tomato plants as a result of silverleaf whitefly feeding. p. 94. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]

814. Mayer, R.T., McCollum, T.G., McDonald, R.E., Polston, J.E., Schroeder, W.J., Sieburth, P.J. and Doostdar, H. 1996. Pant disorders resulting from *Bemisia* feeding. p. 38. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
815. Mazyad, H.M., Aboul-Ata, A.E., Sabik, A., El-Sayed, M., Peters, D., El-Nabawey, M. and Helal, M. 1994. Tomato yellow leaf curl virus in Egypt: epidemiological and management aspects. p. 182. *In* B. Ezzahiri and M. Bouhache (Eds.), Fifth Arab Congress of Plant Protection, November 27 - December 2, 1994, Fez, Morocco. Actes Editions, Rabat, Maroc. [Abstract]
816. McAuslane, H.J. 1996. Leaf pubescence as a factor in antixenosis resistance of soybean to silverleaf whitefly. p. 163. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
817. McAuslane, H.J. 1997. Parasitization of silverleaf whitefly by *Eretmocerus* sp (Hong Kong) on soybean isolines differing in pubescence. p. 159. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
818. McAuslane, H.J., Golub, R. and Schuster, D.J. 1996. Assessing resistance to silverleaf whitefly of tomato mutants for foliar pubescence. p. 162. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
819. McAuslane, H.J., Johnson, F.A. and Colvin, D.L. 1994. Parasitism of sweetpotato whitefly in peanut and soybean in Florida. p. 140. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
820. McAuslane, H.J., Johnson, F.A. and Knauff, D.A. 1993. [peanut, resistance, distribution], p. 121. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
821. McAuslane, H.J., Johnson, F.A. and Knauff, D.A. 1994. Evaluation of peanut breeding lines and cultivars for resistance to sweetpotato whitefly. p. 162. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
822. McAuslane, H.J., Johnson, F.A., Knauff, D.A. and Colvin, D.L. 1993. [peanut, parasite, *Encarsia*, *Eretmocerus*], p. 105. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
823. McAuslane, H.J., Johnson, F.A., Sojack, B.R. and Colvin, D.L. 1995. Parasitism of silverleaf whitefly in soybean isolines varying for foliar pubescence. p. 130. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
824. McAuslane, H.J., Johnson, F.A., Sojack, B.R. and Colvin, D.L. 1995. Resistance to silverleaf whitefly in soybean isolines varying for foliar pubescence. p. 148. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
825. McAuslane, H.J. and Nguyen, R. 1996. Reproductive behavior and biology of a thelytokous *Eretmocerus* sp. (Hong Kong). p. 130. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
826. McAuslane, H.J., Simmons, A.M. and Jackson, D.M. 1997. Parasitization of silverleaf whitefly by *Eretmocerus* sp (Hong Kong) on collard, *Brassica oleracea*, genotypes differing in leaf epidermal wax. p. 158. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
827. McAuslane, H.J., Webb, S.E. and Elmstrom, G.W. 1996. Resistance in germplasm of *Curcubita pepo* to squash silverleaf. p. 164. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
828. McCreight, J.D. 1993. [melon, resistance, ELISA], p. 122. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
829. McCreight, J.D. 1995. Repeatable protocol for evaluation of lettuce for reaction to silverleaf whitefly feeding. p. 149. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
830. McCreight, J.D. 1998. Resistance to lettuce infections yellow virus in melon. *Hortscience* 33(3): 533. [ Abstract]
831. McCreight, J.D., Elmstrom, G., Simmons, A.M. and Wolff, D. 1994. Melon variety trials. p. 163. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
832. McCutcheon, G.S. and Simmons, A.M. 2000. Influence of temperature on parasitism by an indigenous *Eretmocerus* species. p. 92. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
833. McGlashan, D., Polston, J.E. and Bois, D. 1994. Tomato yellow leaf curl geminivirus in Jamaica. *Plant Dis.* 78(12): 1219. [ Abstract]
834. McGrath, M.T., Gilrein, D. and Brown, J.K. 1994. First report of squash silverleaf disorder associated with B-biotype sweetpotato whitefly in New York. *Plant Dis.* 78(6): 641. [ Abstract]
835. McGrath, P.F., Craig, W. and Brown, J.K. 1997. Whitefly-transmitted geminivirus diseases of cotton: an international problem. p. 95. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
836. McKenzie, C.L., Lee, S.D., Doostdar, H., Kokalis-Buelle, N. and Mayer, R.T. 2000. Effect of plant growth promoting rhizobacteria on pathogenesis-related protein induction in cucurbits challenged with different levels of SLWF infestation. p. 116. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
837. McLain, J.M. and Creamer, R. 1996. Physicochemical and ecological characterization of lettuce chlorosis virus. *Phytopathology* 86(11 SUPPL.): S115. [ Abstract]
838. McMillan, R.T., Jr., Hiebert, E. and Abouzid, A.M. 1994. Epidemiology and etiology of BGMV in South Florida. p. 54. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
839. McPherson, R.M. and Severson, R.F. 1993. [soybean, breeding]. p. 123. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]



840. Medina, V., Theodoridis, K. and Markham, P.G. 1999. Geminiviruses: Intracellular pathway and transport in their insect vectors. pp. 36-37. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce(Almeria), Spain. [Abstract]
841. Mehta, P., Wyman, J.A., Nakhla, M.K. and Maxwell, D.P. 1994. Transmission studies of tomato yellow leaf curl geminivirus by the vector *B. tabaci* (Homoptera: Aleyrodidae). p. 63. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
842. Mehta, P., Wyman, J.A., Nakhla, M.K. and Maxwell, D.P. 1994. Use of the polymerase chain reaction to detect viruliferous individuals of *B. tabaci* (Homoptera: Aleyrodidae) with two tomato-infecting geminiviruses. p. 55. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
843. Mehta, P., Wyman, J.A., Nakhla, M.K. and Maxwell, D.P. 1995. Detection of two tomato-infecting geminiviruses in individual *Bemisia tabaci* (Homoptera: Aleyrodidae) adults. p. 43. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
844. Meiners, J.P., Lawson, R.H., Smith, F.F. and Diaz, A.J. 1973. Mechanical transmission of a whitefly-born disease agent of beans in El Salvador. *Phytopathology* 63: 803-804. [Abstract, Cock (1986)]
845. Meister, H. 1996. The effect of "DETUR" on silverleaf whitefly (SLWF) populations in the Imperial Valley of Southern California. p. 82. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
846. Melamed-Madjar, V., Hameiri, J. and Chen, M. 1987. Influence of aldicarb applied at different stages of cotton plant growth on the development of *Bemisia tabaci* populations. *Phytoparasitica* 15: 263. [Abstract]
847. Meyerdirk, D.E. and Coudriet, D.L. 1984. Population densities of *Bemisia tabaci* on cotton, alfalfa, and vegetables in Imperial Valley, California. p. 568. *In* XVII International Congress Entomology. [Abstract]
848. Meyerdirk, D.E., Hennessey, R.D., Wendel, L., Goolsby, J., Ciomperlik, M. and Vacek, D. 1994. Release of exotic natural enemies of *Bemisia tabaci* in the United States. *Phytoparasitica* 22(4): 335. [Abstract]
849. Miller, W.B., Peralta, M.E., Ellis, D.R. and Perkins, H.H., Jr. 1994. Stickiness of individual whitefly honeydew carbohydrates on cotton lint. p. 56. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
850. Minkenberg, O., Bernays, E.A. and Bright, K. 1993. Host plant preference, adult performance, and diet breadth in *Bemisia tabaci*. p. 36. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
851. Minkenberg, O., Kaltenbach, J., Leonard, C., Malloy, R., Simmons, G. and Ziegweid, K. 1994. Development of augmentative biological control of *Bemisia argentifolli* on field and greenhouse crops. *Phytoparasitica* 22(4): 338-339. [Abstract]
852. Minkenberg, O., Leonard, C., Santangelo, R., Zhang, C., Penn, S., Gould, J., Antilla, L. and Staten, R. 1997. Mass-rearing native and exotic *Eretmocerus* wasps for whitefly biological control in greenhouse and field crops. p. 161. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
853. Minkenberg, O., Simmons, G.S., van Schelt, J., Wilson, M.F., Natwick, E.T. and Weddle, R.C. 1993. Development of augmentative biological control of *Bemisia tabaci* on cotton using the parasitoid *Eretmocerus californicus*. p. 106. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
854. Minkenberg, O., Thacker, G.W. and Dennehy, T.J. 1997. Enabling biologically based IPM on cotton: impact of conventional insecticides vs. insect growth regulator buprofezin (Applaud) on parasitoid populations. p. 160. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
855. Minkenberg, O.P.J.M., Berens, R. and Palumbo, J.C. 1994. Field evaluation of insect fungi to control sweetpotato whitefly on lettuce and cole crops. p. 141. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
856. Minkenberg, O.P.J.M., Henter, H.J., Hall, J., Campos, R.M., Leonard, C., Malloy, R., Zhang, C. and Penn, S. 1996. Evaluation of a native *Eretmocerus* for inundative biological control of whitefly on greenhouse poinsettias. p. 131. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
857. Minkenberg, O.P.J.M., Henter, H.J., Kaltenbach, J., Leonard, C., Malloy, R., Tovar-Soto, J., Ziegweid, K., Greatrex, R. and Alcock, K.T. 1995. Evaluation of *Eretmocerus* nr. *californicus* ex Arizona for augmentative biological control of silverleaf whitefly on field and greenhouse crops. p. 131. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
858. Minkenberg, O.P.J.M., Malloy, R., Kaltenbach, J., Leonard, C., Grish, K., Greatrex, R. and Alcock, K.T. 1994. Mass-rearing of *Eretmocerus* nr. *californicus* for augmentative biological control of sweetpotato whitefly in field crops and greenhouse tomatoes. p. 142. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
859. Mirza, M.S. and Aslam, M. 1996. Resistance to yellow mosaic virus disease in soybean. p. 47. *In* M.J. Iqbal and A. Kasana (Eds.), Second International Congress of Entomological Sciences, March 19-21, 1996 Islamabad, Pakistan. [Abstract]
860. Moawad, G.M. and Gerling, D. 1998. Dynamics of whiteflies and their enemies in cotton fields. Implications for pest management. p. 199. *In* U. Kechagia (Ed.), World Cotton Research Conference 2. Sept. 6-12, 1998. Athens Greece. [Abstract]
861. Moghaddam, H.J. and Noori, P. 1998. Comparative estimation of population of *Bemisia tabaci* (Genn.) in different varieties of cotton. p. 178. *In* U. Kechagia (Ed.), World Cotton Research Conference 2. Sept. 6-12, 1998. Athens Greece. [Abstract]
862. Monsef, A.A. 1993. Important factors influencing population fluctuations of cotton whitefly. p. 114. *In* Proceedings 11th Plant Protection Congress of Iran, 28-Aug.-2-Sept., 1993, Rasht, Iran. [Persian, Abstract]
863. Monsef, A.A. and Amin, Gh. 1995. Some biology and ecology observations on *Eretmocerus mundus* (Mercet) a hymenopterous parasite of whitefly in Fars province. p. 298. *In* Proceedings 12th Iranian Plant Protection Congress, 2-7-September 1995, Karadj, Iran. [Persian, Abstract]
864. Moomaw, C., Woolley, J., Rose, M. and Riley, D. 1994. Evaluation of naturally occurring parasitic hymenoptera attacking *Bemisia tabaci* (Genn.) in Texas. p. 143. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]

865. Mor, U. 1983. Cotton yields and quality as affected by *Bemisia tabaci* under different regimes of irrigation and pest control. *Phytoparasitica* 11: 64. [ Abstract Cock (1993)]
866. Mor, U. 1987. *Bemisia tabaci* and cotton physiology: a 5-year summary of the influence of water-stressed plants on the pest population. *Phytoparasitica* 15: 261. [ Abstract]
867. Mor, U. 1994. The effect of *Bemisia tabaci* populations on dryland and irrigated cotton, on the lint stickiness content, and a solution for the detection of stickiness. *Phytoparasitica* 22(4): 321. [ Abstract]
868. Mor, U. and Marani, A. 1984. Relationships between physiology of the cotton plant and development of the tobacco whitefly, *Bemisia tabaci*. *Phytoparasitica* 12: 141. [ Abstract]
869. Mor, U., Marani, A. and Applebaum, S.W. 1982. The relationship between *Bemisia tabaci* populations and cotton physiological conditions. *Phytoparasitica* 10: 295. [ Abstract Cock (1993)]
870. Morales, F.J. and Klass, J. 1999. The ecology and dissemination of whitefly-transmitted viruses in Latin America. p. 77. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce(Almeria), Spain. [Abstract]
871. Muigai, S.G. and Schuster, D.J. 1997. Mechanisms of resistance of wild *Lycopersicon* germplasm to the silverleaf whitefly. p. 183. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
872. Mullins, W. 1994. Principles for the use of imidacloprid (Admire\*) for silverleaf whitefly control. p. 94. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
873. Muniyappa, V., Nateshan, H.M., Jalikop, S.H., Chennarayappa and Ramappa, H.K. 1994. Indian tomato leaf curl geminivirus: characterization and host resistance. *Phytoparasitica* 22(4): 331. [ Abstract]
874. Muñiz, M., Nombela, G. and Beitia, F. 1999. Variation in host response to *Bemisia tabaci* (Homoptera: Aleyrodidae) in tomato plants. p. 81. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce(Almeria), Spain. [Abstract]
875. Muñiz, M. and Rieche, Y. 1999. Settling behavior of *Bemisia tabaci* (Homoptera: Aleyrodidae) on some common weeds in Spain. p. 82. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce(Almeria), Spain. [Abstract]
876. Munthali, D.C. 1994. Field evaluation of three vegetable oils prepared as behavior-interfering formulations against whiteflies. *Phytoparasitica* 22(4): 351-352. [ Abstract]
877. Musana, A.C.Z. 1984. A technique for monitoring whitefly, *Bemisia tabaci* (Genn.) in cotton in Zimbabwe. p. 567. *In* XVII Int. Congress Entomol. [Abstract]
878. Mutschler, M.A., Cobb, E.D., Liedl, B.E. and Shapiro, J.A. 1995. Breeding for acylsugar-mediated multiple-pest resistance in cultivated tomato, *Lycopersicon esculentum*. *Hortscience* 30(4): 801. [ Abstract]
879. Nadeem, A., Nelson, M.R. and Orum, T.V. 1998. GIS and geostatistics: new tools for measuring and analyzing disease epidemics and insect populations in cotton. p. 236. *In* U. Kechagia (Ed.), World Cotton Research Conference 2. Sept. 6-12, 1998. Athens Greece. [Abstract]
880. Nadeem, A., Nelson, M.R. and Xiong, Z. 1994. Molecular characterization and comparison of cotton leaf crumple and cotton leaf curl geminiviruses. p. 179. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
881. Nakhla, M.K., Maxwell, D.P., Martinez, R.T., Carvalho, M.G. and Gilbertson, R.L. 1994. Occurrence of the Eastern Mediterranean strain of tomato yellow leaf curl geminivirus in the Dominican Republic. *Phytopathology* 84: 1072. [ Abstract]
882. Nakhla, M.K., Maxwell, D.P., Martinez, R.T., Carvalho, M.G. and Gilbertson, R.L. 1994. Widespread occurrence of the Eastern Mediterranean strain of tomato yellow leaf curl geminivirus in tomatoes in the Dominican Republic. *Plant Dis.* 78(9): 926. [ Abstract]
883. Naranjo, S. and Hagler, J. 1996. Characterizing and estimating the effect of heteropteran predation. p. 132. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
884. Naranjo, S.E. 1999. Potential interactions between predators and parasitoids of sweetpotato whitefly. p. 78. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
885. Naranjo, S.E. 2000. Effect of an experimental insecticide (NI-25) on natural enemies of sweetpotato whitefly. p. 67. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
886. Naranjo, S.E. 2000. Intraguild predation on whitefly parasitoids. p. 93. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
887. Naranjo, S.E. and Chu, C.C. 1996. Effects of insecticidal management of *Bemisia* on populations of natural enemies. p. 133. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
888. Naranjo, S.E., Chu, C.C., Ellsworth, P.C., Henneberry, T.J., Nichols, R.L., Riley, D.G., Toscano, N.C. and Watson, T.F. 1996. Action thresholds for *Bemisia* in cotton: Results from a multi-site study. p. 9. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
889. Naranjo, S.E. and Ellsworth, P.C. 1999. Cohort-based life table studies of *Bemisia tabaci* in cotton. p. 16. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
890. Naranjo, S.E. and Ellsworth, P.C. 2000. Life table analysis of *Bemisia tabaci* in cotton. p. 22. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
891. Naranjo, S.E. and Ellsworth, P.C. 2000. Whitefly population dynamics: Why use life tables & what do they tell you? pp. 6-8. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
892. Naranjo, S.E., Ellsworth, P.C., Diehl, J., Dennehy, T. and Flint, H. 1994. Validation, refinement and implementation of sampling plans for *Bemisia tabaci* in cotton. *Phytoparasitica* 22(4): 317. [ Abstract]
893. Naranjo, S.E., Ellsworth, P.C. and Diehl, J.W. 1995. Validation and analysis of sampling plans for *Bemisia tabaci* in cotton. p. 12. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
894. Naranjo, S.E., Ellsworth, P.C. and Diehl, J.W. 1997. Partial life table studies of *Bemisia* in cotton fields subject to different management strategies. p. 71. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]

895. Naranjo, S.E., Ellsworth, P.C. and Diehl, J.W. 1998. Comparative life table studies of *Bemisia* under different management strategies in cotton. p. 17. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
896. Naranjo, S.E. and Flint, H.M. 1993. Sampling immature sweetpotato whiteflies in cotton. p. 15. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
897. Naranjo, S.E. and Flint, H.M. 1994. Development and evaluation of sampling plans for immature and adult sweetpotato whitefly in cotton. p. 20. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
898. Naranjo, S.E., Flint, H.M. and Henneberry, T.J. 1995. Integration of irrigation strategies and action thresholds for management of *Bemisia tabaci* in cotton. p. 150. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
899. Naranjo, S.E., Flint, H.M., Henneberry, T.J., Martin, J.M., Parks, N.J. and Jech, L.F. 1994. Comparison of direct and indirect sampling methods for adult sweetpotato whitefly in cotton. p. 21. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
900. Naranjo, S.E. and Hagler, J.R. 1997. Conservation of *Bemisia* natural enemies in relation to conventional and IGR-based management systems. p. 162. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
901. Naranjo, S.E. and Hagler, J.R. 1998. Conservation of *Bemisia* natural enemies in integrated whitefly management systems. p. 74. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
902. Naranjo, S.E., Hagler, J.R. and Ellsworth, P.C. 2000. Conservation of whitefly natural enemies in conventional and IGR-based management systems. p. 94. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
903. Naranjo, S.E. and Henneberry, T.J. 1996. Development of sampling methods for estimating cotton lint stickiness in the field. p. 10. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
904. Naranjo, S.E. and Henneberry, T.J. 1998. Spatial distribution of cotton lint stickiness and preliminary field sampling methods for estimating stickiness. p. 19. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
905. Naranjo, S.E. and Henneberry, T.J. 1999. Evaluation of a high-speed thermodetector for estimating cotton lint stickiness. p. 15. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
906. Naranjo, S.E., Henneberry, T.J. and Chu, C.C. 1997. Progress in development of field sampling methods for estimating cotton lint stickiness. p. 72. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
907. Naranjo, S.E. and Prabhaker, N. 2000. Toxicological studies of two insect growth regulators on the predator *Geocoris punctipes*. p. 66. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
908. Natarajan, K. 1988. Monitoring seasonal activity of cotton whitefly, *Bemisia tabaci*, using yellow sticky traps. p. 61. *In* National Workshop on Pest and Disease Surveillance for IPM, 16-17 September. Indian Council of Agricultural Research. [Abstract]
909. Natwick, E.T. 1993. [cotton, chemical control]. p. 70. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
910. Natwick, E.T. 1994. Control of silverleaf whitefly in fresh market tomatoes using various insecticides, oils, an insect growth regulator, and an insecticidal soap. p. 100. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
911. Natwick, E.T. 1994. Control of silverleaf whitefly on cantaloupe melon using various insecticides, spring, 1992. p. 95. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
912. Natwick, E.T. 1994. Cotton cultivar evaluation for susceptibility to colonization by the silverleaf whitefly. p. 164. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
913. Natwick, E.T. 1994. Silverleaf whitefly control in cotton using insecticides and an insect growth regulator. p. 101. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
914. Natwick, E.T. 1994. Silverleaf whitefly control in cotton using systemic insecticides. p. 98. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
915. Natwick, E.T. 1994. Silverleaf whitefly control in spring planted cantaloupe melons, Imperial County, California, 1993. p. 99. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
916. Natwick, E.T. 1994. Silverleaf whitefly control on broccoli using various insecticide, oil, soap and insect growth regulator treatments. p. 97. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
917. Natwick, E.T. 1994. Silverleaf whitefly control on melons using various insecticides, spring 1992. p. 96. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
918. Natwick, E.T. 1995. Cotton insecticides efficacy for control of silverleaf whitefly. p. 79. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
919. Natwick, E.T. 1995. Efficacy evaluations of insecticides for control of silverleaf whitefly on cantaloupe melon. p. 80. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
920. Natwick, E.T. 1995. IPM and whitefly control from the growers' perspective. p. 169. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]

921. Natwick, E.T. 1996. Control of silverleaf whitefly in cotton using insecticide rotations. p. 84. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
922. Natwick, E.T. 1996. Efficacy evaluation of insecticides for silverleaf whitefly control in cantaloupe. p. 86. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
923. Natwick, E.T. 1996. Efficacy of various insecticides against silverleaf whitefly in cotton. p. 87. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
924. Natwick, E.T. 1997. Effects of silverleaf whitefly feeding on sugar beet yield, percent sugar, and leaf petiole chlorosis. p. 127. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
925. Natwick, E.T. 1997. Silverleaf whitefly control in cantaloupe melons, 1996. p. 124. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
926. Natwick, E.T. 1997. Silverleaf whitefly control in tomato, 1996. p. 125. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
927. Natwick, E.T. 1998. Evaluation of insecticides for silverleaf whitefly control in tomato. p. 41. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
928. Natwick, E.T. 1998. Silverleaf whitefly control in fall planted cantaloupe melons, 1997. p. 42. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
929. Natwick, E.T. 1998. Silverleaf whitefly control in spring planted cantaloupe melons, 1997. p. 43. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
930. Natwick, E.T. 2000. Evaluation of insecticides for silverleaf whitefly control in spring planted cantaloupe melons, 1999. p. 68. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
931. Natwick, E.T., Chu, C.C., Cohen, A.C., Simmons, G. and Brushwood, D. 1997. Effects of cotton variety selection and insecticide treatments on silverleaf whitefly infestation levels. p. 185. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
932. Natwick, E.T., Chu, C.C., Constable, G. and Brushwood, D.E. 1998. Okra-leaf and normal leaf cotton resistance to silverleaf whitefly. p. 210. *In* U. Kechagia (Ed.), World Cotton Research Conference 2. Sept. 6-12, 1998. Athens Greece. [Abstract]
933. Natwick, E.T., Chu, C.C., Henneberry, T.J., Brushwood, D. and Constable, G. 1999. Silverleaf whitefly infestation levels on normal leaf and okra-leaf upland cotton cultivars. p. 106. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
934. Natwick, E.T., Chu, C.C., Henneberry, T.J., Brushwood, D. and Cook, C. 1998. Silverleaf whitefly infestation levels in relation to cotton variety. p. 94. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
935. Natwick, E.T., Chu, C.C., Henneberry, T.J., Cook, C., Gilbertson, R.L. and Brushwood, D. 1998. Silverleaf whitefly infestation and cotton leaf crumple virus symptoms in relation to cotton genotype. p. 95. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
936. Natwick, E.T., Cook, C.G., Gilbertson, R.L. and Seo, Y.S. 1999. Cotton leaf crumple geminivirus disease resistance in upland cotton. p. 105. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
937. Natwick, E.T., Cook, C.G., Gilbertson, R.L., Seo, Y.S. and Turini, T. 2000. Resistance to cotton leaf crumple geminivirus disease in upland cotton. p. 118. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
938. Natwick, E.T. and Engle, C.E. 1997. Evaluation of imidacloprid for silverleaf whitefly control in tomato using various formulations and application methods. p. 123. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
939. Natwick, E.T., Henneberry, T.J. and Brushwood, D. 1998. Silverleaf whitefly control in cotton, 1996. p. 44. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
940. Natwick, E.T., Henneberry, T.J. and Brushwood, D. 1999. Silverleaf whitefly control in cotton, 1997. p. 55. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
941. Natwick, E.T., Henneberry, T.J. and Brushwood, D. 2000. Evaluation of insecticides for silverleaf whitefly control in cotton, 1999. p. 69. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
942. Natwick, E.T. and Leigh, T.F. 1993. [cotton, okra-leaf, defoliation]. p. 124. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
943. Natwick, E.T. and Mayberry, K.S. 1994. Evaluation of floating rowcover materials to exclude silverleaf whitefly from iceberg head lettuce. p. 102. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
944. Natwick, E.T. and Mayberry, K.S. 1994. Evaluation of insecticides for control of silverleaf whitefly on iceberg head lettuce in Southern California. p. 103. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
945. Natwick, E.T. and Mayberry, K.S. 1994. Evaluation of repellents of silverleaf whitefly on iceberg lettuce. p. 166. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]

946. Natwick, E.T. and Mayberry, K.S. 1995. Evaluation of insecticides for control of silverleaf whitefly on iceberg lettuce. p. 81. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
947. Natwick, E.T. and Mayberry, K.S. 1995. Evaluation of insecticides for control of silverleaf whitefly on transplanted cauliflower. p. 82. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
948. Natwick, E.T. and Mayberry, K.S. 1996. Comparison of various formulations of *Beauveria bassiana* with Capture 2EC tank mix with Orthene 75S for control of silverleaf whitefly in cauliflower. p. 83. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
949. Natwick, E.T. and Mayberry, K.S. 1996. Effects of imidacloprid as transplant drench and soil treatments on colonization of silverleaf whitefly, phytotoxicity, plant growth, and marketability of cauliflower. p. 85. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
950. Natwick, E.T. and Mayberry, K.S. 1996. Insecticide efficacy against silverleaf whitefly in broccoli. p. 88. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
951. Natwick, E.T. and Mayberry, K.S. 1996. Insecticide efficacy against silverleaf whitefly in cauliflower. p. 89. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
952. Natwick, E.T. and Mayberry, K.S. 1996. Insecticide efficacy against silverleaf whitefly in tomato. p. 90. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
953. Natwick, E.T. and Mayberry, K.S. 1999. Evaluation of selected insecticides for silverleaf whitefly control in cabbage. p. 53. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
954. Natwick, E.T. and Mayberry, K.S. 1999. Evaluation of selected insecticides for silverleaf whitefly control in iceberg lettuce. p. 56. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
955. Natwick, E.T. and Mayberry, K.S. 1999. Silverleaf whitefly control in spring planted cantaloupe melons, 1998. p. 54. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
956. Natwick, E.T. and Mayberry, K.S. 2000. Efficacy of selected insecticides for silverleaf whitefly control in iceberg lettuce, 1999. p. 70. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
957. Natwick, E.T., Mayberry, K.S. and Laemmlen, F.F. 1994. Protection from phytophagous insects in cantaloupe melons using rowcovers, reflective mulches and reflective netting. p. 165. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
958. Natwick, E.T. and Robinson, F. 1993. Alfalfa cultivar susceptibility to sweetpotato whitefly, strain -B. p. 125. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
959. Natwick, E.T., Robinson, F. and Bell, C. 1993. [alfalfa, irrigation]. p. 126. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
960. Natwick, E.T., Summers, C.G., Chu, C.C., Godfrey, L.D., Mayberry, K.S., Bell, C.E. and Henneberry, T.J. 2000. Hosts of silverleaf whitefly in Imperial and southern San Joaquin Valley, California. p. 23. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
961. Natwick, E.T., Toscano, N.C. and Yates, L. 1994. Correlations of adult whitefly sampling techniques in cotton to whole-plant samples. *Phytoparasitica* 22(4): 319-320. [Abstract]
962. Natwick, E.T., Walker, G., Johnson, D., Chu, C.C., Henneberry, T.J., Brushwood, D. and Constable, G. 2000. Normal leaf and okra-leaf upland cotton cultivars susceptibility to infestation by silverleaf whitefly. p. 117. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
963. Nava-Camberos, U. and Cano Rios, P. 1996. Biología y dinámica poblacional de la mosquita blanca de la hoja plateada *Bemisia argentifolii*, en algodón y hortalizas. [Biology and population dynamics of *Bemisia argentifolii* in cotton and vegetables]. p. 20. *In* Proceedings 4th Scientific and Technological Meeting of Forestry, Agriculture and Husbandry, Saltillo, Coahuila, Mexico, 23-25 Sep 1996. INIFAP. [Spanish, Abstract]
964. Neal, J.W., Severson, R.F., Stephenson, M.G. and Sisson, V.A. 1993. [Nicotiana]. p. 70. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
965. Neal, J.W., Jr. 1996. Phenotype plasticity in whiteflies: an interpretation. p. 39. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
966. Neal, J.W., Jr. 1997. Why polyphenism is important in polyphagous whiteflies, why whiteflies are white, and the relationship of these behaviors by the silverleaf and greenhouse whiteflies to cultivar selection in plant resistance. p. 96. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
967. Neal, J.W., Jr. and Buta, G. 1994. Nicotine does not enhance activity of sucrose esters from *Nicotiana glauca* against whitefly nymphs. p. 104. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
968. Neal, J.W., Jr. and Oliver, J.E. 1993. [broccoli, pheromone]. p. 37. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
969. Nelson, D.R. and Buckner, J.S. 1994. A survey of some species of whitefly occurring in the U.S.A. *Phytoparasitica* 22(4): 310. [Abstract]
970. Nelson, D.R., Buckner, J.S., Freeman, T.P., Gerling, D., Guershon, M. and Walker, G.P. 1998. Surface lipid composition of *Aleyrodes singularis* and comparisons with other whitefly species. p. 18. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
971. Nelson, D.R., Freeman, T.P. and Buckner, J.S. 2000. Characterization of the external waxes and wax producing glands of three species of whiteflies. p. 24. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]

972. Nelson, D.R., Freeman, T.P., Buckner, J.S., Hoelmer, K., Hagler, J. and Jackson, C.G. 1999. Formation of external waxy particles by adult *Bemisia argentifolii* and *Semidalis flinti*. p. 17. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
973. Nelson, M.R. and Orum, T.V. 1993. [geographic information systems, GIS], p. 137. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
974. Nelson, M.R., Orum, T.V., Nadeem, A., Felix, R. and Trinidad, R. 1994. Geostatistical analysis of the regional distribution of viruses transmitted by the sweet potato whitefly. p. 180. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
975. Nguyen, R. 1996. Importation and field release of parasites against silverleaf whitefly, *Bemisia argentifolii* (Bellows and Perring) in Florida from 1990-1995. p. 134. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
976. Nguyen R. and Bennett, F.D. 1994. Importation, release and field recovery of parasites of *Bemisia tabaci* in Florida (1990-1993). p. 144. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
977. Nguyen, R. and Bennett, F.D. 1995. Introduction and establishment of *Eretmocerus* sp. (HK) (Hymenoptera: Aphelinidae), a parasite of *Bemisia argentifolii*, in Florida. p. 132. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
978. Nichols, R.L. 1996. Silverleaf whitefly drives development of sustainable resistance management in Arizona cotton. p. 91. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
979. Nichols, R.L. 1998. Deployment of insecticidal modes of action for resistance management. p. 114. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
980. Nichols, R.L., Bentley, W.J., Chu, C.C., Ellsworth, P.C., Goodell, P.B., Henneberry, T.J., Naranjo, S.E., Riley, D.G., Toscano, N.C. and Watson, T.F. 1996. Determining an action threshold to prevent *Bemisia* outbreaks in cotton. pp. 241-242. *In* D. Gerling and R.T. Mayer (Eds.), *Bemisia* : 1995 Taxonomy, Biology, Damage, Control and Management. Intercept Ltd., Andover, Hants, UK. [Abstract]
981. Nichols, R.L., Chu, C.C., Ellsworth, P.C., Henneberry, T.J., Naranjo, S.E., Riley, D.G., Toscano, N.C. and Watson, T.F. 1994. Determining an action threshold to prevent whitefly outbreaks. *Phytoparasitica* 22(4): 349. [ Abstract]
982. Nicholson, W.F., Senn, R. and Fluckiger, C.R. 1994. Pymetrozine - a novel compound for control of whiteflies. *Phytoparasitica* 22(4): 358. [ Abstract]
983. Nordlund, D.A. and Legaspi, J.C. 1994. Whitefly predators and their possible use in biological control. *Phytoparasitica* 22(4): 333. [ Abstract]
984. Norman, J.W., Jr. 1996. Silverleaf host plant resistance in the U. S.: a status report. p. 165. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
985. Norman, J.W., Jr., Sparks, A.N., Jr. and Riley, D.G. 1995. An integrated approach to sweetpotato whitefly management in the Lower Rio Grande Valley of Texas. p. 170. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
986. O'Doherty, F. 1995. Report of a geminivirus disease complex associated with B-biotype of sweetpotato whitefly in Belize, Central America. *Plant Dis.* 79(12): 1250. [ Abstract]
987. Odom, P., Lublinkhof, J. and Strachan, F. 1995. Control of whitefly with combinations of Ovasyn + Phaser. p. 84. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
988. Oetting, R.D. 1994. *Bemisia* damage expression in commercial greenhouse production. *Phytoparasitica* 22(4): 321-322. [ Abstract]
989. Oliveira, M.R.V. 1998. Impact of *Bemisia argentifolii* Bellows & Perring in Brazil. p. 20. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
990. Oliveira, M.R.V. and Navia, D. 1999. Prospection and evaluation of natural enemies of *Bemisia tabaci* (Biotype B) [Brazil]. p. 79. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
991. Ongprasert, P. 1995. Studies on the efficiency of some insecticides in controlling of tobacco whitefly (*Bemisia tabaci* Gennadius). pp. 47-48. *In* Mae Jo Tobacco Experiment Station, Chiang Mai, Thailand, Annual Report 1993. [Abstract]
992. Onillon, J.C., Braham, M. and Kirk, A. 1996. Preliminary results on the efficacy of *Eretmocerus mundus* (Hymenopt.: Aphelinidae), in the biological control of *Bemisia tabaci* (Homopt.: Aleyrodidae). p. 135. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
993. Onillon, J.C. and Maignet, P. 1995. Preliminary results on the efficacy of *Encarsia pergandiella* (Hymenoptera: Aphelinidae), in the biological control of *Bemisia tabaci* (Homoptera: Aleyrodidae). p. 133. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
994. Or, R. and Gerling, D. 1985. The green lacewing, *Chrysoperla carnea*, as a predator of *Bemisia tabaci*. *Phytoparasitica* 13: 75. [ Abstract]
995. Ordonez-Matzer, L.F. and Yoshii, K. 1979. Evaluacion de perdidas en rendimiento de frijol debidas al mosaico dorado bajo condiciones de campo. [Evaluacion of yield losses in bean due to golden mosaic virus under field conditions]. *Fitopatol. Colombiana* 8(1): 31. [ Spanish, Abstract]
996. Oren, H. and Gerling, D. 1983. Fluctuations in populations of *Trialeurodes vaporariorum* and *Bemisia tabaci* inhabiting the same *Lantana camara* plants. *Phytoparasitica* 11: 64-65. [ Abstract]
997. Orozco-Santos, M., Perez-Zamora, O. and Lopez-Arriaga, O. 1995. Effect of floating rowcover and transparent mulch on insect populations, virus disease, and yield of muskmelon. *Hortscience* 30(4): 828. [ Abstract]
998. Orzolek, M.D., Thomas, C., Berghage, R.D. and Heller, P.R. 1995. Development of a greenhouse IPM program in Pennsylvania. *Hortscience* 30(4): 847. [ Abstract]
999. Osborne, L.S., Jimenez, D.R., Yokomi, R.K. and Shapiro, J.P. 1993. Comparison of B-biotype induced squash silverleaf (SSL) and plant growth regulator(PGR)-induced leaf silencing. p. 38. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]

1000. Osborne, L.S., Storey, G.K., McCoy, C.W. and Walter, J.F. 1990. Potential for controlling the sweetpotato whitefly, *Bemisia tabaci*, with the fungus *Paecilomyces fumosoroseus*. p. 386. *In Proceedings and Abstracts. Vth International Colloquium on Invertebrate Pathology and Microbial Control, Adelaide, Australia, August 20-24, 1990.* Dept. Entomol., Univ. Adelaide, Glen Osmond, Australia. [Abstract, Cock (1993)]
1001. Oster, N. and Gerling, D. 1994. Host killing and time allocation of the parasitoid *Encarsia transvena*. *Phytoparasitica* 22(4): 340. [Abstract]
1002. Ozgür, A.F. and Sekeroglu, E. 1984. Population developments of *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae) on various cotton varieties in Cukurova, Turkey. p. 568. *In XVII International Congress Entomology.* [Abstract]
1003. Pacheco-Covarrubias, J.J. 1995. Response of nival instars of *Bemisia* spp. to imidacloprid. p. 85. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1004. Pacheco-Covarrubias, J.J. 1997. Exponential growth phases of silverleaf whitefly in the Yaqui Valley, Mexico. p. 73. *In Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001).* U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
1005. Pacheco-Covarrubias, J.J. and Hernandez-Jasso, A. 1998. Respuesta del algodón a mosquita blanca de la hoja plateada bajo condiciones de no elección. [Cotton response to silverleaf whitefly under no choice conditions]. p. 139. *In 17th Phytogetic Congress. Proceedings. Scientific Notes, Acapulco, Mexico, 5-9 Oct 1998.* [Spanish, Abstract]
1006. Palaniswami, M.S. 1997. *Bemisia tabaci* Genn. and its natural enemies. p. 163. *In Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001).* U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
1007. Palaniswami, M.S. and Antony, B. 2000. Biology and bionomics of *Encarsia transvena* (Timberlake) on *Bemisia tabaci* Genn. in cassava. p. 95. *In Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan.* U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
1008. Palaniswami, M.S., Antony, B. and Vijayan, L. 1999. Survey and identification of *Bemisia tabaci* Genn. and its natural enemies in India. p. 80. *In Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
1009. Palaniswami, M.S., Antony, B. and Vijayan, L. 2000. Biology, ecology and morphometrics of *Bemisia tabaci* Genn. on cassava, sweet potato, brinjal, cotton and tobacco. p. 25. *In Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan.* U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
1010. Palaniswami, M.S., Vijayan, L. and Antony, B. 1999. Biology of coccinellid predators of *Bemisia tabaci* Genn. p. 81. *In Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
1011. Palumbo, J., Yee, W. and Toscano, N. 1995. Development of seasonal action thresholds for chemical control of silverleaf whitefly on alfalfa. p. 15. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1012. Palumbo, J.C. 1993. [cantaloupe]. p. 16. *In Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1013. Palumbo, J.C. 1993. [chemical control, cantaloupes, cauliflower]. p. 71. *In Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1014. Palumbo, J.C. 1993. [sampling, cantaloupe]. p. 72. *In Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1015. Palumbo, J.C. 1994. Evaluation of Admire for control of sweetpotato whitefly in commercial head lettuce in Arizona. p. 105. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
1016. Palumbo, J.C. 1994. Insecticidal control of sweetpotato whitefly on spring melons. p. 106. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
1017. Palumbo, J.C. 1995. Yield and quality response in iceberg lettuce to whiteflies and imidacloprid. p. 87. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1018. Palumbo, J.C. 1996. Imidacloprid field performance on commercial lettuce in Arizona. p. 92. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1019. Palumbo, J.C., Ellsworth, P.C., Dennehy, T.J. and Umeda, K. 1999. A grower initiated model for sustaining chemical efficacy across commodities. pp. 3-4. *In Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
1020. Palumbo, J.C. and Kerns, D.L. 1996. Imidacloprid formulation and soil placement: effects on colonization by sweetpotato whitefly on head size and incidence of chlorosis in lettuce. p. 93. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1021. Palumbo, J.C. and Riley, D.G. 1995. Interaction of silverleaf whitefly with melon crop quality. p. 13. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1022. Palumbo, J.C. and Sanchez, C.A. 1995. Imidacloprid does not enhance growth and yield of cantaloupe in the absence of whitefly. p. 86. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1023. Palumbo, J.C., Tonhasca, A. and Byrne, D. 1994. Population dynamics of *Bemisia tabaci* in cantaloupe. p. 23. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
1024. Palumbo, J.C., Tonhasca, A. and Byrne, D. 1994. Sampling sweetpotato whitefly populations in cantaloupes. p. 22. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
1025. Palumbo, J.C., Tonhasca, A. and Byrne, D.N. 1996. Comparison of relative sampling methods for estimating adult sweetpotato whitefly abundance on cantaloupes. p. 11. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]

1026. Palumbo, J.C. and Toscano, N.C. 1995. Impact of silverleaf whitefly populations on yield and quality of alfalfa. p. 14. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1027. Parrella, M.P., Heinz, K.M., Nelson, J.M., Brazzle, J. and Pickett, C. 1993. [*Delphastus*, cotton, tomato, poinsettia, *Encarsia*]. p. 107. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1028. Partida, L., Inzunza, J.F., Yanez, M.G. and Acosta, B. 1996. Evaluacion de la preferencia de la mosquita blanca de la hoja plateada *Bemisia argentifolii* Bellows y Perring. Por variedades de brocoli en el Valle de Mexicali, Baja California. [Evaluation of the preference to silverleaf whitefly (*Bemisia argentifolii*) to Brassica oleracea in Mexicali Valley]. p. 60. *In* Proceedings 16th Plant Breeding Congress, 6-11 Oct. 1996, Texcoco, Mexico State. Sociedad Mexicana De Fitogenetica. [Spanish, Abstract]
1029. Pasian, C.C., Struve, D.K. and Lindquist, R. 1996. Painted containers: A new method for imidacloprid application. *Hortscience* 31(4): 655-656. [Abstract]
1030. Pavis, C. and Boissot, N. 1999. Population dynamics of *Bemisia tabaci* B biotype, on continuous pumpkin culture in Guadeloupe (French West Indies). p. 18. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
1031. Percy, R.G., Ellsworth, P. and Moser, H.S. 1996. Field preference of silverleaf whitefly among pima cotton genotypes. p. 166. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1032. Percy, R.G., Ellsworth, P.C. and Moser, H.S. 1997. Silverleaf whitefly resistance screening in Pima cotton genotypes. p. 186. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
1033. Peregrine, D.J. 1983. Developments in the use of amitraz for control of cotton pests. p. 940. *In* 10th International Congress of Plant Protection. Proceedings of a Conference Held at Brighton, England, 20-25 November, 1983. Plant Protection for Human Welfare. British Crop Protection Council, Croydon, UK. [Abstract, Cock (1986)]
1034. Perring, T.M. 1993. [taxonomy, crossing, enzymes, esterase, polymerase chain reaction]. p. 39. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1035. Perring, T.M. 1994. Biological characteristics of *Bemisia tabaci* and closely related species. *Phytoparasitica* 22(4): 309. [Abstract]
1036. Perring, T.M. and Bellows, T.S. 1993. [cantaloupe, cotton, alfalfa, weeds, parasitism]. p. 17. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1037. Perring, T.M., Brown, J., Cooper, A.D., Bedford, I. and Markham, P. 1999. Genetic analysis of *Bemisia* (Homoptera: Aleyrodidae) populations by isoelectric focusing electrophoresis. p. 19. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
1038. Perring, T.M. and Farrar, C.A. 1995. Competitive displacement of sweetpotato whitefly by silverleaf whitefly. p. 44. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1039. Perring, T.M. and Farrar, C.A. 1995. Mating behavior of *Bemisia* and its relationship to competitive advantage. p. 45. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1040. Perring, T.M. and Farrar, C.A. 1998. Sampling protocol for silverleaf whitefly on tomato. p. 21. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
1041. Perring, T.M., Farrar, C.A. and Cooper, A.D. 1994. Mating behavior and competitive displacement in whiteflies. p. 25. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
1042. Perring, T.M., Farrar, C.A. and Vasquez, V. 1994. Silverleaf whitefly on crops and weeds in the Imperial Valley. p. 24. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
1043. Perring, T.M. and Gruenhagen, N.M. 1997. Velvetleaf: a possible candidate for enemy free space in desert agricultural systems. p. 74. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
1044. Perring, T.M., Mayberry, K.S. and Natwick, E.T. 1995. Silverleaf whitefly management in cauliflower using a trap crop. p. 151. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1045. Perry, A.S. 1985. The relative susceptibility to several insecticides of adult whiteflies (*Bemisia tabaci*) from various cotton-growing areas in Israel. *Phytoparasitica* 13: 77-78. [Abstract Cock (1993)]
1046. Pesic-Van Esbroeck, Z., Harris, K.F. and Duffus, J.E. 1995. Immunocytochemical localization of squash leaf curl virus (SLCV) in squash and the sweet potato whitefly. *Phytopathology* 85(10): 1180. [Abstract]
1047. Pesic-Van Esbroeck, Z., Harris, K.F. and Duffus, J.E. 1995. Morphology of the whitefly feeding apparatus relative to noncirculative plant virus transmission. *Phytopathology* 85(10): 1180. [Abstract]
1048. Pesic-Van Esbroeck, Z., Harris, K.F. and Duffus, J.E. 1996. Sweet potato whitefly-squash leaf curl virus immunocytochemistry. p. 40. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1049. Pesic-Van Esbroeck, Z., Harris, K.F. and Duffus, J.E. 1997. Fate of squash leaf curl virus in squash. p. 97. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
1050. Peterkin, D.D. and Hall, R.A. 1994. Effect of accelerated spore germination on virulence of *Paecilomyces fumosoroseus* against the whitefly, *Bemisia tabaci*. *Phytoparasitica* 22(4): 344-345. [Abstract]
1051. Peterlin, O. and Helman, S. 1994. Some aspects of the population dynamics of *Bemisia tabaci* as a cotton pest in Santiago del Estero, NW. Argentina. *Phytoparasitica* 22(4): 318. [Abstract]
1052. Peterschmitt, M., Granier, M., Mekdoud, R., Dalmon, A., Gambin, O., Vayssières, J.F. and Reynaud, B. 1999. First report of tomato yellow leaf curl virus in Reunion Island [Indian Ocean]. *Plant Dis.* 83(3): 303. [Abstract]
1053. Petersen, M.A., Wisler, G.C., Purcifull, D.E. and Duffus, J.E. 1996. Cytopathology of infections caused by the whitefly-transmitted lettuce chlorosis closterovirus. *Phytopathology* 86(11 SUPPL): S71. [Abstract]
1054. Petro, L.O. and Redak, R.A. 2000. Host plant preference and performance of *Bemisia argentifolii* (Homoptera: Aleyrodidae) on poinsettia (*Euphorbia pulcherrima*) in relation to cultivar. p. 119. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]



1055. Pickett, C.H. and Abel, B. 2000. Tracking the impact of released parasites using sentinel plants. p. 98. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
1056. Pickett, C.H., Abel, W.L., Riccomini, C., Simmons, G. and Goolsby, J.A. 1998. Recovery and releases of parasites for biological control of *Bemisia* sp. in the San Joaquin Valley, California. p. 75. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
1057. Pickett, C.H., Goolsby, J.A. and Abel, W.L. 1996. Release of aphelinids for control of silverleaf whitefly in San Joaquin Valley, California. p. 137. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1058. Pickett, C.H., Goolsby, J.A., Abel, W.L. and Boyd, G. 1997. Colonization of parasites for biological control of silverleaf whitefly in the San Joaquin Valley, California. p. 164. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
1059. Pickett, C.H. and Hoelmer, K.A. 1996. Low temperature oviposition rate of aphelinids: a technique for determining winter tolerance of exotic parasites for release in Imperial Valley [CA]. p. 136. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1060. Pickett, C.H., Hoelmer, K.A. and Roltsch, W.J. 1997. Systematics of *Delphastus* and biological control of whiteflies. p. 165. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
1061. Pickett, C.H., Lozano, E. and Overholt, D. 2000. Fall releases of parasites into citrus. p. 97. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
1062. Pickett, C.H. and Overholt, D. 2000. Survivorship of silverleaf whitefly overwintering in citrus in the San Joaquin Valley. p. 26. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
1063. Pickett, C.H., Simmons, G.S. and Goolsby, J.A. 1999. Augmentative biological control using transplants. p. 82. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
1064. Pickett, C.H., Simmons, G.S., Goolsby, J.A. and Overholt, D. 1999. Fall releases of parasites into citrus. p. 83. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
1065. Pickett, C.H., Simmons, G.S., Lozano, E. and Goolsby, J.A. 2000. Augmentative biological control using transplants. p. 96. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
1066. Pittarelli, G.W., Buta, J.G., Chortyk, O.T. and Wilson, S. 1996. Increased production of biopesticides from interspecific hybridization of *Nicotiana* species. p. 139. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1067. Pittarelli, G.W., Buta, J.G., Chortyk, O.T., Wilson, S. and Hossain, A. 1996. The use of chromosome amplification to increase the quantity of biopesticide production from *Nicotiana* species. p. 140. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1068. Pittarelli, G.W., Buta, J.G., Severson, R.F. and Nottingham, S.F. 1996. A novel and rapid bioassay to screen *Nicotiana* species for activity against the adult silverleaf whitefly *Bemisia argentifolii* N. p. 139. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1069. Polston, J.E. 1996. Tomato geminiviruses in the Caribbean: their identification and management. *Phytopathology* 86(11 SUPPL.): S117. [Abstract]
1070. Polston, J.E. 1999. The appearance of tomato yellow leaf curl virus (Geminiviridae, *Begomovirus*) in Florida. pp. 1-2. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
1071. Polston, J.E., Bois, D., Ano, G., Poliakoff, F. and Urbino, C. 1998. Occurrence of a strain of potato yellow mosaic geminivirus infecting tomato in the eastern Caribbean. *Plant Dis.* 82(1): 126. [Abstract]
1072. Polston, J.E., Bois, D., Carmona-Serra, A. and Concepcion, S. 1994. First report of tomato yellow leaf curl-like geminivirus in the Western Hemisphere. *Plant Dis.* 78: 831. [Abstract]
1073. Polston, J.E., Bois, D., Keinath, A.P. and Chellemi, D.O. 1995. Occurrence of tomato mottle geminivirus in South Carolina, Tennessee, and Virginia. *Plant Dis.* 79(5): 539. [Abstract]
1074. Polston, J.E., Hiebert, E., McGovern, R.J., Schuster, D.J. and Scott, J.W. 1993. [tomato, virus]. p. 40. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1075. Poprawski, T. and Ciomperlik, M.A. 1999. Impact of *Beauveria bassiana* on natural populations of sweetpotato whitefly predators. p. 84. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
1076. Poprawski, T.J., Elzen, G.W., Uribe, N.A.S. and Veland, C.J. 1999. *In vitro* and *in vivo* compatibility of selected fungicides with fungal pathogens of *Bemisia* whiteflies. p. 59. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
1077. Poprawski, T.J., Gracia, C.G. and Veland, C.J. 1999. Field evaluation of Mycotrol R (conidia of *Beauveria bassiana* strain GHA), blastospores of *Paecilomyces fumosoroseus* strain 612, the particle film M97-009, and one sugar ester for control of whiteflies on melons. p. 58. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
1078. Poprawski, T.J., Gracia, C.G., Veland, C.J. and De La Garza, F. 1999. Field evaluation of the particle films M-96-018 and M-97-009 for whitefly control on bell peppers and collards. p. 57. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
1079. Poprawski, T.J. and Jones, W.A. 1999. Effects of *Beauveria bassiana* and *Paecilomyces fumosoroseus* on two lines of *Bemisia* whiteflies both reared on two different host plants. p. 60. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]

1080. Poprawski, T.J., Legaspi, J.C. and Parker, P.E. 1998. Effects of entomopathogenic fungi on *Serangium parcesetosum* (Coleoptera: Coccinellidae), an important predator of whiteflies. p. 76. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
1081. Powell, C.A. and Stoffella, P.J. 1997. Control of tomato irregular ripening with imidacloprid. *Phytopathology* 87(6 SUPPL.): S79. [Abstract]
1082. Prabhaker, N. 2000. Factors influencing insecticide resistance in whiteflies: Past, present, and future. *Am. Chem. Soc.* 219(1-2): Agro 29. [Abstract]
1083. Prabhaker, N. and Toscano, N. 1993. [insecticide resistance]. p. 73. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1084. Prabhaker, N., Toscano, N., Castle, S. and Henneberry, T. 1996. Effect of synergists on pyrethroid and organophosphate resistance in *Bemisia tabaci*. p. 94. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1085. Prabhaker, N., Toscano, N., Castle, S. and Henneberry, T. 1996. Evaluation of insecticide rotations and mixtures as a resistance management strategy for whiteflies. p. 95. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1086. Prabhaker, N., Toscano, N., Henneberry, T.J., Lawson, D.S. and Jones, K. 1998. Evaluation of biological activities of CGA-293343 and CGA-215944 (Fulfill) against silverleaf whiteflies on cotton. p. 45. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
1087. Prabhaker, N., Toscano, N.C., Castle, S. and Henneberry, T. 1995. Hydroponic bioassay technique to monitor responses of whiteflies to imidacloprid. p. 89. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1088. Prabhaker, N., Toscano, N.C., Castle, S. and Henneberry, T. 1995. Insecticide rotations as a resistance management strategy for whiteflies. p. 88. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1089. Prabhaker, N., Toscano, N.C., Castle, S.J. and Henneberry, T.J. 1997. Stability of imidacloprid resistance by various preventative strategies in silverleaf whiteflies (Homoptera: Aleyrodidae). p. 128. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
1090. Prabhaker, N., Toscano, N.C. and Henneberry, T.J. 1999. Effect of neem, urea and amitraz on oviposition and immature development of *Bemisia argentifolii* (Homoptera: Aleyrodidae). p. 61. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
1091. Prabhaker, N., Toscano, N.C. and Henneberry, T.J. 1999. Integrating various neonicotinoid insecticides into chemical control practices for sustainable whitefly management. p. 89. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce (Almeria), Spain. [Abstract]
1092. Prabhaker, N., Toscano, N.C. and Henneberry, T.J. 2000. Incorporating various neonicotinoids into chemical control practices for whitefly management. p. 71. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
1093. Prabhaker, N., Toscano, N.C., Henneberry, T.J. and Castle, S. 1994. Monitoring and management of insecticide resistance in the sweetpotato whitefly, *Bemisia tabaci*. p. 107. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
1094. Puri, S.N., Ansingkar, A.S., Ajankar, V.N., Lavekar, R.C., Butler, G.D., Jr. and Henneberry, T.J. 1993. [cotton, leaf morphology]. p. 127. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1095. Puri, S.N., Bhosle, B.B., Fartade, M.K., Kolhal, R.N., Butler, G.D., Jr. and Henneberry, T.J. 1994. Wild brinjal (*Solanum khasianum*) as a potential trap crop for the integrated management of *Bemisia tabaci* in cotton. *Phytoparasitica* 22(4): 358-359. [Abstract]
1096. Puri, S.N., Bhosle, B.B., Ilyas, M.D., Butler, G.D., Jr. and Henneberry, T.J. 1993. [control, soaps, oil, neem, hand spray]. p. 74. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1097. Puthoff, D., Perring, T. and Walling, L. 1998. The tomato defense response to silverleaf whitefly feeding. p. 96. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
1098. Puthoff, D., Walling, L. and Perring, T.M. 1997. The tomato defense response to silverleaf whitefly feeding. p. 98. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
1099. Puthoff, D.P., Perring, T.M. and Walling, L.L. 1998. The size of the silverleaf whitefly (*Bemisia argentifolii* Bellows & Perring) haploid genome. p. 97. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
1100. Puthoff, D.P., Perring, T.M. and Walling, L.L. 1999. Plant-insect interactions: the defense reaction of tomato to whitefly feeding. p. 107. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
1101. Raccach, B. 1994. The impact of international cooperation on the control of whiteflies and aphids. *Phytoparasitica* 22(4): 332. [Abstract]
1102. Rahman, K.A. 1940. Some more hosts of the cotton whitefly. *Indian J. Entomol.* 2: 243. [Abstract, Cock (1986)]
1103. Ramappa, H.K., Muniyappa, V. and Nateshan, H.M. 1994. Integrated management of whitefly-transmitted tomato leaf curl geminivirus disease in India. *Phytoparasitica* 22(4): 356-357. [Abstract]
1104. Ramos, P.L., Guerra, O., Dorestes, V., Ramirez, N., Rivera-Bustamante, R. and Oramas, R. 1996. Detection of TYLCV in Cuba. *Plant Dis.* 80(10): 1208. [Abstract]
1105. Rapid, P. and Gerling, D. 1989. Host habitat finding by *Encarsia deserti*, a parasitoid of *Bemisia tabaci*. *Phytoparasitica* 17(3): 234. [Abstract]
1106. Rathi, Y.P.S. 2000. Epidemiology of mungbean yellow mosaic virus, a yellow plague of kharif pulses. p. 43. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]

- 1107.Rathman, R.J. and Byrne, D.N. 1994. Field examination of migration by *Bemisia tabaci*. p. 26. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
- 1108.Riberiro, S.G., De Avila, A.C., Bezerra, I.C., Fernandes, J.J., Faria, J.C., Lima, M.F., Gilbertson, R.L., Maciel-Zambolim, E. and Zerbini, F.M. 1998. Widespread occurrence of tomato geminiviruses in Brazil, associated with the new biotype of the whitefly vector. *Plant Dis.* 82(7): 830. [ Abstract]
- 1109.Rice, R.P., Jr. and Crane, M. 2000. Susceptibility of poinsettia cultivars to whiteflies. *Hortscience* 35(3): 417. [ Abstract]
- 1110.Riemann, J.G. 1994. Ultrastructural on the nymphal cuticle and the egg membranes of *Bemisia tabaci* and *Trialeurodes vaporariorum*. p. 57. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
- 1111.Riley, D. 1998. Effect of the glabrous leaf trait of whiteflies and melon yield. p. 98. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
- 1112.Riley, D.G. 1993. [cantaloupe, cabbage, tomato, chemical control]. p. 75. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
- 1113.Riley, D.G. 1993. [cantaloupe, crop damage]. p. 138. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
- 1114.Riley, D.G. 1993. [cucurbit, melons]. p. 128. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
- 1115.Riley, D.G. 1993. [sampling]. p. 18. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
- 1116.Riley, D.G. 1994. Insecticide control and resistance management in whitefly populations. p. 108. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
- 1117.Riley, D.G. 1994. Susceptibility of melon cultivars to whiteflies. p. 167. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
- 1118.Riley, D.G. 1994. Thresholds of whiteflies in melons. pp. 27, 145. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
- 1119.Riley, D.G. 1995. Evaluation of sweetpotato whitefly on selected melon cultivars. p. 152. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
- 1120.Riley, D.G. 1995. Imidacloprid applied to cantaloupe. p. 90. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
- 1121.Riley, D.G. 1997. Melon cultivar response to whitefly attack. p. 187. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
- 1122.Riley, D.G. and Allen, J.C. 1994. Population dynamics of *Bemisia tabaci* (and *B. argentifolii*) in agricultural systems. *Phytoparasitica* 22(4): 315. [ Abstract]
- 1123.Riley, D.G. and Palumbo, J.C. 1995. Action thresholds for silverleaf whitefly in cantaloupe. p. 16. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
- 1124.Riley, D.G. and Tan, W. 1995. Bifenthrin resistance studies. p. 91. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
- 1125.Riley, D.G., Tan, W. and Wolfenbarger, D. 1996. [Bifenthrin resistance in the silverleaf whitefly]. p. 167. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
- 1126.Rimon, D. 1982. Chemical methods for the evaluation of stickiness in cotton fibers: *Bemisia tabaci* as a factor in contamination by sugars and in fiber stickiness. *Phytoparasitica* 10: 296-297. [ Abstract]
- 1127.Rimon, D. 1984. *Bemisia tabaci* as a factor in sugars' contamination and stickiness of cotton fibers in the 1983 season. *Phytoparasitica* 12: 139. [ Abstract]
- 1128.Rimon, D., Kaganovski, I. and Altachan, L. 1983. Chemical methods for the evaluation of stickiness in cotton fibers: *Bemisia tabaci* as a factor in contamination by sugars and in fiber stickiness (1982 season). *Phytoparasitica* 11: 66-67. [ Abstract]
- 1129.Ritter, D., Weddle, R.C. and Birdsall, S.L. 1997. Evaluation of CC whitefly trap for monitoring of adult whitefly population densities in an agricultural environment. p. 197. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
- 1130.Roberts, E.J.F., Buck, K.W. and Coutts, R.H.A. 1986. A new geminivirus infecting potatoes in Venezuela. *Plant Dis.* 70: 603. [ Abstract]
- 1131.Roltsch, W. and Pickett, C. 1995. Release and establishment of two new parasitoid species in Imperial Valley [CA]. p. 134. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
- 1132.Roltsch, W. and Pickett, C. 1995. Silverleaf whitefly natural enemy refuges in the Imperial Valley [CA]. p. 135. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
- 1133.Roltsch, W. and Pickett, C. 1996. Evaluation of refuges and new refuge plants for support of silverleaf whitefly natural enemies. p. 141. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
- 1134.Roltsch, W. and Pickett, C. 1998. Annual plants for natural enemy refuges in Imperial Valley, CA. p. 78. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
- 1135.Roltsch, W. and Pickett, C. 1998. Perennial plants for natural enemy refuges in Imperial Valley, CA. p. 79. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
- 1136.Roltsch, W. and Simmons, G. 1997. Release and establishment of exotic natural enemies in home gardens and agricultural field refuges in Imperial Valley, CA. p. 167. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]

1137. Roltsch, W., Simmons, G. and Hoelmer, K. 1998. Establishment of introduced *Encarsia* species in Imperial Valley, CA. p. 80. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
1138. Roltsch, W.J. 1999. Perennial plant refuges for silverleaf whitefly natural enemy conservation in Imperial Valley, CA. p. 85. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
1139. Roltsch, W.J. 2000. *Encarsia sophia* reproduction patterns. p. 100. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
1140. Roltsch, W.J., Andress, E.R., Hoelmer, K.A. and Simmons, G.S. 2000. Establishment of introduced parasitoids of the silverleaf whitefly in Imperial Valley, CA. p. 99. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
1141. Roltsch, W.J. and Brown, J.A. 1999. Nursery plots for establishing exotic parasitoids of the silverleaf whitefly in Imperial Valley, CA. p. 87. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
1142. Roltsch, W.J. and Goolsby, J.A. 1997. Field cage evaluations on non-indigenous parasitoids in desert crops. p. 166. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
1143. Roltsch, W.J. and Goolsby, J.A. 1998. Field cage evaluations of non-indigenous parasitoids in desert crops. p. 77. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
1144. Roltsch, W.J. and Pickett, C.H. 1994. Areawide establishment of *Delphastus pusillus*, a predator of the silverleaf whitefly, in the Imperial Valley. p. 147. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
1145. Roltsch, W.J. and Pickett, C.H. 1994. Silverleaf whitefly natural enemy refuges in Imperial County. p. 146. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
1146. Roltsch, W.J., Simmons, G.S. and Hoelmer, K.A. 1999. Establishment of introduced parasitoids of the silverleaf whitefly in Imperial Valley, CA. p. 86. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
1147. Romo-Ruiz, C. 1993. Evaluación semicomercial del control de mosquita blanca en frijol con endosulfan y karate, en la región de Ures, Son. [Semicomercial evaluation of *Bemisia tabaci* control on beans with endosulfan and karate. Ures region, Sonora State]. p. 81. *In* J.M. Ramirez-Diaz (Ed.), Research Advances. Spring-Summer 1988-1988. Centro De Investigaciones Agrícolas Del Noroeste, Oregon, Mexico. [Spanish, Abstract]
1148. Ronda, M., Adán, A., Cifuentes, D., Cenis, J.L. and Beitia, F. 1999. Laboratory evidence of interbreeding between biotypes of *Bemisia tabaci* (Homoptera, Aleyrodidae) present in Spain. p. 83. *In* VII International Plant Virus Epidemiology Symposium, Aguadulce (Almería), Spain. [Abstract]
1149. Rosado-Lugo, G., Trejo-Rivero, A. and Tun-Suarez, J.M. 1998. Métodos alternativos de control para la mosquita blanca *Bemisia tabaci* Gen. en tomate *Lycopersicon esculentum*. p. 187. *In* Proceedings 9th National Congress of Research and Agricultural Technological Development, Conkal, Yucatan, Mexico, 23-26 Nov 1998. Dir. Gral. Educación Tecnológica Agropecuaria. [Spanish, Abstract]
1150. Rose, M. and Zolnerowich, G. 1998. Systematics of *Eretmocerus*. p. 81. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
1151. Rose, M., Zolnerowich, G. and Hunter, M.S. 1994. Systematics of *Eretmocerus* (Hymenoptera: Aphelinidae), an important parasite of *Bemisia*. *Phytoparasitica* 22(4): 339. [Abstract]
1152. Rosell, R.C., Bedford, I.D., Markham, P.G., Frohlich, D.R. and Brown, J.K. 1994. Morphological variation in *Bemisia* populations. *Phytoparasitica* 22(4): 312. [Abstract]
1153. Rosell, R.C., Bedford, I.D., Markham, P.G., Frohlich, D.R. and Brown, J.K. 1995. Morphological variation in the fourth instar of whitefly populations in the genus *Bemisia*. p. 46. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1154. Rosell, R.C. and Brown, J.K. 1993. Geminivirus acquisition/transmission by the whitefly, *Bemisia tabaci*. p. 140. *In* Second International Symposium on Molecular Insect Science. Flagstaff, Arizona July 17-22, 1993. [Abstract]
1155. Rosell, R.C. and Brown, J.K. 1993. [morphology, virus, vectors]. p. 41. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1156. Rosell, R.C. and Brown, J.K. 1994. Geminivirus acquisition/transmission by *Bemisia tabaci*. p. 58. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
1157. Rosell, R.C. and Brown, J.K. 1994. Mechanisms of geminivirus acquisition and transmission by the whitefly, *Bemisia tabaci* Genn.). *Phytopathology* 84: 1131-1132. [Abstract]
1158. Rosell, R.C., Davidson, E.W., Jancovich, J.K., Brown, J.K. and Hendrix, D.L. 1997. Digestive tract size exclusion in nymphal and adult whiteflies. p. 100. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
1159. Rosell, R.C., Torres-Jerez, I. and Brown, J.K. 1996. Detection of geminivirus in whitefly extracts, saliva, and honeydew by polymerase chain reaction (PCR). p. 41. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1160. Rosell, R.C., Torres-Jerez, I. and Brown, J.K. 1996. Monitoring ingestion, excretion, and transmission of geminiviruses by whiteflies in a feeding chamber assay by polymerase chain reaction. *Phytopathology* 86(11 SUPPL.): S73. [Abstract]
1161. Rosell, R.C., Torres-Jerez, I. and Brown, J.K. 1997. Temporal monitoring of geminivirus DNA in whitefly hemolymph by polymerase chain reaction (PCR). *Phytopathology* 87(6 SUPPL.): S83. [Abstract]
1162. Rosell, R.C., Torres-Jerez, I. and Brown, J.K. 1997. Temporal monitoring of geminivirus in whitefly hemolymph using polymerase chain reaction (PCR). p. 99. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]

1163. Roye, M.E., McLaughlin, W.A., Nakhla, M.K. and Maxwell, D.P. 1999. Genetic diversity among geminiviruses infecting crops and weeds in Jamaica. p. 78. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce (Almeria), Spain. [Abstract]
1164. Rumei, X. 1994. Status and occurrence and distribution of the sweetpotato whitefly (*Bemisia tabaci*) in China. *Phytoparasitica* 22(4): 318. [Abstract]
1165. Russell, D.A., Kranthi, K.R., Jadhav, D., Regupathy, A. and Singh, J. 1999. Insecticide resistance in cotton pests in India. p. 11. *In* Proceedings ICAC-CCRI Regional Consultation, Insecticide Resistance Management in Cotton. Central Cotton Research Institute, Multan, Pakistan. [Abstract]
1166. Sachs, Y. 1993. *Bemisia tabaci* in vegetables - present status. *Phytoparasitica* 21(2): 169-170. [Abstract]
1167. Sachs, Y. 1994. Landmarks for the development of *Bemisia tabaci* infestations in Israel. *Phytoparasitica* 22(4): 347-348. [Abstract]
1168. Salas, J., Arnal, E., Mendoza, O. and Ramos, F. 1994. The sweetpotato whitefly, *Bemisia tabaci*, in Venezuela. *Phytoparasitica* 22(4): 355. [Abstract]
1169. Salas J. and Mendoza, O. 1994. Life cycle of the sweetpotato whitefly, *Bemisia tabaci*. *Phytoparasitica* 22(4): 313. [Abstract]
1170. Salvucci, M.E. and Crafts-Brandner, S.J. 2000. Effects of temperature and dietary sucrose concentration on respiration in the silverleaf whitefly, *Bemisia argentifolii*. p. 27. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
1171. Salvucci, M.E., Hendrix, D.L. and Whittard, C.R. 1996. Uptake and metabolism of plant proteins by the silverleaf whitefly. p. 42. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1172. Salvucci, M.E., Stecher, D.S. and Henneberry, T.J. 1999. Heat shock proteins and sorbitol accumulation as mechanisms for thermotolerance in *Bemisia argentifolii*. p. 20. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
1173. Salvucci, M.E., Wolfe, G. and Hendrix, D.L. 1997. Effect of sucrose concentration on carbohydrate metabolism in *Bemisia argentifolii* biochemical mechanism and physiological role for trehalulose synthesis in the silverleaf whitefly. p. 101. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
1174. Sances, F.V., Ballmer, G., Reuter, L. and Toscano, N. 1993. [chemical control, tomatoes, equipment, hydraulic sprayer, Degania, Berthoud, aircraft, soap, oil, pyrethrum, azadiractin]. p. 76. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1175. Sánchez-Campos, S., Diaz, J.A., Soria, C., Camero, R. and Moriones, E. 1999. Relative importance of tomato yellow leaf curl virus-Is and -Sr species in infections of tomato in Spain. p. 71. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce (Almeria), Spain. [Abstract]
1176. Sánchez-Campos, S., Navas-Castillo, J., Diaz, J.A., Reina, J., Bejarano, E.R. and Moriones, E. 1999. Weeds as reservoirs of tomato yellow leaf curl viruses in Spain. p. 85. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce (Almeria), Spain. [Abstract]
1177. Sanchez, V., Frias, G., Aguirre, L., Bravo, L., Cortez, E., Quezada, J., Ruiz, R., Hernandez, M., Rangel, U., Garzon, H., Rivera, R., Gilbertson, R. and Rojas, M. 1996. Agroecology of a whitefly-transmitted geminivirus in an arid and isolated serrano pepper growing region in northeastern Mexico. *Phytopathology* 86(11 SUPPL.): S117. [Abstract]
1178. Sanderson, J.P., Davis, P.M. and Ferrentino, G.W. 1993. [poinsettia]. p. 19. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1179. Sanderson, J.P. and Ferrentino, G.W. 1993. [*Encarsia*, poinsettia]. p. 108. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1180. Sanderson, J.P., Roush, R.T. and Derksen, R.C. 1993. [poinsettia, chemical control]. p. 77. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1181. Sanjrani, M.W., Baloch, A.A. and Kalroo, A.M. 1996. Studies on the biology of *Chrysoperla carnea* Stephens (Neuroptera: Chrysopidae) on different hosts. p. 6. *In* M.J. Iqbal and A. Kasana (Eds.), Second International Congress of Entomological Sciences, March 19-21, 1996 Islamabad, Pakistan. [Abstract]
1182. Sattayawirut, T., Charoenrak, T. and Sepsawat, P. 1982. Effectiveness of some insecticides against white fly (*Bemisia tabaci* Gennadius) on soybean. p. 139. *In* Research Reports 1980. Department of Agriculture, Entomology and Zoology Division, Bangkok, Thailand. [Thai, Abstract]
1183. Saxena, S., Hallan, V., Singh, B.P. and Sane, P.V. 1998. Leaf curl disease of *Carica papaya* from India may be caused by a bipartite geminivirus. *Plant Dis.* 82(1): 126. [Abstract]
1184. Schaal, D. 1994. Airtec air boom sprayers as a more efficient spray alternative. p. 109. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
1185. Schauff, M.E., Woolley, J.B., Rose, M., Zolnerowich, G., Evans, G. and Williams, R. 1994. Taxonomy of *Bemisia tabaci* parasites. p. 148. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
1186. Schuster, D.J. 1997. Distribution of immature life stages of the silverleaf whitefly on tomato plants. p. 75. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
1187. Schuster, D.J. 1997. Toxicity of biorational pesticides to the lacewing species *Ceraeochrysa cubaba*. p. 129. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
1188. Schuster, D.J., Bennett, F.D., Evans, G.A., Price, J.F. and Dean, D.E. 1993. [parasites, *Eretmocerus*, *Encarsia*, *Delphastus* predators]. p. 109. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1189. Schuster, D.J., Cszinszky, A.A., Dean, D.E. and Polston, J.E. 1996. UV-reflective plastic soil mulches for management of the silverleaf whitefly and tomato mottle geminivirus on tomato. p. 169. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]

- 1190.Schuster, D.J., Osborne, L.S., Price, J.F., Dean, D.E. and Stansly, P.A. 1994. Integration of natural enemies for management of the sweetpotato whitefly. p. 149. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
- 1191.Schuster, D.J., Polston, J.E. and Price, J.F. 1993. [tomato, cabbage, potato, cucumber, squash, weeds]. p. 19. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
- 1192.Schuster, D.J., Scott, J.W., Barten, J.H.M. and Polston, J.E. 1993. [tomato, trichomes, *Lycopersicon*]. p. 129. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
- 1193.Schuster, D.J., Scott, J.W., Thome, C.R. and Polston, J.E. 1994. Host plant resistance for management of the sweetpotato whitefly and tomato mottle geminivirus on tomato. p. 168. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
- 1194.Schuster, D.J. and Stansly, P.A. 1994. Expression of plant damage by *Bemisia*. *Phytoparasitica* 22(4): 322. [ Abstract]
- 1195.Schuster, D.J., Stansly, P.A., Allen, J.C., Brewster, C. and Polston, J.E. 1994. Evaluation of crop associations for management of the sweetpotato whitefly. p. 181. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
- 1196.Schuster, D.J., Stansly, P.A., Dean, D.E. and Polston, J.E. 1996. Potential of companion plantings for managing silverleaf whitefly and tomato mottle geminivirus on tomato. p. 168. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
- 1197.Scott, J.W., Schuster, D.J., Barten, J.H.M., Stevens, M.R., Thome, C.H. and Polston, J.E. 1994. Introgression of resistance to whitefly-transmitted geminiviruses. *Phytoparasitica* 22(4): 329-330. [ Abstract]
- 1198.Seal, D., Baranowski, R., McMillan, R., Jr. and Bryan, H. 1994. Management of whitefly, *Bemisia tabaci* (Gennadius) and its associated silverleaf disorder on squash. p. 110. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
- 1199.Seligman, I.M. 1987. Comparison of responses of three populations of *Bemisia tabaci* to a range of insecticides. *Phytoparasitica* 15: 264-265. [ Abstract]
- 1200.Sereroglu, E., Karut, K., Yildiz, S. and Kazak, C. 1998. Spatial distribution of preimaginal *Bemisia tabaci* (Homoptera: Aleyrodidae) in cotton. p. 171. *In* U. Kechagia (Ed.), World Cotton Research Conference 2. Sept. 6-12, 1998. Athens Greece. [Abstract]
- 1201.Serra, C., Concepcion, S., Polston, J., Ortiz, M. and Benoit, P. 1994. Host range of the tomato leaf curl virus in the northwestern Dominican Republic and attempts at its control. *Phytoparasitica* 22(4): 326-327. [ Abstract]
- 1202.Serra, C.A. 1994. Preliminary results of an intended host -free period in different northwestern Dominican tomato growing areas on the population dynamics of whiteflies and the incidence of geminivirus. p. 28. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
- 1203.Serra, C.A., Ortíz, M., Nuñez, J.B., Schulz, A. and Benoit, P.F. 1996. Potential of entomophagous fungi to control whiteflies on Dominican vegetable and ornamental crops. p. 142. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
- 1204.Serra, C.A., Ortíz, M.O., Nuñez, J.B. and Benoit, P.F. 1996. Survey of indigenous natural enemies of whiteflies in the Dominican Republic. p. 143. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
- 1205.Servin, R., Martinez-Carrillo, J.L., Troyo, E. and Ortega, A. 1995. Susceptibility tests for some common insecticides used for *Bemisia tabaci* (Gennadius) collected from cabbage in La Paz, B.C.S., Mexico. p. 92. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
- 1206.Servin, R., Martinez, J.L., Troyo-Diequez, E. and Ortega, A. 1995. Toxicological assays in *Bemisia tabaci* (Genn.) from cabbage plots in La Paz, Baja California Sur, Mexico. *Hortscience* 30(4): 830. [ Abstract]
- 1207.Severson, R.F., Stephenson, M.G., Chortyk, O.T., Maw, B.W., Neal, J.W., Jr., Pittarelli, G.W. and Buta, J.G. 1993. [*Nicotiana gossei*, extraction]. p. 78. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
- 1208.Severson, R.F., Stephenson, M.G., Chortyk, O.T., Maw, B.W., Neal, J.W. Jr., Pittarelli, G.W., Buta, J.G., Jackson, D.M. and Johnson, A.W. 1994. Field production of *Nicotiana* species. p. 111. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
- 1209.Severson, R.F., Stephenson, M.G., Sisson, V.A., Jackson, D.M., Chortyk, O.T. and Herzog, G.A. 1993. [*Nicotiana*, biorational insecticides]. p. 79. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
- 1210.Shapiro, J.P. 1994. Insect-host plant interactions and expression of damage. *Phytoparasitica* 22(4): 322. [ Abstract]
- 1211.Shapiro, J.P. 1996. Phytochemicals induced in cucurbits by *Bemisia*. p. 43. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
- 1212.Shapiro, J.P., Jimenez, D.R., Yokomi, R.K. and Mayer, R.T. 1993. [proteins, pumpkin, squash]. p. 42. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
- 1213.Shapiro, J.P., Yokomi, R.K., Osborne, L.S. and Jimenez, D.R. 1994. Comparison of sweetpotato whitefly (*Bemisia tabaci*)-induced silverleaf with plant growth regulator-induced leaf silvering. p. 59. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
- 1214.Sharaf, N. and Batta, Y. 1984. Effect of temperature on life history of *Eretmocerus mundus* Mercet (Hymenoptera, Aphelinidae). p. 569. *In* XVII Int. Congress Entomol. [Abstract]
- 1215.Sharp, L.P., Hou, Y.M., Garrido-Ramirez, E.R., Guzman, P. and Gilbertson, R.L. 1999. Synergistic interactions among components of whitefly-transmitted geminiviruses. p. 34. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
- 1216.Shoham, C. 1989. Activity of *Bemisia tabaci* in vegetables in 1987 and 1988 [Israel]. *Phytoparasitica* 17(3): 227-228. [ Abstract]
- 1217.Sim, J. and Valverde, R.A. 1999. Properties of a geminivirus from *Dicliptera sexangularis*. *Phytopathology* 89(6 Suppl.): S72-S73. [ Abstract]
- 1218.Sim, J. and Valverde, R.A. 1999. Whitefly transmission and detection of sweetpotato chlorotic stunt virus. *Phytopathology* 89(6 Suppl.): S73. [ Abstract]

1219. Simmons, A.L. and Dennehy, T.J. 1996. Contrasts of three insecticide resistance monitoring methods for whitefly. p. 96. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1220. Simmons, A.L., Williams, L. III, Dennehy, T.J., Antilla, L., Jech, L.E. and Husman, S. 1997. Investigations of two insect growth regulators against Arizona whitefly populations. p. 168. *In Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001).* U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
1221. Simmons, A.M. 1993. [parasites, beans, *Eretmocerus*, *Encarsia*]. p. 110. *In Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1222. Simmons, A.M. 1994. Ovipositional response to leaf surfaces. p. 60. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
1223. Simmons, A.M. 1995. Abundance of parasitoids of *Bemisia argentifolii* in sweetpotato. p. 136. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1224. Simmons, A.M. 1995. Parasitoids of *Bemisia argentifolii* in vegetable crops with and without Lorsban. p. 137. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1225. Simmons, A.M. 1998. Subsistence of *Bemisia* on upper and lower leaf surfaces of selected vegetables. p. 99. *In Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
1226. Simmons, A.M. and Chortyk, O.T. 1995. *Nicotiana* against *Bemisia argentifolii* in three vegetable crops. p. 93. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1227. Simmons, A.M. and Ciomperlik, M.A. 1996. Exotic parasitoid of *Bemisia argentifolii* in South Carolina. p. 144. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1228. Simmons, A.M. and Ciomperlik, M.A. 1998. Establishment of exotic *Eretmocerus* (Pakistan strain) in South Carolina. p. 82. *In Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
1229. Simmons, A.M. and Elsey, K.D. 1994. Seasonal parasitism in South Carolina. p. 150. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
1230. Simmons, A.M. and Hoelmer, K.A. 1998. Capture of *Bemisia* parasitoids on yellow sticky traps and rate of parasitism. p. 83. *In Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
1231. Simmons, A.M. and Jackson, D.M. 1996. Abundance of parasitoids of *Bemisia argentifolii* in imidacloprid treated vegetables. p. 170. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1232. Simmons, A.M. and Jackson, D.M. 1997. Abundance of native whitefly parasitoids following foliar insecticide treatments of vegetables. p. 169. *In Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001).* U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
1233. Simmons, A.M. and Jackson, D.M. 1997. Ultrasonic fogging device for managing whiteflies. p. 130. *In Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001).* U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
1234. Simmons, A.M. and Jackson, D.M. 1998. Ultrasonic fogging device: managing whiteflies in greenhouses. p. 46. *In Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
1235. Simmons, A.M. and McCreight, J.D. 1994. Breeding melons for resistance. p. 169. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
1236. Simmons, A.M. and McCreight, J.D. 1998. Assays of melons for resistance to *Bemisia*. p. 100. *In Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
1237. Simmons, A.M. and McCutcheon, G.S. 1999. Attractancy of crops to a parasitoid of *Bemisia* and daily foraging [*Encarsia pergandiella*]. p. 88. *In Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
1238. Simmons, A.M., McCutcheon, G.S., Dufault, R.J., Hassell, R.L. and Rushing, J.W. 2000. *Bemisia* and associated parasitoids on species of medicinal herbal plants. p. 28. *In Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan.* U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
1239. Simmons, G.S., Hoelmer, K., Jaronski, S. and Lord, J. 1996. Effect of *Beauveria bassiana* GHA (Mycotrol WP) on parasitoids of *Bemisia* in spring melons. p. 145. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1240. Simmons, G.S., Hoelmer, K., Staten, R. and Boratynski, T. 1995. Biological control of *Bemisia* in spring melons. p. 138. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1241. Simmons, G.S., Hoelmer, K., Staten, R. and Boratynski, T. 1996. Seasonal inoculations in spring melons with parasitoids of *Bemisia*. p. 146. *In Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1242. Simmons, G.S., Hoelmer, K., Staten, R. and Boratynski, T. 1998. Biological control of silverleaf whitefly infesting cantaloupe with large scale releases of exotic parasitoids in the Imperial Valley of California. p. 84. *In Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan.* U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
1243. Simmons, G.S., Hoelmer, K., Staten, R., Boratynski, T. and Natwick, E. 1997. Seasonal inoculative biological control with parasitoids of *Bemisia* infesting cantaloupe in the Imperial Valley of California: a report on three years of investigations. p. 170. *In Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001).* U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]

1244. Simmons, G.S. and Minkenberg, O.P.J.M. 1994. Development of augmentative biological control for sweetpotato whiteflies on cotton and field vegetables in the Imperial Valley. p. 151. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
1245. Simone, G.W., Brown, J.K., Hiebert, E. and Cullen, R.C. 1990. Geminiviruses associated with epidemics in Florida tomatoes and peppers. *Phytopathology* 80: 1063. [Abstract]
1246. Singh, J., Sohi, A.S., Brar, D.S., Denholm, I. and Russell, D. 1998. Management of cotton leaf curl virus disease in India. p. 273. *In* U. Kechagia (Ed.), World Cotton Research Conference 2. Sept. 6-12, 1998. Athens Greece. [Abstract]
1247. Sippell, D.W., Bindra, O.S. and Khalifa, H. 1983. Resistance in cotton to whitefly (*Bemisia tabaci*). p. 841. *In* 10th International Congress of Plant Protection, Proceedings of a Conference Held at Brighton, England 20-25 November 1983. Plant Protection for Human Welfare. British Crop Protection Council, Croydon, UK. [Abstract, Cock (1986)]
1248. Sisson, V.A. and Severson, R.F. 1993. [*Nicotiana*, biorational insecticide]. p. 80. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1249. Sivasupramaniam, S., Dennehy, T.J. and Williams, L. III. 1997. Management of pyrethroid-resistant whiteflies in Arizona cotton: selection, cross-resistance, and dynamics. p. 171. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
1250. Sivasupramaniam, S., Johnson, S., Watson, T.F., Osman, A.A. and Jassim, R. 1996. A four-year study on monitoring resistance of the silverleaf whitefly to organophosphorus+pyrethroid, cyclodiene, and pyrethroid insecticides in Arizona. p. 97. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1251. Sivasupramaniam, S. and Watson, T.F. 1996. Selection for fenpropathrin and fenpropathrin+acephate resistance in the silverleaf whitefly (Homoptera: Aleyrodidae) and reversion of fenpropathrin+acephate resistance. p. 98. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1252. Skinner, R.H. and Cohen, A.C. 1994. Morphological and physiological parameters associated with sweetpotato whitefly host selection. p. 61. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
1253. Smith, M.T. 1993. [parasites, *Encarsia*]. p. 110. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1254. Smith, M.T. 1995. Evaluation of exotic parasitoid species from areas of *Bemisia tabaci* origin. p. 139. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1255. Smith, M.T. 1996. Efficacy of parasitoid species against silverleaf whitefly. p. 147. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1256. Smith, M.T., Alfred, M. and Ciomperlik, M. 1996. Identification of the whitefly and associated parasitoid species in Mississippi. p. 148. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1257. Smith, M.T., Allred, M., Fieselman, D., Ciomperlik, M., Layton, B., Jarratt, J., Harris, P., Milling, A. and Snyder, R. 1997. Survey and identification of the whitefly and associated parasitoid species in Mississippi. p. 172. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
1258. Smith, W.J., Smith, C.W., Meagher, R.L. and Norman, J.W. 1993. [cotton, breeding]. p. 130. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1259. Snyder, J.C. and Simmons, A.M. 1995. Day length affects oviposition preference of *Bemisia* on *Lycopersicon hirsutum*. p. 153. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1260. Sosa-Coronel, J. 1995. Response of broccoli cultivars to *Bemisia argentifolii* Bellows and Perring. p. 154. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1261. Sosa-Gómez, D.R., Moscardi, F. and Santos, M. 1997. *Bemisia* spp. na cultura da soja: ocorrência, controle químico e incidência de fungo entomopatogênico *Paecilomyces* spp. p. 144. *In* 16th Congresso Brasileiro de Entomologia, Resumos. SEB/EMBRAPA-CNPMPF, Salvador, Brazil. [Abstract]
1262. Srivastava, K.P., Varma, A. and Ahlawat, Y.S. 1995. Biology of whitefly, *Bemisia tabaci*, and its role as a vector of plant viruses. p. 46. *In* Symposia on National Water Policy, Vector Biology and Integrated On-farm and Off-farm Employment, 2nd Agricultural Science Congress 19-21 January. National Academy of Agricultural Sciences, Andhra Pradesh Agricultural University, Rajendranagar, Hyderabad, India. [Abstract]
1263. Sserubombwe, W.S., Thresh, J.M., Otim-Nape, G.W. and Osiru, D.S.O. 1999. The effect of varietal mixtures on the progress of cassava mosaic virus disease grown under epidemic conditions in Uganda. p. 87. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce (Almeria), Spain. [Abstract]
1264. Sseruwagi, P., Legg, J.P. and Otim-Nape, G.W. 1999. An overview of the incidence of cassava mosaic disease in East Africa, 1998 update. pp. 87-88. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce (Almeria), Spain. [Abstract]
1265. Stansly, P.A. 1994. *B. tabaci* control & tomato mottle geminivirus (TMoV) in FL [Florida] staked tomato. p. 112. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
1266. Stansly, P.A. 1999. Control of silverleaf whitefly and turnip aphid on collards with foliar and soil-applied systemic insecticides. p. 62. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
1267. Stansly, P.A. 1999. Management of geminivirus epidemics of field-grown tomato in Florida and the Dominican Republic. pp. 71-72. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce (Almeria), Spain. [Abstract]
1268. Stansly, P.A. 2000. Impact and management of tomato yellow leafcurl virus in Southwest Florida. p. 44. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
1269. Stansly, P.A. and Cawley, B.M. 1993. [tomato, chemical control]. p. 81. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]



1270. Stansly, P.A. and Liu, T.X. 1994. Bioassays of biorational insecticides for sweetpotato whitefly control. p. 113. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
1271. Stansly, P.A., Liu, T.X. and Schuster, D.J. 1994. Role of biorational insecticides in the management of the silverleaf whitefly, *Bemisia argentifolii*. *Phytoparasitica* 22(4): 350-351. [Abstract]
1272. Stansly, P.A., Liu, T.X. and Schuster, D.J. 1995. The rationality of biorational insecticides for control of silverleaf whitefly (*Bemisia argentifolii*). p. 94. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1273. Stavely, J.R., McMillan, R.T., Jr., Beaver, J.S. and Miklas, P.N. 1998. Three McCasain type, indeterminate, rust and golden mosaic resistant snap bean germplasm lines, Beldade-RGMR-4, -5, and -6. p. 101. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
1274. Steinberg, S. and Prag, H. 1994. Efficacy of the fungus *Aschersonia aleyrodis* and a coccinellid predator *Delphastus pusillus*, used to control *Bemisia tabaci* in greenhouse cucumber. *Phytoparasitica* 22(4): 341. [Abstract]
1275. Stenseth, C. and Singh, H.M. 1989. [Buprofezin against *Trialeurodes vaporariorum* and *Bemisia tabaci*]. p. 239. *In* Nordic Plant Protection Conference 1989: Section of Botany, Zoology, Nematology, Virology, Weed and Chemistry, Helsingør, Denmark, 5-6 Dec 1989. Statens Planteavltsforsoeg, Lyngby, Denmark. [Norwegian, Abstract]
1276. Stephenson, M.G., Severson, R.F., Maw, B.W. and Pittarelli, G.W. 1993. *Nicotiana*. p. 82. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1277. Stern, T.U. and Gerling, D. 1993. The influence of whitefly (*Bemisia tabaci*) density on fitness of *Delphastus pusillus* (Coleoptera: Coccinellidae). *Phytoparasitica* 21(2): 173. [Abstract]
1278. Stout, J.T., Liu, H.T., Polston, J.E., Gilbertson, R.L., Nakhla, M.K., Hanson, S.F. and Maxwell, D.P. 1997. Engineered rep gene-mediated resistance to tomato mottle geminivirus in tomato. *Phytopathology* 87(6 SUPPL.): S94. [Abstract]
1279. Summer, H.R., Womac, A.R., Herzog, G.A. and Chandler, L.D. 1993. [chemical control, sprayer, Hagie, Proptec, Electrostatic, Berthoud, Degania, Hydrapak, peanuts]. p. 83. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1280. Summers, C.G. and Estrad, D. 1996. Chlorotic streak of bell pepper: a new toxicogenic disorder induced by feeding of the silverleaf whitefly, *Bemisia argentifolii*. *Plant Dis.* 80(7): 822. [Abstract]
1281. Syed, T.S., Abro, G.H., Khuro, R.D. and Dhaunroo, M.H. 1996. Relative resistance of cotton varieties against sucking insect pests. p. 52. *In* M.J. Iqbal and A. Kasana (Eds.), Second International Congress of Entomological Sciences, March 19-21, 1996 Islamabad, Pakistan. [Abstract]
1282. Taher, M.M. 1994. Introduction to the whitefly *Bemisia tabaci* problem and its control in the Near East region. p. 13. *In* B. Ezzahiri and M. Bouhache (Eds.), Fifth Arab Congress of Plant Protection, November 27 - December 2, 1994, Fez, Morocco. Actes Editions, Rabat, Maroc. [Abstract]
1283. Teppidargarn, P., Jeerajunya, K. and Kittibunya, S. 1984. Relationship between adult whitefly in sticky yellow trap and its larval population in cotton field. p. 19. *In* Research Report in 1984: Fiber Crop, Sugarcane, Tobacco. Department of Agriculture, Bangkok, Thailand. [Thai, Abstract]
1284. Teuber, L.R., Gibbs, L.K. and Taggard, K.L. 1999. Progress in breeding alfalfa for resistance to the silverleaf whitefly. p. 3. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
1285. Tonhasca, A., Palumbo, J.C. and Byrne, D.N. 1994. Impact of pesticide applications on the distribution of *Bemisia tabaci*. p. 29. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
1286. Torres, V. and Zapata, M. 2000. Bacterial isolations from common insects of pepper (*Capsicum annuum*). *Phytopathology* 90(Suppl. 6): S124. [Abstract]
1287. Toscano, N., Ballmer, G., Reuter, L. and Sances, F. 1993. [lettuce, tomato, chemical control]. p. 84. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1288. Toscano, N., Prabhaker, N., Zhou, S. and Ballmer, G. 1998. Toxicity of Applaud and Knack against silverleaf whiteflies from Southern California: Implications for susceptibility monitoring. p. 48. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
1289. Toscano, N.C., Ballmer, G.R., Beehler, L.L. and Henneberry, T.J. 1994. Determination of temporal activity and directional infestation gradients of silverleaf whitefly, *Bemisia argentifolii*. p. 62. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
1290. Toscano, N.C., Ballmer, G.R., Beehler, L.L. and Henneberry, T.J. 1994. Evaluation of broad spectrum vs. biorational insecticides. p. 114. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
1291. Toscano, N.C., Castle, N.P., Castle, S. and Henneberry, T.J. 1996. Monitoring insecticide resistance in the silverleaf whitefly (*Bemisia argentifolii*) populations. p. 184. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1292. Toscano, N.C. and Henneberry, T. 1994. Population dynamics of *Bemisia* species (*B. argentifolii* and *B. tabaci* in agricultural systems. p. 14. *In* B. Ezzahiri and M. Rouhache (Eds.), Fifth Arab Congress of Plant Protection, November 27 - December 2, 1994, Fez, Morocco. Actes Editions, Rabat, Maroc. [Abstract]
1293. Toscano, N.C., Henneberry, T.J., Prabhaker, N. and Castle, S. 1994. Formulate management strategies to extend the effectiveness of chemicals needed for silverleaf whitefly control. *Phytoparasitica* 22(4): 346-347. [Abstract]
1294. Toscano, N.C., Prabhaker, N., Castle, S.J. and Henneberry, T. 1999. Managing whiteflies (*Bemisia tabaci*), strain B in an agricultural system. p. 169. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce (Almeria), Spain. [Abstract]
1295. Toscano, N.C., Prabhaker, N., Castle, S.J., Henneberry, T.J., Ballmer, G., Natwick, E. and Kallenbach, R. 1997. Areawide resistance monitoring of sweetpotato and silverleaf whiteflies, *Bemisia tabaci* and *B. argentifolii* (Homoptera, Aleyrodidae) in California. p. 131. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
1296. Toscano, N.C., Yoshida, H.A. and Henneberry, T.J. 1998. Responses to azadirachtin and pyrethrum by two species of *Bemisia* (Homoptera: Aleyrodidae). p. 47. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]

1297. Tsai, J.H. and Wang, K. 1995. Characteristics of bean golden mosaic virus transmitted by silverleaf whitefly (*Bemisia argentifolii*) in South Florida. p. 47. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1298. Tsai, J.H. and Wang, K. 1995. Development and reproduction of silverleaf whitefly *Bemisia argentifolii* (Homoptera: Aleyrodidae) on five vegetable crops. p. 17. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1299. Ulubilir, A. and Yabas, C. 1996. Population fluctuations, natural enemies and chemical control possibilities of cotton whitefly, *Bemisia tabaci* Genn., on vegetables in Cukurova. [Turkish, English summary]. *Zirai Mucadele Arastirma Yilligi* 28-29: 18. [Abstract]
1300. Umeda, K., Chu, C.C. and Henneberry, T.J. 1996. Efficacy and trap comparison in monitoring adult whitefly population densities in cantaloupe. p. 44. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1301. Umeda, K., Chu, C.C. and Henneberry, T.J. 1998. Comparison of leaf-turn and CC Trap methods to estimate adult whitefly densities in commercial spring melon fields. p. 49. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
1302. Urbino, C., Caruana, M.L., Sauvion, N. and Pavis, C. 1999. Potato yellow mosaic virus (PYMV) on tomato in Guadeloupe: characterization of the virus and its vector [French West Indies]. p. 35. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
1303. Vacek, D.C. 1993. [genetic markers, genome, polymerase chain reaction]. p. 43. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1304. Vacek, D.C. and Ruiz, R.A. 1995. RAPD-PCR identification of natural enemies of SPWF. p. 140. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1305. Vacek, D.C., Ruiz, R.A. and Wendel, L.E. 1996. RAPD-PCR identification of natural enemies of SPWF. p. 149. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1306. Vacek, D.C., Ruiz, R.A. and Wendel, L.E. 1997. RAPD-PCR identification of natural enemies of SPWF. p. 173. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
1307. Valderrama, H., Velasquez, A. and Fernandez, O. 1994. Transmission y ambitos de hospederos de un geminivirus asociado con *B. tabaci* en tomate. [Transmission and circuit of host of a geminivirus associated with *B. tabaci* in tomatoes]. p. 17. *In* Jornada Agropecuaria Region Oriental, Panama, 25 Ago 1994. Instituto De Investigacion Agropecuaria De Panama, Divisa, Panama. [Spanish, Abstract]
1308. Valenzuela, M.A., Gomez, C.O., Brown, J.K. and Lambe, R.C. 1988. Evaluation of a perforated polyethylene floating row cover for virus control in two squash hybrids. *Biological and Cultural Tests for Control of Plant Diseases. Am. Phytopathol. Soc.* 3: 27. [Abstract]
1309. Valverde, R.A. 1999. Whitefly-transmitted viruses infecting sweetpotato in the United States. pp. 69-70. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce (Almeria), Spain. [Abstract]
1310. van de Ven, W.T.G., LeVesque, C.S., Cooper, A., Perring, T.M. and Walling, L.L. 1997. Accumulation and reduction of mRNA transcripts in squash leaves upon which silverleaf whitefly has fed. p. 102. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
1311. van de Ven, W.T.G., LeVesque, C.S., Perring, T.M. and Walling, L.L. 2000. Characterization of two squash genes induced by silverleaf whitefly infestation. p. 120. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
1312. van de Ven, W.T.G., Perring, T.M. and Walling, L.L. 1998. Genes involved in silverleaf whitefly induced squash leaf silencing. p. 102. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
1313. van de Ven, W.T.G., Perring, T.M. and Walling, L.L. 1999. Genes modulated in silverleaf whitefly infested squash. p. 108. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
1314. van Giessen, W.A. and Jackson, D.M. 1996. Insect adaptation effects on plant resistance to silverleaf whitefly, *Bemisia argentifolii* Bellows & Perring. p. 171. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1315. van Giessen, W.A., Mollema, C. and Elsey, K.D. 1995. Development of a simulation model for evaluating plant germplasm for antibiotic resistance to whitefly. p. 155. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1316. van Iersel M. and Oetting, R. 1999. Imidacloprid applications for whitefly control on poinsettia using ebb-and-flow systems. *Hortscience* 34(3): 540-541. [Abstract]
1317. van Lenteren, J.C., Brasch, K. and Henter, H. 1994. Biological control of *Bemisia tabaci* with *Encarsia formosa*: a realistic opinion? *Phytoparasitica* 22(4): 338. [Abstract]
1318. van Schelt, J. 1994. The use of *Macrolophus caliginosus* as a whitefly predator in protected crops. *Phytoparasitica* 22(4): 334. [Abstract]
1319. Veenstra, K.H. and Byrne, D.N. 1995. Physiological adaptations for dispersal in the sweet potato whitefly. p. 18. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1320. Vega, F.E., Jackson, M.A. and McGuire, M.R. 1997. Germination of conidia and blastospores of the fungal entomopathogen *Paecilomyces fumosoroseus* on silverleaf whiteflies. p. 174. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
1321. Vega, F.E., McGuire, M.R., Jackson, M.A. and Cliquet, S. 1996. Formulations of the fungal entomopathogen *Paecilomyces fumosoroseus*. p. 150. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]

- 1322.Veierov, D., Aharonson, N., Eliyahu, M., Fenigstein, A., Cohen, S. and Kletter, E. 1994. Non-toxic formulations for the control of the sweetpotato whitefly (*Bemisia tabaci*). *Phytoparasitica* 22(4): 351. [ Abstract]
- 1323.Veierov, D., Aharonson, N., Kletter, E., Eliyahu, M., Fenigstein, A., Cohen, S. and Ben-Joseph, R. 1993. Non-toxic behaviour -interfering formulations for the control of the tobacco whitefly, *Bemisia tabaci*. *Phytoparasitica* 21(2): 177-178. [ Abstract]
- 1324.Veierov, D. and Fenigstein, A. 1989. The type and behaviour of foliar residue, as related to *Bemisia tabaci* management. *Phytoparasitica* 17(3): 231. [ Abstract]
- 1325.Veierov, D., Fenigstein, A., Bar-Joseph, R. and Cohen, S. 1990. Interference of foliar residues of fenprothrin and of several other toxic and non-toxic chemicals with the landing behavior of adult tobacco whitefly (*Bemisia tabaci*) on cotton. p. 353. *In* H. Frehse, E. Kesseler-Schimitz and S. Conway (Eds.), Seventh International Congress of Pesticide Chemistry. Hamburg, Germany. [Abstract]
- 1326.Vidal, C., Lacey, L., Kirk, A. and Fargues, J. 1996. Intraspecific variability of *Paecilomyces fumosoroseus* (Deuteromycotiana: Hyphomycetes): Pathogenic activity of *Bemisia tabaci* (Homoptera: Aleyrodidae). p. 151. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
- 1327.Vidal, C., Lacey, L.A. and Fargues, J. 1996. Intraspecific variability of *Paecilomyces fumosoroseus*: vegetative growth as a function of temperature. p. 152. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
- 1328.Vidavsky, S. and Czosnek, H. 1999. Tomato breeding lines derived from *Lycopersicon hirsutum* that are immune and tolerant to tomato yellow leaf curl virus (TYLCV). p. 73. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce(Almeria), Spain. [Abstract]
- 1329.Vir, S. 1983. Assessment of yield loss in mothbean and cowpea crops due to insect pests, and their control. p. 1215. *In* 10th International Congress of Plant Protection: Proceedings of a Conference Held at Brighton, England 20-25 November 1983. Plant Protection for Human Welfare. British Crop Protection Council, Croydon, UK. [Abstract, Cock (1986)]
- 1330.Vir, S. 1999. Ecology and population dynamics of *Bemisia tabaci* Genn (Aleyrodidae) an extent of damage both by vector and yellow mosaic virus [India]. p. 21. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
- 1331.Vir, S. 1999. Integrated management of *Bemisia tabaci* Genn (Aleyrodidae) and yellow mosaic virus in mothbean (*Vigna aconitifolia*) crop [India]. p. 124. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Second Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1999-01. [Abstract]
- 1332.Vonghirunpinyo, L., Kittipakorn, K. and Choopanya, D. 1981. Study on mungbean yellow mosaic [transmitted by *Bemisia*]. pp. 15-16. *In* Research Reports, 1978. Department of Agriculture, Plant Pathology and Microbiology Division, Bangkok, Thailand. [Thai, Abstract]
- 1333.Wagner, T.L. 1994. Temperature-dependent reproduction of sweetpotato whitefly on cotton. p. 30. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
- 1334.Walker, G.P. 1997. Feeding behavior of adult *Bemisia argentifolii* on host and non-host plants. p. 103. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
- 1335.Walker, G.P. and Johnson, D.D. 1998. Feeding behavior may explain why nonpersistent viruses are transmitted primarily by aphids not whiteflies. p. 30. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
- 1336.Walker, G.P., Johnson, D.D. and Costa, H. 2000. Behavioral response of silverleaf whitefly adult females to plant species varying in host suitability. p. 121. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
- 1337.Walker, G.P. and Perring, T.M. 1994. Feeding and oviposition behavior of whiteflies. p. 64. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
- 1338.Walker, G.P. and Perring, T.M. 1995. Correlations of silverleaf whitefly feeding and oviposition behaviors with AC electronic feeding monitor waveforms. p. 49. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
- 1339.Walling, L.L. 2000. New genes and new signals: The *Bemisia argentifolii* - squash interaction. pp. 3-4. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
- 1340.Walling, L.L., Puthoff, D.P. and Perring, T.M. 2000. Whitefly feeding induces local and systemic changes in tomato gene expression. p. 122. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
- 1341.Watson, T.F., Sivasupramaniam, S., Johnson, S., Osman, A.A. and Jassim, R. 1995. Development of a resistance-management strategy for the silverleaf (sweetpotato) whitefly. p. 95. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
- 1342.Watson, T.F., Tellez, M.A., Johnson, S.E., Sivasupramaniam, S. and Brown, P.W. 1995. The sweetpotato whitefly, *Bemisia tabaci*, (Gennadius) in Arizona: ecological factors affecting outbreaks and control in cotton. p. 19. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
- 1343.Weaver, J.S. and Ciomperlik, M. A. 2000. Biological control of *Bemisia argentifolii* (Aleyrodidae) infesting poinsettias: evaluation of *Encarsia formosa*, Nile Delta strain, (Aphelinidae) and *Serangium parcesetosum* (Coccinellidae). p. 102. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
- 1344.Weaver, J.S. and Ciomperlik, M.A. 2000. Dispersal of *Serangium parcesetosum* (Coccinellidae) on poinsettia infested with *Bemisia argentifolii* (Aleyrodidae) in greenhouse trials. p. 101. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
- 1345.Webb, S.E. and Polston, J.E. 1996. Transmission of tomato mottle geminivirus by *Bemisia argentifolii*. p. 45. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
- 1346.Weddle, L., Weddle, R. and Culver, G. 1995. Survey of *Bemisia tabaci* on the Colorado Desert, Imperial Valley, California. p. 20. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]

1347. Wee, L.Y., Toscano, N.C., Hendrix, D.L. and Henneberry, T.J. 1997. Effect of insecticide application on *Bemisia argentifolii* (Homoptera: Aleyrodidae) densities, honeydew production, and cotton yields. p. 77. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
1348. Weisz, E. and Gerling, D. 1994. Leaf-settling behaviour of *Bemisia tabaci*. *Phytoparasitica* 22(4): 314. [Abstract]
1349. Wendt, K., Wyatt, S.D. and Brown, J.K. 1994. Detection of DNA components of two strains of squash leaf curl virus using polymerase chain reaction. *Phytopathology* 84: 1104. [Abstract]
1350. Wessel-Beaver, L. 1997. Resistance in *Cucurbita* spp. to silverleaf and sweetpotato whitefly. *Hortscience* 32(3): 450. [Abstract]
1351. Williams, L., III, Dennehy, T.J. and Palumbo, J.C. 1996. Whitefly control in Arizona: development of a resistance management program for imidacloprid. p. 99. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1352. Williams, L., III, Dennehy, T.J. and Palumbo, J.C. 1997. Defining the risk of resistance to imidacloprid in Arizona populations of whitefly. p. 175. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
1353. Williams, R.J. 1975. A whitefly transmitted mosaic of lima bean in Nigeria. *Trop. Grain Legume Bull.* 1(1): 11. [Abstract, Cock (1986)]
1354. Wintermantel, W.M. and Cortez, A.A. 2001. Complementation for transmission by non-vector whiteflies among tomato-infecting criniviruses. *Phytopathology* 91(Suppl. 6): S96. [Abstract]
1355. Wisler, G.C. and Cortez, A.A. 2000. Differential transmission characteristics among four whitefly vectors of tomato chlorosis crinivirus. p. 45. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
1356. Wisler, G.C., Duffus, J.E. and Liu, H.Y. 1999. Expansion of tomato-infecting criniviruses into new areas. p. 70. *In* Plant virus epidemiology: Current status and future prospects. VII International Plant Virus Epidemiology Symposium, Aguadulce (Almeria), Spain. [Abstract]
1357. Wisler, G.C., Duffus, J.E., Liu, H.Y., Li, R.H., Simone, G.W. and Hochmuth, R.C. 1996. A new, whitefly-transmitted virus infecting tomato from Florida. *Phytopathology* 86 (11 SUPPL): S71-72. [Abstract]
1358. Wisler, G.C., Li, R.H., Liu, H.Y. and Duffus, J.E. 1997. Partial molecular and cytological analyses of tomato chlorosis virus. *Phytopathology* 87: S104. [Abstract]
1359. Wisler, G.C., Li, R.H., Liu, H.Y., Duffus, J.E., Simone, G.W., Hochmuth, R.C. and Knight, J.R. 1996. Tomato chlorosis virus (ToCV) is a new closterovirus distinct from other whitefly-transmitted closteroviruses. *Phytopathology* 86(11 SUPPL.): S118. [Abstract]
1360. Wisler, G.C., Liu, H.Y., Li, R. and Duffus, J.E. 1996. Partial characterization and diagnosis of the lettuce chlorosis virus of lettuce. p. 46. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1361. Wolfe, G.R., Hendrix, D.L. and Salvucci, M.E. 1998. Sorbitol metabolism in *Bemisia argentifolii*: Cloning of the NADPH-dependent ketose reductase. p. 22. *In* Silverleaf Whitefly: National Research, Action, and Technology Transfer Plan, 1997-2001: First Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. 1998-01. [Abstract]
1362. Wolfenbarger, D.A. 1993. [chemical control, cotton]. p. 85. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1363. Wolfenbarger, D.A. 1994. Field and glass vial bioassays of insecticides against the sweetpotato whitefly. p. 115. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
1364. Wolfenbarger, D.A. 1995. Insecticide tests. p. 96. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1365. Wolfenbarger, D.A. 1996. Insecticides in laminated plastic. p. 100. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1366. Womac, A.R., Mulrooney, J.E. and Howard, K.D. 1993. [Degania, Berthoud, Proptec, electrostatic, Turbo-Thrush, Hardi, chemical control, cotton, peanuts]. p. 86. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1367. Wool, D. 1989. Genetic variation in Israeli populations of *Bemisia tabaci*. *Phytoparasitica* 17(3): 229. [Abstract]
1368. Wraight, S., Bradley, C., Jaronski, S., Galaini-Wraight, S., Garza, F., De La Garza, F., Munguia, O., De Anda, M., Rosales, A. and Becerra, M. 1997. Efficacy of Mycotrol (*Beauveria bassiana* strain GHA) against silverleaf whitefly on cucurbit field crops in the Lower Rio Grande Valley of Texas. p. 176. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
1369. Wraight, S., Carruthers, R., Bradley, C. and Jaronski, S. 1996. Efficacy of *Beauveria bassiana* against silverleaf whitefly on field crops in South Texas. p. 153. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1370. Wraight, S., Carruthers, R., Jaronski, S., Bradley, C., Galaini-Wraight, S., Underwood, N., Wood, P., Garza, J. and Britton, J. 1995. Efficacy of fungal pathogens against silverleaf whitefly on field crops in South Texas. p. 141. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1371. Wright, J.E., Bouse, L.F., Kirk, I.W., Carlton, J.B., Franz, E., Lathief, M.A. and Reitorik, R. 1993. [*Beauveria*, *Naturalis-L*, cotton]. p. 111. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1372. Yasui, Y., Ellsworth, P., Lublinkhof, J. and Comer, D. 1997. Monitoring *Bemisia argentifolii* susceptibility to buprofezin. p. 132. *In* Silverleaf Whitefly, 1997 Supplement to the 5-Year National Research and Action Plan: Progress Review, Technology Transfer, and New Research and Action Plan (1997-2001). U.S. Dept. Agric., Agric. Res. Serv. 1997-02. [Abstract]
1373. Yee, W., Toscano, N. and Palumbo, J. 1995. Silverleaf whitefly tritrophic interaction instable ecosystem. p. 156. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1374. Yee, W.L., Hendrix, D.L., Toscano, N.C., Chu, C.C. and Henneberry, T.J. 1995. Continuous honeydew production by silverleaf whitefly nymphs on cotton. p. 21. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]

1375. Yee, W.L., Toscano, N.C., Chu, C.C., Henneberry, T.J. and Nichols, R.L. 1996. Photosynthesis of cotton in relation to action thresholds for whitefly control. p. 12. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1996 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1996-01. [Abstract]
1376. Yilmaz, M.A., Ozaslan, M. and Ozaslan, D. 1989. Cucumber vein yellowing virus in Cucurbitaceae in Turkey. *Plant Dis.* 73(7): 610. [Abstract, Cock (1993)]
1377. Yokomi, R.K., Jimenez, D.R., Osborne, L.S., Shapiro, J.P. and Schroeder, W.J. 1994. Plant biochemical regulators (PBRs) to mediate interactions of the sweetpotato whitefly. p. 170. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
1378. Yokomi, R.K., Jimenez, D.R. and Shapiro, J.P. 1993. [esterase isozyme analysis, cucurbits, squash silverleaf]. p. 44. *In* Sweetpotato Whitefly: 1993 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 112. [Abstract]
1379. Yokomi, R.K., Jimenez, D.R., Shapiro, J.P., Duffus, J.E., Brown, J.K. and Bird, J. 1992. A new biotype of *Bemisia tabaci*: interactions with plants and virus epidemiology. p. 297. *In* Proc. XIX Int. Congress Entomol, Beijing, China, June 28-July 4, 1992. [Abstract]
1380. Yokomi, R.K. and Osborne, L.S. 1995. Mediation of whitefly feeding behavior. p. 157. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1995 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. 1995-2. [Abstract]
1381. Yoldas, Z. 1998. Effectiveness of *Encarsia formosa* (Hymenoptera, Aphelinidae) on whiteflies in greenhouses in Izmir (Turkey). p. 567. *In* Proceedings 6th European Congress of Entomology, Ceske Budejovice, Czech Republic, 23-29 Aug 1998. [Abstract]
1382. Yoshida, H.A., Blua, M.J. and Toscano, N.C. 1994. Oviposition preference and nymphal performance of two *Bemisia* species. p. 65. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]
1383. Yuki, V.A. and Costa, A.S. 1976. Comparacao entre tipos de armadilhas, colocadas a diferentes alturas, para a coleta da mosca branca *Bemisia tabaci*, em campo. [Comparison among traps, placed at different heights, to collect white flies under field conditions]. *Fitopatologia* 11(1/2): 34. [Portuguese, Abstract]
1384. Zachrisson, B. 1992. Manejo integrado de la mosca blanca. [Integrated management of the white fly]. pp. 26-27. *In* Jornada Agropecuaria, Los Santos, Panama. 16-18 Nov 1992. Instituto De Investigacion Agropecuaria De Panama, Divisa, Panama. [Spanish, Abstract]
1385. Zahra, S.O. 1998. Screening watermelon for resistance against watermelon chlorotic stunt virus in the area of Gezira, Sudan. p. 50. *In* A.M. Tronsmo and A.A. Ergon (Eds.), 16th Nordic Postgraduate Course in Plant Pathology, Resistance to Plant Pathogens, 18-25 April 1998, Honne Konferansesenter, Biri, Norway. Nordic Veterinary and Agricultural University, Nova. [Abstract]
1386. Zarokosta, M. and Copland, M.J.W. 1998. Effect of leaf age and source food plant on the life table parameters of 'B' strain of *B. tabaci* (Gennadius) (Homoptera, Aleyrodidae). pp. 484-485. *In* Proceedings 6th European Congress of Entomology, Ceske Budejovice, Czech Republic, 23-29 Aug 1998. [Abstract]
1387. Zarrabi, M. 1995. Aleyrodid species on citrus plants of Iran. p. 217. *In* Proceedings 12th Iranian Plant Protection Congress, 2-7-September 1995, Karadj, Iran. [Persian, Abstract]
1388. Zeidan, M. and Czosnek, H. 1993. Transmission of the tomato yellow leaf curl virus by *Bemisia tabaci* fed on infected plants, viral DNA and *Agrobacterium*. *Phytoparasitica* 21(2): 175. [Abstract]
1389. Zhang, J., Oleson, A.E., Vacek, D.C., Ciomperlik, M.A., Gould, J.R., Heilmann, L.J. and Nelson, D.R. 2000. Whitefly biocontrol agents: differentiation of parasitoid wasps by satellite DNA-targeted hybridization. p. 103. *In* Silverleaf Whitefly, National Research, Action, and Technology Transfer Plan, 1997-2001: Third Annual Review of the Second 5-Year Plan. U.S. Dept. Agric., Agric. Res. Serv. July 2000. [Abstract]
1390. Zidan, Z.H., Zidan, A.A., Hussein, M.I., Abdel-Fattah, M.S. and Gabr, A.M. 1990. Knock-down and residual activities of certain insecticides against the cotton whitefly, *Bemisia tabaci* (Genn.) on tomato seedlings under laboratory conditions. p. 17. *In* 3rd National Conference of Pests and Diseases of Vegetables and Fruits in Egypt and Arab Countries, Ismaileyah, Egypt, 24-26 Oct 1989. Suez Canal University, Ismaileyah, Egypt. [Abstract]
1391. Zidan, Z.H., Zidan, A.A., Hussien, M.I., Abdel-Fattah, M.S. and Gabr, A.M. 1990. Performance of certain insecticide regimes on the population density of the cotton white fly, *Bemisia tabaci* (Genn.) in tomato field, with special references to phytotoxicity and virus infestation. p. 18. *In* 3rd National Conference of Pests and Diseases of Vegetables and Fruits in Egypt and Arab Countries, Ismaileyah, Egypt, 24-26 Oct 1989. Suez Canal University, Ismaileyah, Egypt. [Abstract]
1392. Ziegweid, K., Minkenber, O.P.J.M. and Hennessey, R.D. 1994. Pre-introduction selection of parasitoids for augmentative biological control of the sweetpotato whitefly on greenhouse tomatoes. p. 152. *In* Silverleaf Whitefly (Formerly Sweetpotato Whitefly, Strain B): 1994 Supplement to the 5-Year National Research and Action Plan. U.S. Dept. Agric., Agric. Res. Serv. No. 125. [Abstract]

## Appendix C: Meeting Agenda

### FIFTH ANNUAL PROGRESS REVIEW FIVE-YEAR SILVERLEAF WHITEFLY RESEARCH, ACTION & TECHNOLOGY TRANSFER PLAN Feb. 10-12, 2002

#### Sunday, February 10, 2002

10:00 a.m.	Registration (PACIFIC FOYER) Poster set-up begins (HARBORSIDE ROOM)	
1:00 p.m.	Welcome and Announcements (PACIFIC BALLROOM A)	<i>Walker Jones &amp; Thomas M. Perring</i>
1:15 p.m.	Charge to the Conference	<i>Robert Faust</i>
1:30 p.m.	Section A Paper Session	<i>Jackie Blackmer &amp; David Byrne</i>
3:35 p.m.	Section A Discussion	
4:05 p.m.	Break	
4:20 p.m.	Section F Paper Session	<i>Steve Castle &amp; Peter Ellsworth</i>
6:00 p.m.	Section F Discussion	
6:30 p.m.	Mixer and Poster session (HARBORSIDE ROOM)	

#### Monday, February 11, 2002

7:00 a.m.	Continental breakfast (PACIFIC BALLROOM A)	
8:00 a.m.	Section C Paper Session	<i>John Palumbo &amp; Shirley Taylor</i>
9:50 a.m.	Section C Discussion	
10:20 a.m.	Break	
10:30 a.m.	Section E Paper Session	<i>Cindy McKenzie &amp; Greg Walker</i>
11:45 a.m.	Section E Discussion	
12:15 p.m.	Lunch Break	
2:30 p.m.	Section D Paper Session	<i>Bill Roltsch &amp; Greg Simmons</i>
4:00 p.m.	Break	
4:15 p.m.	Section D Paper Session (cont.)	
4:50 p.m.	Section D Discussion	
5:35 p.m.	Depart for Boat Taxi to Coronado Island for Dinner (HOTEL LOBBY)	

National Whitefly Conference Agenda

Page 2

#### Tuesday, February 12, 2002

7:00 a.m.	Continental breakfast (PACIFIC BALLROOM A)	
8:00 a.m.	Section B Paper Session	<i>Judith K. Brown &amp; Robert L. Gilbertson</i>
10:15 a.m.	Section B Discussion	
10:45 a.m.	Break	
11:00 a.m.	Progress Review - Sections A, B, C	<i>Moderated by Section Co-Chairs</i>
12:30 p.m.	Lunch Break	
2:00 p.m.	Progress Review - Sections D, E, F	<i>Moderated by Section Co-Chairs</i>
3:30 p.m.	General Discussion	

Immediately following the General Discussion, the Working Group will meet. The Program Planning and Review Committee will meet immediately following the Working Group.

## Section A: Biology, Ecology, and Population Dynamics

Co-Chairs: Jackie Blackmer and David Byrne

Sunday Afternoon 1:30-4:20

- 1:30 Keynote Address. Dale B. Gelman, Michael B. Blackburn, Jing S. Hu, and D. Gerling. Whitefly metamorphosis - timing, regulation, and influences on the development of its parasitoid, *Encarsia formosa*.
- 2:00 Jackie Blackmer and D. Cross. Response of *Eretmocerus eremicus* to skylight and plant cues in a vertical flight chamber.
- 2:15 Luis A. Canas, Steven E. Naranjo, Peter C. Ellsworth. Seasonal ecology of *Bemisia tabaci* in Arizona: Low temperature and host plant effects on field populations and associated mortality factors.
- 2:30 Thomas P. Freeman, James S. Buckner, and Dennis R. Nelson. What I know about silverleaf whitefly nymph feeding and what I would like to know.
- 2:45 Dan Gerling and Dale B. Gelman. Physiological considerations in the penetration of *Eretmocerus mundus* into *Bemisia* nymphs.
- 3:00 M.S. Palaniswami. Isozyme, vector capability and cross breeding studies to confirm biotypes of *B. tabaci* in India.
- 3:15 Historical Presentation. David N. Byrne. Recounting whitefly biology research since 1992.
- 3:35 - 4:05 Discussion
- 4:05 - 4:20 Break

### **Posters:**

- Thomas P. Freeman, James S. Buckner, and Dennis R. Nelson. Silverleaf whitefly nymph stylets and feeding characteristics.
- M. S. Palaniswami. Life table studies of *Bemisia tabaci* Genn. on cassava and laboratory evaluation of potentiality of predators.
- Dennis R. Nelson. Comparison of the feeding process of nymph and adult whiteflies.
- Bahram Tafaghodinia. Engineering-based computer simulation for modeling greenhouse whitefly population growth.

## Section F: Integrated and Areawide Pest Management Approaches, and Crop Management Systems

Co Chairs: Steve Castle and Peter Ellsworth

Sunday Afternoon 4:20-6:30

- 4:20 C.C. Chu, C.G. Jackson, P.J. Alexander, and T.J. Henneberry. A light-emitting diode equipped CC trap for greenhouse insects.
- 4:35 Robert T. McMillan, Jr. Tomato yellow leafcurl gemini virus management in south Florida.
- 4:50 Keynote Address. Steve Castle. Outbreak occurrences: Factors that contribute and tactics that suppress.
- 5:20 Historical Presentation. Tom Henneberry. Assembling a multi-agency response to *Bemisia* outbreaks: a retrospective.
- 5:40 Historical Presentation. Nick Toscano. Achievements and management successes in the whitefly wars.
- 6:00 - 6:30 Discussion

### **Posters:**

- Jennifer Jones, Peter Ellsworth, John Palumbo, Kai Umeda, and Pat Clay. Cross commodity research and outreach program.
- C.C. Chu, A.M. Simmons, P.J. Alexander, and T.J. Henneberry. A light-emitting diode equipped yellow card sticky trap for greenhouse insects.

### **Section C: Chemical Control, Biopesticides, Resistance Management, and Application Methods**

**Co Chairs: John Palumbo and Shirley Taylor**

**Monday Morning 8:00 - 10:20**

- 8:00 D.H. Akey and T.J. Henneberry. Use of sucrose esters as biorational agents to control silverleaf whitefly in Arizona upland cotton.
- 8:15 D.J. Schuster, L.D. Ortega, S. Thompson, and J.E. Polston. Laboratory comparison of potential repellents for silverleaf whitefly adults.
- 8:30 Eric Natwick. Comparison of four neonicotinoid insecticides for whitefly control.
- 8:45 N.C. Toscano, N. Prabhaker, S. Castle, T. Henneberry. Inter-regional differences in baseline toxicity of *Bemisia argentifolii* to buprofezin and pyriproxyfen.
- 9:00 Keynote Address. Nilima Prabhaker. Insecticide resistance in *Bemisia tabaci*: Past expectations and present reality
- 9:30 Historical Presentation. John C. Palumbo. Chemical control of *Bemisia* whiteflies: Summary of research accomplishments and technology transfer.
- 9:50 - 10:20 Discussion
- 10:20 - 10:30 Break

#### **Posters:**

- D.J. Schuster, and S. Thompson. Comparing the susceptibility of the silverleaf whitefly to imidacloprid on tomato in Florida.
- Cindy McKenzie. Efficacy of sucrose octanoate to whitefly using a plant-based bioassay.
- T.J. Henneberry, L. Forlow Jech, D.L. Hendrix, T. de la Torre, and J. Maurer. Effects of Applaud and Knack on sweetpotato whitefly nymph mortality and adult and nymph honeydew production in the laboratory.

### **Section E: Host Plant Resistance, Physiological Disorders, and Host-Plant Interactions**

**Co Chairs: Cindy McKenzie and Greg Walker**

**Monday Morning 10:30 - 12:15pm**

- 10:30 Keynote Address. Gregory P. Walker. Overview of whitefly feeding behavior: What we know and what we need to know.
- 11:00 Yun-Shu Chen, Wilhelmina T.G. van de Ven and Linda L. Walling. Expression of *SLW3* (silverleaf whitefly-induced) gene in the transgenic plants.
- 11:15 Laura Petro, Rick Redak,, Jim Bethke and Thomas M. Perring. Preference and performance of silverleaf whitefly on selected poinsettia cultivars.
- 11:30 Alvin M. Simmons. Movement of whitefly crawlers on several types of vegetable plants.
- 11:45-12:15 Discussion

#### **Posters:**

- Eric Natwick. Cotton host plant resistance to whitefly transmitted cotton leaf crumple disease.
- Gregory P. Walker and Eric Natwick. High level of resistance to silverleaf whitefly in the cotton relative, *Gossypium thurberi*



**Section D: Natural Enemy Ecology and Biological Control**  
**Co Chairs: Bill Roltsch and Greg Simmons**  
**Monday Afternoon 2:30-5:20**

- 2:30 Keynote Address. Don C. Vacek. Application of genetic diagnostics to biological control of silverleaf whitefly.
- 3:00 Steven E. Naranjo. Intraguild predation on *Bemisia tabaci* parasitoids by three generalist predators.
- 3:15 Jesusa C. Legaspi. Parasitism of the silverleaf whitefly in north Florida.
- 3:30 William Roltsch and Earl Andress. Silverleaf whitefly biological control in Imperial Valley, CA.
- 3:45 Gregory S. Simmons, Kim Hoelmer, Charles Pickett, Eric Natwick, and Earl Andress. Augmentative biological control of silverleaf whitefly infesting melons in California with releases of exotic *Eretmocerus* species, a review of the progress.
- 4:00 - 4:15 Break
- 4:15 Charles H. Pickett. Releases of exotic parasites in central California.
- 4:30 Historical Presentation. William Roltsch. The value of a collective effort in the biological control of silverleaf whitefly.
- 4:50 - 5:20 Discussion

**Posters:**

- Earl Andress. Exotic parasitoids in four crops in the Imperial Valley.
- Larry J. Heilmann. DNA based species identification test for parasitic wasps of the whitefly.
- James Buckner. Marking pheromones for female *Eretmocerus mundus*, a parasitoid of *Bemisia argentifolii*.

**Section B: Viruses, Epidemiology, & Virus-Vector Interactions**  
**Co Chairs: Judith K. Brown and Robert L. Gilbertson**  
**Tuesday Morning 8:00-10:50**

- 8:00 Keynote Address. Judith K. Brown: A twenty year legacy of Begomoviruses and the whitefly vector in the U.S. sunbelt states: 1981-2001.
- 8:25 Keynote Address. Robert L. Gilbertson: Tomato yellow leaf curl virus in the Dominican Republic - A case study.
- 8:55 William M. Wintermantel and Arturo A. Cortez. Complementation for transmission by non-vector whiteflies among tomato-infecting criniviruses.
- 9:10 Thomas M. Perring, and Charles A. Farrar. Whitefly -borne virus epidemiology: Lessons learned from the greenhouse whitefly and tomato infectious chlorosis virus.
- 9:25 Jane E. Polston Lack of transovarial transmission of begomoviruses by *Bemisia tabaci* biotype B.
- 9:45 Judith K. Brown and A.M. Idris. Cloning and sequencing of *Cotton leaf crumple virus*—a begomovirus infecting cotton in the Sonoran Desert.
- 10:00 Y.P.S. Rathi. Mungbean yellow mosaic virus-epidemiology and management.
- 10:15 - 10:45 Discussion

**Posters:**

- Judith K. Brown, A.M. Idris, and J. Bird. Cloning and molecular characterization of the A and B components for *Macrotium mosaic virus* from Puerto Rico.

## Appendix D: List of Registered Meeting Participants

David Akey  
Research Entomologist  
USDA-ARS-WCRL  
4135 E. Broadway Road  
Phoenix, AZ 85040-8803  
Ph: 602-437-0121 x 245  
Fax: 602-437-1274  
[dakey@wrcr.ars.usda.gov](mailto:dakey@wrcr.ars.usda.gov)

Pamela Anderson  
CIAT  
1380 NW 78th Ave.  
Miami, FL 33126  
Ph: 650-833-6625  
Fax: 650-833-6626  
[p.anderson@cgiar.org](mailto:p.anderson@cgiar.org)

Earl Andress  
Entomologist  
USDA-APHIS  
4151 Hwy 86, Bldg. 10  
Brawley, CA 92227  
Ph: 760-344-7857  
Fax: 760-351-0532  
[eandress@quix.net](mailto:eandress@quix.net)

Stephen Birdsall  
Imperial County Agricultural Commissioner  
Imperial County Ag. Commission  
150 S. 9th Street  
El Centro, CA 92243  
Ph: 760-482-4314  
Fax: 760-353-9420  
[agcomm@imperialcounty.net](mailto:agcomm@imperialcounty.net)

Jackie Blackmer  
Research Entomologist  
WCRL-USDA  
4135 E. Broadway Road  
Phoenix, AZ 85040-8803  
Ph: 602-437-0121 x 246  
Fax: 602-437-1274  
[jblackmer@wrcr.ars.usda.gov](mailto:jblackmer@wrcr.ars.usda.gov)

Judy Brown  
Associate Professor  
Department of Plant Sciences  
Forbes Bld. Room 303  
The University of Arizona  
Tucson, AZ 85721 USA  
Ph: 520-621-1402  
Fax: 520-621-8839  
[jbrown@ag.arizona.edu](mailto:jbrown@ag.arizona.edu)

James Buckner  
Research Biochemist  
USDA-ARS-Bioscience Research Lab  
1605 Albrecht Boulevard  
Fargo, ND 58105  
Ph: 701-239-1280  
Fax: 701-239-1348  
[bucknerj@fargo.ars.usda.gov](mailto:bucknerj@fargo.ars.usda.gov)

John Byers  
USDA-ARS-WCRL  
4135 E. Broadway Rd.  
Phoenix, AZ 85040  
Ph: 602-437-0121  
Fax: 602-437-1274  
[jbyers@wrcr.ars.usda.gov](mailto:jbyers@wrcr.ars.usda.gov)

David Byrne  
The University of Arizona  
Department of Entomology  
Tucson, AZ 85721  
Ph: 520-621-7169  
Fax: 520-615-4855  
[byrne@ag.arizona.edu](mailto:byrne@ag.arizona.edu)

Luis Canas  
Research Associate  
University of Arizona  
37860 W. Smith-Enke Rd.  
Maricopa, AZ 85239  
Ph: 520-568-2273 x 251  
Fax: 520-568-2556  
[lcanas@ag.arizona.edu](mailto:lcanas@ag.arizona.edu)

Steve Castle  
Research Entomologist  
USDA-ARS-WCRL  
4135 E. Broadway Rd.  
Phoenix, AZ 85040-8803  
Ph: 602-437-0121 x 238  
Fax: 602-437-1274  
[scastle@wrcr.ars.usda.gov](mailto:scastle@wrcr.ars.usda.gov)

Yun-Shu Chen  
Graduate Student  
Department of Botany & Plant Sciences  
University of California, Riverside  
Riverside, CA 92521  
Ph: 909-787-4687  
Fax: 909-787-4437  
[yunsch@hotmai.com](mailto:yunsch@hotmai.com)

C.C. Chu  
Plant Physiologist  
USDA-ARS - WCRL  
4135 E. Broadway Road  
Phoenix, AZ 85040-8803  
Ph: 602-437-0121 x240  
Fax: 602-437-1274  
[cchu@wcrf.ars.usda.gov](mailto:cchu@wcrf.ars.usda.gov)

Amy Corley  
University of California  
107 Entomology Annex 1  
Riverside, CA 92521  
Ph: 909-787-4562  
Fax: 909-787-3681

Jolene Dessert  
Imperial County Entomologist  
Imperial County Ag. Commission  
150 S. 9th Street  
El Centro, CA 92243  
Ph: 760-482-4649  
Fax: 760-482-4124  
[jolenedessert@imperialcounty.net](mailto:jolenedessert@imperialcounty.net)

Jeff Dobbs  
Olympic Horticultural Products  
1045 Applecross Drive  
Roswell, GA 30075  
Ph: 770-992-0121  
Fax: 770-992-5564  
[dobbs-jh@bellsouth.net](mailto:dobbs-jh@bellsouth.net)

Peter Ellsworth  
University of Arizona  
37860 W. Smith-Enke Rd.  
Maricopa, AZ 85239  
Ph: 520-568-2273  
Fax: 520-568-2556  
[peterell@ag.arizona.edu](mailto:peterell@ag.arizona.edu)

Thomas Freeman  
Electron Microscopy Center, NCSL  
North Dakota State University  
Fargo, ND 58105  
Ph: 701-231-8234  
Fax: 701-239-1395  
[thomas.freeman@ndsu.nodak.edu](mailto:thomas.freeman@ndsu.nodak.edu)

Dale Gelman  
Research Entomologist  
Insect Biocontrol Lab, USDA ARS  
Bldg 011A, Rm. 214 BARC West  
Beltsville, MD 20705  
Ph: 301-504-8909  
Fax: 301-504-5104  
[gelmand@ba.ars.usda.gov](mailto:gelmand@ba.ars.usda.gov)

Dan Gerling  
Department of Zoology  
Tel Aviv University  
Ramat Aviv 69978  
ISRAEL  
Ph: 972-3-640-8611  
Fax: 972-3-640-9403  
[dang@ccsg.tau.ac.il](mailto:dang@ccsg.tau.ac.il)

Robert L. Gilbertson  
University of California  
1 Shields Avenue  
Davis, CA 95616  
Ph: (530) 752-3163  
Fax: (530) 752-5674  
[rlgilbertson@ucdavis.edu](mailto:rlgilbertson@ucdavis.edu)

Celso Goseco  
Director, Pesticide Control  
Del Monte Fresh Produce  
P.O. Box 149222  
Coral Gables, FL 33114-9222  
Ph: 305-520-8101  
Fax: 305-445-7612  
[cgooseco@freshdelmonte.com](mailto:cgooseco@freshdelmonte.com)

Larry Heilman  
Research Associate  
North Dakota State University, Dept. of Biochemistry  
Loftsgard Hall  
Fargo, ND 58105  
Ph: 701-231-9683  
Fax: 701-231-8324  
[larry.heilmann@ndsu.nodak.edu](mailto:larry.heilmann@ndsu.nodak.edu)

T.J. Henneberry  
Laboratory Director  
USDA-ARS, WCRL  
4135 E. Broadway Road  
Phoenix, AZ 85040-8803  
Ph: 602-437-0121 x236  
Fax: 602-437-1274  
[thenneberry@wcrf.ars.usda.gov](mailto:thenneberry@wcrf.ars.usda.gov)

Charles Jackson  
Research Entomologist  
USDA-ARS, WCRL  
4135 E. Broadway Road  
Phoenix, AZ 85040-8803  
Ph: 602-437-0121 x 239  
Fax: 602-437-1274  
[gjackson#wcrf.ars.usda.gov](mailto:gjackson#wcrf.ars.usda.gov)

Lynn Jech  
Biological Tech  
USDA-ARS - WCRL  
4135 E. Broadway Road  
Phoenix, AZ 85040-8803  
Ph: 602-437-0121  
Fax: 602-437-1274  
[ljech@wcr1.ars.usda.gov](mailto:ljech@wcr1.ars.usda.gov)

Walker Jones  
Supervisory Research Entomologist  
Beneficial Insects Research Unit, ARS - USDA  
2413 E. Hwy 83  
Weslaco, TX 78596  
Ph: 956-969-4852  
Fax: 956-969-4888  
[wjones@weslaco.ars.usda.gov](mailto:wjones@weslaco.ars.usda.gov)

Jennifer Jones  
University of Arizona  
37860 W. Smith-Enke Rd.  
Maricopa, AZ 85239  
Ph: 520-568-2273  
Fax: 520-568-2556  
[jjones@ag.arizona.edu](mailto:jjones@ag.arizona.edu)

Marla Lawrence  
Secretary  
USDA-ARS  
4135 E. Broadway Road  
Phoenix, AZ 85040-8803  
Ph: 602-437-0121 x236  
Fax: 602-437-1274  
[mlawrence@wcr1.ars.usda.gov](mailto:mlawrence@wcr1.ars.usda.gov)

Jesusa Legaspi  
Research Entomologist  
USDA-ARS  
310 Perry-Paige Bldg., South  
Tallahassee, FL 32307  
Ph: 850-412-7061  
Fax: 850-412-7263  
[jlegaspi@gainesville.usda.ufl.edu](mailto:jlegaspi@gainesville.usda.ufl.edu)

Richard Lindquist  
Senior Technical Manager  
Olympic Horticultural Products  
1594 Sunset Lane  
Wooster, OH 44691-1824  
Ph: 330-345-5570  
Fax: 330-345-1311  
[hindquist@olympichort.com](mailto:hindquist@olympichort.com)

Tongxian Liu  
Associate Professor  
Texas Ag. Exp. Station, Texas A&M  
2415 E. Highway 83  
Weslaco, TX 78596-8399  
Ph: 956-968-5585  
Fax: 956-968-0641  
[tx-liu@tamu.edu](mailto:tx-liu@tamu.edu)

Cindy McKenzie  
Research Entomologist  
USDA, ARS, US Horticultural Research Laboratory  
2001 South Rock Road  
Ford Pierce, FL 34945  
Ph: 561-462-5917  
Fax: 561-462-5986  
[cmckenzie@ushrl.ars.usda.gov](mailto:cmckenzie@ushrl.ars.usda.gov)

Robert McMillan  
Professor of Plant Pathology  
University of Florida, IFAS, TREC  
18905 SW 280 Street  
Homestead, FL 33031  
Ph: 305-246-7001 ext. 270  
Fax: 305-246-7003  
[rtmcm@mail.ifas.ufl.edu](mailto:rtmcm@mail.ifas.ufl.edu)

Steve Naranjo  
Research Entomologist  
USDA-ARS  
4135 E. Broadway Road  
Phoenix, AZ 85040-8803  
Ph: 602-437-0121  
Fax: 602-437-1274  
[snaranjo@wcr1.ars.usda.gov](mailto:snaranjo@wcr1.ars.usda.gov)

Eric Natwick  
Entomology Advisor  
UC Cooperative Extension  
1050 E. Holton Road  
Holtville, CA 92250  
Ph: 760-352-9474  
Fax: 760-352-0846  
[etnatwick@ucdavis.edu](mailto:etnatwick@ucdavis.edu)

Dennis Nelson  
Research Leader  
USDA-ARS  
1605 Albrecht Boulevard  
Fargo, ND 58105  
Ph: 701-239-1286  
Fax: 701-239-1202  
[nelsond@fargo.ars.usda.gov](mailto:nelsond@fargo.ars.usda.gov)

M.S. Palaniswami  
Head, Division of Crop Protection  
Division of Crop Protection  
Central Tuber Crops Research Institute  
Trivandrum 695 017, INDIA  
Ph: 91-0471-598551  
Fax: 91-471-590063  
[raja@ctcri.ren.nic.in](mailto:raja@ctcri.ren.nic.in)

John M. Palumbo  
University of Arizona  
Dept. of Entomology  
Yuma Agricultural Center  
6425 W. 8th Street  
Yuma, AZ 85264  
Ph: 520-782-3836  
Fax: 520-782-1940  
[jpalumbo@ag.arizona.edu](mailto:jpalumbo@ag.arizona.edu)

Thomas Perring  
University of California  
107 Entomology Annex 1  
Riverside, CA 92521  
Ph: 909-787-4562  
Fax: 909-787-3681  
[thomas.perring@ucr.edu](mailto:thomas.perring@ucr.edu)

Charles Pickett  
CDFA-Biol. Control Prog.  
3288 Meadowview Road  
Sacramento, CA 95832  
Ph: 916-262-2053  
Fax: 916-262-2059  
[cpickett@cdfa.ca.gov](mailto:cpickett@cdfa.ca.gov)

Jane Polston  
University of Florida  
GCREC, 5007 60th St. E  
Bradenton, FL 34203  
[jep@mail.ifas.ufl.edu](mailto:jep@mail.ifas.ufl.edu)

Nilima Prabhaker  
University of California  
4135 E. Broadway Rd.  
Phoenix, AZ 85045  
Ph: 602-437-0121 x 248  
Fax: 602-437-1274  
[castle@ucr1.ucr.edu](mailto:castle@ucr1.ucr.edu)

Jesse Richardson  
9330 10<sup>th</sup> Ave  
Hesperia, CA 92345-3443  
Ph: 760-949-2565

William Roltsch  
California Dept. of Food & Agric.,  
Biological Control Program  
3288 Meadowview Rd.  
Sacramento, CA 95832  
Ph: 916-262-2055  
Fax: 916 262-2059  
[wroltsch@cdfa.ca.gov](mailto:wroltsch@cdfa.ca.gov)

Alan Rudy  
Assistant Professor of Sociology  
Michigan State University  
Department of Sociology, 316 Berkey Hall  
East Lansing, MI 48824  
Ph: 517-353-0745  
Fax: 517-432-2856  
[alan.rudy@ssc.msu.edu](mailto:alan.rudy@ssc.msu.edu)

David Schuster  
University of Florida, IFAS  
Gulf Coast Research and Education Center  
5007 60th St. East  
Bradenton, FL 34203  
Ph: 941-751-7636  
[dschuster@mail.ifas.ufl.edu](mailto:dschuster@mail.ifas.ufl.edu)

Alvin Simmons  
Research Entomologist  
USDA-ARS, U.S. Vegetable Laboratory  
2875 Savannah Highway  
Charleston, SC 29414  
Ph: 843-556-0840  
Fax: 843-763-7013  
[asimmons@awod.com](mailto:asimmons@awod.com)

Greg Simmons  
USDA-APHIS PPQ  
Oswell St. Biological Control Facility  
Bakersfield, CA  
[Gregory.S.Simmons@aphis.usda.gov](mailto:Gregory.S.Simmons@aphis.usda.gov)

Robert Steffens  
Bayer Corporation  
8400 Hawthorne Rd.  
Kansas City, MO 84720  
Ph: 816-242-2261  
Fax: 816-242-2753

Guido Sterk  
Field and Development Manager  
Biobest N.V.  
Ilse Velden 18  
2260 Westerlo, BELGIUM  
32-14-257980  
32-14-257982  
[guido.sterk@biobest.be](mailto:guido.sterk@biobest.be)

Shirley Taylor  
Bayer Corporation  
5020-D Baltimore Dr., #362  
La Mesa, CA 91941  
Ph: 619-466-6294  
E-mail: [s.taylor.b@bayer.com](mailto:s.taylor.b@bayer.com)

Nick Toscano  
University of California  
Entomology Dept.  
Riverside, CA 92521  
Ph: 909-787-6373  
Fax: 909-787-3086  
[nick.toscano@ucr.edu](mailto:nick.toscano@ucr.edu)

Don Vacek  
USDA-APHIS-PPQ  
Mission Plant Protection Center  
P.O. Box 2140  
Mission, TX 78573-2140  
Ph: 956-580-7301  
Fax: 956-580-7300  
[Don.C.Vacek@usda.gov](mailto:Don.C.Vacek@usda.gov)

Greg Walker  
University of California  
Entomology Dept.  
Riverside, CA 92521  
Ph: 909-787-5808  
Fax: 909-787-3086  
[walker@citrus.ucr.edu](mailto:walker@citrus.ucr.edu)

Linda Walling  
Professor of Genetics  
Department of Botany & Plant Sciences  
University of California, Riverside  
Riverside, CA 92521  
Ph: 909-787-4687  
Fax: 909-787-4437  
[lwalling@citrus.ucr.edu](mailto:lwalling@citrus.ucr.edu)

Ian Wedderspoon  
President  
Ian Industries Inc.  
13015 SW 89 Place, #203  
Kendall, FL 33176  
Ph: 305-255-6706  
Fax: 305-255-1317  
[Lairdian@worldnet.att.net](mailto:Lairdian@worldnet.att.net)

William Wintermantel  
Research Plant Pathologist  
USDA-ARS  
1636 E. Alisal St.  
Salinas, CA 93905  
Ph: 831-755-2824  
Fax: 831-755-2814  
[wwintermantel@pw.ars.usda.gov](mailto:wwintermantel@pw.ars.usda.gov)

Sonia Zarate  
Graduate Student  
Department of Botany & Plant Sciences  
University of California, Riverside  
Riverside, CA 92521  
Ph: 909-787-4687  
Fax: 909-787-4437  
[sonia@mail.ucr.edu](mailto:sonia@mail.ucr.edu)

## Appendix E. Minutes of the PPRC

### Minutes of the PPRC

Minutes  
PPRC Meeting  
Holiday Inn on the Bay  
2/10/02  
11:30 a.m.

Meeting was chaired by Tom Perring. PPRC members in attendance were:

Jacke Blackmer	David Byrne	Steve Castle
Thomas Henneberry	Walker Jones	Marla Lawrence
Cindy McKenzie	Tom Perring	Shirley Taylor

Tom Perring discussed the general overview of the program and reminded Section Co-chairs of their assignments during the meeting which included: 1) keeping the scheduled talks on time, 2) leading discussions, and 3) updating year 3 tables during the progress review.

He reminded Section Co-chairs that after the meeting they must submit section summaries with technology transfer and completed year 3 tables to Marla Lawrence.

Marla Lawrence will send co-chairs all copies of abstracts by February 26<sup>th</sup>.

Deadlines for abstracts submission or changes will be February 25<sup>th</sup> and Section Chair information is due to Marla Lawrence by March 10<sup>th</sup>.

**Five-Year National Research and Action Plan Priority Tables, Research Needs, and Yearly Goals (1997-2001)**

**Table A. Biology, Ecology, and Population Dynamics**

Approaches/Goals	Year 1	Year 2	Year 3	Year 4	Year 5
<b>Determine life cycle vulnerabilities (life tables)<sup>a</sup>, population development and natural mortality factors, natural enemies on major crops, urban plantings, weeds and predict overwintering potential.</b>	Whitefly and natural enemy sampling in cultivated crops, urban planting and weed hosts.	Determine potential of intercrop weed host & urban planting, movement of whiteflies and natural enemies.	Identify potential low population manipulation on vital host links for survival.	Initiate studies to manipulate host sequences to determine potential influence on whitefly population.	Continue 4 and finalize analysis of the potential of habitat modification as a management tool.
<b>Develop sampling methodology, action and economic thresholds for all major crops. Sampling methods and thresholds modified in light of natural enemy levels and existing management strategies.</b>	Initiate whitefly to identify spatial and temporal distributions in major cultivated crops.	Analysis and identification of needed additional sampling research to develop appropriate sampling protocol.	Validate and refine sampling methods.	Implement sampling protocols through cooperative extension outlets and other technology transfer methods.	Finalization, implementation and use in IPM systems.
<b>Develop population models to describe and predict whitefly population growth and spatial and temporal distribution. Develop simple day-degree sub-models for estimating phenology and temporal patterns of whitefly, natural enemies and host crops.</b>	Summarize whitefly biology, ecology and plant phenology to identify whitefly host plant interfaces.	Begin model development to include all biological and plant phenology data in simulation development.	Provide model simulation of whitefly populations and multiple cropping systems.	Identify weak points and needed information to improve model simulations.	Validate and expand effort to provide predictive models capabilities for whitefly population development and crop interfaces.
<b>Develop sampling methods for quality of cotton lint, vegetables and other commodities.</b>	Initiate sampling of seed cotton in the field during the season, at harvest, after picking, moduling and ginning.	Based on 1, expand and repeat sampling protocols as described.	Develop sampling protocol for field and harvest and processing sampling and determine interrelationships.	Extend sampling protocols to textile mill and verify field findings in relation to mill problems.	Modify, refine and complete sticky cotton sampling protocols from the field to the mill.
<b>Quantify whitefly and natural enemy dispersals and contribution to population dynamics.</b>	Review and analyze existing knowledge of whitefly dispersal.	Validate times of whitefly dispersal, environmental factors and identify modifying factors.	Determine proportion of whitefly population that are migratory and their reproductive potential.	Quantify the role of dispersal in population dynamics on different crop systems.	Formulate theory for manipulating and/or using dispersal as a tool in IPM.
<b>Define mating behavior, reproductive isolation, species, biotypes.</b>	Initiate studies on mating, oviposition and other behavior.	Define interspecies interbiotype mating interactions.	Define factors involved in mating, cues, feedback mechanisms, etc.	Develop potential methods of utilizing behavioral information in management strategies.	Review, summarize and propose additional needed research.



**Table A. Biology, Ecology, and Population Dynamics**

Approaches/Goals	Year 1	Year 2	Year 3	Year 4	Year 5
<b>Validate <i>Bemisia</i> taxa morphology, genetic, biochemical, and biology characteristics.</b>	Continue examination of <i>Bemisia</i> sp. for distinct morphological character differences.	Develop genetic molecular level and acceptable species level separation.	Discuss results, plan additional research, arrive at a consensus decision.	Publish verification of new species or other appropriate taxa.	-----
<b>Define role of endosymbionts in metabolism, host adaptation, nutrition and survival.</b>	Identify endosymbionts in whitefly.	Determine role of endosymbionts in whitefly biological functioning.	Determine potential for manipulating, interfering with or inhibiting endosymbiont function.	Determine associated enzymes and/or other endosymbionts and whitefly relationships.	Summarize and implement findings with suggestion for additional research.
<b>Characterize nutrient uptake and metabolism</b>	Identify the major carbohydrates, amino acids and other nutrients essential for whitefly growth and development.	Determine the biochemical pathways for metabolism of compounds essential for whitefly development.	Determine the physical and biochemical processes involved in uptake of carbohydrates, amino acids and other essential nutrients.	Determine the potential for blocking key steps in nutrient uptake and/or metabolism.	Implement findings by developing inhibitors of nutrient uptake and/or metabolism.
<b>Develop whitefly artificial diets and natural enemy mass-rearing.</b>	Identify whitefly nutritional components in plant tissue.	Develop whitefly artificial feeding systems.	Conduct addition, deletion studies to identify essential nutritional needs.	Evaluate developed diets on whitefly fecundity/longevity biology, behavioral characteristics.	Develop whitefly rearing system and adapt for production of natural enemies.
<b>**Pursue specific genetic and biological basis for variability in whitefly biotypes, strains, and species; determine impact of different genotypes/phenotypes on whitefly-mediated transmission and on the epidemiology of virus diseases.</b>	Identify differences in species, strains and biotypes with respect to transmission, host range, mating compatibilities, molecular variability, and map the biogeographic distribution of distinct types within the <i>B. tabaci</i> species complex.	Continue to study differences in species/ strains/biotypes with respect to transmission, host range, mating compatibilities, molecular variability. Determine molecular basis of observed variability in biological, molecular, & genetic terms. Infer molecular phylogenies from molecular markers.	Continue with work from previous years. Study impact of biotypes, strains, and species differences in the disease spread, crop damage, and specific control measures to reduce whitefly vector populations. Linkages with biological and chemical control sections.	Identify potential factors related to specific genetic and biological variability that may be manipulated to reduce disease spread. Develop molecular approaches to track biotypes, strains, and species relative to disease spread, based on differential molecular markers.	Summarize results, identify new research needs and make recommendations for implementation or expansion of research.

<sup>a</sup> Natural enemy research complements from Section D, see Table D.

<sup>b</sup> Action and economic thresholds also apply in Section C, see Table C.

<sup>c</sup> Sampling technology applicable to all other sections, see Tables B to F.

\*\* Revised 3/31/2000

**Table B. Viruses, Epidemiology, and Virus-Vector Interactions**

Approaches/Goals	Year 1	Year 2	Year 3	Year 4	Year 5
<b>Identification and characterization of new or emerging whitefly-transmitted viruses and strains</b>	Monitor crops for presence of whitefly-transmitted diseases, and determine relative disease incidence. Begin virus identification and strain differentiation.	Virus identification and characterization. Develop methods for identifying causal agents and for tracking viruses and strains using molecular methods.	Continue etiological studies and virus characterization. Apply molecular diagnostics to virus identification and evaluation of disease incidence and virus distribution.	Continue etiological studies and virus characterization efforts. Apply molecular diagnostics to virus identification and evaluation of disease incidence and virus distribution.	Summarize and review results. Determine areas of new research.
<b>Molecular epidemiology: identification of economic viruses, host plants, and reservoirs, and determination of geographic distribution of viruses.</b>	Monitor and identify host plants, virus reservoirs in affected areas. Linkages to diagnostic methods for virus ID and tracking.	Continue field studies. Determine economic input of diseases on crop production and associated losses.	Establish geographic distribution of viruses and identify sources of inoculum. Assess role of alternative host virus reservoirs on spread of diseases.	Identify and characterize virus involvement in disease establishment and spread. Assess potential methods of reducing virus reservoirs as a method of reducing disease.	Review and make recommendations for further research and potential implementation of results.
<b>Virus-vector interactions, factors affecting virus transmission, and basis for virus-vector specificity; determination of endosymbiont involvement in whitefly-mediated transmission</b>	Initiate studies on virus-vector interactions and on basis for the specificity of whitefly-mediated geminivirus transmission.	Determine specific cellular and molecular factors involved in virus transmission. Study role of endosymbionts in virus acquisition and transmission.	Continue studies in progress to determine specific factors involved in virus transmission, and the role of endosymbionts in virus acquisition and transmission.	Continue virus-vector interactions studies toward the development of approaches for disease control.	Summarize findings and suggest new research needs; implementation of existing knowledge.
<b>Strategies to reduce virus spread by management of cropping systems, reduced transmission frequencies, and other potentially effective approaches.</b>	Develop approaches to managing cropping systems to reduce vector densities to decrease transmission frequency and inoculum sources, taking into account weed and crop reservoirs in disease incidence and distribution.	Continue studies of management approaches for disease abatement. Interdisciplinary studies in conjunction with whitefly control methods in Sections B and C.	Continue studies of management approaches for disease abatement. Focus on interdisciplinary studies in conjunction with whitefly control methods in Sections B and C.	Evaluate strategies for crop management and impact on disease epidemiology.	Evaluate approaches and identify areas of future research for disease control by management of cropping systems. Linkages with IPM approaches.

**Table B. Viruses, Epidemiology, and Virus-Vector Interactions (continued)**

Approaches/Goals	Year 1	Year 2	Year 3	Year 4	Year 5
<p><b>Control of virus diseases: development of virus resistant germplasm through conventional and engineered/molecular approaches. Define prospective strategies for selecting candidate viruses, identifying specific virus diseases to target, and prioritize specific crops and cultivars for protection approaches.</b></p>	<p>Define strategies for resistance efforts. Identify target viruses. Identify germplasm with virus resistance. Initiate efforts toward defining prospective engineered resistance strategies. Identify candidate crops and recipient cultivars.</p>	<p>Continue to define suitable strategies for determining target viruses. Isolate and characterize virus-resistant germplasm. Continue work toward engineered resistance in target crops and selected viruses.</p>	<p>Further identification of resistant germplasm and develop new methods of incorporating resistance into crop plants. Evaluate resistance strategies with respect to broad spectrum or virus-specific protection.</p>	<p>Continue development of resistant varieties. Evaluate resistance strategies with respect to broad spectrum or virus-specific protection. Define mechanisms of resistance.</p>	<p>Evaluate resistant plants in greenhouse and field experimentation, and identify additional research. Molecular-based monitoring of transgenes in environment.</p>

\*\* Revised 3/31/2000

**Table C. Chemical Control, Biopesticides , Resistance Management, and Application Methods**

Approaches/Goals	Year 1	Year 2	Year 3	Year 4	Year 5
<b>Improve insecticide efficacy:</b>					
<ul style="list-style-type: none"> <li><b>Develop, test, and assist in the registration of insecticides, biorationals, and natural products.</b></li> </ul>	Develop new chemistries and natural products. Develop improved techniques for evaluating efficacy of insecticides. Support registration of desirable new products by providing information to regulatory agencies.	Same as Year 1. Determine new modes of action of effective materials. Elucidate biochemical pathways of synthesis and degradation of natural products.	Same as Year 2. Evaluate the potential for transforming plants with natural product genes.	Same as Year 3.	Same as Year 4.
<ul style="list-style-type: none"> <li><b>Develop improved methods of application including formulation and delivery of materials to improve control.</b></li> </ul>	Develop spray systems for better underleaf coverage. Evaluate rates, timing, placement in relation to efficacy. Consider formulation, UV protectants, and other means to improve efficacy. Develop improved methods to evaluate application efficacy. Field test under commercial conditions for technology transfer.	Same as Year 1.	Same as Year 2.	Same as Year 3.	Same as Year 4.
<b>Conserve insecticide efficacy:</b>					
<ul style="list-style-type: none"> <li><b>Relate action thresholds to insecticide usage patterns.</b></li> </ul>	Refine action thresholds based on insecticide efficacy and input from other control strategies.	Same as Year 1.	Same as Year 2.	Same as Year 3.	Same as Year 4. Summarize and recommend in IPM systems.

**Table C. Chemical Control, Biopesticides , Resistance Management, and Application Methods (continued)**

Approaches/Goals	Year 1	Year 2	Year 3	Year 4	Year 5
<ul style="list-style-type: none"> <li><b>Elucidate the role of genetic, biochemical and ecological factors leading to insecticide resistance.</b></li> </ul>	<p>Establish whitefly strains resistant and susceptible to various classes of insecticide. Conduct studies to determine the genetics and biochemistry of resistance and cross resistance to different classes of insecticide.</p>	<p>Same as Year 1. Evaluate the role of refuge habitats (weeds, tolerant crops, urban areas) to assure input of susceptible genes in whitefly population.</p>	<p>Conduct studies to determine the genetics and biochemistry of resistance and cross resistance to different classes of insecticide. Evaluate the role of refuge habitats (weeds, tolerant crops, urban areas) to assure input of susceptible genes in whitefly population. Evaluate the influence of host plant on susceptibility to insecticides.</p>	<p>Same as Year 3.</p>	<p>Same as Year 4.</p>
<p><b>Improve insecticide efficacy:</b></p>					
<p><b>Improve techniques for monitoring resistance.</b></p>	<p>Establish baseline data on toxicogenic responses of whitefly populations to new insecticides.</p>	<p>Same as Year 1. Expand comparative studies of resistance levels in diverse agro-ecosystems. Evaluate relationship between monitoring results and field efficacy.</p>	<p>Same as Year 2. Summarize, analyze, and produce standardized comparable monitoring systems.</p>	<p>Same as Year 3. Develop standard systems for general use including user friendly techniques to assist growers and extension agents to evaluate susceptibility of whitefly populations to commonly used insecticides.</p>	<p>Same as Year 4.</p>
<p><b>Develop, evaluate and refine resistance management systems</b></p>	<p>Evaluate the effects of mixtures and rotations of new and old chemistries to mitigate selection for resistance.</p>	<p>Same as Year 1. Develop methods to evaluate and augment the beneficial influence of refuges as sources of susceptible genes to the population pool.</p>	<p>Same as Year 2. Develop criteria for integration of successful strategies in agricultural systems. Field test resistance management systems as long range components of successful IPM.</p>	<p>Same as Year 3.</p>	<p>Same as Year 4. Technology transfer.</p>

**Table C. Chemical Control, Biopesticides , Resistance Management, and Application Methods (continued)**

Approaches/Goals	Year 1	Year 2	Year 3	Year 4	Year 5
<b>Integrate chemical control with other tactics.</b>	Evaluate selectivity of synthetic insecticides and natural products to key whitefly natural enemies.	Same as Year 1. Test compatibility of biological control with selective synthetic or natural product insecticides as required.	Same as Year 2. Integrate systems with host plant resistance and cultural controls.	Test compatibility of biological control with selective synthetic or natural product insecticides as required. Integrate systems with host plant resistance and cultural controls.	Integrate systems with host plant resistance and cultural controls. Summarization and technology transfer.

<sup>a</sup> See Table A for complementary research on thresholds.

<sup>a</sup> See Table B for complementary research on virus/vector interactions.

<sup>a</sup> See Table D for complementary research on biological control.

<sup>b</sup> See Tables E and F for complementary research on systems management.

**Table D. Natural Enemy Ecology and Biological Control**

Approaches/Goals <sup>a</sup>	Year 1	Year 2	Year 3	Year 4	Year 5
<b>Natural control and conservation:</b>					
<ul style="list-style-type: none"> <li>• <b>Develop natural enemy conservation practices to reduce mortality to indigenous and introduced natural enemies.</b></li> </ul>	<p>Conduct life table analyses of indigenous and introduced natural enemies to identify key mortality factors of natural enemy populations.</p>	<p>Identify the spatial scale upon which the key mortality agents are acting.</p>	<p>Conduct manipulative experiments to evaluate the impact of each natural enemy mortality agent on whitefly suppression.</p>	<p>Conduct a feasibility study and economic assessment of altered crop management practices that may enhance the impact of indigenous natural enemies.</p>	<p>Develop and evaluate area wide programs to facilitate full implementation.</p>
<ul style="list-style-type: none"> <li>• <b>Evaluate potential of alternate plants it act as in-field refuges or insectaries for natural enemies.</b></li> </ul>	<p>Identify potential plants for natural enemy population development and assess risks of these plants to foster additional pest problems.</p>	<p>Determine refugia plant phenology in relation to cultivated crop phenology.</p>	<p>Conduct field tests to assess whether refuges act of natural enemy and whitefly sinks or sources to adjacent cropping systems.</p>	<p>Conduct field tests to evaluate spacing of refuges necessary to achieve satisfactory whitefly suppression.</p>	<p>Conduct a feasibility study and economic assessment of alternate plantings in terms of an entire crop management program.</p>
<ul style="list-style-type: none"> <li>• <b>Assess cues used by natural enemies to locate whitefly to identify potential methods for enhancing natural enemy activity.</b></li> </ul>	<p>Conduct laboratory tests to identify cues used by natural enemies to locate and attack whitefly.</p>	<p>Determine potential methods for manipulating cues as part of a whitefly management program.</p>	<p>Conduct small scale trials to enhance whitefly suppression by manipulating natural enemy location and attack of whitefly.</p>	<p>Conduct large scale field trials and evaluate product development for commercial investment as necessary.</p>	<p>Transfer technology (as needed) to commercial interests for full implementation.</p>
<b>Augmentation of natural enemies:</b>					
<ul style="list-style-type: none"> <li>• <b>Develop natural enemy mass-rearing systems.</b></li> </ul>	<p>Identify natural enemies with the highest potential for controlling whitefly in key cropping systems.</p>	<p>Determine nutritional, physiological, and ecological requirements for mass-rearing.</p>	<p>Develop rearing systems on selected hosts and on artificial diets. Determine economic feasibility of the procedure.</p>	<p>Evaluate rearing system effects on natural enemy life history characteristics, behavior, and ability to suppress whitefly populations.</p>	<p>Facilitate transfer of mass-rearing technology to commercial interests as necessary.</p>

**Table D. Natural Enemy Ecology and Biological Control (continued)**

Approaches/Goals <sup>a</sup>	Year 1	Year 2	Year 3	Year 4	Year 5
<b>Importation biological control:</b>					
<ul style="list-style-type: none"> <li>• <b>Develop release technologies to maximize the effectiveness of mass-reared natural enemies in the field.</b></li> </ul>	<p>Identify natural enemies with the highest potential for controlling whitefly in key cropping systems and that may be economically mass produced.</p>	<p>Evaluate the fate of natural enemy life stages under field conditions to identify the appropriate developmental stage to be released.</p>	<p>Develop necessary technology for release of the appropriate natural enemy life stage.</p>	<p>Evaluate release technology effects on natural enemy life history characteristics, behavior, and ability to suppress whitefly populations.</p>	<p>Facilitate transfer of mass-rearing technology to commercial interests as necessary.</p>
<ul style="list-style-type: none"> <li>• <b>Evaluate augmentative parasitoid, predator, or pathogen releases.</b></li> </ul>	<p>Initiate studies on natural enemy augmentation with identified high potential natural enemies.</p>	<p>Conduct releases on selected crop systems at various rates of release.</p>	<p>Identify optimal release strategies for key cropping systems.</p>	<p>Continue evaluation of releases, determine need for additional releases. Compare results in different cropping systems and environments.</p>	<p>Analyze information and make recommendation regarding need for expansion of the approach.</p>
<ul style="list-style-type: none"> <li>• <b>Evaluate the ability of exotic natural enemies to suppress whitefly populations under field conditions.</b></li> </ul>	<p>Identify sites suitable for the release and subsequent evaluation of each candidate natural enemy. Conduct inoculative releases of natural enemies.</p>	<p>Evaluate establishment of exotic natural enemies within target release area. Determine if additional releases are necessary.</p>	<p>Assess spread of established natural enemies and their ability to suppress whitefly populations.</p>	<p>Continue to assess the spread of established natural enemies and their ability to suppress whitefly populations. Evaluate program progress and determine if additional strategies are necessary.</p>	<p>Complete program analysis. Publish program assessment and conduct an economic assessment.</p>
<ul style="list-style-type: none"> <li>• <b>Clarify systematics of predators, parasitoids and pathogens.</b></li> </ul>	<p>Conduct taxonomic studies of species within targeted release sites. Verify taxonomic purity of mass-reared natural enemies. Complete taxonomic work on poorly characterized but important groups. Assist in determining most suitable natural enemies for release through biogeographical analysis.</p>	<p>Provide taxonomic support for importation and mass-rearing programs. Publish keys to assist in species identifications.</p>	<p>Provide taxonomic support for importation and mass-rearing programs.</p>	<p>Provide taxonomic support for importation and mass-rearing programs.</p>	<p>Provide taxonomic support for importation and mass-rearing programs.</p>



**Table D. Natural Enemy Ecology and Biological Control (continued)**

Approaches/Goals <sup>a</sup>	Year 1	Year 2	Year 3	Year 4	Year 5
<b>Systematics, ecology, and population dynamics of natural enemies<sup>b</sup>:</b>					
<ul style="list-style-type: none"> <li>• <b>Determine <i>Bemisia</i> - natural enemy-host plant (Tritrophic) interactions.</b></li> </ul>	Initiate studies to identify mechanisms involved in <i>Bemisia</i> - and natural enemy plant attraction.	Study plant characteristics mediating whitefly population densities.	Study compatibility of characteristics of plant traits mediating whitefly populations with the abilities of natural enemies to suppress whitefly populations.	Assess the implementability of favorable tritrophic interactions within the context of an whitefly management program.	Implement and evaluate large scale crop management programs for suppression of whitefly populations.
<ul style="list-style-type: none"> <li>• <b>Identify the attributes of natural enemy biology and population level interactions to explain biological control successes and failures.</b></li> </ul>	Assess the value of the <i>Bemisia</i> biological control research to evaluate key issues to the science of biological control.	In conjunction with field evaluations, validate predictions made by behavioral and population models important to biological control.	Assess deviations between theoretical predictions and field data.	Evaluate behavioral or population level parameters that may explain observed deviations.	Quantify the impact of basic research on the development of feasible biological control programs for <i>Bemisia</i> and the advancement of the field as a science.

<sup>a</sup> See Table C for complementary research.

<sup>b</sup> See Table A for complementary research.

**Table E. Host Plant Resistance, Physiological Disorders, and Host Plant Interactions**

Approaches/Goals	Year 1	Year 2	Year 3	Year 4	Year 5
<b>Characterize resistance mechanisms and identify chemical/morphological components, and study effects of insect adaptation.</b>	Identify potential sources of germplasm for disease, plant disorders and whitefly resistance. <sup>a</sup>	Determine physiological and/or morphological basis for resistance, & effects of host-plant history and insect adaptation on plant resistance to whiteflies. Continue to identify resistant germplasm.	Elucidate biochemical and molecular basis for resistance. Continue to identify resistant germplasm.	Determine potential for transfer of resistance traits.	Evaluate potential for incorporating <i>Bemisia</i> , plant disorder and disease resistance into acceptable plant type.
<b>Develop molecular level techniques to produce resistant germplasm.</b>	Identify physiological processes of whiteflies to target for inhibition.	Identify natural products for inhibiting processes.	Isolate the relevant biosynthetic enzymes that encode for natural products inhibiting processes.	Insert genes into plants via plant transformation. <sup>b</sup>	Evaluate potential of newly transformed germplasm.
<b>Incorporate resistance traits into commercial genotypes.</b>	Identify and isolate genetic sources of resistance for transformation and/or breeding.	Insert genes into plants <sup>b</sup> via plant transformation.	Evaluate potential of newly transformed germplasm.	Continue to refine resistance factors to improve resistance in newly transformed germplasm.	Incorporate other desirable plant characteristics for crop production.
<b>Determine influence of host plant morphology, physiology and phenology on feeding behavior and competition.<sup>c</sup></b>	Characterize nutritional and other preference properties of various host plants.	Determine the biochemical mechanism regulating adaptation to host plants.	Determine changes in whitefly gene expression in response to host manipulation.	Relate changes in gene expression to whitefly physiology.	Summarize and disseminate results.
<b>Define whitefly feeding and oviposition behavior and investigate approaches for interrupting whitefly feeding and digestion.<sup>d</sup></b>	Investigate approaches for interruption of feeding, assimilation, development and reproduction.	Identify physiological and morphological mechanisms regulating processes.	Determine biochemical and molecular basis for inhibiting processes.	Determine potential for transfer of resistance traits.	Insert genes into plants <sup>a</sup> via plant transformation.
<b>Study whitefly toxicogenic plant reactions.</b>	Determine effects of whitefly feeding on host plant physiology, morphology and anatomy.	Determine biochemical basis for physiological response of plant.	Elucidate changes in plant gene expression.	Identify resistance germplasm.	Evaluate potential for transferring new germplasm.

<sup>a</sup> See Table B for additional plant disease resistance research.

<sup>b</sup> Progress at this point may extend to several year research.

<sup>c</sup> See Section A.

<sup>d</sup> See Section A, approach #9.

**Table F. Integrated and Areawide Pest Management Approaches and Crop Management Systems**

Approaches/Goals <sup>a</sup>	Year 1	Year 2	Year 3	Year 4	Year 5
<b>Development:</b>					
<b>Study whitefly-crop interactions<sup>b</sup> as cultural components that affect population dynamics, e.g., water, nutrients, plant population, planting/ termination/harvest dates, other farm practices, intercrop relationships.</b>	Identify potential beneficial or exacerbating farm practices or inputs for testing.	Determine nature and character of relationship between interaction and whitefly population dynamics.	Identify mechanisms governing relationship and alter or manipulate factors that suppress whitefly dynamics.	Refine system, add other compatible components, evaluate economic impact; conduct field testing and evaluations.	Conduct economic analyses and determine next level of IPM/ICM systems evaluation. Develop recommendations of best management practices.
<b>Develop behavioral barriers<sup>b</sup> to whitefly colonization and population development, e.g., mulches, trap crops, intercropping, row covers, etc.</b>	Review potential behavioral disrupters and evaluate as potential IPM components.	Conduct field-level trials; quantify impact to crop and whitefly dynamics	Apply promising technologies to high-value crop systems; field test and evaluate.	Refine system, add other components, and conduct economic feasibility analyses.	Summarize and evaluate results; prepare crop systems-specific recommendations.
<b>Integration:</b>					
<b>Develop Integrated Pest Management<sup>c</sup> systems using dual or multiple control tactics, e.g., cultural, biological, chemical, host plant resistance, etc.</b>	Identify candidate dual or multiple control tactic systems, e.g., IGRs and natural enemy conservation.	Initiate field testing of candidate systems.	Continue field testing & evaluate feasibility of large scale testing; add components as necessary.	Initiate large-scale experiments; incorporate economic evaluation.	Evaluate multiple component system as potential deliverable; prepare recommendations.
<b>Integrate sampling with other key components of IPM systems, e.g., thresholds, economics, decision-making, biological control, etc.</b>	Develop or modify sampling systems for new crops; integrate with thresholds and decision-making.	Establish practical utility of system through economic analyses; field efficiencies and costs.	Integrate additional control components into sampling, threshold & decision-making systems	Evaluate in whole field systems. Identify weaknesses; target improvements.	Evaluate redesigned decision systems; continue field testing and economic analyses.
<b>Delivery and Implementation:</b>					
<b>Elevate single field/farm practices to areawide community-based contexts; develop methodology for installing and evaluating areawide control technologies and their impact.</b>	Identify agricultural communities amenable to areawide management; conduct thorough pre-implementation evaluation.	Install control technologies into community; develop systems for evaluation.	Identify additional IPM/ICM compatible components. Re-assess and adapt program. Conduct areawide economic analyses.	Formulate clientele surveys; develop & begin to implement protocols for evaluating areawide technologies.	Refine, reevaluate and identify weaknesses. Formulate recommendations for future areawide management systems. Conduct surveys.

**Table F. Integrated and Areawide Pest Management Approaches and Crop Management Systems**

Approaches/Goals <sup>a</sup>	Year 1	Year 2	Year 3	Year 4	Year 5
<b>Implement and deliver Integrated Pest Management and Integrated Crop Management systems or system components to clientele.</b>	Develop and distribute provisional IPM & ICM recommendations.	Conduct whole farm/operation demonstrations of IPM systems.	Expand sites of testing with grower cooperators; conduct validation studies.	Incorporate new information and economics into recommendations.	Validate new components; finalize recommendations; expand to new crops.

<sup>a</sup> See Tables A to E for additional complementary research.

<sup>b</sup> See Table A for additional complementary research.

<sup>c</sup> See Table E for additional complementary research.

