## **SECTION 3**

### **TARGET SPECIES**

The primary objectives of this section are to: (1) discuss the purpose of using target species, (2) describe the criteria used by the 1993 EPA Fish Contaminant Workgroup to select target species, and (3) provide lists of recommended target species. Target species recommended for freshwater and estuarine/marine ecosystems are discussed in Sections 3.3 and 3.4, respectively.

### 3.1 PURPOSE OF USING TARGET SPECIES

The use of target species allows comparison of fish, shellfish, and turtle tissue contaminant monitoring data among sites over a wide geographic area. Differences in habitat, food preferences, and rate of contaminant uptake among various fish, shellfish, and turtle species make comparison of contaminant monitoring results within a state or among states difficult unless the contaminant data are from the same species. It is virtually impossible to sample the same species at every site, within a state or region or nationally, due to the varying geographic distributions and environmental requirements of each species. However, a limited number of species can be identified that are distributed widely enough to allow for collection and comparison of contaminant data from many sites.

Three aims are achieved by using target species in screening studies. First, states can cost-effectively compare contaminant concentrations in their state waters and then prioritize sites where tissue contaminants exceed human health screening values. In this way, limited monitoring resources can be used to conduct intensive studies at sites exhibiting the highest degree of tissue contamination in screening studies. By resampling target species used in the screening study in Phase I intensive studies and sampling additional size classes and additional target species in Phase II intensive studies as resources allow, states can assess the magnitude and geographic extent of contamination in species of commercial, recreational, or subsistence value. Second, the use of common target species among states allows for more reliable comparison of sampling information. Such information allows states to design and evaluate their own contaminant monitoring programs more efficiently, which should further minimize overall monitoring costs. For example, monitoring by one state of fish tissue contamination levels in the upper reaches of a particular river can provide useful information to an adjacent state on tissue contamination levels that might be anticipated in the same target species at sampling sites downstream. Third, the use of a select group of target fish, shellfish, and freshwater turtle species will allow for the development of a national database for tracking the magnitude and

geographic extent of pollutant contamination in these target species nationwide and will permit analyses of trends in fish, shellfish, and turtle contamination over time.

### 3.2 CRITERIA FOR SELECTING TARGET SPECIES

The appropriate choice of target species is a key element of any chemical contaminant monitoring program. Criteria for selecting target species used in the following national fish and shellfish contaminant monitoring programs were reviewed by the 1993 EPA Fish Contaminant Workgroup to assess their applicability for use in selecting target species for state fish contaminant monitoring programs:

- National Study of Chemical Residues in Fish (U.S. EPA)
- National Dioxin Study (U.S. EPA)
- 301(h) Monitoring Program (U.S. EPA)
- National Pesticide Monitoring Program (U.S. FWS)
- National Contaminant Biomonitoring Program (U.S. FWS)
- National Status and Trends Program (NOAA).
- National Water Quality Assessment Program (USGS).

The criteria used to select target species in many of these programs are similar although the priority given each criterion may vary depending on program aims.

According to the 1993 EPA Fish Contaminant Workgroup, the most important criterion for selecting target fish, shellfish, and turtle species for state contaminant monitoring programs assessing human consumption concerns was that the species were commonly consumed in the study area and were of commercial, recreational, or subsistence fishing value. Two other criteria of major importance are that the species have the potential to bioaccumulate high concentrations of chemical contaminants and have a wide geographic distribution. EPA recommends that states use the same criteria to select species for both screening and intensive site-specific studies.

In addition to the three primary criteria for target species selection, it is also important that the target species be easy to identify taxonomically because there are significant species-specific differences in bioaccumulation potential. Because many closely related species can be similar in appearance, reliable taxonomic identification is essential to prevent mixing of closely related species with the target species. **Note:** Under no circumstance should individuals of more than one species be mixed to create a composite sample (U.S. EPA, 1991e). It is also both practical and cost-effective to sample target species that are abundant, easy to capture, and large enough to provide adequate tissue samples for chemical analyses.

It cannot be overemphasized that final selection of target species will require the expertise of state fisheries biologists with knowledge of local species that best meet the selection criteria and knowledge of local human consumption patterns. Although, ideally, all fish, shellfish, or turtle species consumed from a given waterbody by the local population should be monitored, resource constraints may dictate that only a few of the most frequently consumed species be sampled.

In the next two sections, lists of recommended target species are provided for freshwater ecosystems (inland fresh waters and the Great Lakes) and estuarine/marine ecosystems (Atlantic, Gulf, and Pacific waters), and the methods used to develop each list are discussed.

### 3.3 FRESHWATER TARGET SPECIES

As part of the two-tiered sampling strategy proposed for state fish contaminant monitoring programs, EPA recommends that states collect one bottom-feeding fish species and one predator fish species at each freshwater screening study site. Some suggested target species for use in state fish contaminant monitoring programs are shown in Table 3-1 for inland fresh waters and in Table 3-2 for Great Lakes waters.

The lists of target species recommended by the 1993 EPA Fish Contaminant Workgroup for freshwater ecosystems were developed based on a review of species used in the following national monitoring programs:

- National Study of Chemical Residues in Fish (U.S. EPA)
- National Dioxin Study (U.S. EPA)
- National Pesticide Monitoring Program (U.S. FWS)
- National Contaminant Biomonitoring Program (U.S. FWS)
- National Water Quality Assessment Program (USGS)

and on a review of fish species cited in state fish consumption advisories or bans (RTI, 1993). Separate target species lists were developed for inland fresh waters (Table 3-1) and Great Lakes waters (Table 3-2) because of the distinct ecological characteristics of these waters and their fisheries. Each target species list has been reviewed by regional and state fisheries experts.

Use of two distinct ecological groups of finfish (i.e., bottom-feeders and predators) as target species in freshwater systems is recommended. This permits monitoring of a wide variety of habitats, feeding strategies, and physiological factors that might result in differences in bioaccumulation of contaminants. Bottom-feeding species may accumulate high contaminant concentrations from direct physical contact with contaminated sediment and/or by consuming benthic invertebrates and epibenthic organisms that live in contaminated sediment. Predator species are also good indicators of persistent pollutants (e.g., mercury or DDT and its metabolites) that may be biomagnified through several trophic levels of the food web. Species used in several federal programs to assess the

Family name	Common name	Scientific name
Percichthyidae	White bass	Morone chrysops
Centrarchidae	Largemouth bass Smallmouth bass Black crappie White crappie	Micropterus salmoides Micropterus dolomieui Pomoxis nigromaculatus Pomoxis annularis
Percidae	Walleye Yellow perch	Stizostedion vitreum Perca flavescens
Cyprinidae	Common carp	Cyprinus carpio
Catostomidae	White sucker	Catostomus commersoni
lctaluridae	Channel catfish Flathead catfish	lctalurus punctatus Pylodictis olivaris
Esocidae	Northern pike	Esox lucius
Salmonidae	Lake trout Brown trout Rainbow trout	Salvelinus namaycush Salmo trutta Oncorhynchus mykissª

 Table 3-1. Recommended Target Species for Inland Fresh Waters

<sup>a</sup>Formerly *Salmo gairdneri*.

Family name	Common name	Scientific name
Percichthyidae	White bass	Morone chrysops
Centrarchidae	Smallmouth bass	Micropterus dolomieui
Percidae	Walleye	Stizostedion vitreum
Cyprinidae	Common carp	Cyprinus carpio
Catostomidae	White sucker	Catostomus commersoni
lctaluridae	Channel catfish	lctalurus punctatus
Esocidae	Muskellunge	Esox masquinongy
Salmonidae	Chinook salmon	Oncorhynchus tschawytscha
	Lake trout	Salvelinus namaycush
	Brown trout	Salmo trutta
	Rainbow trout	Oncorhynchus mykiss <sup>a</sup>

### Table 3-2. Recommended Target Species for Great Lakes Waters

<sup>a</sup>Formerly *Salmo gairdneri*.

extent of freshwater fish tissue contamination nationwide are compared in Table 3-3.

In addition to finfish species, states should consider monitoring the tissues of freshwater turtles for environmental contaminants in areas where turtles are consumed by recreational, subsistence, or ethnic populations. Interest has been increasing in the potential transfer of environmental contaminants from the aquatic food chain to humans via consumption of freshwater turtles. Turtles may bioaccumulate environmental contaminants in their tissues from exposure to contaminated sediments or via consumption of contaminated prey. Because some turtle species are long-lived and occupy a medium to high trophic level of the food chain, they have the potential to accumulate high concentrations of chemical contaminants from their diets (Hebert et al., 1993). Some suggested target turtle species for use in state contaminant monitoring programs are listed in Table 3-4.

The list of target turtle species recommended for freshwater ecosystems was developed based on a review of turtle species cited in state consumption advisories or bans (RTI, 1993) and a review of the recent scientific literature. The recommended target species list has been reviewed by regional and state experts.

### 3.3.1 Target Finfish Species

### 3.3.1.1 Bottom-Feeding Species

EPA recommends that, whenever practical, states use common carp (*Cyprinus*) carpio), channel catfish (Ictalurus punctatus), and white sucker (Catostomus commersoni) in that order as bottom-feeding target species in both inland fresh waters (Table 3-1) and in Great Lakes waters (Table 3-2). These bottom-feeders have been used consistently for monitoring a wide variety of contaminants including dioxins/furans (Crawford and Luoma, 1993; U.S. EPA, 1992c, 1992d; Versar Inc., 1984), organochlorine pesticides (Crawford and Luoma, 1993; Schmitt et al., 1983, 1985, 1990; U.S. EPA, 1992c, 1992d), and heavy metals (Crawford and Luoma, 1993; Lowe et al., 1985; May and McKinney, 1981; Schmitt and Brumbaugh, 1990; U.S. EPA, 1992c, 1992d). These three species are commonly consumed in the areas in which they occur and have also demonstrated an ability to accumulate high concentrations of environmental contaminants in their tissues as shown in Tables 3-5 and 3-6. Note: The average contaminant concentrations shown in Tables 3-5 and 3-6 for fish collected for the EPA National Study of Chemical Residues in Fish (U.S. EPA, 1992c, 1992d) were derived from concentrations in fish from undisturbed areas and from areas expected to have elevated tissue contaminant concentrations. The mean contaminant concentrations shown, therefore, may be higher or lower than those found in the ambient environment because of site selection criteria used in this study.

	U.S. EPA National	U.S. FWS	U.S. EPA	USGS
	Dioxin Study	NPMP and NCBP	NSCRF	NWQAP
BOTTOM FEEDERS			1	
Family <i>Cyprinidae</i> Carp ( <i>Cyprinus carpio</i> )	•	•	•	•
Family <i>lcataluridae</i> Channel catfish ( <i>lctalurus punctatus</i> )	•	• Or other ictalurid	•	•
Family Catostomidae White sucker (Catastomus commersoni)	•	Or other catostomid	•	•
Longnose sucker (C. catostromus)				•
Largescale sucker (C.macrocheilus)				
Spotted sucker (Minytrema melanops)			•	
Redhorse sucker (Moxostoma sp.) included variety of species: Silver redhorse (M. anisurum) Grey redhorse (M. congetum) Black redhorse (M. duquesnei) Golden redhorse (M. erythrurum) Shorthead redhorse (M. macrolepidotum) Blacktail redhorse (M. poecilurum)			•	
PREDATORS				
Family Salmoridae Rainbow trout (Oncortynchus mykiss) [formerly Salmo gairdneni] Brown trout (Salmo trutta)	•	•	•	•
Brook trout (Salvelinus fontinalis)	•	•		•
Lake trout (Salmo namaycush)	•	•		
Family <i>Percidae</i> Walleye ( <i>Stizostedion vitreum</i> )	● Or other pericid	Or other pericid	•	
Sauger (Stizostedion canadense)	О	0		
Yellow perch (Perca flavescans)	О	0		
Family Percichthyidae White bass (Morone chrysops)			•	
Family <i>Centrarchidae</i> Largemouth bass <i>(Micropterus salmoides)</i>	• Or other centrarchid	● Or other centrarchid	•	•
Smallmouth bass (Micropterus dolomieui)			•	
Black crappie (Pomoxis nigromaculatus)	О	0		
White crappie (Pomoxis annularis)	О	0	•	
Bluegill sunfish (Lepornis macrochirus)	О	0		•
Family <i>Esocidae</i> Northern pike <i>(Esox lucius)</i>			•	
Family Ictaluridae Flathead catfish (Pylodictis olivaris)			•	

### Table 3-3. Comparison of Freshwater Finfish Species Used in Several National **Fish Contaminant Monitoring Programs**

• Recommended target species

O Alternate target species

NPMP = National Pesticide Monitoring Program

NCBP = National Contaminant Biomonitoring Program NSCRF = National Study of Chemical Residues in Fish NWQAP = National Water Quality Assessment Program

Sources: Versar, Inc., 1984; Schmitt et al., 1990; Schmitt et al., 1983; May and McKinney, 1981; U.S. EPA, 1992c, 1992d; Crawford and Luoma, 1993.

Family name	Common name	Scientific name
Chelydridae	Snapping turtle	Chelydra serpentina
Emydidae	Yellow-bellied turtle Red-eared turtle River cooter Suwanee cooter Slider Texas slider Florida cooter Peninsula cooter	Trachemys scripta scripta Trachemys scripta elegans Pseudemys concinna concinna Pseudemys concinna suwanniensis Pseudemys concinna hieroglyphica Pseudemys concinna texana Pseudemys floridana floridana Pseudemys floridana penisularis
Trionychidae	Smooth softshell Eastern spiny softshell Western spiny softshell Gulf Coast spiny softshell Florida softshell	Apalone muticus Apalone spinifera spinifera Apalone spinifera hartwegi Apalone spinifera aspera Apalone ferox

 Table 3-4.
 Freshwater Turtles Recommended for Use as Target Species

In addition, these three species are relatively widely distributed throughout the continental United States, and numerous states are already sampling these species in their contaminant monitoring programs. A review of the database *National Listing of State Fish and Shellfish Consumption Advisories and Bans* (RTI, 1993) indicated that the largest number of states issuing advisories for specific bottom-feeding species did so for carp (21 states) and channel catfish (22 states), with eight states issuing advisories for white suckers (see Table 3-7). Appendix D lists the freshwater fish species cited in consumption advisories for each state as of 1998.

### 3.3.1.2 Predator Species

EPA recommends that, whenever practical, states use predator target species listed in Tables 3-1 and 3-2 for inland fresh waters and Great Lakes waters. respectively. Predator species, because of their more definitive habitat and water temperature preferences, generally have a more limited geographic distribution. Thus, a greater number of predator species than bottom feeders have been used in national contaminant monitoring programs (Table 3-3) and these are recommended for use as target species in freshwater ecosystems. Predator fish that prefer relatively cold freshwater habitats include many members of the following families: Salmonidae (trout and salmon), Percidae (walleye and yellow perch), and *Esocidae* (northern pike and muskellunge). Members of the Centrarchidae (large- and smallmouth bass, crappie, and sunfish), Percichthyidae (white bass), and Ictaluridae (flathead catfish) families prefer relatively warm water habitats. Only two predator species (brown trout and largemouth bass) were used in all four of the national monitoring programs reviewed by the 1993 EPA Fish Contaminant Workgroup (Table 3-3). However, most of the other predator species recommended as target species have been used in at least one national monitoring program. To identify those predator species with a known ability to bioaccumulate contaminants in their tissues, the 1993 EPA Workgroup reviewed average tissue concentrations of xenobiotic contaminants for major

	Alpha-	Gamma-						Heptachlor			Oxychlor	
	BHC	BHC	Biphenyl	Chlorpyritos	Dicotol	Dieldrin	Endrin	ерохід		Mercury Mirex		PCBS
Bottom Feeders" Carn	3 10	4 34	4.38	8 23	0.88	44 75	1 40	4 00		0.11 3.70		2941 13
White cueber	0.00	1 66		1 75	0.00	00.7E					0.10	•
	0.0	00 	07.1			26.13	4 C C	01.0				
	7.87	3.17	1.24	0.97	60.0	44.CI	9.07	nc:n		PC.41 P.0.0		-
Redhorse sucker	0.82	0.41	1.25	0.35	QN	5.35	0.97	DN	5	.27 0.57	7 2.37	
Spotted sucker	1.45	2.63	3.35	0.56	0.05	5.52	Q	QN	5	0.12 1.7		133.90
Predators"							!					
Largemouth bass	0.15	0.07	0.38	0.23	0.20	5.01	QN	0.30	5			
Smallmouth bass	0.36	0.15	0.33	0.08	QN	2.34	QN	0.07	5			
Walleye	DN	DN	0.40	0.04	QN	3.73	QN	0.21	J			368.65
Brown trout	1.59	DN	0.81	QN	0.94	20.13	QN	2.08	J	0.14 43.98		2434.07
White bass	0.34	0.79	0.62	1.32	QN	9.35	QN	1.40	J	0.35 0.11		288.35
Northern pike	0.55	DN	0.59	11.43	0.31	9.04	QN	DN	J			
Flathead cat	0.92	0.58	0.60	22.57	1.28	37.38	3.45	0.57	J			
White crappie	0.23	DN	0.21	QN	DN	DN	QN	DN	J	0.22 ND	QN	
Bluefish	0.38	0.12	0.20	DN	QN	2.87	QN	DN	J	0.22 0.13		368.06
	Penta-	Penta-										Hexa-
Fish Species	chloro- anisole	chloro- benzene	e DDE	Total Chlordane	Total e Nonachlor	-	23 TCB 1	24 TCB	135 TCB	1234 TECB	Trifluralin	chloro- benzene
Bottom Feeders <sup>b</sup>												
Carp	16.50	1.04	415.43		63.15		54	4.77	0.08	0.30	12.55	3.5
White sucker	90.6	0.39	78.39	18.43	20.83		0.16	0.30	0.14	0.15	QN	3.62
Channel cat	39.60	1.32	627.77	54.39	66.28		0.14	0.37	DN	0.88	1.00	2.36
Redhorse sucker	2.87	0.02	87.25	16.48	30.73		0.55	6.48	0.08	0.09	QN	0.58
Spotted sucker	17.68	0.02	75.31	12.33	15.0(		34	12.00	1.00	0.09	QN	0.0
l argemouth bass	0.57	0 0	55 72	2 89	4 21		22	0 19	0.03	0.01	CIN	0
Smallmouth hass	0.23	20.0	33.63	4 01	7 82		0 70	0.59	0.04	0.04	S	0.36
Walleve	0.76	Q	34.00	3.62	8.04		0.29	0.38	ND	0,004	Q	0.11
Brown trout	0.09	0.60	158.90		32.60		1.10	0.98	QN	0.09	QN	3.0
White bass	0.93	QN	17.44		16.00		0.21	0.10	QN	0,01	QN	0.83
Northern pike	1.51	0.09	59.50	5.45	13.88		0.30	0.23	DN	0.01	QN	0.2
Flathead catfish	0.31	QN	755.18		14.04		0.10	0.18	DN	DN	44.37	0.85
White crappie	0.33	Q	10.04	0.34	0.28		0.08	0.08	0.08	QN	QN	QN
Bluefish	0.05	QN	29.13	2.74	2.56		6.25	4.66	4.66	ND	QN	ND

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Source: US. EPA, 1991h.

Table 3-6. Average Fish Tissue Concentrations (ppt) of Dioxins and Furans for Major Finfish Species

			Sampl	npled in	n the Né	ed in the National Study of Chemical Residues in	Study	of Che	emical	Resid	ues in	Fish <sup>a</sup>		•		
Fish Species	2378 TCDD	12378 PeCDD	123478 HxCDD	123678 HxCDD	123789 HxCDD	1234678 HpCDD	2378 TCDF	12378 PeCDF	23478 PeCDF	123478 HxCDF	123678 HxCDF	123789 HxCDF	234678 HxCDF	1234678 HpCDF	1234789 HpCDF	ТЕО
Bottom Feeders <sup>b</sup>																
Carp	7.76	3.63	2.16	6.81	1.54	22.29	10.15	1.31	4.01	2.54	1.91	1.16	1.20	2.49	1.22	13.06
White sucker	8.08	2.05	1.03	1.96	0.88	3.72	22.89	1.10	2.64	2.21	1.29	1.06	1.09	1.23	1.13	12.79
Channel catfish	11.56	2.37	1.61	5.62	1.29	9.40	2.22	0.52	2.91	2.41	1.41	1.38*	1.62	2.55	1.26	14.80
Redhorse sucker	4.65	1.50	1.40	2.36	0.84	4.94	30.09	0.75	1.28	2.10	1.16	1.19*	1.50	1.57	1.36*	9.22
Spotted sucker	1.73	2.34	1.70	12.08	1.14	17.48	7.49	2.12	2.06	2.22	1.79	1.28*	1.78	1.77	1.08	6.23
Predators <sup>b</sup>																
Largemouth bass	1.73	0.59	1.12	1.28	0.64	2.48	2.18	0.37	0.47	1.24	1.23	1.21*	0.88	0.82	1.21*	1.91
Smallmouth bass	0.72	0.50*	1.13*	0.79	0.64*	0.67	1.93	0.36*	0.51	1.28	1.23	1.26*	0.89*	0.69	1.30*	0.65*
Walleye	0.88	0.54*	0.99*	0.73	0.62*	0.88	1.83	0.35*	0.38	1.04	1.09*	1.07*	0.75	0.74	1.21*	0.79*
Brown trout	2.52	1.01	1.07*	0.98	0.68*	1.18	3.74	09.0	1.36	1.47	1.12*	1.09*	0.94*	0.67*	1.16*	3.31
White bass	3.00	0.66	1.05*	0.78	0.61*	1.01	5.07	0.40	0.49	1.04	1.16*	1.13*	0.81*	0.63	1.17*	3.44
Northern pike	0.77	0.46*	1.23*	0.91	0.69*	0.73	1.01	0.44	0.66	1.41*	1.42*	1.38*	0.98*	0.56	1.30*	0.66
Flathead cat	0.78	0.43	06.0	1.06	0.50	1.67	1.63	0.40	0.56	1.05	1.20*	1.17*	0.61*	0.56	1.10*	0.99
White crappie	2.13	09.0	1.29*	1.03*	0.83*	1.33	10.46	0.54	0.67	1.33*	1.33*	1.30*	0.95*	0.96*	1.34*	3.80
Bluefish	0.85	0.56	1.23*	0.98*	0.69*	0.65	2.11	0.41	0.59	1.42*	1.42*	1.39*	0.98*	0.72*	1.31*	1.41
<sup>a</sup> These average fish tissue concentrations may be higher or lower than those found in the ambient environment because of site selection criteria used in this study. <sup>b</sup> Values were calculated using whole-body samples for bottom feeders and fillet samples for predators. Values below detection limit have been replaced by one-half detection limit for the given sample. Asterisk indicates all values below detection limit. Unlits = ppt (pg/g) wet weight basis. TEQ = Toxicity equivalency was based on TEF-89 toxicity weighting values; however, octachlorodibenzo- <i>p</i> -dioxin and octachlorodibenzofurans were not analyzed; therefore, the TEQ value does not include these two compounds.	th tissue co lated usin Asterisk i luivalency slude these	oncentrati g whole-b ndicates a was base two com	ons may b oody sampl all values t d on TEF-{ pounds.	e higher or es for botto selow detec 89 toxicity v	lower than im feeders tion limit. I veighting v	those foun and fillet sa Jnits = ppt alues; howε	d in the ar imples for (pg/g) wei »ver, octa	mbient er r predator t weight t chlorodib	rvironmen rs. Values aasis. enzo- <i>p</i> -dic	t because below de xin and o	of site sel tection lim ctachlorod	əction crite it have bee ibenzofura	ria used in en replaced ns were nc	this study. I by one-ha ot analyzed	lf detection ; therefore,	limit for the TEQ

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Source: U.S. EPA, 1991h.

predator fish species sampled in the National Study of Chemical Residues in Fish. Unlike the bottom feeders (common carp, channel catfish, and white suckers), no single predator species or group of predator species consistently exhibited the highest tissue concentrations for the contaminants analyzed (Tables 3-5 and 3-6). However, average fish tissue concentrations for some contaminants (i.e., mercury, mirex, chlorpyrifos, DDE, 1,2,3-trichlorobenzene [123-TCB], and trifluralin) were higher for some predator species than for the bottom feeders despite the fact that only the fillet portion rather than the whole body was analyzed for predator species. This finding emphasizes the need for using two types of fish (i.e., bottom feeders and predators) with different habitat and feeding strategies as target species.

The existence of fish consumption advisories for these predator target species was further justification for their recommended use. As was shown for the bottom-feeder target species, states were already sampling the recommended predator target species listed in Table 3-7. The largest number of states issuing advisories in 1993 for specific predator species did so for largemouth bass (15), lake trout (10), white bass (10), smallmouth bass (9), brown trout (9), walleye (9), rainbow trout (8), yellow perch (8), chinook salmon (7), northern pike (7), black crappie (5), flathead catfish (4), and muskellunge (4) (RTI, 1993). For comparison, the number of states reporting advisories for each species in 1998 is also presented in Table 3-7.

Because some freshwater finfish species (e.g., several Great Lake salmonids) are highly migratory, harvesting of these species may be restricted to certain seasons because sexually mature adult fish (i.e., the recommended size for sampling) may make spawning runs from the Great Lakes into tributary streams. EPA recommends that spawning populations not be sampled in fish contaminant monitoring programs. Sampling of target finfish species during their spawning period should be avoided because contaminant tissue concentrations may decrease during this time (Phillips, 1980) and because the spawning period is generally outside the legal harvest period. **Note:** Target finfish may be sampled during their spawning period, however, if the species can be legally harvested at this time.

State personnel, with their knowledge of site-specific fisheries and human consumption patterns, must be the ultimate judge of the species selected for use in freshwater fish contaminant monitoring programs within their jurisdiction.

### 3.3.2 Target Turtle Species

EPA recommends that states in which freshwater turtles are consumed by recreational, subsistence, or ethnic populations consider monitoring turtles to assess the level of environmental contamination and whether they pose a human health risk. In all cases, the primary criterion for selecting the target turtle species is whether it is commonly consumed. To identify those turtle species with a known ability to bioaccumulate contaminants in their tissues, the 1993 EPA Workgroup reviewed turtle species cited in state consumption advisories and those species identified

			Number of state	es with advisories⁵
Family name	Common name	Scientific name	1993	1998
Percichthyidae	White bass	Morone chrysops	10	17
	Striped bass	Morone saxatilis	6	12
	White perch	Morone americana	4	7
Centrarchidae	Largemouth bass	Micropterus salmoides	15	33
	Smallmouth bass	Micropterus dolomieui	9	18
	Black crappie	Pomoxis nigromaculatus	5	18
	White crappie	Pomoxis annularis	2	11
	Bluegill sunfish	Lepomis macrochirus	5	11
	Rock bass	Ambloplites rupestris	3	5
Percidae	Yellow perch	Perca flavescens	8	12
	Sauger	Stizostedion canadense	4	9
	Walleye	Stizostedion vitreum	9	12
Cyprinidae	Common carp	Cyprinus carpio	21	25
Acipenseridae	Shovelnose sturgeon	Scaphirhynchus platorynchus	1	3
	Lake sturgeon	Acipenser fulvescens	2	3
Catostomidae	Smallmouth buffalo	Ictiobus bubalus	4	5
	Bigmouth buffalo	Ictiobus cyprinellus	4	6
	Shorthead redhorse	Moxostoma macrolepidotum	2	3
	White sucker	Catostomus commersoni	8	11
	Quillback carpsucker	Carpiodes cyprinus	2	5
Ictaluridae	White catfish	Ictalurus catus	5	6
	Channel catfish	lctalurus punctatus	22	26
	Flathead catfish	Pylodictis olivaris	4	11
	Black bullhead	lctalurus melas	2	3
	Brown bullhead	lctalurus nebulosus	7	10
	Yellow bullhead	Ictalurus natalis	2	8
Sciaenidae	Freshwater drum	Aplodinotus grunniens	3	13
Esocidae	Northern pike	Esox lucius	7	10
	Muskellunge	Esox masquinongy	4	4
Salmonidae	Coho salmon	Oncorhynchus kisutch	6	8
	Chinook salmon	Oncorhynchus tschawytscha	7	7
	Brown trout	Salmo trutta	9	11
	Lake trout	Salvelinus namaycush	10	12
	Rainbow trout	Oncorhynchus mykiss <sup>c</sup>	8	12
	Brook trout	Salvelinus fontinalis	3	4 7
	Lake whitefish	Coregonus clupea formis	2	7
Anguillidae	American eel	Anguilla rostrata	6	7

## Table 3-7. Principal Freshwater Fish Species Cited in State Fish Consumption Advisories<sup>a</sup>

<sup>a</sup> Species in boldface are EPA-recommended target species for inland fresh waters (see Table 3-1) and the Great Lakes waters (Table 3-2).
 <sup>b</sup> Many states did not identify individual species of finfish in their advisories.

<sup>c</sup> Formerly *Salmo gairdneri*.
 Sources: RTI, 1993; U.S. EPA, 1999c (NLFWA).

in the scientific literature as having accumulated high concentrations of environmental contaminants.

Based on information in state advisories and a number of environmental studies using turtles as biological indicators of pollution, one species stands out as an obvious choice for a target species, the common snapping turtle (*Chelydra serpentina*). This turtle has been recommended by several researchers as an important bioindicator species (Bishop et al., 1996; Bonin et al., 1995; Olafsson et al., 1983; Stone et al., 1980) and has the widest geographic distribution of any of the North American aquatic turtles (see Figure 3-1). In addition, this species is highly edible, easily identified, easily collected, long-lived (>20 years), grows to a large size, and has been extensively studied with respect to a variety of environmental contaminants. Other turtle species that should be considered for use as target species are listed in Table 3-4.

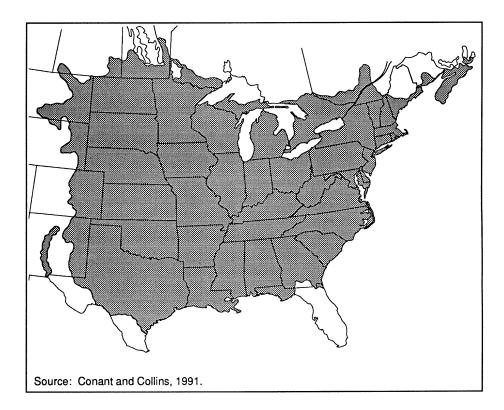


Figure 3-1. Geographic range of the common snapping turtle (*Chelydra serpentina*).

Four states (Arizona, Massachusetts, Minnesota, and New York) currently have consumption advisories in force for various turtle species (U.S. EPA, 1999c; New York State Department of Health, 1994). The species cited in the state advisories and the pollutants identified in turtle tissues as exceeding acceptable levels of contamination with respect to human health are listed in Table 3-8. New York

Family name	Common name	Scientific name	Pollutant	State
Chelydridae	Snapping turtle <sup>a</sup>	Chelydra serpentina	Mercury	MN
	Snapping turtle <sup>a</sup> (and other unspecified turtle species)	Chelydra serpentina	PCBs	MA
	Snapping turtle <sup>b</sup>	Chelydra serpentina	PCBs	NY
Trionychidae	Western spiny softshell <sup>a</sup>	Apalone spiniferus	DDT toxaphene, chlordane, dieldrin	AZ

 Table 3-8. Principal Freshwater Turtle Species Cited in State Consumption Advisories

PCB = Polychlorinated biphenyls. DDT = 1,1,1-trichloro-2,2 bis(p-chlorophenyl)ethane.

<sup>a</sup>Source: U.S. EPA 1999c (NLFWA).

<sup>b</sup>Source: New York State Department of Health, 1994.

state has a statewide advisory directed specifically at women of childbearing age and children under 15 and advises these groups to avoid eating snapping turtles altogether. The advisory also recommends that members of the general population who wish to consume turtle meat should trim away all fat and discard the liver tissue and eggs of the turtles prior to cooking the meat or preparing other dishes. These three tissues (fat, liver, and eggs) have been shown to accumulate extremely high concentrations of a variety of environmental contaminants in comparison to muscle tissue (Bishop et al., 1996; Bonin et al., 1995; Bryan et al., 1987; Hebert et al., 1993; Olafsson et al 1983; 1987; Ryan et al., 1986; Stone et al., 1980). The Minnesota advisory also recommends that consumers remove all fat from turtle meat prior to cooking as a risk-reducing strategy (Minnesota Department of Health, 1994). States should consider monitoring pollutant concentrations in all three tissues (fat, liver, and eggs) in addition to muscle tissue if resources allow. If residue analysis reveals the presence of high concentrations of any environmental contaminant of concern, the state should consider making the general recommendation to consumers to discard these three highly lipophilic tissues (fat, liver, and eggs) to reduce the risk of exposure particularly to many organic chemical contaminants.

To identify those freshwater turtle species with a known ability to bioaccumulate chemical contaminants in their tissues, several studies were reviewed that identified freshwater turtle species as useful biomonitors of PCBs (Bishop et al., 1996; Bonin et al., 1995; Bryan et al., 1987; Hebert et al., 1993; Helwig and Hora, 1983; Olafsson et al., 1983; 1987; Safe, 1985; and Stone et al., 1980), dioxins and dibenzofurans (Bishop et al., 1996; Rappe et al., 1981; Ryan et al., 1986), organochlorine pesticides (Bishop et al., 1996; Bonin et al., 1995; Hebert et al., 1983; Stone et al., 1980), heavy metals (Bonin et al., 1995; Helwig and Hora, 1983; Stone et al., 1980), and radioactive nuclides (cesium-137 and strontium-90) (Lamb et al., 1991; Scott et al., 1986). The turtle species used in these studies, the pollutants monitored, and the reference sources are summarized in Table 3-9.

Species	Pollutant monitored	Source
Snapping turtle ( <i>Chelydra serpentina)</i>	PCBs, total DDT, mirex	Hebert et al., 1993
Snapping turtle ( <i>Chelydra serpentina)</i>	PCBs	Olafsson et al., 1987 Olafsson et al., 1983
Snapping turtle ( <i>Chelydra serpentina)</i>	PCBs	Safe, 1987
Snapping turtle ( <i>Chelydra serpentina)</i>	PCBs	Bryan et al., 1987
Snapping turtle ( <i>Chelydra serpentina)</i>	Dioxins/Furans	Ryan et al., 1986
Snapping turtle ( <i>Chelydra serpentina)</i>	PCBs, mercury, cadmium	Helwig and Hora, 1983
Snapping turtle ( <i>Chelydra serpentina)</i>	Furans	Rappe et al., 1981
Snapping turtle ( <i>Chelydra serpentina</i> )	Organochlorine pesticides (DDE, dieldrin, hexachlorobenzene, heptachlor epoxide, mirex), PCBs, cadmium, mercury	Stone et al., 1980
Snapping turtle ( <i>Chelydra serpentina)</i>	29 Organochlorine pesticides, 39 PCB congeners, mercury	Bonin et al., 1995
Snapping turtle eggs	4 Organochlorine pesticides (DDE, dieldrin, mirex, hexachloro- benzene), PCBs, dioxins/furans	Bishop et al., 1996
Yellow-bellied turtle ( <i>Trachemys scripta)</i>	Cesium-137 Strontium-90	Lamb et al., 1991
Yellow-bellied turtle ( <i>Trachemys scripta)</i>	Cesium-137 Strontium-90	Scott et al., 1986

#### Table 3-9. Studies Using Freshwater Turtles as Biomonitors of Environmental Contamination

PCBs = Polychlorinated biphenyls.

DDT = 1,1,1-Trichloro-2,2 bis(p-chlorophenyl)ethane.

DDE = 1,1-Dichloro-2,2-bis(p-chlorophenyl)-ethylene.

State personnel, with their knowledge of site-specific fisheries and human consumption patterns, must be the ultimate judge of the turtle species selected for use in contaminant monitoring programs within their jurisdictions. Because several turtle species are becoming less common as a result of habitat loss or degradation or overharvesting, biologists need to ensure that the target species selected for the state toxics monitoring program is not of special concern within their jurisdiction or designated as a threatened or endangered species. For example, two highly edible turtle species, the Alligator snapping turtle (*Macroclemys temmincki*) and the Northern diamondback terrapin (*Malaclemys terrapin terrapin*) are protected in some states or designated as species of concern within portions of their geographic range and are also potential candidates for federal protection (Sloan and Lovich, 1995). Although protected to varying degrees by several states, George (1987) and Pritchard (1989) concluded that the Alligator snapping turtle should receive range-wide protection

from the federal government as a threatened species under the Endangered Species Act. Unfortunately, basic ecological and life history information necessary to make environmental management decisions (i.e., federal listing as endangered or threatened species) is often not available for turtles and other reptiles (Gibbons, 1988).

Several species of freshwater turtles already have been designated as endangered or threatened species in the United States including the Bog turtle (*Clemmys muhlenbergii*), Plymouth red-bellied turtle (*Pseudemys rubriventris bangsi*), Alabama red-bellied turtle (*Pseudemys alabamensis*), Flattened musk turtle (*Stemotherus depressus*), Ringed map (=sawback) turtle (*Graptemys oculifera*), and the Yellow-blotched map (=sawback) turtle (*Graptemys flavimaculata*) (U.S. EPA, 1994; U.S. Fish and Wildlife Service, 1994). In addition, all species of marine sea turtles including the Green sea turtle (*Chelonia mydas*), Hawksbill sea turtle (*Eretmochelys imbricata*), Kemp's ridley sea turtle (*Lepidochelys kempii*), Olive ridley sea turtle (*Lepidochelys olivacea*), Loggerhead sea turtle (*Caretta caretta*), and the Leatherback sea turtle (*Dermochelys coriacea*) have been designated as endangered (U.S. EPA, 1994; U.S. Fish and Wildlife Service, 1994).

### 3.4 ESTUARINE/MARINE TARGET SPECIES

EPA recommends that states collect **either** one shellfish species (preferably a bivalve mollusc) and one finfish species **or** two finfish species at each estuarine/marine screening site. In all cases, the primary criterion for selecting the target species is that it is commonly consumed. Ideally, one shellfish species and one finfish species should be sampled; however, if no shellfish species from the recommended target species list meets the primary criterion, EPA recommends that states use two finfish species selected from the appropriate regional estuarine/marine target species lists. If two finfish are selected as the target species, one should be a bottom-feeding species.

EPA recommends that, whenever practical, states use target species selected from fish and shellfish species identified in Tables 3-10 through 3-16 for the following specific estuarine/marine coastal areas:

- Northeast Atlantic region (Maine through Connecticut)—Table 3-10
- Mid-Atlantic region (New York through Virginia)—Table 3-11
- Southeast Atlantic region (North Carolina through Florida)—Table 3-12
- Gulf Coast region (west coast of Florida through Texas)—Table 3-13
- Pacific Northwest region (Alaska through Oregon)—Table 3-14
- Northern California waters (Klamath River through Morro Bay)—Table 3-15
- Southern California waters (Santa Monica Bay to Tijuana Estuary)— Table 3-16.

Family name	Common name	Scientific name
Finfish Species		
Anguillidae	American eel	Anguilla rostrata
Percichthyidae	Striped bass	Morone saxatilis
Pomatomidae	Bluefish	Pomatomus saltatrix
Sparidae	Scup	Stenotomus chrysops
Sciaenidae	Weakfish	Cynoscion regalis
Bothidae	Summer flounder	Paralichthys dentatus
	Four-spotted flounder	Paralichthys oblongus
Pleuronectidae	Winter flounder	Pseudopleuronectes americanus
	Yellowtail flounder	Limanda ferruginea
	American dab	Hippoglossoides platessoides
Shellfish Species		
Bivalves	Soft-shell clam	Mya arenaria Mercenaria mercenaria Arctica islandica Spisula solidissima Mytilus edulis
Crustaceans	American lobster Eastern rock crab	Homarus americanus Cancer irroratus

# Table 3-10. Recommended Target Species for Northeast Atlantic Estuaries and Marine Waters (Maine through Connecticut)

Family name	Common name	Scientific name
Finfish Species		
Anguillidae	American eel	Anguilla rostrata
Ictaluridae	Channel catfish	Ictalurus punctatus
	White catfish	Ictalurus catus
Percichthyidae	White perch	Morone americana
	Striped bass	Morone saxatilis
Pomatomidae	Bluefish	Pomatomus saltatrix
Sparidae	Scup	Stenotomus chrysops
Sciaenidae	Weakfish	Cynoscion regalis
	Spot	Leistomus xanthurus
	Atlantic croaker	Micropogonias undulatus
	Red drum	Sciaenops ocellatus
Bothidae	Summer flounder	Paralichthys dentatus
Pleuronectidae	Winter flounder	Pseudopleuronectes americanus
Shellfish Species		
Bivalves	Hard clam	Mercenaria mercenaria
	Soft-shell clam	Mya arenaria
	Ocean quahog	Arctica islandica
	Surf clam	Spisula solidissima
	Blue mussel	Mytilus edulis
	American oyster	Crassostrea virginica
Crustaceans	Blue crab	Callinectes sapidus
	American lobster	Homarus americanus
	Eastern rock crab	Cancer irroratus

## Table 3-11. Recommended Target Species for Mid-Atlantic Estuaries and Marine Waters (New York through Virginia)

Family name	Common name	Scientific name
infish Species		
Anguillidae	American eel	Anguilla rostrata
Ictaluridae	Channel catfish White catfish	lctalurus punctatus Ictalurus catus
Percichthyidae	White perch Striped bass	Morone americana Morone saxatilis
Sciaenidae	Spot	Leistomus xanthurus
	Atlantic croaker	Micropogonias undulatus
	Red drum	Sciaenops ocellatus
Bothidae	Southern flounder	Paralichthys lethostigma
	Summer flounder	Paralichthys dentatus
nellfish Species		
Bivalves	Hard clam	Mercenaria mercenaria
	American oyster	Crassostrea virginica
Crustaceans	West Indies spiny lobster	Panulirus argus
	Blue crab	Callinectes sapidus

## Table 3-12. Recommended Target Species for Southeast Atlantic Estuaries and Marine Waters (North Carolina through Florida)

Family name	Common name	Scientific name
Finfish Species		
Ictaluridae	Blue catfish	Ictalurus furcatus
	Channel catfish	Ictalurus punctatus
Ariidae	Hardhead catfish	Arius felis
Sciaenidae	Spotted seatrout	Cynoscion nebulosus
	Spot	Leistomus xanthurus
	Atlantic croaker	Micropogonias undulatus
	Red drum	Sciaenops ocellatus
Bothidae	Gulf flounder	Paralichthys albigutta
	Southern flounder	Paralichthys lethostigma
Shellfish Species		
Bivalves	American oyster	Crassostrea virginica
	Hard clam	Mercenaria mercenaria
Crustaceans	White shrimp	Penaeus setiferus
	Blue crab	Callinectes sapidus
	Gulf stone crab	Menippe adina
	West Indies spiny lobster	Panulirus argus

## Table 3-13. Recommended Target Species for Gulf of Mexico Estuaries and Marine Waters (West Coast of Florida through Texas)

Family name         Common name         Scientific name				
Finfish Species				
Embiotocidae	Redtail Surfperch	Amphistichus rhodoterus		
Scorpaenidae	Copper rockfish	Sebastes caurinus		
	Black rockfish	Sebastes melanops		
Bothidae	Speckled sanddab	Citharichthys stigmaeus		
	Pacific sanddab	Citharichthys sordidus		
Pleuronectidae	Starry flounder	Platichthys stellatus		
	English sole	Parophrys vetulus		
Salmonidae	Coho salmon	Onchorhynchus kisutch		
	Chinook salmon	Onchorhynchus tshawytscha		
Shellfish Species				
Bivalves	Blue mussel	Mytilus edulis		
	California mussel	Mytilus californianus		
	Pacific oyster	Crassostrea gigas		
	Horseneck clam	Tresus capax		
	Pacific littleneck clam	Protothaca staminea		
	Soft-shell clam	Mya arenaria		
	Manila clam	Venerupis japonica		
Crustaceans	Dungeness crab	Cancer magister		
	Red crab	Cancer productus		

## Table 3-14. Recommended Target Species for Pacific NorthwestEstuaries and Marine Waters (Alaska through Oregon)

Family name         Common name         Scientific name			
Finfish Species			
Triakidae	Leopard shark	Triakis semifasciata	
Sciaenidae	White croaker	Genyonemus lineatus	
Embiotocidae	Redtailed surfperch	Amphistichus rhodoterus	
	Striped seaperch	Embiotoca lateralis	
Scorpaenidae	Black rockfish	Sebastes melanops	
	Yellowtail rockfish	Sebastes flavidus	
	Bocaccio	Sebastes paucispinis	
Bothidae	Pacific sanddab	Citharichthys sordidus	
	Speckled sanddab	Citharichthys stigmaeus	
Pleuronectidae	Starry flounder	Platichthys stellatus	
	English sole	Parophrys vetulus	
Salmonidae	Coho salmon	Onchorhynchus kisutch	
	Chinook salmon	Onchorhynchus tshawytscha	
Shellfish Species			
Bivalves	Blue mussel	Mytilus edulis	
	California mussel	Mytilus californianus	
	Pacific littleneck clam	Protothaca staminea	
	Soft-shell clam	Mya arenaria	
Crustaceans	Dungeness crab	Cancer magister	
	Red crab	Cancer productus	
	Pacific rock crab	Cancer antennarius	

## Table 3-15. Recommended Target Species for Northern California Estuaries and Marine Waters (Klamath River through Morro Bay)

Family name         Common name         Scientific name		
Finfish Species		
Serranidae	Kelp bass	Paralabrax clathratus
	Barred sand bass	Paralabrax nebulifer
Sciaenidae	White croaker	Genyonemus lineatus
	Corbina	Menticirrhus undulatus
Embiotocidae	Black perch	Embiotoca jacksoni
	Walleye surf perch	Hyperprosopan argenteum
	Barred surfperch	Amphistichus argenteus
Scorpaenidae	California scorpionfish	Scorpaena guttata
	Widow rockfish	Sebastes entomelas
	Blue rockfish	Sebastes mystinus
	Bocaccio	Sebastes paucispinis
Pleuronectidae	Diamond turbot	Hypsopetta guttulata
	Dover sole	Microstomus pacificus
Shellfish Species		
Bivalves	Blue mussel	Mytilus edulis
	California mussel	Mytilus californianus
	Pacific littleneck clam	Protothaca staminea
Crustaceans	Pacific rock crab	Cancer antennarius
	Red crab	Cancer productus
	California rock lobster	Panulirus interruptus

Table 3-16. Recommended Target Species for Southern	California
Estuaries and Marine Waters (Santa Monica Bay to Tijuar	na Estuary)

The seven separate regional lists of target species recommended by the 1993 EPA Workgroup for estuarine/marine ecosystems were developed because of differences in species' geographic distribution and abundance and the nature of the regional fisheries and were developed based on a review of species used in the following national monitoring programs:

- National Dioxin Study (U.S. EPA)
- Section 301(h) Monitoring Program (U.S. EPA)
- National Status and Trends Program (NOAA)
- National Study of Chemical Residues in Fish (U.S. EPA).

Because some of these programs identified some fish and shellfish species that are not of commercial, sportfishing, or subsistence value, several additional literature sources identifying commercial and sportfishing species were also reviewed (Table 3-17). Some sources included information on seasonal distribution and abundance of various life stages (i.e., adults, spawning adults, juveniles) of fish and shellfish species. This information was useful in delineating seven regional estuarine/marine areas nationwide. The 1993 EPA Workgroup also reviewed fish and shellfish species cited in state consumption advisories for estuarine/marine waters (Appendix D). Each of the final regional lists of target species has been reviewed by state, regional, and national fisheries experts.

Use of two distinct ecological groups of organisms (shellfish and finfish) as target species in estuarine/marine systems is recommended. This permits monitoring of a wide variety of habitats, feeding strategies, and physiological factors that might result in differences in bioaccumulation of contaminants. Estuarine/marine species used in several national contaminant monitoring programs reviewed by the 1993 EPA Workgroup are compared in Table 3-18.

### 3.4.1 Target Shellfish Species

Selection of shellfish species (particularly bivalve molluscs) as target species received primary consideration by the 1993 EPA Workgroup because of the commercial, recreational, and subsistence value of shellfish in many coastal areas of the United States. Bivalve molluscs (e.g., oysters, mussels, and clams) are filter feeders that accumulate contaminants directly from the water column or via ingestion of contaminants adsorbed to phytoplankton, detritus, and sediment particles. Bivalves are good bioaccumulators of heavy metals (Cunningham, 1979) and polycyclic aromatic hydrocarbons (PAHs) and other organic compounds (Phillips, 1980; NOAA, 1987) and, because they are sessile, they may reflect local contaminant concentrations more accurately than more mobile crustacean or finfish species.

Three bivalve species—the blue mussel (*Mytilus edulis*), the California mussel (*Mytilus californianus*), and the American oyster (*Crassostrea virginica*)—were recommended and/or used in three of the national monitoring programs reviewed by the 1993 EPA Workgroup. Two other bivalve species—the soft-shell clam (*Mya arenaria*) and the Pacific oyster (*Crassostrea gigas*)—were also recommended and/or used in two national programs. Although no bivalve species was identified by name in state fish and shellfish consumption advisories (Appendix D), seven coastal states issued advisories in 1993 for unspecified bivalves or shellfish species that may have included these and other bivalve species. All three species are known to bioaccumulate a variety of environmental contaminants (Phillips, 1988). The wide distribution of these three species makes them useful for comparison within a state or between states sharing coastal waters (Figure 3-2). Because these three species met all of the selection criteria, they were recommended as target species for use in geographic areas in which they occur.

Geographic area	Source
Atlantic Coast	<ul> <li>National Marine Fisheries Service. 1987. Marine Recreational Fishery Statistics Survey, Atlantic and Gulf Coasts, 1986. Current Fishery Statistics Number 8392. National Oceanic and Atmospheric Administration, U.S. Department of Commerce, Rockville, MD.</li> <li>Leonard, D.L., M.A. Broutman, and K.E. Harkness. 1989. The Quality of Shellfish Growing Waters on the East Coast of the United States. Strategic Assessment Branch, National Oceanic and Atmospheric Administration, U.S. Department of Commerce, Rockville, MD.</li> <li>Nelson, D.M., M.E. Monaco, E.A. Irlandi, L.R. Settle, and L. Coston-Clements. 1991. Distribution and Abundance of Fishes and Invertebrates in Southeast Estuaries. ELMR Report No. 9. Strategic Assessment Division. National Oceanic and Atmospheric Administration, U.S. Department of Commerce, Rockville, MD.</li> <li>Stone, S.L., T.A. Lowery, J.D. Field, C.D. Williams, D.M. Nelson, S.H. Jury, M.E. Monaco, and L. Andreasen 1994. Distribution and Abundance of Fishes and Invertebrates in Southeast Estuaries in Mid-Altantic Estuaries. ELMR Rep. No. 12 NOAA/NOS Strategic Environmental Assessments Division, SIlver Spring, MD.</li> <li>Jury, S.H., J.D. Field, S.L. Stone, D.M. Nelson, and M.E. Monaco. 1994. Distribution and Abundance of Fishes and Invertebrates in North Atlantic Estuaries. ELMR Rep. No. 13. NOAA/NOS Strategic Environmental Assessments Division, SIlver Spring, MD.</li> </ul>
Gulf Coast	<ul> <li>National Marine Fisheries Service. 1987. Marine Recreational Fishery Statistics Survey, Atlantic and Gulf Coasts, 1986. Current Fishery Statistics Number 8392. National Oceanic and Atmospheric Administration, U.S. Department of Commerce, Rockville, MD.</li> <li>Broutman, M.A., and D.L. Leonard. 1988. The Quality of Shellfish Growing Waters in the Gulf of Mexico. Strategic Assessment Branch, National Oceanic and Atmospheric Administration, Rockville, MD.</li> <li>Monaco, M.E., D.M. Nelson, T.C. Czapla, and M.E. Patillo. 1989. Distribution and Abundance of Fishes and Invertebrates in Texas Estuaries. ELMR Report No. 3. Strategic Assessment Branch, National Oceanic and Atmospheric Administration, U.S. Department of Commerce, Rockville, MD.</li> <li>Williams, C.D., D.M. Nelson, M.E. Monaco, S.L. Stone, C. Iancu, L. Coston-Clements, L.R. Settle, and E.A. Irlandi. 1990. Distribution and Abundance of Fishes and Invertebrates in Eastern Gulf of Mexico Estuaries. ELMR Report No. 6. Strategic Assessment Branch, National Oceanic and Atmospheric Administration, U.S. Department of Commerce, Rockville, MD.</li> <li>Czapla, T.C., M.E. Patillo, D.M. Nelson, and M.E. Monaco. 1991. Distribution and Abundance of Fishes and Invertebrates in Central Gulf of Mexico Estuaries. ELMR Report No. 7. Strategic Assessment Branch, National Oceanic and Atmospheric Administration, U.S. Department of Commerce, Rockville, MD.</li> <li>Czapla, T.C., M.E. Patillo, D.M. Nelson, and M.E. Monaco. 1991. Distribution and Abundance of Fishes and Invertebrates in Central Gulf of Mexico Estuaries. ELMR Report No. 7. Strategic Assessment Branch, National Oceanic and Invertebrates in Gulf of Mexico Estuaries ELMR Rep. No. 10. NOAA/NOS Strategic Environmental Assessments Division, Rockville, MD.</li> <li>Patillo, M.E., T.E. Czapla, D.M. Nelson, and M.E. Monaco. 1997. Distribution and Abundance of Fishes and Invertebrates in Gulf of Mexico Estuaries. Vol. II: Species Life History Summaries. ELMR Rep. No. 14. NOAA/NOS Strategic Environmental Assessme</li></ul>
West Coast	<ul> <li>National Marine Fisheries Service. 1987. Marine Recreational Fishery Statistics Survey, Pacific Coast, 1980.</li> <li>Current Fishery Statistics Number 8393. National Oceanic and Atmospheric Administration, U.S. Departmer of Commerce, Rockville, MD.</li> <li>Leonard, D.L., and E.A. Slaughter. 1990. The Quality of Shellfish Growing Waters on the West Coast of the United States. Strategic Assessment Branch, National Oceanic and Atmospheric Administration, U.S.</li> <li>Department of Commerce, Rockville, MD.</li> <li>Monaco, M.E., D.M. Nelson, R.L. Emmett, and S.A. Hinton. 1990. Distribution and Abundance of Fishes and Invertebrates in West Coast Estuaries. Volume I: Data Summaries. ELMR Report No. 4. Strategic Assessment Branch, National Oceanic and Atmospheric Administration, Brockville, MD.</li> <li>Emmett, R.L., S.A. Hinton, S.L. Stone, and M.E. Monaco. 1991. Distribution and Abundance of Fishes and Invertebrates in West Coast Estuaries. Volume II: Life History Summaries. ELMR Report No. 8. Strategic Environmental Assessment Division, Rockville, MD.</li> </ul>

## Table 3-17. Sources of Information on Commercial and SportfishingSpecies in Various Coastal Areas of the United States

	U.S. EPA National Dioxin Study <sup>a</sup>	NOAA Status and Trends	U.S. EPA 301(h) Program	U.S. EPA NSCRF⁵
FINFISH				
Family <i>Acipenseridae</i> White sturgeon ( <i>Acipenser transmontanus</i> )				•
Family <i>Ariidae</i> Hardhead catfish <i>(Arius felis)</i>				•
Family <i>Percichthyidae</i> White perch <i>(Morone americana)</i>				•
Family <i>Pomatomidae</i> Bluefish <i>(Pomatomus saltatrix)</i>				•
Family <i>Lutjanidae</i> Red snapper <i>(Lutjanus campechanus)</i>				•
Family <i>Sparidae</i> Sheepshead (Archosargus probatocephalus)				•
Family (Sciaenidae) Spotted seatrout (Cynoscion nebulosus) Weakfish (Cynoscion regalis) Spot (Leiostomus xanthurus) White croaker (Genyonemus lineatus) Atlantic craoker (Micropogonias undulatus) Black drum (Pogonias cromis) Red drum (Sciaenops ocellatus)			•	
Family <i>Serranidae</i> Barred sand bass <i>(Paralabrax nebulifer)</i>		•		
Family <i>Mugilidae</i> Striped mullet <i>(Mugil cephalus)</i>				•
Family <i>Bothidae</i> Southern flounder ( <i>Paralichthys lethostigma</i> )				•
Windowpane flounder (Scophthalmus aquosus)		•		
Family Pleuronectidae Pacific sanddab (Citharichthys sordidus) Flathead sole (Hippoglossoides elassodon) Diamond turbot (Hypsopsetta guttulata) Starry flounder (Platichthys stellatus) Hornyhead turbot (Pleuronichthys verticalis) Winter flounder (Pseudopleuronectes americanus) English sole (Parophrys vetulus)		•	•	•
Dover sole ( <i>Microstomus pacificus</i> ) See notes at end of table.				(continued)

# Table 3-18. Estuarine/Marine Species Used in Several National Fish and Shellfish Contaminant Monitoring Programs

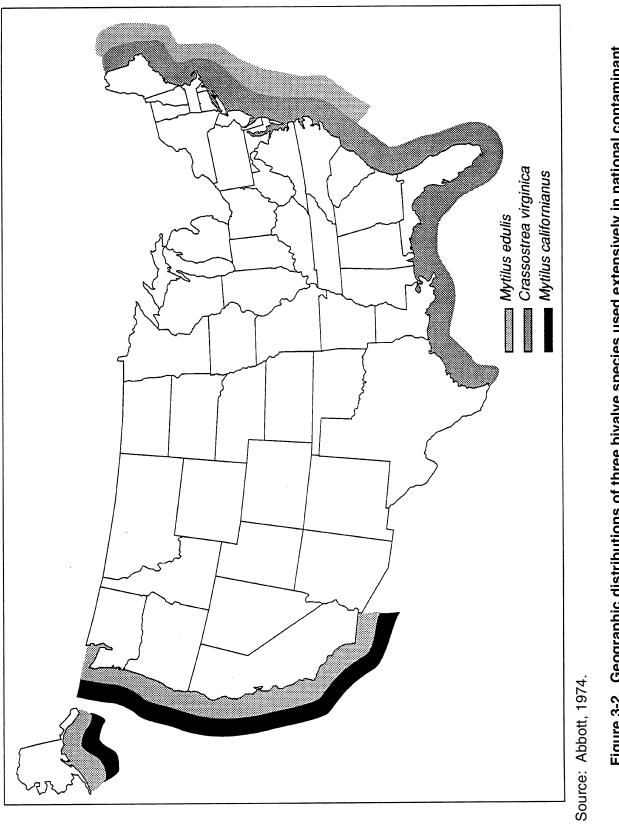
See notes at end of table.

(continued)

	U.S. EPA National	NOAA Status and	U.S. EPA 301(h)	U.S. EPA
	Dioxin Study <sup>a</sup>	Trends	Program	NSCRF⁵
SHELLFISH				
Bivalves Hard clam (Mercenaria mercanaria) Soft-shell clam (Mya arenaria) Ocean quahog (Arctica islandia) Surf clam (Spisula solidissima) Blue mussel (Mytilus edulis) California mussel (Mytilus californianus) American oyster (Crassostrea virginica) Hawaiian oyster (Ostrea sandwichensis) Pacific oyster (Crassostrea gigas) Bent-nosed macoma (Macoma nasuta) Baltic macoma (Macoma baltica) White sand macoma (Macoma secta)	•	• • •		•
Crustaceans American lobster (Homarus americanus) West Indies spiny lobster (Panulirus argus) California rock lobster (Panulirus interruptus) Hawaiian spiny lobster (Panulirus penicillatus) Eastern rock crab (Cancer irroratus) Dungeness crab (Cancer magister) Pacific rock crab (Cancer antennarius) Yellow crab (Cancer anthonyi) Red crab (Cancer productus)				•

### Table 3-18. (continued)

NSCRF = National Study of Chemical Residues in Fish.
 <sup>a</sup> Only freshwater finfish were identified as target species; bivalves were identified as estuarine/marine target species.
 <sup>b</sup> Species listed were those collected at more than one site nationally; *Salmonidae* were not listed because they were included on freshwater lists.





In addition, several species of edible clams were added to the various estuarine/ marine target species lists based on recommendations received from specific state and regional fisheries experts.

Crustaceans are also recommended as target species for estuarine/marine sampling sites. Many crustaceans are bottom-dwelling and bottom-feeding predator and/or scavenger species that are good indicators of contaminants that may be biomagnified through several trophic levels of the food web. Several species of lobsters and crabs were recommended in one national monitoring program, and the Dungeness crab was recommended in two national monitoring programs (Table 3-18). These crustaceans, although of fishery value in many areas, are not as widely distributed nationally as the three bivalve species (Figure 3-2). However, they should be considered for selection as target species in states where they are commonly consumed.

Only two crustaceans—the American lobster (*Homarus americanus*) and the blue crab (*Callinectes sapidus*)—were specifically identified in state advisories (RTI, 1993). However, in 1993, seven coastal states reported advisories in estuarine/ marine waters for unspecified shellfish species that may have included these and other crustacean species (Table 3-19). All of the shellfish species cited in state advisories are included as EPA-recommended target species on the appropriate estuarine/marine regional lists.

### 3.4.2 Target Finfish Species

Two problems were encountered in the selection of target finfish species for monitoring fish tissue contamination at estuarine/marine sites regionally and nationally. First is the lack of finfish species common to both Atlantic and Gulf Coast waters as well as Pacific Coast waters. Species used in several federal fish contaminant monitoring programs are compared in Table 3-18. Members of the families Sciaenidae (seven species), Bothidae (two species), and Pleuronectidae (eight species) were used extensively in these programs. Bottomdwelling finfish species (e.g., flounders in the families Bothidae and Pleuronectidae) may accumulate high concentrations of contaminants from direct physical contact with contaminated bottom sediments. In addition, these finfish feed on sedentary infaunal or epifaunal organisms and are at additional risk of accumulating contaminants via ingestion of these contaminated prey species (U.S. EPA, 1987a). For finfish species, two Atlantic coast species, spot (Leiostomus xanthurus) and winter flounder (Pseudopleuronectes americanus), are recommended and/or used in three of the national monitoring programs, and the Atlantic croaker (Micropogonias undulatus) is recommended and/or used in two national monitoring programs. Three Pacific coast species, Starry flounder (Platichthys stellatus), English sole (Parophrys vetulus), and Dover sole (Microstomus pacificus), are recommended or used in two of the national monitoring programs.

Species group name	Common name	Scientific name	Number of states with advisories in 1993	Number of states with advisories in 1998
Finfish				
Percichthyidae	Striped bass White perch	<i>Morone saxatilis</i> Morone americana	<b>5</b> 3	<b>6</b> 3
Centrarchidae	Largemouth bass Smallmouth bass	Micropterus salmoides Micropterus dolomieui	0 0	3 1
lctaluridae	White catfish Channel catfish	lctalurus catus Ictalurus punctatus	4 5	2 2
Anguillidae	American eel	Anguilla rostrata	6	5
Elopidae	Ladyfish	Elops saurus	0	1
Carangidae	Crevalle jack	Caranx hippos	0	1
Pomatomidae	Bluefish	Pomatomus saltatrix	4	6
Labridae	Tautog	Tautoga onitis	0	1
Sparidae	Scup	Stenotomus chrysops	0	1
Sciaenidae	Spotted sea trout Atlantic croaker Red drum Black drum Silver perch	Cynoscion nebulosus Micropogonias undulatus Sciaenops ocellatus Pogonias cromis Bairdiella chrysoura	0 0 0 0 0	2 1 1 1 1
Scombridae	King mackerel Spanish mackerel	Scomberomorus cavalla Scomberomorus maculatus	0 0	5 1
Ariidae	Gafftopsail catfish	Bagre marinus	0	1
Belonidae	Atlantic needlefish	Strongylura marina	1	1
Serranidae	Kelp bass	Paralabrax clathratus	1	1
Sciaenidae	Black croaker <b>White croaker</b> Queenfish <b>Corbina</b>	Cheilotrema saturnum <b>Genyonemus lineatus</b> Seriphus politus <b>Menticirrhus undulatus</b>	1 1 1	1 1 1
Shellfish				
Crustaceans <sup>c</sup>	American lobster Blue crab	Homarus americanus Callinectes sapidus	1 3	5 4

### Table 3-19. Principal Estuarine/Marine Fish and Shellfish Species Cited in State Consumption Advisories<sup>a,b</sup>

а Species in boldface are EPA-recommended target species for regional estuarine/marine waters (see Tables 3-10 through 3-16). <sup>b</sup> Many coastal states issued advisories for fish and shellfish species and thus did not identify specific finfish and

shellfish species in their advisories.

с Eight coastal states (California, Georgia, Hawaii, Louisiana, Massachusetts, North Carolina, Texas, and Washington) and the U.S. territory of American Samoa report advisories for unspecified shellfish or bivalve species.

Sources: RTI, 1993, EPA 1999a (NLFWA).

Second, because some estuarine/marine finfish species are highly migratory, harvesting of these species may be restricted to certain seasons because sexually mature adult fish (i.e., the recommended size for sampling) may enter the estuaries only to spawn. EPA recommends that neither spawning populations nor undersized juvenile stages be sampled in fish contaminant monitoring programs. Sampling of target finfish species during their spawning period should be avoided as contaminant tissue concentrations may decrease during this time (Phillips, 1980) and because the spawning period is generally outside the legal harvest period. **Note:** Target finfish species may be sampled during their spawning period if the species can be legally harvested at this time. Sampling of undersized juveniles of species that use estuaries as nursery areas is precluded by EPA's recommended monitoring strategy because juveniles may not have had sufficient time to bioaccumulate contaminants or attain harvestable size.

Because of these problems, the 1993 EPA Workgroup consulted with regional and state fisheries experts and reviewed the list of state fish consumption advisories and bans to determine which estuarine/marine finfish species should be recommended as target species. As shown in Table 3-19, the largest number of states issuing advisories in 1993 for specific estuarine and marine waters did so for the American eel (6), channel catfish (5), striped bass (5), bluefish (4), white catfish (4), and white perch (3). Several other estuarine/marine species were cited in advisories for one state each (Table 3-19). Many coastal states did not identify individual finfish species by name in their advisories (see Appendix D); however, almost all of the species that have been cited in state advisories are recommended as target species by EPA (see Tables 3-10 through 3-16). The listing of estuarine fish and shellfish cited in state advisories in 1998 is also shown in Table 3-19.

These seven regional lists of recommended estuarine/marine target species are provided to give guidance to states on species commonly consumed by the general population. state personnel, with their knowledge of site-specific fisheries and human consumption patterns, must be the ultimate judge of the species selected for use in estuarine/marine fish contaminant monitoring programs within their jurisdiction.